REVIEW Open Access

# G-protein coupled receptor, PI3K and Rho signaling pathways regulate the cascades of Tau and amyloid-β in Alzheimer's disease



Smita Eknath Desale<sup>1,2†</sup>, Hariharakrishnan Chidambaram<sup>1,2†</sup> and Subashchandrabose Chinnathambi<sup>1,2\*</sup>

## **Abstract**

Alzheimer's disease is a progressive neurodegenerative disease characterized by the presence of amyloid- $\beta$  plaques in the extracellular environment and aggregates of Tau protein that forms neurofibrillary tangles (NFTs) in neuronal cells. Along with these pathological proteins, the disease shows neuroinflammation, neuronal death, impairment in the immune function of microglia and synaptic loss, which are mediated by several important signaling pathways. The PI3K/Akt-mediated survival-signaling pathway is activated by many receptors such as G-protein coupled receptors (GPCRs), triggering receptor expressed on myeloid cells 2 (TREM2), and lysophosphatidic acid (LPA) receptor. The signaling pathway not only increases the survival of neurons but also regulates inflammation, phagocytosis, cellular protection, Tau phosphorylation and A $\beta$  secretion as well. In this review, we focused on receptors, which activate PI3K/Akt pathway and its potential to treat Alzheimer's disease. Among several membrane receptors, GPCRs are the major drug targets for therapy, and GPCR signaling pathways are altered during Alzheimer's disease. Several GPCRs are involved in the pathogenic progression, phosphorylation of Tau protein by activation of various cellular kinases and are involved in the amyloidogenic pathway of amyloid- $\beta$  synthesis. Apart from various GPCR signaling pathways, GPCR regulating/ interacting proteins are involved in the pathogenesis of Alzheimer's disease. These include several small GTPases, Ras homolog enriched in brain, GPCR associated sorting proteins,  $\beta$ -arrestins, etc., that play a critical role in disease progression and has been elaborated in this review.

Keywords: Alzheimer's disease, Tau, PI3K/Akt, GPCR, TREM2, LPA, Rho GTPase

#### Introduction

Alzheimer's disease (AD) is a progressive neurodegenerative disease that causes degeneration, neuronal loss and dementia [1]. Intracellular aggregates of Tau and extracellular amyloid- $\beta$  plaques are the pathological hallmarks of AD [2, 3]. Amyloid precursor proteins (APP) are membrane proteins that are cleaved by  $\beta$ - and  $\gamma$ -

secretases to form amyloid- $\beta$  fragments under AD conditions. Tau is a microtubule-associated protein that binds tubulin and promotes microtubule stability [4, 5]. In AD brains, Tau is highly disordered and self-aggregates to form filaments. Several post-translational modifications of Tau are reported in AD brain that majorly includes phosphorylation, acetylation, glycation, glycosylation, methylation, sumoylation, etc., which limit its affinity to bind to microtubules [6–9]. In recent research days, Tau based therapy is attaining importance over amyloid- $\beta$  and acetylcholine hypotheses [10–13]. Several drugs and molecules are being reported that inhibits Tau pathology, i.e., reversal of post-translational

<sup>&</sup>lt;sup>1</sup>Neurobiology Group, Division of Biochemical Sciences, CSIR-National Chemical Laboratory (CSIR-NCL), Dr. Homi Bhabha Road, Pune 411008, India <sup>2</sup>Academy of Scientific and Innovative Research (AcSIR), Ghaziabad 201002, India



© The Author(s). 2021 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

<sup>\*</sup> Correspondence: s.chinnathambi@ncl.res.in

<sup>&</sup>lt;sup>†</sup>Smita Eknath Desale and Hariharakrishnan Chidambaram contributed equally to this work.

Desale et al. Molecular Biomedicine (2021) 2:17 Page 2 of 18

modifications, aggregation inhibition and disaggregation of pre-formed filaments, etc. [10-26]. In Alzheimer's disease, several pathological signaling cascades are activated on exposure to these aggregated proteins which interacts with several membrane receptors of neuronal and glial cells. Signaling cascades mediated by various cell surface receptors in neurons as well as glial cells is a highly dynamic and interconnected system. Cellular processes such as phagocytosis, cell motility, immune function, metabolism and polarity are tightly regulated by cellsurface receptors and their signaling pathways [26–32]. Most of the cell surface receptors have an extracellular ligand-binding domain, a transmembrane region and an intracellular effector region, which is usually essential in transmitting the extracellular signal to the cell [33]. The intracellular transmission of signals is associated with kinase activities as a response to ligand binding.

One of which is receptor tyrosine kinase (RTK) that consists of intracellular tyrosine kinase domain irrespective of extracellular ligand-binding domain and a transmembrane domain [34]. Cytokine receptor type 1 and 2, multisubunit antigen receptors of B, T as well as macrophage cells comes under RTK family [35, 36]. Apart from RTKs, GPCRs are the largest family of surface receptors, which regulate physiological functions such as glucose and lipid metabolism, homeostasis, neurotransmission, immune responses, and other cellular functions [37]. GPCR consists of extracellular domain, a transmembrane region and intracellular domain that transmit the signal via various G-proteins (small GTPases). Gα and Gby subunits activate downstream effector pathways such as cAMP, protein kinase C (PKC), Ca<sup>2+</sup> influx, PI3K, MAPK as well as ERK pathway [38]. In the downstream signaling cascades of receptors, protein kinase C (PKC) signaling acts as a feedback signaling for RTKs like insulin receptors, Met, Kit, EGFR, etc. Increase in intracellular Ca<sup>2+</sup> levels by the production of diacylglycerol (DAG) via PLCy is a well-known mechanism to activate PKC pathway [39]. The downstream signaling of GPCRs include activation of small GTPase Ras, PI3Ks, PKC, tyrosine kinase and arrestin, which follows ERK1/2 signaling pathway [40]. In Alzheimer's disease, several GPCRs such as calcium sensing receptors, muscarinic acetyl choline receptors are activated by amyloid-β and Tau exposure and are involved in pathological signaling process that ultimately leads to neurodegeneration [41-44]. The different roles of GPCRs in amyloid-β synthesis, Tau phosphorylation, and Tau aggregation are clearly been reviewed [45-47]. Apart from the role of different GPCRs and their signaling through various G-protein dependent and independent pathways, there are various other GPCR regulating/ interacting proteins that take part in cellular signaling pathways for their growth and development. These proteins include small GTPases, Ras homolog enriched in brain (Rheb), GPCR associated sorting proteins (GASPs), receptor activity modifying proteins (RAMPs), *etc.* Among various small GTPases, the Ras superfamily of GTPases is well studied and are classified as five different sub-families: Ras, Rho, Rab, Ran, and Arf GTPases [48].

## PI3K/Akt signaling

The PI3K/Akt/mTOR is the central pathway for cell metabolism, migration, immune function, and cell survival which is governed by many cell surface receptors. It involves formation of 3'- phosphorylated inositol phosphate, which activates Akt or mTOR ahead of PI3K activation. GPCRs activate PI3Kγ by regulating its catalytic subunit p110, the chemokine-induced cell migration of immune cells-mediated by this pathway [49]. In this study, we further discuss majorly the PI3K/Akt cell survival signaling and the other signaling partners associated with GPCRs.

# PI3K/Akt signaling-mediated by GPCRs

GPCRs are the type of integral membrane proteins, which transmit extracellular signals to intracellular space. These receptors form a superfamily of proteins, which bear a similar structural motif of seven transmembrane (TM) helices and are connected to three extracellular and intracellular loops [50]. The extracellular region of receptors act as an access point for ligands, and those include a range of molecules such as amines, lipids, proteins, peptides, nucleotides and even photons [51]. TM regions act as a structural core, which binds to ligands and transmits information to the intracellular space, via conformational changes. Intracellular region acts as a platform, which provides an interface for cytosolic signaling molecules. Despite of structural similarities, the signal-transducing mechanism varies as G protein-dependent and G protein-independent signaling pathways. The GPCRs significantly interact with G proteins while exerting their effector functions. G proteins that are coupled to GPCR include  $G_s$ ,  $G_{g/11}$ ,  $G_{12/13}$ , and G<sub>i</sub> [52, 53]. Stimulation of GPCR not only activates heterotrimeric G protein but also regulate intracellular effector signaling pathways such as activation of phospholipases, mitogen activated protein-kinases (MAPK), opening/closing of ion channels, and activation/inhibition of adenylyl and guanylyl cyclases [54, 55]. Various GPCRs are directly associated with small Gproteins and efficiently modulate downstream GPCR signaling network. GPCRs widely activate PI3K/Akt pathway via Gα and Gβγ, whereas Gβγ directly binds PI3K and activates the heterotrimeric protein consisting of either p110 $\beta$  or p110 $\gamma$  [55]. Along with authentic agonist, GPCR can recognize and respond to a wide array of ligands, which are known to activate PI3K/Akt signaling

Desale et al. Molecular Biomedicine (2021) 2:17 Page 3 of 18

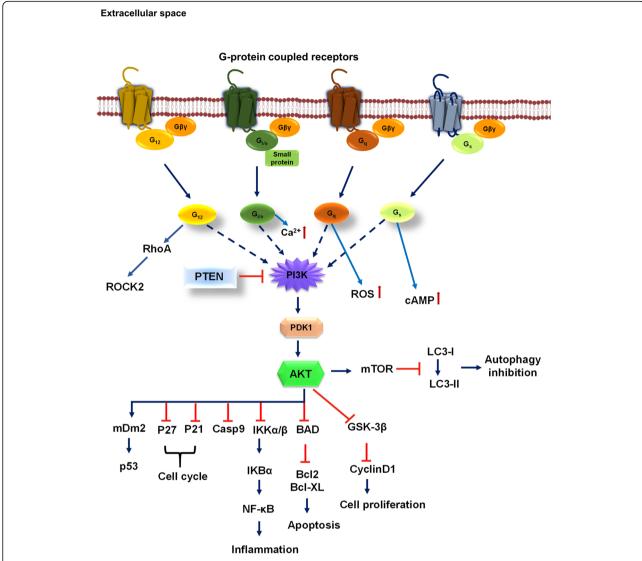
pathway. PI3K/Akt signaling is crucial since it regulates cell survival as well as metabolism [56]. Cells respond to a wide variety of stimuli via both GPCR and receptortyrosine kinases (RTK) under physiological conditions. Significant role of PI3K/Akt in cell proliferation, migration, apoptosis carried out by GPCR alone or in association with RTKs, is poorly recognized [57]. Human serotonin 5-HT receptor activates PI3K/Akt and MAPK signaling similar to VEGF via GPCR, which is important in angiogenesis [58]. Serotonin 5-HT receptor might activate dopaminergic neuron activity in a regiondependent manner via activating PI3K/Akt signaling through GPCR and extensively contributes to neurological functions. GPCR/PI3K/Akt signaling cascade is believed to be an important therapeutic target to understand neurodegenerative diseases [56]. Several medicinal herbs have been studied to initiate neuroprotective effects via PI3K/Akt pathway that include curcumin, danshensu, puerarin, etc [56]. Docosahexaenoic acid (DHA), a polyunsaturated fatty acid directly influences the oligomerization of GPCR by changing the cell membrane composition. GPCR signaling is modulated by DHA and it is observed to increase neuronal survival by phosphorylating Akt [59, 60]. Ratios of n-6/n-3 polyunsaturated fatty acids (PUFA) significantly influence PI3K/Akt signaling and inhibits inflammation [61]. The free fatty acids can increase glucose uptake by cell via PI3K/Akt signaling [62]. On activation, PI3K generates secondary lipid mediator phosphatidylinositol 3,4,5- triphosphate (PIP3), which phosphorylates Thr308 on Akt via phosphoinositide-dependent kinase 1 and at Ser473 via rapamycin complex 2. Dual phosphorylation of Akt is necessary for complete activation, however Thr308 is indicative of PI3K activity [63, 64]. Up-to date, several GPCR activating ligands have been identified such as sphingosine 1-phosphate, lysophosphatidic acid, stromal cell-derived factor, prostaglandin E2, carbachol, isoproterenol, thyroid-stimulating hormone (TSH), follicle stimulating hormone (FSH), and luteinizing hormone (LH)/choriogonadotropin (CG), etc. [57]. Activation of PI3K signaling pathway is majorly carried out in response to extracellular signal which is mediated by cellsurface receptors such as GPCRs, RTKs, integrin, growth factor receptors, where Akt plays a crucial role in activating this pathway [65]. The receptor-mediated activaof PI3K results in the production phosphatidylinositol 3,4,5- triphosphate (PIP3) from phosphatidylinositol 4,5 bisphosphate (PIP2). PIP3 can be reversed to PIP2 by the enzyme, phosphatase and tensin homolog (PTEN) and SH-domain-containing inositol polyphosphate 5-phosphatase (SHIP). Both Akt and phosphoinositide-dependant kinase (PDK) are enriched at the plasma membrane via PIP3 through their pleckstrin homology domain [66] (Fig. 1).

PI3K/Akt/GSK-3ß pathway plays a crucial role in Alzheimer's disease (AD), as it is one of the leading causes of Tau hyperphosphorylation. Glycogen synthase kinase-3β (GSK-3β) regulates the neuronal stress response via CREB, which regulates BDNF an important neuropeptide for memory retention, synaptic plasticity, etc. [55, 67]. Tau hyperphosphorylation and aggregation is generally led by several kinases, which include GSK3B, CDK-5, p38, p38-MAPK etc. PI3K/Akt pathway impairment majorly deals with the increased activity of GSK-3ß and hence Tau aggregation, which ultimately forms neurofibrillary tangles (NFTs) [68, 69]. Aβ-induced hyperphosphorylation of Tau is mediated through defective Akt/ GSK- $3\beta$  pathway, which provides a therapeutic link [70]. Anomalously, downregulation of Akt and subsequent elevated GSK-3β activity is related to brain dysfunctions, since Akt is a key regulator of cell survival and apoptosis [68]. Beside several signaling pathways, GPCRs are found to bind to β- and y-secretase, the key enzymes in amyloid protein precursor (APP) processing and amyloid plaque deposition [46]. In the GPCR superfamily, M1mAChR, A<sub>2A</sub> receptor, and δ-opioid receptor are known to influence BACE1 activity, which is important Aβ production. Along with receptors, GPCRassociated sorting proteins (GASPs), small G proteins such as Rabs, ADP-ribosylation factor 6 (ARF6) mediate BACE1 activity [47]. Muscarinic acetylcholine receptor (mAChR), especially M1 and M3 are found to increase soluble APP *via* α-secretase activity and reduces Aβ production. The release of sAPPα is assumed to be *via* PKC activation or interaction with calcium released via IP3 pool and DAG [71]. The opioid system, which is involved in neuroendocrine function as well as immune response is associated with abnormal Aβ production upon disrupted signaling. The enhancement of BACE1 and secretases activity are also regulated by opioid signaling of delta-opioid receptor (DOR) and naltrindole (NTI) [72, 73]. Adenosine receptor present on glia and neurons function in synaptic transmission and neuronal excitability. Adenosine A1 receptor found to be involved in Tau phosphorylation and also in Aβ production. These receptors are found to be colocalized with NFTs and amyloid plaques in the AD brain [45, 74]. Along with GPCRs, small GTPases such as Ras, Rac/cdc42/ Rho, Rab, Sar1/ARF and Ran family have a significant role in AD pathogenesis [75] (Fig. 1).

### PI3K/Akt survival signaling pathway by TREM2 receptor

Triggering receptor expressed on myeloid cells 2 (TREM2) that belongs to immunoglobulin superfamily is an innate immune receptor, which is exclusively expressed in myeloid cells such as immature dendritic cells, tissue macrophages, microglia and osteoclasts. The structure comprises of extracellular single v-type immunoglobulin

Desale et al. Molecular Biomedicine (2021) 2:17 Page 4 of 18



domain, short ectodomain, transmembrane region and short cytoplasmic tail [76, 77]. TREM2 contributes to various cellular processes such as regulation of inflammatory cytokine production, phagocytosis, migration, proliferation, etc. [78]. With recent studies, it has been shown that TREM2 is associated with the adaptor protein, DNAX activating protein 12 (DAP12) and DAP10. Upon ligand binding, these adaptor proteins undergo phosphorylation to activate downstream signaling cascades. DAP12 is known to mediate activation of spleen tyrosine kinase (Syk), and DAP10 propagates the signal by recruiting

PI3K. TREM2 binds to DAP12 or DAP10 and forms heterodimers [79]. DAP10 is critical in the activation of Akt and extracellular signal-regulated kinase (ERK), whereas DAP12 is essential for Ca<sup>2+</sup> mobilization [79, 80]. DAP12 is a type I transmembrane adaptor protein, which shares immunoreceptor tyrosine-based activation motifs (ITAM) in cytoplasmic domain, charged acidic transmembrane region, and also bears capacity to recruit src-homology domain-2 (SH2) on tyrosine phosphorylation. DAP12 is homodimeric, where cysteine residues (Cys33 and Cys35) at extracellular domain allow homodimer formation.

Desale et al. Molecular Biomedicine (2021) 2:17 Page 5 of 18

Phosphorylation of tyrosine residues on ITAM by Src protein tyrosine kinase provides docking site for SH2 domain such as spleen tyrosine kinase (Syk) [81-84]. TREM2 is activated through a wide variety of ligands such as bacterial products, lipoproteins (LDL), phospholipids (phosphatidylserine, cardiolipin), glycolipids, lipopolysaccharides (LPS), APOE and AB [85, 86]. Similarly, clearance of extracellular bacteria carried out through pattern recognition receptors (PRR), TREM2 and its adaptor protein DAP12 implement binding and uptake of a varied class of gram-negative and positive bacteria [87]. The activation of TREM2/DAP12 successively recruits PI3K after interaction between DAP12 and p85 subunit of PI3K, suggesting its involvement in TREM2 signaling [88]. PI3K targets Akt activation that further phosphorylates downstream targets, which include IKK, p21, caspase 9 and mTOR, depending upon the response. TREM2-mediated activation of PI3K/Akt is involved in reactive oxygen species (ROS) generation necessary to kill the internalized pathogens in macrophages [89]. The signaling pathway controls microglial survival by proliferation and reduced apoptosis by stabilizing β-catenin via Akt/GSK3β mechanism, which initiates the expression of responsible genes Cyclin D1, c-Myc and Bcl-2 [90]. TREM2 upregulates expression of CCR7, promotes chemokine-mediated response and enhances phagocytosis via ERK-dependent pathway. The signaling pathway is a legitimate mediator to balance proinflammatory and anti-inflammatory activity by microglia [78]. TREM2 signaling is directed towards antiinflammatory response, which profusely antagonizes TLR-4-mediated inflammatory response by modulating JNK and NF-κB pathway [91]. TREM2 and DAP12 overexpression is observed to suppress the IL-6 and IL-1β response, anticipate the role of TREM2 signaling in the suppression of LPS-mediated inflammatory response [78, 92]. Further, the capacity of TREM2-DAP12 expression to influence the phagocytosis has been linked with enrichment of microglia-mediated cytoskeleton rearrangements. pathway follows tyrosine phosphorylation of DAP12, Syk, Rac1 and cdc42. DAP12 signal mediates through Vav guanine nucleotide exchange factor family protein Vav2 and Vav3, which have the potential of activating Rho, Rac and cdc42. The pathway to exerbate phagocytosis via TREM2-DAP12 includes Src-Syk-Vav2/3-Rac1/cdc42-Arp2/3, which initiates necessary cytoskeleton rearrangement for phagocytosis [93, 94]. Hence, TREM2 mediated PI3K/Akt activation is a key pathway to control inflammatory response, apoptosis, survival pathway, and phagocytosis by microglia [81, 95] (Fig. 2).

# TREM2-mediated signaling in Alzheimer's disease

TREM2 is constitutively expressed in microglia which are surrounded by amyloid plaque for the phagocytic clearance in AD. Amyloid- $\beta$  oligomers found to bind

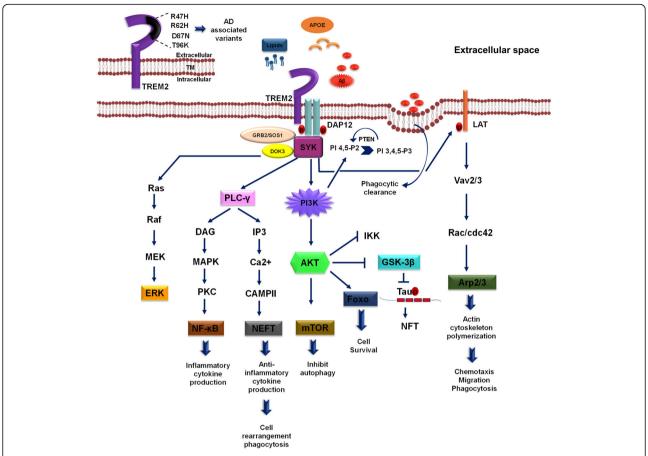
with TREM-2 with nanomolar affinity, however ADrelated mutations in TREM2, reduce its affinity towards binding and clearance of Aβ oligomers [96]. Aβ oligomers induce TREM2-mediated activation of microglial signaling pathways and clustering of microglia around the plaque regions in brain [97]. The dysfunction of TREM2-DAP12 axis is one of the major reason for impaired clearance of plagues, elevated pro-inflammatory activity by microglia in AD [98]. The heterozygous diseased mutants R47H and R62H of TREM2 recently has been identified as a risk factor for AD [99, 100] (Fig. 2). The TREM2 diseased variant drastically impedes the phagocytosis and the capacity to avert NF-κB activation in microglia. Aß oligomers have a high affinity towards TREM2, which activate nuclear factor for activated T cells (NFAT). However, in TREM2 AD variant, a partial loss of TREM2 functioning has been observed [101]. TREM2 deficiency have been observed to reduce microglial viability, proliferation, microgliosis and decrease stability of β-catenin, an important factor of canonical Wnt signaling pathways. TREM-2-mediated stability of β-catenin is maintained by inhibiting its degradation via Akt/GSK-3ß pathway. TREM2 knockdown decreases the level of AKT<sup>S473</sup> and GSK-3β<sup>S9</sup>, which indicates that TREM-2 mediated stabilization of β-catenin is *via* AKT/ GSK-3β signaling pathway [90, 102]. Disrupted PI3K/ Akt signaling affects downstream factors as well, such as mTOR, a regulator of autophagy. Decreased activation of mTOR elevates excessive accumulation of autophagosomes, metabolic impairment and inability of microglia to clear Aβ plaques [103]. The recent finding of diseaseassociated microglia (DAM) phenotype importantly stresses upon the TREM2-PI3K-Akt signaling axis for its development [104]. TREM2 mutation dramatically hampers TREM-2-mediated phagocytosis, which is mediated by SYK/PI3K/Akt/PLCy signaling pathway. The mutation also decreases the inhibition of NF-KB-mediated inflammatory response [95]. Hence, TREM2 deliberately reduce neuroinflammation and improves neuronal functions via PI3K/Akt-mediated signaling cascade [105]. Effect of TREM2 deficiency also elevates Tau pathology, but the mechanism is still needs to be addressed [106] (Fig. 2).

# Role of GPCR regulating/ interacting proteins in Alzheimer's disease

# **Small GTPases**

Small GTPases belongs to the group of G proteins that are capable of hydrolysing GTPs to form GDPs [48]. Small GTPases constitute more than 100 different proteins and are broadly classified into five families, i.e. Ras family of GTPases, Rab family of GTPases, Rho family of GTPases, Ran GTPases and Arf GTPases [48]. GTPases are regulated by three main regulatory proteins, Guanine

Desale et al. Molecular Biomedicine (2021) 2:17 Page 6 of 18



**Fig. 2** PI3K/Akt signaling-mediated by TREM2. DAP12-mediated TREM2 signaling have been discussed in the figure. The possible ligands with respect to Alzheimer's disease for TREM2 are lipids, APOE as well as amyloid-β. In the physiological condition, upon tyrosine phosphorylation of DAP12 ITAM leads to recruitment of Syk kinase along with the GRB/SOS1 and DOK3, which activates downstream signaling pathways of ERK, PLC-γ, PI3K/Akt as well as Vav. The activation of PLC-γ induces inflammatory cytokine response *via* DAG stimulation and anti-inflammatory cytokine response along with cell rearrangement and phagocytosis via elevating Ca<sup>2+</sup> response in the cell through IP3 activation. The main survival pathway of cell PI3K/Akt primarily activates mTOR pathway and Foxo that inhibit autophagy and induce cell survival, inhibits GSK-3β that eventually reduce Tau phosphorylation in Alzheimer's disease and suppress IKK, which is involved in the inflammatory response. PI3K activation increase formation of PI 3,4,5-P3, an important phosphoinositide in the process of phagocytosis and migration. Syk recruitment activates Vav2/3 molecule *via* LAT phosphorylation, which induces actin cytoskeleton polymerization important for chemotaxis, migration, and phagocytosis. TREM2 is one of the disease-linked genes in Alzheimer's disease, the AD-related variant generally present in Ig-like extracellular domain of the receptor, which includes R47H, R62H, D87N, T96K

nucleotide exchange factors (GEFs), GTPase activating proteins (GAPs) and Guanosine nucleotide dissociation inhibitors (GDIs) [107–109]. GEFs are involved in the activation of GTPases by catalyzing the exchange of GTPs for GDPs, whereas GAPs are involved in the inactivation of small GTPases by promoting GTP hydrolysis [107, 108]. GDIs maintain the inactive form of GTPases by preventing its membrane localization [109]. GTPases respond to various extracellular stimuli and are involved in cell survival via various downstream signaling cascades that regulate gene expression [48]. Rho (RhoA, Rac1 and Cdc42) signaling cascades play a crucial role in actin and tubulin dynamics of neuronal cells [110, 111]. Several membrane receptors such as  $G_{12/13}$ -protein coupled GPCRs, tyrosine kinase receptors,

integrin receptors are involved in the activation of Rho signaling pathway [112–114]. The receptors are mainly involved in the activation of different Rho-GEFs: p115-RhoGEF, PDZ-RhoGEF, leukaemia-associated RhoGEF and lymphoid blast crisis-RhoGEF, [115, 116] which further activates Rho-GTPases by the exchange of GTP for a GDP molecule. In the human brain, Rho GTPases are involved in a wide variety of functions such as learning and memory formation (by maturation of dendritic spines), neuronal survival, cell migration via actin cytoskeleton dynamics, microtubule stabilization, etc., [111, 117–119]. Rab-GTPases are the key regulators of intracellular membrane trafficking and are involved in several vital functions of CNS development [120–123]. Arf GTPases are involved in the vesicular transport similar

Desale et al. Molecular Biomedicine (2021) 2:17 Page 7 of 18

to Rab proteins [124]. Ran GTPases regulate the nucleocytoplasmic transport of proteins and RNA in the cell [125]. GTPases are involved in a wide variety of functions ranging from vesicular trafficking, membrane transport, actin networking, etc., In the human brain, small GTPases are involved in the regulation of various functions such as phagocytosis and vesicular trafficking by Rab and Arf family of GTPases in microglia; actin remodelling and microglial migration by Rho GTPases; maturation of dendritic spines and neuronal survival by Rho GTPases, etc. The signaling of these different GTPases and their pathological role in Alzheimer's disease have been widely described.

# Rho family of GTPases in amyloid-β and Tau pathology

Rho family of GTPases are primarily involved in actin cytoskeleton remodelling and constitutes around 20 members that include RhoA, Rac1, Cdc42, etc., [117, 126]. Activation of Rho GTPases is GEF-mediated that are regulated by  $G_{12/13}$  proteins.  $G_{12/13}$  proteins are involved in the translocation of Rho-GEF from cytosolic region to the plasma membrane for its activation. Activated membrane-bound Rho-GEF in turn catalyzes the activation of Rho GTPases. Similarly, Rho-GTPases are also activated by different receptor tyrosine kinases and integrin receptors. 8 Rho-GTPases among 20 are known to be activated by RTKs [113]. Upon activation, each of these GTPases has a distinct set of effector kinases that are involved in different physiological functions of the cell. For example, RhoA effector kinases include Rhoassociated protein kinases (ROCK1 and ROCK2), Citron, PKN/PRK1, etc. Cdc42 effector kinases include myotonic dystrophy kinase-related CDC42-binding (MRCK), activated Cdc42 kinase (ACK), mixed lineage kinases (MLK), p-21 activated kinases (PAK), etc. Rac1 effector kinases include mixed lineage kinases (MLK) and p-21 activated kinases (PAK) which act as common effectors for Rac1 and Cdc42. In AD, synaptic degeneration and impaired actin cytoskeletal network has been observed which is mainly mediated by the pathological effects of these Rho GTPases [127-129].

### **RhoA GTPases**

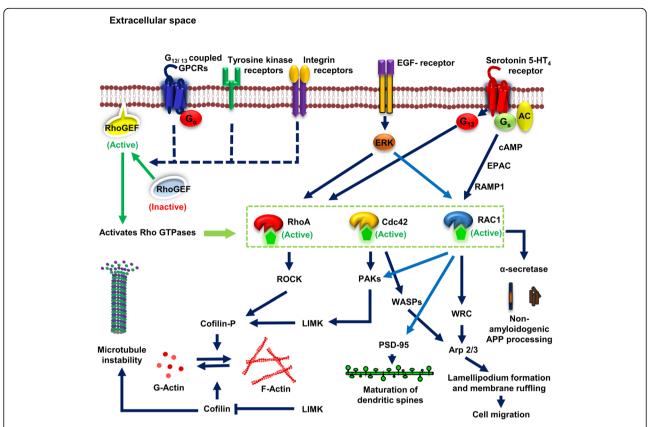
RhoA GTPases are majorly associated with synapses and dendritic microtubules [130].  $G_{12/13}$  coupled receptors are involved in the activation of RhoA GTPases mediated by Rho-GEFs. Several  $G_{12/13}$ -protein linked GPCRs, tyrosine kinase receptors, integrin receptors are involved in the activation of RhoA GTPases. One such receptor is serotonin 5-HT<sub>4</sub> receptor which is reported to play a key role in learning, memory, behaviour and synaptic plasticity. Schill *et al.* 2020 reported that serotonin 5-HT<sub>4</sub> receptor activates RhoA-mediated signaling via  $G_{13}$  proteins that leads to the phosphorylation of cofilin by ROCK kinases [131].

Cofilin is a key molecule that has a significant role in stabilization and reorganization of the actin cytoskeleton. RhoA activation by 5-HT<sub>4</sub> receptor agonist in primary cultures of hippocampal neurons promoted the formation of filamentous actin and maturation of dendritic spines [131] (Fig. 3). In neurodegenerative diseases such as AD, RhoA GTPases are playing a crucial role in promoting the progression of the disease. The activity of p-Tyr42 Rho was observed to be significantly higher in AD mice models and human brain of AD patients [132]. Amyloid-β disrupts microtubule stability and actin polymerization via activation of RhoA GTPases [132, 133]. Amyloid-β promoted GTP-bound RhoA in human SH-SY5Y cells and the cellular localization of RhoA was increased in dystrophic neurites of APP mice models [130, 133]. Amyloidβ exposure on SH-SY5Y cells also showed increased expression levels of collapsin response mediator protein-2A (CRMP-2A) and its phosphorylated form (phospho-threonine) [133]. CRMPs are a class of microtubule-associated proteins that are involved in a wide variety of neuronal functions such as neuronal development, dendritic formation, axonal guidance and also in cell migration [134]. Phosphorylation of CRMP-2A mediated by amyloid-β/ RhoA-GTP in Tg2576 mice models showed reduced levels of tubulin binding to CRMP-2A, which could prevent neuritic growth and plasticity [133]. Protein tyrosine phosphatases 1B (PTP1B) which are involved in neuronal survival are also inhibited by amyloid-β activated RhoA-GTPases [135]. Histone deacetyl transferace-6 (HDAC6) is involved in the deacetylation of tubulin and Tau [136, 137]. P-Tyr42 RhoA activates ROCK, which in turn inhibits HDAC6 activation and promotes tubulin and Tau acetylation, which would lead to microtubule instability and Tau aggregation respectively [132]. Immunohistological analysis revealed that RhoA colocalized with hyperphosphorylated Tau in human AD cortex and hippocampus [130]. Cap et al. 2020 demonstrated the role of RhoA/ROCK signaling towards the phosphorylation of Tau, expression of NADPH oxidase and subsequent generation of ROS [138]. Amyloid-β exposure activated RhoA (p-Tyr42) which in turn activates GSK-3β, a serinethreonine kinase majorly involved in hyperphosphorylation of Tau species [138]. At higher concentrations of amyloid-β, nuclear translocation of P-Tyr42 RhoA was observed and regulated the expression of NAD kinase by binding to NADK promoter which was similar to ROS exposure [138]. Amyloid-β mediated Rho/ROCK signaling also mediates retrograde flow of actin cytoskeleton that leads to growth cone collapse and microtubule destabilization [132] (Fig. 4).

# Rac1 and Cdc42 GTPases

Rac GTPases (Rac1 and Rac3) are mainly involved in the formation of dendritic spines and play a vital role in

Desale et al. Molecular Biomedicine (2021) 2:17 Page 8 of 18

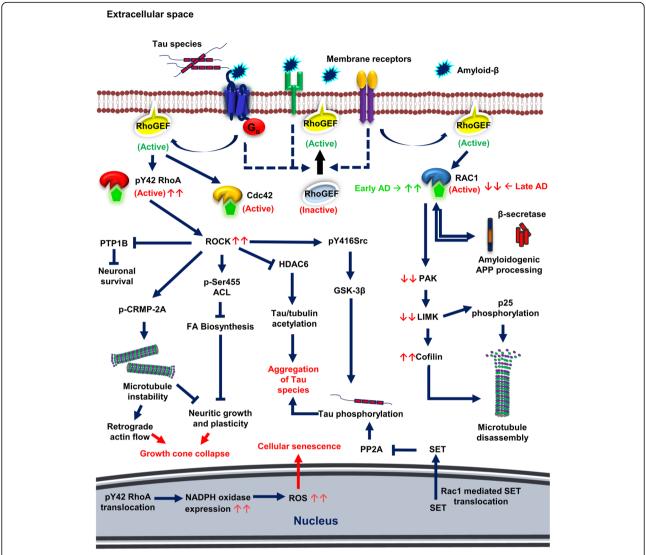


**Fig. 3** Physiological activation and signaling of Rho-GTPases. Several membrane receptors, such as  $G_{12/13}$  coupled GPCRs, Tyrosine kinase receptors, Integrin receptors, etc., are involved in the activation of Rho-GEFs. The active form of Rho-GEF binds to the membrane and are involved in activation of Rho-GTPases (mostly RhoA, Rac1 and Cdc42) through GTP catalysis. Epidermal growth factor receptor is involved in ERK-mediated phosphorylation of RhoA and Rac1 and its activation. Serotonin 5-HT4 receptor activates cAMP, EPAC and RAMP1 *via*  $G_5$  signaling and activates Rac1 signaling that is involved in the promotion of non-amyloidogenic APP processing, dendritic spine formation (PSD-95 expression) in mature neurons, lamellipodium formation and membrane ruffling for migration. Serotonin 5-HT4 receptor also activates RhoA *via*  $G_{13}$  proteins. Activated RHO-GTPases are involved in several vital functions such as cell migration, maturation of dendritic spines, neuritic outgrowth, learning, memory, *etc.*, RhoA GTPase activates ROCK kinase that is involved in the phosphorylation of cofilin. Cofilin is involved in the formation of globular actin from filamentous actin (actin depolymerization), and this process is inhibited and reversed upon cofilin phosphorylation. Cofilin also binds tubulin and promotes microtubule instability. Mixed lineage kinases (MLK) and p-21 activated kinases (PAK) are the common effectors of Cdc42 and Rac1. PAKs are involved in the activation of LIM kinase, which also involved in the phosphorylation of cofilin. Wiskott–Aldrich syndrome protein (WASP) and WAVE regulatory complex (WRC), the downstream effectors of Cdc42 and Rac1 respectively activate Arp 2/3 complex which promotes membrane ruffling during migration

learning, memory and synaptic plasticity [139]. Rac1 and Cdc42 are involved in the polymerization of actin cytoskeletal network, formation of lamellipodia structures and membrane ruffling and play a significant role in cell migration [118, 140]. Also, Rac1 plays an essential role in axonal growth, guidance and promotes neuronal survival in the central and peripheral nervous system [119]. p21-activated kinases (PAKs) are serine/threonine kinases that are direct effectors of Rac1/Cdc42-GTPases, which are majorly involved in actin remodelling [141]. Phosphorylation of regulatory light chains (RLC) on serine19 by PAKs leads to actin network stabilization and dendritic spine morphogenesis [142]. LIM kinase (LIMK), a serine-threonine kinase is another PAK effector that is involved in the promotion of filamentous

actin and microtubule disassembly [143]. Phosphorylation of cofilin by LIMK inhibits its activity which leads to actin polymerization [143]. LIMK also inhibits p25 activity (promotes tubulin polymerization) by phosphorylation, thus leading to microtubule disassembly [143, 144] (Fig. 3). In AD brains, PAKs levels and activity are depleted due to amyloid- $\beta$  and hence, active levels of cofilin increases, which ultimately leads to microtubule instability [145, 146]. It is also reported that activated cofilin directly binds tubulin, displaces Tau from tubulin/microtubules, and inhibits Tau-induced microtubule assembly [145]. Rac1 GTPases are reported to be involved in the non-amyloidogenic APP processing through serotonin 5-HT4 receptor activation followed by cAMP/Epac/Rap1/Rac1 signaling pathway [147, 148].

Desale et al. Molecular Biomedicine (2021) 2:17 Page 9 of 18



**Fig. 4** Pathological signaling of Rho GTPases in Alzheimer's disease. In Alzheimer's disease, the extracellular space is filled with senile plaques of amyloid-β and aggregated Tau species. These disordered proteins interact with several surface receptors, which on activation lead to pathological signaling of Rho-GTPases. Rho-GTPases are highly activated (phosphorylated on Y42) on amyloid-β exposure. Hyperactivation of Rho-GTPases leads to increased activation of ROCK kinases. ROCK kinase phosphorylates Src at Tyrosine 416 and activates to activate GSK-3β, an enzyme involved in Tau hyperphosphorylation. ROCK also inhibits HDAC6 activity which ultimately promotes Tau and tubulin acetylation. All these post-translational modifications of Tau lead to aggregation of Tau species. ROCK phosphorylates and inactivates CRMP-2A and ATP citrate lyase (p-Ser 455) (ACL). CRMP-2A are microtubule-associated proteins, and its inactivation led to microtubule instability and retrograde flow of actin filaments. Fatty acid biosynthesis is inhibited by ATP-citrate lyase phosphorylation which ultimately inhibits neurite growth, plasticity and leads to growth cone collapse in neuronal cells. Activated ROCK kinases also inhibit protein tyrosine phosphatase 1B and neuronal survival. Phosphorylated RhoA translocates to the nucleus and promotes the expression of NADPH oxidase, increases ROS levels and cellular senescence. Rac1 GTPase expression level varies at different stages and promotes amyloidogenic APP processing during the later stages of Alzheimer's disease leading to more amyloid-β secretion and *vice versa*. At later stages, Rac1 expression level is reduced, and subsequent effectors PAK and LIMK activities are also downregulated. This condition leads to the imbalance in cofilin (active) and its phosphorylated form (inactive). Excessive cofilin activity causes actin depolymerization and microtubule instability. Under AD conditions, Rac1 also promotes translocation of SET proteins from the nucleus to the cytoplasm that acts as ph

Both Rac1 and Cdc42 are involved in the negative regulation of RhoA by activating Rho-GAPs [149, 150]. The levels of Rac1 has been significantly altered in the frontal cortex and plasma of AD patients at different stages of disease progression when compared with age-matched

controls [151]. Under pathological conditions, Rac1 GTPases are involved in amyloidogenic pathway of APP processing and promotes the formation of amyloid- $\beta$  peptides [148, 151]. The Rac1-specific inhibitor NSC23766, decreased the levels of Rac1 and APP in a

Desale et al. Molecular Biomedicine (2021) 2:17 Page 10 of 18

concentration-dependent manner [148]. Rac1 peptide treatment in primary cortical neurons and human SH-SY5Y neuroblastoma cell lines enhanced the immunore-activity of amyloid-β peptides and/or its precursor, APP [151]. However, controversial results were observed during the later stages of disease progression. Rac1 level decreased in 7 months old 3xTg-AD mice models and Rac1 administration enhanced the expression of the post-synaptic marker, PSD-95Rac1, rescues spine loss and ameliorates synaptic abnormalities [151]. Rac1 GTPases also promotes phosphorylation of Tau protein by enhancing the translocation of nuclear oncoprotein, SET to the cytosol that acts as a protein phosphatase 2A (PP2A) inhibitor [151] (Fig. 4).

# Rab family of GTPases

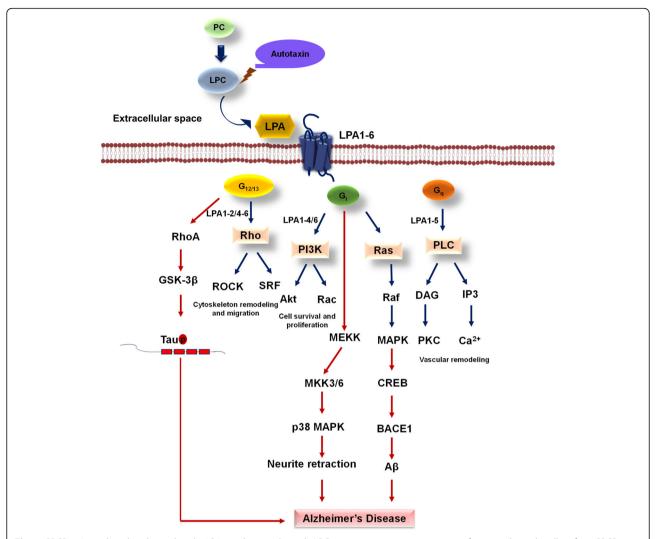
Ras-like GTPases in brain (Rab) are the family of small GTPases with 61 members and are involved in regulating intracellular vesicular transport that includes both endocytosis and exocytosis [152]. In the brain, Rab GTPases are involved in several vital functions such as CNS development, polarized neurite growth, endocytosis and axonal retrograde transport, synaptic vesicle exocytosis, etc., (briefly reviewed by Ling NG et al. 2008) [120]. Several Rab GTPases such as Rab5, Rab7A, Rab10, Rab11A are reported to be associated with AD [153-155]. The levels of Rab5 and Rab7 were significantly upregulated in the frontal cortex and hippocampus of AD brain [156]. Rab5 GTPases are involved in trafficking and fusion of early endosomes and the Rab5 positive early endosomes are found enlarged with increased endocytosis under AD conditions [156-158]. It is also reported that Rab5-mediated APP processing led to apoptosis in AD neurons [158]. Rab11 GTPases are colocalized with BACE-1 during the late onset of AD that are involved in axonal sorting of BACE-1 [154, 159]. Rab7 GTPases are colocalized with BACE-1, the enzyme responsible for amyloidogenic APP processing and synthesis of amyloid-β peptides [72]. Rab7A is involved in multiple roles that include regulation of late endosomes and lysosomal degradation pathway [160]. In AD brain, Rab7A level is elevated and is reported that it regulates exosome-mediated Tau secretion [161]. Rab7A deletion on primary cortical neuronal cells and HeLa cells overexpressing Tau downregulated Tau secretion which clearly indicated the pathological role of Rab7A in Alzheimer's disease and Tauopathy [161, 162].

## GPCR-mediated lysophosphatidic acid signaling

Lysophospholipids are the part of membrane phospholipid that comprises of a polar head group followed by a hydrophobic carbon chain. It comes under lyso-type glycerolipid having glycerol backbone with ester-linked fatty acids chain (acyl chain at position 1 or 2) and a

phosphate group [163]. In humans, 16:0 LPA, 18:1 LPA, 18:2 LPA are commonly observed that are majorly expressed in astrocytes, microglia, neural progenitor cells and oligodendrocytes in the nervous system [164]. There are four pathways of LPA formation among which autotaxin (ATX) is majorly involved in cellular signaling cascades. Autotaxin is a pyrophosphatase/phosphodiesterase enzyme (ENPP), which hydrolyses phosphobond of lysophospholipids, nucleoside diester triphosphates, and choline phosphate esters [165]. ATX importantly converts lysophospholipids into lysophosphatidic acid (LPA) for signaling in the cell [166]. ATX mediates cleavage of lysophosphatidylcholine (LPC) into LPA and choline and similarly, lysophosphatidylethanolamine, and lysophosphatidylserine into LPA and respective secondary group for the signaling pathway. LPA is also produced from phosphatidic acid (PA) via enzyme phospholipase A1 $\alpha$  or  $\beta$  [167]. LPA-mediated physiological effects include nervous system development, immune system function, inflammation, and wound healing, which is executed by currently known six LPA receptors LPAR1-LPAR6 [168, 169]. LPA formed extracellularly act as a ligand for cognate GPCRs to activate signaling cascade, and the signal proceeds via  $G\alpha$  subunits such as  $G_{12/13}$ ,  $G_{q/11}$ ,  $G_{i/o}$ , and  $G_s$ [170] (Fig. 5).

Neuronal progenitor cells (NPC) are the preliminary cells that initiate neurogenesis by differentiating into neurons and other glial cells as restricted cell types. LAP mediate neurogenesis of NPC via LPA1, LPA2, LPA4 receptors [171]. LPA1 plays an important role in modulating neurogenesis, i.e., the process of differentiation is driven by LPA1 and partly by LPA2. LPA exerbate neurotransmitter like stimuli of ionic conductance hence arbitrate cortical development [172, 173]. LPA1 function through  $G_{i/o}$  protein-mediated pathway, which implement Rho-mediated actin rearrangement, microtubule formation important for cell morphology and motility. LPA1 also initiates phospholipase C (PLC) activation via pertussis toxin (PTX)-insensitive pathway mediated by G<sub>q</sub> proteins, making LPAR1 a multifunctional LPA receptor [174]. LPA1 especially induces synaptic plasticity that includes protein kinase c (PKC) activation, with Rho and Rac-dependent actin-based changes [175, 176]. Similarly, LPA2 also functions through G<sub>i</sub> and G<sub>q</sub> mediated pathways, which activates PLC, and increases intracellular Ca2+ and inositol phosphate production [177]. Extension and branching neurites are important for the formation of neuronal network, which requires actin rearrangements. The neurite retraction and growth cone collapse are induced by all LPA receptors except LPA3. One anticipated function of LPA3 is to increase intracellular Ca<sup>2+</sup> levels, however, it Desale et al. Molecular Biomedicine (2021) 2:17 Page 11 of 18



**Fig. 5** GPCR in Lysophosphatidic acid cycle. LPA production through LPC via autotaxin enzyme exerts its function through cell surface GPCR mainly G12/13, Gi/o, and Gαq protein-mediated signals. Amongst six different LPA, LPA1-2/4-6 exert their function through  $Ga_{12/13}$  and follows Rho-mediated cytoskeleton remodeling which assists cellular migration. LPA1-4/6 exert their function through Gi/o, which is responsible for activating Pl3K and Ras-mediated MAPK signaling for cell survival and proliferation. Gαq protein-mediated signal activate the PLC pathway results in vascular remodeling on stimulation by LPA1-5. Defect in signaling through  $Ga_{12/13}$  in AD abnormally increases active GSK-3β, which increases phospho-Tau. Gi leads to dysregulation of BACE1, which impose abnormal Aβ and neurite retraction via p38 MAPK kinases

induces axonal branching and neurite branching through  $G_q$  and the Rho family GTPase 2 (Rnd2) family [178, 179]. LPA4 receptor is linked with  $G_{q^{\prime}}$   $G_{12/13}$  and to some extent with  $G_s$  protein. LPA4 reduces the LPA driven cell motility and invasion by negatively modulating PI3K pathways, Rho-dependent migration. It also enhances cell adhesion and aggregation [180]. Similarly, LPA5 is also an anti-migratory receptor, which signals through PKA-CAMP pathway and also responsible for reduced PIP3 levels at the plasma membrane that supports reduced migration [181]. However, LPA6 is recently characterized, but the mutation of this receptor leads to hair loss in humans [182]. Its function related to CNS still needs to addressed.

In AD, amyloidogenic pathway sequentially functions through  $\beta$ -secretase and  $\gamma$ -secretase that leads to the production of toxic A $\beta$  peptide.  $\beta$ -secretase activity, also known as  $\beta$ -site APP-cleaving enzyme 1 (BACE1), is the rate limiting step in production of A $\beta$  [182]. Additionally, vascular factors such as oxidized low-density lipoprotein (oxLDL) have been reported in AD pathogenesis. Human cerebrospinal fluid (CSF) consists of a high level of lipoprotein in AD, the occurrence of levels of A $\beta$  and oxLDL indicated a significant role of oxLDL in A $\beta$  production [183]. LPA is a major bioactive component of oxLDL, which are known to disrupt bloodbrain barrier (BBB) and imparts various pathologies related to AD [184]. BACE expression increases by LPA

Desale et al. Molecular Biomedicine (2021) 2:17 Page 12 of 18

via cAMP response element binding protein (CREB) mediated mechanism. LPA increases the binding affinity of CREB to CRE site of the BACE1 promoter sequence. Hence there is upregulation of BACE1 upon LPA response. The signaling pathway involves phosphorylation of PKCδ, MEK, MAPK and p90RSK, which mediate CREB phosphorylation leading to upregulation of BACE1 [166]. LPA induce Tau phosphorylation and neurite retraction, the major target of LPA to cause Tau phosphorylation is glycogen synthase kinase- 3B (GSK-3β) [185]. Increased activity of GSK-3β in response to LPA is carried out through  $G_{\alpha 12/13}$ -mediated Rho/ROCK pathway. Rho-GTPase has been observed to cause Tau pathology, where Rho/ROCK pathway phosphorylates Tau at various epitopes such as Thr245 and Ser409 [186]. LPA-induced neurite retraction is carried out after phosphorylation of Tau by GSK-3B, PKA, and additionally with activation of p38 MAPK [187] (Fig. 5).

# Other GPCR interacting proteins in amyloid- $\beta$ and Tau pathology

G-protein coupled receptor kinases (GRKs) and βarrestins are the first identified GPCR/ G-protein reguproteins that are involved in desensitization and endocytosis [188]. GRKs are involved in phosphorylation of receptor upon ligand activation [189–191]. Arrestin binds to phosphorylated GPCRs in order to prevent its coupling to G-proteins, ultimately leading to receptor desensitization and endocytosis [192, 193]. β-arrestin 1 and 2 are majorly involved in receptor desensitization, followed by internalization which mediates several G-protein independent signaling pathways that includes activation of cellular kinases, transactivation, gene expression, etc., [194, 195]. The internalized GPCRs further undergoes endosomal or lysosomal sorting for dephosphorylation/resensitization or lysosomal degradation respectively [196, 197]. The expression level of  $\beta$ -arrestin varies during the progression of AD.  $\beta$ arrestin 2 expression was observed to be elevated in AD patients [198]. Several GPCRs such as GPR3,  $\beta_2$  adrenergic receptor and δ-opioid receptors are involved in Gprotein independent regulation of amyloid-β synthesis via γ-secretase activity [72, 199, 200]. Later, Thathiah et al. 2013 demonstrated that these receptors are involved in regulation of amyloid-β production and γsecretase activity by directly activating  $\beta$ -arrestin 2 [198]. Gene knock-out studies for β-arrestin 2 failed to produce amyloid-β, which indicated β-arrestin-mediated GPCR signaling [198]. β-arrestin 1 is also involved in elevated amyloid-β synthesis mediated by γ-secretase activity [201]. Both  $\beta$ -arrestin 1 and 2 are reported to bind directly to anterior pharynx-defective 1α (APH-1α) subunit of  $\gamma$ -secretase for its activation [198, 201]. There are other GPCR regulating proteins that are involved in the pathogenic progression of Alzheimer's disease. Ras homolog enriched in brain (Rheb) are another group of small GTPases that are highly expressed in frontal cortex and hippocampus [202]. Rheb GTPases are the key regulators of mTOR signaling pathways and are involved in the regulation of BACE1 activity and degradation [203, 204]. Overexpression of Rheb in cultured neuronal cells attenuated BACE1 and AB levels in mTOR-independent pathway [204]. In AD brains, the levels of Rheb is downregulated, which lead to the enhanced activation of BACE1 and Aβ production [204]. GPCR-associated sorting proteins (GASPs) are the group of sorting proteins that promotes lysosomal degradation of internalized receptors [205]. p60TRP (transcription regulator protein) is a GASP that are involved in neuronal survival and rescues cells from death [206, 207]. Overexpression of p60TRP in P-12 cells promoted dephosphorylation of APP and downregulated BACE1 activity [207].

# **Conclusion**

Approximately, 481 drug molecules (~34%) are approved by the FDA act that targets GPCRs. 320 drugs are under clinical trials among which ~36% could be potentially novel GPCR targets. Among CNS diseases, around 130 drugs (27%) are FDA approved that are potential GPCR targets [208]. The mis-regulation of signaling pathways in Alzheimer's disease have serious impact on amyloid-β secretion, Tau phosphorylation, neurotransmitter signaling, glial function which eventually ameliorate the disease condition. The cell surface receptors, GPCR and the subsequent GPCR related signaling molecule are involved in regulation of BACE1, γ-secretase and eventually affect Tau phosphorylation in Alzheimer's disease. Several drug molecules have been developed in the field of Alzheimer's disease targeting a wide variety of receptors such as G-protein coupled receptors, kinases/ enzymes, ionotropic channels, hormone receptors, etc., that largely failed during clinical trials [209]. Allosteric modulators could be an alternative and effective therapeutic agent for Alzheimer's disease. For example, positive allosteric modulators (PAM) of M1 muscarinic acetyl choline receptor are proved to be effective in AD therapy [210]. Hence, understanding the role of Gprotein coupled receptor signaling pathways and the molecules involved in the pathway is necessary to design a particular drug target to treat the disease. With the evolved knowledge regarding cell surface receptors, GPCR as well as interacting GTPase might provide alternative strategies to modulate the AD pathogenesis.

#### **Abbreviations**

A $\beta$ : Amyloid- $\beta$ ; AD: Alzheimer's disease; ACK: Activated Cdc42 kinase; APH-1 $\alpha$ : Anterior pharynx-defective 1 $\alpha$ ; APP: Amyloid precursor protein; Arf: ADP-ribosylation factor; ATX: Autotaxin; BACE1:  $\beta$ -site APP-cleaving enzyme 1; BBB: Blood-brain barrier; CNS: Central nervous system; CREB: CAMP response

Desale et al. Molecular Biomedicine (2021) 2:17 Page 13 of 18

element binding protein; CRMP-2A: Collapsin response mediator protein-2A; CSF: Cerebrospinal fluid; DHA: Docosahexaenoic acid; DOR: Delta-opioid receptor; FSH: Follicle stimulating hormone; GAP: GTPase activating protein; GASP: GPCR-associated sorting protein; GDI: Guanosine nucleotide dissociation inhibitor; GDP: Guanosine diphosphate; GEF: Guanine nucleotide exchange factor; GPCR: G-protein coupled receptor; GSK3β: Glycogen synthase kinase 3β; GTP: Guanosine triphosphate; HDAC6: Histone deacetyl transferase 6; ITAM: Immunoreceptor tyrosine-based activation motifs; LIMK: LIM kinase; LPA: Lysophosphatidic acid; LPC: Lysophosphatidylcholine; LPS: Lipopolysaccharides; MAPK: Mitogen activated protein-kinases; MLK: Mixed lineage kinase; MRCK: Myotonic dystrophy kinase-related CDC42binding kinase; NADK: Nicotinamide adenine dinucleotide kinase; oxLDL: Oxidized low-density lipoprotein; PAK: p-21 activated kinase; PDK: Phosphoinositide-dependent kinase; PIP3: Phosphatidylinositol 3,4,5triphosphate; PKC: Protein kinase C; PKN/PRK: Protein kinase-C related kinase; PP2A: Protein phosphatase 2A; PUFA: Polyunsaturated fatty acids; PIP2: Phosphatidylinositol 4,5 bisphosphate; PTEN: Phosphatase and tensin homologue; PTP1B: Protein tyrosine phosphatase 1B; Rab: Ras-like proteins in brain; Ras: Ras Sarcoma oncoproteins; Ran: Ras-like nuclear proteins; Rheb: Ras homolog enriched in brain; Rho: Ras homologous proteins; RLC: Regulatory light chain; ROCK: Rho-associated protein kinase; ROS: Reactive oxygen species; RTK: Receptor tyrosine kinase; SHIP: SHdomain-containing inositol polyphosphate 5-phosphatase; Syk: Spleen tyrosine kinase; TREM2: Triggering receptor expressed on myeloid cells 2; WASP: Wiskott-Aldrich syndrome protein; WRC: WAVE regulatory complex

#### Acknowledgements

We are grateful to Chinnathambi's lab members for their scientific discussions, helpful suggestions and critical reading of the manuscript.

#### Authors' contributions

SD, HC and SC performed the literature search and wrote the manuscript. SC conceived the idea of the work and supervised the project. We are grateful to Chinnathambi's lab members for their critical readings and fruitful discussions. All authors approved the final manuscript.

#### Funding

This project is supported in part by grant from in-house CSIR-National Chemical Laboratory grant MLP029526 and CSIR-FBR mission mode, MLP107026. Hariharakrishnan Chidambaram and Smita Eknath Desale acknowledges the fellowship from Department of Biotechnology, Government of India.

# Availability of data and materials

Not applicable.

#### **Declarations**

# Ethics approval and consent to participate

Not applicable.

#### Consent for publication

Not applicable.

#### Competing interests

The authors declare that they have no competing interests.

# Received: 2 December 2020 Accepted: 18 March 2021 Published online: 10 June 2021

#### References

- DeTure MA, Dickson DW. The neuropathological diagnosis of Alzheimer's disease. Mol Neurodegener. 2019;14(1):1–18 S10.1186/s13024-019-0333-5.
- Cvetković-Dožić D, Skender-Gazibara M, Dožić S. Neuropathological hallmarks of Alzheimer's disease. Arch Oncol. 2001;9(3):195–9 UDC: 616-091.
- Serrano-Pozo A, Frosch MP, Masliah E, Hyman BT. Neuropathological alterations in Alzheimer disease. Cold Spring Harb Perspect Med. 2011;1(1): a006189. https://doi.org/10.1101/cshperspect.a006189.
- Binder LI, Frankfurter A, Rebhun LI. The distribution of tau in the mammalian central nervous system. J Cell Biol. 1985;101(4):1371–8. https://doi.org/10.1083/jcb.101.4.1371.

- Conde C, Cáceres A. Microtubule assembly, organization and dynamics in axons and dendrites. Nat Rev Neurosci. 2009;10(5):319–32. https://doi.org/1 0.1038/nrn2631.
- Martin L, Latypova X, Terro F. Post-translational modifications of tau protein: implications for Alzheimer's disease. Neurochem Int. 2011;58(4):458–71. https://doi.org/10.1016/j.neuint.2010.12.023.
- Balmik AA, Chinnathambi S. Multi-faceted role of melatonin in neuroprotection and amelioration of Tau aggregates in Alzheimer's disease. J Alzheimers Dis. 2018;62(4):1481–93. https://doi.org/10.3233/JAD-170900.
- Gorantla NV, Chinnathambi S. Tau protein squired by molecular chaperones during Alzheimer's disease. J Mol Neurosci. 2018;66(3):356–68. https://doi. org/10.1007/s12031-018-1174-3.
- Sonawane SK, Chinnathambi S. Prion-like propagation of post-translationally modified tau in Alzheimer's disease: a hypothesis. J Mol Neurosci. 2018; 65(4):480–90. https://doi.org/10.1007/s12031-018-1111-5.
- Brunden KR, Trojanowski JQ, Lee VM-Y. Advances in tau-focused drug discovery for Alzheimer's disease and related tauopathies. Nat Rev Drug Discov. 2009;8(10):783–93. https://doi.org/10.1038/nrd2959.
- Congdon EE, Sigurdsson EM. Tau-targeting therapies for Alzheimer disease. Nat Rev Neurol. 2018;14(7):399–415. https://doi.org/10.1038/s41582-018-0013-z.
- Noble W, Jimenez-Sanchez M, Perez-Nievas BG, Hanger DP. Considerations for future tau-targeted therapeutics: can they deliver? Expert Opin Drug Discov. 2020;15(3):265–7. https://doi.org/10.1080/17460441.2020.1685977.
- Panza F, Solfrizzi V, Seripa D, Imbimbo BP, Lozupone M, Santamato A, et al. Tau-centric targets and drugs in clinical development for the treatment of Alzheimer's disease. Biomed Res Int. 2016;2016. https://doi.org/10.1155/201 6/3245935.
- Balmik AA, Das R, Dangi A, Gorantla NV, Marelli UK, Chinnathambi S. Melatonin interacts with repeat domain of Tau to mediate disaggregation of paired helical filaments. Biochim Biophys Acta Gen Subj. 1864;2020(3): 129467. https://doi.org/10.1016/j.bbagen.2019.129467.
- Gorantla NV, Das R, Chidambaram H, Dubey T, Mulani FA, Thulasiram HV, et al. Basic Limonoid modulates Chaperone-mediated Proteostasis and dissolve Tau fibrils. Sci Rep. 2020;10(1):1–19. https://doi.org/10.1038/s41598-020-60773-1
- Sonawane SK, Chidambaram H, Boral D, Gorantla NV, Balmik AA, Dangi A, et al. EGCG impedes human Tau aggregation and interacts with Tau. Sci Rep. 2020;10(1):1–17. https://doi.org/10.1038/s41598-020-69429-6.
- Das R, Balmik AA, Chinnathambi S. Melatonin reduces GSK3β expression and tau phosphorylation via Nrf2 nuclear translocation. BioRxiv. 2019: 861229. https://doi.org/10.1101/861229.
- Gorantla NV, Landge VG, Nagaraju PG, Priyadarshini CGP, Balaraman E, Chinnathambi S. Molecular cobalt (II) complexes for tau polymerization in Alzheimer's disease. ACS omega. 2019;4(16):16702–14. https://doi.org/10.1 021/acsomega.9b00692.
- Balmik AA, Chidambaram H, Dangi A, Marelli UK, Chinnathambi S. HDAC6 ZnF UBP as the Modifier of Tau Structure and Function. Biochemistry. 2020; 59(48):4546–62. https://doi.org/10.1021/acs.biochem.0c00585.
- 20. Das R, Balmik AA, Chinnathambi S. Effect of Melatonin on Tau aggregation and Tau-mediated cell surface morphology. Int J Biol Macromol. 2020;152: 30–9. https://doi.org/10.1016/j.ijbiomac.2020.01.296.
- Gorantla NV, Balaraman E, Chinnathambi S. Cobalt-based metal complexes prevent Repeat Tau aggregation and nontoxic to neuronal cells. Int J Biol Macromol. 2020;152:171–9. https://doi.org/10.1016/j.ijbiomac.2020.02.278.
- Gorantla NV, Das R, Balaraman E, Chinnathambi S. Transition metal nickel prevents Tau aggregation in Alzheimer's disease. Int J Biol Macromol. 2020; 156:1359–65. https://doi.org/10.1016/j.ijbiomac.2019.11.176.
- Gorantla NV, Das R, Mulani FA, Thulasiram HV, Chinnathambi S. Neem derivatives inhibits tau aggregation. J Alzheimers Dis Rep. 2019;3(1):169–78. https://doi.org/10.3233/ADR-190118.
- Sonawane SK, Balmik AA, Boral D, Ramasamy S, Chinnathambi S. Baicalein suppresses Repeat Tau fibrillization by sequestering oligomers. Arch Biochem Biophys. 2019;675:108119. https://doi.org/10.1016/j.abb.2 019.108119.
- Desale SE, Dubey T, Chinnathambi S. α-Linolenic acid inhibits Tau aggregation and modulates Tau conformation. Int. J Biol Macromol. 2020; 166:687–93.
- Desale SE, Chinnathambi S. α-Linolenic acid induces clearance of Tau seed via Actin-remodeling in Microglia. Mol Biomed. 2020;2:4. https://doi.org/1 0.1186/s43556-021-00028-1.

Desale et al. Molecular Biomedicine (2021) 2:17 Page 14 of 18

- Das R, Balmik AA, Chinnathambi S. Phagocytosis of full-length Tau oligomers by Actin-remodeling of activated microglia. J Neuroinflammation. 2020;17(1):1–15. https://doi.org/10.1186/s12974-019-1694-y.
- Das R, Chinnathambi S. Microglial priming of antigen presentation and adaptive stimulation in Alzheimer's disease. Cell Mol Life Sci. 2019;76(19): 3681–94. https://doi.org/10.1007/s00018-019-03132-2.
- Das R, Chinnathambi S. Actin-mediated Microglial Chemotaxis via G-Protein Coupled Purinergic Receptor in Alzheimer's Disease. Neuroscience. 2020;448: 325–36. https://doi.org/10.1016/j.neuroscience.2020.09.024.
- Desale SE, Chinnathambi S. Role of dietary fatty acids in microglial polarization in Alzheimer's disease. J Neuroinflammation. 2020;17(1):1–14. https://doi.org/10.1186/s12974-020-01742-3.
- Desale SE, Chinnathambi S. Phosphoinositides signaling modulates microglial actin remodeling and phagocytosis in Alzheimer's disease.Cell Commun Signal. 2021;24;19(1):28. https://doi.org/10.1186/s12964-021-00715-0.
- Chidambaram H, Das R, Chinnathambi S. Interaction of Tau with the chemokine receptor, CX3CR1 and its effect on microglial activation, migration and proliferation. Cell Biosci. 2020;10(1):1–9. https://doi.org/10.11 86/s13578-020-00474-4.
- ElAli A, Rivest S. Microglia ontology and signaling. Front Cell Dev Biol. 2016;
  4:72. https://doi.org/10.3389/fcell.2016.00072.
- Schlessinger J. Cell signaling by receptor tyrosine kinases. Cell. 2000;103(2): 211–25. https://doi.org/10.1016/s0092-8674(00)00114-8.
- Renauld J-C. Class II cytokine receptors and their ligands: key antiviral and inflammatory modulators. Nat Rev Immunol. 2003;3(8):667–76. https://doi. org/10.1038/nri1153.
- Samelson LE. Immunoreceptor signaling. Cold Spring Harb Perspect Biol. 2011;3(12):a011510. https://doi.org/10.1101/cshperspect.a011510.
- 37. Pierce KL, Premont RT, Lefkowitz RJ. Seven-transmembrane receptors. Nat Rev Mol Cell Biol. 2002;3(9):639–50. https://doi.org/10.1038/nrm908.
- Heldin C-H, Lu B, Evans R, Gutkind JS. Signals and receptors. Cold Spring Harb Perspect Biol. 2016;8(4):a005900. https://doi.org/10.1101/cshperspect.a 005900.
- Newton AC, Bootman MD, Scott JD. Second messengers. Cold Spring Harb Perspect Biol. 2016;8(8):a005926. https://doi.org/10.1101/cshperspect.a005926.
- Gutkind JS. Regulation of mitogen-activated protein kinase signaling networks by G protein-coupled receptors. Sci STKE. 2000;2000(40):re1-re. https://doi.org/10.1126/stke.2000.40.re1.
- Chiarini A, Armato U, Liu D, Dal Pl. Calcium-sensing receptor antagonist NPS 2143 restores amyloid precursor protein physiological non-amyloidogenic processing in Aβ-exposed adult human astrocytes. Sci Rep. 2017;7(1):1–15. https://doi.org/10.1038/s41598-017-01215-3.
- Gómez-Ramos A, Díaz-Hernández M, Cuadros R, Hernández F, Avila J. Extracellular tau is toxic to neuronal cells. FEBS Lett. 2006;580(20):4842–50. https://doi.org/10.1016/j.febslet.2006.07.078.
- Gómez-Ramos A, Díaz-Hernández M, Rubio A, Díaz-Hernández JI, Miras-Portugal MT, Avila J. Characteristics and consequences of muscarinic receptor activation by tau protein. Eur Neuropsychopharmacol. 2009;19(10): 708–17. https://doi.org/10.1016/j.euroneuro.2009.04.006.
- Gomez-Ramos A, Diaz-Hernandez M, Rubio A, Miras-Portugal M, Avila J. Extracellular tau promotes intracellular calcium increase through M1 and M3 muscarinic receptors in neuronal cells. Mol Cell Neurosci. 2008;37(4):673–81. https://doi.org/10.1016/j.mcn.2007.12.010.
- Chidambaram H, Chinnathambi S. G-Protein coupled receptors and taudifferent roles in alzheimer's disease. Neurosci. 2020. https://doi.org/10.1016/ j.neuroscience.2020.04.019.
- Thathiah A, De Strooper B. The role of G protein-coupled receptors in the pathology of Alzheimer's disease. Nat Rev Neurosci. 2011;12(2):73–87. https://doi.org/10.1038/nrn2977.
- Zhao J, Deng Y, Jiang Z, Qing H. G protein-coupled receptors (GPCRs) in Alzheimer's disease: a focus on BACE1 related GPCRs. Front Aging Neurosci. 2016;8:58. https://doi.org/10.3389/fnagi.2016.00058.
- 48. Wennerberg K, Rossman KL, Der CJ. The Ras superfamily at a glance. J Cell Sci. 2005;118(5):843–6. https://doi.org/10.1242/jcs.01660.
- Coso OA, Chiariello M, Yu J-C, Teramoto H, Crespo P, Xu N, et al. The small GTP-binding proteins Rac1 and Cdc42regulate the activity of the JNK/SAPK signaling pathway. Cell. 1995;81(7):1137–46. https://doi.org/10.1016/s0092-8674(05)80018-2.
- Venkatakrishnan A, Deupi X, Lebon G, Tate CG, Schertler GF, Babu MM. Molecular signatures of G-protein-coupled receptors. Nature. 2013; 494(7436):185–94. https://doi.org/10.1038/nature11896.

- Rosenbaum DM, Rasmussen SG, Kobilka BK. The structure and function of G-protein-coupled receptors. Nature. 2009;459(7245):356–63. https://doi. org/10.1038/nature08144.
- Ferguson SS. Phosphorylation-independent attenuation of GPCR signalling. Trends Pharmacol Sci. 2007;28(4):173–9. https://doi.org/10.1016/j.tips.2007. 02.008
- 53. Ritter SL, Hall RA. Fine-tuning of GPCR activity by receptor-interacting proteins. Nat Rev Mol Cell Biol. 2009;10(12):819–30. https://doi.org/10.1038/nrm2803.
- Bhattacharya M, Babwah A, Ferguson S. Small GTP-binding protein-coupled receptors. Biochem Soc Trans. 2004;32(6):1040-4. https://doi.org/10.1042/ BST0321040.
- New DC, Wu K, Kwok AW, Wong YH. G protein-coupled receptor-induced Akt activity in cellular proliferation and apoptosis. FEBS J. 2007;274(23):6025–36. https://doi.org/10.1111/j.1742-4658.2007.06116.x.
- Nakano N, Matsuda S, Ichimura M, Minami A, Ogino M, Murai T, et al. PI3K/ AKT signaling mediated by G protein-coupled receptors is involved in neurodegenerative Parkinson's disease. Int J Mol Med. 2017;39(2):253–60. https://doi.org/10.3892/ijmm.2016.2833.
- Law NC, White MF, Hunzicker-Dunn ME. G protein-coupled receptors (GPCRs) that signal via protein kinase A (PKA) cross-talk at insulin receptor substrate 1 (IRS1) to activate the phosphatidylinositol 3-kinase (PI3K)/AKT pathway. J Biol Chem. 2016;291(53):27160–9. https://doi.org/10.1074/jbc.M116.763235.
- Zamani A, Qu Z. Serotonin activates angiogenic phosphorylation signaling in human endothelial cells. FEBS Lett. 2012;586(16):2360–5. https://doi.org/1 0.1016/j.febslet.2012.05.047.
- Yang R-H, Lin J, Hou X-H, Cao R, Yu F, Liu H-Q, et al. Effect of docosahexaenoic acid on hippocampal neurons in high-glucose condition: involvement of PI3K/AKT/nuclear factor-κB-mediated inflammatory pathways. Neurosci. 2014;274:218–28. https://doi.org/10.1016/j. neuroscience.2014.05.042.
- Guixà-González R, Javanainen M, Gómez-Soler M, Cordobilla B, Domingo JC, Sanz F, et al. Membrane omega-3 fatty acids modulate the oligomerisation kinetics of adenosine A 2A and dopamine D 2 receptors. Sci Rep. 2016;6: 19839. https://doi.org/10.1038/srep19839.
- Zhuang J-Y, Chen Z-Y, Zhang T, Tang D-P, Jiang X-Y, Zhuang Z-H. Effects of Different Ratio of n-6/n-3 Polyunsaturated Fatty Acids on the PI3K/Akt Pathway in Rats with Reflux Esophagitis. Med Sci Mon Int Med J Exp Clin Res. 2017;23:542. https://doi.org/10.12659/msm.898131.
- Pu J, Liu P. Fatty acids stimulate glucose uptake by the PI3K/AMPK/Akt and PI3K/ERK1/2 pathways. Protein Phosphorylation Hum Health. 2012;129. https://doi.org/10.5772/52456.
- Vanhaesebroeck B, Alessi DR. The PI3K–PDK1 connection: more than just a road to PKB. Biochem. 2000;346(3):561–76. 10698680.
- Pearce LR, Komander D, Alessi DR. The nuts and bolts of AGC protein kinases. Nat Rev Mol Cell Biol. 2010;11(1):9–22. https://doi.org/10.1038/nrm2822.
- Hanada M, Feng J, Hemmings BA. Structure, regulation and function of PKB/ AKT—a major therapeutic target. Biochim Biophys Acta Proteins Proteomics. 2004;1697(1-2):3–16. https://doi.org/10.1016/j.bbapap.2003.11.009.
- Alessi DR, James SR, Downes CP, Holmes AB, Gaffney PR, Reese CB, et al. Characterization of a 3-phosphoinositide-dependent protein kinase which phosphorylates and activates protein kinase Ba. Curr Biol. 1997;7(4):261–9. https://doi.org/10.1016/s0960-9822(06)00122-9.
- Wu H, Lu D, Jiang H, Xiong Y, Qu C, Li B, et al. Simvastatin-mediated upregulation of VEGF and BDNF, activation of the PI3K/Akt pathway, and increase of neurogenesis are associated with therapeutic improvement after traumatic brain injury. J Neurotrauma. 2008;25(2):130–9. https://doi.org/10.1 089/neu.2007.0369.
- Kitagishi Y, Nakanishi A, Ogura Y, Matsuda S. Dietary regulation of Pl3K/AKT/ GSK-3β pathway in Alzheimer's disease. Alzheimer's Res Ther. 2014;6(3):35. https://doi.org/10.1186/alzrt265.
- Lee CW-C, Lau K-F, Miller CC, Shaw P-C. Glycogen synthase kinase-3β-mediated tau phosphorylation in cultured cell lines. Neuroreport. 2003;14(2): 257–60. https://doi.org/10.1097/00001756-200302100-00020.
- Zempel H, Thies E, Mandelkow E, Mandelkow E-M. Aβ oligomers cause localized Ca2+ elevation, missorting of endogenous Tau into dendrites, Tau phosphorylation, and destruction of microtubules and spines. J Neurosci. 2010;30(36):11938–50. https://doi.org/10.1523/JNEUROSCI.2357-10.2010.
- Nitsch RM, Slack BE, Wurtman RJ, Growdon JH. Release of Alzheimer amyloid precursor derivatives stimulated by activation of muscarinic acetylcholine receptors. Science. 1992;258(5080):304–7. https://doi.org/1 0.1126/science.1411529.

Desale et al. Molecular Biomedicine (2021) 2:17 Page 15 of 18

- Teng L, Zhao J, Wang F, Ma L, Pei G. A GPCR/secretase complex regulates β-and γ-secretase specificity for Aβ production and contributes to AD pathogenesis. Cell Res. 2010;20(2):138–53. https://doi.org/10.1038/cr.2010.3.
- Kieffer BL, Gavériaux-Ruff C. Exploring the opioid system by gene knockout. Prog Neurobiol. 2002;66(5):285–306. https://doi.org/10.1016/s0301-0082(02)00008-4.
- Angulo E, Casadó V, Mallol J, Canela El, Viñals F, Ferrer I, et al. A1 adenosine receptors accumulate in neurodegenerative structures in Alzheimer's disease and mediate both amyloid precursor protein processing and tau phosphorylation and translocation. Brain Pathol. 2003;13(4):440–51. https:// doi.org/10.1111/j.1750-3639.2003.tb00475.x.
- 75. Takai Y, Sasaki T, Matozaki T. Small GTP-binding proteins. Physiol Rev. 2001; 81(1):153–208. https://doi.org/10.1152/physrev.2001.81.1.153.
- Bouchon A, Dietrich J, Colonna M. Cutting edge: inflammatory responses can be triggered by TREM-1, a novel receptor expressed on neutrophils and monocytes. J Immunol. 2000;164(10):4991–5. https://doi.org/10.4049/ iimmunol.164.10.4991.
- Ulrich JD, Holtzman DM. TREM2 function in Alzheimer's disease and neurodegeneration. ACS Chem Nerosci. 2016;7(4):420–7. https://doi.org/10.1 021/acschemneuro.5b00313.
- Takahashi K, Rochford CD, Neumann H. Clearance of apoptotic neurons without inflammation by microglial triggering receptor expressed on myeloid cells-2. J Exp Med. 2005;201(4):647–57. https://doi.org/10.1084/jem.2 0041611.
- Peng Q, Malhotra S, Torchia JA, Kerr WG, Coggeshall KM, Humphrey MB. TREM2-and DAP12-dependent activation of PI3K requires DAP10 and is inhibited by SHIP1. Sci Signal. 2010;3(122):ra38-ra. https://doi.org/10.1126/ scisignal.2000500.
- Deczkowska A, Weiner A, Amit I. The physiology, pathology, and potential therapeutic applications of the TREM2 signaling pathway. Cell. 2020;181(6): 1207–17. https://doi.org/10.1016/j.cell.2020.05.003.
- Mecca C, Giambanco I, Donato R, Arcuri C. Microglia and aging: the role of the TREM2–DAP12 and CX3CL1-CX3CR1 axes. Int J Mol Sci. 2018;19(1):318. https://doi.org/10.3390/ijms19010318.
- J-i S. Molecular network of microRNA targets in Alzheimer's disease brains. Exp Neurol. 2012;235(2):436–46. https://doi.org/10.1016/j. expneurol.2011.09.003.
- 83. Turnbull IR, Colonna M. Activating and inhibitory functions of DAP12. Nat Rev Immunol. 2007;7(2):155–61. https://doi.org/10.1038/nri2014.
- Ji S, Tabunoki H, Ishida T, Yagishita S, Jinnai K, Futamura N, et al. Phosphorylated Syk expression is enhanced in Nasu-Hakola disease brains. Neuropathology. 2012;32(2):149–57. https://doi.org/10.1111/j.1440-1789.2 011.01256.x.
- Hsieh CL, Koike M, Spusta SC, Niemi EC, Yenari M, Nakamura MC, et al. A role for TREM2 ligands in the phagocytosis of apoptotic neuronal cells by microglia. J Neurochem. 2009;109(4):1144–56. https://doi.org/10.1111/j.14 71-4159.2009.06042.x.
- Kober DL, Brett TJ. TREM2-ligand interactions in health and disease. J Mol Biol. 2017;429(11):1607–29. https://doi.org/10.1016/j.jmb.2017.04.004.
- 87. Charles JF, Humphrey MB, Zhao X, Quarles E, Nakamura MC, Aderem A, et al. The innate immune response to Salmonella enterica serovar Typhimurium by macrophages is dependent on TREM2-DAP12. Infect Immun. 2008;76(6):2439–47. https://doi.org/10.1128/IAI.00115-08.
- Romashkova JA, Makarov SS. NF-kB is a target of AKT in anti-apoptotic PDGF signalling. Nature. 1999;401(6748):86–90. https://doi.org/10.103 8/43474.
- Zhu M, Li D, Wu Y, Huang X, Wu M. TREM-2 promotes macrophagemediated eradication of Pseudomonas aeruginosa via a PI3K/Akt pathway. Scand J Immunol. 2014;79(3):187–96. https://doi.org/10.1111/sji.12148.
- Zheng H, Jia L, Liu C-C, Rong Z, Zhong L, Yang L, et al. TREM2 promotes microglial survival by activating Wnt/β-catenin pathway. J Neurosci. 2017; 37(7):1772–84. https://doi.org/10.1523/JNEUROSCI.2459-16.2017.
- Hamerman JA, Yu YL, Shen KY, Lowell CA, Lanier LL, Chu CL. Increased TLR responses in dendritic cells lacking the ITAM-containing adapters Dap12 and FcRy. Eur J Immunol. 2009;38(1):166-73. https://doi.org/10.1002/eji.20073 7600.
- Zhong L, Chen X-F, Zhang Z-L, Wang Z, Shi X-Z, Xu K, et al. DAP12 stabilizes the C-terminal fragment of the triggering receptor expressed on myeloid cells-2 (TREM2) and protects against LPS-induced pro-inflammatory response. J Biol Chem. 2015;290(25):15866–77. https://doi.org/10.1074/jbc. M115.645986.

- N'Diaye E-N, Branda CS, Branda SS, Nevarez L, Colonna M, Lowell C, et al. TREM-2 (triggering receptor expressed on myeloid cells 2) is a phagocytic receptor for bacteria. J Cell Biol. 2009;184(2):215–23. https://doi.org/10.1083/ icb.200808080.
- 94. Painter MM, Atagi Y, Liu C-C, Rademakers R, Xu H, Fryer JD, et al. TREM2 in CNS homeostasis and neurodegenerative disease. Mol Neurodegener. 2015; 10(1):1–10. https://doi.org/10.1186/s13024-015-0040-9.
- Yao H, Coppola K, Schweig JE, Crawford F, Mullan M, Paris D. Distinct signaling pathways regulate TREM2 phagocytic and NFkB antagonistic activities. Front Cell Neurosci. 2019;13. https://doi.org/10.3389/fncel.2019.004
- Zhao Y, Wu X, Li X, Jiang L-L, Gui X, Liu Y, et al. TREM2 is a receptor for βamyloid that mediates microglial function. Neuron. 2018;97(5):1023–31. e7. https://doi.org/10.1016/j.neuron.2018.01.031.
- Zhong L, Wang Z, Wang D, Wang Z, Martens YA, Wu L, et al. Amyloid-beta modulates microglial responses by binding to the triggering receptor expressed on myeloid cells 2 (TREM2). Mol Neurodegener. 2018;13(1):1–12. https://doi.org/10.1186/s13024-018-0247-7.
- Xiang X, Werner G, Bohrmann B, Liesz A, Mazaheri F, Capell A, et al. TREM2 deficiency reduces the efficacy of immunotherapeutic amyloid clearance. EMBO Mol Med. 2016;8(9):992–1004. https://doi.org/10.15252/emmm.201 606370.
- Jonsson T, Stefansson H, Steinberg S, Jonsdottir I, Jonsson PV, Snaedal J, et al. Variant of TREM2 associated with the risk of Alzheimer's disease. N Engl J Med. 2013;368(2):107–16. https://doi.org/10.1056/NEJMoa1211103.
- Dean HB, Roberson ED, Song Y. Neurodegenerative disease—Associated variants in TREM2 destabilize the apical ligand-binding region of the immunoglobulin domain. Front Neurol. 2019;10:1252. https://doi.org/10.33 89/fneur.2019.01252.
- Lessard CB, Malnik SL, Zhou Y, Ladd TB, Cruz PE, Ran Y, et al. High-affinity interactions and signal transduction between Aβ oligomers and TREM 2. EMBO Mol Med. 2018;10(11):e9027. https://doi.org/10.15252/emmm.201 809027.
- 102. Rivera EJ, Goldin A, Fulmer N, Tavares R, Wands JR, de la Monte SM. Insulin and insulin-like growth factor expression and function deteriorate with progression of Alzheimer's disease: link to brain reductions in acetylcholine. J Alzheimers Dis. 2005;8(3):247–68. https://doi.org/10.15252/emmm.201 809027.
- Ulland TK, Song WM, Huang SC-C, Ulrich JD, Sergushichev A, Beatty WL, et al. TREM2 maintains microglial metabolic fitness in Alzheimer's disease. Cell. 2017;170(4):649–63. e13. https://doi.org/10.1016/j.cell.2017.07.023.
- 104. Keren-Shaul H, Spinrad A, Weiner A, Matcovitch-Natan O, Dvir-Szternfeld R, Ulland TK, et al. A unique microglia type associated with restricting development of Alzheimer's disease. Cell. 2017;169(7):1276–90. e17. https://doi.org/10.1016/j.cell.2017.05.018.
- 105. Chen S, Peng J, Sherchan P, Ma Y, Xiang S, Yan F, et al. TREM2 activation attenuates neuroinflammation and neuronal apoptosis via PI3K/Akt pathway after intracerebral hemorrhage in mice. J Neuroinflammation. 2020;17:1–16. https://doi.org/10.1186/s12974-020-01853-x.
- 106. Bemiller SM, McCray TJ, Allan K, Formica SV, Xu G, Wilson G, et al. TREM2 deficiency exacerbates tau pathology through dysregulated kinase signaling in a mouse model of tauopathy. Mol Neurodegener. 2017;12(1):74. https://doi.org/10.1186/s13024-017-0216-6.
- Bernards A, Settleman J. GAP control: regulating the regulators of small GTPases. Trends Cell Biol. 2004;14(7):377–85. https://doi.org/10.1016/j.tcb.2 004.05.003.
- Schmidt A, Hall A. Guanine nucleotide exchange factors for Rho GTPases: turning on the switch. Genes Dev. 2002;16(13):1587–609. https://doi.org/1 0.1101/gad.1003302.
- Seabra MC, Wasmeier C. Controlling the location and activation of Rab GTPases. Curr Opin Cell Biol. 2004;16(4):451–7. https://doi.org/10.1016/j.ceb.2 004 06 014
- Sit S-T, Manser E. Rho GTPases and their role in organizing the actin cytoskeleton. J Cell Sci. 2011;124(5):679–83. https://doi.org/10.1242/jcs.064964.
- Wojnacki J, Quassollo G, Marzolo M-P, Cáceres A. Rho GTPases at the crossroad of signaling networks in mammals: impact of Rho-GTPases on microtubule organization and dynamics. Small GTPases. 2014;5(1):e28430. https://doi.org/10.4161/sgtp.28430.
- Clark EA, King WG, Brugge JS, Symons M, Hynes RO. Integrin-mediated signals regulated by members of the rho family of GTPases. J Cell Biol. 1998; 142(2):573–86. https://doi.org/10.1083/jcb.142.2.573.

Desale et al. Molecular Biomedicine (2021) 2:17 Page 16 of 18

- Schiller MR. Coupling receptor tyrosine kinases to Rho GTPases—GEFs what's the link. Cell Signal. 2006;18(11):1834–43. https://doi.org/10.1016/j. cellsig.2006.01.022.
- Siehler S. Regulation of RhoGEF proteins by G12/13-coupled receptors. Br J Pharmacol. 2009;158(1):41–9. https://doi.org/10.1111/j.1476-5381.2009.00121 x.
- Dutt P, Jaffe AB, Merdek KD, Hall A, Toksoz D. Gaz inhibits serum response factor-dependent transcription by inhibiting Rho signaling. Mol Pharmacol. 2004;66(6):1508–16. https://doi.org/10.1124/mol.104.002949.
- Fukuhara S, Chikumi H, Gutkind JS. RGS-containing RhoGEFs: the missing link between transforming G proteins and Rho? Oncogene. 2001;20(13): 1661–8. https://doi.org/10.1038/sj.onc.1204182.
- 117. Etienne-Manneville S, Hall A. Rho GTPases in cell biology. Nature. 2002; 420(6916):629–35. https://doi.org/10.1038/nature01148.
- Nobes CD, Hall A. Rho GTPases control polarity, protrusion, and adhesion during cell movement. J Cell Biol. 1999;144(6):1235–44. https://doi.org/10.1 083/icb.1446.1235.
- 119. Hua ZL, Emiliani FE, Nathans J. Rac1 plays an essential role in axon growth and guidance and in neuronal survival in the central and peripheral nervous systems. Neural Dev. 2015;10(1):21. https://doi.org/10.1186/s13064-015-0049-3.
- 120. Ng EL, Tang BL. Rab GTPases and their roles in brain neurons and glia. Brain Res Rev. 2008;58(1):236–46. https://doi.org/10.1016/j.brainresrev.2008.04.006.
- 121. Stenmark H. Rab GTPases as coordinators of vesicle traffic. Nat Rev Mol Cell Biol. 2009;10(8):513–25. https://doi.org/10.1038/nrm2728.
- Wandinger-Ness A, Zerial M. Rab proteins and the compartmentalization of the endosomal system. Cold Spring Harb Perspect Biol. 2014;6(11):a022616. https://doi.org/10.1101/cshperspect.a022616.
- 123. Zhen Y, Stenmark H. Cellular functions of Rab GTPases at a glance. J Cell Sci. 2015;128(17):3171–6. https://doi.org/10.1242/jcs.166074.
- Memon AR. The role of ADP-ribosylation factor and SAR1 in vesicular trafficking in plants. Biochim Biophys Acta Biomembr. 2004;1664(1):9–30. https://doi.org/10.1016/j.bbamem.2004.04.005.
- 125. Weis K. Regulating access to the genome: nucleocytoplasmic transport throughout the cell cycle. Cell. 2003;112(4):441–51. https://doi.org/10.1016/ s0092-8674(03)00082-5.
- 126. Hunter MP, Russo A, O'Bryan JP. Emerging roles for intersectin (ITSN) in regulating signaling and disease pathways. Int J Mol Sci. 2013;14(4):7829–52. https://doi.org/10.3390/ijms14047829.
- 127. Aguilar BJ, Zhu Y, Lu Q. Rho GTPases as therapeutic targets in Alzheimer's disease. Alzheimer's Res Ther. 2017;9(1):97. https://doi.org/10.1186/s13195-017-0320-4.
- Bamburg JR, Bloom GS. Cytoskeletal pathologies of Alzheimer disease. Cell Motil Cytoskeleton. 2009;66(8):635–49. https://doi.org/10.1002/cm.20388.
- Penzes P, VanLeeuwen J-E. Impaired regulation of synaptic actin cytoskeleton in Alzheimer's disease. Brain Res Rev. 2011;67(1-2):184–92. https://doi.org/10.1016/j.brainresrev.2011.01.003.
- 130. Huesa G, Baltrons MA, Gómez-Ramos P, Morán A, García A, Hidalgo J, et al. Altered distribution of RhoA in Alzheimer's disease and AβPP overexpressing mice. J Alzheimers Dis. 2010;19(1):37–56. https://doi.org/1 0.3233/JAD-2010-1203.
- 131. Schill Y, Bijata M, Kopach O, Cherkas V, Abdel-Galil D, Böhm K, et al. Serotonin 5-HT 4 receptor boosts functional maturation of dendritic spines via RhoA-dependent control of F-actin. Commun Biol. 2020;3(1):1–16. https://doi.org/10.1038/s42003-020-0791-x.
- 132. Tsushima H, Emanuele M, Polenghi A, Esposito A, Vassalli M, Barberis A, et al. HDAC6 and RhoA are novel players in Abeta-driven disruption of neuronal polarity. Nat Commun. 2015;6(1):1–14. https://doi.org/10.1038/ncomms8781.
- 133. Petratos S, Li Q-X, George AJ, Hou X, Kerr ML, Unabia SE, et al. The β-amyloid protein of Alzheimer's disease increases neuronal CRMP-2 phosphorylation by a Rho-GTP mechanism. Brain. 2008;131(1):90–108. https://doi.org/10.1093/brain/awm260.
- Nakamura F, Ohshima T, Goshima Y. Collapsin response mediator proteins: their biological functions and pathophysiology in neuronal development and regeneration. Front Cell Neurosci. 2020;14:188. https://doi.org/10.3389/ fncel.2020.00188.
- 135. Chacon PJ, Garcia-Mejias R, Rodriguez-Tebar A. Inhibition of RhoA GTPase and the subsequent activation of PTP1B protects cultured hippocampal neurons against amyloid  $\beta$  toxicity. Mol Neurodegener. 2011;6(1):1–11. https://doi.org/10.1186/1750-1326-6-14.
- 136. Cohen TJ, Guo JL, Hurtado DE, Kwong LK, Mills IP, Trojanowski JQ, et al. The acetylation of tau inhibits its function and promotes pathological tau

- aggregation. Nat Commun. 2011;2(1):1–9. https://doi.org/10.1038/ncomms1255.
- 137. Hubbert C, Guardiola A, Shao R, Kawaguchi Y, Ito A, Nixon A, et al. HDAC6 is a microtubule-associated deacetylase. Nature. 2002;417(6887):455–8. https://doi.org/10.1038/417455a.
- 138. Cap KC, Jung Y-J, Choi BY, Hyeon SJ, Kim J-G, Min J-K, et al. Distinct dual roles of p-Tyr42 RhoA GTPase in tau phosphorylation and ATP citrate lyase activation upon different Aβ concentrations. Redox Biol. 2020;101446. https://doi.org/10.1016/j.redox.2020.101446.
- Corbetta S, Gualdoni S, Ciceri G, Monari M, Zuccaro E, Tybulewicz VL, et al. Essential role of Rac1 and Rac3 GTPases in neuronal development. FASEB J. 2009;23(5):1347–57. https://doi.org/10.1096/fi.08-121574.
- 140. Kurokawa K, Itoh RE, Yoshizaki H, Nakamura YOT, Matsuda M. Coactivation of Rac1 and Cdc42 at lamellipodia and membrane ruffles induced by epidermal growth factor. Mol Biol Cell. 2004;15(3):1003–10. https://doi.org/1 0.1091/mbc.e03-08-0609.
- 141. Szczepanowska J. Involvement of Rac/Cdc42/PAK pathway in cytoskeletal rearrangements. Acta Biochim Pol. 2009;56(2):225-34.
- Zhang H, Webb DJ, Asmussen H, Niu S, Horwitz AF. A GIT1/PIX/Rac/PAK signaling module regulates spine morphogenesis and synapse formation through MLC. J Neurosci. 2005;25(13):3379–88. https://doi.org/10.1523/ JNEUROSCI.3553-04.2005.
- 143. Bernard O. Lim kinases, regulators of actin dynamics. Int J Biochem. 2007; 39(6):1071–6. https://doi.org/10.1016/j.biocel.2006.11.011.
- 144. Acevedo K, Li R, Soo P, Suryadinata R, Sarcevic B, Valova VA, et al. The phosphorylation of p25/TPPP by LIM kinase 1 inhibits its ability to assemble microtubules. Exp Cell Res. 2007;313(20):4091–106. https://doi.org/10.1016/j. yexcr.2007.08.012.
- Woo J-A, Liu T, Fang CC, Cazzaro S, Kee T, LePochat P, et al. Activated cofilin exacerbates tau pathology by impairing tau-mediated microtubule dynamics. Commun Biol. 2019;2(1):1–15. https://doi.org/10.1038/s42003-019-0359-9
- 146. Zhao L, Ma Q-L, Calon F, Harris-White ME, Yang F, Lim GP, et al. Role of p21-activated kinase pathway defects in the cognitive deficits of Alzheimer disease. Nat Neurosci. 2006;9(2):234–42. https://doi.org/10.1038/nn1630.
- 147. Maillet M, Robert SJ, Cacquevel M, Gastineau M, Vivien D, Bertoglio J, et al. Crosstalk between Rap1 and Rac regulates secretion of sAPPa. Nat Cell Biol. 2003;5(7):633–9. https://doi.org/10.1038/ncb1007.
- 148. Wang PL, Niidome T, Akaike A, Kihara T, Sugimoto H. Rac1 inhibition negatively regulates transcriptional activity of the amyloid precursor protein gene. J Neurosci Res. 2009;87(9):2105–14. https://doi.org/10.1002/jnr.22039.
- Nimnual AS, Taylor LJ, Bar-Sagi D. Redox-dependent downregulation of Rho by Rac. Nat Cell Biol. 2003;5(3):236–41. https://doi.org/10.1038/ncb938.
- Sander EE, ten Klooster JP, van Delft S, van der Kammen RA, Collard JG. Rac downregulates Rho activity: reciprocal balance between both GTPases determines cellular morphology and migratory behavior. J Cell Biol. 1999; 147(5):1009–22. https://doi.org/10.1083/jcb.147.5.1009.
- 151. Borin M, Saraceno C, Catania M, Lorenzetto E, Pontelli V, Paterlini A, et al. Rac1 activation links tau hyperphosphorylation and A $\beta$  dysmetabolism in Alzheimer's disease. Acta Neuropathol Commun. 2018;6(1):61. https://doi.org/10.1186/s40478-018-0567-4.
- Pereira-Leal JB, Seabra MC. Evolution of the Rab family of small GTP-binding proteins. J Mol Biol. 2001;313(4):889–901. https://doi.org/10.1006/jmbi.2001.5072.
- 153. Ridge PG, Karch CM, Hsu S, Arano I, Teerlink CC, Ebbert MT, et al. Linkage, whole genome sequence, and biological data implicate variants in RAB10 in Alzheimer's disease resilience. Genome Med. 2017;9(1):1–14. https://doi.org/10.1186/s13073-017-0486-1.
- 154. Udayar V, Buggia-Prévot V, Guerreiro RL, Siegel G, Rambabu N, Soohoo AL, et al. A paired RNAi and RabGAP overexpression screen identifies Rab11 as a regulator of β-amyloid production. Cell Rep. 2013;5(6):1536–51. https://doi.org/10.1016/j.celrep.2013.12.005.
- Vardarajan BN, Bruesegem SY, Harbour ME, George-Hyslop PS, Seaman MN, Farrer LA. Identification of Alzheimer disease-associated variants in genes that regulate retromer function. Neurobiol Aging. 2012;33(9):2231. e15–30. https://doi.org/10.1016/j.neurobiolaging.2012.04.020.
- 156. Ginsberg SD, Mufson EJ, Counts SE, Wuu J, Alldred MJ, Nixon RA, et al. Regional selectivity of rab5 and rab7 protein upregulation in mild cognitive impairment and Alzheimer's disease. J Alzheimers Dis. 2010;22(2):631–9. https://doi.org/10.3233/JAD-2010-101080.
- 157. Ginsberg SD, Alldred MJ, Counts SE, Cataldo AM, Neve RL, Jiang Y, et al. Microarray analysis of hippocampal CA1 neurons implicates early

Desale et al. Molecular Biomedicine (2021) 2:17 Page 17 of 18

- endosomal dysfunction during Alzheimer's disease progression. Biol Psychiatry. 2010;68(10):885–93. https://doi.org/10.1016/j.biopsych.2010.05.030.
- Laifenfeld D, Patzek LJ, McPhie DL, Chen Y, Levites Y, Cataldo AM, et al. Rab5 mediates an amyloid precursor protein signaling pathway that leads to apoptosis. J Neurosci. 2007;27(27):7141–53. https://doi.org/10.1523/ JNEUROSCI.4599-06.2007.
- 159. Buggia-Prévot V, Fernandez CG, Riordan S, Vetrivel KS, Roseman J, Waters J, et al. Axonal BACE1 dynamics and targeting in hippocampal neurons: a role for Rab11 GTPase. Mol Neurodegener. 2014;9(1):1–18. https://doi.org/10.1186/1750-1326-9-1.
- 160. Guerra F, Bucci C. Multiple roles of the small GTPase Rab7. Cells. 2016;5(3): 34. https://doi.org/10.3390/cells5030034.
- Rodriguez L, Mohamed NV, Desjardins A, Lippé R, Fon EA, Leclerc N. Rab7A regulates tau secretion. J Neurochem. 2017;141(4):592–605. https://doi.org/1 0.1111/jnc.13994.
- 162. Zafar S, Younas N, Correia S, Shafiq M, Tahir W, Schmitz M, et al. Strain-specific altered regulatory response of Rab7a and Tau in Creutzfeldt-Jakob disease and Alzheimer's disease. Mol Neurobiol. 2017;54(1):697–709. https://doi.org/10.1007/s12035-016-9694-8.
- Aikawa S, Hashimoto T, Kano K, Aoki J. Lysophosphatidic acid as a lipid mediator with multiple biological actions. J Biochem. 2015;157(2):81–9. https://doi.org/10.1093/jb/mvu077.
- 164. Anliker B, Choi JW, Lin ME, Gardell SE, Rivera RR, Kennedy G, et al. Lysophosphatidic acid (LPA) and its receptor, LPA1, influence embryonic schwann cell migration, myelination, and cell-to-axon segregation. Glia. 2013;61(12):2009–22. https://doi.org/10.1002/glia.22572.
- Zimmermann H, Zebisch M, Sträter N. Cellular function and molecular structure of ecto-nucleotidases. Purinergic Signal. 2012;8(3):437–502. https://doi.org/10.1007/s11302-012-9309-4.
- 166. Ramesh S, Govindarajulu M, Suppiramaniam V, Moore T, Dhanasekaran M. Autotaxin–lysophosphatidic acid signaling in Alzheimer's disease. Int J Mol Sci. 2018;19(7):1827. https://doi.org/10.3390/ijms19071827.
- Pagès C, Simon M-F, Valet P, Saulnier-Blache JS. Lysophosphatidic acid synthesis and release. Prostaglandins Other Lipid Mediat. 2001;64(1-4):1–10. https://doi.org/10.1016/s0090-6980(01)00110-1.
- Yung YC, Stoddard NC, Mirendil H, Chun J. Lysophosphatidic acid signaling in the nervous system. Neuron. 2015;85(4):669–82. https://doi.org/10.1016/j. neuron.2015.01.009.
- Choi JW, Chun J. Lysophospholipids and their receptors in the central nervous system. Biochim Biophys Acta. 2013;1831(1):20–32. https://doi.org/1 0.1016/j.bbalip.2012.07.015.
- Frisca F, Sabbadini RA, Goldshmit Y, Pébay A. Biological effects of lysophosphatidic acid in the nervous system. Int Rev Cell Mol Biol. Elsevier. 2012;273–322. https://doi.org/10.1016/B978-0-12-394307-1.00005-9.
- Dubin AE, Herr DR, Chun J. Diversity of lysophosphatidic acid receptormediated intracellular calcium signaling in early cortical neurogenesis. J Neurosci. 2010;30(21):7300–9. https://doi.org/10.1523/JNEUROSCI.6151-09.2010.
- 172. Kingsbury MA, Rehen SK, Contos JJ, Higgins CM, Chun J. Non-proliferative effects of lysophosphatidic acid enhance cortical growth and folding. Nat Neurosci. 2003;6(12):1292–9. https://doi.org/10.1038/nn1157.
- 173. Dubin AE, Bahnson T, Weiner JA, Fukushima N, Chun J. Lysophosphatidic acid stimulates neurotransmitter-like conductance changes that precede GABA and L-glutamate in early, presumptive cortical neuroblasts. J Neurosci. 1999;19(4):1371–81. https://doi.org/10.1 523/JNEUROSCI.19-04-01371.1999.
- 174. Ishii I, Fukushima N, Ye X, Chun J. Lysophospholipid receptors: signaling and biology. Annu Rev Biochem. 2004;73(1):321–54. https://doi.org/10.1146/annurev.biochem.73.011303.073731.
- Pilpel Y, Segal M. Activation of PKC induces rapid morphological plasticity in dendrites of hippocampal neurons via Rac and Rho-dependent mechanisms. Eur J Neurosci. 2004;19(12):3151–64. https://doi.org/10.1111/j. 0953-816X.2004.03380.x.
- Pilpel Y, Segal M. The role of LPA1 in formation of synapses among cultured hippocampal neurons. J Neurochem. 2006;97(5):1379–92. https://doi.org/1 0.1111/i.1471-4159.2006.03825.x.
- An S, Bleu T, Zheng Y, Goetzl EJ. Recombinant human G protein-coupled lysophosphatidic acid receptors mediate intracellular calcium mobilization. Mol Pharmacol. 1998;54(5):881–8. https://doi.org/10.1124/mol.54.5.881.
- 178. Hecht JH, Weiner JA, Post SR, Chun J. Ventricular zone gene-1 (vzg-1) encodes a lysophosphatidic acid receptor expressed in neurogenic regions of the developing cerebral cortex. J Cell Biol. 1996;135(4):1071–83. https://doi.org/10.1083/jcb.135.4.1071.

- Furuta D, Yamane M, Tsujiuchi T, Moriyama R, Fukushima N.
  Lysophosphatidic acid induces neurite branch formation through LPA3. Mol Cell Neurosci. 2012;50(1):21–34. https://doi.org/10.1016/j.mcn.2012.03.006.
- Lee Z, Cheng C-T, Zhang H, Subler MA, Wu J, Mukherjee A, et al. Role of LPA4/p2y9/GPR23 in negative regulation of cell motility. Mol Biol Cell. 2008; 19(12):5435–45. https://doi.org/10.1091/mbc.e08-03-0316.
- 181. Jongsma M, Matas-Rico E, Rzadkowski A, Jalink K, Moolenaar WH. LPA is a chemorepellent for B16 melanoma cells: action through the cAMP-elevating LPA 5 receptor. PLoS One. 2011;6(12):e29260. https://doi.org/10.1371/journal. pone.0029260.
- 182. Pasternack SM, von Kügelgen I, Al Aboud K, Lee Y-A, Rüschendorf F, Voss K, et al. G protein–coupled receptor P2Y5 and its ligand LPA are involved in maintenance of human hair growth. Nat Genet. 2008;40(3):329–34. https://doi.org/10.1038/ng.84.
- 183. Sun Y-X, Minthon L, Wallmark A, Warkentin S, Blennow K, Janciauskiene S. Inflammatory markers in matched plasma and cerebrospinal fluid from patients with Alzheimer's disease. Dement Geriatr Cogn Disord. 2003;16(3): 136–44. https://doi.org/10.1159/000071001.
- 184. Frisardi V, Panza F, Seripa D, Farooqui T, Farooqui AA. Glycerophospholipids and glycerophospholipid-derived lipid mediators: a complex meshwork in Alzheimer's disease pathology. Prog Lipid Res. 2011;50(4):313–30. https://doi.org/10.1016/j.plipres.2011.06.001.
- 185. Takahashi M, Tomizawa K, Kato R, Sato K, Uchida T, Fujita SC, et al. Localization and developmental changes of  $\tau$  protein kinase l/glycogen synthase kinase-3 $\beta$  in rat brain. J Neurochem. 1994;63(1):245–55. https://doi.org/10.1046/j.1471-4159.1994.63010245.x.
- Sayas CL, Moreno-Flores MT, Avila J, Wandosell F. The neurite retraction induced by lysophosphatidic acid increases Alzheimer's disease-like Tau phosphorylation. J Biol Chem. 1999;274(52):37046–52. https://doi.org/10.1 074/jbc.274.52.37046.
- 187. Sun Y, Kim N-H, Yang H, Kim S-H, Huh S-O. Lysophosphatidic acid induces neurite retraction in differentiated neuroblastoma cells via GSK-3 $\beta$  activation. Mol Cells. 2011;31(5):483. https://doi.org/10.1007/s10059-011-103
- 188. Krupnick JG, Benovic JL. The role of receptor kinases and arrestins in G protein–coupled receptor regulation. Annu Rev Pharmacol. 1998;38(1):289–319. https://doi.org/10.1146/annurev.pharmtox.38.1.289.
- Freedman NJ, Lefkowitz RJ. Desensitization of G protein-coupled receptors.
  Recent Prog Horm Res. 1996;51:319–51 discussion 52. PMID: 8701085.
- Pitcher JA, Freedman NJ, Lefkowitz RJ. G protein–coupled receptor kinases. Annu Rev. 1998. https://doi.org/10.1146/annurev.biochem.67.1.653.
- Bünemann M, Hosey MM. G-protein coupled receptor kinases as modulators of G-protein signalling. J Physiol. 1999;517(1):5–23. https://doi.org/10.1111/ i1469-7793199900057x
- 192. Wilden U, Hall S, Kühn H. Phosphodiesterase activation by photoexcited rhodopsin is quenched when rhodopsin is phosphorylated and binds the intrinsic 48-kDa protein of rod outer segments. Proc Natl Acad Sci. 1986; 83(5):1174–8. https://doi.org/10.1073/pnas.83.5.1174.
- Carman CV, Benovic JL. G-protein-coupled receptors: turn-ons and turn-offs. Curr Opin Neurobiol. 1998;8(3):335–44. https://doi.org/10.1016/s0959-43 88(98)80058-5.
- DeWire SM, Ahn S, Lefkowitz RJ, Shenoy SK. Beta-arrestins and cell signaling. Annu Rev Physiol. 2007;69:483. https://doi.org/10.1146/annurev.physiol.69. 022405.154749.
- 195. Jean-Charles P-Y, Kaur S, Shenoy SK. GPCR signaling via β-arrestin-dependent mechanisms. J Cardiovasc Pharmacol. 2017;70(3):142. https://doi.org/10.1097/FJC.0000000000000482.
- Mohan LM, Vasudevan NT, Gupta MK, Martelli EE, Naga Prasad SV. G-protein coupled receptor resensitization-appreciating the balancing act of receptor function. Curr Mol Pharmacol. 2012;5(3):350–61 PMID: 22697395.
- Marchese A, Paing MM, Temple BR, Trejo J. G protein–coupled receptor sorting to endosomes and lysosomes. Annu Rev Pharmacol Toxicol. 2008;48: 601–29. https://doi.org/10.1146/annurev.pharmtox.48.113006.094646.
- 198. Thathiah A, Horre K, Snellinx A, Vandewyer E, Huang Y, Ciesielska M, et al. β-arrestin 2 regulates Aβ generation and γ-secretase activity in Alzheimer's disease. Nat Med. 2013;19(1):43–9. https://doi.org/10.1038/nm.3023.
- 199. Ni Y, Zhao X, Bao G, Zou L, Teng L, Wang Z, et al. Activation of  $\beta$  2-adrenergic receptor stimulates  $\gamma$ -secretase activity and accelerates amyloid plaque formation. Nat Med. 2006;12(12):1390–6. https://doi.org/10.1038/nm1485.
- 200. Thathiah A, Spittaels K, Hoffmann M, Staes M, Cohen A, Horré K, et al. The orphan G protein–coupled receptor 3 modulates amyloid-beta peptide

Desale et al. Molecular Biomedicine (2021) 2:17 Page 18 of 18

- generation in neurons. Science. 2009;323(5916):946–51. https://doi.org/10.1126/science.1160649.
- 201. Liu X, Zhao X, Zeng X, Bossers K, Swaab DF, Zhao J, et al. β-arrestin1 regulates γ-secretase complex assembly and modulates amyloid-β pathology. Cell Res. 2013;23(3):351–65. https://doi.org/10.1038/cr.2012.167.
- Yamagata K, Sanders LK, Kaufmann WE, Yee W, Barnes CA, Nathans D, et al. rheb, a growth factor-and synaptic activity-regulated gene, encodes a novel Ras-related protein. J Biol Chem. 1994;269(23):16333–9 PMID: 8206940.
- Neuman NA, Henske EP. Non-canonical functions of the tuberous sclerosis complex-Rheb signalling axis. EMBO Mol Med. 2011;3(4):189–200. https://doi.org/10.1002/emmm.201100131.
- 204. Shahani N, Pryor W, Swarnkar S, Kholodilov N, Thinakaran G, Burke RE, et al. Rheb GTPase regulates  $\beta$ -secretase levels and amyloid  $\beta$  generation. J Biol Chem. 2014;289(9):5799–808. https://doi.org/10.1074/jbc.M113.532713.
- Moser E, Kargl J, Whistler JL, Waldhoer M, Tschische P. G protein-coupled receptor-associated sorting protein 1 regulates the postendocytic sorting of seven-transmembrane-spanning G protein-coupled receptors. Pharmacology. 2010;86(1):22–9. https://doi.org/10.1159/000314161.
- Heese K, Yamada T, Akatsu H, Yamamoto T, Kosaka K, Nagai Y, et al. Characterizing the new transcription regulator protein p60TRP. J Cell Biochem. 2004;91(5):1030–42. https://doi.org/10.1002/jcb.20010.
- 207. Mishra M, Heese K. P60TRP interferes with the GPCR/secretase pathway to mediate neuronal survival and synaptogenesis. J Cell Mol Med. 2011;15(11): 2462–77. https://doi.org/10.1111/j.1582-4934.2010.01248x.
- Hauser AS, Attwood MM, Rask-Andersen M, Schiöth HB, Gloriam DE. Trends in GPCR drug discovery: new agents, targets and indications. Nat Rev Drug Discov. 2017;16(12):829–42. https://doi.org/10.1038/nrd.2017.178.
- Wallach J, Colestock T, Adejare A. Receptor targets in Alzheimer's disease drug discovery. Drug Discov Approaches Treat Neurodegener Disord. Elsevier. 2017:83–107. https://doi.org/10.1016/B978-0-12-802810-0.00006-4.
- 210. Moran SP, Dickerson JW, Cho HP, Xiang Z, Maksymetz J, Remke DH, et al. M 1-positive allosteric modulators lacking agonist activity provide the optimal profile for enhancing cognition. Neuropsychopharmacology. 2018;43(8): 1763–71. https://doi.org/10.1038/s41386-018-0033-9.

# **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.