RESEARCH

Open Access

Impact of sex differences on thrombin-induced hydrocephalus and white matter injury: the role of neutrophils

Kang Peng^{1,2}, Sravanthi Koduri¹, Fan Xia¹, Feng Gao¹, Ya Hua¹, Richard F. Keep¹ and Guohua Xi^{1*}

Abstract

Background Thrombin has been implicated in playing a role in hydrocephalus development following intraventricular hemorrhage (IVH). However, the mechanisms underlying the sex differences to the detrimental effects of thrombin post-IVH remain elusive.

Method Three-month old male and female Sprague-Dawley rats underwent unilateral intracerebroventricular (ICV) injections of 3U or 5U thrombin, or saline, to examine differences in thrombin-induced hydrocephalus and white matter injury. Mortality, and lateral ventricle volume and white matter injury were measured on magnetic resonance imaging evaluation at 24 h post-injection. In addition, male rats were pretreated with 17- β estradiol (E2, 5 mg/kg) or vehicle at 24 and 2 h prior to ICV injection of 3U thrombin. All rats were euthanized at 24 h post-injection for histology and immunohistochemistry.

Results ICV injection of 5U thrombin caused 100 and 0% mortality in female and male rats, respectively. 3U of thrombin resulted in significant ventricular dilation and white matter damage at 24 h in both male and female rats, but both were worse in females (p < 0.05). Furthermore, neutrophil infiltration into choroid plexus and periventricular white matter was enhanced in female rats and may play a critical role in the sex difference in brain injury. Pre-treating male rats with E2, increased thrombin (3U)-induced hydrocephalus, periventricular white matter injury and neutrophil infiltration into the choroid plexus and white matter.

Conclusions ICV thrombin injection induced more severe ventricular dilation and white matter damage in female rats compared to males. Estrogen appears to contribute to this difference which may involve greater neutrophil infiltration in females. Understanding sex differences in thrombin-induced brain injury may shed light on future interventions for hemorrhagic stroke.

Keywords Thrombin, Male, Female, Neutrophils, Hydrocephalus, White matter injury

*Correspondence:

Guohua Xi

guohuaxi@umich.edu

¹ Department of Neurosurgery, University of Michigan, R5018 Biomedical Science Research Building, 109 Zina Pitcher Place, Ann Arbor, MI 48109-2200, USA

² Department of Neurosurgery, Xiangya Hospital, Central South University, Changsha, China

Background

Intraventricular hemorrhage (IVH) is a common complication of intracerebral hemorrhage (ICH) and subarachnoid hemorrhage (SAH) in adults, and for germinal matrix hemorrhage in newborns [1-3]. Hydrocephalus occurs frequently after IVH and it is correlated with higher hemorrhage-associated mortality [4, 5]. Ventricular zone disruption and ependymal denudation or discontinuities have been observed in human neonates with IVH or communicating hydrocephalus [6, 7].



© The Author(s) 2021, corrected publication 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data. Furthermore, accumulating evidence implicates hydrocephalus as a potential cause of axonal and myelin damage in the periventricular white matter (WM), which may contribute to poor behavioral outcomes after IVH [8, 9].

Thrombin is an essential component of the coagulation cascade and is produced in the brain immediately after a hemorrhagic event to induce hemostasis [10]. Thrombin has been shown to participate in brain injury after both ICH and cerebral ischemia [11–13]. Our previous studies have also demonstrated that thrombin is involved in hydrocephalus development after IVH, and intracerebroventricular (ICV) injection of thrombin can cause severe hydrocephalus and induce neuroinflammation within the choroid plexus [14, 15].

Sex is an influential factor impacting the severity of brain injury following hemorrhagic stroke. Several studies have reported a higher incidence of mortality following IVH in premature male infants, compared to female infants [16, 17]. Additionally, our previous studies have shown that female mice have reduced cerebral edema and improved behavioral recovery following ICH [18]. Furthermore, estrogen has been found to reduce ironmediated intracerebral edema and neuronal death [19]. These findings suggest that the female sex may be protected after IVH and ICH. However, previous studies have shown that women had higher morbidity and worse brain injury after suffering SAH [20]. In addition, some clinical investigations indicate that being female is an independent factor for higher incidence of hydrocephalus after SAH [21, 22]. The current study investigates whether there are sex differences in thrombin-induced hydrocephalus and periventricular WM injury, and further elucidates the underlying mechanisms.

Methods

Animal preparation and intraventricular injection

All animal protocols were approved by the University of Michigan Institutional Animal Care and Use Committee. A total of forty-eight 3-month-old Sprague-Dawley rats (Charles River Laboratories, Portage, MI), including 30 male and 18 female rats, were used in this study. Animals were anesthetized with pentobarbital (50 mg/kg intraperitoneally) and core body temperature was maintained at 37.5 °C during intraventricular injection and recovery from anesthesia. Rats received an ICV infusion of saline or thrombin into the right lateral ventricle (coordinates: 0.6 mm posterior, 4.5 mm ventral, and 1.6 mm lateral to the bregma).

Experimental groups

This study included 3 parts: (1) male and female rats (n=6 for each) underwent an unilateral ICV injection of 5 units (5U) rat thrombin (Sigma-Aldrich) dissolved

in saline (50 µl); (2) rats of both sexes received an unilateral ICV injection of 3 units (3U) thrombin dissolved in saline or saline alone (50 µl) for a total of four groups (n=6 for each group); (3) male rats were pretreated with 17- β estradiol (E2; 5 mg/kg dissolved into saline with 1% gelatin, subcutaneous) or vehicle (saline with 1% gelatin) at 24 and 2 h prior to the ICV injection of 3U thrombin (n=6 for each group). All injections were performed over 10 min. Following MRI at 24 h post-ICV injection, rats were euthanized for immunohistochemistry analysis.

MRI scanning and measurement

All rats underwent MRI at 24 h after saline or thrombin ICV injections. MRI was performed in a 7.0-T Varian MR scanner (Varian Inc.) with a T2 fast spin-echo sequence using a field of view of 35mm×35 mm and obtaining a total of 25 coronal slices. Ventricle volume and WM damage area were measured as described previously [23–25]. Briefly, bilateral lateral ventricles as well as WM T2-hyperintensity area were outlined and quantified all slices using Image J (National Institutes of Health, USA) by a blinded observer.

Hematoxylin & eosin and immunohistochemistry staining

Rats were euthanized with pentobarbital (100 mg/kg, intraperitoneally) and perfused with 4% paraformaldehyde. Brains were removed and placed in 4% paraformaldehyde for 24 h at 4 °C prior to immersion in 30% sucrose for 48-72 h at 4 °C. Brains were then embedded in optimal cutting temperature compound (Sakura Finetek USA) and sectioned on a cryostat (18-µm-thick sections). Hematoxylin and eosin (H&E) staining was used to measure the ventricular wall damage as well as neutrophil infiltration (by morphology). Immunohistochemical studies were performed using the avidin-biotin complex technique as previously described [26]. The primary antibody was rabbit anti-myeloperoxidase (MPO; 1:200; Invitrogen) and the secondary antibody was goat anti-rabbit (1:500; Invitrogen). Negative controls omitted the primary antibody. Hematoxylin was used for contrast staining.

Cell counting and immunoactivity determination

To assess the severity of ventricular wall damage and avoid the area of direct damage due to the ICV injection, the temporal horns of the lateral ventricles (coronal brain section between -3.8 and -4.3 mm posterior to bregma) were chosen and the ratio of disrupted ependymal surface to total ventricular wall length within one slice was calculated as a percentage. For MPO positive cells counts, three random high-power images (×40 magnification) were obtained of the choroid plexus (based on the coronal brain sections between -0.4 and -0.9 mm posterior

to bregma) and periventricular WM from bilateral lateral ventricles, and the number of MPO positive cells is represented as cells per square millimeter. All analyses were completed with ImageJ by a blinded observer.

Statistical analysis

Values are given as means \pm SD. Student *t*-test and twoway ANOVA were used for analysis. Differences were considered significant at p < 0.05.

Results

Intracerebroventricular injection of 5U of rat thrombin caused 100% mortality in female rats (6/6) but there were no deaths in male rats (0/6). When 3U thrombin or saline were injected ICV, there were no deaths in either female or male rats. Our previous study showed that ICV injection of thrombin (3U) resulted in ventricular enlargement as determined by MRI [14]. Therefore, we used 3U thrombin in the remainder of this study.

Intracerebroventricular injection of thrombin induced

ventricular dilation and white matter damage in male rats. Intracerebroventricular thrombin (3U) increased lateral ventricle volume in male rats compared to saline injection at 24 h post-injection (Fig. 1A). Additionally, compared to the saline group, greater ventricular wall damage as determined based on the disruption of the ependymal layer, sparse WM and cellular shedding, was observed in male rats with ICV thrombin (Fig. 1B). Neuroinflammation within the choroid plexus may contribute to the occurrence of hydrocephalus. Our previous study revealed that ICV thrombin injection mimics the choroid plexus inflammation that was observed after subarachnoid hemorrhage [15]. In the current study, neuroinflammation within the choroid plexus was detected at 24 h after ICV thrombin injection by H&E staining with marked polymorphonuclear neutrophil infiltration in male rats. MPO staining confirmed that ICV thrombin injection induced choroid plexus neutrophil infiltration. Even within the ventricular wall, we also found there were numerous neutrophils infiltration within the ependymal cells layer, which may contribute to the ventricular wall damage (Fig. 1C).

Periventricular T2-hyperintensity, a marker of WM injury, has been shown to be induced in rats after ICV thrombin injection, but not saline injection [27]. In the current experiments, periventricular T2-hyperintensity area was also found to be larger in male rats who underwent thrombin injection than with saline injection at 24 h post-injection (Fig. 2A). To gain further understanding of the potential mechanism of thrombin-induced WM damage, MPO staining was performed and showed marked neutrophil infiltration into the periventricular WM injury region following ICV injection of thrombin but not saline in male rats (Fig. 2B). Based on the above data, neutrophils may play a role in thrombin-induced periventricular WM injury.

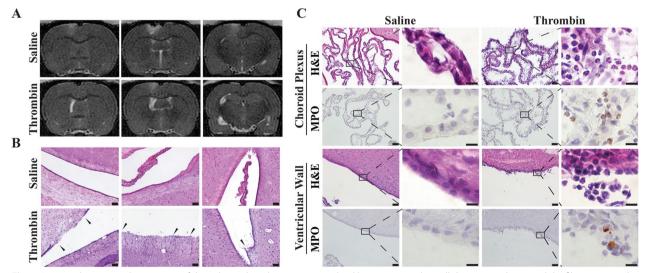


Fig. 1 Intracerebroventricular injection of thrombin induced severe ventricular dilation, ventricular wall damage, and neutrophil infiltration in male rats. **A** T2 weighted MRI showing ventricular volume at 24 h after ICV injection of 50 μ I of saline or thrombin (3U) in male rats. The bottom left image of this panel has been published previously [15]. **B** Representative images of H&E staining showing ependymal denudation and rupture (arrows) at 24 h in the thrombin (3U) but not the saline group. Scale bar = 50 μ m. **C** Representative H&E and myeloperoxidase (MPO) staining of the choroid plexus and ventricle wall 24 h after thrombin or saline injection. Note the neutrophil infiltration into the choroid plexus and the ventricular wall damage in the thrombin injection group. Lower magnification, scale bar = 50 μ m; higher magnification, scale bar = 10 μ m

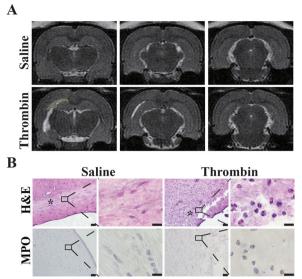


Fig. 2 Intracerebroventricular injection of thrombin induced white matter damage and neutrophil infiltration in male rats. **A** T2 weighted MRI showing white matter hyperintensity (outline) at 24 h after ICV injection of thrombin (3U) injection but not saline injection in male rats. **B** Representative H&E and MPO staining images showing neutrophil infiltration into the white matter injury area at 24 h in the thrombin injection group. ***** indicates white matter area. Lower magnification, scale bar = $50 \ \mu m$; higher magnification, scale bar = $10 \ \mu m$

Intracerebroventricular injection of thrombin induced more severe ventricle dilation and white matter damage in female rats compared to males

To investigate differences in injury due to sex, 3U of thrombin was injected ICV in female rats. Similar to male rats, ICV thrombin induced ventricular dilation in female rats at 24 h ($36.1 \pm 12.5 \text{ mm}^3$) compared to saline injection ($9.3 \pm 1.9 \text{ mm}^3$, p < 0.01; Fig. 3A and C). Furthermore, ventricular volume after ICV thrombin was significantly higher in female rats compared to males ($36.1 \pm 12.5 \text{ vs}$. $23.1 \pm 5.1 \text{ mm}^3$ in male group, p < 0.05; Fig. 3A and C). There was no significant difference in ventricular volume between female and male saline-injected groups ($9.3 \pm 1.9 \text{ vs}$. $8.5 \pm 0.8 \text{ mm}^3$, respectively; p > 0.05, Fig. 3A and C).

Interestingly, there was a very robust increase in polymorphonuclear neutrophil infiltration within the choroid plexus at 24 h after ICV thrombin injection in female rats (Fig. 3B). MPO staining confirmed that ICV thrombin-induced neutrophil infiltration into the choroid plexus of female rats (938±313 vs. 0 per mm² in saline group, p < 0.01; Fig. 3B and D). When quantified, there was almost a 2-fold higher density of neutrophils within the choroid plexus of female rats compared to male rats at 24 h after thrombin injection (938±313 vs. 576±216 per mm² in male rats, p < 0.01; Fig. 3D).

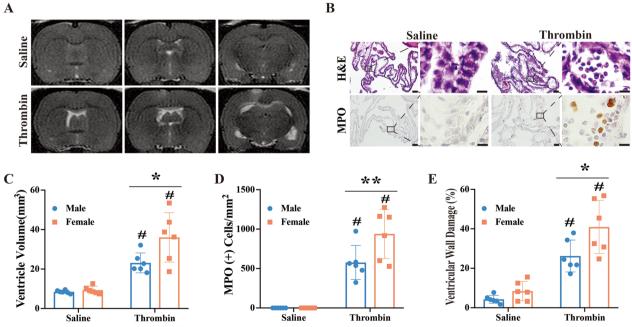


Fig. 3 Intracerebroventricular injection of thrombin induced greater ventricular dilation, worse ventricular wall damage, and more neutrophil infiltration into choroid plexus in female rats compared to males. **A** T2 weighted MRI showing ventricular volume at 24 h after ICV injection of 50 μ l of saline or thrombin (3U) in female rats. **B** Representative H&E and myeloperoxidase (MPO) staining images showing neutrophil infiltration into the choroid plexus after ICV injection of thrombin but not saline. Lower magnification, scale bar = 50 μ m; higher magnification, scale bar = 10 μ m. **C** Quantification of ventricular dilation in the male and female rats. **D** Quantification of MPO positive cell infiltration into the choroid plexus in male and female groups. **E** Quantification of ventricular wall damage in the male and female groups. Values are expressed as the means ± SD, n = 6 in each group, **#** p < 0.01 vs. saline, *****p < 0.05, ******p < 0.01

Similarly, ICV thrombin caused marked ventricular wall damage in female rats ($40.9 \pm 8.5\%$ vs. $8.4 \pm 4.9\%$ in saline group, p < 0.01; Fig. 3E) and this damage was greater than that observed in male rats at 24 h after thrombin injection ($40.9 \pm 8.5\%$ vs. $26.3 \pm 8.1\%$ in male group, p < 0.05; Fig. 3E).

Compared to the saline group, female rats injected ICV with thrombin had a much larger periventricular T2-hyperintensity lesion at 24 h (12.8 ± 8.2 vs. 1.1 ± 0.7 mm^3 in saline group, p<0.01; Fig. 4A). The female rat thrombin injection group also had more WM injury than the male rat thrombin injection group (12.8 ± 8.2) vs. 3.6 ± 2.1 mm³ in male rats, p<0.05, Fig. 4B). Similar to male rats, female rats injected with thrombin ICV showed a robust increase in neutrophils within the region of WM injury when compared to saline injected rats (Fig. 4C). The number of MPO positive cells was calculated within the WM injury region and were drastically higher following thrombin injection $(1092 \pm 680 \text{ vs. 0 per})$ mm^2 in saline group, p<0.01; Fig. 4D). In female rats, the number of neutrophils within the WM injury after ICV thrombin was significantly higher than in male rats $(1092 \pm 680 \text{ vs.} 281 \pm 284 \text{ per mm}^2 \text{ in male group, } p < 0.05;$ Fig. 4D). Altogether, these data indicate that ICV thrombin induced more severe brain damage in female rats compared to male rats and that neutrophils may contribute to this difference. It should be noted that thrombininduced ventricular injury was bilateral.

17-β estradiol pre-treatment in male rats exacerbates the ventricular dilation, WM injury and neutrophil infiltration induced by ICV injection of thrombin

To further understand the potential mechanism underlying the sex differences noted above, male rats were treated with 17- β estradiol via subcutaneous injection at 24 and 2 h before ICV thrombin injection. Thrombin injection induced greater ventricular enlargement in the estradiol pre-treatment group at 24 h post-injection compared to the vehicle pre-treatment group (36.0±9.8 vs. 23.4±3.5 mm³ in vehicle, p<0.05; Fig. 5A).

This increase in ventricular volume was accompanied by a concomitant increase in neutrophil infiltration into choroid plexus in both lateral ventricles in the estradiol pretreatment group compared to the vehicle pre-treatment group (Fig. 5B). Myeloperoxidase staining, which was consistent with neutrophil identification on H&E staining,

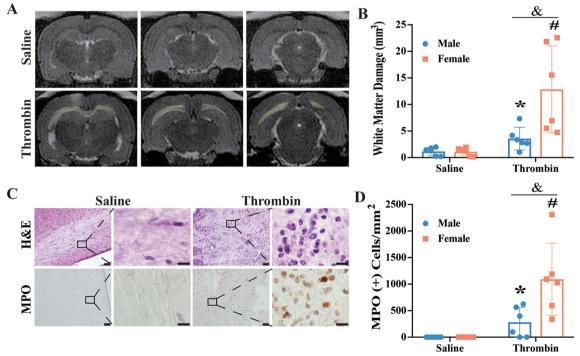


Fig. 4 Intracerebroventricular injection of thrombin induces worse white matter damage and more white matter neutrophil infiltration in female compared to male rats. **A** Representative T2 weighted MRI showing white matter hyperintensity (outline) at 24 h after ICV injection of thrombin (3U) but not saline injection in female rats. **B** Quantification of white matter injury in male and female rats 24 h after ICV injection of 3U thrombin or saline. **C** Representative H&E and MPO staining images showing neutrophil infiltration into the white matter injury region at 24 h after ICV injection of 3U thrombin or saline. **C** Representative H&E and MPO staining images showing neutrophil infiltration of MPO positive cells within the white matter injury region in male and female rats 24 h after ICV injection of 3U thrombin or saline. **D** Quantification of MPO positive cells within the white matter injury region in male and female rats 24 h after ICV injection of 3U thrombin or saline. Values are expressed as the means ± SD, n = 6 in each group; *p < 0.05, #p < 0.01 vs. saline. [&]p < 0.05. Lower magnification, scale bar = 50 µm; higher magnification, scale bar = 10 µm

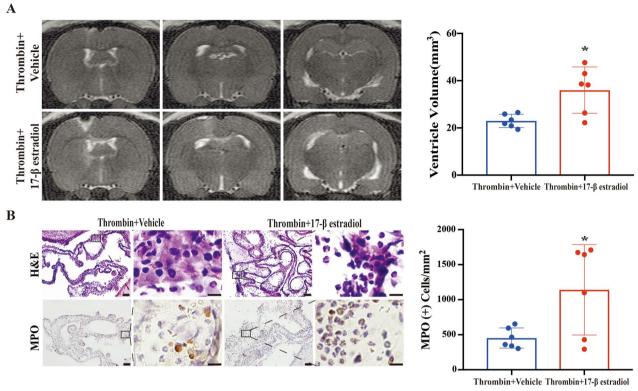


Fig. 5 Pre-treatment with 17- β estradiol in male rats leads to greater ventricular dilation and more neutrophil infiltration into choroid plexus after ICV thrombin injection. **A** T2 weighted MRI showing ventricular volume 24 h after ICV thrombin injection in male rats pretreated with 17- β estradiol or vehicle. Quantification of ventricular volume for each group is shown in the bar chart. **B** Representative H&E and MPO staining images showing neutrophil infiltration into the choroid plexus 24 h after ICV injection of thrombin in male rats pretreated with 17- β estradiol or vehicle. Quantification of ventricular volume for each group is shown in the bar chart. **B** Representative H&E and MPO staining images showing neutrophil infiltration into the choroid plexus 24 h after ICV injection of thrombin in male rats pretreated with 17- β estradiol or vehicle. Quantification of the number of MPO positive cells in each group is shown in the bar chart. Values are expressed as the means ± SD, n = 6, *p < 0.05 vs. vehicle treatment group. Lower magnification, scale bar = 50 µm; higher magnification, scale bar = 10 µm

showed a greater than 2-fold increase in MPO (+) cells in the estradiol pre-treatment group compared to the vehicle group (1142 ± 646 vs. 452 ± 144 per mm² in vehicle, p<0.05; Fig. 5B).

In addition, 17- β estradiol pre-treatment in male rats also resulted in greater thrombin-induced WM damage on MRI compared to the vehicle group (10.5±7.2 vs. 2.5±3.2 mm³ in vehicle, p<0.05; Fig. 6A). This increased damage with estradiol pre-treatment was accompanied by an increase in the number of polymorphonuclear neutrophils after ICV thrombin in the estradiol pre-treated group (Fig. 6B). This was confirmed by an increased number of MPO positive cells in the WM injury region in 17- β estradiol treated male rats (721±500 vs. 205±249 per mm² in vehicle, p<0.05; Fig. 6B). These data indicate that 17- β estradiol treatment worsens WM damage in the male ICV thrombin injection model.

Neutrophil infiltration correlates with ventricular dilation and WM injury induced by unilateral ICV injection of thrombin

To further elucidate whether neutrophil infiltration contributes to ventricular dilation and WM injury, linear regression analysis was performed in all rats injected with thrombin ICV. There was a positive correlation between ventricular dilation and neutrophil infiltration within the choroid plexus bilaterally (r=0.65, p<0.0005, linear regression analysis, Fig. 7A). Furthermore, WM injury also had a positive correlation with neutrophil infiltration within the WM injury region bilaterally (r=0.84, p<0.0001, linear regression analysis, Fig. 7B).

Taken together, these data further suggest that neutrophils play a prominent role in the sex differences noted after ICV thrombin injection. This role may be related to hormonal control, particularly with respect to estrogen.

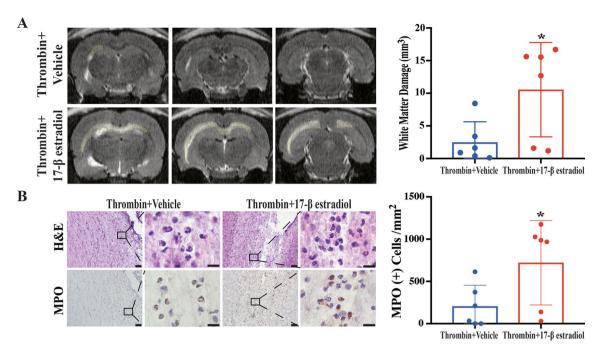


Fig. 6 Pre-treatment with 17- β estradiol in male rats leads to increased white matter damage and more neutrophil infiltration after ICV injection of thrombin. **A** T2 weighted MRI showing white matter hyperintensity (outline) at 24 h after thrombin intraventricular injection in male rats pretreated with 17- β estradiol or vehicle. Quantification of the white matter damage in each group is shown in the bar chart. **B** Representative H&E and MPO staining images showing more neutrophil infiltration into the white matter injury region 24 h post thrombin-injection in male rats pretreated with 17- β estradiol or vehicle. Quantification of the number of MPO positive cells in each group is shown in the bar chart. Values are expressed as the means ± SD, n = 6 in each group; *p < 0.05 vs. male vehicle treatment group. Lower magnification, scale bar = 50 µm; higher magnification, scale bar = 10 µm

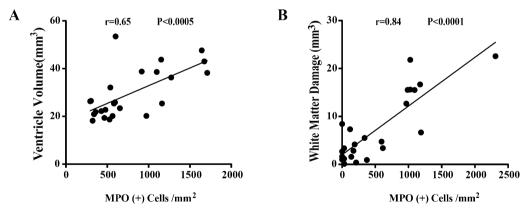


Fig. 7 Neutrophil infiltration correlates with the ventricular dilation and WM injury induced by ICV injection of thrombin. **A** Linear regression analysis showing thrombin-induced ventricular dilation correlates with neutrophil infiltration of the choroid plexus. n = 24, p < 0.0005. **B** Linear regression analysis showing thrombin-induced WM injury correlates with neutrophil infiltration of the WM injury area. n = 24, p < 0.0001

Discussion

Based on the data presented above, there are three major findings: (1) unilateral ICV thrombin injection causes higher mortality and results in more severe ventricular enlargement and periventricular WM damage in female than male rats; (2) $17-\beta$ estradiol pre-treatment

aggravates thrombin-induced hydrocephalus and WM injury in male rats; and (3) neutrophils may play a central role in thrombin-induced brain damage as well as in the mechanism leading to sex differences of this pathology. Our prior study showed that thrombin plays a central role in the development of hydrocephalus after IVH,

and that ICV injection of thrombin causes significant ventricular dilatation in male rats [14]. This study replicated those results in male rats and extended the finding to female rats and further elucidated the potential mechanisms underlying those pathological changes.

Thrombin is produced immediately after a brain hemorrhage. It is known that 100 µl of whole blood can produce about 26 to 36 units of thrombin [28]. In this study, ICV injection of 5U thrombin in female rats caused 100% mortality but 0% mortality in male rats. Similarly, ICV injection of 3U thrombin induced significantly greater ventricular enlargement, more severe ventricular wall damage, and worse WM injury in female than male rats. Male rats who underwent pretreatment with 17-β estradiol also had greater ventricular enlargement, more severe ventricular wall damage, and worse WM injury than untreated male rats. The degree of ventricular enlargement, however, has been modest compared to the severe ventriculomegaly seen in human IVH. This is thought to be secondary to the localized injection of thrombin into the right lateral ventricle, whereas IVH is diffuse. Also, these injections create a bolus of thrombin at a specific time point rather than the gradual release of thrombin that is observed in IVH. Both of these deviations from human physiology may be causal for this modest ventriculomegaly found in the animal model. Overall, these data indicate that females have worse outcome after ICV thrombin injection with estradiol playing a significant role in the mechanism generating this difference.

Ependymal denudation and discontinuities are the dominant alterations in the damage process associated with IVH and hydrocephalus and may potentially contribute to the development of hydrocephalus [6, 7]. An interesting phenomenon found in the *hyh* mutant mouse, which has congenital moderate communicating hydrocephalus, is the presence of ventral ependymal denudation early in fetal life, even prior to the occurrence of communicating hydrocephalus. This leads to the possibility that ventral ependymal denudation may contribute to the mechanism underlying the development of communicating hydrocephalus [29]. In the current study, ependymal denudation and regions of sparse WM were observed after ICV thrombin injection, consistent with previous reports. A recent study successfully established a preterm IVH model in vitro that allows for the exploration of potential pathophysiologic alterations within the ventricular zone [30]. Further study of the effect of thrombin on preterm infants is needed and can be examined in such a model.

The impact of sex on different types of brain injury is complex. There have been multiple studies indicating female animals sustain less severe brain damage after ischemic and hemorrhagic stroke (e.g. [18, 31–33]).

Page 8 of 10

Several clinical studies have also demonstrated that, in the human neonatal population, male neonates suffer a higher rate of mortality and secondary hydrocephalus following IVH, especially severe IVH (grades 3 and 4) [16, 17]. In contrast, other studies have shown that there is no significant difference in the development of hydrocephalus after IVH between male and female neonates [34, 35]. However, previous studies have revealed that being female is an independent risk factor associated with hydrocephalus after SAH [21, 22]. In fact, a study from our group indicated that tamoxifen, a selective estrogen receptor modulator, caused hydrocephalus in male rats after ICH [36]. Estrogen administration enhances thrombin generation in the rat model and 17- β estradiol administration has been shown to lead to platelet aggregation and reduce tolerance to thrombin [37, 38]. These studies suggest that estrogen may play a role in promoting the toxic effects of thrombin, in line with the results of the current study. Nonetheless, the exact mechanisms that underlie this sex difference in thrombin-induced hydrocephalus are still unclear, though neutrophils may play an important role.

Neutrophils are a first line of defense of the innate immune system against external microbes [39]. However, several reports have demonstrated that neutrophils can play a deleterious role in intracerebral injury by releasing extracellular traps (NETs). Depletion of neutrophils and subsequent decreased formation of NETs can reduce neuronal death and improve outcomes after ischemic and hemorrhagic stroke [40–42]. Our previous studies have shown that neutrophils may participate in the development of post-hemorrhagic hydrocephalus [43, 44]. However, within the current literature on hydrocephalus, there has been limited investigation of the mechanisms involving neutrophils.

In the current study, a greater than 2-fold increase in neutrophil infiltration into choroid plexus and regions of WM damage was observed at 24 h after ICV thrombin injection in female rats compared to male rats, and male rats pre-treated with $17-\beta$ estradiol compared to vehicle, suggesting that neutrophils may play a crucial role in this sex difference. Thrombin, acting as a damage-associated molecular patterns (DAMPs), may enhance the expression of inflammatory adhesion molecules (e.g. VCAM-1, ICAM-1, P-selectin, etc.), which are essential proteins in the process of neutrophil infiltration [45]. 17-ß estradiol administration may reduce tolerance to thrombin and therefore, aggravate the resultant toxic effects [37]. Additionally, an *in vitro* study showed that $17-\beta$ estradiol could promote neutrophil extracellular trap formation, which has been shown to be a mechanism for causing intracerebral injury [46]. Further investigations of the role of neutrophils in generating the sex differences found in this study are needed.

It should be noted that multiple mechanisms are involved in IVH-, SAH- and ICH-induced brain injury. Thus, for example, neuroinflammation after hemorrhagic stroke involves multiple types of leukocytes and resident microglia. The impact of being female or male, and the role of sex hormones may depend on which injury pathways are activated in particular forms of hemorrhagic stroke.

There are several limitations to this study. Only adult rats were used in this study. Therefore, there is a need for more investigation to fully understand the effect of thrombin on preterm and term infants. Only the effect of estrogen was investigated and there may be other sex hormones involved in creating the observed sex differences. Only two doses of thrombin were given ICV and the form of administration does not fully reflect the human pathophysiology where thrombin is gradually released from an IVH. In addition, only acute experiments within a 24-hour timeframe were completed and therefore, it is unclear whether there may be a recovery back to normal ventricular size and repair of the associated damage or if the damage is permanent. Finally, functional outcomes were not examined, so longerterm studies are needed with behavioral testing to understand the chronic implications of ICV thrombin injection.

Conclusions

In conclusion, ICV injection of thrombin induced higher mortality, increased ventricular dilation and worsened WM injury in female rats compared to male rats. Those effects were mimicked by pre-treating male rats with estrogen and appear, at least in part, to be related to differences in neutrophil infiltration. These results indicate there may be sex-related targets for reducing IVH-induced brain injury and hydrocephalus.

Abbreviations

DAMP Damage-associated molecular patterns

- E2 17-β estradiol
- H&E Hematoxylin and eosin
- ICH Intracerebral hemorrhage ICV Intracerebroventricular
- IVH Intraventricular hemorrhage
- MPO Myeloperoxidase
- MRI Magnetic resonance imaging
- NETS Neutrophil extracellular traps
- SAH Subarachnoid hemorrhage
- SD Standard deviation
- U Units
- WM White matter

Acknowledgements

Not applicable.

Authors' contributions

KP, FG performed the experiments and tissue staining. KP, FX analyzed the data. KP, SK and FX wrote the manuscript. GX, YH and RFK conceived the original idea and supervised the project. All authors read and approved the final manuscript.

Funding

Y.H., R.F.K. and G.X. were supported by Grants NS-096917, NS-106746, NS-112394 and NS-116786; SK was supported by NS-007222 from the National Institutes of Health.

Availability of data and materials

The datasets used and/or analyzed in this study are available from the corresponding author (G.X.) upon reasonable request.

Declarations

Ethics approval and consent to participate

The animal use protocols were approved by the Committee on the Use and Care of Animals of the University of Michigan.

Consent for publication

Not applicable.

Competing interests

RFK is a co-Editor-in-Chief of Fluids and Barriers of the CNS.

Received: 24 June 2021 Accepted: 10 August 2021 Published: 16 August 2021

References

- Rosen DS, Macdonald RL, Huo D, Goldenberg FD, Novakovic RL, Frank JI, et al. Intraventricular hemorrhage from ruptured aneurysm: clinical characteristics, complications, and outcomes in a large, prospective, multicenter study population. J Neurosurg. 2007;107(2):261–5.
- Bhattathiri PS, Gregson B, Prasad KS, Mendelow AD, Investigators S. Intraventricular hemorrhage and hydrocephalus after spontaneous intracerebral hemorrhage: results from the STICH trial. Acta Neurochir Suppl. 2006;96:65–8.
- Hanley DF. Intraventricular hemorrhage: severity factor and treatment target in spontaneous intracerebral hemorrhage. Stroke. 2009;40(4):1533–8.
- Diringer MN, Edwards DF, Zazulia AR. Hydrocephalus: a previously unrecognized predictor of poor outcome from supratentorial intracerebral hemorrhage. Stroke. 1998;29(7):1352–7.
- Nieuwkamp DJ, de Gans K, Rinkel GJ, Algra A. Treatment and outcome of severe intraventricular extension in patients with subarachnoid or intracerebral hemorrhage: a systematic review of the literature. J Neurol. 2000;247(2):117–21.
- Dominguez-Pinos MD, Paez P, Jimenez AJ, Weil B, Arraez MA, Perez-Figares JM, et al. Ependymal denudation and alterations of the subventricular zone occur in human fetuses with a moderate communicating hydrocephalus. J Neuropathol Exp Neurol. 2005;64(7):595–604.
- McAllister JP, Guerra MM, Ruiz LC, Jimenez AJ, Dominguez-Pinos D, Sival D, et al. Ventricular zone disruption in human neonates with intraventricular hemorrhage. J Neuropathol Exp Neurol. 2017;76(5):358–75.
- 8. Yeo SS, Choi BY, Chang CH, Jung YJ, Ahn SH, Son SM, et al. Periventricular white matter injury by primary intraventricular hemorrhage: a diffusion tensor imaging study. Eur Neurol. 2011;66(4):235–41.
- Whitelaw A. Intraventricular haemorrhage and posthaemorrhagic hydrocephalus: pathogenesis, prevention and future interventions. Semin Neonatol. 2001;6(2):135–46.
- Ye F, Garton HJL, Hua Y, Keep RF, Xi G. The role of thrombin in brain injury after hemorrhagic and ischemic stroke. Transl Stroke Res. 2021;12(3):496–511.
- 11. Chen B, Cheng Q, Yang K, Lyden PD. Thrombin mediates severe neurovascular injury during ischemia. Stroke. 2010;41(10):2348–52.

- 12. Xi G, Keep RF, Hoff JT. Mechanisms of brain injury after intracerebral haemorrhage. Lancet Neurol. 2006;5(1):53–63.
- Keep RF, Hua Y, Xi G. Intracerebral haemorrhage: mechanisms of injury and therapeutic targets. Lancet Neurol. 2012;11(8):720–31.
- Gao F, Liu F, Chen Z, Hua Y, Keep RF, Xi G. Hydrocephalus after intraventricular hemorrhage: the role of thrombin. J Cereb Blood Flow Metab. 2014;34(3):489–94.
- Wan Y, Hua Y, Garton HJL, Novakovic N, Keep RF, Xi G. Activation of epiplexus macrophages in hydrocephalus caused by subarachnoid hemorrhage and thrombin. CNS Neurosci Ther. 2019;25(10):1134–41.
- Tioseco JA, Aly H, Essers J, Patel K, El-Mohandes AA. Male sex and intraventricular hemorrhage. Pediatr Crit Care Med. 2006;7(1):40–4.
- Vassilyadi M, Tataryn Z, Shamji MF, Ventureyra EC. Functional outcomes among premature infants with intraventricular hemorrhage. Pediatr Neurosurg. 2009;45(4):247–55.
- Nakamura T, Xi G, Hua Y, Schallert T, Hoff JT, Keep RF. Intracerebral hemorrhage in mice: model characterization and application for genetically modified mice. J Cereb Blood Flow Metab. 2004;24(5):487–94.
- 19. Gu Y, Xi G, Liu W, Keep RF, Hua Y. Estrogen reduces iron-mediated brain edema and neuronal death. Acta Neurochir Suppl. 2010;106:159–62.
- Kongable GL, Lanzino G, Germanson TP, Truskowski LL, Alves WM, Torner JC, et al. Gender-related differences in aneurysmal subarachnoid hemorrhage. J Neurosurg. 1996;84(1):43–8.
- Sheehan JP, Polin RS, Sheehan JM, Baskaya MK, Kassell NF. Factors associated with hydrocephalus after aneurysmal subarachnoid hemorrhage. Neurosurgery. 1999;45(5):1120–7. discussion 7–8.
- Dorai Z, Hynan LS, Kopitnik TA, Samson D. Factors related to hydrocephalus after aneurysmal subarachnoid hemorrhage. Neurosurgery. 2003;52(4):763–9. discussion 9–71.
- Okauchi M, Hua Y, Keep RF, Morgenstern LB, Xi G. Effects of deferoxamine on intracerebral hemorrhage-induced brain injury in aged rats. Stroke. 2009;40(5):1858–63.
- Okubo S, Strahle J, Keep RF, Hua Y, Xi G. Subarachnoid hemorrhageinduced hydrocephalus in rats. Stroke. 2013;44(2):547–50.
- Chen Z, Gao C, Hua Y, Keep RF, Muraszko K, Xi G. Role of iron in brain injury after intraventricular hemorrhage. Stroke. 2011;42(2):465–70.
- Wu J, Hua Y, Keep RF, Nakamura T, Hoff JT, Xi G. Iron and iron-handling proteins in the brain after intracerebral hemorrhage. Stroke. 2003;34(12):2964–9.
- Ni W, Gao F, Zheng M, Koch LG, Britton SL, Keep RF, et al. Effects of aerobic capacity on thrombin-induced hydrocephalus and white matter injury. Acta Neurochir Suppl. 2016;121:379–84.
- Lee KR, Colon GP, Betz AL, Keep RF, Kim S, Hoff JT. Edema from intracerebral hemorrhage: the role of thrombin. J Neurosurg. 1996;84(1):91–6.
- Jimenez AJ, Tome M, Paez P, Wagner C, Rodriguez S, Fernandez-Llebrez P, et al. A programmed ependymal denudation precedes congenital hydrocephalus in the hyh mutant mouse. J Neuropathol Exp Neurol. 2001;60(11):1105–19.
- Castaneyra-Ruiz L, McAllister JP 2nd, Morales DM, Brody SL, Isaacs AM, Limbrick DD. Jr. Preterm intraventricular hemorrhage in vitro: modeling the cytopathology of the ventricular zone. Fluids Barriers CNS. 2020;17(1):46.
- Nakamura T, Hua Y, Keep RF, Park JW, Xi G, Hoff JT. Estrogen therapy for experimental intracerebral hemorrhage in rats. J Neurosurg. 2005;103(1):97–103.
- Zhang YQ, Shi J, Rajakumar G, Day AL, Simpkins JW. Effects of gender and estradiol treatment on focal brain ischemia. Brain Res. 1998;784(1–2):321–4.
- Friedrich V, Bederson JB, Sehba FA. Gender influences the initial impact of subarachnoid hemorrhage: an experimental investigation. PLoS One. 2013;8(11):e80101.
- Flores G, Amaral-Nieves N, De Jesus O, Vigo J. Risk factors for developing hydrocephalus in neonatal intraventricular hemorrhage. P R Health Sci J. 2020;39(1):55–7.
- Lee IC, Lee HS, Su PH, Liao WJ, Hu JM, Chen JY. Posthemorrhagic hydrocephalus in newborns: clinical characteristics and role of ventriculoperitoneal shunts. Pediatr Neonatol. 2009;50(1):26–32.
- Xie Q, Guan J, Wu G, Xi G, Keep RF, Hua Y. Tamoxifen treatment for intracerebral hemorrhage. Acta Neurochir Suppl. 2011;111:271–5.
- Solov'ev VG, Byshevskii A, Karpova IA. The effect of estrogens and progestagens on biochemical components of hemostasis, platelets, continuous

intravascular coagulation and tolerance to thrombin: correction of their effect of antioxidants. Biomed Khim. 2012;58(4):429–37.

- Ohashi R, Sugimura M, Kanayama N. Estrogen administration enhances thrombin generation in rats. Thromb Res. 2003;112(5–6):325–8.
- 39. Segal AW. How neutrophils kill microbes. Annu Rev Immunol. 2005;23:197–223.
- 40. Kang L, Yu H, Yang X, Zhu Y, Bai X, Wang R, et al. Neutrophil extracellular traps released by neutrophils impair revascularization and vascular remodeling after stroke. Nat Commun. 2020;11(1):2488.
- Herz J, Sabellek P, Lane TE, Gunzer M, Hermann DM, Doeppner TR. Role of neutrophils in exacerbation of brain injury after focal cerebral ischemia in hyperlipidemic mice. Stroke. 2015;46(10):2916–25.
- Auriat A, Plahta WC, McGie SC, Yan R, Colbourne F. 17beta-Estradiol pretreatment reduces bleeding and brain injury after intracerebral hemorrhagic stroke in male rats. J Cereb Blood Flow Metab. 2005;25(2):247–56.
- 43. Tan X, Chen J, Keep RF, Xi G, Hua Y. Prx2 (peroxiredoxin 2) as a cause of hydrocephalus after intraventricular hemorrhage. Stroke. 2020;51(5):1578–86.
- Chen T, Tan X, Xia F, Hua Y, Keep RF, Xi G. Hydrocephalus induced by intraventricular peroxiredoxin-2: the role of macrophages in the choroid plexus. Biomolecules. 2021;11(5).
- Popovic M, Smiljanic K, Dobutovic B, Syrovets T, Simmet T, Isenovic ER. Thrombin and vascular inflammation. Mol Cell Biochem. 2012;359(1–2):301–13.
- Yasuda H, Sonoda A, Yamamoto M, Kawashima Y, Takishita Y, Morita A, et al. 17-beta-estradiol enhances neutrophil extracellular trap formation by interaction with estrogen membrane receptor. Arch Biochem Biophys. 2019;663:64–70.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.