

RESEARCH

Open Access

Targeted cancer immunotherapy via combination of designer bispecific antibody and novel gene-engineered T cells

Katarzyna Urbanska², Rachel C Lynn², Caitlin Stashwick², Archana Thakur³, Lawrence G Lum³
and Daniel J Powell Jr^{1,2*}

Abstract

Background: Redirection of T lymphocytes against tumor antigens can induce dramatic regression of advanced stage malignancy. The use of bispecific antibodies (BsAbs) that bind both the T-cell receptor (TCR) and a target antigen is one promising approach to T-cell redirection. However, BsAbs indiscriminately bind all CD3+ T-cells and trigger TCR activation in the absence of parallel costimulatory signals required to overcome T-cell unresponsiveness or anergy.

Methods: To address these limitations, a combination platform was designed wherein a unique BsAb referred to as frBsAb exclusively engages T-cells engineered to express a novel chimeric receptor comprised of extracellular folate receptor fused to intracellular TCR and CD28 costimulatory signaling domains in tandem; a BsAb-binding immune receptor (BsAb-IR). As a surrogate TCR, the BsAb-IR allows for concomitant TCR and costimulatory signaling exclusively in transduced T-cells upon engagement with specific frBsAbs, and can therefore redirect T-cells on command to desired antigen. Human primary T-cells were transduced with lentiviral vector and expanded for 14–18 days. BsAb-IRs were harvested and armed with frBsAbs to test for redirected cytotoxicity against CD20 positive cancer cell lines.

Results: Using frBsAbs specific for CD20 or HER2, the lytic activity of primary human T-cells expressing the BsAb-IR was specifically redirected against CD20+ leukemic cells or HER2+ epithelial cancer cells, respectively, while non-engineered T-cells were not activated. Notably, elimination of the CD28 costimulatory domain from the BsAb-IR construct significantly reduced frBsAb-redirectioned antitumor responses, confirming that frBsAbs are capable of delivering simultaneous TCR activation and costimulatory signals to BsAb-IR T-cells.

Conclusion: In summary, our results establish the proof of concept that the combination of BsAbs with optimized gene-engineered T-cells provides the opportunity to specify and augment tumor antigen-specific T-cell activation and may improve upon the early success of conventional BsAbs in cancer immunotherapy.

Keywords: Immunotherapy, Adoptive T cell transfer, Chimeric immunoreceptor, Cancer, Bispecific antibody, Trastuzumab, Rituximab

* Correspondence: poda@mail.med.upenn.edu

¹Department of Pathology and Laboratory Medicine, Abramson Cancer Center, Perelman School of Medicine, University of Pennsylvania, 3400 Civic Center Blvd, Bldg 421, Smilow CTR, Rm 08-103, Philadelphia, PA 19104-5156, USA

²Department of Obstetrics and Gynecology, Ovarian Cancer Research Center, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA, USA

Full list of author information is available at the end of the article

Introduction

Antigen-specific monoclonal antibodies (mAbs) are established as immunotherapeutic agents for the treatment of human malignancies such as non-Hodgkin lymphoma (NHL), CD30-positive lymphoma [1,2], EGFR-expressing advanced bowel cancer, metastatic colorectal carcinoma [3-6]. However, the therapeutic efficacy of tumor antigen-specific mAbs can be limited in cancer therapy due to their poor recruitment of the adaptive immune response. To address this, other strategies were employed, including the development of bispecific antibodies (BsAbs) [7,8]. While a mAb recognizes a single antigen target and closely resembles a naturally-occurring antibody, a BsAb is a synthetic construct that aligns two antigen-specific binding potentials within one molecule enabling the linking of two distinct antigens [9]. BsAbs generally couple T-cells, through a T-cell receptor (TCR)-CD3-specific antibody, with target cells, via an antigen-specific antibody. As a result, cancer cells are killed when cytotoxic T lymphocytes are engaged to antigen-expressing tumor cells and simultaneously activated by the arm of the BsAb that triggers TCR activation [10,11]. Most BsAbs rely on re-direction of cytotoxic T-cells, the most powerful effector cells of the immune system [12], where the BsAb indiscriminately engages all available TCR CD3 molecules and overrides the natural antigen-specificity of T-cells.

While TCR activation alone by BsAbs can activate T-cells, stimulation of T-cell activity is a complex, sophisticated process regulated by an assortment of molecules that provide activating, inhibiting or costimulatory signals to T-cells. One fundamental tenet of T-cell immunobiology is that sustained stimulation via TCR CD3 (signal 1) without parallel costimulatory signals, such as those provided by CD28 receptor, results in impaired T-cell activation with induction of anergy or apoptosis [13]. Accordingly, CD3-based immunotherapy with BsAbs may be improved by provision of accessory costimulation *ex vivo* or *in vivo* to elicit potent, long-lasting antitumoral effects. This can be achieved by *ex vivo* activation of cytotoxic T-cells [14,15], or by systemic administration of IL-2 cytokine [16,17]. Alternatively, technological advances have led to the development of new BsAb strategies which simultaneously trigger the activation of costimulatory receptors (e.g., CD28, 4-1BB, OX40) in conjugation with conventional BsAbs treatment [18,19]. Parallel costimulatory signaling can also be provided by combining BsAbs with an agonistic anti-CD28 mAb to mediate a synergistic effect in eliciting an antitumor response *in vitro* [20,21]. Similarly, 4-1BB-mediated costimulation at the tumor site can enhance T-cell activation mediated by a BsAb [22,23], as evidenced by increased T-cell cytokine release, activation marker expression, and proliferation. While it is increasingly evident that BsAb approaches that incorporate parallel costimulation are more effective than

conventional BsAb, the undefined optimal stoichiometry of multiple receptor engagement and the indiscriminant nature of T-cell engagement represent still represent challenges to the field.

Here, we sought to establish a proof of concept that the needs for costimulation, fixed stoichiometry and T-cell specification of conventional BsAbs can be resolved through the use of advanced T-cell engineering strategies. We and others have previously shown that human T-cells engineered to express a chimeric antigen receptor (CAR) containing an extracellular tumor antigen-specific antibody fused to intracellular TCR CD3 and costimulatory domains in tandem receive dual TCR (signal 1) and costimulatory (signal 2) upon antigen encounter that reinforce T-cell activation, proliferation and cancer killing [24-26]. Based upon this principle, we have designed a novel platform that combines the application of a BsAb with T-cells that are genetically engineered to express a unique BsAb-binding immune receptor (BsAb-IR). Here, the BsAb-IR is comprised of a portion of an extracellular folate receptor (FR; 231aa) fused to intracellular TCR and CD28 costimulatory signaling domains in tandem, and can be bound and activated by an anti-FR antibody arm of a unique BsAb that bridges FR and tumor antigen (frBsAb). Using frBsAbs of diverse antigen specificities, we show that tumor antigen-specific frBsAbs specifically bind target antigen on human tumor cells and, upon co-engagement of the BsAb-IR on engineered T-cells, delivers simultaneous TCR CD3 activation and CD28 costimulation signals in a target dependent manner, resulting in the selective augmentation of activation, proliferation and antitumor activity of BsAb-IR T-cell subset.

Materials and methods

BsAb-binding immune receptor (BsAb-IR) construction

Folate Receptor alpha (FR) DNA sequence was amplified using primers: 5'-AAAAGCCTAGGATCC-3' and 5'-A ACCGCGCTAGCAAA-3'. After amplification and the insertion of 3'-Bam-HI and 5'-Nhe-1 restriction sites, PCR product was digested with Bam-HI and NheI enzymes and ligated into pELNS, a third generation self-inactivating lentiviral expression vector, containing human CD3z or CD28-CD3z signaling endodomains, under an EF-1 α promoter. The resulting constructs were designated pELNS FBIR-zeta and pELNS FBIR-28z, respectively.

Recombinant lentivirus production

High-titer replication-defective lentiviral vectors were produced and concentrated as previously described [27,28]. Briefly, 293 T human embryonic kidney cells were transfected with pVSV-G (VSV glycoprotein expression plasmid), pRSV.REV (Rev expression plasmid), pMDLg/p.RRE (Gag/Pol expression plasmid), and pELNS transfer plasmid using Lipofectamine 2000 (Invitrogen). The viral

supernatant was harvested at 24 and 48 h post-transfection. Viral particles were concentrated and resuspended in 0.5 ml by ultracentrifugation for 2.5 h at 25,000 rpm with a Beckman SW28 rotor (Beckman Coulter, Fullerton, CA).

T-cells

Primary human CD4⁺ and CD8⁺ T-cells were isolated from healthy volunteer donors following leukapheresis by negative selection, and purchased from the Human Immunology Core at University of Pennsylvania. All specimens were collected under a University Institutional Review Board-approved protocol, and written informed consent was obtained from each donor. T-cells were cultured in complete media (RPMI 1640 supplemented with 10% heat inactivated fetal bovine serum (FBS), 100 U/ml penicillin, 100 µg/ml streptomycin sulfate, 10-mM HEPES), and stimulated with anti-CD3 and anti-CD28 mAbs coated beads (Invitrogen) as described. 24 hr after activation, T-cells were transduced with lentiviral vectors at MOI of ~5-10. Human recombinant interleukin-2 (IL-2; Novartis) was added every other day to 50 IU/ml final concentration and a 0.5×10^6 cells/ml cell density was maintained. Rested engineered T-cells were adjusted for identical transgene expression prior to functional assays. For the investigation into the effects of dose of folic acid, T-cells were cultured in RPMI 1640, 10% heat inactivated fetal bovine serum (FBS), 100 U/ml penicillin, 100 µg/ml streptomycin sulfate, 10-mM HEPES) supplemented with 40 mg/l folic acid (Sigma-Aldrich).

Cell lines

Lentivirus packaging was performed in the immortalized normal fetal renal 293 T cell line purchased from ATCC. Human cell lines used in immune based assays include CD20 positive cell lines Ramos and Daudi. 293 T cells and tumor cell lines were maintained in RPMI-1640 (Invitrogen) supplemented with 10% (v/v) heat-inactivated FBS, 2 mM L-glutamine, and 100 µg/mL penicillin and 100U/mL streptomycin. All cell lines were purchased from ATCC.

BsAb heteroconjugation

The BsAbs were engineered by heteroconjugating cross-linked mAbs. Anti-CD3 (Orthoclone OKT-3, Orthobiotec, Bridgewater, NJ, USA) or anti-FR alpha (MOV18 Ab, Enzo Life Sciences) was cross-linked with 2-iminothiolane HCl (Pierce, Rockford, IL, USA) or trastuzumab (Herceptin, Genentech, San Francisco, CA) and rituximab (Rituxan; Genentech, South San Francisco, CA, USA) was cross-linked with sulphosuccinimidyl 4-(N-maleimidomethyl) cyclohexane-1-carboxylate (Pierce), purified on PD-10 columns (BioRad Laboratories, Hercules, CA, USA),

and heteroconjugated overnight at 4°C and analyzed by non-reducing SDS-PAGE as described earlier [29,30].

Flow cytometric analysis

The following mAbs were used for phenotypic analysis: APC-Cy7 Mouse Anti-Human CD3; FITC-anti-human CD4; APC-anti-human CD8; (BD Biosciences). Tumor cell surface expression of HER2 was detected by trastuzumab antibody (Herceptin, Genentech, San Francisco, CA), CD20 by biotinylated rituximab (Rituxan; Genentech, South San Francisco, CA, USA) followed by incubation with Streptavidin-APC. BsAb-IR expression was detected by Mov18/ZEL antibody (Enzo Life Sciences) followed by anti-mouse FITC or APC conjugated antibody (LifeBioscience). The isotype controls mIgG-APC-Cy7, mIgG-FITC, mIgG-APC, mIgG-PerCpCy5, mIgG-Pacific-Blue-A, and mIgG-PE-Cy7 were from eBiosciences and anti-mouse-AF488 from Invitrogen. Flow cytometric data were analyzed by FlowJo software. For intracellular staining, cells were permeabilized using the FoxP3 staining buffer set (eBioscience), according to manufacturer's instructions and labeled with: anti-human IFNγ-PacificBlue, anti-human IFNγ-PerCpCy5, anti-human TNFα-APC, anti-human IL2-PeCy7 (BD Biosciences).

Cytokine release assays and intracellular cytokine staining

Cytokine release assays were performed by co-culture of 1×10^5 BsAb-IR T-cells with immobilized MOV18Ab (anti-FR alpha) or IgG1 (100 ng/ml). For co-culture experiments against tumor cells, 1×10^5 target cells were labeled with frBsAbs at 100 ng/ 10^6 cells for 30 min at 4°C, per well in triplicate in 96-well round bottom plates, T-cells were added into the culture at the E:T 1:1 ratio, in a final volume of 200µl of T-cell media. After 16 h, co-culture supernatants were assayed for presence of IFNγ using an ELISA Kit, according to manufacturer's instructions (Biolegend). Values represent the mean of triplicate wells. IFNγ, IL2, IL-4, TNFα and MIP-1α cytokines were measured by flow cytometry using Cytokine Bead Array, according to manufacturer's instructions (BD Biosciences). For cytokine staining cells were stimulated in 37°C for around 6 h in the presence of Golgistop (BD Biosciences) and monensin (Sigma). Cells were washed, Fc blocked, and stained with MOV18AB-FITC then permeabilized and stained with anti-IFNγ, TNFα, or isotype controls in permeabilization buffer for 30 min.

Cytotoxicity assays

^{51}Cr release assays were performed as described [31]. Target cells were labeled with frBsAbs at 100 ng per 10^6 cells for 30 min at 37°C in PBS/2%FBS. Next, antibody labeled cells were labeled with 100µCi ^{51}Cr at 37°C for 1.5 hours. Target cells were washed three times in PBS, resuspended in CM at 10^5 viable cells/mL and

100uL added per well of a 96-well V-bottom plate. Effector cells were washed twice in CM and added to wells at the given ratios. Plates were quickly centrifuged to settle cells, and incubated at 37°C in a 5% CO₂ incubator for 4 hours. The supernatants were harvested, transferred to a lumaplate (Packard) and counted using a 1450 Microbeta Liquid Scintillation Counter (Perkin-Elmer). Spontaneous ⁵¹Cr release was evaluated in target cells incubated with medium alone. Maximal ⁵¹Cr release was measured in target cells incubated with SDS at a final concentration of 2% (v/v). Percent specific lysis was calculated as (experimental - spontaneous lysis/maximal - spontaneous lysis) times 100.

Statistical analysis

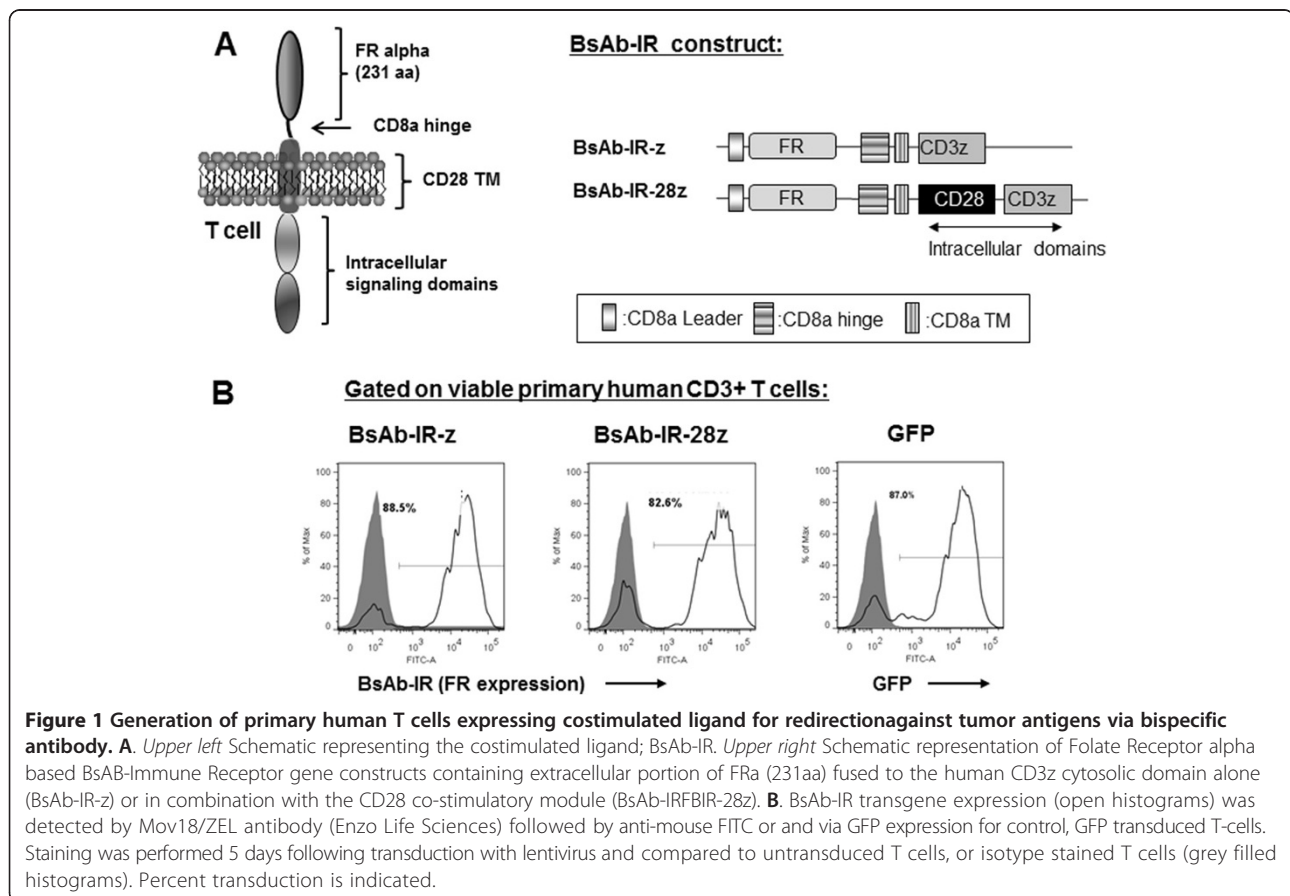
Student's *t* test was used to evaluate differences in T-cells specific cytolysis. GraphPad Prism 4.0 (GraphPad Software) was used for the statistical calculations. *P* < 0.05 was considered significant.

Results

Construction of a novel bispecific antibody platform

We sought to design and construct a combination platform that merges advantages of BsAb technology with that of gene-engineered T-cell therapy. Based upon its limited endogenous expression by human T-cells, folate

receptor alpha (FR) was selected for use in immunoreceptor construction. We constructed a folate binding immunoreceptor construct comprised of the extracellular portion of FR (1-231aa) linked to a CD8α hinge and transmembrane region, followed by a CD3z signaling moiety alone (BsAb-IR-z; Figure 1A Schematic). In order to incorporate simultaneous CD28-mediated signaling and evaluate whether a functional secondary signal potentiates T-cell activity, a second-generation BsAB-IR that includes a chimeric fusion of CD28 and CD3-z intracellular domains was constructed (BsAb-IR-28z). GFP transduced, or non-transduced T-cells were used as a specificity control in all experiments. BsAb-IR-z and BsAb-IR-28z immunoreceptor sequences were transduced into freshly isolated human primary T-cells utilizing lentiviral vectors. Five days following lentiviral transduction, BsAb-IR cell surface expression of was measured via detection of surface FR expression by flow cytometric analysis utilizing an anti-FRa antibody (MOV18) with a FITC-labeled anti-mouse secondary antibody. As shown in Figure 1B, transduction efficiencies were >80% for all constructs. For all functional assays, transduced T-cells were equilibrated to 70% by adding untransduced T-cells. In all T-cell cultures the ratio of CD4 and CD8 T cells was approximately 30% CD4 and 70% CD8 (data not shown).



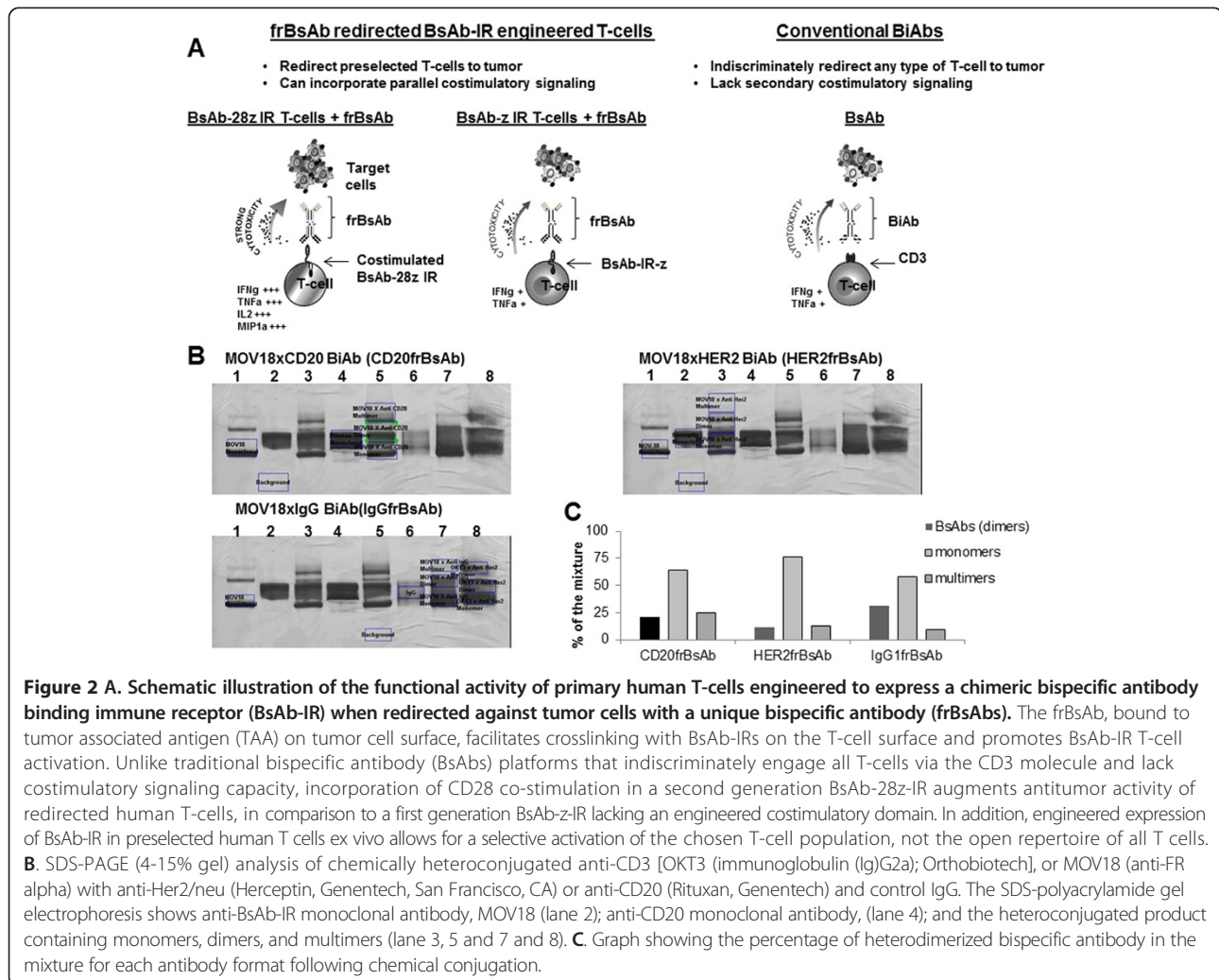
Heteroconjugation of BsAb-IR bispecific antibodies

The BsAb-IR engagers, referred to as frBsAbs, were produced by chemical heteroconjugation of two mAbs (Additional file 1: Figure S1), one that binds the extracellular FR portion of the BsAb-IR (MOV18Ab) on engineered T-cells and another that targets a chosen tumor-associated antigen (TAA) (Figure 2A). mAb against CD20 molecule (rituximab) was used for tumor targeting. Antibody recognizing HER2 (trastuzumab) and an isotype IgG1 were used as specificity controls in the construction of frBsAbs. Figure 2B shows Western Blot analysis of the cross-linked mAbs, following overnight heteroconjugation at 4°C. The products of the heteroconjugation were resolved by SDS-PAGE (4–15% gradient) and stained with Gelcode Blue (Pierce). By densitometric quantitation using Gel Doc analysis and Quantity One software (Bio-Rad), the mixture consisted of 21% of the desired conjugated heterodimers, 64% unconjugated monomers and 25% multimers for the CD20-frBsAb preparation. The HER2frBsAb mixture consisted of 11% conjugated heterodimers, 76%

monomers and 13% multimers, and the IgGfrBsAb control mixture was 31%, 58%, and 9%, respectively as represented on the graph (Figure 2C). Previous preclinical and clinical studies have shown no significant difference in binding and specificity between the purified and unpurified heteroconjugates [29]. Therefore, the unpurified heteroconjugated material was used for arming of the BsAb-IR engineered T-cells.

Primary human T-cells expressing BsAb-IR are specifically activated by immobilized anti-BsAb-IR antibody

To determine whether crosslinking of BsAb-IRs on the T-cell surface results in a specific immune-activation of engineered T-cells, BsAb-IR T-cells were cultured in plates coated with immobilized anti-BsAb-IR antibody (MOV18 Ab), control IgG1 or rituximab, or left uncoated. BsAb-IR-z expressing T-cells exclusively produced pro-inflammatory cytokine interferon-g (IFN γ) in response to anti-BsAb-IR antibody (MOV18 Ab) stimulation, but not to control antibodies; anti-CD20 (rituximab)

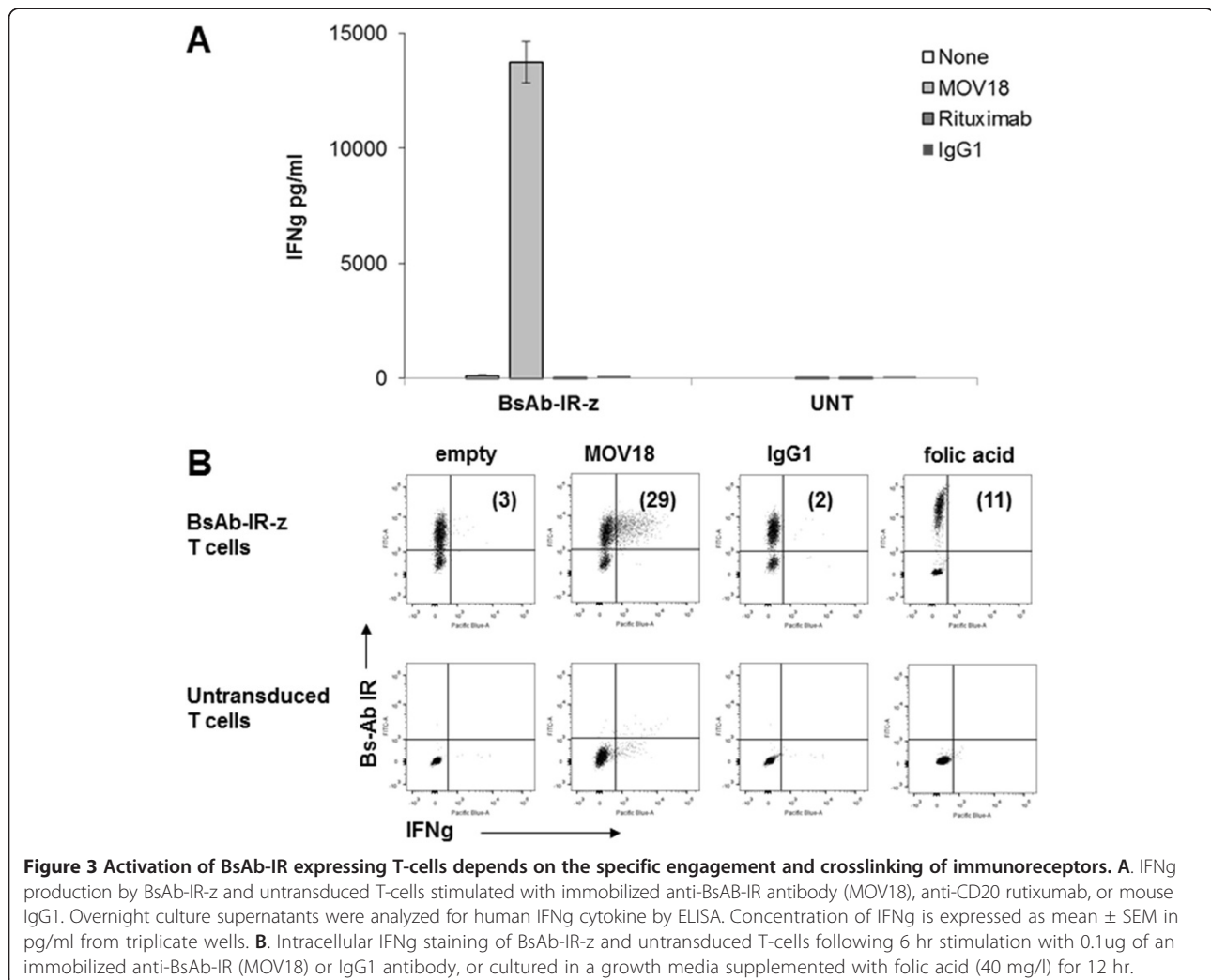


and IgG1 (Figure 3A). Importantly, untransduced T cells did not show any significant immune-reactivity. Single cell analysis by intracellular cytokine production confirmed that IFN γ production was restricted to T cells expressing BsAb-IR-z; T cells lacking BsAb-IR did not (Figure 3B). Since the BsAb-IR comprise extracellular portion FRa which binds folic acid with high affinity, we tested whether high levels of folic acid can mediate BsAb-IR T-cell activation. As shown by intracellular cytokine analysis, BsAb-IR T-cells did not produce IFN γ in the presence of supra-physiological levels of folic acid.

Costimulatory properties of BsAb-binding immune receptor (BsAb-IR)

Production of multiple proinflammatory cytokines can improve cytolytic function and contribute to *in vivo* activity [32]. Costimulation promotes T-cell activation, proliferation and effector function and therefore represents a key aspect in potentiating T-cell responses in cancer immunotherapy. The costimulatory properties of

BsAB-IR were therefore evaluated in an experimental setting where the first signal required for T-cell activation and the second costimulatory signal are triggered by the engagement of BsAb-IR-28z by frBsAb. CD28 is one of the best-characterized costimulatory molecules expressed by T-cells, providing signals critical for optimal T-cell activation, cytokine production, proliferation, and survival [33]. Having incorporated the CD28 endodomain into the BsAB-IR, we evaluated the contribution of CD28 costimulatory signals in BsAB-induced activation of human T-cells. We first tested whether cross-linking of the costimulated BsAb-IR-28z receptor induces more potent T-cell activity in comparison to activation triggered by the first generation BsAb-IR-z immunoreceptor, which lacks a costimulatory signaling domain. BsAb-IR-28z, BsAb-IR-z or untransduced T-cells were added to wells containing immobilized anti-FR antibody (MOV18) or IgG isotype antibody, and incubated for 16 hours before culture supernatants were collected and measured for type and amount of proinflammatory cytokines secreted in



response to specific stimulation. BsAb-IR-28z and BsAb-IR-z T-cells both secreted IFN γ , interleukin-2 (IL-2) and tumor necrosis factor-alpha (TNF α) cytokines when stimulated with anti-FR, but not control, antibody (Figure 4). Little to no IL-4 or IL-10 cytokines was detected suggesting a preferential Th1-cytokine response. GFP transduced control T-cells did not secrete cytokine under either antibody condition. More notably, primary human T-cells expressing a BsAb-IR containing the CD28 endodomain secreted higher levels of proinflammatory cytokines compared to T-cells with the BsAb-IR-z alone (n = 4). Similar increases in the levels of secreted IFN γ , IL-2 and TNF α cytokines occurred in the BsAb-IR-28z group. In all cases, cytokine secretion by BsAb-IR T-cells was specific, and dependent on the engagement of the BsAb-IR, demonstrated by little constitutive cytokine production in the absence of immobilized anti-FR antibody, or in the presence of the isotope control antibody. Importantly, stimulation of BsAb-IR T-cells with immobilized antibodies including: trastuzumab, rituximab and/or IgG1 did not induce secretion of proinflammatory cytokines (data not shown).

Evaluating BsAb-directed antitumor responses of BsAb-IR+ T-cells *in vitro*

Finally, we investigated the cytotoxic potential of BsAb-IR T-cells. To evaluate whether BsAb-IR-mediated activation of T-cells by cancer cell bound frBsAb facilitates specific target cell killing, BsAb-IRs T cells were assayed for their capacity to specific lyse Ramos, Daudi and AE17 cell lines. We first evaluated the potency of the combination of a CD20frBsAb and BsAb-IR T-cells. Cell surface expression of CD20 on lymphoma cell targets

was confirmed by flow cytometry. Human Burkitt's lymphoma cell lines, Ramos and Daudi, expressed high levels of CD20, with a specific mean fluorescence intensity (MFI) of 2189 and 2602 respectively (Figure 5A). The mouse mesothelioma cell line, AE17 did not express CD20 antigen on cell surface and was used as an antigen specificity control. Target cells were exposed to frBsAb bearing either an anti-CD20 (CD20frBsAb), irrelevant anti-HER2 (HER2frBsAb) or control IgG (IgGfrBsAb) antibody arm at 100 ng per 10⁶ cells for 30 minutes, labeled with ⁵¹Cr, and washed before culture with T-cells. Cytotoxicity was measured at indicated E:T ratios of 1, 3, 10 and 30 to 1 in standard 4-hour ⁵¹Cr-release assays. BsAb-IR T-cells exhibited specific redirected cytotoxic activity against CD20+ lymphoma cells when using CD20frBsAbs at E:T ratios >10:1. Cytotoxic activity was not detected in co-cultures where anti-HER2 or IgG control frBsAbs were used for redirection (Figure 5B). After 4 hours, BsAb-IR transduced T-cells specifically lysed CD20frBsAbs targeted Ramos (20% \pm 3%, 30:1) and Daudi (30% \pm 4%, 30:1) cell targets (P < 0.05), but not when redirected with control HER2 or IgG1frBsAbs (5% \pm 1.5% and 4.6% \pm 2%, respectively at 30:1; n = 3). CD20frBsAb redirected cytotoxicity of the CD20-negative AE17 cell line was not observed at any E:T ratio (Additional file 2: Figure S2).

In parallel studies, BsAb-IR-28z T-cells, but not control GFP T-cells, produced high levels of IFN-g in co-culture with Ramos cells but only when redirected with a CD20frBsAb (Figure 6A). Redirection with other frBsAbs had no impact on cytokine secretion. Culture supernatants were pooled and measured for various T-cell produced cytokines by cytokine bead array

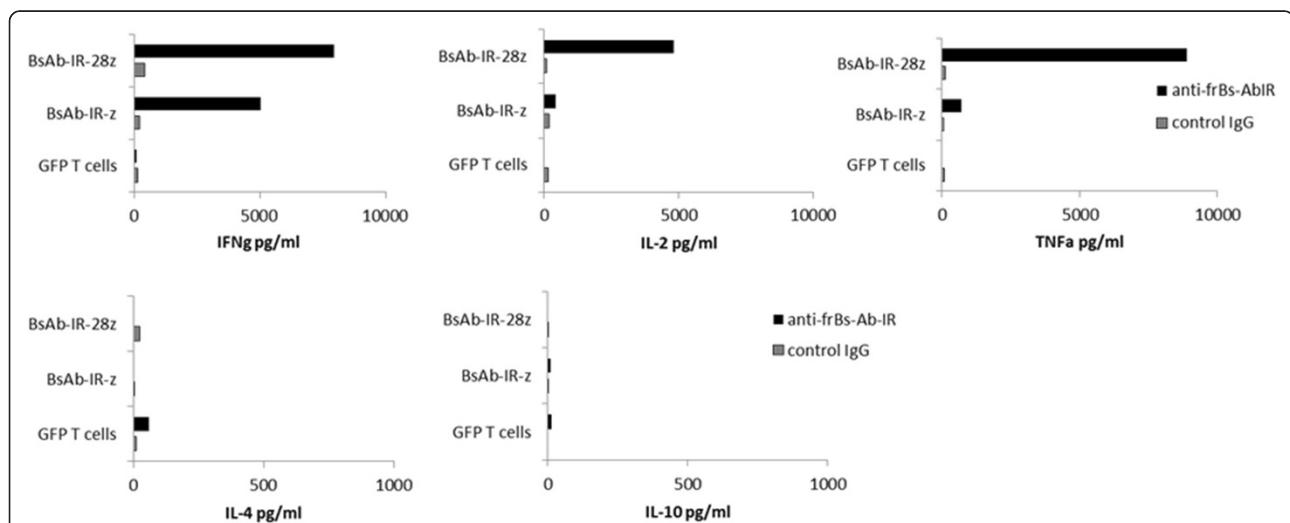


Figure 4 The amount of cytokines produced by BsAb-IR+ T-cells is dependent on the presence of the CD28 costimulatory domains within the immune receptor construct. Comparison of cytokine production by T-cells transduced with different BsAb-IR constructs. Pooled supernatant from triplicate co-cultures were measured for proinflammatory cytokine secretion by cytokine bead assay. IFN γ , IL-2, TNF α , IL-4 and IL-10 cytokine concentrations (pg/ml) were measured from cultures of BsAb-IR-z, BsAb-IR-28z and control GFP transduced T-cells stimulated with 0.1 μ g of an immobilized anti-BsAb-IR (MOV18) antibody. Representative data from 4 independent assays are shown.

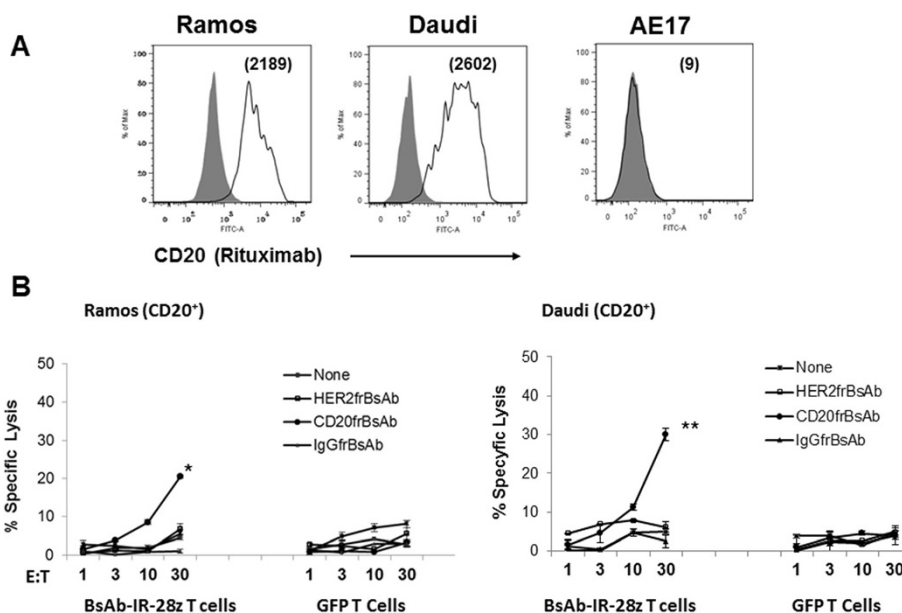


Figure 5 frBsAb-mediated activation of T cells by antigen-bound bispecific antibody (frBsAb) facilitates specific target cell killing. A. The CD20 cell surface expression (open histograms) on cancer cell lines was detected with biotinylated anti-CD20 mAb rituximab and evaluated by flow cytometry. Numbers represent mean fluorescence intensity (MFI). **B.** Antigen-specific tumor killing by CD20frBsAb redirected BsAb-IR T-cells. Primary human T-cells transduced to express BsAb-IR-28z or GFP (control) were co-cultured with Cr⁵¹-labeled Ramos, and Daudi tumor cells, pretargeted with indicated frBsAb for 4 hrs at the indicated effector to target ratio. Percent specific target cell lysis was calculated as (experimental - spontaneous release) ÷ (maximal - spontaneous release) × 100. Data represent the means ± SD for 3 different experiments. * P = 0.0082 for BsAb-IR-28z T-cells/CD20frBsAb versus IgGfrBsAb redirected BsAb-IRs against Ramos cells. ** P = 0.0166 for BsAb-IR-28z T-cells/CD20frBsAb versus IgGfrBsAb redirected BsAb-IRs against Daudi cells.

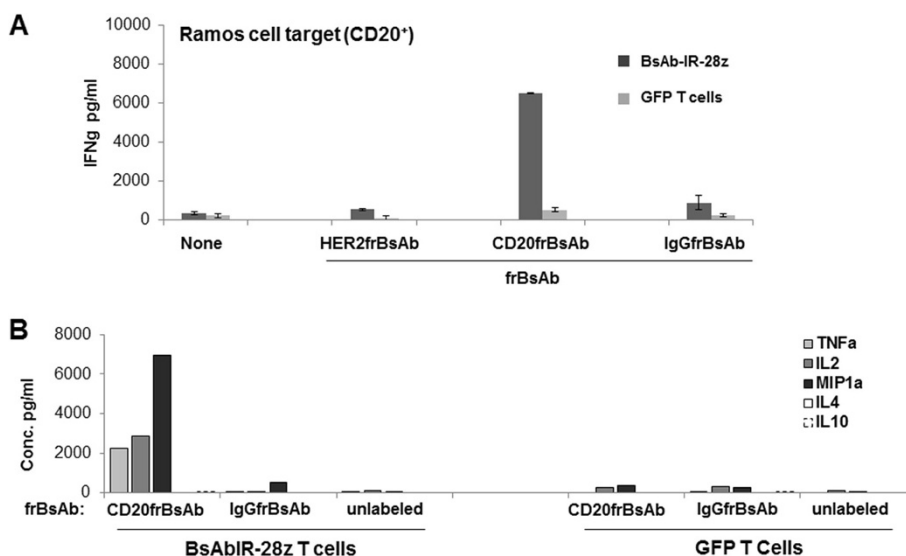


Figure 6 Anti-tumor activity of BsAb-IR-28z engineered T-cells. A. BsAb-IR-28z + T lymphocytes produce inflammatory cytokines when redirected with antigen-bound frBsAbs. CD20 redirected BsAb-IR, but not GFP (control) T-cells produce high levels of IFN γ against CD20-positive Ramos tumor cells. BsAb-IR-28z or GFP transduced T-cells were cultured overnight with an equal number of CD20frBsAb, or control frBsAbs pretargeted CD20-positive Ramos cancer cells. Overnight culture supernatants were analyzed for human IFN γ cytokine by ELISA. Concentration of IFN γ is expressed as mean ± SEM in pg/ml from triplicate wells. **B.** Cytokine bead-array analysis of cytokine production (pg/ml) by BsAb-IR-28z + T-cells or GFP+ T-cells. Supernatants from three independent cultures were harvested 16 h post-stimulation, pooled and assessed for measurable cytokine.

assay. Similar to IFN γ , IL-2, TNF α and MIP-1 α cytokines were all detected in supernatants from the co-culture of BsAb-IR-28z T-cells, but not GFP T-cells, and Ramos cells exposed to CD20frBsAb (Figure 6B). IL-4 and IL-10 secretion were not detected. Again, no substantial cytokine production by BsAb-IR-28z T-cells or GFP T-cells was observed in the presence of antigen specificity control frBsAbs.

Discussion

Tumor cells express various proteins on their surfaces that differentiate them from healthy tissues, either by levels of expression or by revealing novel antigens. Multiple antibodies against tumor associated antigens (TAAs) have been identified and exploited against different types of cancers though their mode of activity is largely independent of T-cell involvement. In order to enlist and mobilize of T-cells against cancer and maximize antibody targeted therapy, innovative methods have been designed to allow T-cells to be redirected against TAAs via TAA-specific antibodies. In one approach, relying on genetic retargeting of cytotoxic T-cells with antibody [34,35], a chimeric gene constructed that encodes an extracellular antibody specific for a TAA in the form of a single-chain variable fragment (scFv) in tandem with a gene sequence that encodes intracellular T-cell signaling domains, such as those from the CD3zeta subunit of the T cell receptor complex for T-cell activation and from costimulatory molecules to support T-cell effector function, proliferation and survival. The gene encoding this chimeric antigen receptor (CAR) is then introduced into T-cells for adoptive T-cell therapy [36]. Transfer of autologous T-cells engineered to express CARs with costimulatory domains has shown remarkable anti-tumor activity in CD19 $^{+}$ hematological malignancies [37,38]. Another strategy to target T-cells to epitopes recognized by antibodies is to develop dual specificity antibody-like molecules that possess antigen-binding specificity for both a protein expressed on the target tumor cell and a protein expressed by effector T-cells, resulting in their coupling. Commonly, those molecules, known as a Bispecific Antibodies (BsAbs), recruit and activate T-cells through binding a CD3 subunit of the T cell receptor complex for redirected lysis of tumor cells. This approach has been used to redirect T-cells toward a variety of TAAs, including CD19, Her2, and CD20 targets. Perhaps the most advanced BsAb is Blinatumomab [39], an investigational Bispecific T cell engager (BiTE $^{\circ}$) antibody construct designed to direct T-cells against target cells expressing CD19 expressing B-cell derived leukemias and lymphomas, which has received orphan drug designation from the FDA for a wide array of CD19 expressing hematological malignancies. While BsAbs have shown promising results in liquid tumors and in preclinical studies of solid tumors, their clinical activity has been limited in solid

malignancy but may be enhanced with more robust activation of effector T-cells.

Here, we sought to establish proof of concept for a unique approach that combines the advantages of BsAb therapies and gene-modified T-cell therapy. To address the issue of insufficient lymphocyte activation mediated by BsAbs, we designed, constructed and functionally characterized genetically-modified human T-cells expressing a unique co-stimulated immunoreceptor for bispecific antibody engagement, referred to as a BsAb-IR. Analogous to conventional BsAbs, our novel platform allows for the redirection of both CD8 $^{+}$ and CD4 $^{+}$ T-cells expressing a BsAb-IR against tumor targets via interaction with unique frBsAb intermediate, however, incorporation of a costimulatory signaling domain within the BsAb-IR construct results in heightened T-cell function. frBsAb-mediated activation of costimulated BsAb-IR-28z T-cells elicited enhanced production of multiple Th1-type proinflammatory cytokines, including IFN- γ , TNF- α and IL-2, in comparison to activated BsAb-IR-z T-cells that lack a costimulatory signaling endodomain. These results are consistent with the notion that BsAb-IR T-cells have the capacity to receive simultaneous TCR and costimulatory signals via crosslinking of BsAb-IRs on the T-cell surface upon frBsAb engagement. In addition to cytokine secretion, antigen-specific target cell lysis occurs when BsAb-IR T-cells are redirected by TAA-specific frBsAb against TAA-expressing tumor cells. To the best of our knowledge, this strategy of utilizing primary human T-cells engineered to express a co-stimulated immune receptor for BsAb-mediated anti-tumor activity has not been previously reported. Although the specific tumor lysis mediated by this first generation of redirected BsAb-IR T-cells was less robust than previously reported for T-cells redirected with conventional BsAb [30,40], this platform is highly tunable for optimization. One likely explanation for this modest lytic activity is the low concentration of functionalized frBsAb heterodimer used in these assays, which was less than 20% of total antibodies mixture following heteroconjugation. Accordingly, selective isolation of functional heterodimers from the antibody mixture may significantly improve in vitro lytic activity of BsAb-IR T-cells.

Activation of T-cells is triggered by signals transmitted through the T-cell receptor/signal transduction complex (TCR/CD3 signal 1). Other costimulatory receptor-ligand interactions between T-cells are needed, however, for comprehensive T-cell stimulation (signal 2). One of the best-characterized costimulatory molecules is CD28, which plays a complex role in regulating T-cell function after interaction with its ligands, B7-1 (CD80) and B7-2 (CD86) [41,42]. Costimulation through CD28 and TCR/CD3 complex leads to high-level production of IL-2 and regulates T-cell proliferation [33]. Moreover, CD28 costimulation is thought to be required not only

to activate T-cells but also to protect them from apoptosis mediated by the CD95 (APO-1/Fas) death receptor [43]. As reported, the lack or loss of CD28 ligands expression by tumors may, therefore, lead to insufficient T cell activation [44,45], where continued signaling through the T-cell receptor alone (CD3) can lead to anergy or antigen induced cell death (AICD) [46,47]. Thus, T-cells redirected and activated by a conventional CD3-signaling BsAb bound to the cancer cell surface are unlikely to obtain necessary secondary costimulatory signals, and may result in inefficient BsAb-mediated T cell activation and antitumor activity in patients. In previous studies, co-administering anti-CD28 antibodies or B7-scFv fusion proteins was shown to deliver a costimulatory signals [48,49]. However, this kind of systemic activation of the costimulatory ligand was accompanied by unexpected systemic toxicities *in vivo*, as observed after superagonistic anti-CD28 mAb TGN1412 application [50,51]. For this reason, and in order to localize the immunologic response to the tumor, others have attempted to transfer costimulatory ligands into the tumor cells. Indeed, 4-1BBL expression by tumor cells inhibited tumor outgrowth and induced strong cytotoxic T-cell response followed by long-term immunity in diverse mouse models [52]. However, the clinical applicability of such a strategy is challenged by the need for selective, efficient and safe gene transfer into the tumor cells. Thus, the generation of a novel, costimulated and specific receptor composed of a short binding moiety linked to T-cell signaling domains, CD3z and CD28, and construction of a novel bispecific antibody against a TAA and the extracellular domain of the costimulatory immune-receptor, may circumvent some of the potential major limitations associated with infusing co-stimulatory ligands together with BsAbs, specifically by permitting simultaneous activation and costimulation of a pre-selected T cell population in localized antigen-rich environment. As a proof of concept, we chose the extracellular portion of folate receptor alpha (FRa) as a BsAB partner. FRa is a 38 kDa glycosylphosphatidylinositol (GPI)-anchored protein that binds folic acid with high affinity and transports folate by receptor-mediated endocytosis [53]. FRa exhibits limited normal tissue distribution, with measurable expression restricted largely to the apical surfaces of a few epithelia, predominantly in the kidney cortex, lung and choroid plexus [54]. Restricted distribution, and importantly, the lack of FRa expression on T-cells, makes this antigen a good candidate for a creation of a unique T-cell surface CD28 costimulated engager for bispecific antibody, the frBsAb-IR, yet other binding partners are under investigation.

In these principle-establishing studies, we generated bispecific antibodies by chemical conjugation (Additional file 1: Figure S1) and demonstrated that these hybrid molecules can redirect specific cytotoxicity of BsAb-IR engineered

T-cells toward tumor cells expressing CD20 *in vitro*. These BsAbs were comprised of clinical-grade bivalent antibodies with low heterodimerism. While bivalency may increase IR crosslinking, limited heterodimers may reduce overall potency. To address this, future studies that incorporate newer BsAb formats, such as BiTE (bi-specific T cell engager) [55] and DART (dual affinity retargeting) [56], which have proven to be more potent than chemically conjugated BsAbs, are warranted.

In line with the known role of costimulatory signals in T-cell function [33], we found that BsAb-IR-28z T-cells produced at least three Th1 type cytokines, IFN- γ , IL-2 and TNF- α , as well as MIP-1 α , at levels that were significantly higher than those produced by BsAb-IR-z T-cells, following the engagement by frBsAb. This is of special importance, since the release of multiple cytokines and chemokines may recruit and activate endogenous immune cells to enhance the local antitumor response. Taken together, our *in vitro* data illustrate the high potential of a costimulated BsAb-IR approach, and supports its further optimization and evaluation in preclinical models. Our *in vitro* findings are in line with previously published data showing an enhanced BsAb-mediated antitumor response by provision of a CD137 signal in trans using a second recombinant Ab directed against a non-tumor specific cellular target [57]. In contrast to this approach, BsAb-IR has the unique advantage to simultaneously and locally activates selected population of gene engineered T-cells, and trigger the co-stimulatory signals upon cross-linkage of the BsAb-IRs to its target cell by application of one single bispecific molecule.

Conclusions

In conclusion, a strategy of using genetically modified primary human T-cells expressing a co-stimulated immune receptor and armed with bispecific molecules offers the possibility of a straightforward method for delivery of co-stimulatory signals to antigen-redirection cytotoxic T-cells. This approach extends upon our experience in bridging tumor cells with T-cells using biotinylated antibodies that engage an avidin-based immunoreceptor on the engineered T-cell surface [58], however, by comparison, the BsAb-IR described here is fully human in composition reducing the risk for transgene immunogenicity, and can be bound with antibodies with affinities higher than that obtainable using dimeric avidin binding to biotin (KD = 0.5 +/- 0.1 nM). BsAb-IR T-cells encountering antigen-bound frBsAbs undergo potent activation that is enhanced by coordinate costimulatory signaling stemming from the BsAb-IR itself. Since CD28 costimulatory signals may not only bolster augmented T-cell function but also prevent T-cell AICD, we believe that the principles established in this study will significantly enhance anti-tumor activity of targeted BsAb-based tumor immunotherapy.

Additional files

Additional file 1: Figure S1. Schematic illustration of the construction of anti-BsAb-IR × TAA bispecific antibody (frBsAb). Anti-BsAb-IR (MOV18 Ab) is cross-linked with Traut's reagent and anti-TAA is cross-linked with SulfoSMCC [sulfosuccinimidyl 4-(N-maleimidomethyl) cyclohexane-1-carboxylate; before heteroconjugation overnight under the conditions described.

Additional file 2: Figure S2. Tumor killing by redirected BsAb-IR T-cells is antigen-specific. CD20frBsAb redirected cytotoxicity of the CD20-negative AE17 cell line by BsAb-IR T-cells was not observed. Primary human T-cells transduced to express BsAb-IR-28z or GFP (control) were co-cultured with Cr⁵¹-labeled CD20-negative AE17, mouse mesothelioma cell line, pretargeted with indicated frBsAb for 4 hrs at the indicated effector to target ratio. Percent specific target cell lysis was calculated as (experimental - spontaneous release) ÷ (maximal - spontaneous release) × 100. Data represent the means ± SD for 3 different experiments.

Abbreviations

TCR: T-cell Receptor; IFN γ : Interferon gamma; IL2: Interleukin 2; GM-CSF: Granulocyte-macrophage colony-stimulating protein; TNF α : Tumor necrosis factor alpha; MIP-1a: Macrophage inhibitory protein; BsAb-IR, TAA: Tumor Associated Antigen; CAR: Chimeric Antigen Receptor; BsAb: Bispecific antibody.

Competing interests

DJP, KU, RL, CS, and AT have no competing interests. LGL is a co-founder of Transtarget Inc.

Authors' contributions

KU participated in the design of the study, performed experiments, analyzed data and statistics, and preparation of the manuscript. RL, CS participated in immunologic assays. AT and LGL participated in antibody heteroconjugation. DJP participated in the design of the study and in manuscript preparation. All authors read and approved the final manuscript.

Acknowledgements

This work was supported by grants from the NIH (RO1-CA168900) and the Joint Fox Chase Cancer Center and University of Pennsylvania Ovarian Cancer SPORE (P50 CA083638).

Author details

¹Department of Pathology and Laboratory Medicine, Abramson Cancer Center, Perelman School of Medicine, University of Pennsylvania, 3400 Civic Center Blvd, Bldg 421, Smilow CTR, Rm 08-103, Philadelphia, PA 19104-5156, USA. ²Department of Obstetrics and Gynecology, Ovarian Cancer Research Center, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA, USA. ³Department of Oncology, Wayne State University, Detroit, MI, USA.

Received: 23 September 2014 Accepted: 29 November 2014

Published online: 13 December 2014

References

1. Younes A, Bartlett NL, Leonard JP, Kennedy DA, Lynch CM, Sievers EL, Forero-Torres A: **Brentuximab vedotin (SGN-35) for relapsed CD30-positive lymphomas.** *N Engl J Med* 2010, **363**:1812–1821.
2. Ansell SM, Horwitz SM, Engert A, Khan KD, Lin T, Strair R, Keler T, Graziano R, Blanset D, Yellin M, Fischkoff S, Assad A, Borchmann P: **Phase I/II study of an anti-CD30 monoclonal antibody (MDX-060) in Hodgkin's lymphoma and anaplastic large-cell lymphoma.** *J Clin Oncol* 2007, **25**:2764–2769.
3. Van Cutsem E, Peeters M, Siena S, Humblet Y, Hendlisz A, Neyns B, Canon JL, Van Laethem JL, Maurel J, Richardson G, Wolf M, Amado RG: **Open-label phase III trial of panitumumab plus best supportive care compared with best supportive care alone in patients with chemotherapy-refractory metastatic colorectal cancer.** *J Clin Oncol* 2007, **25**:1658–1664.
4. Hurwitz H, Tebbutt NC, Kabbinavar F, Giantonio BJ, Guan ZZ, Mitchell L, Waterkamp D, Taberner J: **Efficacy and safety of Bevacizumab in metastatic colorectal cancer: pooled analysis from seven randomized controlled trials.** *Oncologist* 2013, **18**(9):1004–1012.
5. Ducreux M, Adenis A, Pignon JP, Francois E, Chaffert B, Ichante JL, Boucher E, Ychou M, Pierga JY, Montoto-Grillot C, Conroy T: **Efficacy and safety of bevacizumab-based combination regimens in patients with previously untreated metastatic colorectal cancer: final results from a randomised phase II study of bevacizumab plus 5-fluorouracil, leucovorin plus irinotecan versus bevacizumab plus capecitabine plus irinotecan (FNCLCC ACCORD 13/0503 study).** *Eur J Cancer* 2013, **49**:1236–1245.
6. Slamon DJ, Leyland-Jones B, Shak S, Fuchs H, Paton V, Bajamonde A, Fleming T, Eiermann W, Wolter J, Pegram M, Baselga J, Norton L: **Use of chemotherapy plus a monoclonal antibody against HER2 for metastatic breast cancer that overexpresses HER2.** *N Engl J Med* 2001, **344**:783–792.
7. Weiner GJ, Hillstrom JR: **Bispecific anti-idiotypic/anti-CD3 antibody therapy of murine B cell lymphoma.** *J Immunol* 1991, **147**:4035–4044.
8. Renner C, Jung W, Sahin U, Denfeld R, Pohl C, Trummer L, Hartmann F, Diehl V, van Lier R, Pfreundschuh M: **Cure of xenografted human tumors by bispecific monoclonal antibodies and human T cells.** *Science* 1994, **264**:833–835.
9. Segal DM, Weiner GJ, Weiner LM: **Bispecific antibodies in cancer therapy.** *Curr Opin Immunol* 1999, **11**:558–562.
10. Jain P, O'Brien S: **Anti-CD20 monoclonal antibodies in chronic lymphocytic leukemia.** *Expert Opin Biol Ther* 2013, **13**:169–182.
11. Younes A, Bartlett NL, Leonard JP, Kennedy DA, Lynch CM, Sievers EL, Forero-Torres A: **Brentuximab vedotin (SGN-35) for relapsed CD30-positive lymphomas.** *N Engl J Med* 2010, **363**:1812–1821.
12. Kontermann RE: **Recombinant bispecific antibodies for cancer therapy.** *Acta Pharmacol Sin* 2005, **26**:1–9.
13. Appleman LJ, Boussiotis VA: **T cell energy and costimulation.** *Immunol Rev* 2003, **192**:161–180.
14. Nitta T, Sato K, Yagita H, Okumura K, Ishii S: **Preliminary trial of specific targeting therapy against malignant glioma.** *Lancet* 1990, **335**:368–371.
15. Canevari S, Stoter G, Arienti F, Bolis G, Colnaghi MI, Di Re EM, Eggermont AM, Goey SH, Gratama JW, Lamers CH: **Regression of advanced ovarian carcinoma by intraperitoneal treatment with autologous T lymphocytes retargeted by a bispecific monoclonal antibody.** *J Natl Cancer Inst* 1995, **87**:1463–1469.
16. Kroesen BJ, Buter J, Sleijfer DT, Janssen RA, van der Graaf WT, The TH, de Leij L, Mulder NH: **Phase I study of intravenously applied bispecific antibody in renal cell cancer patients receiving subcutaneous interleukin 2.** *Br J Cancer* 1994, **70**:652–661.
17. Gillies SD, Reilly EB, Lo KM, Reisfeld RA: **Antibody-targeted interleukin 2 stimulates T-cell killing of autologous tumor cells.** *Proc Natl Acad Sci U S A* 1992, **89**:1428–1432.
18. Bauer S, Renner C, Juwana JP, Held G, Ohnesorge S, Gerlach K, Pfreundschuh M: **Immunotherapy of human tumors with T-cell-activating bispecific antibodies: stimulation of cytotoxic pathways in vivo.** *Cancer Res* 1999, **59**:1961–1965.
19. Guo H, Jiang W, Liu W, Gao Y, Yang M, Zhou Y, Wang J, Qi J, Cheng X, Zhu Z, Yang C, Xiong D: **Extracellular domain of 4-1BBL enhanced the antitumor efficacy of peripheral blood lymphocytes mediated by anti-CD3 × anti-Pgp bispecific antibody against human multidrug-resistant leukemia.** *Cell Immunol* 2008, **251**:102–108.
20. Otz T, Grosse-Hovest L, Hofmann M, Rammensee HG, Jung G: **A bispecific single-chain antibody that mediates target cell-restricted, supra-agonistic CD28 stimulation and killing of lymphoma cells.** *Leukemia* 2009, **23**:71–77.
21. Cochlovius B, Kipriyanov SM, Stassar MJ, Schuhmacher J, Benner A, Moldenhauer G, Little M: **Cure of Burkitt's lymphoma in severe combined immunodeficiency mice by T cells, tetraivalent CD3 × CD19 tandem antibody, and CD28 costimulation.** *Cancer Res* 2000, **60**:4336–4341.
22. Muller D, Frey K, Kontermann RE: **A novel antibody-4-1BBL fusion protein for targeted costimulation in cancer immunotherapy.** *J Immunother* 2008, **31**:714–722.
23. Liu R, Jiang W, Yang M, Guo H, Zhang Y, Wang J, Zhu H, Shi R, Fan D, Yang C, Zhu Z, Xie Y, Xiong D: **Efficient inhibition of human B-cell lymphoma in SCID mice by synergistic antitumor effect of human 4-1BB ligand/anti-CD20 fusion proteins and anti-CD3/anti-CD20 diabodies.** *J Immunother* 2010, **33**:500–509.
24. Song DG, Ye Q, Poussin M, Harms GM, Figini M, Powell DJ Jr: **CD27 costimulation augments the survival and antitumor activity of redirected human T cells in vivo.** *Blood* 2012, **119**:696–706.
25. Carpenito C, Milone MC, Hassan R, Simonet JC, Lakhai M, Suhoski MM, Varela-Rohena A, Haines KM, Heitjan DF, Albelda SM, Carroll RG, Riley JL,

- Pastan I, June CH: Control of large, established tumor xenografts with genetically retargeted human T cells containing CD28 and CD137 domains. *Proc Natl Acad Sci U S A* 2009, **106**:3360–3365.
26. Lanitis E, Poussin M, Hagemann IS, Coukos G, Sandaltzopoulos R, Scholler N, Powell DJ Jr: Redirected antitumor activity of primary human lymphocytes transduced with a fully human anti-mesothelin chimeric receptor. *Mol Ther* 2012, **20**:633–643.
27. Song DG, Ye Q, Carpenito C, Poussin M, Wang LP, Ji C, Figini M, June CH, Coukos G, Powell DJ Jr: In vivo persistence, tumor localization, and antitumor activity of CAR-engineered T cells is enhanced by costimulatory signaling through CD137 (4-1BB). *Cancer Res* 2011, **71**:4617–4627.
28. Perez EE, Riley JL, Carroll RG, von Laer D, June CH: Suppression of HIV-1 infection in primary CD4 T cells transduced with a self-inactivating lentiviral vector encoding a membrane expressed gp41-derived fusion inhibitor. *Clin Immunol* 2005, **115**:26–32.
29. Sen M, Wankowski DM, Garlie NK, Siebenlist RE, Van Epps D, LeFever AV, Lum LG: Use of anti-CD3 x anti-HER2/neu bispecific antibody for redirecting cytotoxicity of activated T cells toward HER2/neu + tumors. *J Hematother Stem Cell Res* 2001, **10**:247–260.
30. Gall JM, Davol PA, Grabert RC, Deaver M, Lum LG: T cells armed with anti-CD3 x anti-CD20 bispecific antibody enhance killing of CD20+ malignant B cells and bypass complement-mediated rituximab resistance in vitro. *Exp Hematol* 2005, **33**:452–459.
31. Johnson LA, Heemskerk B, Powell DJ Jr, Cohen CJ, Morgan RA, Dudley ME, Robbins PF, Rosenberg SA: Gene transfer of tumor-reactive TCR confers both high avidity and tumor reactivity to nonreactive peripheral blood mononuclear cells and tumor-infiltrating lymphocytes. *J Immunol* 2006, **177**:6548–6559.
32. Kannanganat S, Ibegbu C, Chennareddi L, Robinson HL, Amara RR: Multiple-cytokine-producing antiviral CD4 T cells are functionally superior to single-cytokine-producing cells. *J Virol* 2007, **81**:8468–8476.
33. Appleman LJ, Berezovskaya A, Grass I, Boussiotis VA: CD28 costimulation mediates T cell expansion via IL-2-independent and IL-2-dependent regulation of cell cycle progression. *J Immunol* 2000, **164**:144–151.
34. Scholler J, Brady TL, Binder-Scholl G, Hwang WT, Plesa G, Hege KM, Vogel AN, Kalos M, Riley JL, Deeks SG, Mitsuyasu RT, Bernstein WB, Aronson NE, Levine BL, Bushman FD, June CH: Decade-long safety and function of retroviral-modified chimeric antigen receptor T cells. *Sci Transl Med* 2012, **4**:132ra153.
35. Rosenberg SA, Dudley ME: Cancer regression in patients with metastatic melanoma after the transfer of autologous antitumor lymphocytes. *Proc Natl Acad Sci U S A* 2004, **101**(Suppl 2):14639–14645.
36. Kandalaft LE, Powell DJ Jr, Coukos G: A phase I clinical trial of adoptive transfer of folate receptor-alpha redirected autologous T cells for recurrent ovarian cancer. *J Transl Med* 2012, **10**:157.
37. Porter DL, Levine BL, Kalos M, Bagg A, June CH: Chimeric antigen receptor-modified T cells in chronic lymphoid leukemia. *N Engl J Med* 2011, **365**:725–733.
38. Kalos M, Levine BL, Porter DL, Katz S, Grupp SA, Bagg A, June CH: T cells with chimeric antigen receptors have potent antitumor effects and can establish memory in patients with advanced leukemia. *Sci Transl Med* 2011, **3**:95ra73.
39. Klinger M, Brandl C, Zugmaier G, Hijazi Y, Bargou RC, Topp MS, Gokbuget N, Neumann S, Goebeler M, Viardot A, Stelljes M, Bruggemann M, Hoelzer D, Degenhard E, Nagorsen D, Baeuerle PA, Wolf A, Kufer P: Immunopharmacologic response of patients with B-lineage acute lymphoblastic leukemia to continuous infusion of T cell-engaging CD19/CD3-bispecific BiTE antibody blinatumomab. *Blood* 2012, **119**:6226–6233.
40. Davol PA, Smith JA, Kouttab N, Elfenbein GJ, Lum LG: Anti-CD3 x anti-HER2 bispecific antibody effectively redirects armed T cells to inhibit tumor development and growth in hormone-refractory prostate cancer-bearing severe combined immunodeficient beige mice. *Clin Prostate Cancer* 2004, **3**:112–121.
41. Dong H, Zhu G, Tamada K, Chen L: B7-H1, a third member of the B7 family, co-stimulates T-cell proliferation and interleukin-10 secretion. *Nat Med* 1999, **5**:1365–1369.
42. Yao S, Zhu Y, Zhu G, Augustine M, Zheng L, Goode DJ, Broadwater M, Ruff W, Flies S, Xu H, Flies D, Luo L, Wang S, Chen L: B7-h2 is a costimulatory ligand for CD28 in human. *Immunity* 2011, **34**:729–740.
43. McLeod JD, Walker LS, Patel YI, Boulougouris G, Sansom DM: Activation of human T cells with superantigen (staphylococcal enterotoxin B) and CD28 confers resistance to apoptosis via CD95. *J Immunol* 1998, **160**:2072–2079.
44. Stopeck AT, Gessner A, Miller TP, Hersh EM, Johnson CS, Cui H, Frutiger Y, Grogan TM: Loss of B7.2 (CD86) and intracellular adhesion molecule 1 (CD54) expression is associated with decreased tumor-infiltrating T lymphocytes in diffuse B-cell large-cell lymphoma. *Clin Cancer Res* 2000, **6**:3904–3909.
45. Seliger B, Marincola FM, Ferrone S, Abken H: The complex role of B7 molecules in tumor immunology. *Trends Mol Med* 2008, **14**:550–559.
46. Chai JG, Lechler RI: Immobilized anti-CD3 mAb induces anergy in murine naive and memory CD4+ T cells in vitro. *Int Immunol* 1997, **9**:935–944.
47. Yamamoto T, Hattori M, Yoshida T: Induction of T-cell activation or anergy determined by the combination of intensity and duration of T-cell receptor stimulation, and sequential induction in an individual cell. *Immunology* 2007, **121**:383–391.
48. Hunig T: Manipulation of regulatory T-cell number and function with CD28-specific monoclonal antibodies. *Adv Immunol* 2007, **95**:111–148.
49. Holliger P, Manzke O, Span M, Hawkins R, Fleischmann B, Qinghua L, Wolf J, Diehl V, Cochet O, Winter G, Bohlen H: Carcinoembryonic antigen (CEA)-specific T-cell activation in colon carcinoma induced by anti-CD3 x anti-CEA bispecific diabodies and B7 x anti-CEA bispecific fusion proteins. *Cancer Res* 1999, **59**:2909–2916.
50. Suntharalingam G, Perry MR, Ward S, Brett SJ, Castello-Cortes A, Brunner MD, Panoskaltsis N: Cytokine storm in a phase 1 trial of the anti-CD28 monoclonal antibody TGN1412. *N Engl J Med* 2006, **355**:1018–1028.
51. Hanke T: Lessons from TGN1412. *Lancet* 2006, **368**:1569–1570. author reply 1570.
52. Melero I, Bach N, Hellstrom KE, Aruffo A, Mittler RS, Chen L: Amplification of tumor immunity by gene transfer of the co-stimulatory 4-1BB ligand: synergy with the CD28 co-stimulatory pathway. *Eur J Immunol* 1998, **28**:1116–1121.
53. Shen F, Zheng X, Wang J, Ratnam M: Identification of amino acid residues that determine the differential ligand specificities of folate receptors alpha and beta. *Biochemistry* 1997, **36**:6157–6163.
54. Weitman SD, Lark RH, Coney LR, Fort DW, Frasca V, Zurawski VR Jr, Kamen BA: Distribution of the folate receptor GP38 in normal and malignant cell lines and tissues. *Cancer Res* 1992, **52**:3396–3401.
55. Baeuerle PA, Reinhardt C: Bispecific T-cell engaging antibodies for cancer therapy. *Cancer Res* 2009, **69**:4941–4944.
56. Moore PA, Zhang W, Rainey GJ, Burke S, Li H, Huang L, Gorlatov S, Veri MC, Aggarwal S, Yang Y, Shah K, Jin L, Zhang S, He L, Zhang T, Ciccarone V, Koenig S, Bonvini E, Johnson S: Application of dual affinity retargeting molecules to achieve optimal redirected T-cell killing of B-cell lymphoma. *Blood* 2011, **117**:4542–4551.
57. Hornig N, Kermer V, Frey K, Diebold P, Kontermann RE, Muller D: Combination of a bispecific antibody and costimulatory antibody-ligand fusion proteins for targeted cancer immunotherapy. *J Immunother* 2012, **35**:418–429.
58. Urbanska K, Lanitis E, Poussin M, Lynn RC, Gavin BP, Kelderman S, Yu J, Scholler N, Powell DJ Jr: A universal strategy for adoptive immunotherapy of cancer through use of a novel T-cell antigen receptor. *Cancer Res* 2012, **72**:1844–1852.

doi:10.1186/s12967-014-0347-2

Cite this article as: Urbanska et al.: Targeted cancer immunotherapy via combination of designer bispecific antibody and novel gene-engineered T cells. *Journal of Translational Medicine* 2014 **12**:347.