# RESEARCH

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# Assessment of the ecological status of an ancient reservoir using macroinvertebrate assemblage and water quality parameters

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# Abstract

**Background** Atori Reservoir, located in the heart of southwestern Nigeria, is a tribute to the region's historical significance and natural splendor. Its establishment as a reservoir in 1935 marked a water resource management watershed in the region, transforming Atori into one of Nigeria's oldest and most important waterbodies. Despite its ancient age and indisputable importance, the exact ecological state of Atori Reservoir has been shrouded in mystery for decades. This study was necessitated by a paucity of information on the ecological status of Atori Reservoir.

**Results** The study identified 953 macroinvertebrates belonging to four classes, distributed across nine orders and nineteen families within Atori Reservoir. Among these, the class Insecta exhibited the highest diversity, while *Melanoides tuberculata*, a member of the Mollusca class, emerged as the dominant species. Despite the diverse macroinvertebrate community, the physicochemical parameters of the water raised concerns. Elevated values of total dissolved solids and conductivity indicated poor water quality, which was further reflected in the predominance of pollution-tolerant species and the absence of pollution-sensitive ones. Canonical correspondence analysis highlighted potential correlations between macroinvertebrates and water quality variables, yet statistical significance was lacking, as demonstrated by the Monte Carlo permutation test.

**Conclusion** This study sheds light on the ecological state of Atori Reservoir, revealing a diverse macroinvertebrate community but highlighting concerns regarding water quality. The study also emphasizes the pressing need for improved management practices to safeguard the ecological health of Atori Reservoir, given the critical role it plays in the region's ecosystem and local communities.

Keywords Ecological state, Macroinvertebrates, Diversity, Water quality, Pollution-tolerant, Atori Reservoir

# Background

Reservoirs are generally considered essential components of the freshwater ecosystem as they constitute a major habitat for critical aquatic resources (Schofield et al., 2018; Zwahlen, 2022). However, many reservoirs are being threatened by the growing human population, urbanization and other anthropogenically induced stressors. This has brought about a growing concern about the sustainability of the natural, biological and functional dynamics of freshwater ecosystems. The situation is especially critical in developing countries like Nigeria, where little or no control measures are in place to manage and regulate activities around important waterbodies (Garba et al., 2022). Natural habitat destruction and the introduction of pollutants into aquatic environments are common and widespread phenomena in Nigeria. This has not only affected the ecological balance of many important waterbodies but has led to a great loss of biodiversity within such habitats (Ghali et al., 2020). Furthermore,



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these activities have practically led to the loss of the capacity of such waterbodies to self-purify to a large extent. As such, water quality monitoring is essential for ensuring public health and maintaining the ecological balance of waterbodies.

One of the major causes of degradation in the aquatic environment is the intensification of agricultural activities (Umara et al., 2019). The constant growth of the human population has placed a huge burden on the necessity of increased production and improvements in the yield of agricultural products. This has informed the conventional use of synthetic fertilizers and chemicals for improved yield as well as for weed and pest control (Yung-Chul et al., 2016). However, these activities have serious ecological implications for surrounding waterbodies as the residues from the chemicals are dispersed indiscriminately into the aquatic environment. This catalyzes excessive nutrient enrichment and pollution in the waterbodies. Pollution in the aquatic environment causes degradation and fragmentation of local habitats, which ultimately alter the equilibrium and community structure of aquatic organisms in such waterbodies. Pollutants such as nitrogen and phosphorus from runoff may also cause increased biological productivity, resulting in the depletion of dissolved oxygen (DO) and eutrophication in reservoirs and other lentic waterbodies. The direct consequence of this is the reduction of habitat heterogeneity and the alteration of the ecological equilibrium of such waterbodies (Saleh et al., 2021).

In assessing the ecological status of waterbodies, one method that has become globally acceptable is bio-assessment using metrics of the community structure of aquatic organisms (Makumbe et al., 2022). The species richness and community assemblage of macroinvertebrates in a waterbody can provide useful and reliable information on the health of the waterbody. The species richness and community assemblage of macroinvertebrates reflect the number of species and individuals in a community and how evenly spread they are in such a community. Pollution leads to a decrease in the number of sensitive species, while hardy species thrive and outcompete less resilient ones. Assessment of water quality using physicochemical parameters of water is one of the traditional methods that have also been commonly used. The objectivity of this method is based on the fact that the physicochemical qualities of the substratum on which the macroinvertebrates are found determine their occurrence and distribution. Variation in the physicochemical parameters of water may lead to distortions in biological activities, the transfer of energy and the migration of aquatic insects (Amusan & Balogun, 2019). Furthermore, the nature of the distribution of physicochemical parameters of water provides firsthand information on the ecological processes and environmental variables that govern the populations and community structure of macroinvertebrates in a given waterbody (Kumar et al., 2022). Therefore, a combination of bio-assessment and the determination of the physicochemical properties of water will capture the whole spectrum of ecological stressors, thus providing a more reliable estimate of the health of waterbodies (Gouissi et al., 2019).

Most studies on water quality assessment in Nigeria have focused on either the use of physicochemical parameters or the macroinvertebrate community structure. However, this study aimed at assessing the health of Atori Reservoir using a combination of macroinvertebrate community structure and physicochemical parameters of the waterbody as indicators.

Atori Reservoir, one of the oldest in Nigeria, was impounded in 1935, yet no documented effort has been made to assess its health status since then. Located in Iseyin, southwestern Nigeria, the reservoir lies between Longitude 003036'E and Latitude 07058'N, at an altitude of 329 m amsl (Fig. 1). It boasts a capacity of about 580 million  $m^3$ , covering a surface area of 4.7 km<sup>2</sup>, with a mean depth of 5.2 m. The embankment stretches approximately 120 m in length and 40 m in width, featuring a mechanical spillway that automatically releases water when it exceeds the maximum operational capacity. Unique to Atori Reservoir is its water source, which primarily comes from seepage from the ground. The reservoir serves various purposes, including providing potable water and other ancillary functions to the people in Iseyin and neighboring communities. Iseyin, an ancient city in Oyo State, lies approximately 100 km north of Ibadan, encompassing a total land mass of 1,419 km<sup>2</sup> (Adewuyi et al., 2018). Despite its historical significance, the reservoir faces varying degrees of anthropogenic disturbances, mainly from runoff from agricultural lands, fishing activities and domestic waste. In addition to its physical attributes, Atori Reservoir's surrounding landscape is characterized by extensive agricultural lands, where food and cash crops such as maize, cassava, yam, citrus, cashew and cocoa are cultivated. The area also supports animal husbandry, with abundant grasslands to sustain livestock. Notable tree species like *Elaeis* guineensis (Oil palm), Cocos nucifera (Coconut), Adonsonia digitata (Baobab) and Albizia lebbeck (Ayunre) are found in the vicinity, contributing to the region's biodiversity. Overall, while Atori Reservoir plays a vital role in providing water resources to the local communities and supporting agricultural activities, it faces environmental challenges due to human activities (Oladimeji et al., 2024). Understanding its health status is crucial for effective management and conservation efforts in the region.



Fig. 1 Map of Atori Reservoir showing the sampling stations numbered St1-St5

Hence, this study aims to evaluate the health status of the waterbody by examining macroinvertebrate community structure and the physicochemical properties of water as indicators. The specific objectives include analyzing the composition, diversity and abundance of macroinvertebrate species, monitoring variations in physicochemical properties during the wet and dry seasons across various sampling locations and assessing potential relationships between physicochemical properties and macroinvertebrate composition in the reservoir.

## Methods

#### Sampling program and selection of sampling stations

The study area lies within the tropical monsoon climate zone, characterized by two distinct seasons: the wet (May–October) and the dry (November–April). As such, the sampling program was designed to cover both the wet and dry seasons of an annual cycle. Sampling was carried out between December 2019 and October 2020. The reservoir was visited once every two months, thus making a total of six sampling periods for the collection of macroinvertebrates and water samples. We selected five (5) sampling stations based on the reservoir reaches and to ensure that the numerous microhabitats within the reservoir are covered. Information on the grid location, elevation and mean depth for the sampling stations is presented in Table 1.

**Table 1**Summary of the sampled stations with their gridlocations, elevations and mean depth

S/No.	Latitude (N)	Longitude (E)	Elevation (m)	Mean depth (m)
1	0758.393	003 36.203	300	4.7
2	0758.378	003 36.171	298	4.2
3	0758.331	003 36.161	299	7.1
4	0758.373	003 36.228	298	5.3
5	0758.367	003 36.202	294	4.2

# Collection, sorting and identification of macroinvertebrates

Sampling of macroinvertebrates was carried out three consecutive times per station during every sampling period aboard a canoe using an improvised grab sampler of 0.04 m<sup>2</sup> (0.2 m  $\times$  0.2 m). The grabbed sediment samples were then gently sieved through a 0.5 mm mesh sieve using the reservoir water (Strayer, 2009). The macroinvertebrates retained in the sieve were picked up using a pair of long forceps. Apart from this method, macroinvertebrates were also collected thrice per sampling site using a long-handled D-frame net. This net was used to scoop the open water, submerged and emergent vegetation zones in the reservoir. The sorting of the collected specimens was done on a white enamel tray. Sorted specimens were preserved in specimen bottles containing 70% ethanol solution (APHA, 2005) and appropriately labeled. The specimens were identified using appropriate taxonomic keys and guides such as Brown (1980), Madsen (1985), Schneider (1990), Water Research Commission WRC (2003) and Verma (2006).

#### **Collection of water samples**

Two water samples per site were collected from clear water in sterilized 1-L plastic bottles for the analysis of the physicochemical properties of the water during every sampling period. Water temperature, dissolved oxygen (DO), pH, electrical conductivity (EC) and total dissolved solids (TDS) were determined on the field using a multi-3630 IDS digital meter, precision meters and a waterproof pH meter (version pH-689). However, the water samples were taken to the laboratory for the analysis of other parameters, including biological oxygen demand (BOD), acidity, total alkalinity, chloride, calcium, magnesium, nitrate and phosphate, according to APHA (2005).

#### Data analysis

All the data sets were subjected to a normality test. Based on the normality test, appropriate parametric and nonparametric statistical tests were used to analyze the data. Physicochemical parameters were analyzed with one-way analysis of variance (ANOVA), while a Kruskal–Wallis test was used to analyze the community structure of the macroinvertebrates. Margalef, Shannon–Wiener and Simpson's diversity index were used to measure diversity, and Pielou's evenness index was used to assess species evenness. Canonical correspondence analysis (CCA) was used to determine the relationship between the community structure of macroinvertebrates and physicochemical parameters. All statistical tests were carried out using Microsoft Excel and PAST (Hammer et al., 2001).

#### Results

## Physicochemical parameters of water in Atori Reservoir Spatial variations

There were variations in the values obtained for the physicochemical parameters of the water across the sampling stations, but the variations were not statistically significant (p > 0.05) as shown in Table 2. The pH and phosphate values were similar across the sampling stations. The highest values for EC, TDS and alkalinity were recorded in Station 1, while acidity, calcium and hardness were highest in Station 3. The highest BOD value was obtained in Station 2, while magnesium was highest in Station 4.

Table 2 Spatial variation in physicochemical parameters of water in Atori Reservoir, (December 2019–October 2020)

Parameters	$Mean \pm SD$					ANOVA	
	Station 1	Station 2	Station 3	Station 4	Station 5	F	Р
Temperature (°C)	26.37±1.10	26.87±1.02	27.02±1.19	27.30±1.44	27.50±0.73	0.9068	0.4751
Dissolved oxygen (Mg/L)	$7.37 \pm 0.890$	$7.25 \pm 1.024$	$7.28 \pm 0.929$	$7.27 \pm 0.768$	$7.31 \pm 0.752$	0.01687	0.9994
Biological demand oxygen (Mg/L)	$2.05 \pm 1.19$	$2.60 \pm 1.14$	$2.17 \pm 0.65$	$2.01 \pm 1.38$	$2.47 \pm 1.27$	0.3077	0.8701
Conductivity (µs/cm)	$650 \pm 44.2$	$589 \pm 51.0$	$611 \pm 77.5$	$590 \pm 79.6$	616±46.7	0.9732	0.4397
Total dissolved solids (Mg/L)	$462 \pm 22$	$408 \pm 45$	$435 \pm 41$	$415 \pm 56$	437±33	1.574	0.2122
рН	$7.5 \pm 0.20$	$7.5 \pm 0.13$	$7.5 \pm 0.16$	$7.5 \pm 0.20$	$7.5 \pm 0.24$	0.1588	0.9572
Acidity	7.3±13.2	$6.0 \pm 14.7$	$10.2 \pm 12.3$	4.3±8.7	$7.3 \pm 11.4$	0.1843	0.9444
Alkalinity	$275 \pm 24.6$	$248 \pm 36.5$	$271 \pm 23.8$	$256 \pm 32.1$	271±19.6	0.9973	0.4274
Magnesium	$5.20 \pm 6.65$	$9.62 \pm 12.86$	$7.17 \pm 4.41$	$10.03 \pm 13.6$	$7.19 \pm 11.50$	0.2171	0.9264
Calcium	$58.00 \pm 39.12$	$48.04 \pm 32.3$	$62.34 \pm 38.28$	$50.25 \pm 33.7$	$58.49 \pm 37.73$	0.1651	0.9541
Water hardness	$166.3 \pm 115.6$	$159.6 \pm 108.2$	$185.1 \pm 103.4$	166.7±117.8	175.7±120.4	0.04596	0.9957
Phosphate	$0.32 \pm 0.004$	$0.32 \pm 0.004$	$0.32 \pm 0.004$	$0.32 \pm 0.005$	$0.32 \pm 0.005$	0.1445	0.9637
Nitrate	$6.00 \pm 4.33$	$7.22 \pm 8.52$	$5.75 \pm 6.30$	$6.52 \pm 5.68$	$8.82 \pm 8.90$	0.1871	0.9429
Chloride	$39.5 \pm 10.6$	$43.4 \pm 9.2$	$43.0 \pm 10.5$	$39.9 \pm 10.4$	$50.0 \pm 19.1$	0.6766	0.6145

A good number of the investigated parameters, such as DO, EC, TDS, alkalinity, calcium and hardness, had their minimum values in Station 2 (Table 2).

#### Seasonal variations

The mean values obtained for DO, TDS, pH, magnesium, calcium, water hardness, phosphate, nitrate and chloride were higher in the wet season, while water temperature, BOD, EC, acidity and alkalinity had higher values in the dry season (Table 3). Only six (6) parameters in the seasonal variations of physicochemical characteristics of water in Atori Reservoir were significantly different (*T*-test, p < 0.05, p < 0.01, p < 0.001), according to a paired-two sample *T*-test (p < 0.05, p < 0.01, p < 0.01, p < 0.001). The physicochemical parameters that were significantly different include DO concentration, acidity, alkalinity, calcium, water hardness and phosphate (Table 3).

#### Taxonomic composition and distribution of macroinvertebrates

A total of 953 macroinvertebrate individuals (ind.) distributed in three phyla, four classes, nine orders, 19 families and 32 genera were reported in this study (Table 4). Four families comprised the non-insect group (Hirudidae, Phanorbidae, Thiaridae and Pisauridae), while the remaining fifteen (15) were insects, and they were the dominant macroinvertebrates group collected. The identified insect orders include Hemiptera, Coleoptera, Diptera, Ephemeroptera and Odonata. The dominant species recorded was *Melanoides tuberculata* (Muller, 1774), with 321 macroinvertebrate individuals (33.68%). Other species that were recorded in relatively high numbers include *Notonecta* sp. (Insecta) and *Biomphalaria* sp. (Gastropoda), with 191 (20%) and 138 (14%), respectively. More macroinvertebrates were recorded in the dry period (569) (59.7%) as against 384 (40.3%) macroinvertebrate individuals recorded in the wet (Table 4). The stations did not differ significantly with respect to the abundance of macroinvertebrates. However, the highest number of macroinvertebrates was recorded at Station 5, while the least was recorded at Station 2. In terms of taxa richness, Station 2 had the lowest, while Station 1 had the highest (Tables 5 and 6). Margalef and Shannon–Wiener (H) indices showed that Station 1 had the highest diversity of macroinvertebrates, while Station 2 had the least diversity (Table 5). Pielou's evenness index was highest in Station 3, while the least was recorded in Station 2.

# Relationship between the macroinvertebrates and physicochemical parameters

Canonical correspondence analysis (CCA) of the relationship between macroinvertebrates and water physicochemical parameters revealed that macroinvertebrate abundance in stations 2 and 4 was associated with increasing temperatures, magnesium and pH, whereas abundance in the other stations was associated with increasing values for all other parameters (Fig. 2). Furthermore, the distribution of macroinvertebrates in the triplot of the first and second CCA axes showed that macroinvertebrates (*Hydrophilus* sp., *Bulinus* sp. and *Limnogonus* sp.) that were sensitive to optimum concentrations of physicochemical variables of water were also observed to be associated with Stations 3 and 5 on the triplots (Fig. 2). Aside from this,

Table 3 Seasonal water variations at Atori Reservoir, Iseyin, Nigeria (December 2019–October 2020)

Parameters	MEAN±SD		t-TEST	
	DRY	WET	Т	Р
Temperature (°C)	27.14±1.27	26.88±0.95	0.63308	0.53182
Dissolved oxygen (Mg/L)	$6.92 \pm 0.92$	$7.67 \pm 0.49$	- 2.7945	0.0092765**
Biological demand oxygen (Mg/L)	$2.27 \pm 1.42$	$2.25 \pm 0.69$	0.053921	0.95738
Conductivity (µs/cm)	616.6±66.06	$605.33 \pm 58.66$	0.49391	0.62522
Total dissolved solids (Mg/L)	424.73±33.65	$437.47 \pm 50.32$	-0.81465	0.42215
Ph	$7.44 \pm 0.19$	$7.56 \pm 0.15$	- 2.0391	0.05979
Acidity	$11.8 \pm 14.90$	$2.27 \pm 1.98$	2.4563	0.0205*
Alkalinity	276.5±31.52	251.8±17.41	2.6603	0.012774*
Magnesium	$6.60 \pm 11.40$	9.08±8.27	-0.68438	0.49936
Calcium	$32.77 \pm 34.04$	$78.07 \pm 12.68$	-4.8299	4.4159*10 <sup>-5***</sup>
Water hardness	108.9±118.2	$232.5 \pm 30.5$	- 3.9191	0.00052184***
Phosphate	$0.31 \pm 0.004$	$0.32 \pm 0.001$	-6.204	1.0588*10 <sup>-6***</sup>
Nitrate	$4.72 \pm 2.39$	$9.00 \pm 8.59$	- 1.8576	0.073772
Chloride	40.66±39.76	45.66±46.81	- 1.1246	0.27033

Implies p < 0.05\*, p < 0.01\*\*, p < 0.001\*\*\*

Table 4	Taxonomic composition of	aquatic macroinverter	orates in Atori Re	servoir, Iseyin, S	Southwestern N	ligeria (Decerr	nber 2019–
October	2020)						

Dry     Wet     Tot       Annelida     Clitellata     Arhynchobdellida     Hirudidae     Hirudio sp.     0     1     1       Mollusca     Gastropoda     Basommatophora     Planorbidae     Biomphalaria sp.     100     38     138       Bulinus sp.     197     124     323       Neotaenioglossa     Thiaridae     Melanoides tuberculara     197     124     323       Arthropoda     Arachnida     Araneae     Pisauridae     Dolomedes sp.     1     0     1       Insecta     Hemiptera     Belostomatidae     Apasus sp.     17     0     7       Neogerris sp.     14     10     24     24       Rhagadotarus sp.     1     7     8       Torndogonus sp.     1     1     2       Naucoridae     Maarocoris sp.     1     1     2       Naucoridae     Anisops sp.     0     2     2       Naucoridae     Anisops sp.     2     0     2       Naucoridae     Anisops sp.     2     0     2       Naucoridae     Anisops sp.     2     0     2       Coleoptera     Chrysomelidae     Anisops sp.     3     0     3       Coleoptera     Chrysomelii	Phylum	Class	Order	Family	Genus/Species	Season		
Annelida Clitellata Arthynchobdellida Hirudidae Hirudo sp. 0 1 1 1 Mollusca Gastropoda Basommatophora Planorbidae Biomphalaria sp. 100 38 138 Bullinus sp. 24 9 33 Neotaenioglossa Thiairidae Melanoides tuberculata 197 124 321 Arthropoda Arachnida Araneae Pisauridae Dolomedes sp. 1 0 1 1 Insecta Hemiptera Belostomatidae Appaus sp. 41 9 50 Gerridae Limnogonus sp. 3 0 3 Limnogonus sp. 17 0 177 Neogeris sp. 14 10 24 Rhagadotarus sp. 1 7 0 3 Naucoridae Macrocoris sp. 1 1 1 2 Naucoris sp. 0 3 3 Naucoridae Macrocoris sp. 1 1 1 2 Neomacrocoris sp. 0 2 2 Entrihares sp. 4 0 4 Notonectidae Apiss sp. 4 0 4 Notonectidae Agaiseles p. 3 0 3 Naucoridae Macrocoris sp. 0 1 1 1 Neejidae Ranatra sp. 2 0 2 Entrihares sp. 4 0 4 Notonectidae Agaiseles p. 3 0 3 Dytiscicae Chrysomelidae Agaiseles p. 3 0 3 Dytiscicae Agaiseles p. 3 0 3 Dytiscicae Arisops p. 8 13 21 Hydrophilidae Arantra sp. 2 0 2 Ephemeroptera Chrysomelidae Agaiseles p. 8 13 21 Hydrophilidae Arantra sp. 2 0 2 Ephemeroptera Baetidae Arantra sp. 2 0 2 Ephemeroptera Baetidae Agaiseles p. 8 13 21 Hydrophilidae Antaro sp. 2 0 2 Ephemeroptera Baetidae Aranonus sp. 2 0 2 Ephemeroptera Baetidae Antaro sp. 2 0 2 Ephemeroptera Aestinide Antaro sp. 2 0 2 Ephemeroptera Baetidae Antaro sp. 2 0 2 Ephemoroptera Agaitae Antaro sp. 2 0 2 Ephemoroptera Aestinide Antaro sp. 2 0 2 Ephemoroptera Aestinide Antaro sp. 2 0 2 Ephemoroptera Aestinide Antaro sp. 2 0 2 Baetidae Atoromotoporo sp. 3 3 0 5 Colonata: Zygoptera Coenagrionidae Agrocore sp. 2 0 2 Preudos sp. 8 16 24						Dry	Wet	Total
Mollusca     Gastropoda     Basommatophora     Planorbidae     Biomphalaria sp.     100     38     138       Neotaenioglossa     Thairidae     Melanoides tuberculata     17     12     32       Arthropoda     Arachnida     Araneae     Plaauridae     Dolomedes sp.     10     1     30     30       Insecta     Hemiptera     Belostomatidae     Apasus sp.     41     0     3       Medanoides tuberculata     Gerridae     Limnogorus sp.     3     0     3       Medanoides tuberculata     Sp.     10     0     2       Medanoides tuberculata     Araneae     Plaauridae     Apasus sp.     10     0     3       Marchnida     Araneae     Gerridae     Apasus sp.     10     0     2       Macoridae     Macorocoris sp.     1     0     2     2       Naucoridae     Ranatra sp.     2     0     2     2       Notonectidae     Anisops sp.     3     0     3     3       Macorolae     Chrysomelidae     Anisops sp.     3     0     3       Notonectidae     Chrysomelidae     Apatines sp.     3     3     3       Macorolae     Apasophilus sp.     8     1     9 <tr< td=""><td>Annelida</td><td>Clitellata</td><td>Arhynchobdellida</td><td>Hirudidae</td><td>Hirudo sp.</td><td>0</td><td>1</td><td>1</td></tr<>	Annelida	Clitellata	Arhynchobdellida	Hirudidae	Hirudo sp.	0	1	1
Arthropoda       Arachnida       Neotaenioglossa       Thiaridae       Melonoides tuberculata       197       124       321         Arthropoda       Arachnida       Araneae       Pisauridae       Dolomedes sp.       1       9       50         Insecta       Hemiptera       Belostomatidae       Appasus sp.       17       0       1       10       24         Brandgotarus sp.       17       0       1       10       24         Madadotarus sp.       16       0       1       12       2         Macorids sp.       1       1       2       2         Naucoridae       Marcoris sp.       1       1       2       2         Naucorids sp.       0       1       1       2 <td>Mollusca</td> <td>Gastropoda</td> <td>Basommatophora</td> <td>Planorbidae</td> <td>Biomphalaria sp.</td> <td>100</td> <td>38</td> <td>138</td>	Mollusca	Gastropoda	Basommatophora	Planorbidae	Biomphalaria sp.	100	38	138
Neotaenioglossa         Thiaridae         Melanoides tuberculata         197         124         321           Arthropoda         Arachnida         Araneae         Pisauridae         Dolomedes sp.         1         0         1           Insecta         Hemiptera         Belostomatidae         Appauss sp.         17         0         3           Gerridae         Limnoporus sp.         17         0         1         1         2           Rhagadorus sp.         14         0         2         3         3         3         3         3           Natocridae         Reagerins sp.         1         1         2         3         3         3         3         3         3           Natocridae         Rantaros sp.         0         1         1         2					Bulinus sp.	24	9	33
Arthropoda       Arachnida       Araeae       Pisauridae       Belostomatidae       Appasus sp.       41       9       50         Insecta       Hemiptera       Belostomatidae       Appasus sp.       3       0       3         Innogorus sp.       10       0       17       No       17         Neogerris sp.       14       10       2       16       3         Rhagadotarus sp.       14       10       2       3       3         Naucoridae       Rhagadotarus sp.       10       3       3         Naucoridas pp       0       1       1       2         Naucoridae       Ranatra sp.       1       1       2         Notonectidae       Ranatra sp.       2       2       2         Notonectidae       Anisops pp.       3       1       3         Notonectidae       Anisops pp.       2       2       2       2         Notonectidae       Anisops pp.       3       1       3       3         Optiscidae       Laccophilus sp.       3       3       3       3       3         Notonectidae       Anaspi sp.       3       3       3       3       3 <td< td=""><td></td><td></td><td>Neotaenioglossa</td><td>Thiaridae</td><td>Melanoides tuberculata</td><td>197</td><td>124</td><td>321</td></td<>			Neotaenioglossa	Thiaridae	Melanoides tuberculata	197	124	321
InsectaHemipteraBelostomatidaeAppasus sp.41950GerridaeLinnogonus sp.303Linnoporus sp.17017Neogenris sp.1778Rhagadotarus sp.1778Renagogonus sp.178NaucoridaeMacrocaris sp.112Neogenris sp.1122Neoraris sp.1122Neucoris sp.0112NotonectridaeRanatra sp.222NotonectridaeAnisops sp.222NotonectridaeAnisops sp.311ColeopteraChrysomelidaeAgasicles sp.311Odonata: AnisopteraChrionomidaeCorectogrus sp.101HydrophilidaeDipteraChrionomidaeChroinomis sp.202EphemeropteraBaetidaeAnax sp.2022Odonata: AnisopteraAeshnidaeAnax sp.202Odonata: AnisopteraCoreagrioninge,3111Diplacodes sp.31112Diplacodes sp.31212Peudagrion sp.20222Peudagrion sp.2022Peudagrion sp.3112Diplacodes	Arthropoda	Arachnida	Araneae	Pisauridae	Dolomedes sp.	1	0	1
GerridaeLimnogonus sp.303Limnoporus sp.17017Neogeris sp.141024Rhagadotarus sp.033Panagoonus sp.033NaucoridaeMacrocris sp.112NaucoridaeMacrocris sp.011NepidaeRanatra sp.222NotonectidaeAnisops sp.222Notonecta sp.4043QuiscidasAnisops sp.3033QuiscidaeAnisops sp.3043Notonecta sp.61151133QuiscidaeAnisops sp.3033QuiscidaeCacephilus sp.3011HydrophilidaeAmphilops sp.81321HydrophilidaeAnax sp.2022EphemeropteraBaetidaePotmocleoor sp.22Qonata: AnisopteraAeshnidaeAnax sp.334LibellulidaeAnizoptera3111Diplacodes sp.30112Qonata: ZigopteraCeenagrinoidaeAcisoma sp.363QuisciaeCeenagrinoidaeAcisoma sp.3633QuisciaeCeenagrinoidaeAcisoma sp.3633Quisciae<		Insecta	Hemiptera	Belostomatidae	Appasus sp.	41	9	50
Limnoporus sp.       17       0       17         Neogerris sp.       14       10       24         Rhagadotarus sp.       1       7       8         Inagogonus sp.       0       3       3         Naucoridae       Macrocoris sp.       0       2       2         Naucoridae       Macrocoris sp.       0       1       1         Nepidae       Ranatra sp.       2       0       2       2         Notonectidae       Anisops sp.       0       2				Gerridae	Limnogonus sp.	3	0	3
Neigents       14       10       24         Rhagadotarus       1       7       8         Tenagogonus       0       3       3         Naucoridae       Marcocris       1       1       2         Naucoris       Naucoris       1       1       2         Naucoris       Naucoris       1       1       1       2         Naucoris       Ranatra       0       1       1       1       1         Nepidae       Ranatra       2       0       2					Limnoporus sp.	17	0	17
Rhagadotarus sp.         1         7         8           Tenagogonus sp.         0         3         3           Naucoridae         Macrocoris sp.         1         1         2           Naucorids sp.         0         2         2         2           Naucorids sp.         0         1         1         1           Nepidae         Ranatra sp.         2         0         2           Notonectidae         Anisops sp.         20         2         2           Notonecta sp.         76         115         19           Optiscidae         Agasicles sp.         3         3         3           Dytiscidae         Chironomidae         Chironomus sp.         4         0         4           Hydrophilus sp.         3         1         1         1         1           Hydrophilus sp.         8         1         2         2         2         2           Diptera         Chironomidae         Chironomus sp.         2         0         2         2         2         2         2         2         2         2         2         2         2         2         2         2         2         2 <td></td> <td></td> <td></td> <td></td> <td>Neogerris sp.</td> <td>14</td> <td>10</td> <td>24</td>					Neogerris sp.	14	10	24
Image is a set of the set of					Rhagadotarus sp.	1	7	8
Naucoridae         Macrocoris sp.         1         1         2           Naucoris sp.         0         2         2           Nepidae         Ranatra sp.         0         1         1           Notonectidae         Ranatra sp.         20         2         22           Inithares sp.         4         0         4         2         22           Inithares sp.         6         115         19           Notonectidae         Anisops sp.         6         115         19           Ocleoptera         Chrysomelidae         Agasicles sp.         3         0         4           Qytiscidae         Laccophilus sp.         1         0         1         1           Hydrophilidae         Amphiops sp.         8         13         21           Hydrophilidae         Potamocloeon sp.         2         0         2           Odonata: Anisoptera         Aeshnidae         Anax sp.         0         1         1           Ibiellulidae         Acisoma sp.         3         3         6         2           Odonata: Zygoptera         Cenagrionidae         Agriocnemis sp.         3         6         2           Ibiellulidae					Tenagogonus sp.	0	3	3
Naucoris sp.       0       2       2         Nepidae       Ranatra sp.       0       1       1         Notonectidae       Anisops sp.       20       2       2         Interes sp.       4       0       4       2       2         Notonectidae       Anisops sp.       20       2       2       2         Interes sp.       4       0       4				Naucoridae	Macrocoris sp.	1	1	2
Nepidae       Nepidae       Ranatra sp.       0       1       1         Notonectidae       Ranatra sp.       20       2       22         Initianes sp.       4       0       2       2         Notonectidae       Anisops sp.       20       2       2         Initianes sp.       4       0       4       2         Notonecta sp.       3       115       19         Optiscidae       Laccophilus sp.       4       0       4         Optiscidae       Cretogyrus sp.       1       0       1         Hydrophilidae       Orectogyrus sp.       8       13       21         Hydrophilus sp.       8       13       21         Iptera       Chironomidae       Netonocloon sp.       8       13       21         Iptera       Chironomidae       Potamocloon sp.       2       0       2         Ipleneroptera       Baetidae       Anax sp.       2       0       2         Iplecodes sp.       2       0       1       1         Iplecodes sp.       1       1       1       1         Iplecodes sp.       5       1       1       1 <t< td=""><td></td><td></td><td></td><td></td><td>Naucoris sp</td><td>0</td><td>2</td><td>2</td></t<>					Naucoris sp	0	2	2
Nepidae       Ranatra sp.       2       0       2         Notonectidae       Anisops sp.       20       2       22         Enithares sp.       4       0       4         Notonectidae       Agasicles sp.       3       115       191         Notonecta sp.       3       0       4       4       4         Ocleoptera       Chrysomelidae       Agasicles sp.       3       0       4         Oytiscidae       Laccophilus sp.       4       0       4       4         Oytiscidae       Orectogyrus sp.       1       0       1       4					Neomacrocoris sp.	0	1	1
Notonectidae         Anisops sp.         20         2         22           Enithares sp.         4         0         4           Notonecta sp.         76         115         191           Coleoptera         Chrysomelidae         Agasicles sp.         3         0         4           Dytiscidae         Laccophilus sp.         4         0         4         1           Gyrinidae         Orectogyrus sp.         1         0         1         1           Hydrophilidae         Amphiops sp.         8         13         21           Hydrophilidae         Amphiops sp.         8         13         21           Ephemeroptera         Baetidae         Notanecloeon sp.         2         0         2           Odonata: Anisoptera         Aeshnidae         Anax sp.         0         1         1           Diplacodes sp.         8         16         24         24         24           Odonata: Anisoptera         Coenagrionidae         Anix sp.         0         1         1           Diplacodes sp.         8         16         24         24         24         24         24         24         24         24         24         <				Nepidae	<i>Ranatra</i> sp.	2	0	2
Enithares sp.         4         0         4           Notonecta sp.         76         115         191           Coleoptera         Chrysomelidae         Agasicles sp.         3         0         4           Dytiscidae         Laccophilus sp.         4         0         4         1           Gyrinidae         Orectogyrus sp.         1         0         1         1           Hydrophilidae         Amphiops sp.         8         13         21           Hydrophilidae         Chironomus sp.         8         13         21           Diptera         Chironomidae         Chironomus sp.         2         0         2           Codonata: Anisoptera         Aeshnidae         Anax sp.         2         0         2           Odonata: Anisoptera         Aeshnidae         Acisoma sp.         1         1         1           Diplacodes sp.         3         6         3         6         3         6           Odonata: Zygoptera         Coenagrionidae         Agriconemis sp.         3         6         2         2         2         2         2         2         2         2         2         2         2         2         3				Notonectidae	Anisops sp.	20	2	22
Coleoptera       Chrysomelidae       Agasicles sp.       3       0       3         Dytiscidae       Laccophilus sp.       4       0       4         Gyrinidae       Orectogyrus sp.       1       0       1         Hydrophilidae       Amphiops sp.       8       13       21         Hydrophilidae       Amphiops sp.       8       13       21         Diptera       Chironomidae       Chironomus sp.       2       0       2         Codonata: Anisoptera       Baetidae       Potamocloeon sp.       2       0       2         Odonata: Anisoptera       Coenagrionidae       Anax sp.       0       1       1         Diplacodes sp.       3       3       2 <t< td=""><td></td><td></td><td></td><td></td><td>Enithares sp.</td><td>4</td><td>0</td><td>4</td></t<>					Enithares sp.	4	0	4
Coleoptera       Chrysomelidae       Agasicles sp.       3       0       3         Dytiscidae       Laccophilus sp.       4       0       4         Gyrinidae       Orectogyrus sp.       1       0       1         Hydrophilidae       Amphiops sp.       8       13       21         Hydrophilus sp.       8       13       21         Hydrophilus sp.       8       13       21         Diptera       Chironomidae       Amonologoon sp.       2       0       2         Odonata: Anisoptera       Aeshnidae       Anax sp.       2       0       2         Ibielluidae       Araix sp.       8       16       2         Odonata: Zygoptera       Coenagrionidae       Agriocnemis sp.       3       3       4         Odonata: Zygoptera       Coenagrionidae       Agriocnemis sp.       5       0       5         Ceriagrion sp.       2       0       2       2       2       2         Magrion sp.       2       0       2       3       4       3         Diplacoders sp.       5       0       2       2       2       2       2       2       2       2       2       <					Notonecta sp.	76	115	191
Dytiscidae         Laccophilus sp.         4         0         4           Gyrinidae         Orectogyrus sp.         1         0         1           Hydrophilidae         Amphiops sp.         8         13         21           Hydrophilidae         Maphiops sp.         8         13         21           Hydrophilus sp.         8         10         9           Diptera         Chironomidae         Chironomus sp.         2         0         2           Ephemeroptera         Baetidae         Potamocloeon sp.         2         0         2           Odonata: Anisoptera         Aeshnidae         Anax sp.         2         0         2           Libellulidae         Acisoma sp.         0         1         1           Diplacodes sp.         8         3         6           Odonata: Zygoptera         Coenagrionidae         Agriocnemis sp.         3         3         6           Criagrion sp.         2         0         2         2         2         2         2         2         2         2         2         2         2         2         2         2         3         4         3         3         4         3			Coleoptera	Chrysomelidae	Agasicles sp.	3	0	3
Gyrinidae       Orectogyrus sp.       1       0       1         Hydrophilidae       Amphiops sp.       8       13       21         Hydrophilus sp.       8       1       9         Diptera       Chironomidae       Chironomus sp.       2       0       2         Ephemeroptera       Baetidae       Potamocloeon sp.       2       0       2         Odonata: Anisoptera       Aeshnidae       Anax sp.       0       1       1         Libellulidae       Arisoma sp.       0       1       1       2         Odonata: Anisoptera       Cenagrionidae       Acisoma sp.       0       1       1         Diplacodes sp.       8       16       24<				Dytiscidae	Laccophilus sp.	4	0	4
Hydrophilidae       Amphiops sp.       8       13       21         Hydrophilus sp.       8       10       9         Diptera       Chironomidae       Chironomus sp.       2       0       2         Ephemeroptera       Baetidae       Potamocloeon sp.       2       0       2         Odonata: Anisoptera       Aeshnidae       Anax sp.       2       0       2         Libellulidae       Acisoma sp.       0       1       1         Diplacodes sp.       8       16       24         Odonata: Zygoptera       Coenagrionidae       Agriocnemis sp.       3       6         Greiagrion sp.       2       0       2       2         Paeudagrion sp.       2       0       2       2         Coenagrionidae       Agriocnemis sp.       3       0       2         Paeudagrion sp.       2       0       2       2         Paeudagrion sp.       20       2       2       2         Paeudagrion sp.       20       28       3       3         Paeudagrion sp.       569       384       95				Gyrinidae	Orectogyrus sp.	1	0	1
Hydrophilus sp.819DipteraChironomidaeChironomus sp.202EphemeropteraBaetidaePotamocloeon sp.202Odonata: AnisopteraAeshnidaeAnax sp.202LibellulidaeAcisoma sp.011Diplacodes sp.81624Odonata: ZygopteraCoenagrionidaeAgriocnemis sp.336Ceriagrion sp.20222Pseudagrion sp.202848Total56938495333				Hydrophilidae	Amphiops sp.	8	13	21
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EphemeropteraBaetidaePotamocloeon sp.202Odonata: AnisopteraAeshnidaeAnax sp.202LibellulidaeAcisoma sp.011Diplacodes sp.81624Tetrathemis sp.336Odonata: ZygopteraCoenagrionidaeAgriocnemis sp.502Pseudagrion sp.202848Total569384953			Diptera	Chironomidae	Chironomus sp.	2	0	2
Odonata: AnisopteraAeshnidaeAnax sp.202LibellulidaeAcisoma sp.011Diplacodes sp.81624Tetrathemis sp.336Odonata: ZygopteraCoenagrionidaeAgriocnemis sp.505Ceriagrion sp.202848Total569384953			Ephemeroptera	Baetidae	Potamocloeon sp.	2	0	2
Libellulidae       Acisoma sp.       0       1       1         Diplacodes sp.       8       16       24         Tetrathemis sp.       3       6         Odonata: Zygoptera       Coenagrionidae       Agriocnemis sp.       5       0       5         Ceriagrion sp.       20       28       48         Total       569       384       953			Odonata: Anisoptera	Aeshnidae	Anax sp.	2	0	2
Diplacodes sp.       8       16       24         Tetrathemis sp.       3       6         Odonata: Zygoptera       Coenagrionidae       Agriocnemis sp.       5       0       5         Ceriagrion sp.       20       28       48         Total       569       384       953				Libellulidae	Acisoma sp.	0	1	1
Tetrathemis sp.       3       3       6         Odonata: Zygoptera       Coenagrionidae       Agriocnemis sp.       5       0       5         Ceriagrion sp.       2       0       2         Pseudagrion sp.       20       28       48         Total       569       384       953					Diplacodes sp.	8	16	24
Odonata: Zygoptera       Coenagrionidae       Agriocnemis sp.       5       0       5         Ceriagrion sp.       2       0       2         Pseudagrion sp.       20       28       48         Total       569       384       953					Tetrathemis sp.	3	3	6
Ceriagrion sp.       2       0       2         Pseudagrion sp.       20       28       48         Total       569       384       953			Odonata: Zygoptera	Coenagrionidae	Agriocnemis sp.	5	0	5
Pseudagrion sp.         20         28         48           Total         569         384         953					Ceriagrion sp.	2	0	2
<b>Total</b> 569 384 955					Pseudagrion sp.	20	28	48
					Total	569	384	953

Table 5         Diversity of macroinvertebrates among sampled
stations in Atori Reservoir, Iseyin, southwestern Nigeria
(December 2019–October 2020)

Metric	ST. 1	ST. 2	ST. 3	ST. 4	ST. 5
Taxa richness	23	15	17	18	18
Abundance	199	156	210	167	221
Margalef diversity index	4.16	2.77	2.99	3.32	3.15
Shannon–Wiener diversity index (H)	2.22	1.77	2.05	2.03	2.09
Simpson's diversity index (1-D)	0.82	0.76	0.82	0.79	0.82
Pielou's evenness index	0.40	0.39	0.46	0.42	0.45
ST STATION					

Agriocnemis sp., Macrocoris sp., Rhagadotarus sp. and Anisops were associated with increases in magnesium and temperature in Station 2. However, Anax sp., Enithares sp. and Potamocloeon sp. were associated with Station 4 where pH was the only increased variable while Pseudagrion sp., Naucoris sp. and Agasicles sp associated with high levels of BOD.

The percentage of variance that each axis of the CCA explains is that axes 1 and 2 accounted for 47.38% and 24.18%, respectively (Table 7).

Station	Shannon-	Shannon–Wiener		t-TEST			t-TEST	
	DRY	WET	т	Р	DRY	WET	T	Р
1	2.071	2.025	0.302	0.7631	0.795	0.823	0.743	0.458
2	1.647	1.700	-0.335	0.738	0.737	0.770	0.803	0.423
3	2.056	1.680	2.771	0.001*	0.829	0.737	-2.402	0.0175*
4	1.908	1.698	1.315	0.190	0.775	0.748	- 0.595	0.5528
5	2.118	1.806	2.302	0.0225*	0.837	0.767	-1.814	0.0719

 Table 6
 Comparison of two diversity indices during the dry and wet seasons



**Fig. 2** Graph CCA representation of macroinvertebrates, water variables and sampling stations in Atori Reservoir, Nigeria (December 2019– October 2020). Keys: Tem = temperature, DO = dissolved oxygen, BOD = biological oxygen demand, EC = electrical conductivity, TDS = total dissolved solids, Aci = acidity, Alk = alkalinity, Mag = magnesium, Cal = calcium, WaH = water hardness, Pho = phosphate, Nit = nitrate, Chl = chloride, Bio = Biomphalaria, Bul = Bulinus, Mel = Melanoides tuberculata, App = Appasus, Limg = Limnogonus, Limp = Limnoporus, Neo = Neogerris, Rha = Rhagadotarus, Mac = Macrocoris, Nau = Naucoris, Ani = Anisops, Eni = Enithares, Not = Notonecta, Aga = Agasicles, Amp = Amphiops, Hyd = Hydrophilus, Chil = Chironomus, Pot = Potamocloeon, Anax = Anax, Dip = Diplacodes, Tetr = Tetrathemis, Agr = Agriocnemis, Pse = Pseudagrion

**Table 7** Eigenvalues and percentage of total inertia of the first four axes of the canonical correspondence analysis of physicochemical parameters of water and macroinvertebrates in Atori Reservoir, Iseyin, Southwestern Nigeria

Axis	Eigenvalue	% of total inertia
1	0.10115	47.39
2	0.051616	24.17
3	0.044694	20.94
4	0.015693	7.351

# Discussion

## Physicochemical parameters of water

Atori Reservoir, a vital water source in Nigeria, serves multiple purposes including providing potable water, irrigation and domestic uses. However, its health is subject to scrutiny because the reservoir receives farm runoff from agricultural lands around the reservoir catchment basin while fishing, bathing and washing of agricultural equipment were also evident. These activities can potentially cause a deterioration of the water quality in the reservoir. As such, the high values reported for EC, TDS, turbidity, water hardness and chloride are markers of human-induced stress on the reservoir (Akamagwuna et al., 2019).

Although the measured water quality variables of Atori Reservoir were generally within the optimum range for the survival and growth of aquatic life in the tropics (Godswill, 2012; Mugo, 2010), the parameters were within the permissible limits of the National Environmental Standards and Regulatory Enforcement Agency (NESREA, 2007), the United States Environmental Protection Agency (USEPA, 2005) and the World Health Organization (WHO, 2004) standard for standing waters.

Spatially, variations in physicochemical parameters across sampled stations were observed but not statistically significant (p > 0.05), indicating microhabitat homogeneity. However, higher values in Station 2 may signal increased stress compared to others due to its closer proximity to agricultural farm lands. These findings align with studies on some selected similar waterbodies in Nigeria, such as Owalla Reservoir (Aduwo et al., 2019), Opa Reservoir (Aliu et al., 2020) and Aiba Reservoir (Atobatele & Ugwumba, 2008), in which there were no significant variations in the spatial distribution of the physicochemical parameters of the waterbodies.

Seasonal variations in physicochemical properties in Atori Reservoir were more pronounced and showed statistical significance for some properties such as DO, acidity, alkalinity, water hardness and nutrient levels. During the wet season, heightened precipitation and runoff introduced into the reservoir probably caused an increase in pH nutrient content, DO and water hardness (Zhang et al., 2020). Elevated temperatures enhance metabolic rates, influencing aquatic organism growth and reproduction. Conversely, the dry season brings increased temperatures, reduced water levels and flow rates, lowering dissolved oxygen.

Dissolved oxygen concentration is an important indicator of the ecological condition of aquatic ecosystems as it reflects the physical and biological processes taking place in such an environment (Kumar et al., 2011a; Ridanovic et al., 2010). The range of values obtained for the dissolved oxygen concentration in the reservoir was considered satisfactory. The relatively high values of DO could be attributed to the aeration rate and photosynthetic activities (Jaji et al., 2007). The distribution of DO in waterbodies has been known to be majorly influenced by a balance between some factors such as precipitation, photosynthesis activities, inputs from the atmosphere, losses by chemicals and biotic oxidations (Muralidharan et al., 2010; Nkwoji et al., 2010). The relatively higher concentration of DO in the wet season is not surprising, as it aligns with previous reports such as Amusan et al. (2018), Bhawsar and Vyas (2022), and Yusoff et al. (2002), in which higher concentrations of DO were reported. The observed higher concentration of DO in the wet season could be as a result of lower water temperature and increased algal productivity that produces oxygen in the season (Venkatesharaju et al., 2010).

The pH is an important parameter that not only measures the acid–base balance of water but also determines its suitability for various purposes (Sharma et al., 2011). The relatively high pH values recorded for the water indicated that the reservoir was slightly alkaline. The relatively high pH may be attributed to intense photosynthetic activities of aquatic plants, which reduce the free carbon dioxide concentrations in water (Gouissi et al., 2019). The reduction of free carbon dioxide concentration has been identified as one of the key indicators of higher pH in water bodies. Atori Reservoir had the majority of its surface area covered with emergent and submerged vegetation, as such a large volume of carbon dioxide is used up by the plants. Similar reports of slightly high pH values have been reported for Ikere Gorge Reservoir (Aiwerioghene & Ayoade, 2016), Opa Reservoir (Adedeji et al., 2020) and Watari Reservoir (Rabiu et al., 2018).

Water hardness is largely determined by the calcium and magnesium concentrations in the water. The hardness of water describes the effects of dissolved minerals and determines its suitability for drinking, domestic and industrial purposes (Kumar et al., 2011a). The relatively high values recorded for total hardness of Atori Reservoir water may be attributed to the discharge of sewage, washing and bathing with soaps and detergents as observed in this study.

Electrical conductivity (EC) is a measure of the capacity of the water to conduct electrical current and is chiefly determined by the concentrations of ions and nutrient load. The EC values reported in this study were considered relatively high when compared with similar water bodies in this part of the country. The high values recorded for conductivity in this study may be linked to the stagnant water in the reservoir. This is because conductivity has been reported to increase with stagnancy in water (Atobatele & Ugwumba, 2008). Aside from this, the circulation of water, which neutralizes salts in domestic and industrial wastes, could have also contributed to the high values of conductivity recorded in this study (Popoola et al., 2019).

Conductivity is also a good measure of the total dissolved solids (TDS) of water bodies. A water body with high TDS values is expected to have correspondingly high conductivity values. The TDS values recorded in this study could be considered relatively high for a tropical inland water body. The high values could possibly be due to high decaying vegetation, which would have increased the amount of dissolved solids in the water (Tanimu et al., 2011; Wang et al., 2007).

The nutrients were relatively low when compared with similar water bodies in tropical areas where low values have been reported for nutrients. The low concentration of the nutrients could be attributed to decreased organic matter due to mixing by runoff in the wet season, which reduced algal bloom in the water body (Popoola et al., 2019).

#### Taxonomic composition of macroinvertebrates

The abundance and diversity of macroinvertebrates recorded in Atori Reservoir can be considered relatively high and it compares favorably with studies on other water bodies in Nigeria such as Aduwo et al. (2019), Akindele et al. (2019), Aliu et al. (2020) and Amusan et al. (2018) in which high abundance and diversity of macroinvertebrates have been reported. The high diversity of macroinvertebrates reported in this study may be attributed to the stable substrata and nutrient availability in the reservoir (Aliu et al., 2020). The dominant taxon, Melanoides tuberculata, has been reportedly known to be dominant in water bodies around Nigeria. Previous studies such as Aduwo et al. (2019), Amusan et al. (2018), Anyanwu et al. (2019) and Aliu et al. (2020) have all reported M. tuberculata as the dominant taxon in their respective studies. The dominance of this taxon in most water bodies in the country has been attributed to their ability to survive in moderately impacted waters (Aliu et al., 2020).

The richness and abundance of macroinvertebrates exhibited notable seasonal variations, with considerably higher numbers recorded during the dry season. This trend aligns with previous findings by Ayoade and Aiwerioghene (2017), Makumbe et al. (2022), Kahlon et al. (2018) and Ogidiaka (2012). The lower abundance observed in the wet season could be attributed to several factors. The instability of bottom sediment due to increased rainfall and runoff during this season may have dislodged macroinvertebrates from their microhabitats (Yusuf, 2019). Additionally, higher levels of total suspended solids in the water column during the wet season reduce visibility, making macroinvertebrate collections more challenging.

Among various taxa, distinct differences in abundance were evident between the dry and wet seasons. For instance, *Biomphalaria* sp. displayed higher abundance during the dry season, while *Notonecta* sp. thrived more in the wet season. These patterns underscore the strong influence of seasonal environmental factors on macroinvertebrate distribution and abundance.

The observed variations suggest that increased precipitation during the wet season creates favorable conditions for certain species, leading to higher abundance. Conversely, fluctuations in water levels and nutrient availability during the dry season may impact macroinvertebrate communities differently. Moreover, the exclusive presence of certain species during specific seasons indicates their reliance on seasonal conditions for breeding or foraging, highlighting specialized adaptations to seasonal environmental cues.

The range of values for the Margalef and Shannon– Wiener diversity indices in clean water ranges between 3 and 4 and less than 1 in polluted water bodies (Akindele & Adeniyi, 2013; Akindele et al., 2013; Yusuf, 2019). The values recorded for the Margalef diversity index ranged from 2.77 to 4.16 while the Shannon–Wiener index values ranged from 1.77 to 2.22. This indicated that the reservoir was slightly stressed at the time the study was carried out.

# Relationship between the macroinvertebrates and water parameters.

A total of 71.96% of the variables were explained by the first two axes, with Axis 1 of the CCA triplot explaining 47.39% of the total inertia and Axis 2 explaining 24.17% of the total inertia. Axis 1 of the CCA triplot accounted for six environmental variables, i.e., TDS, acidity, alkalinity, magnesium, calcium and water hardness. Axis 2, on the other hand, accounted for a total of three environmental variables, i.e., water temperature, nitrate and phosphate. A good number of the macroinvertebrates showed positive relationships with the physicochemical parameters of the water, but canonical correspondence analysis (CCA) revealed that the relationships were not statistically significant (Monte Carlo permutation test). The dominant species, Melanoides tuberculata, showed positive relationship with some of the physicochemical parameters such as electrical conductivity, alkalinity, TDS, DO and calcium. This implied that elevated concentrations of these factors positively impacted the population of this taxon. This is not shocking, as studies have reported positive correlation between these physicochemical parameters and Melanoides tuberculata (Efitre et al., 2001; Mwabvu & Sasa, 2009). These studies revealed that most tropical waters with moderately high conductivity, calcium and DO concentrations have recorded abundance of Melanoides sp. Conductivity and calcium are essential in shell development (Mitchell et al., 2007; Alkandari et al., 2020); as such, elevated concentrations are considered favorable to the Melanoides sp. This may as well explain their abundance and dominance in the tropical inland waters.

*Pseudagrion* sp., *Naucoris* sp. and *Agasicles* sp. were related to the elevated levels of BOD while *Anax* sp. was closely related to increased nitrate values. *Anisops* sp., *Tetrathamis* sp. and *Enithares* sp. were related to magnesium values while *Biomphalaria* sp. and *Hydrophilus* sp. were both related to organic carbon and organic matter values. *Bulinus* sp., *Macrocoris* sp. and *Neogeris* sp. were also related to manganese and *Limnogonus* chloride concentrations. Furthermore, the canonical correspondence analysis (CCA) revealed that the reservoir harbors a greater number of facultative species. These species are capable of thriving across a wide range of physicochemical concentrations found in the water quality parameters, including both higher and lower concentrations. It has previously been reported that facultative organisms can thrive in aquatic environments with both good and moderately impacted water quality (Akindele et al., 2019; Voshell, 2002).

## Conclusions

This study discovered signs of water quality degradation in Atori Reservoir as a result of anthropogenic disturbances and a lack of adequate management practices within the water body's catchment basin. The dominance of pollution-tolerant species, the rarity of sensitive species and the values obtained for the water quality variables reflected that the reservoir was constantly subjected to certain stress factors. The baseline information on the ecological condition of Atori Reservoir provided in this study should serve as a basis for monitoring structures for policymakers as well as preventive measures to prevent further deterioration of the water quality in the reservoir.

#### Abbreviations

- ANOVA Analysis of variance BOD Biological oxygen demand CCA Canonical correspondence analysis
- DO Dissolved oxygen
- EC Electrical conduct
- EC Electrical conductivity TDS Total dissolved solids

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#### Author contributions

SA performed the practical section, collected the samples and analyzed the data. SA, BA and AK conceived the idea, helped in designing the study, supervised the preparation of the experiment and helped in writing the manuscript. All authors read and approved the final manuscript.

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#### Availability of data and materials

Data are available on reasonable request.

#### Declarations

# Ethics approval and consent to participate

Not applicable for this kind of research.

#### **Consent for publication**

Not applicable.

#### **Competing interests**

The authors declare that they have no competing interests.

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