

RESEARCH ARTICLE

Open Access



RDWN6^{XB}, a major quantitative trait locus positively enhances root system architecture under nitrogen deficiency in rice

Galal Bakr Anis^{1,2}, Yingxin Zhang¹, Anowerul Islam¹, Yue Zhang¹, Yongrun Cao¹, Weixun Wu¹, Liyong Cao^{1*} and Shihua Cheng^{1*}

Abstract

Background: Nitrogen (N) is a major input cost in rice production, in addition to causing severe pollution to agricultural and ecological environments. Root dry weight has been considered the most important component related to crop yields than the other root traits. Therefore, development of rice varieties/lines with low input of N fertilizer and higher root traits are essential for sustainable rice production.

Results: In this context, a main effect quantitative trait locus *qRDWN6^{XB}* on the long arm of chromosome 6 which positively confers tolerance to N deficiency in the *Indica* rice variety XieqingzaoB, was identified using a chromosomal segment substitution line (CSSL) population. *qRDWN6^{XB}* was determined to be located near marker InD90 on chromosome 6 based on association analysis of phenotype data from three N levels and 120 polymorphic molecular markers. The target chromosomal segment substitution line CSSL45, which has the higher root dry weight (RDW) than *indica* cultivar Zhonghui9308 and carry *qRDWN6^{XB}*, was selected for further study. A BC₅F_{2:3} population derived from a cross between CSSL45 and Zhonghui9308 was constructed. To fine-map *qRDWN6^{XB}*, we used the homozygous recombinant plants and ultimately this locus was narrowed to a 52.3-kb between markers ND-4 and RM19771, which contains nine candidate genes in this region. One of these genes, *LOC_Os06g15910* as a potassium transporter was considered a strong candidate gene for the *RDWN6^{XB}* locus.

Conclusions: The identification of *qRDWN6^{XB}* provides a new genetic resource for breeding rice varieties and a starting point to improve grain yield despite the decreased input of N fertilizers. The newly developed and tightly linked InDel marker ND-4 will be useful to improve the root system architecture under low N by marker-assisted selection (MAS) in rice breeding programs.

Keywords: Rice, *qRDWN6^{XB}*, Quantitative trait locus, Nitrogen deficiency tolerance

Background

Rice (*Oryza sativa* L.) is one of the most important three major cereal crops for billions of people, most of who live in rural and urban areas of tropical and subtropical Asian countries [1, 2]. Therefore, rice production is playing an important role in ensuring food security and poverty alleviation in rice-eating countries in addition

farmers' incomes for the majority of the world population [3]. According to previous estimates, the world's population will reach 8 billion by 2030 [4], and 9 billion by 2050 [5]. The dramatically increasing population will be exhausted the sustainable agricultural production system, therefore rice production must be increased by 50% in order to meet the growing demand and food needs in the future [4].

Nitrogen (N) is the most essential nutrient plant need in crop developments and high quantity. It is often the most yield-limiting nutrient in rice crop production in

* Correspondence: caoliyong@caas.cn; chengshihua@caas.cn

¹Key Laboratory for Zhejiang Super Rice Research and State Key Laboratory of Rice Biology, China National Rice Research Institute, Hangzhou 310006, China

Full list of author information is available at the end of the article



many countries [6, 7]. Nitrogen improves rice grain yield and grain quality by improving; root system, panicle numbers, leaf area, filled grains numbers, thousand grain weight, grain formation, and protein synthesis [8]. Of the total rice production costs, nitrogen fertilizers are major input cost and cause environmental pollution in case of excessive quantities used [9]. The problems and negative environmental effects of excessive nitrogen (N) quantities are well documented [10–12]. Nitrogen (N) pollution from fertilizers is the biggest environmental disaster affecting human health in multiple ways. Therefore, it is necessary to develop new rice lines/varieties with improved N use efficiency and high responses to N deficiency stress for rice breeding programs and sustainable rice production [13, 14]. Nitrogen deficiency conditions lead to a reduction in many agronomic traits and, as a result, growth rate, grain and biomass yield decrease [15]. Additionally, most of the physiological processes, nitrogen metabolism, carbon metabolism, hormone metabolism and photosynthetic, are negatively affected [16]. Thus, developing rice genotypes with high tolerance to N deficiency is a highly desired characteristic for sustainable rice production [17].

Root traits are the important nitrogen-deficiency tolerance measurements, water uptake and as the major interface between rice plant and various biotic and abiotic stresses in natural soil [18]. Numerous studies have measured root traits as the ratio of the trait values under low nitrogen to those under normal, with a criterion for selecting genotypes for the tolerance of low nitrogen. Root number, root length, root volume, root thickness, dry weight of roots, dry weight of the shoot and the total dry weight, are important traits for studying of nitrogen-deficiency tolerance in rice breeding programs. Studying like these traits is difficult to handle in soil environment. To meet these limitations, many of studies grow rice seedlings in hydroponic system using diverse mapping populations to identify the QTLs responses to nitrogen deficiency tolerance in rice [19–21]. Examination of N deficiency tolerance in rice using hydroponic systems as controlled conditions is an alternative approach.

Several quantitative trait loci (QTLs) associated with nitrogen-deficiency tolerance in rice were detected in the field and greenhouse experiments using diverse mapped populations, e.g. in Chromosomal Segment Substitution Lines [22–24], Recombinant Inbred Lines [19, 25], Introgression Lines [26] and Backcross Recombinant Lines [27]. Feng et al. [28] detected seven QTLs associated with N-deficiency tolerance on chromosomes 1, 2, 3 and 8 at seedling stage. Zhao et al. [29] mapped 28 QTLs under two N levels and 16 QTLs of their relative traits for seedling traits related to low nitrogen tolerance in rice. Main effect QTLs on chromosome 3 were mapped using

a DH population under three nitrogen conditions [30]. Obara et al. [20] mapped *qRL6.1*, a major QTL on chromosome 6 for root elongation of rice seedling under different N levels. Many genes associated with utilization of nitrogen (N) for root seedling traits have been identified in rice [31–33]. Studying and understanding the mechanisms of rice N utilization at a molecular level may help to improve rice varieties for N deficiency tolerance.

The super hybrid rice ‘Xieyou9308’ is one of the earliest hybrids occupied greater areas in China [34]. Building on our previous study, some QTLs were identified under N deficiency using a set of chromosomal segment substitution lines (CSSLs) derived from XieqingzaoB/Zhonghui9308 as two parents of the hybrid ‘Xieyou9308’. Here in this study, the objectives were to validate and fine map a major QTL, *qRDWN6^{XB}*, for root dry weight under N deficiency using $F_{2:3}$ populations derived from a cross between the target CSSL45 and genetic background parent Zhonghui9308.

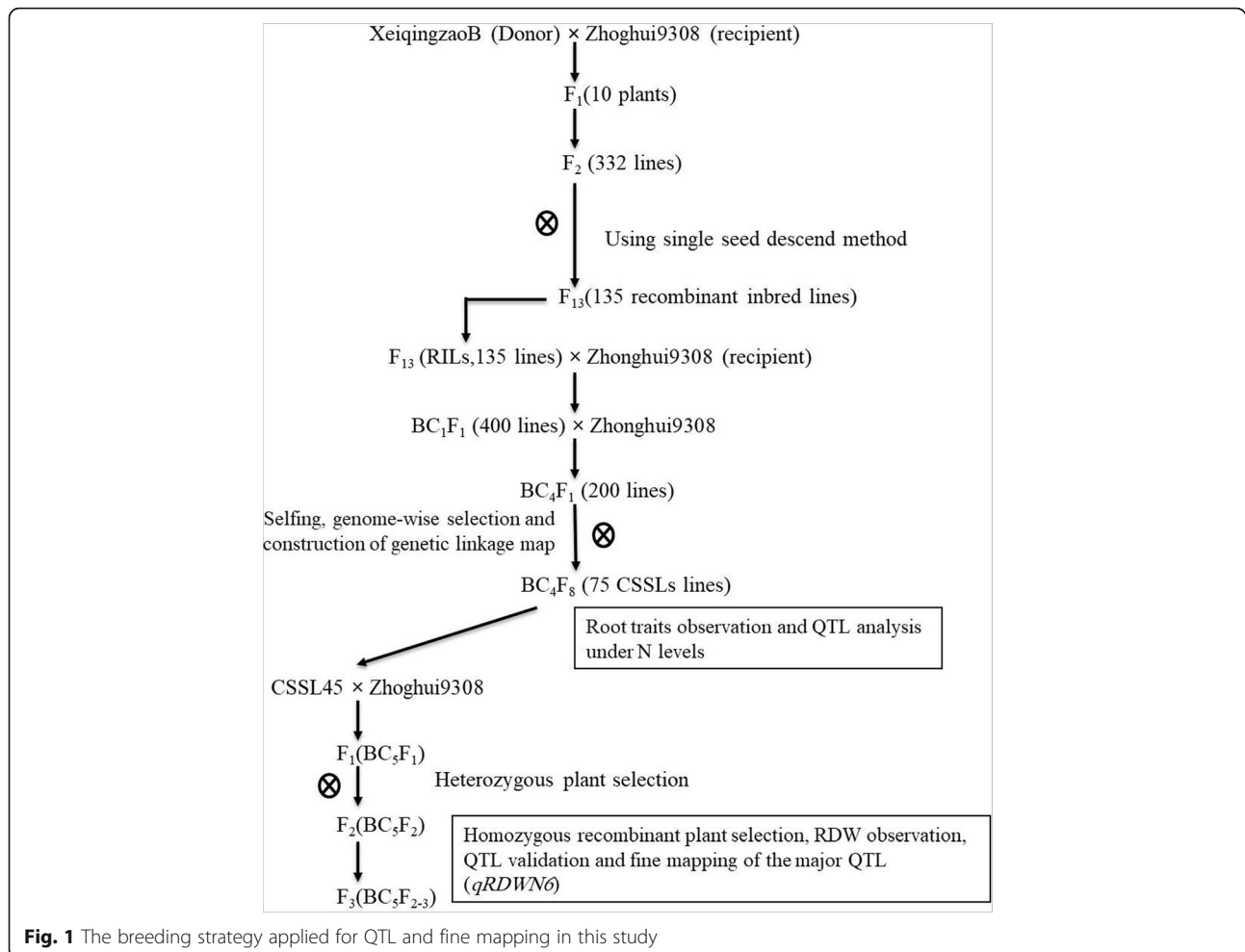
Methods

Rice materials and fine mapping population

In our previous study, a set of 75 Chromosome Segment Substitution Lines (CSSLs) were evaluated for root dry weight and other related root traits under N deficiency at the seedling. The donor parent XieqingzaoB (XQZB) and recipient parent Zhonghui9308 (ZH9308) as parental lines of this population exhibited high and low of RDW and other related root traits, respectively, under N deficiency [24]. For this study, one line of CSSLs population, namely CSSL45, which showed high root dry weight (RDW) values compared with Zhonghui9308 under N⁻ deficiency and harboring the *qRDWN6^{XB}* a root dry weight conditioning locus, was crossed with the genetic background parent Zhonghui9308, and a total of 2200 $F_{2:3}$ (BC₅F_{2:3}) seedlings obtained from the cross were evaluated to perform fine mapping of the *qRDWN6^{XB}* locus under low nitrogen on chromosome 6 as reported by our previous study [24]. Therefore, XieqingzaoB, Zhonghui9308, CSSL45 and $F_{2:3}$ (BC₅F_{2:3}) population derived from a cross between CSSL45/Zhonghui9308 were used in this study. The development scheme of rice populations used here is shown in Fig. 1.

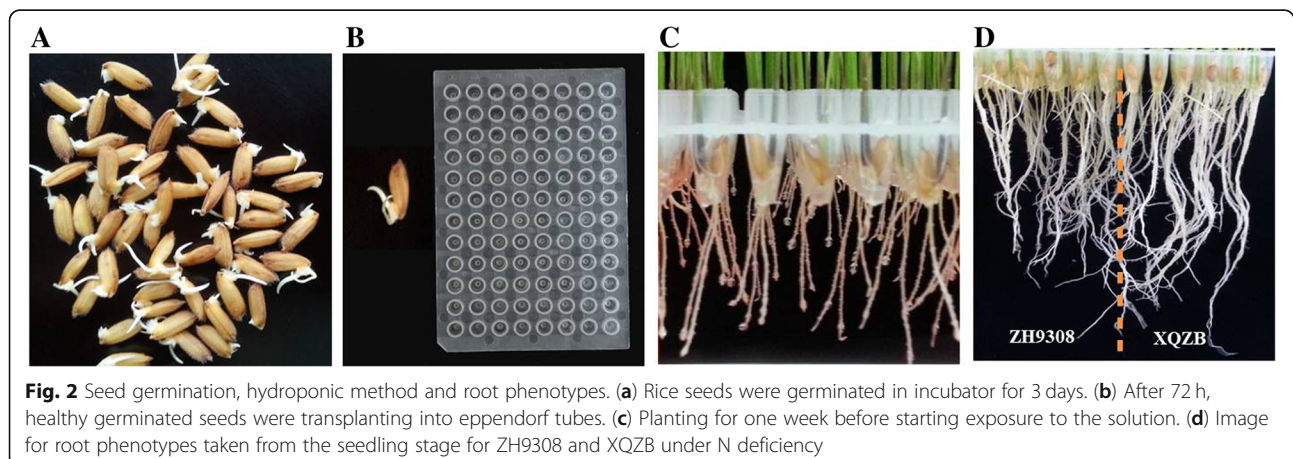
Growth conditions in hydroponic culture

Seeds of XieqingzaoB, Zhonghui9308, CSSL45 and $F_{2:3}$ (BC₅F_{2:3}) populations were soaked in sterilized distilled water for 2 days at 30 °C and then incubated at 29 °C for 24 h (Fig. 2a). The healthy germinated seeds were transplanting into Eppendorf tubes for 1 week before starting exposure to the solution (Fig. 2b, c). The experiment was conducted at China National Rice Research Institute (CNRRI) at Hangzhou, China, during 2017 and 2018. For confirming and comparison of root dry weight and other related root traits of XieqingzaoB, Zhonghui9308 and



CSSL45, fifteen seedlings of these parental lines were evaluated under both (N^+) normal and (N^-) deficiency conditions. Yoshida rice nutrient solution was used with some minor modifications [35]. For nitrogen (N^-) deficiency condition, the concentration of $(NH_4)_2SO_4$ decreased to 0.42 (mg/l). The nutrient solution was prepared in distilled

water and renewed every 5 days. The plants were grown and screened in the chamber with a $30 \pm 3/21 \pm 2^\circ C$ day/night temperature and 70% relative humidity. The $F_{2,3}(BC_5F_{2,3})$ plants were evaluated only under nitrogen deficiency (N^-) treatment, and root dry weight under nitrogen deficiency (N^-) was measured as RDWN. The root



dry weight and other related traits were recorded after 5 weeks in a chamber (Fig. 2d).

Sampling and measurements

At seedling stage and after 5 weeks from growing, plants were sampled for trait measurements and genotypic analysis. Multiple root traits including root number, root length, shoot length, shoot dry weight, root dry weight and total dry weight of parental lines (XieqingzaoB, Zhonghui9308 and CSSL45) were measured under both (N⁺) normal and (N⁻) deficiency conditions. Root dry weight of F_{2:3}(BC₅F_{2:3}) plants was measured as the weight of root mass after 2 days of drying at 70 °C.

DNA extraction and development of molecular markers

DNA was extracted from the fresh leaves of each plants of F_{2:3} (BC₅F_{2:3}) population and their parents using the CTAB method described by Luo et al. [36]. PCR protocol was performed in a 10-μL reaction volume containing 1.5 μL of 20.0 ng/μL template genomic DNA, 1.0 μL of 10× PCR buffer, 0.25 μL of 1.0 pmol/μL dNTPs, 1.5 μL of 2.0 pmol/μL primer pairs, 0.06 μL of 5.0 U/μL Taq DNA polymerase and 5.69 μL of ddH₂O. The amplification protocol consisted of an initial denaturation step (94 °C for 5 min), followed by 32 cycles of 94 °C for 30 s, 55 °C for 30 s, and 72 °C for 1 min. The reactions were completed with a final extension step of 72 °C for 7 min. The PCR products were separated by electrophoresis on 8% non-denaturing polyacrylamide gels and then visualized by silver staining [37]. New InDel markers in the specific genomic region were designed according to genome sequence differences between the *indica* cv. XieqingzaoB and the *indica* cv. Zhonghui9308. The sequences of these new markers used in this investigation are listed in Table 1.

Statistical analysis of data

Mean phenotypic values for seedling root traits were compared using the Student's *t*-test. Previously, a total of

120 SSR and Insertion/Deletion markers were distributed along the rice genome and used for identifying the Ici-Mapping QTLs in 75 CSSLs [24]. Phenotypic correlations were calculated using a generalized linear model implemented within the SAS statistical software package.

Results

Characterization of XQZB and ZH9308 under different N levels

To identify the major QTLs responsible for nitrogen deficiency tolerance in rice, a set of 75 CSSLs population derived from an *Indica* c.v XieqingzaoB as the donor parent and *Indica* c.v Zhonghui9308 as the recurrent parent were used [24]. The phenotypic values for six seedling root traits of XieqingzaoB and Zhonghui9308 under both low and normal N are shown in Fig. 3. There were significant differences between XieqingzaoB and Zhonghui9308 for all studied root traits (root number, root length, root dry weight, shoot dry weight and total dry weight). The two parents, XieqingzaoB and Zhonghui9308 exhibited no differences in their shoot length under both N levels (Fig. 3b). Zhonghui9308 was significantly inhibited reductions in most of the root traits compared with XieqingzaoB under N⁻ deficiency level. The donor parent XieqingzaoB showing consistently higher values than Zhonghui9308 in two N conditions, thereby suggesting that Zhonghui9308 showed a smaller response to N deficiency than XieqingzaoB (Fig. 3).

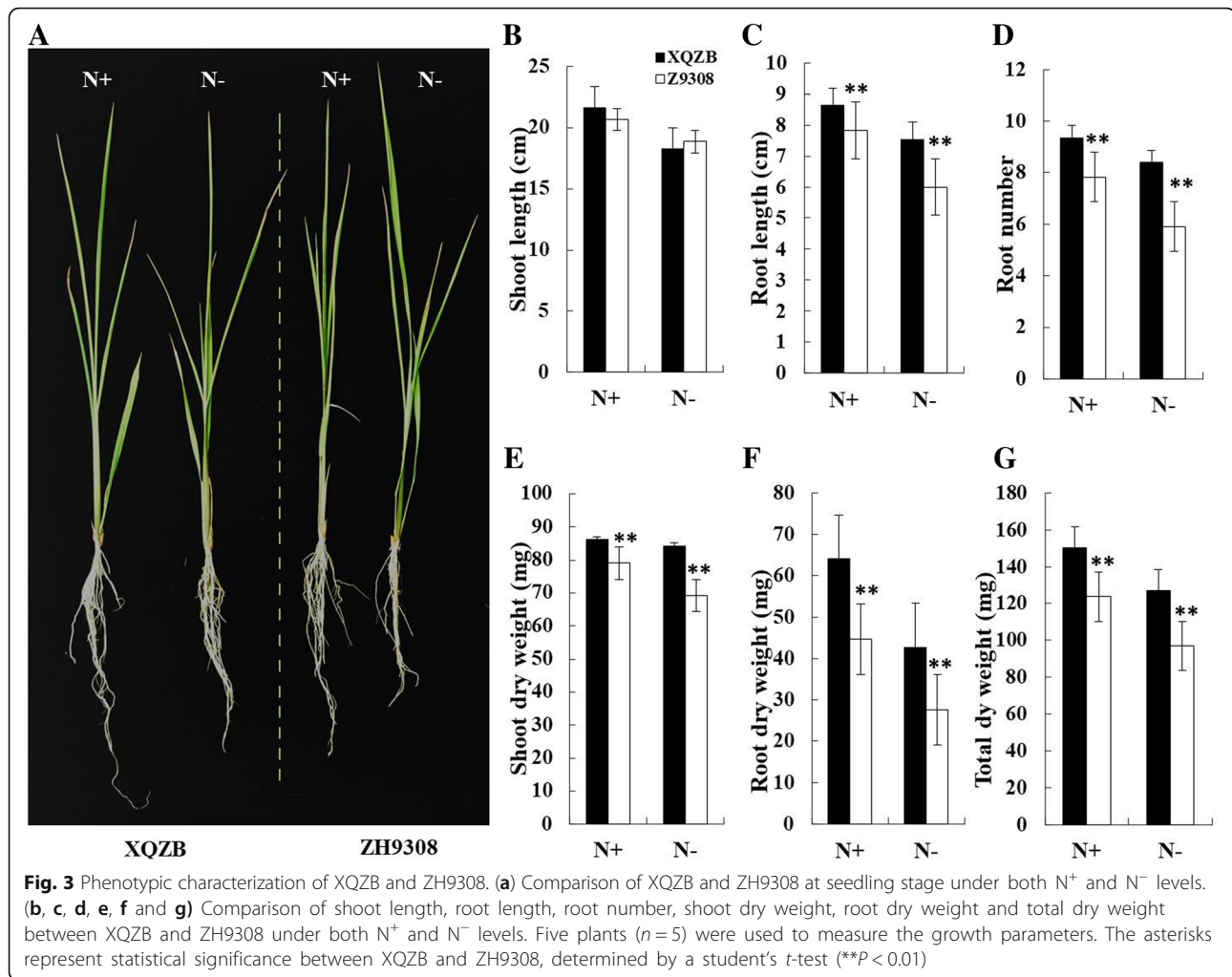
Correlation analysis among root traits

Phenotypic correlations coefficient analysis between root dry weight and other related root traits like root length and root number under both N conditions were performed (Fig. 4). Data by red color correspond to correlations under low N⁻ level and data by black color correspond to correlations under normal N⁺ level. As shown in Fig. 4, three phenotypic seedling root traits were significant and highly significant correlation coefficients across two N levels except for correlations between root

Table 1 DNA primers sequences for polymorphic markers designed for fine mapping of *qRDWN6^{XB}* on chromosome 6

Marker	Marker type	Forward primer sequence	Reverse primer sequence	Purpose	Size
RM5963	SSR	TCAAGTTACGGAAATGTGTGG	CTGCCTAGCTTCCGTTTCTCC	Primary mapping	139
InD90	InDel	CCTCATCCAGGGGTCATGTA	CGGTCAAGTGTCCATCCAGGT	Primary mapping	19
RM20069	SSR	GCGAGCGAGAGAGAGATAGACG	CGAATTCGGCACGAGTAATAGGG	Primary mapping	157
ND-3	InDel	AGACGGTGATATCGGTGAGT	GGAGTTTAGTGGCTGCATCA	Fine mapping	258
ND-4	InDel	AAAACACCAAAGAATCCGGC	AGGATAGGAAAACCGTGCAA	Fine mapping	282
ND-5	InDel	GCTTTAGCTACGGTTTCCGA	TTTGACTCGTCCCAATAGGC	Fine mapping	225
ND-11	InDel	CCGAGTAGCGAAGCTCAAAT	CTAGCATGGACGAACGGATG	Fine mapping	103
ND-13	InDel	ATTGAGCGTTCCTTAACCCG	GACAGAGTCGAGAAAACCGTG	Fine mapping	116
ND-15	InDel	AGATCTCACATGATTATATCCGA	TCTGCAACAAAGTGAATCCT	Fine mapping	117

SSR Simple Sequence Repeat, InDel Insertion/deletion



dry weight and root number, and between root length and roots number under normal nitrogen (N⁺). The strongest phenotypic correlations were found between root dry weight and both root number and root length under N deficiency level, with the correlation coefficients of 0.647** and 0.616**, respectively. Simultaneously, there was a highly significant correlation coefficient between root length and root number with the value of 0.442** under N⁻ level. On the other hand, the significant correlation between root length and root dry weight was found with the value of 0.377* under N⁺ level.

qRDWN6^{XB} mapping using a CSSL population

In our previous study and by using 75 CSSLs population derived from a cross between two parents (XieqingzaoB as a donor and Zhonghui9308 as a recipient) differ markedly for seedling root traits under N⁻ deficiency, *qRDWN6^{XB}* as a major QTL for root dry weight (RDW) against N-deficiency was detected. Using 120 simple sequence repeat (SSR) and insertion/deletion (InDel) markers, which were distributed along the whole rice

genome according to previously reported linkage maps and known polymorphisms between the parents were used to identify the QTLs in CSSLs population based on a cut-off LOD value ≥ 2.0 . Analysis of variance based on genotype, treatment, and genotype \times treatment interaction effects on root dry weight (RDW) revealed highly significant variance between the two N levels in the CSSL population (Table 2). RDW displayed a continuous variation and followed a normal frequency distribution under low N level (Fig. 5). This locus (named *qRDWN6^{XB}*) detected in the region between RM5963 and RM20069 markers on the long arm of chromosome 6. It had the highest LOD score of 3.00, PVE% (phenotypic variance explained) of 16.84% and additive value of 6.14, which indicated that the *qRDWN6^{XB}* is likely the main effect QTL (Table 3). The positive allele from XieqingzaoB increased the root dry weight under N⁻ deficiency condition. One CSSL namely CSSL45, which had higher root dry weight values compared with Zhonghui9308 under N⁻ deficiency and harboring the *qRDWN6^{XB}*, was selected for advanced investigation and their genotypes were shown in Fig. 6.

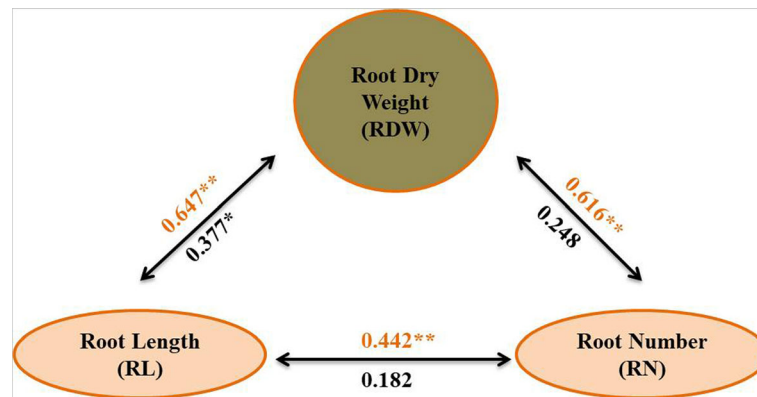


Fig. 4 Phenotypic correlations among root dry weight and other related traits. The red and black numbers represent Person correlation coefficients between the traits under N^- and N^+ , respectively. Single and double asterisk represent statistical significant at $P < 0.05$ and 0.01 , respectively

Characterization of CSSL45 compared to ZH9308 under N^- deficiency

To confirm the effect of $qRDWN6^{XB}$ for root dry weight under N deficiency, one chromosome segment substitution line (CSSL) namely CSSL45, harboring this QTL and have the segment from XieqingzaoB between the markers RM5754 and RM162 (Fig. 7b, c), was crossed with the background parent Zhonghui9308. When Zhonghui9308 and CSSL45 in addition to XieqingzaoB were evaluated under N deficiency in hydroponic condition, the root dry weight in the CSSL45 was significantly higher than that of Zhonghui9308 (Fig. 7a, f). For other related root parameters and compared to Zhonghui9308, CSSL45 showed a longer root length and higher root number under low nitrogen (Fig. 7d, e). Therefore, the CSSL45 was backcrossed with the parent ‘Zhonghui9308’ and a total of 2200 $F_{2:3}$ ($BC_5F_{2:3}$) seedlings obtained from the cross were screened for a fine mapping of $qRDWN6^{XB}$ under N deficiency using the homozygous recombinant plants (Fig. 8).

Fine mapping of the $qRDWN6^{XB}$ to a 52.3-kb genomic region

For genetic analysis and fine mapping of $qRDWN6^{XB}$, a total of 2200 $F_{2:3}$ individual plants derived from CSSL45/ZH9308 were screened and subjected to marker analysis by scanning RM5963 and RM20069 flanking $qRDWN6^{XB}$

in the preliminary mapping (Fig. 9a). Out of 65 Simple Sequence Repeats (SSR) markers located in this genomic region (<http://www.gramene.org/>), nine markers were identified as polymorphic between the parents CSSL45 and ZH9308 (Fig. 9b). An analysis of RM5963 detected 98 recombination events between the marker and $qRDWN6^{XB}$ on one side, while an analysis of RM20069 identified 169 recombination events between the marker and $qRDWN6^{XB}$ on the other side. Simultaneously, the SSR markers RM20034, RM19986, InD90, RM19844, RM19840, RM19834, RM19831, RM19807 and RM19804 revealed 158, 155, 148, 121, 102, 99, 90, 86 and 48 recombinants, respectively, while RM19765 showed 51 recombinants on the other side (Fig. 9b). Therefore, the $qRDWN6^{XB}$ was mapped to a 1003-kb genomic region between the interval RM19765 and RM19804. In this genomic region and out of 13 SSR markers, only four markers exhibited polymorphism between the two parents and used to narrow the region of $qRDWN6^{XB}$ using the homozygous recombinant plants. The SSR markers RM19766, RM19771, and RM19776 were found to co-segregate with $qRDWN6^{XB}$ and nine recombinant plants were revealed at RM19771, therefore $qRDWN6^{XB}$ was mapped in the 247.3-kb interval of RM19766 to RM19776 (Fig. 9c). To narrow the targeted region of $qRDWN6^{XB}$, fifteen new Insertion/Deletion (InDel) markers located at the substituted interval were designed and of which six markers exhibited polymorphism

Table 2 Analysis of variance to determine the contribution to root dry weight (RDW) of genotype, treatment and their interaction in CSSLs population

Source of variance (S.O.V)	Degree of freedom (df)	Mean square (MS)	F-value (F)	P-value (P)
Genotype	75	43.795	362.30	**
Treatment ^a	1	4291.216	86.55	**
Genotype x treatment	75	55.696	328.19	**
Residual	152	24.87		

^a Including two N levels (normal N^+ and low N^-)

**Significant at the 0.01 level

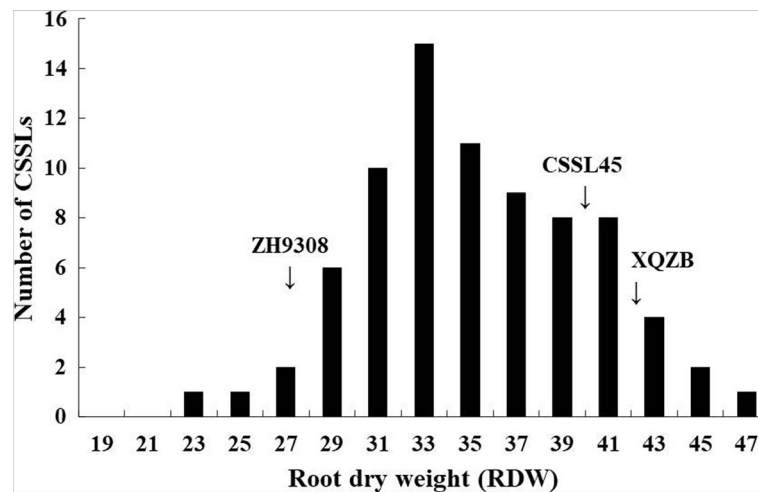


Fig. 5 Frequency distribution of root dry weight (RDW) in CSSL population under nitrogen deficiency condition (N⁻). The arrows show average RDW of CSSL45, XQZB and ZH9308

between the parents CSSL45 and ZH9308 (Table 1). Further recombinant screening with the newly designed Insertion/Deletion markers as well as RM19766, RM19771, and RM19776 showed that the gene was narrowed between ND-4 and RM19771 (Fig. 9d). With these newly designed markers and selected the homozygous recombinant plants, we evaluated the gene effect and then validated the phenotypic performance of root dry weight and two closely related traits, which varied from 28.9–41.4 mg for root dry weight, 11.3–13.8 cm for root length and 8–12 for root number. Finally and on the basis of the phenotypic and genetic analysis, the targeted region of *qRDWN6^{XB}* was localized to a 52.3-kb between the ND-4 and RM19771 markers (Fig. 9d).

Candidate genes at the *RDWN6^{XB}* locus

Based on Rice Genome Annotation Project Database (<http://rice.plantbiology.msu.edu/cgi-bin/gbrowse/rice/>) and within the 8,981,538–9,033,871 bp in the target genomic region of *RDWN6^{XB}* on chromosome 6, there are nine predicted genes in this 52.3-kb region (Table 4). *LOC_Os06g15830* and *LOC_Os06g15890* expressed protein, *LOC_Os06g15840* and *LOC_Os06g15850* transposon protein, *LOC_Os06g15860*, *LOC_Os06g15870* and *LOC_Os06g15880* retrotransposon protein, *LOC_Os06g15900* conserved hypothetical protein and *LOC_Os06g15910* potassium transporter 24. *LOC_Os06g15910* has been reported previously as a major potassium transporter gene *OsHAK24*, so it was nominated and considered the most

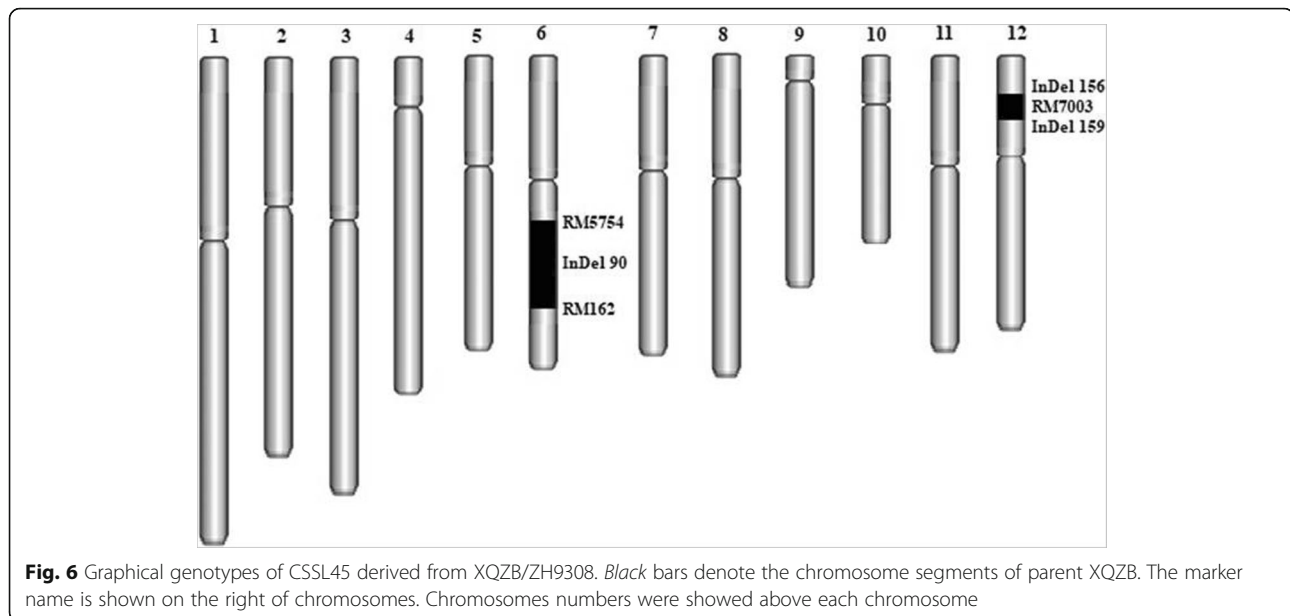
likely and strong candidate gene for the *RDWN6^{XB}* locus associated with nitrogen deficiency tolerance.

Discussion

Genetic populations played an important role in quantitative trait loci (QTLs) identifying and gene mapping. With different genetic segregating populations, such as F₂ populations, chromosome segment substitution lines (CSSLs), double haploid lines (DH), near-isogenic lines (NILs) and recombinant inbred lines (RILs), hundreds genomic regions associated with several root-related traits under macro-nutrients deficiency have been identified in rice, but few have been fine mapped or cloned [38]. CSSLs have been a successful and effective tool for QTL mapping and positional cloning [39, 40]. Recently, there have been an increasing number of published studies on the genetic, molecular and physiological regulation for root architecture as related to rice nutrient efficiency. A number of genes which are involved in changing the root development and root system architecture to facilitate enhanced nutrient acquisition have been identified in rice [33, 41, 42]. Two major QTLs, *TOND1*, confers tolerance to N deficiency tolerance in rice [33], *DEP1*, increases the tolerance to N deficiency [42], have been cloned. In our previous study, we analyzed the genomic regions in the XQZB/ZH9308 CSSLs population and found three putative QTLs under N deficiency during seedling stage using hydroponic culture. Among them, one strong QTL (*qRDWN6^{XB}*) had the highest LOD value and that corresponds to the root dry

Table 3 Descriptive statistics for *qRDWN6^{XB}* mapped under low N in the CSSL population derived from XQZB and ZH9308

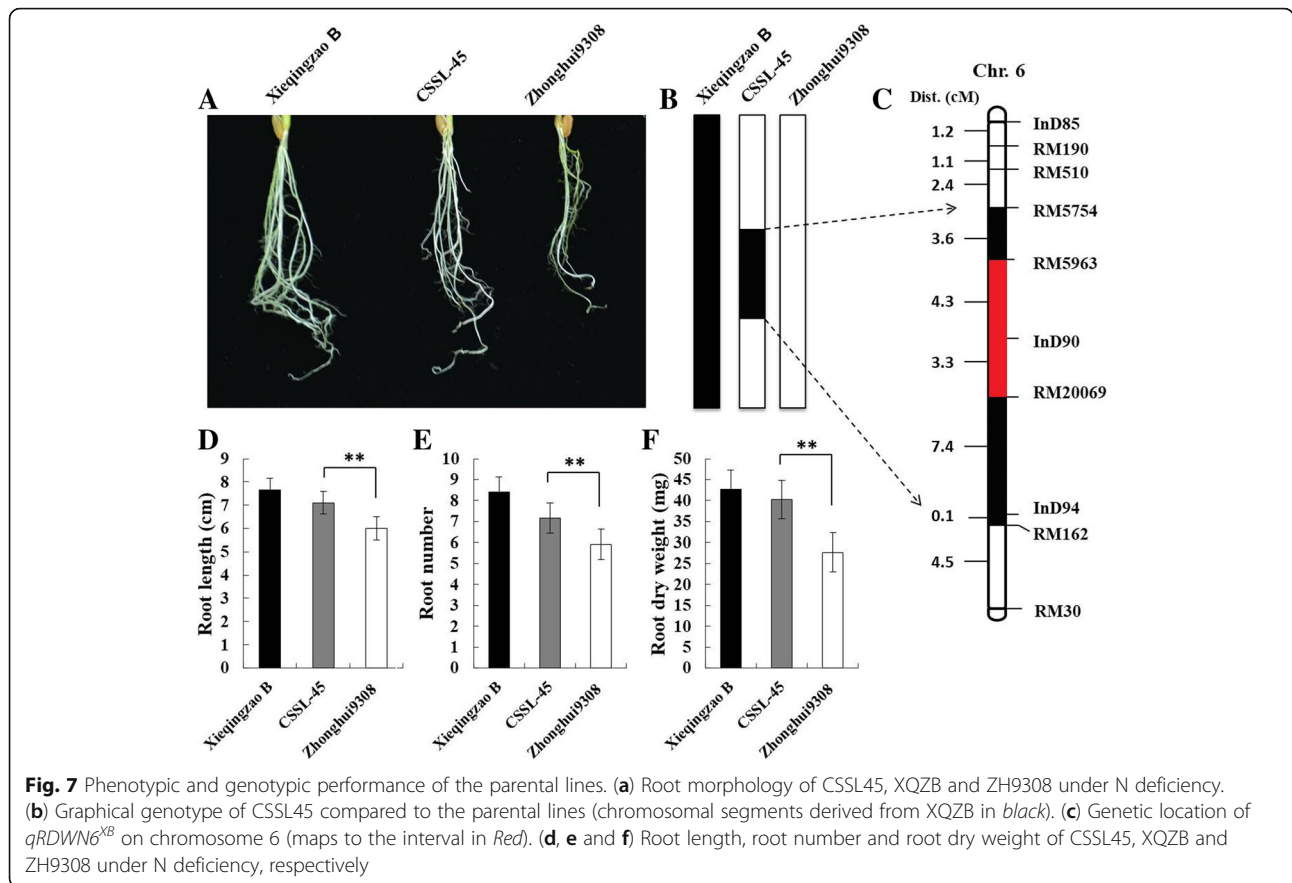
Trait	Nitrogen level	QTL	Chr.	Marker name	LOD score	PVE (%)	Add
RDW	N ⁻	<i>qRDWN6^{XB}</i>	6	InD90	3.00	16.84	6.14



weight was identified in the genomic region of RM5963-RM20069 on the long arm of chromosome 6 [24]. In the same chromosome, major QTLs for some root traits were also identified [19, 20, 26, 43] and affecting root system architecture, suggesting $qRDWN6^{XB}$ might be a common genetic factor for root traits in various rice genotypes. Previous studies revealed that root dry weight (RDW) was significantly correlated with the other root traits under N deficiency, such as root number (RN), root length (RL), root thickness (RT) and root volume (RV), suggesting that these traits might be controlled and inherited by common genes [44, 45]. Root dry weight has been considered the most important component associated with crop yields than the other root traits. Therefore, significantly correlations analysis between root dry weight (RDW) and other related root traits like root number (RN) and root length (RL) under N deficiency were observed (Fig. 4). In this study, we delimited the chromosome segment containing $qRDWN6^{XB}$ and fine mapped to a 52.3-kb for the first time. One CSSL, CSSL45- $qRDWN6^{XB}$, which consistently produced higher root dry weight was selected for advanced study and crossed with the genetic background parent Zhonghui9308 and a total of 2200 $F_{2:3}$ ($BC_5F_{2:3}$) seedlings obtained from the cross were screened for a fine mapping of $qRDWN6^{XB}$ under N^- deficiency condition. In addition, CSSL45 as a line harboring this locus was significantly higher than Zhonghui9308 in root dry weight (RDW) under two nitrogen levels (N^+ and N^-), indicating $qRDWN6^{XB}$ could stably express in different conditions and there were more CSSLs with high RDW under N^- deficiency than that of the two parents (Fig. 5). This transgressive segregation of root traits including

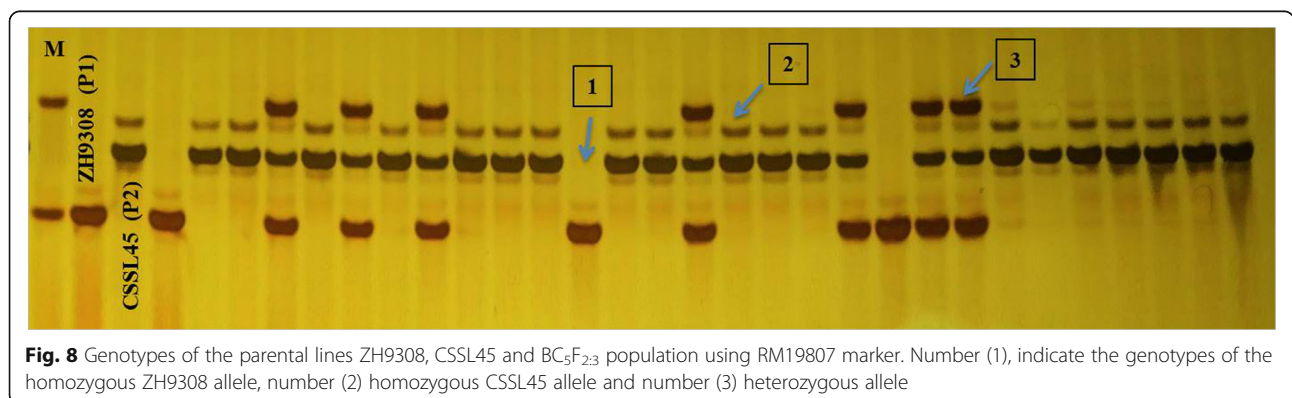
RDW under different N treatments was also observed in introgression lines [26] and recombinant inbred line (RIL) population [28].

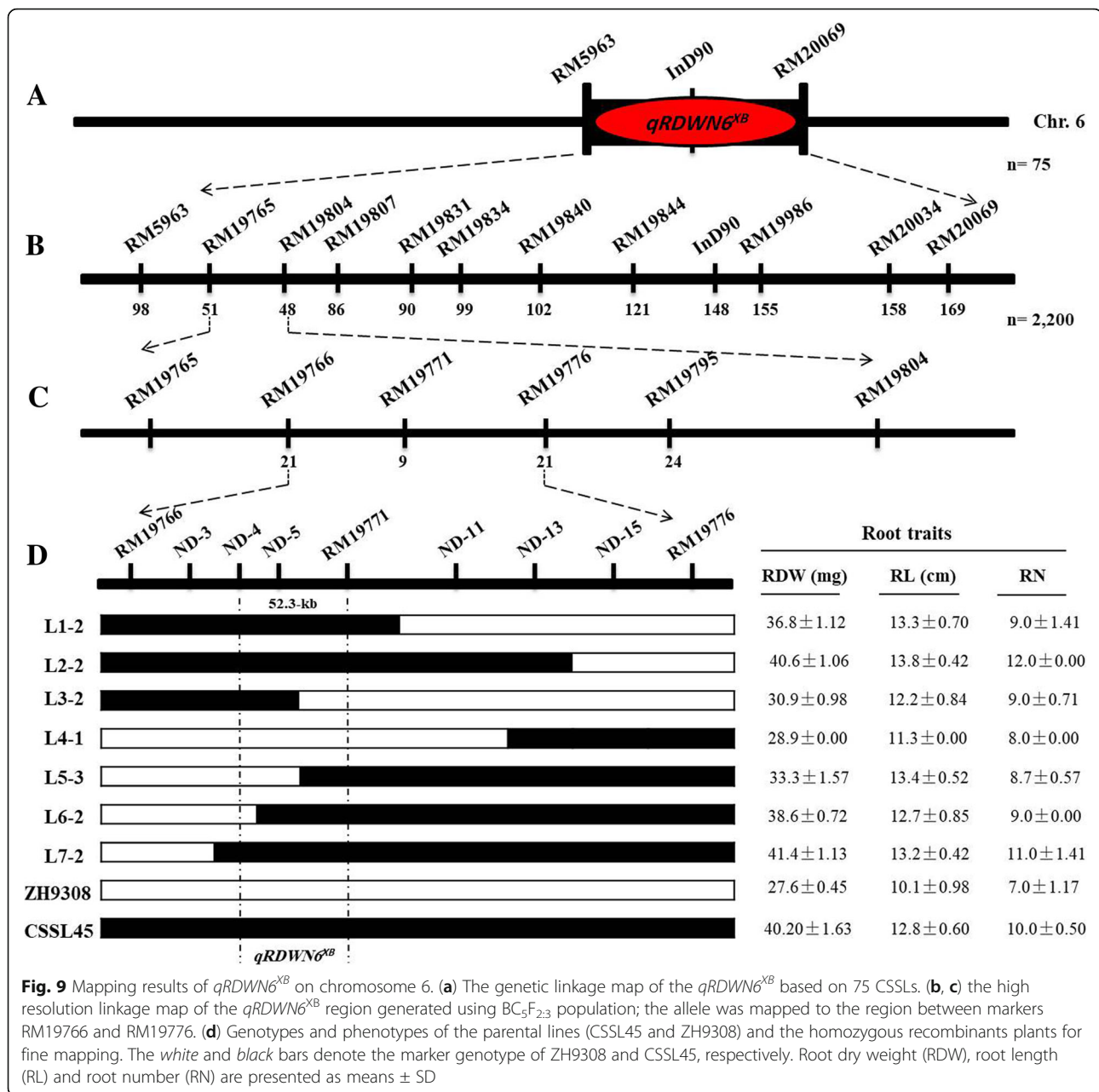
In rice crop, the important functions of the root system are the responses to low N availability and mainly through enhanced root traits like root number, root length, root density and root thickness to absorb water and primarily nutrients [23, 46–48]. Root dry weight is better related to crop yields and important measurement because it is a summary of all root traits. In this investigation, we identified $qRDWN6^{XB}$, a major QTL enhancing tolerance of N deficiency in rice. This QTL is the first locus fine mapped for root dry weight under N deficiency in the target region 8,981,538 - 9,033,871 bp on chromosome 6. Numerous studies have identified several QTLs/genes for nitrogen use efficiency on the same chromosome. For example, Song et al. [49] identified *AspAT3* gene within the 23,738,029 - 23,738,154 bp region, which is contributed to nitrogen and carbon metabolism in rice. Around the waxy gene on chromosome 6, Shan et al. [50] mapped *qNUEp-6* as a major QTL for nitrogen utilization in the 1.76–2.09 Mb regions. Under normal and low nitrogen conditions and within the 28.13–29.63 Mb regions, Tong et al. [22] identified QTLs associated with dry weight and grain yield on the chromosome 6. Using rice chromosome segment substitution lines (CSSLs) a novel QTL of effective panicle and yield in the range of 2.29–2.83 Mb was detected by Wang et al. [51]. Also, a novel locus related to NUE was identified using a genome-wide association analysis of 184 rice genotypes at 4,845,258 - 4,845,375 bp and near the SSR marker RM314 [52]. Yang et al. [53] delimited the *qNUE6* locus in 8,647,275 -



8,913,783 bp region. However, these QTLs/genes are not the same loci as *qRDWN6^{XB}*; therefore, this QTL is a newly discovered locus controlling nitrogen deficiency tolerance in the target region. Among the nine predicted genes in the fine-mapped region 52.3-kb, only one gene *LOC_Os06g15910* has been reported previously as a major potassium transporter gene *OsHAK24* [54], it was considered the most likely and strong gene for the *RDWN6^{XB}* locus and might be the candidate gene responsible for nitrogen deficiency tolerance. Previous studies have demonstrated different genes controlling

nitrogen use efficiency and nitrogen deficiency tolerance in rice. In the concentrations of low and high nitrate ions, two genes *NRT1.1B* had a nitrate-transporting activity and *NRT2* a high-affinity transporter were found by Hu et al. [55] but *NRT2* can't transfer NO^{-3} independently. *OsNRT2.3b* able to increase the uptake of nitrogen and phosphorus, nitrogen use efficiency and improving the grain yield [56]. At different levels of nitrate, Yan et al. [57] showed that *OsNAR2.1* is able to interact with *OsNRT2-1*, *OsNRT2-2*, and *OsNRT2-3* genes, and can enhance nitrate uptake by rice roots at





different levels of nitrate. *TOND1* as a major QTL was detected on chromosome 12 and its over-expression enhanced the nitrogen deficiency tolerance in rice [33]. *OsAMT1-1*, *OsAMT1-2*, and *OsAMT1-3* are the major three ammonium transporter genes were identified in rice and played a pivotal role in absorbing NH_4^+ [58–60]. These results have greatly enhanced our understanding of the regulation of nitrogen use efficiency and nitrogen deficiency tolerance in rice. Here, we report the characterization and identification of a novel QTL, $qRDWN6^{XB}$, that regulates root dry weight in rice to a region of 52.3-kb (Fig. 9d), and identified nine predicted genes as viable candidates for it. One of them,

LOC_Os06g15910 previously reported as a major potassium transporter gene *OsHAK24* [54] might play an important role in nitrogen deficiency tolerance in rice. The annual consumption of N fertilizers increases year after year, causing severe pollution to agricultural and ecological environments. Only 30–45% of N fertilizer is efficiently used by rice plants [61]. Therefore, the rice cultivars harbor the $qRDWN6^{XB}$ may significantly decrease the use of N fertilizers, reduce the cost of rice production, alleviate pollution and protect the environment. $qRDWN6^{XB}$ is expected to have wide use potential in rice breeding programs and will play an important role in the ‘Second Green Revolution’.

Table 4 Candidate genes located within 52.3-kb physical regions for *qRDWN6^{XB}* on chromosome 6

Gene ID	Gene start (bp)	Gene end (bp)	Putative function
<i>LOC_Os06g15830</i>	8,986,168	8,987,101	Expressed protein
<i>LOC_Os06g15840</i>	8,992,763	8,996,095	Transposon protein, putative, CACTA, En/Spm sub-class, expressed
<i>LOC_Os06g15850</i>	8,999,300	9,004,290	Transposon protein, putative, CACTA, En/Spm sub-class, expressed
<i>LOC_Os06g15860</i>	9,006,419	9,007,843	Retrotransposon protein, putative, unclassified, expressed
<i>LOC_Os06g15870</i>	9,009,481	9,014,615	Retrotransposon protein, putative, unclassified, expressed
<i>LOC_Os06g15880</i>	9,017,523	9,018,636	Retrotransposon protein, putative, unclassified
<i>LOC_Os06g15890</i>	9,019,348	9,019,788	Expressed protein
<i>LOC_Os06g15900</i>	9,021,414	9,022,376	Conserved hypothetical protein
<i>LOC_Os06g15910</i>	9,033,862	9,037,775	Potassium transporter 24, putative, expressed

Conclusions

In summary, a major QTL, *qRDWN6^{XB}* was identified in the rice response to N deficiency and found that it enhances a positive regulator of root system architecture. Based on the target CSSL and homozygous recombinant population, *qRDWN6^{XB}* was delimited to a 52.3-kb region on chromosome 6. Nine candidate genes were identified in the target region using gene annotation information. *LOC_Os06g15910* previously reported as a potassium transporter gene in this region and might play an important role in nitrogen deficiency tolerance of rice. The identification of *qRDWN6^{XB}* provides a new genetic resource for breeding rice varieties and a starting point to improve grain yield despite the decreased input of N fertilizers. The newly developed and tightly linked InDel marker ND-4 will be useful to improve the root system architecture by marker-assisted selection (MAS) in rice breeding programs. But to understand how *RDWN6^{XB}* affects the agronomic characteristics of rice, further study is needed to clarify their molecular and biological functions through cloning and transgenic approaches.

Abbreviations

CSSLs: Chromosome Segment Substitution Lines; DH: Double haploid lines; InDel: Insertion/deletion; LOD: Logarithm of the odds; MAS: Marker assisted selection; N⁻: Low nitrogen; N⁺: Normal nitrogen; NILs: Near-isogenic lines; PVE: Phenotypic variance explained; QTL: Quantitative trait loci; RDW: Root dry weight; RDWN: Root dry weight under N deficiency; RILs: Recombinant inbred lines; RL: Root length; RN: Root Number; SSR: Simple Sequence Repeat; XQZB: XieqingzaoB; ZH9308: Zhonghui9308

Acknowledgements

The authors would like to thank Dr. Nehal Elekhtyar (Agricultural Research Center, Egypt) for excellent assistance with data analysis. The authors also thank Yuyu Chen, Adil Abbas and Aamir Riaz for their efforts and suggestions. Lastly, the authors thank the reviewers for their valuable comments to improve the quality of the manuscript.

Funding

This work was supported by the National Key Research and Development Program (2016YFD0101801), the National Rice Technology System (CARS-01), the Natural Science Foundation of China (31521064), and the Agricultural Science and Technology Innovation Program of the Chinese Academy of Agricultural Science (CAAS-ASTIP-2013-CNRI). These funding bodies had no role in the experimental design, data collection, analysis, interpretation of data and preparation of the manuscript.

Availability of data and materials

The datasets generated and analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

YZX, LC and SC designed the research. GBA, YXZ and WW prepare materials. GBA performed research. GBA, YXZ, AI, YZ, YC and WW analyzed data. GBA and YXZ wrote the drafting of manuscript. All authors GBA, YXZ, AI, YZ, YC, WW, LC and SC have read, revised and approved the final manuscript.

Ethics approval and consent to participate

The rice seed, XieqingzaoB and Zhonghui9308 are common and broadly cultivated varieties in China. The seed of all materials was provided by China National Rice Research Institute (Hangzhou, China). Our present work didn't use transgenic technology or material therefore it does not require ethical approval. The experimental research on plants performed in this study complies with institutional, national and international guidelines. The field study was conducted in accordance with local legislation.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details

¹Key Laboratory for Zhejiang Super Rice Research and State Key Laboratory of Rice Biology, China National Rice Research Institute, Hangzhou 310006, China. ²Rice Research and Training Center, Field Crops Research Institute, Agriculture Research Center, Kafrelsheikh 33717, Egypt.

Received: 24 September 2018 Accepted: 27 December 2018

Published online: 08 January 2019

References

- Zhou Y, Zhu JY, Li ZY, Yi CD, Liu J, Zhang HG, et al. Deletion in a quantitative trait gene *qPE9-1* associated with panicle erectness improves plant architecture during rice domestication. *Genet.* 2009;183(1):315–24.
- Cottyn B, Debode J, Regalado E, Mew TW, Swings J. Phenotypic and genetic diversity of rice seed-associated bacteria and their role in pathogenicity and biological control. *J Appl Microbiol.* 2009;107(3):885–97.
- Miura K. The role of QTLs in the breeding of high-yielding rice. *Trends Plant Sci.* 2011;16(6):319–26.
- Khush GS, Brar DS. Biotechnology for rice breeding: progress and potential impact. In: proceeding of the 20th session of the international rice commission, 23th - 26th July, 2002; Bangkok,Thailand.
- Gregory PJ, George TS. Feeding nine billion: the challenge to sustainable crop production. *J Exp Bot.* 2011;62(15):5233–9.

6. Epstein E, Bloom A. Mineral nutrition of plants 2. Principles and Perspectives. Sunderland: Sinauer; 2005.
7. Samonte S, Wilson LT, Medley JC, Pinson SRM, Mc-Clung A, Lales JS. Nitrogen utilization efficiency: relationships with grain yield, grain protein, and yield-related traits in rice. *Agron J*. 2006;98:168–76.
8. Fageria NK. Yield physiology of rice. *J Plant Nutr*. 2007;30:843–79.
9. Tirol-Padre A, Ladha JK, Singh U, Laureles E, Punzalan G, Akita S. Grain yield performance of rice genotypes at suboptimal levels of soil N as affected by N uptake and utilization efficiency. *Field Crops Res*. 1996;46(1–3):127–43.
10. Raun WR, Johnson GV. Improving nitrogen use efficiency for cereal production. *Agron J*. 1999;91(3):357–63.
11. Guo JH, Liu XJ, Zhang Y, Shen JL, Han WX, Zhang WF, et al. Significant acidification in major Chinese croplands. *Sci*. 2010;327(5968):1008–10.
12. Glass ADM. Nitrogen use efficiency of crop plants: physiological constraints upon nitrogen absorption. *Crit Rev Plant Sci*. 2003;22(5):453–70.
13. Cho Y, Jiang WZ, Chin JH, Piao ZZ, Cho YG, McCouch SR, et al. Identification of QTLs associated with physiological nitrogen use efficiency in rice. *Mol Cells*. 2007;23(1):72–9.
14. Selvaraj MG, Valencia MO, Ogawa S, Lu YZ, Wu LY, Downs C, et al. Development and field performance of nitrogen use efficient rice lines for Africa. *Plant Biotechnol J*. 2017;15(6):775–87.
15. Borrell AK, Garside AL, Fukai S, Reid DJ. Season, nitrogen rate, and plant type affect nitrogen uptake and nitrogen use efficiency in rice. *Aust J Agric Res*. 1998;49(5):829–43.
16. Novoa R, Loomis RS. Nitrogen and plant production. *Plant Soil*. 1981;58:177–204.
17. Hirel B, Le Gouis J, Ney B, Gallais A. The challenge of improving nitrogen use efficiency in crop plants: towards a more central role for genetic variability and quantitative genetics within integrated approaches. *J Exp Bot*. 2007;58(9):2369–87.
18. Smith S, Smet ID. Root system architecture: insights from Arabidopsis and cereal crops. *Philos Trans R Soc Lond Ser B Biol Sci*. 2012;367(1595):1441–52.
19. Lian XM, Xing YZ, Yan H, Xu CG, Li XH, Zhang QF. QTLs for low nitrogen tolerance at seedling stage identified using a recombinant inbred line population derived from an elite rice hybrid. *Theor Appl Genet*. 2005;112(1):85–96.
20. Obara M, Tamura W, Ebitani T, Yano M, Sato T, Yamaya T. Fine mapping of *qRL6.1*, a major QTL for root length of rice seedlings grown under a wide range of NH₄⁺ concentrations in hydroponic conditions. *Theor Appl Genet*. 2010;121(3):535–47.
21. Ogawa S, Valencia MO, Ishitani M, Selvaraj MG. Root system architecture variation in response to different NH₄⁺ concentrations and its association with nitrogen-deficient tolerance traits in rice. *Acta Physiol Plant*. 2014;36(9):2361–72.
22. Tong HH, Mei HW, Yu XQ, Xu XY, Li MS, Zhang SQ, et al. Identification of related QTLs at late developmental stage in rice (*Oryza sativa* L.) under two nitrogen levels. *Acta Genet Sin*. 2006;33(5):458–67.
23. Yu P, Li X, White PJ, Li C. A large and deep root system underlies high nitrogen-use efficiency in maize production. *PLoS One*. 2015;10(5):1–17.
24. Anis G, Zhang YX, Xu XM, Fiaz S, Wu WX, Rahman MH, et al. QTL analysis for rice seedlings under nitrogen deficiency using chromosomal segment substitution lines. *Pak J Bot*. 2018;50:537–44.
25. Wei D, Cui K, Ye G, Pan J, Xiang J, Huang J, et al. QTL mapping for nitrogen-use efficiency and nitrogen-deficiency tolerance traits in rice. *Plant Soil*. 2012;359:281–95.
26. Kim PS, Kim DM, Kang JW, Lee HS, Ahn SN. QTL mapping of Rice root traits at different NH₄⁺ levels in hydroponic condition. *Plant Breed Biotech*. 2015; 3(3):244–52.
27. Obara M, Takeda T, Hayakawa T, Yamaya T. Mapping quantitative trait loci controlling root length in rice seedlings grown with low or sufficient supply using backcross recombinant lines derived from a cross between *Oryza sativa* L. and *Oryza glaberrima* Steud. *Soil Sci and Plant Nutr*. 2011;57(1):80–92.
28. Feng Y, Cao LY, Wu WM, Shen XH, Zhan XD, Zhai RR, et al. Mapping QTLs for nitrogen-deficiency tolerance at seedling stage in rice (*Oryza sativa* L.). *Plant Breed*. 2010;129:652–6.
29. Zhao C, Zhao L, Zhang Y, Chen T, Zhao Q, Zhu Z, et al. QTL mapping for seedling traits related to low nitrogen tolerance in rice. *Acta Agric Bor-Sin*. 2015;30(6):1–7.
30. Senapathy S, Kunnummal KV, Palaniappan M, Marappa M. QTL and QTLxenvironment effects on agronomic and nitrogen acquisition traits in rice. *J Integr Plant Biol*. 2008;50(9):1108–17.
31. Tang Z, Fan XR, Li Q, Feng HM, Miller AJ, Shen QR, et al. Knock down of a rice stellar nitrate transporter alters long distance translocation but not root influx. *Plant Physiol*. 2012;160(4):2052–63.
32. Li SM, Shi WM. Quantitative characterization of nitrogen regulation of *OsAMT1;1*, *OsAMT1;2*, and *OsAMT2;2* expression in rice seedlings. *Russ J Plant Physiol*. 2006;53(6):837–43.
33. Zhang YJ, Tan LB, Zhu ZF, Yuan LX, Xie DX, Sun CQ. *TOND1* confers tolerance to nitrogen deficiency in rice. *Plant J*. 2015;81(3):367–76.
34. Cheng SH, Cao LY, Zhuang JY, Chen SG, Zhan XD, Fan YY, et al. Super hybrid rice breeding in China: achievements and prospects. *J Integr Plant Biol*. 2007;49(6):805–10.
35. Yoshida S, Forno DA, Cock JH, Gomez KA. Laboratory Manual for physiological studies of rice. Las Bano. Laguna: IRRRI; 1976. p. 83.
36. Luo ZY, Zhou G, Chen XH, Lu QH, Hu WX. Isolation of high-quality genomic DNA from plants. *Bull Hunan Med Univ*. 2001;26:178–80.
37. Creste S, Tulmann Neto A, Figueira A. Detection of single sequence repeat polymorphisms in denaturing polyacrylamide sequencing gels by silver staining. *Plant Mol Biol Rep*. 2001;19(4):299–306.
38. Courtois B, Ahmadi N, Khawaja F, Price AH, Rami J-F, Frouin J, et al. Rice root genetic architecture: meta-analysis from a drought QTL database. *Rice*. 2009;2(2–3):115–28.
39. Xiao YH, Liu LL, Jiang L, Lu CG, Yu CY, Zhang WW, et al. Development of the chromosome segment substitution lines (CSSL) derived from a hybrid rice cross, PA64s/9311 with super high yield potential. *Rice Genet Newsl*. 2005;22:17–9.
40. Wan XY, Wan JM, Jiang L, Wang JK, Zhai HQ, Weng JF, et al. QTL analysis for rice grain length and fine mapping of an identified QTL with stable and major effects. *Theor Appl Genet*. 2006;112(7):1258–70.
41. Wu P, Wang X. Role of *OsPHR2* on phosphorus homeostasis and root hairs development in rice (*Oryza sativa* L.). *Plant Signal Behav*. 2008;3(9):674–5.
42. Sun HY, Qian Q, Wu K, Luo J, Wang S, Zhang C, et al. Heterotrimeric G proteins regulate nitrogen-use efficiency in rice. *Nat Genet*. 2014;46(6):652–6.
43. Anis GB, Zhang YX, Wang H, Li ZH, Wu WX, Sun LP, et al. Genomic regions analysis of seedling root traits and their regulation in responses to phosphorus deficiency tolerance in CSSL population of elite super hybrid rice. *Int J Mol Sci*. 2018;19(5):1460.
44. Wang HM, Xu XM, Zhan XD, Zhai RR, Wu WM, Shen XH, et al. Identification of *qRL7*, a major quantitative trait locus associated with rice root length in hydroponic conditions. *Breed Sci*. 2013;63(3):267–74.
45. Zhao C, Zhou L, Zhang Y, Zhu Z, Chen T, Zhao Q, et al. QTL mapping for seedling traits associated with low-nitrogen tolerance using a set of advanced backcross introgression lines of rice. *Plant Breed*. 2014;133(2):189–95.
46. Wang XB, Wu P, Hu B, Chen QS. Effects of nitrate on the growth of lateral root and nitrogen absorption in rice. *Acta Bot Sin*. 2002;44:678–83.
47. Ju CX, Buresh RJ, Wang ZQ, Zhang H, Liu LJ, Yang JC, et al. Root and shoot traits for rice varieties with higher grain yield and higher nitrogen use efficiency at lower nitrogen rates application. *Field Crop Res*. 2015;175:47–55.
48. Rasmussen IS, Dresboll DB, Thorup-Kristensen K. Winter wheat cultivars and nitrogen (N) fertilization-effects on root growth, N uptake efficiency and N use efficiency. *Eur J Agron*. 2015;68:38–49.
49. Song J, Yamamoto K, Shomura A, Yano M, Minobe Y, Sasaki T. Characterization and mapping of cDNA encoding aspartate aminotransferase in rice, *Oryza sativa* L. *DNA Res*. 1996;3(5):303–10.
50. Shan YH, Wang YL, Pan XB. Mapping of QTLs for Nitrogen use efficiency and related traits in rice (*Oryza sativa* L.). *J Integr Agric*. 2005;4:721–7.
51. Wang Y, Sun YJ, Chen DY, Yu SB. Analysis of quantitative trait LOCI in response to nitrogen and phosphorus deficiency in rice using chromosomal segment substitution lines. *Acta Agronom Sin*. 2009;35(4):580–7.
52. Liu Z, Zhu C, Jiang Y, Tian Y, Yu J, An H, et al. Association mapping and genetic dissection of nitrogen use efficiency-related traits in rice (*Oryza sativa* L.). *Funct Integr Genomics*. 2016;16(3):323–33.
53. Yang X, Xia X, Zhang Z, Nong B, Zeng Y, Xiong F, et al. QTL mapping by whole genome re-sequencing and analysis of candidate genes for nitrogen use efficiency in Rice. *Front Plant Sci*. 2017;8:1634.
54. Yang ZF, Gao QS, Sun CS, Li WJ, Gu SL, Xu CW. Molecular evolution and functional divergence of HAK potassium transporter gene family in rice (*Oryza sativa* L.). *J Genet Genomics*. 2009;36(3):161–72.
55. Hu B, Wang W, Ou S, Tang J, Li H, Che R, et al. Variation in NRT1.1B contributes to nitrate-use divergence between rice subspecies. *Nat Genet*. 2015;47(7):834–8.
56. Fan X, Tang Z, Tan Y, Zhang Y, Luo B, Yang M, et al. Overexpression of a pH-sensitive nitrate transporter in rice increases crop yields. *Proc Natl Acad Sci*. 2016;113(26):7118–23.
57. Yan M, Fan X, Feng H, Miller AJ, Shen Q, Xu G. Rice *OsNAR2.1* interacts with *OsNRT2.1*, *OsNRT2.2* and *OsNRT2.3a* nitrate transporters to provide uptake over high and low concentration ranges. *Plant Cell Environ*. 2011;34(8):1360–72.

58. Sonoda Y, Ikeda A, Saiki S, Von WN, Yamaya T, Yamaguchi J. Distinct expression and function of three ammonium transporter genes (*OsAMT1;1* – *1;3*) in rice. *Plant Cell Physiol.* 2003;44(7):723–34.
59. Ferreira LM, Souza V, Tavares O, Zonta E, Santa-catarina C, Souza SR, et al. *OsAMT1.3* expression alters rice ammonium uptake kinetics and root morphology. *Plant Biotechnol Rep.* 2015;9(4):221–9.
60. Yang S, Hao D, Cong Y, Jin M, Su Y. The rice *OsAMT1;1* is a proton-independent feedback regulated ammonium transporter. *Plant Cell Rep.* 2015;34(2):321–30.
61. Peng SB, Buresh RJ, Huang JL, Yang JC, Zou YB, Zhong XH, et al. Strategies for overcoming low agronomic nitrogen use efficiency in irrigated rice systems in China. *Field Crop Res.* 2006;96(1):37–47.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

