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Thionin-like peptide from *Capsicum annuum* fruits: mechanism of action and synergism with fluconazole against *Candida* species

Gabriel B. Taveira¹, André O. Carvalho¹, Rosana Rodrigues², Fernanda G. Trindade³, Maura Da Cunha³ and Valdirene M. Gomes^{1*}

Abstract

Background: Thionins are a family of plant antimicrobial peptides (AMPs), which participate in plant defense system against pathogens. Here we describe some aspects of the *Ca*Thi thionin-like action mechanism, previously isolated from *Capsicum annuum* fruits. Thionin-like peptide was submitted to antimicrobial activity assays against *Candida* species for IC₅₀ determination and synergism with fluconazole evaluation. Viability and plasma membrane permeabilization assays, induction of intracellular ROS production analysis and *Ca*Thi localization in yeast cells were also investigated.

Results: *Ca*Thi had strong antimicrobial activity against six tested pathogenic *Candida* species, with IC₅₀ ranging from 10 to 40 µg.mL⁻¹. *Ca*Thi antimicrobial activity on *Candida* species was candidacidal. Moreover, *Ca*Thi caused plasma membrane permeabilization in all yeasts tested and induces oxidative stresses only in *Candida tropicalis*. *Ca*Thi was intracellularly localized in *C. albicans* and *C. tropicalis*, however localized in nuclei in *C. tropicalis*, suggesting a possible nuclear target. *Ca*Thi performed synergistically with fluconazole inhibiting all tested yeasts, reaching 100 % inhibition in *C. parapsilosis*. The inhibiting concentrations for the synergic pair ranged from 1.3 to 4.0 times below *Ca*Thi IC₅₀ and from zero to 2.0 times below fluconazole IC₅₀.

Conclusion: The results reported herein may ultimately contribute to future efforts aiming to employ this plant-derived AMP as a new therapeutic substance against yeasts.

Keywords: Antimicrobial peptides, Thionin, Synergistic activity, Fluconazole, *Candida*

Background

Currently, a significant global public health threat is the emergence of pathogenic bacteria, fungi, and yeasts that are resistant to multiple antimicrobial agents. Indeed, few or no effective chemotherapies are available for infections caused by some of these resistant microorganisms [1, 2].

Alternatives to chemotherapies include antimicrobial peptides (AMPs), small molecules produced by all living organisms, which have gained considerable attention

because of their potent antimicrobial activity against a broad range of microbes, including viruses, bacteria, protozoa, and fungi [3, 4]. Moreover, some kill microorganisms rapidly, are able to synergize with other AMPs and clinical antibiotics, have low toxicity to mammalian cells, and exert their microbial inhibitory activity at low concentrations. These molecules have multiple targets in the plasma membrane and also in intracellular components, which is thought to make an increase in microbial resistance more difficult [2, 5].

Promising AMPs include plant-derived thionins, a family of basic, low molecular weight (~5 kDa), cysteine-rich peptides. Various family members have high sequence similarity and structure [6–8]. Many of them

* Correspondence: valmg@uef.br

¹Laboratório de Fisiologia e Bioquímica de Microrganismos, Centro de Biociências e Biotecnologia, Universidade Estadual do Norte Fluminense, Campos dos Goytacazes 28013-602, RJ, Brazil

Full list of author information is available at the end of the article

are toxic against yeasts, pathogenic fungi, Gram-positive and Gram-negative bacteria, protozoa, and insects [7–10]. Like other AMPs, thionins' antimicrobial activity relies on their interaction with phospholipids to cause membrane instability [10].

Infections caused by *Candida* species have increased substantially over the last 30 years due to the rise of AIDS, ageing population, numbers of immunocompromised patients and the extensive use of indwelling prosthetic devices [1, 11]. *Candida albicans* is the main cause of candidiasis, however, other *Candida* species such as *C. tropicalis*, *C. parapsilosis*, and *C. glabrata* are now frequently identified as human pathogens [11–13]. Antifungals, especially fluconazole (FLC), have been used with some success for the treatment of *Candida* infections; however, there are numerous reports on the emergence of strains resistant to azoles that overexpress multidrug efflux transporters [14, 15].

In a previous report [8], our research team isolated a plant-derived thionin, named *CaThi*, with strong antimicrobial activity against two pathogenic *Candida* species, as well as *Escherichia coli* and *Pseudomonas aeruginosa*. FLC in combination with AMPs resulting in promising therapeutic results against important human pathogens, such as *C. albicans* and *Cryptococcus neoformans*, has been demonstrated [16, 17]. In this work we investigated whether the AMP *CaThi* could act synergistically with FLC. This synergistic strategy could result in a more efficient response against six *Candida* strains of clinical importance, avoiding the cytotoxic effects commonly exhibited by thionins against mammalian cells [10] by using low concentrations of this AMP. We were also interested in understanding the mechanism by which plant-derived thionins affect *Candida* species, which remains partially unknown [10]. These questions are addressed in the present study.

The results reported herein may ultimately contribute to future efforts aiming to develop this plant-derived AMP as a new therapeutic substance against these pathogenic *Candida* species as well as other yeast infections.

Results

Determination of IC₅₀ for *CaThi* and FLC

Initially we performed growth inhibition assays of six *Candida* species using different concentrations of FLC and thionin *CaThi* to determine the IC₅₀ of these substances. The lowest IC₅₀ for FLC was found for *C. buinensis* (0.125 µg.mL⁻¹) and the highest for *C. pelliculosa* (5 µg.mL⁻¹). In the case of *CaThi*, 10 µg.mL⁻¹ was sufficient to cause 50 % inhibition of growth of *C. albicans*, *C. tropicalis*, *C. parapsilosis*, and *C. buinensis* but 40 µg.mL⁻¹ was necessary to achieve IC₅₀ for *C. pelliculosa*. Thus, higher concentrations of both FLC and *CaThi* were needed to affect the growth of *C. pelliculosa*.

Moreover, although the antimicrobial activity of *CaThi* against *Candida* species is indeed relevant, our data showed it to be lower than that observed for FLC (Table 1).

Viability assay

CaThi induced viability loss in all yeasts cells tested (Fig. 1a). The most susceptible species to *CaThi* were *C. buinensis*, *C. parapsilosis* and *C. albicans* with 99.2, 98.9 and 80.3 % of viability loss, respectively, and the less susceptible was *C. tropicalis* with 47.9 % of viability loss (Fig. 1b). These results indicated that inhibitory effect of *CaThi* was candidicidal.

Plasma membrane permeabilization

Candida species cells were tested to determine the membrane permeabilization by Sytox green dye. All yeasts showed Sytox green fluorescence when grown for 24 h in the presence of *CaThi* IC₅₀. As with other AMPs, it is likely that *CaThi* acts on the plasma membrane of these *Candida* species, compromising it structurally and allowing the permeabilization of the labeling dye (Fig. 2). The membrane permeabilization percentage of the treated yeasts with *CaThi* was assessed (Table 2). A higher number of *C. albicans* and *C. pelliculosa* cells presented higher Sytox green fluorescence percentage, suggesting that *CaThi* is more effective at permeabilizing the membrane of these cells than the other *Candida* species analyzed.

ROS induction assay

Endogenous production of ROS was analyzed by incubating the yeasts for 24 h with *CaThi* IC₅₀. Increased ROS production was observed only in *C. tropicalis* (Fig. 3), suggesting that a *CaThi*-induced increase in oxidative stress may underlie the growth inhibitory effect on this yeast. Nevertheless, oxidative stresses were not detected for other *Candida* species, implicating that we could not associate the *CaThi* role and ROS production with growth inhibition of *Candida*, at least for the concentrations tested.

Table 1 IC₅₀^a (µg.mL⁻¹) of fluconazole and *CaThi* in different species of *Candida* respectively

Yeasts	Fluconazole	<i>CaThi</i>
<i>Candida albicans</i> (CE022)	1.0	10.0
<i>Candida tropicalis</i> (CE017)	1.0	10.0
<i>Candida parapsilosis</i> (CE002)	0.5	10.0
<i>Candida pelliculosa</i> (3974)	5.0	40.0
<i>Candida buinensis</i> (3982)	0.125	10.0
<i>Candida mogii</i> (4674)	2.5	20.0

^a represents the concentration of a drug that is required for 50 % inhibition *in vitro*








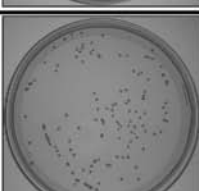
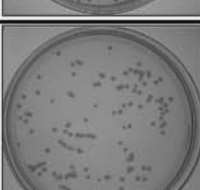
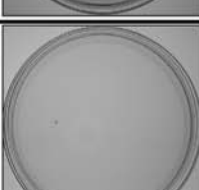
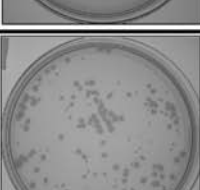
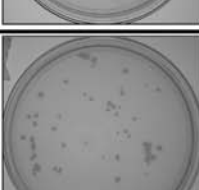
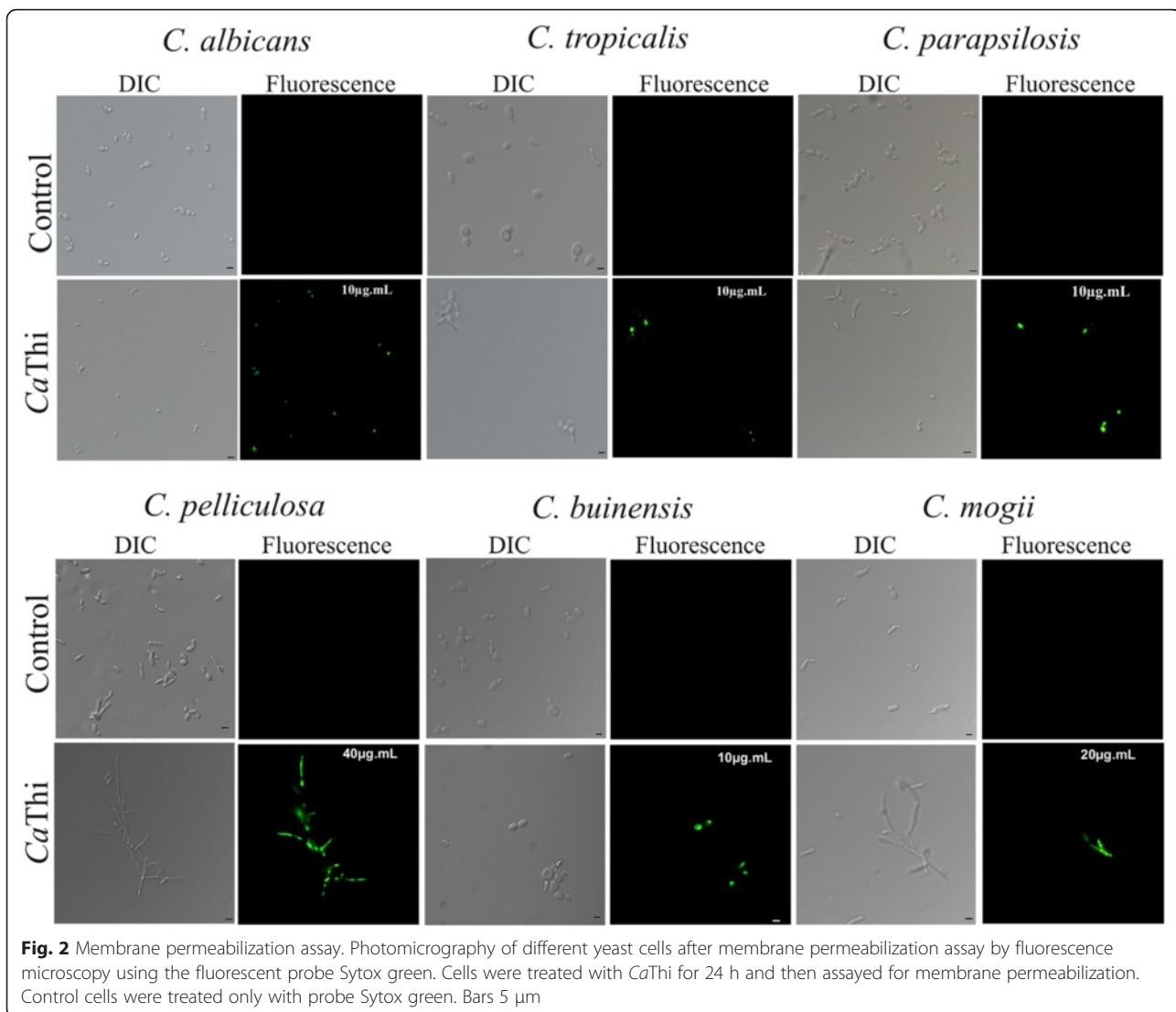
	A		B		
	Control	CaThi	Sample	CFU*	Viability loss (%)
<i>C. albicans</i>			control	395.3±20.7	0
			CaThi	77.6±3.7	80.3
<i>C. tropicalis</i>			control	344.3±28.9	0
			CaThi	179.3±5.5	47.9
<i>C. parapsilosis</i>			control	190.6±9.9	0
			CaThi	2.0±2.1	98.9
<i>C. pelliculosa</i>			control	439.3±33.3	0
			CaThi	130.3±9.2	70.2
<i>C. butinensis</i>			control	131.6±8.9	0
			CaThi	1.0±0.8	99.2
<i>C. mogii</i>			control	158.3±18.4	0
			CaThi	61.3±4.4	61.6

Fig. 1 Cell viability loss. **a** Photographs of the Petri dishes showing the viability of yeasts cells after the treatment with IC_{50} CaThi for 24 h. **b** The table shows the percentage of viability loss of yeasts cells after the treatment with IC_{50} CaThi for 24 h. CFU = Colony forming unit. (*) Indicates significance by the *T* test ($P < 0.05$) among the experiments and their respective controls. The experiments were carried out in triplicate

Localization of CaThi in yeast cells

We also investigated whether CaThi was actually internalized in *C. albicans* and *C. tropicalis* cells. These yeasts were chosen because they are known to be the most opportunistic pathogens among *Candida* species. Another important point is that *C. tropicalis* was the only yeast that presented membrane permeabilization and induction of ROS by CaThi in this work. To perform the test, we used $10 \mu\text{g}\cdot\text{mL}^{-1}$ of

FITC-tagged CaThi to search for intracellular signal fluorescence. We also treated the cells with DAPI for nuclei labeling. Intracellular signal fluorescence of CaThi-FITC was observed in both of these *Candida* species. However, while CaThi-FITC labeling of *C. tropicalis* produced a specific and intense spot of fluorescence inside the cells, *C. albicans* cells showed a more diffuse fluorescence. Overlapping of these CaThi-FITC images with DAPI nuclei labeling



indicated a co-localization of these fluorescent signals in *C. tropicalis* but not in *C. albicans* cells (Fig. 4). These data suggest that, at least for *C. tropicalis*, *CaThi* may have an intracellular target, possibly located in the nucleus.

Synergism assay

Given the increase in *Candida* infections, particularly among immunocompromised patients, searches for antifungal therapeutic alternatives are warranted. This concern and the aforementioned data prompted us to investigate whether FLC and *CaThi* could act synergistically to improve therapeutic results against *Candida* species. The combination of FLC and *CaThi* showed an increase in inhibitory activity of all of the *Candida* species tested, suggestive of synergistic activity (Table 3). Interestingly, although *C. pelliculosa* had the highest IC_{50} for both substances, when we combined FLC at one-fold

below its IC_{50} and *CaThi* at threefold below its IC_{50} , we observed 57 % increase in growth inhibition of this yeast. Similarly, in *C. parapsilosis* cells, when IC_{50} FLC was combined with *CaThi* threefold below its IC_{50} , we obtained 100 % growth inhibition of this yeast. Combined use of FLC and *CaThi* also strongly inhibited (96 %) *C. tropicalis*, although when used separately the inhibition achieved with these substances did not reach 12 %. Taken together, these data suggest that in combination FLC and *CaThi* could have an important synergistic action resulting in very effective control of *Candida* species.

Morphological alterations of *CaThi* and FLC plus *CaThi* on yeast growth

Investigation regarding the possible morphological alterations in yeast cells grown in the presence of FLC, *CaThi*, or a combination of both substances after the inhibition assays (Fig. 5a) was performed. Optical

Table 2 Fluorescent cell percentage of yeasts treated with CaThi^a

Yeasts species	Sample	Cell number viewed in DIC	Cell number viewed in fluorescence	% of fluorescence cells ^b
<i>C. albicans</i>	control	62.0 ± 9.3	0.6 ± 0.8	0.9
	CaThi	20.0 ± 5.0	16.6 ± 5.3	83.0
<i>C. tropicalis</i>	control	41.2 ± 4.2	0.8 ± 1.3	1.9
	CaThi	9.8 ± 4.9	4.8 ± 4.6	48.9
<i>C. parapsilosis</i>	control	79.2 ± 12.1	0	0
	CaThi	18.2 ± 5.4	6.2 ± 1.4	34.0
<i>C. pelliculosa</i>	control	23.8 ± 3.5	0	0
	CaThi	7.2 ± 1.9	6.6 ± 2.3	91.6
<i>C. buinensis</i>	control	43.6 ± 9.0	0.8 ± 1.3	1.8
	CaThi	13.6 ± 6.2	6.2 ± 1.3	45.5
<i>C. mogii</i>	control	18.6 ± 3.2	0.6 ± 0.8	3.2
	CaThi	8.6 ± 2.0	4.4 ± 2.7	51.1

^aCells number determination in five random fields of the DIC and fluorescence views of the samples obtained from Plasma membrane permeabilization assay. The total cell number in DIC of each yeast (in control and test) was assumed as 100 %

^bIndicates significance by the *T* test ($P < 0.05$) among the experiments and their respective controls

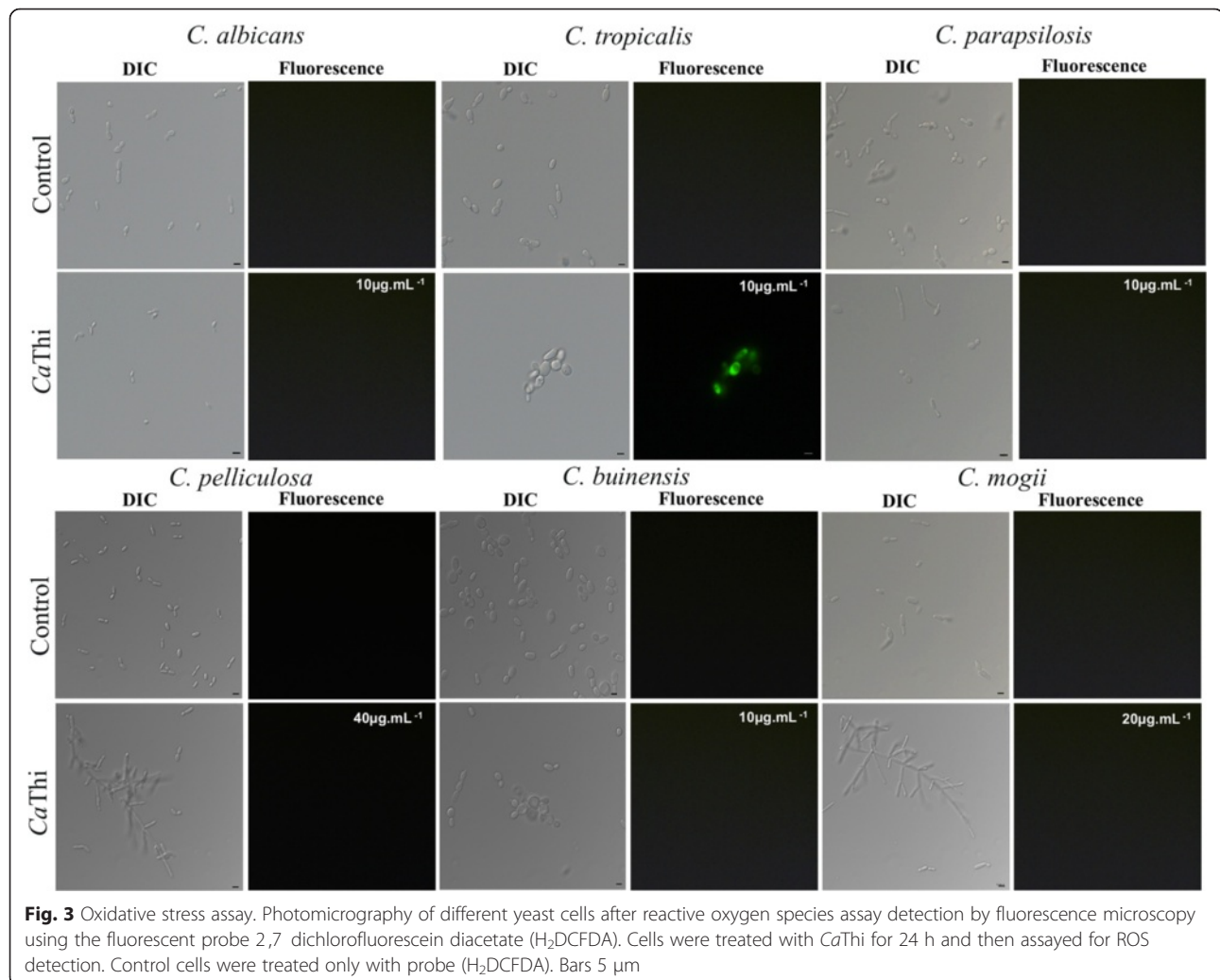


Fig. 3 Oxidative stress assay. Photomicrography of different yeast cells after reactive oxygen species assay detection by fluorescence microscopy using the fluorescent probe 2,7 dichlorofluorescein diacetate (H₂DCFDA). Cells were treated with CaThi for 24 h and then assayed for ROS detection. Control cells were treated only with probe (H₂DCFDA). Bars 5 µm

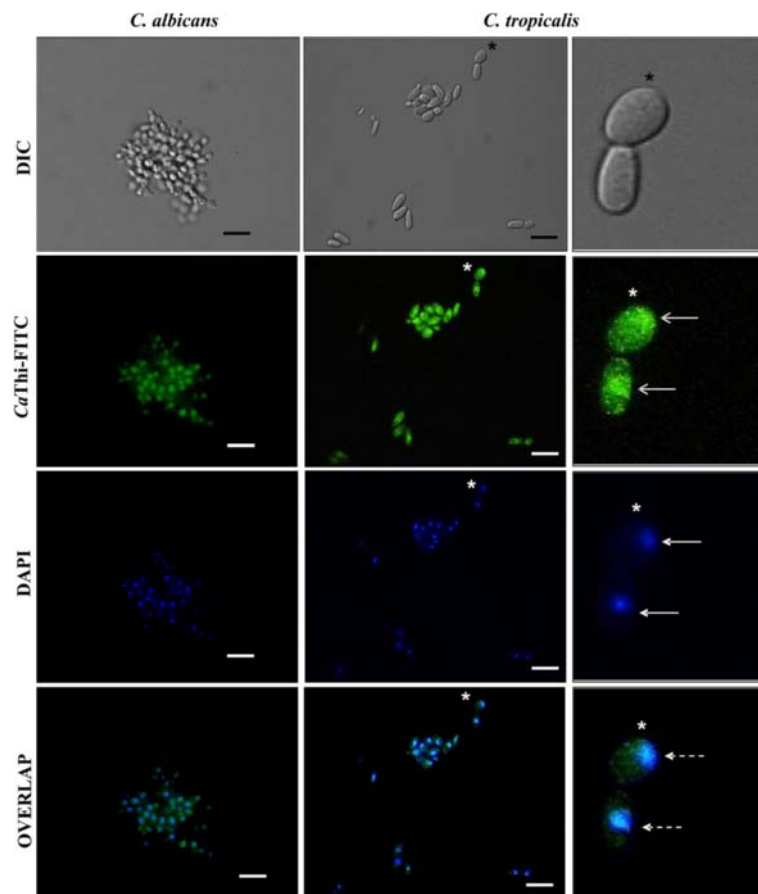


Fig. 4 Localization of *CaThi* in yeast cells. Photomicrography of *Candida albicans* and *Candida tropicalis* cells incubated for 24 h with $10 \mu\text{g}\cdot\text{mL}^{-1}$ *CaThi*-FITC (green fluorescence, open arrows) by fluorescence microscopy. Nuclei were visualized by 4,6-diamidino-2-phenylindole dihydrochloride (DAPI) after the *CaThi*-FITC incubation period (blue fluorescence, filled arrows). Overlap of the DAPI and FITC images (dotted arrows). Bars $20 \mu\text{m}$. (*) Indicates the position of digital enlargement

Table 3 Inhibition percentage of yeast species treated with *CaThi* and FLC alone and in combination showing synergism effect *in vitro*

Yeasts species	Sample	Concentration ($\mu\text{g}\cdot\text{mL}^{-1}$) ^a	Inhibition (%)	Combination inhibition (%) (<i>CaThi</i> + FLC) ^b
<i>C. albicans</i>	<i>CaThi</i>	3.5	2.93	77.5
	FLC	0.5	24.48	
<i>C. tropicalis</i>	<i>CaThi</i>	3.5	0	96.26
	FLC	0.5	11.55	
<i>C. parapsilosis</i>	<i>CaThi</i>	3.5	4.0	100.0
	FLC	0.5	50	
<i>C. pelliculosa</i>	<i>CaThi</i>	15.0	2.63	57.45
	FLC	2.5	7.8	
<i>C. buinensis</i>	<i>CaThi</i>	5.0	19.0	67.01
	FLC	0.06	5.45	
<i>C. mogii</i>	<i>CaThi</i>	10.0	17.19	61.05
	FLC	1.0	22.80	

^a*CaThi* concentrations ranging 1.3 to 4.0 times below its IC_{50} and FLC concentrations 2.0 times below its IC_{50} or at its IC_{50}

^bIndicates significance by the ANOVA test ($P < 0.05$) which were calculated by the absorbance values of synergism among the experiments and their respective controls

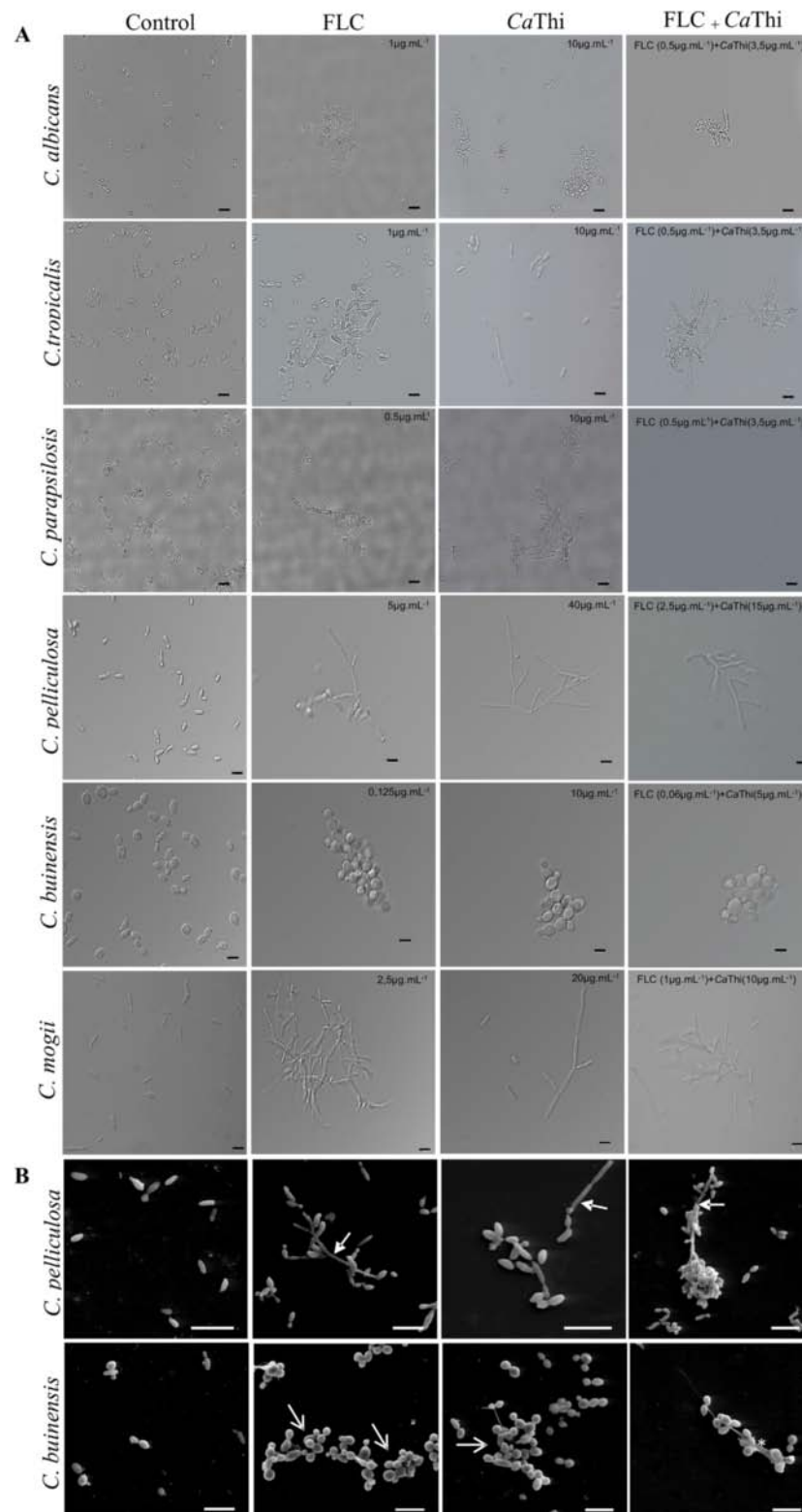


Fig. 5 Effect of CaThi, FLC, and CaThi plus FLC on yeast growth. **a** Photomicrography of *Candida albicans*, *Candida tropicalis*, *Candida parapsilosis*, *Candida pelliculosa*, *Candida buinensis*, and *Candida mogii* cells by light microscopy after the growth inhibition assay. Bars 5 μm . **b** Scanning electron microscopy of *Candida pelliculosa* and *Candida buinensis*. Filled arrow (formation of pseudohyphae); open arrow (cell agglomeration); asterisk (amorphous material). Bars 10 μm . Cells grown in the presence of Fluconazole (FLC), CaThi, and FLC plus CaThi

microscopy analysis revealed that FLC, CaThi, and the combined treatment caused changes in the morphology of cells of *Candida* species. *C. tropicalis*, *C. parapsilosis*, *C. pelliculosa*, and *C. mogii* exhibited an apparent difficulty in releasing buds thus leading to the formation of pseudohyphae when grown in the presence of CaThi. Moreover, in the presence of either substance, *C. tropicalis* cells presented hyper branching of pseudohyphae. For *C. albicans* and *C. buinensis*, an intense cellular agglomeration in all treatments was observed. Further, the combination of CaThi and FLC caused a shrunken appearance in *C. albicans* cells.

Scanning Electronic Microscopy (SEM) of *C. pelliculosa* reinforces the optical microscopy observations, corresponding to intense cell agglomeration and pseudohyphae formation in all treatments. For *C. buinensis*, all treatments showed intense cellular agglomeration and apparent difficulty in bud release, but not in the formation of pseudohyphae. For this yeast an amorphous material among cells in all treatments was also observed (Fig. 5b). These results show that CaThi is capable of causing morphological changes similar to FLC, an azole antifungal agent, commonly used in treatment of infections caused by *Candida* species. Importantly, we were able to demonstrate that the combination of these substances potentiates the therapeutic effects against these opportunistic species of *Candida*.

Discussion

Plant-derived thionins exhibit toxic effects against a wide range of plant pathogens including bacteria and fungi [18–21]. However, there is a gap regarding the mode of action of plant-derived thionins against human pathogens. Prompted by the considerable increase in the incidence of human infections by *Candida* species, we investigated the potential of CaThi, a plant-derived thionin peptide, as a novel therapeutic drug against six *Candida* strains of clinical interest: *C. albicans*, *C. tropicalis*, *C. parapsilosis*, *C. pelliculosa*, *C. buinensis*, and *C. mogii*.

The growth inhibition assay was not done with RPMI 1640 medium, which is generally indicated by Clinical and Laboratory Standards Institute guidelines, because it has in its composition a large amount of inorganic salts and it is well known in the literature that the presence of salts, such as sodium chloride and magnesium sulfate, can decrease the inhibitory activity of antimicrobial peptides since it is necessary electrostatic approximation of the peptide with the membrane of microorganisms and the presence of salts disrupts this initial interaction [22–24]. Therefore, as explained above, our growth inhibition assay was done with Sabouraud broth which is a common used medium to growth of fungi, including *Candida albicans*.

Our growth inhibition assays of the six *Candida* species tested revealed that $10 \mu\text{g}\cdot\text{mL}^{-1}$ was IC_{50} for CaThi to inhibit the growth of *C. albicans*, *C. tropicalis*, *C. parapsilosis*, and *C. buinensis* but $40 \mu\text{g}\cdot\text{mL}^{-1}$ was necessary to achieve IC_{50} for *C. pelliculosa* (Table 1), and this inhibitory effect was candidacidal inducing viability loss in all yeast cells tested (Fig. 1). Thi 2.1, a thionin from *Arabidopsis thaliana*, achieved 80 % inhibition of *C. albicans* with $2.5 \mu\text{g}\cdot\text{mL}^{-1}$ [25]. Although Thi 2.1 showed stronger antimicrobial activity against *C. albicans* than CaThi, these authors did not test it against non-*albicans* species. Thus, whether this thionin would affect other species with similar strength remains unknown.

Sytox green is a dye that only penetrates cells when the plasma membrane is structurally compromised. All yeast species tested showed Sytox green fluorescence (Fig. 2), however CaThi was more effective in plasma membrane permeabilization in *C. albicans* and *C. pelliculosa* (Table 2). Antimicrobial activity against the fungus *Neurospora crassa* by α -Hodothionin, isolated from barley seeds, also occurs via a mechanism involving membrane permeabilization, resulting in the inward flux of Ca^{2+} and K^{+} efflux and consequent potential membrane collapse [26]. Another plant-derived thionin isolated from *Viscum album*, named VtA₃, interacts with the plasma membrane of the fungus *Fusarium solani*, causing its permeabilization and thus inhibiting the growth of this microorganism [27]. Indeed, several studies suggest that most of the biological effects of thionins result from the interaction of these peptides with the target cell membrane. Three mechanisms have been proposed: formation of an ion-selective channel; formation of patches or carpets of peptides; and loss of membrane phospholipids [10].

AMPs have been demonstrated to play a direct role in membrane permeabilization, causing a loss of membrane potential [28]. As cells depend on membrane potential to fulfill their physiological functions, its restoration is mandatory and demands cellular energy. One possible consequence of this process is ROS generation by activated mitochondria [29]. Therefore, we analyzed whether this primary membrane-permeabilizing event in *Candida* species induced by CaThi was followed by oxidative stress. Interestingly, CaThi only induced production of ROS in *C. tropicalis* (Fig. 3). We speculate that CaThi binds to a specific domain of the *C. tropicalis* membrane, which triggers the increase in oxidative stress response through ROS production. However, further studies are needed to establish this. Reports show that increase in ROS production by the target organisms is a recurring mode of action employed by thionins and other AMPs [27, 30, 31]. Indeed, increased death of the fungus *Fusarium solani* subjected to VtA₃ was connected with boosted ROS production by these organisms

[27]. More recently, a peptide similar to thionin isolated from the beetle *Psacotheta hilaris* also provoked an increase in the levels of endogenous ROS in *C. albicans* [32]. Our study is the first to report an increase in ROS production by a plant-derived thionin as an antimicrobial mechanism against a human pathogen, *C. tropicalis*. Therefore, *CaThi* seems to employ a sophisticated mechanism to inhibit the growth of this opportunistic yeast involving not only membrane permeabilization but also induction of oxidative stress response.

Some AMPs are able to enter the cell, after the initial cell membrane interaction [33, 34]. Accordingly, the next experiments were designed to analyze whether *CaThi* was able to actively enter *C. albicans* and *C. tropicalis*. In the approach used, FITC-tagged *CaThi* was monitored by fluorescence microscopy. Because *CaThi* entered *C. albicans* and *C. tropicalis* cells, we suggest that a possible intracellular target for this thionin might be part of a complex mechanism responsible for the death of *Candida* species. FITC-tagged *CaThi* overlapped with DAPI staining, indicating that, in *C. tropicalis*, this target is nuclear (Fig. 4). Giudici et al. [27] showed that VtA₃ entered and accumulated in the fungus *F. solani*. These authors also demonstrated that this entry was related to the sphingolipid composition of the plasma membrane of this fungus. Our study is the first to show that a plant-derived thionin (*CaThi*) is able to enter human pathogens (*C. albicans* and *C. tropicalis*) and to suggest an intracellular target for it. Our work opens new perspectives regarding the antimicrobial mechanism of plant-derived thionins as it suggests that these peptides' toxicity may not be restricted to the plasma membrane.

The real mode of action of AMPs has not been fully elucidated, but much of the described AMPs to date target the plasma membrane of microorganisms causing pore formation and leading to imbalance in cellular homeostasis [33, 35]. However, some studies showed that not only is permeabilization the cause of a particular microorganism death, as they may have multiple targets [36] after the interaction with the membrane causing, for example, ROS induction [31, 37]; inhibition of protein synthesis [38, 39]; inhibition of mitochondrial activity [40, 41], and also may trigger signaling cascades that lead to apoptosis [42, 43]. Thus it is difficult to identify the most important factor for the candidicidal effect of *CaThi* and is technically challenging to characterize the steps leading up to cell death, however evidence supports that all events described in the manuscript may have a crucial role in the death of the tested yeasts.

The continuous emergence of resistance of fungal strains to conventional antibiotics and antifungals,

especially *Candida* species, has become an important medical issue and has spurred the demand for new therapeutic alternatives. This concern prompted us to investigate whether FLC and *CaThi* could act synergistically to improve therapeutic results against *Candida* species. Here we show that the combination of these two substances was effective against all *Candida* species tested (Table 3), causing drastic morphological changes in these cells (Fig. 5). Interestingly, we show that the inhibitory effect of this combination was more effective for *C. albicans*, *C. parapsilosis*, and *C. tropicalis*, which are the major yeast species recovered from infections in immunocompromised patients [13]. The azoles mode of action occurs by inhibition of the enzyme lanosterol 14 α -demethylase, blocking ergosterol incorporation and leading to the accumulation of intermediate sterols. These intermediate sterols do not have the same configuration and physical properties of ergosterol, therefore they cause the plasma membrane to form with altered properties, changing in fluidity, permeability and impairing nutrient uptake, which ultimately lead to cell toxicity [44, 45]. In regard to the synergistic effect of FLC and *CaThi*, we hypothesize that permeabilization is firstly caused by *CaThi* (Fig. 2) facilitating the entrance of FLC into the cell cytoplasm, triggering structural alterations in the plasma membrane which feedback positively to the entrance of more *CaThi* and FLC. This entrance creates potential for toxic effects, which were experimentally observed in the lower IC₅₀ used for both substances in the combinatory treatment (Table 3). Additionally, secondary toxicity effects were demonstrated, such as the induction of ROS to *C. tropicalis*, the *CaThi* presence in *C. albicans* cell cytoplasm and in *C. tropicalis* nuclei. These localizations suggest that *CaThi* may have cytoplasmic targets, where interference could consolidate the inhibitory effect. However, more studies are necessary to clearly unravel the antimicrobial mechanism of *CaThi* against *Candida* species as well as the mechanism of synergy between *CaThi* and FLC.

Conclusions

Investigating a plant-derived thionin mode of action against opportunistic human pathogenic yeasts is relevant and advisable, whereas most studies involving plant-derived thionins focus their effects against plant pathogenic microorganisms as experimental models. In this report, we demonstrated that *CaThi* has strong candidicidal activity against six pathogenic *Candida* species, and it works by permeabilizing the membrane and inducing oxidative stress response in these yeasts. Additionally, we present evidence to suggest a nuclear intracellular target for *CaThi*. Finally, our results show that FLC and *CaThi* combined causes dramatic morphological changes in these yeasts, effective against all

Candida species tested. The combined treatment of *Ca*Thi and FLC is a strong candidate for clinical studies aiming to improve therapeutic results against resistant strains of *Candida* species. Studies involving drug combinations should be reinforced due to the possibility of synergistic effects that increase the toxic effect of the drugs combined when compared to monotherapy. Moreover, drug combinations can broaden the spectrum of antimicrobial activity, minimizing resistant microorganisms selection, increasing security and tolerance using lower drugs doses.

Methods

Biological materials

Capsicum annuum L. fruits (accession UENF1381) were provided by *Laboratório de Melhoramento Genético Vegetal*, from *Centro de Ciências e Tecnologias Agropecuárias*, *Universidade Estadual do Norte Fluminense Darcy Ribeiro* (UENF), Campos dos Goytacazes, Rio de Janeiro, Brazil.

The yeasts *Candida albicans* (CE022), *Candida tropicalis* (CE017), and *Candida parapsilosis* (CE002) were obtained from *Departamento de Biologia*, *Universidade Federal do Ceará*, Fortaleza, Brazil. The yeasts *Candida pelliculosa* (3974), *Candida buinensis* (3982), and *Candida mogii* (4674) were obtained from *Micoteca URM* from *Universidade Federal de Pernambuco*, Recife, Pernambuco, Brazil. Yeasts were maintained on Sabouraud agar (1 % peptone, 2 % glucose, and 1.7 % agar-agar) (Merck) in the *Laboratório de Fisiologia e Bioquímica de Microrganismos*, from *Centro de Biociência e Biotecnologia*, UENF, Campos dos Goytacazes, Rio de Janeiro, Brazil.

*Ca*Thi

Extraction and purification of the thionin *Ca*Thi from *Capsicum annuum* fruits by chromatographic methods were performed as described by Taveira et al. [8]. The retention time to recover the thionin *Ca*Thi during the reversed-phase chromatography in column μ RPC C2/C18 (ST 4.6/100) (GE Healthcare) was 37.87 min.

Preparation of yeast cells and determination of IC_{50} for *Ca*Thi and fluconazole

For the preparation of yeast cell cultures, an inoculum from each stock of *Candida albicans*, *Candida tropicalis*, *Candida parapsilosis*, *Candida pelliculosa*, *Candida buinensis*, and *Candida mogii* was transferred to Petri dishes containing Sabouraud agar and allowed to grow at 30 °C for 48 h. After this time, each cell aliquot was added to 10 mL sterile culture medium (Sabouraud broth, Merck). The cells were quantified in a Neubauer chamber (Optik Labor) with the aid of an optical microscope (Axiovision 4, Zeiss). The assay for checking the

growth inhibition of yeast cells was performed according to Broekaert et al. [46] with modifications. Initially yeast cells (1×10^4 cells mL^{-1}) were incubated in Sabouraud broth containing *Ca*Thi at concentrations ranging from 100 $\mu g \cdot mL^{-1}$ to 1 $\mu g \cdot mL^{-1}$ and fluconazole (FLC) at concentrations ranging from 20 $\mu g \cdot mL^{-1}$ to 0.125 $\mu g \cdot mL^{-1}$, with the final volume adjusted to 200 μL . The assay was performed in 96-well microplates (Nunc) at 30 °C for 24 h. Optical readings at 620 nm were recorded at zero hour and every 6 h interval for 24 h. Control cells were grown in the absence of *Ca*Thi and FLC. The optical densities were plotted against the concentration of *Ca*Thi and FLC, and then the concentration of the drug (*Ca*Thi and FLC) required for 50 % inhibition (IC_{50}) *in vitro* of the tested yeasts was determined. Experiments were performed in triplicate.

Viability assay

To assay the effect of *Ca*Thi on the cell viability of yeasts, 1×10^4 cells mL^{-1} in Sabouraud broth culture medium and at the corresponding IC_{50} of *Ca*Thi values for each yeast were incubated at 30 °C for 24 h in 96-well microplates (Nunc). To determine the control viability, the control cells (without *Ca*Thi) were washed once and diluted 1,000-fold in Sabouraud broth culture medium, and an aliquot of 100 μL from this dilution was spread over the surface of a Sabouraud agar medium (contained in a Petri dish) with a Drigalski loop and grown at 30 °C for 48 h. At the end of this period, colonies forming units (CFU) were determined for each yeast species, and the Petri dishes were photographed. The same procedure was followed with yeasts treated with *Ca*Thi. The experiments were carried out in triplicate, and the results are shown assuming that the control represents 100 % viability. Calculations of the standard deviation and *T* test were performed with Prism software (version 5.0).

Plasma membrane permeabilization assay

The plasma membrane permeabilization of yeast cells was measured by Sytox green uptake, according to the methodology described by Thevissen et al. [47], with some modifications. Each of the different species of yeasts was incubated with *Ca*Thi at the concentration required to inhibit 50 % growth (IC_{50}) of the respective yeast cells for 24 h. After this time, a 100 μL aliquot of each yeast cell suspension was incubated with 0.2 μM of Sytox green in 1.5 mL microcentrifuge tubes for 30 min at 25 °C with constant agitation. Cells were analyzed by DIC optical microscope (Axiovision 4, Zeiss) equipped with a fluorescent filter set for detection of the fluorescein (excitation wavelength, 450–490 nm, emission wavelength 500 nm). To indicate membrane permeabilization, the percentage of fluorescent cells was determined by counting the DIC and fluorescent views

for each yeast ($n = 5$). The total cell number in DIC view of each yeast (in control and test) was assumed as 100 %. The experiments were carried out in triplicate. Calculations of T test were performed with Prism software (version 5.0).

Determining the induction of intracellular ROS in yeast cells

To evaluate whether the mechanism of action of *Ca*Thi involves induction of oxidative stress, the fluorescent probe 2', 7'-dichlorofluorescein diacetate (H_2DCFDA) was used to measure intracellular reactive oxygen species (ROS) according to the methodology described by Mello et al. [31]. Each of the different species of yeasts was incubated with the respective IC_{50} for *Ca*Thi in 96-well microplates for 24 h at 30 °C; after this incubation an aliquot of 50 μ L of each of yeast cell suspension was incubated with 200 μ M of H_2DCFDA in microcentrifuge tubes of 1.5 mL for 1 h at 25 °C with constant agitation at 500 rpm. Cells were analyzed by DIC optical microscope (Axiovision 4, Zeiss) equipped with a fluorescent filter set for detection of the fluorescein (excitation wavelength, 450–490 nm, emission wavelength 500 nm). The experiments were carried out in triplicate.

*Ca*Thi conjugated to FITC localization for optical microscopy

*Ca*Thi at 100 μ g was resuspended in 100 μ L of 750 mM sodium carbonate-sodium bicarbonate buffer, pH 9.5 containing FITC at 50 μ g.mL⁻¹ (previously solubilized in DMSO). This solution was incubated with constant agitation at 500 rpm for 2 h at 30 °C. After this incubation, the sample was submitted to gel filtration chromatography on Sephadex G25 column (Sigma) for elimination of free FITC and recovery *Ca*Thi-FITC. The column was equilibrated and run with 20 mM Tris-HCl, pH 8.0 at flow rate of 0.3 mL.min⁻¹. After coupling, 10 μ g.mL⁻¹ of *Ca*Thi-FITC was incubated with cells of *C. albicans* and *C. tropicalis* for 24 h in 96-well microplates. After this time an aliquot of each cell suspension was removed and incubated with 50 μ g.mL⁻¹ of 4',6-diamidino-2-phenylindole dihydrochloride (DAPI) for 10 min for nuclei stain. Cells were analyzed by DIC optical microscope (Axiovision 4, Zeiss) equipped with a fluorescent filter set for detection of the fluorescein (excitation wavelength, 450–490 nm, emission wavelength 500 nm). The entire assay was performed protected from light.

Synergism assay

To verify the synergistic activities, we combined FLC with *Ca*Thi. Initially yeast cells (1×10^4 cells mL⁻¹) were incubated in Sabouraud broth containing an IC_{50}

concentration or less than that for FLC and *Ca*Thi concentrations ranging from 1.3 to 4.0 times below the IC_{50} for the respective yeast and the final volume adjusted to 200 μ L *in vitro*. The assay was performed in 96-well microplates (Nunc) at 30 °C for 24 h. Optical readings at 620 nm were taken at zero hour and every 6 h for the following 24 h. Control cells were: 1) grown in the absence of *Ca*Thi and FLC; 2) grown in the presence of FLC; 3) grown in the presence of *Ca*Thi. The synergistic activity was deduced comparing optical densities of each control and combined drugs (FLC plus *Ca*Thi) considering each yeast strain tested. We define synergism as the combination action of the AMP with other substance that causes an enhanced decrease in the growth of the microorganism, compared with the growth inhibition of the single substances. After synergism, assay cells (controls and tests) were analyzed by DIC optical microscope (Axiovision 4, Zeiss). The data were obtained from experiments performed in triplicate. The data were evaluated using a one-way ANOVA. Mean differences at $p < 0.05$ were considered to be significant. All statistical analyses were performed using the GraphPad Prism software (version 5.0 for Windows).

Scanning electron microscopy

C. pelliculosa and *C. buinensis* cells were submitted to scanning electron microscopy (SEM) analysis. Yeast cells were grown for 24 h in Sabouraud broth in the presence of FLC (5 μ g.mL⁻¹ and 0.125 μ g.mL⁻¹, respectively), *Ca*Thi (40 μ g.mL⁻¹ and 10 μ g.mL⁻¹, respectively), and FLC plus *Ca*Thi (2.5 μ g.mL⁻¹ + 15 μ g.mL⁻¹, 0.06 μ g.mL⁻¹ + 5 μ g.mL⁻¹, respectively) or absence of these drugs, then were fixed for 30 min at 30 °C in a solution containing 2.5 % glutaraldehyde and 4.0 % formaldehyde in 0.1 M cacodylate buffer, pH 7.0. Subsequently, the cells were rinsed three times in 0.1 M cacodylate buffer, pH 7.0; post-fixed for 30 min at 30 °C with 1.0 % osmium tetroxide diluted in 0.1 M cacodylate buffer, pH 7.0; and rinsed again with this same buffer. The yeast cells were gradually dehydrated in alcohol solution (15, 30, 50, 70 to 90 % and finally 100 % alcohol), critical point dried in CO₂, covered with 20 nm gold and observed in a DSEM 962 Zeiss SEM.

Abbreviations

AMP: Antimicrobial peptide; FLC: Fluconazole; ROS: Reactive oxygen species; *Ca*Thi: *Capsicum annuum* Thionin; DIC: Differential interference contrast; FITC: Fluorescein isothiocyanate; SEM: Scanning Electron Microscopy.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

Conceived and designed the experiments: GBT, AOC, RR, FGT, MC and VMG. Performed the experiments: GBT and FGT. Analyzed the data: GBT, AOC, FGT, MC and VMG. Contributed reagents/materials/analysis tools: AOC, RR, MC

and VMG. Wrote the paper: GBT, AOC and VMG. All authors read and approved the final manuscript.

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Author details

¹Laboratório de Fisiologia e Bioquímica de Microrganismos, Centro de Biotecnologia, Universidade Estadual do Norte Fluminense, Campos dos Goytacazes 28013-602, RJ, Brazil. ²Laboratório de Melhoramento Genético Vegetal, Centro de Ciências e Tecnologias Agropecuárias, Universidade Estadual do Norte Fluminense, Campos dos Goytacazes 28013-602, RJ, Brazil. ³Laboratório de Biologia Celular e Tecidual, Centro de Biotecnologia, Universidade Estadual do Norte Fluminense, Campos dos Goytacazes 28013-602, RJ, Brazil.

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