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Sigma-1 receptor expression in sensory neurons and the effect of painful peripheral nerve injury

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Abstract

Background: The sigma-1 receptor (σ 1R), an endoplasmic reticulum chaperone protein, is widely distributed and regulates numerous intracellular processes in neurons. Nerve injury alters the structure and function of axotomized dorsal root ganglion (DRG) neurons, contributing to the development of pain. The σ 1R is enriched in the spinal cord and modulates pain after peripheral nerve injury. However, σ 1R expression in the DRG has not been studied. We therefore characterized σ 1R expression in DRGs at baseline and following spinal nerve ligation (SNL) in rats.

Results: Immunohistochemical (IHC) studies in DRG sections show σ 1R in both neuronal somata and satellite glial cells. The punctate distribution of σ 1R in the neuronal cytoplasm suggests expression in the endoplasmic reticulum. When classified by neuronal size, large neurons (>1300 μ m) showed higher levels of σ 1R staining than other groups (700-1300 μ m, <700 μ m). Comparing σ 1R expression in neuronal groups characterized by expression of calcitonin gene-related peptide (CGRP), isolectin-B4 (IB4) and neurofilament-200 (NF-200), we found σ 1R expression in all three neuronal subpopulations, with highest levels of σ 1R expression in the NF-200 group. After SNL, lysates from L5 DRGs that contains axotomized neurons showed decreased σ 1R protein but unaffected transcript level, compared with Control DRGs. IHC images also showed decreased σ 1R protein expression, in SNL L5 DRGs, and to a lesser extent in the neighboring SNL L4 DRGs. Neurons labeled by CGRP and NF-200 showed decreased σ 1R expression in L5 and, to a lesser extent, L4 DRGs. In IB4-labeled neurons, σ 1R expression decreased only in axotomized L5 DRGs. Satellite cells also showed decreased σ 1R expression in L5 DRGs after SNL.

Conclusions: Our data show that σ 1R is present in both sensory neurons and satellite cells in rat DRGs. Expression of σ 1R is down-regulated in axotomized neurons as well as in their accompanying satellite glial cells, while neighboring uninjured neurons show a lesser down-regulation. Therefore, elevated σ 1R expression in neuropathic pain is not an explanation for pain relief after σ 1R blockade. This implies that increased levels of endogenous σ 1R agonists may play a role, and diminished neuroprotection from loss of glial σ 1R may be a contributing factor.

Keywords: Sigma-1 receptor, Neuropathic pain, Peripheral nerve injury, Endoplasmic reticulum, Sensory neuron, Dorsal root ganglion

Background

Sigma-1 receptor (σ 1R), a non-opioid receptor residing at the mitochondria-associated endoplasmic reticulum (ER) membrane [1], is widely distributed in various tissues including heart, liver, immune cells, as well as the nervous system [2-6]. Cellular functions are regulated by σ 1R binding through a wide range of signaling pathways including protein kinases, ion channels, and transcription factors [7-10]. The σ 1Rs are abundant in the central nervous

system, where they may contribute to psychostimulant addiction [11,12], neurodegenerative disorders [13], neuroinflammation [14] and neuroprotection [15,16]. We have previously demonstrated that the σ 1R mediates (+)-morphine induced anti-analgesia through actions at both supraspinal and spinal sites [17-19]. Other reports show that opioid ligand-induced antinociception is attenuated by σ 1R agonists and potentiated by σ 1R antagonists [20-23]. Further, σ 1R antagonists effectively attenuate painful behavior in neuropathic and inflammatory pain models [24-29]. Although much remains to be learned about the role of the σ 1R in health and disease, these various findings together point toward a significant role in pain modulation.

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Peripheral nerve injury initiates multiple structural and functional changes in peripheral, spinal cord, and supraspinal sites that contribute to long-term neuropathic pain [30-34]. Our previous findings demonstrate that injured sensory neurons have diminished voltage-gated Ca^{2+} influx and reduced intracellular Ca^{2+} stores [35-37]. These features are of particular relevance to a potential role of σ 1R in neuropathic pain since store depletion activates σ 1R [1,38], and σ 1R activation reduces I_{Ca} in other types of neurons [6,39]. Furthermore, the dorsal root ganglion (DRG), site of the somata of peripheral sensory neurons, generates neurosteroids that are σ 1R ligands [40-42]. Although this evidence suggests that peripheral sensory neuron σ 1Rs may contribute to the complex of changes that result in chronic neuropathic pain, expression of σ 1Rs in this neuronal population has not been explored. In the present study, we therefore characterize the expression and distribution of σ 1R in sensory neurons. Since elevated σ 1R expression could contribute to the generation of neuropathic pain, we also examine the influence of painful nerve injury. We found that σ 1Rs are present in both neurons and satellite glial cells (SGCs) in the DRG, and that σ 1Rs are down-regulated after peripheral nerve injury.

Results

A total of 31 rats were used in the present study, of which 16 were Control animals and 15 had painful neuropathy induced by spinal nerve ligation (SNL). All SNL animals demonstrated hyperalgesic responses, whereas no animals in the Control group did. The frequency of hyperalgesia responses from noxious punctate mechanical stimulation in SNL rats ($32 \pm 3\%$) was greater than in control rats ($0 \pm 0\%$; $P < 0.001$). All SNL animals used in this study had greater than 20% hyperalgesia responses (at least 6 responses to a total of 30 pin stimulations from 3 sensory test sessions). Within a single testing session, we see no pattern of accumulating sensitivity or accommodation to the stimuli. The anatomical accuracy of the SNL surgery was confirmed at the time of tissue harvest in all SNL animals.

Both DRG neurons and satellite cells exhibit σ 1R immunofluorescence

We first used immunohistochemical (IHC) methods to identify if σ 1R protein is present in the DRG. Fluorescent images of σ 1R staining in DRG sections revealed the presence of σ 1R in neuronal somata in a punctate distribution in the cytoplasm (Figure 1A,B), consistent with its location in the ER [1,43]. Double staining for σ 1R combined with NeuN, a neuron-specific marker, revealed expression of σ 1R in all neurons (Figure 1A). Using glutamine synthetase [44,45] as a marker for astrocyte-like SGCs [46], sections also demonstrated expression of σ 1R in SGCs (Figure 1B). This finding is consistent with prior reports of the presence

of σ 1R in glial cells such as astrocytes and Schwann cells [3,47]. When classified into different neuronal subpopulations by size, large neurons comparatively higher levels of σ 1R expression (Figure 1C). This suggests that there may be neuron subtype-specific expression of σ 1R. Accordingly, we compared σ 1R expression in neuronal groups characterized by expression of CGRP, a marker of small unmyelinated peptidergic primary sensory neurons, by binding of IB4, a marker of small unmyelinated non-peptidergic sensory neurons, and by expression of NF200, a marker of large, myelinated neurons [48,49]. This analysis confirmed comparatively higher σ 1R expression in the NF200 group (Figure 1D, E), consistent with their larger size.

Effect of injury on σ 1R Expression in DRG

Transcripts for σ 1R have previously been found in neuronal tissues [50,51], but their presence has not been examined in peripheral sensory neurons. We identified σ 1R at the transcript level in DRG lysates (Figure 2A), confirming the IHC findings of its expression there. As an initial examination of the effect of injury on sensory neuron σ 1R expression, we also examined lysates from animals that developed painful neuropathy following SNL. Examination of transcript level (Figure 2A) did not show any significant changes. However, Western immunoblotting of lysates using a commercially available antibody (Invitogen) demonstrated significantly down-regulated σ 1R protein expression in SNL L5 DRGs compared to Control, while neighboring SNL L4 DRGs showed an intermediate level of σ 1R protein expression (Figure 2B,C). These Western findings were duplicated using another σ 1R antibody (gift from Drs. T. P. Su and T. Hayashi at NIDA; data not shown).

Effect of injury on σ 1R immunofluorescence in subpopulations of DRG neurons

To further evaluate the effect of injury on σ 1R expression, we quantified IHC images from DRG sections following SNL (Figure 3A). Immunofluorescent intensity of σ 1R was attenuated in both axotomized SNL L5 DRGs and, to a lesser extent, in the neighboring SNL L4 DRG (Figure 3B). The finding is consistent with our Western blotting result (Figure 2), in which σ 1R protein expression decreased more in SNL L5 than in SNL L4 DRGs. This pattern was found in small and medium size groups (Figure 3C), although large neurons failed to show an effect on the SNL L5 group, which were few in number.

For the neuronal subpopulation labeled with CGRP (Figure 4A), we found that σ 1R expression decreased not only in SNL L5 but also in neighboring uninjured L4 DRG neurons, compared to Control (Figure 4B). IB4 showed plasma membrane and perinuclear staining (Figure 5A), consistent with a previous description [52]. Double staining for σ 1R with IB4 revealed that expression of σ 1R decreased in SNL L5 DRGs but not in SNL

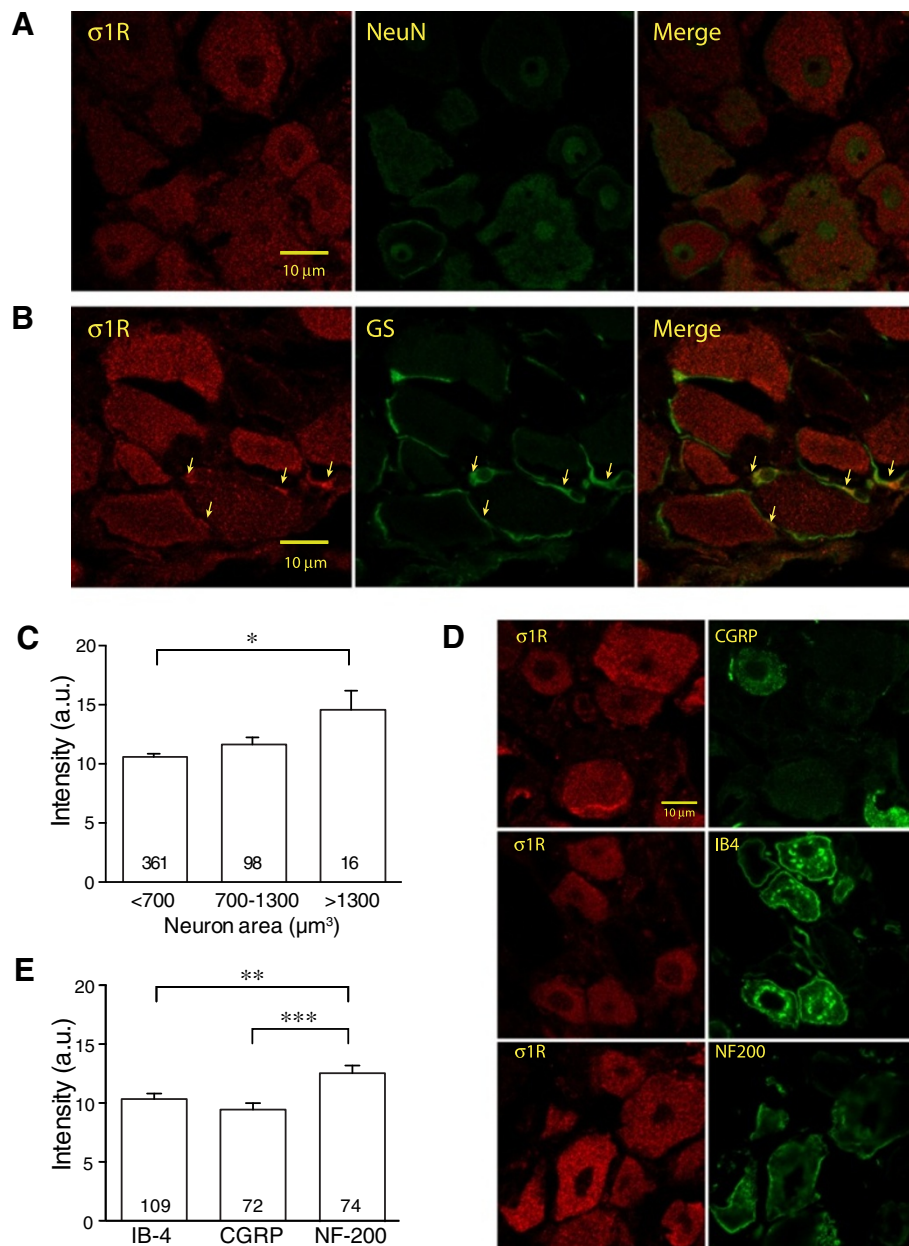


Figure 1 σ 1 Receptor (σ 1R) in various Control dorsal root ganglion (DRG) cell populations. (A) Staining for σ 1R was uniformly distributed within cytoplasmic areas of cells identified as neuronal profiles through co-staining with NeuN. No specific σ 1R staining was found in neuronal nuclei. **(B)** σ 1R was also found in cellular components identified as satellite glial cells by their expression of glutamine synthase (GS). Identically placed arrows in the left and middle panels indicate cellular areas clearly identifiable as satellite glial cell cytoplasm that expresses σ 1R. **(C)** Classification of control DRG neurons by size (<700, 700-1300 and >1300 μ m³) showed greater σ 1R intensity (arbitrary units, recorded using standardized image acquisition) in large neurons compared to others. **(D)** σ 1Rs expression is compared in neuronal subtypes identified by their expression of calcitonin-gene related peptide (CGRP), isolectin B-4 (IB4), and neurofilament-200 (NF-200). **(E)** Higher fluorescent intensity for σ 1Rs was found in the subpopulation of control neurons expressing NF-200. Mean \pm SEM; number in bars represents the sample size; * p < 0.05, ** p < 0.01, *** p < 0.001.

L4 neurons (Figure 5B). Double staining for σ 1R with NF200 (Figure 6A) showed decreased σ 1R expression in both L4 and L5 DRGs after SNL, compared to Control (Figure 6B).

Effect of injury on σ 1R immunofluorescence in DRG satellite glial cells

Using immunostaining of glutamine synthetase to detect satellite glial cells, we observed diminished σ 1R

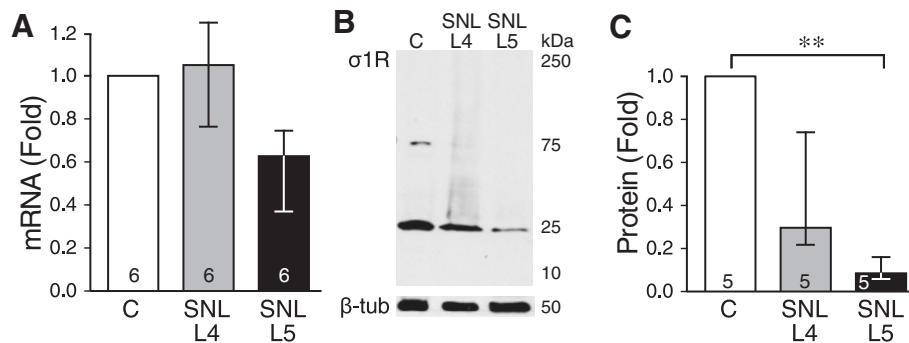


Figure 2 Effect of injury on σ_1 Receptor (σ_1 R) expression in dorsal root ganglia (DRG). **(A)** Transcript measured by quantitative real time PCR showed no significant differences in σ_1 R level after spinal nerve ligation (SNL) in both the axotomized 5th lumbar (L5) dorsal root ganglion and the adjacent 4th lumbar (L4) ganglion, compared to control (C) ganglia. **(B)** Protein measurement by Western blot showed a dominant band at the expected molecular weight that was decreased in both L4 and L5 ganglia after SNL compared to control DRGs. β -Tubulin 1 (β -Tub) was used as loading control. **(C)** Summary data of Western blot showed a significant decrease of σ_1 R protein expression in axotomized L5 DRG. Both graphs show Median and interquartile range for fold difference from normalized controls; numbers in bars represents the sample size; $**p < 0.01$.

immunofluorescent intensity within the satellite glial cells of the L5 DRG following SNL (Figure 7A,B).

Discussion

Several key findings emerge from our examination of σ_1 R expression in control and injured sensory neurons.

We demonstrated that σ_1 Rs were present in both sensory neurons and SGCs. In line with previous reports [5,43,53,54], we found that σ_1 Rs are dispersed throughout the cytoplasm of DRG neurons in all size and histochemical subgroups. Expression of the σ_1 R protein decreases in both axotomized L5 neurons as well as

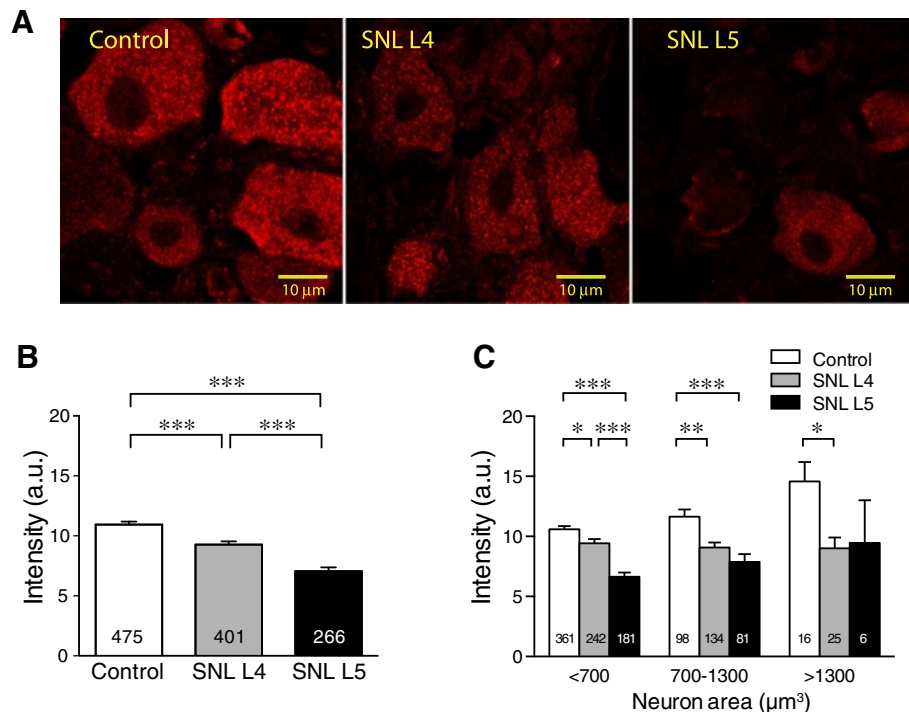


Figure 3 Effect of injury on σ_1 receptor (σ_1 R) expression in sensory neurons. **(A)** Decreases of σ_1 receptor (σ_1 R) immunofluorescence intensity were found in both spinal nerve ligation (SNL) 4th lumbar (L4) and 5th lumbar (L5) neurons compared with that in Control animals. **(B)** Summary data showed decrease of intensity of staining in SNL L4 and L5 neurons. Number in bars represents the n for neurons. **(C)** Classified by neuronal size (<700, 700-1300 and >1300 μm^3), SNL L4 DRG neurons showed lower σ_1 R intensity (arbitrary units, recorded using standardized image acquisition) than Control in all three size groups, and SNL L5 neurons in the small and medium size groups showed lower σ_1 R intensity than Controls. The data for Control groups are the same as shown in Figure 1C. Mean \pm SEM; $*p < 0.05$, $**p < 0.01$, $***p < 0.001$.

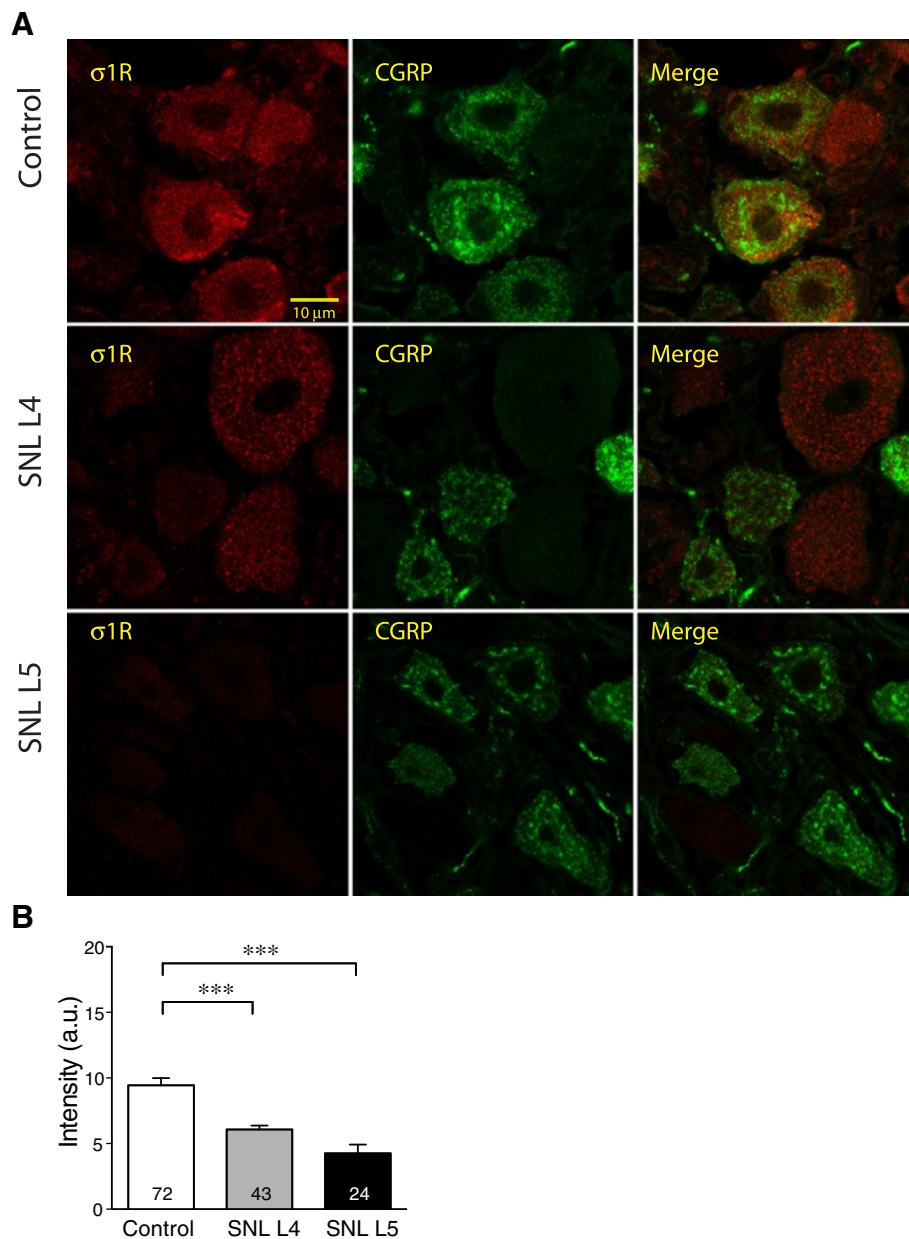


Figure 4 Effect of injury on $\sigma 1$ receptor ($\sigma 1R$) expression in sensory neurons identified by calcitonin-gene related peptide (CGRP) staining. (A) Staining of $\sigma 1R$ was distributed within cytoplasmic areas of all CGRP co-expressing neurons. $\sigma 1R$ immunofluorescence intensity was decreased in both spinal nerve ligation (SNL) 4th lumbar (L4) and 5th lumbar (L5) neurons compared with Control. **(B)** Summary data showed decrease of intensity of $\sigma 1R$ staining in CGRP-positive SNL L4 and L5 neurons. Mean \pm SEM; number in bars represents the sample size; *** $p < 0.001$.

neighboring L4 neurons, and this decrease occurred in all neuronal subpopulations. These observations confirm the presence of $\sigma 1R$ in the DRG, which was previously suspected on the basis of ligand binding studies [53,55]. Furthermore, our findings also support a role for primary sensory neuron $\sigma 1R$ s in pain generation after injury.

In the present study, we determined the average $\sigma 1R$ expression in relevant DRG neuronal subgroups and the

effect of SNL. In prior IHC analysis by Hammond et al. [52], it was shown that SNL also affects the number of neurons in these subgroup populations. Specifically, axotomy decreases the number of neurons in both the CGRP and IB4 subgroups in L5 DRGs, whereas SNL L4 neurons are unaffected [52]. These populations recover over the ensuing 20 weeks, and partial recovery occurs by the 3-week time point that we used in the present study. This downward shift in neuron population size,

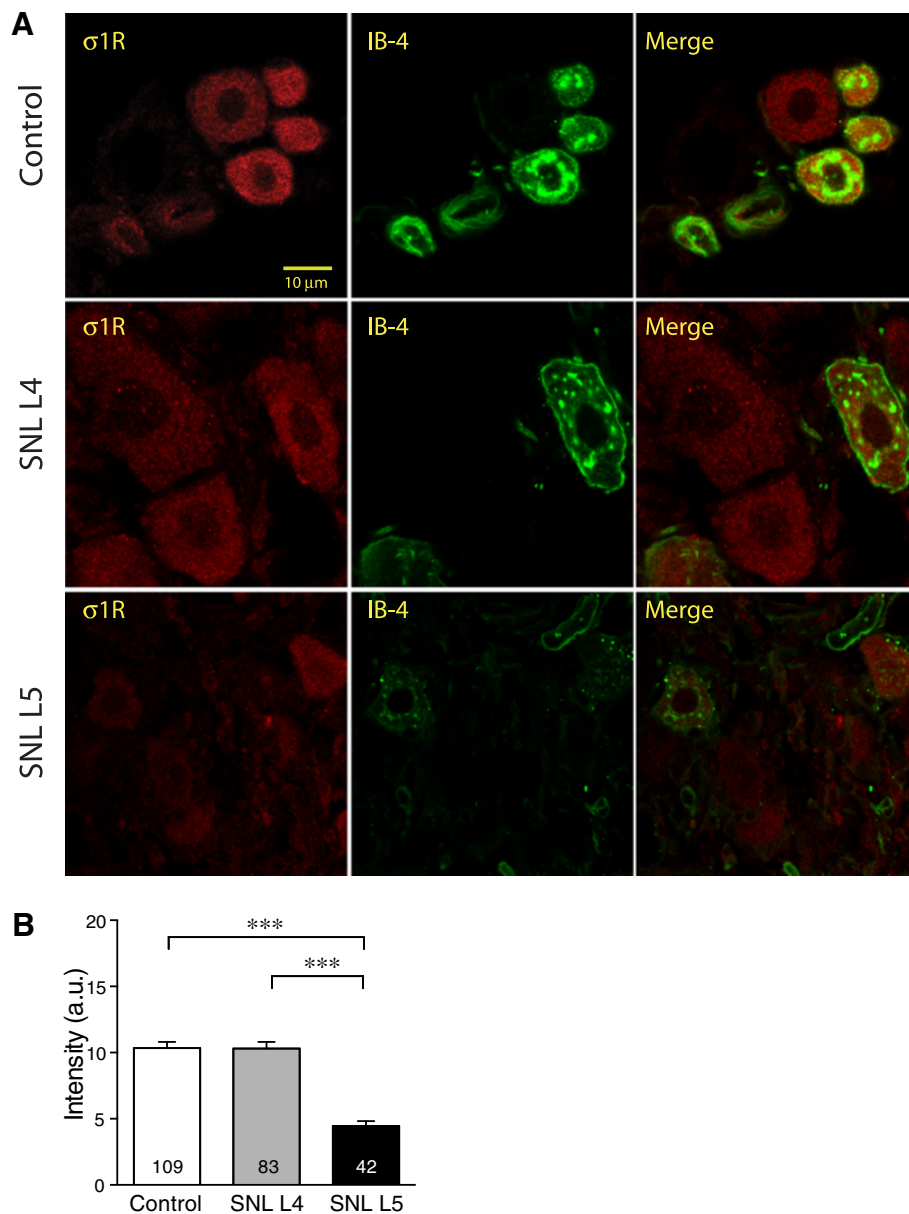


Figure 5 Effect of injury on σ 1 receptor (σ 1R) expression in sensory neurons identified by IB4 staining. **(A)** Staining of σ 1R was distributed within cytoplasmic areas of all IB4 co-expressing neurons. σ 1R immunofluorescence intensity was decreased in spinal nerve ligation (SNL) 5th lumbar (L5) neurons compared with Control. **(B)** Summary data showed decrease of intensity of σ 1R staining in IB4-positive SNL L5 neurons. Mean \pm SEM; number in bars represents the sample size; *** p < 0.001.

when combined with our observation of decreased σ 1R expression per neuron, could amplify the functional effect of depression of σ 1R in neurons of the CGRP and IB4 subgroups (typically small nociceptors). Our data do not provide any insight into the cause of depressed σ 1R levels after nerve injury. However, it is possible that accelerated axonal transport of σ 1R from the sensory neuron soma may account for receptor depletion there. Specifically, σ 1R bound to the ER membrane is activated by ER stress, such as occurs with ER Ca^{2+} depletion that

follows nerve injury [35]. Thereupon, σ 1R is relocated to targeted cell compartments [38,56,57], which may include non-somatic regions.

We found that σ 1R expression was depressed in SNL L4 DRGs, although not to the same extent as in axotomized L5 neurons and not in the IB4 population. It has been shown that L5 axotomy results in changes of a variety of signaling peptides and cellular function in the uninjured L4 DRG neurons [58,59]. In common with these phenotypic shifts, decreased σ 1R expression in

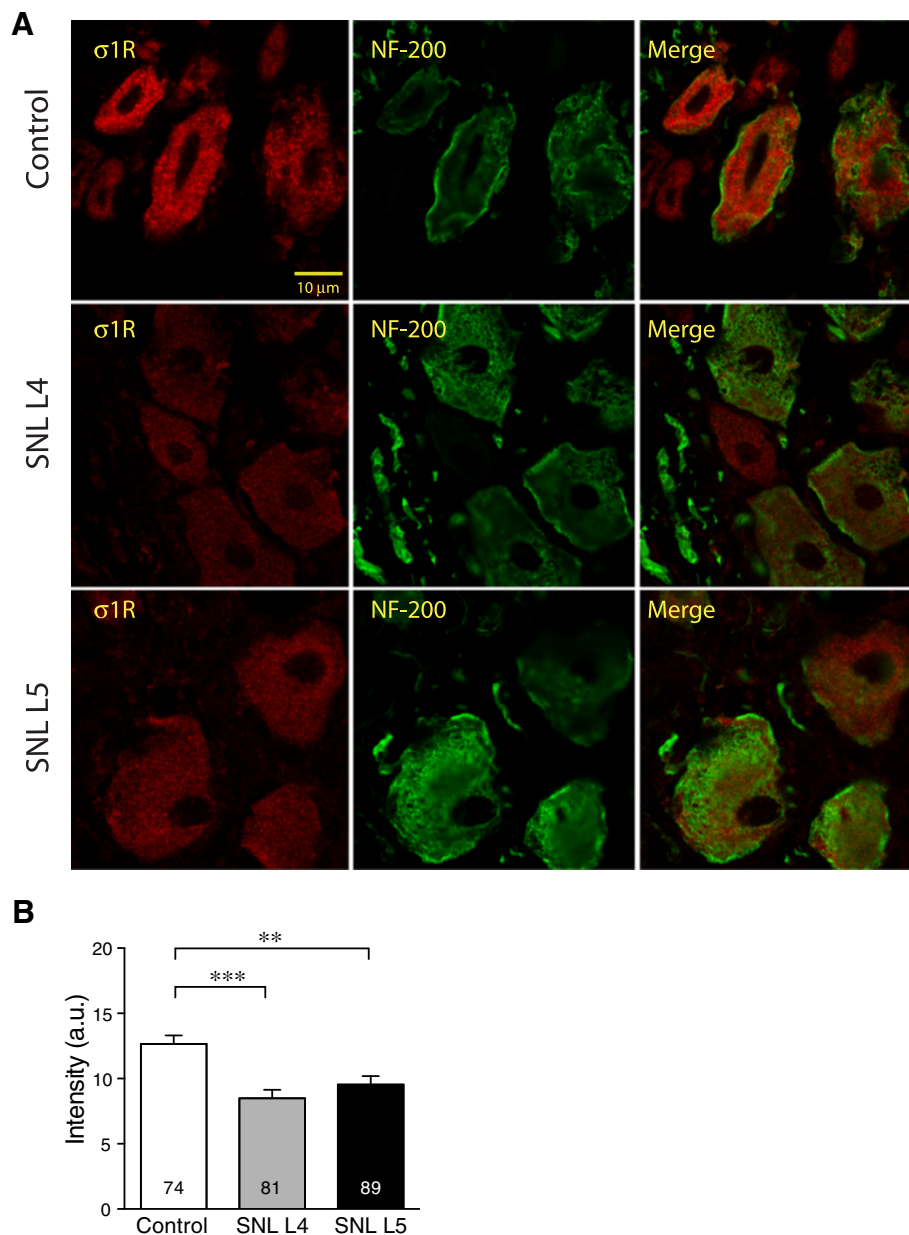


Figure 6 Effect of injury on σ 1 receptor (σ 1R) expression in sensory neurons identified by neurofilament-200 (NF-200) staining.

(A) Staining of σ 1R was distributed within cytoplasmic areas of all NF-200 co-expressing neurons. σ 1R immunofluorescence intensity was decreased in both spinal nerve ligation (SNL) 4th lumbar (L4) and 5th lumbar (L5) neurons compared with Control. **(B)** Summary data showed decrease of intensity of σ 1R staining in IB4-positive SNL L4 and L5 neurons. Mean \pm SEM; number in bars represents the sample size; ** p < 0.01, *** p < 0.001.

SNL L4 neurons may result from inflammatory processes triggered by the degeneration of axotomized distal segments of L5 neurons within the distal sciatic nerve [60-62].

We must consider what our present findings may indicate for the role of σ 1R in pain processing. It is clear that σ 1R activation potentiates nociceptive responses under both baseline conditions and in the setting of facilitated pain models [20,22,24,63,64]. In the specific

setting of nerve injury, responses to mechanical and thermal stimuli are potentiated by σ 1R agonists and attenuated by σ 1R antagonists or genetic elimination of σ 1R [65]. Implicated signaling pathways include nitric oxide, PKC, PKA, and NMDA receptor in the spinal cord [24-29]. One possible explanation of these events would be injury-induced overexpression of σ 1R. Our results, however, clearly indicate that σ 1R expression is down-regulated in primary sensory neurons in the

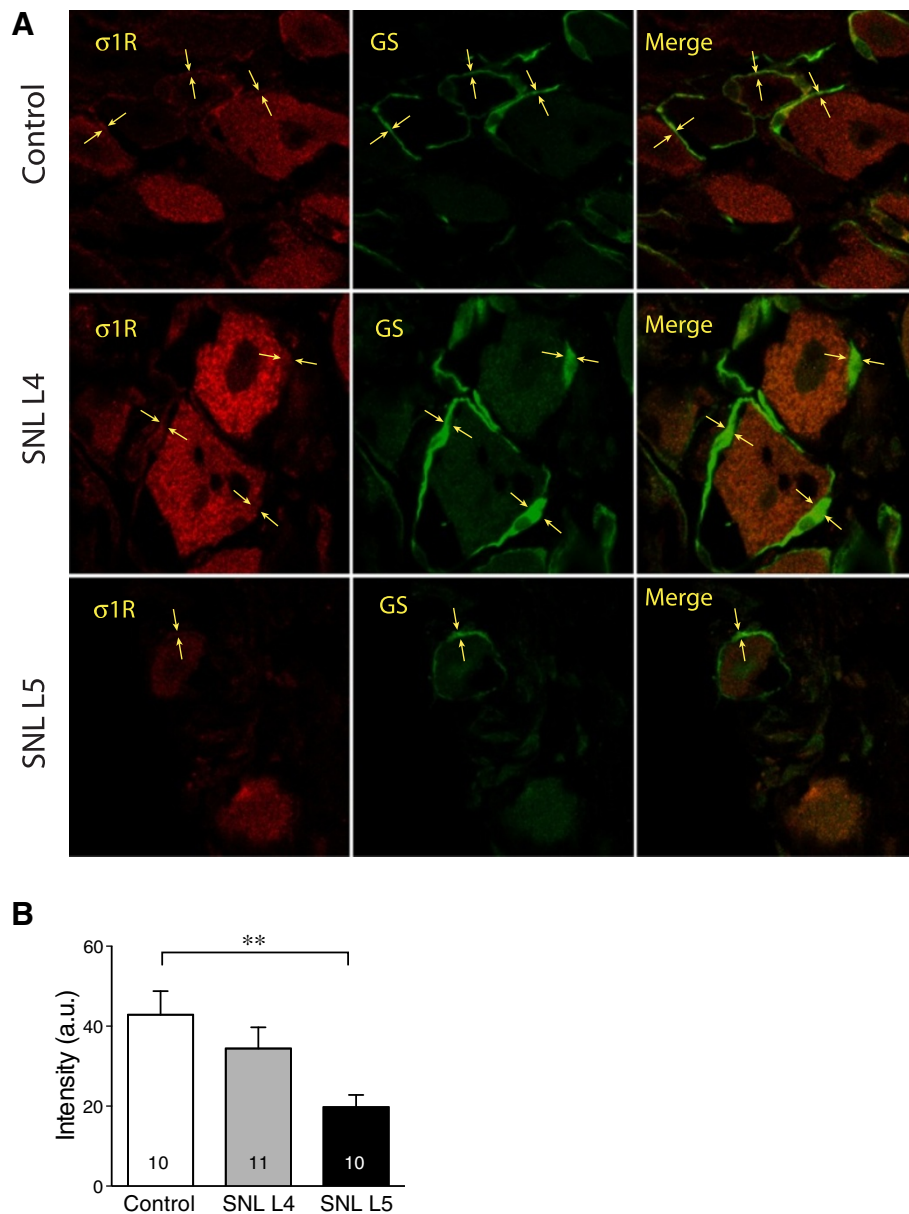


Figure 7 Effect of injury on $\sigma 1R$ expression in satellite glial cells identified by glutamine synthase (GS) staining.

(A) Intensity of $\sigma 1R$ was decreased in cytoplasmic areas of satellite glial cells in 5th lumbar (L5) DRGs after spinal nerve ligation. (B) Summary data for average intensity in satellite glial areas showed decrease of intensity of $\sigma 1R$ staining in GS-positive glial cells in SNL L5 dorsal root ganglia. Mean \pm SEM; number in bars represents the section sample size; ** $p < 0.01$.

chronic phase after axotomy, although we did not examine earlier time points. In contrast to our DRG findings, spinal cord dorsal horn homogenates show elevated $\sigma 1R$ protein expression early (3 days) after sciatic nerve injury [25], but since expression returns to control level by day 7, a shift in dorsal horn $\sigma 1R$ levels is also unlikely to contribute chronic neuropathic pain. It therefore appears that the $\sigma 1R$ activation and blockade influence pain signaling either through the relatively unaffected L4 neurons, or that the $\sigma 1R$ remaining in the axotomized

population is sufficient for these modulators to alter pain generation. Likewise, there may be a pathogenic role for $\sigma 1R$ in neuropathic pain if levels of endogenous neurosteroid agonists such as dehydroepiandrosterone [66] and pregnenolone increase sufficiently to produce elevated $\sigma 1R$ effects despite lowered $\sigma 1R$ expression.

Recent studies have shown that $\sigma 1R$ s are present in oligodendroglia and astrocytes in the CNS [4,47,67,68] and in Schwann cells of the sciatic nerve [3]. Since SGCs in the DRG play a similar role as astrocytes in CNS [46]

and come from the same embryological origin as Schwann cells [69], our finding of σ 1R in SGCs is consistent with these previous reports. After ischemic brain injury, σ 1Rs are upregulated in reactive astrocytes as well as neurons [67], which reduces glutamate accumulation and inhibits interleukin-1 β expression and microglial migration to abate ischemia-induced inflammatory reaction [70,71] (reviewed in [7]). Our data show that peripheral nerve injury differs from ischemic brain injury since σ 1R decreases in the ganglia in which axotomized neurons reside. This may deprive such neurons of the evident protective effects of σ 1R modulation of inflammatory processes and indirectly contribute to the generation of chronic pain after nerve injury.

Conclusions

Our findings indicate that σ 1Rs are present in sensory neurons, thus supporting these neurons as a possible site of action in enhancing pain after intrathecal σ 1R activation and reducing pain with σ 1R blockade [25,27,72]. After peripheral nerve injury, σ 1R expression is decreased in axotomized L5 and to a lesser extent in neighboring L4 DRG neurons in all tested neuronal subpopulations, which eliminates injury-induced change of neuronal receptor expression as a pathogenic mechanism. Overexpression of endogenous σ 1R agonists may account for sensitivity of neuropathic pain to σ 1R blockade, and loss of glial σ 1R may be a contributing factor.

Methods

Animals

All methods and use of animals were approved by the Medical College of Wisconsin Institutional Animal Care and Use Committee. Male Sprague-Dawley rats (Taconic Farms Inc., Hudson, NY) were housed individually in a room maintained at $22 \pm 0.5^\circ\text{C}$ and constant humidity ($60 \pm 15\%$) with an alternating 12 hr light-dark cycle. Food and water were available *ad libitum* throughout the experiments.

Injury model

Rats weighing 150 to 180 g were subjected to SNL modified from the original technique [73]. Specifically, rats were anesthetized with 2% isoflurane in oxygen and the right paravertebral region was exposed. The L6 transverse process was removed, after which the L5 and L6 spinal nerves were ligated with 6-0 silk suture and transected distal to the ligature. To minimize non-neural injury, no muscle was removed, muscles and intertransverse fascia were incised only at the site of the two ligations, and articular processes were not removed. The muscular fascia was closed with 4-0 resorbable polyglactin sutures and the skin closed with staples. Control animals received skin incision and closure only. After surgery, rats were returned to their

cages and kept under normal housing conditions with access to pellet food and water *ad lib*.

Sensory testing

We measured the incidence of a pattern of hyperalgesic behavior that we have previously documented to be associated with conditioned place avoidance [74,75]. Briefly, on day 10, 12, and 17 after surgery, right plantar skin was touched (10 stimuli/session) with a 22G spinal needle with adequate pressure to indent but not penetrate the skin. Control animals respond with only a brief reflexive withdrawal, whereas rats subjected to SNL may display a complex hyperalgesia response that includes licking, chewing, grooming and sustained elevation of the paw, which peaks in frequency by day 17. Because of day-to-day variability in individual rat responses, the average frequency of hyperalgesia responses over the 3 testing days was tabulated for each rat. After SNL, only rats that displayed a hyperalgesia-type response after at least 20% of stimuli were used further in this study.

Quantitative real-time PCR analysis

Total RNA was isolated from the homogenized L4 and L5 DRG of control animals ($n = 6$), and separately from the L4 and L5 DRGs of SNL rats ($n = 6$) harvested 21 days after surgery, following the manufacturer's (Invitrogen, Carlsbad, CA) instructions using Trizol reagent (from aqueous phase). After DNase treatment, cDNA was synthesized from amounts of RNA that were standardized for each experiment ranging from 152 to 232 ng for different experiments, using random hexamer primers (Superscript III first strand synthesis kit; Invitrogen, Grand Islands, NY). Quantitative real time PCR (qPCR) analysis was carried out using IQ Syber Green supermix (Biorad Laboratories, Hercules, CA) and specific primers to quantify the cDNA levels of the σ 1R genes (forward primer (FP), 5'-CCTGCTGGCATTTCGGGGCTC-3'; reverse primer (RP), 5'-TCACAGCTGCAGGCCGAACG G-3'; NCBI reference sequence, NM_030996.1). A preliminary gel showed that PCR products had the expected molecular weight and a melting curve was run in each sample to confirm single product in every run (data not shown). Normalization was carried out using the geometric mean of two reference genes, glyceraldehyde 3-phosphate dehydrogenase (GAPDH; FP, 5'-AGACAGCC GCATCTTCTTGT-3'; RP, 5'-TGATGGCAACAATGT CCACT-3') and mitogen-activated protein kinase 6 (MAPK6; FP, 5'-TAAAGCCATTGACATGTGGG-3'; RP, 5'-TCGTGCACAACAGGGATAGA-3'), which were chosen for their stability in the context of SNL injury [76]. For each sample, two inter-run determinations were averaged and the fold differences in expression (SNL L4, and SNL L5 DRG) were compared to that of the Control samples using the comparative C_T method. Statistical

analysis of $\Delta\Delta C_T$ values used for $\sigma 1R$, normalized with MAPK6/GAPDH, was performed using Kruskal-Wallis test ($p = 0.073$) with *Post hoc* by Dunn's test. Graphs were plotted using $2^{-\Delta\Delta C_T}$ values for representation of $\sigma 1R$ gene expressions in different groups.

Western blot analysis

Total protein was isolated from individual L4 and L5 DRGs of Control animals, and separately from L4 and L5 DRGs of SNL animals. Harvested ganglia were homogenized in 150 μ l of RIPA lysis buffer containing protease inhibitors (Roche Diagnostics, Indianapolis, IN) and phosphatase inhibitors (Thermo Scientific, Rockford, IL), and incubated on ice for 30 minutes. Lysates were spun down at 14000 rpm for 10 min at 4°C. The supernatant was used for protein estimation using Pierce bicinchoninic acid protein assay kit (Thermo Scientific, Rockford, IL). Equal amounts of protein (30 μ g) were separated on 4-15% sodium dodecyl sulphate polyacrylamide gel electrophoresis gel (SDS-PAGE; Bio-Rad, Hercules, CA) and transferred onto a polyvinylidene fluoride membrane. After blocking with 5% milk in TBS-T (Tris-buffered saline plus 0.1% Tween 20), blots were sequentially probed overnight at 4°C with one of two anti- $\sigma 1R$ rabbit polyclonal antibodies, either a commercial product (1:500, Invitrogen, Camarillo, CA, catalog number 42-3300; Lot No. 797152A, polyclonal affinity-purified antibody raised against a synthetic peptide derived from the C-terminus region (amino acid 139-155) of the rat $\sigma 1R$), or another that was a kind gift from Drs. Su and Hayashi of NIDA (1:1000, Lot No. 6405, polyclonal affinity-purified antibody raised against amino acid 52-69 from the C-terminus of rat $\sigma 1R$) [6,77]. Loading control was used anti- β -tubulin I mouse monoclonal antibody (1:5000, Sigma-Aldrich, St. Louis, MO, catalog number T7816, Lot No. 088 K4795, monoclonal antibody raised against a synthetic peptide corresponding to the C-terminus sequence of β -tubulin isotype I, conjugated to BSA) [45]. Three 15 min washes in TBS-T followed before probing with secondary antibody of horseradish peroxidase-conjugated goat anti-rabbit antibody (1:2000, Pierce, Rockford, IL). Enhanced chemiluminescence (ECL plus; Amersham, Piscataway, NJ) was used to detect protein bands. Western blot restore stripping buffer (Thermo Scientific, Rockford, IL) was used to strip antibodies from the membrane. The bands obtained were quantified using ImageJ (U.S. National Institutes of Health, Bethesda, MD) program, and β -tubulin I was used to normalize the protein loading. The normalized values (to β -tubulin I) of Control, SNL L4 and SNL L5 isoform were used for the statistical analysis. Although the antibody from NIDA showed a strong expression band at the expected molecular weight (25 kDa), the blot also showed non-specific bands as well (data not shown). The more selective Invitrogen antibody was used for immunohistochemistry.

Immunohistochemistry

Twenty-one days post-surgery, the Control, SNL L4 and SNL L5 DRGs were harvested and cryoprotected in 4% paraformaldehyde with 15% sucrose in 0.1 M PBS for 1 h, followed by incubation in 30% sucrose 0.1 M PBS overnight [49]. Tissues embedded in Tissue-Tek optimal cutting temperature compound (Ted Pella, Inc., Redding, CA) were sectioned (10 μ m) with a Leica cryostat (Jung CM 1800; Vienna, Austria), plated onto subbed slides (Superfrost Plus Gold; Fisher Scientific, Pittsburgh, PA), and post-fixed in 4% paraformaldehyde with 4% sucrose for 10 min. After blocking with 10% normal goat serum for 1 h at room temperature, the tissue was incubated overnight with anti- $\sigma 1R$ rabbit polyclonal antibody (1:100, Invitrogen, same antibody used Western) [77]. After four washes with PBST, sections were incubated with Alexa Fluor 568 Goat anti-rabbit antibody (1:500; Invitrogen, Camarillo, CA) for 2 h. To determine co-localization of $\sigma 1R$ with the neuron-specific nuclear protein (NeuN), neurofilament 200 (NF-200), CGRP, and glutamine synthetase, the sections were washed four times with PBST followed by incubated with anti-NeuN mouse monoclonal antibody (1:100, Millipore, Billerica, MA, catalog number MAB377, Lot No. LV1616015, monoclonal antibody raised against purified cell nuclei from mouse brain) [45], anti-NF-200 mouse monoclonal antibody (1:1000, Abcam, Cambridge, UK, catalog number ab28029, monoclonal antibody raised against a non phosphorylated epitope from 200kD Neurofilament Heavy of most mammalian species) [49], anti-CGRP mouse polyclonal antibody (1:50, Santa Cruz Biotechnology, Santa Cruz, CA, catalog number SC-28920, Lot No. H1407, epitope corresponding to amino acid 81-128 mapping at the C-terminus of CGRP of human origin) [49], or anti-glutamine synthetase rabbit polyclonal antibody (1:500, Santa Cruz Biotechnology, Santa Cruz, CA, catalog number SC-6640, Lot No. H1407, polyclonal affinity purified antibody raised against a peptide mapping at the C-terminus of glutamine synthetase of human origin) [45] for 2 h. Following four washes with PBST, sections were incubated with Alexa Fluor 488 goat anti-mouse antibody or Alexa Fluor 488 goat anti-rabbit antibody (1:1000; Molecular Probes, Camarillo, CA) for 1 h. To determine co-localization of $\sigma 1R$ with IB4, sections were incubated with Alexa Fluor 488 conjugate IB₄ (1:50, Invitrogen, Camarillo, CA, catalog number I21411, Lot No. 743633,) [78] after four washes with PBST. Sections were washed four times with PBST and covered with Prolong Gold Antifade mounting medium (Invitrogen, Camarillo, CA). Sections were examined by confocal microscopy. The expression level of $\sigma 1R$ protein was represented by the image intensity that was captured using standardized camera parameters (Metamorph, Downingtown, PA), and cell area was determined by outlining the neuronal profile by excluding its nucleus. A neuron was considered positive when intensity in

cells of interest was two-fold greater than in background in sections incubated without targeting primary antibody. At least 3 sections from each DRG were randomly chosen for fluorescence intensity measurement, except in a case of SNL L5 DRG with σ 1R/CGRP double staining, for which 2 sections were evaluated. The fluorescence intensity of all cells in the section was quantified. The individual who measured the fluorescence intensity was not completely blinded to the treatment due to obvious markers of axotomy such as eccentric location of the nucleus. Average fluorescence intensity of neurons was measured in traced cytoplasmic areas of interest after subtracting background fluorescence [49,79]. Satellite glial cell σ 1R average intensity was derived by creating a mask from a thresholded image of glutamine synthetase immunofluorescence to isolate areas of interest in the image of σ 1R immunofluorescence using Photoshop (Adobe Systems Inc., New York City, NY).

Statistical analysis

Prism (version 6.1, GraphPad Software, Inc., San Diego, CA) was used to perform paired or unpaired Student's *t*-test or one-way ANOVA. Data were derived from at least 3 DRGs for every group. Non-parametric Kruskal-Wallis test with *Post hoc* by Dunn's test was used for analyzing the influence of injury on σ 1R in qPCR and Western immunoblot experiments. In IHC experiments, main effects identified by ANOVA were further analyzed by Tukey's test (for comparisons between all groups) to compare relevant means. Data are reported as median with interquartile range for qPCR and Western immunoblot experiments, and as mean \pm SEM for IHC experiments. A *P* value less than 0.05 was considered significant.

Abbreviations

SNL: Spinal nerve ligation; CNS: Central nervous system; DRG: Dorsal root ganglion; SGC: Satellite glial cells; σ 1R: Sigma-1 receptor; IB4: Isolectin-4; CGRP: Calcitonin gene-related protein; NeuN: Neuron-specific nuclear protein; NF-200: Neurofilament 200; IHC: Immunohistochemical.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

BML designed quantitative real time PCR experiment, acquired, analyzed and interpreted data and also contributed to western blotting and IHC studies; DM contributed to confocal microscope study in analyzing and interpreted IHC data; VZ contributed to IHC studies; QBT contributed to western blotting and IHC study designs; QH participated in interpretation of the data, and revising the manuscript; HEW contributed to the conception and design of the studies, interpretation of the data, writing and revising the manuscript. All authors read and approved the final manuscript.

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References

1. Hayashi T, Su TP: Sigma-1 receptor chaperones at the ER-mitochondrion interface regulate Ca(2+) signaling and cell survival. *Cell* 2007, **131**:596-610.
2. Matsumoto RR: σ Receptors: historical perspective and background. In *Sigma Receptors*. 1st edition. Edited by Matsumoto RR, Bowen WD, Su T-P. New York, NY: Springer; 2007:1-23.
3. Palacios G, Muro A, Verdu E, Pumarola M, Vela JM: Immunohistochemical localization of the sigma1 receptor in Schwann cells of rat sciatic nerve. *Brain Res* 2004, **1007**:65-70.
4. Palacios G, Muro A, Vela JM, Molina-Holgado E, Guitart X, Ovalle S, Zamanillo D: Immunohistochemical localization of the sigma1-receptor in oligodendrocytes in the rat central nervous system. *Brain Res* 2003, **961**:92-99.
5. Alonso G, Phan V, Guillemain I, Saunier M, Legrand A, Anoaï M, Maurice T: Immunocytochemical localization of the sigma(1) receptor in the adult rat central nervous system. *Neuroscience* 2000, **97**:155-170.
6. Tchédre KT, Huang RQ, Dibas A, Krishnamoorthy RR, Dillon GH, Yorio T: Sigma-1 receptor regulation of voltage-gated calcium channels involves a direct interaction between sigma-1 receptors and the voltage-gated L-type calcium channel. *Invest Ophthalmol Vis Sci* 2008, **49**:4993-5002.
7. Hayashi T, Tsai SY, Mori T, Fujimoto M, Su TP: Targeting ligand-operated chaperone sigma-1 receptors in the treatment of neuropsychiatric disorders. *Expert Opin Ther Targets* 2011, **15**:557-577.
8. Maurice T, Su TP: The pharmacology of sigma-1 receptors. *Pharmacol Ther* 2009, **124**:195-206.
9. Tagashira H, Zhang C, Lu YM, Hasegawa H, Kanai H, Han F, Fukunaga K: Stimulation of sigma(1)-receptor restores abnormal mitochondrial Ca(2+) mobilization and ATP production following cardiac hypertrophy. *Biochim Biophys Acta* 1830, **2013**:3082-3094.
10. Megalizzi V, Le Mercier M, Decaestecker C: Sigma receptors and their ligands in cancer biology: overview and new perspectives for cancer therapy. *Med Res Rev* 2012, **32**:410-427.
11. Hayashi T, Justinova Z, Hayashi E, Cormaci G, Mori T, Tsai SY, Barnes C, Goldberg SR, Su TP: Regulation of sigma-1 receptors and endoplasmic reticulum chaperones in the brain of methamphetamine self-administering rats. *J Pharmacol Exp Ther* 2010, **332**:1054-1063.
12. Hiranita T, Soto PL, Tanda G, Katz JL: Reinforcing effects of sigma-receptor agonists in rats trained to self-administer cocaine. *J Pharmacol Exp Ther* 2010, **332**:515-524.
13. Villard V, Espallergues J, Keller E, Alkam T, Nitta A, Yamada K, Nabeshima T, Vamvakides A, Maurice T: Antiamnesic and neuroprotective effects of the aminotetrahydrofuran derivative ANAVEX1-41 against amyloid beta(25-35)-induced toxicity in mice. *Neuropsychopharmacology* 2009, **34**:1552-1566.
14. Yao H, Yang Y, Kim KJ, Bethel-Brown C, Gong N, Funa K, Gendelman HE, Su TP, Wang JQ, Buch S: Molecular mechanisms involving sigma receptor-mediated induction of MCP-1: implication for increased monocyte transmigration. *Blood* 2010, **115**:4951-4962.
15. Ajmo CT Jr, Vernon DO, Collier L, Pennypacker KR, Cuevas J: Sigma receptor activation reduces infarct size at 24 hours after permanent middle cerebral artery occlusion in rats. *Curr Neurovasc Res* 2006, **3**:89-98.
16. Cuevas J, Behensky A, Deng W, Katnik C: Afobazole modulates neuronal response to ischemia and acidosis via activation of sigma-1 receptors. *J Pharmacol Exp Ther* 2011, **339**:152-160.
17. Tseng LF, Hogan QH, Wu HE: (+)-Morphine attenuates the (-)-morphine-produced tail-flick inhibition via the sigma-1 receptor in the mouse spinal cord. *Life Sci* 2011, **89**:875-877.
18. Terashvili M, Wu HE, Moore RM, Harder DR, Tseng LF: (+)-Morphine and (-)-morphine stereoselectively attenuate the (-)-morphine-produced tail-flick inhibition via the naloxone-sensitive sigma receptor in the ventral periaqueductal gray of the rat. *Eur J Pharmacol* 2007, **571**:1-7.
19. Wu HE, Hong JS, Tseng LF: Stereoselective action of (+)-morphine over (-)-morphine in attenuating the (-)-morphine-produced antinociception via the naloxone-sensitive sigma receptor in the mouse. *Eur J Pharmacol* 2007, **571**:145-151.

20. Chien CC, Pasternak GW: **Selective antagonism of opioid analgesia by a sigma system.** *J Pharmacol Exp Ther* 1994, **271**:1583–1590.
21. Chien CC, Pasternak GW: **Sigma antagonists potentiate opioid analgesia in rats.** *Neurosci Lett* 1995, **190**:137–139.
22. Mei J, Pasternak GW: **Sigma1 receptor modulation of opioid analgesia in the mouse.** *J Pharmacol Exp Ther* 2002, **300**:1070–1074.
23. Mei J, Pasternak GW: **Modulation of brainstem opiate analgesia in the rat by sigma 1 receptors: a microinjection study.** *J Pharmacol Exp Ther* 2007, **322**:1278–1285.
24. Kim HW, Kwon YB, Roh DH, Yoon SY, Han HJ, Kim KW, Beitz AJ, Lee JH: **Intrathecal treatment with sigma1 receptor antagonists reduces formalin-induced phosphorylation of NMDA receptor subunit 1 and the second phase of formalin test in mice.** *Br J Pharmacol* 2006, **148**:490–498.
25. Roh DH, Kim HW, Yoon SY, Seo HS, Kwon YB, Kim KW, Han HJ, Beitz AJ, Na HS, Lee JH: **Intrathecal injection of the sigma(1) receptor antagonist BD1047 blocks both mechanical allodynia and increases in spinal NR1 expression during the induction phase of rodent neuropathic pain.** *Anesthesiology* 2008, **109**:879–889.
26. Romero L, Zamanillo D, Nadal X, Sanchez-Arroyos R, Rivera-Arconada I, Dordal A, Montero A, Muro A, Bura A, Segales C, et al: **Pharmacological properties of S1RA, a new sigma-1 receptor antagonist that inhibits neuropathic pain and activity-induced spinal sensitization.** *Br J Pharmacol* 2012, **166**:2289–2306.
27. Roh DH, Kim HW, Yoon SY, Seo HS, Kwon YB, Kim KW, Han HJ, Beitz AJ, Lee JH: **Intrathecal administration of sigma-1 receptor agonists facilitates nociception: Involvement of a protein kinase C-dependent pathway.** *J Neurosci Res* 2008, **86**:3644–3654.
28. Roh DH, Choi SR, Yoon SY, Kang SY, Moon JY, Kwon SG, Han HJ, Beitz AJ, Lee JH: **Spinal nNOS activation mediates sigma-1 receptor-induced mechanical and thermal hypersensitivity in mice: involvement of PKC-dependent NR1 phosphorylation.** *Br J Pharmacol* 2011, **163**:1707–1720.
29. Yoon SY, Roh DH, Seo HS, Kang SY, Han HJ, Beitz AJ, Lee JH: **Intrathecal injection of the neurosteroid, DHEAS, produces mechanical allodynia in mice: involvement of spinal sigma-1 and GABA receptors.** *Br J Pharmacol* 2009, **157**:666–673.
30. Zimmermann M: **Pathobiology of neuropathic pain.** *Eur J Pharmacol* 2001, **429**:23–37.
31. von Hehn CA, Baron R, Woolf CJ: **Deconstructing the neuropathic pain phenotype to reveal neural mechanisms.** *Neuron* 2012, **73**:638–652.
32. Cai YQ, Chen SR, Pan HL: **Upregulation of nuclear factor of activated T-cells by nerve injury contributes to development of neuropathic pain.** *J Pharmacol Exp Ther* 2013, **345**:161–168.
33. Inquimbert P, Bartels K, Babaniyi OB, Barrett LB, Tegeder I, Scholz J: **Peripheral nerve injury produces a sustained shift in the balance between glutamate release and uptake in the dorsal horn of the spinal cord.** *Pain* 2012, **153**:2422–2431.
34. Takeuchi Y, Yamasaki M, Nagumo Y, Imoto K, Watanabe M, Miyata M: **Rewiring of afferent fibers in the somatosensory thalamus of mice caused by peripheral sensory nerve transection.** *J Neurosci* 2012, **32**:6917–6930.
35. Rigaud M, Gemes G, Weyker PD, Cruikshank JM, Kawano T, Wu HE, Hogan QH: **Axotomy depletes intracellular calcium stores in primary sensory neurons.** *Anesthesiology* 2009, **111**:381–392.
36. Hogan QH, McCallum JB, Sarantopoulos C, Aason M, Mynlieff M, Kwok WM, Bosnjak ZJ: **Painful neuropathy decreases membrane calcium current in mammalian primary afferent neurons.** *Pain* 2000, **86**:43–53.
37. McCallum JB, Kwok WM, Sapunar D, Fuchs A, Hogan QH: **Painful peripheral nerve injury decreases calcium current in axotomized sensory neurons.** *Anesthesiology* 2006, **105**:160–168.
38. Hayashi T, Maurice T, Su TP: **Ca(2+) signaling via sigma(1)-receptors: novel regulatory mechanism affecting intracellular Ca(2+) concentration.** *J Pharmacol Exp Ther* 2000, **293**:788–798.
39. Zhang H, Cuevas J: **Sigma receptors inhibit high-voltage-activated calcium channels in rat sympathetic and parasympathetic neurons.** *J Neurophysiol* 2002, **87**:2867–2879.
40. Hayashi T, Su TP: **Sigma-1 receptor ligands: potential in the treatment of neuropsychiatric disorders.** *CNS Drugs* 2004, **18**:269–284.
41. Maurice T, Gregoire C, Espallergues J: **Neuro(steroid)s actions at the neuromodulatory sigma1 (sigma1) receptor: biochemical and physiological evidences, consequences in neuroprotection.** *Pharmacol Biochem Behav* 2006, **84**:581–597.
42. Schaeffer V, Meyer L, Patte-Mensah C, Eckert A, Mensah-Nyagan AG: **Sciatic nerve injury induces apoptosis of dorsal root ganglion satellite glial cells and selectively modifies neurosteroidogenesis in sensory neurons.** *Glia* 2010, **58**:169–180.
43. Hayashi T, Su TP: **Regulating ankyrin dynamics: Roles of sigma-1 receptors.** *Proc Natl Acad Sci USA* 2001, **98**:491–496.
44. Weick M, Cherkas PS, Hartig W, Pannicke T, Uckermann O, Bringmann A, Tal M, Reichenbach A, Hanani M: **P2 receptors in satellite glial cells in trigeminal ganglia of mice.** *Neuroscience* 2003, **120**:969–977.
45. Gemes G, Bangaru ML, Wu HE, Tang Q, Weihrach D, Koopmeiners AS, Cruikshank JM, Kwok WM, Hogan QH: **Store-operated Ca2+ entry in sensory neurons: functional role and the effect of painful nerve injury.** *J Neurosci* 2011, **31**:3536–3549.
46. Hanani M: **Satellite glial cells in sensory ganglia: from form to function.** *Brain Res Brain Res Rev* 2005, **48**:457–476.
47. Hayashi T, Su TP: **Sigma-1 receptors at galactosylceramide-enriched lipid microdomains regulate oligodendrocyte differentiation.** *Proc Natl Acad Sci USA* 2004, **101**:14949–14954.
48. Xing J, Lu J, Li J: **Acid-sensing ion channel subtype 3 function and immunolabelling increases in skeletal muscle sensory neurons following femoral artery occlusion.** *J Physiol* 2012, **590**:1261–1272.
49. Zoga V, Kawano T, Liang MY, Bienengraeber M, Weihrach D, McCallum B, Gemes G, Hogan Q, Sarantopoulos C: **KATP channel subunits in rat dorsal root ganglia: alterations by painful axotomy.** *Mol Pain* 2010, **6**:6.
50. Monnet FP, Debonnel G, Junien JL, De Montigny C: **N-methyl-D-aspartate-induced neuronal activation is selectively modulated by sigma receptors.** *Eur J Pharmacol* 1990, **179**:441–445.
51. Son JS, Kwon YB: **Sigma-1 Receptor Antagonist BD1047 Reduces Allodynia and Spinal ERK Phosphorylation Following Chronic Compression of Dorsal Root Ganglion in Rats.** *Korean J Physiol Pharmacol* 2010, **14**:359–364.
52. Hammond DL, Ackerman L, Holdsworth R, Elzey B: **Effects of spinal nerve ligation on immunohistochemically identified neurons in the L4 and L5 dorsal root ganglia of the rat.** *J Comp Neurol* 2004, **475**:575–589.
53. Aanonsen LM, Seybold VS: **Phencyclidine and sigma receptors in rat spinal cord: binding characterization and quantitative autoradiography.** *Synapse* 1989, **4**:1–10.
54. McCann DJ, Su TP: **Haloperidol competitively inhibits the binding of (+)-[3H]SKF-10,047 to sigma sites.** *Eur J Pharmacol* 1990, **180**:361–364.
55. Gundlach AL, Largent BL, Snyder SH: **Autoradiographic localization of sigma receptor binding sites in guinea pig and rat central nervous system with (+)3H-3-(3-hydroxyphenyl)-N-(1-propyl)piperidine.** *J Neurosci* 1986, **6**:1757–1770.
56. Su TP, Hayashi T, Maurice T, Buch S, Ruoho AE: **The sigma-1 receptor chaperone as an inter-organelle signaling modulator.** *Trends Pharmacol Sci* 2010, **31**:557–566.
57. Kourrich S, Su TP, Fujimoto M, Bonci A: **The sigma-1 receptor: roles in neuronal plasticity and disease.** *Trends Neurosci* 2012, **35**:762–771.
58. Fukuoka T, Tokunaga A, Kondo E, Miki K, Tachibana T, Noguchi K: **Change in mRNAs for neuropeptides and the GABA(A) receptor in dorsal root ganglion neurons in a rat experimental neuropathic pain model.** *Pain* 1998, **78**:13–26.
59. Fukuoka T, Tokunaga A, Kondo E, Noguchi K: **The role of neighboring intact dorsal root ganglion neurons in a rat neuropathic pain model.** In *Proceedings of the 9th world congress on pain*. 1st edition. Edited by Dever M, Rowbotham MC, Weisenfeld-Hallin Z. Seattle, WA: IASP Press; 2000:137–146.
60. Murinson BB, Griffin JW: **C-fiber structure varies with location in peripheral nerve.** *J Neuropathol Exp Neurol* 2004, **63**:246–254.
61. Wu G, Ringkamp M, Hartke TV, Murinson BB, Campbell JN, Griffin JW, Meyer RA: **Early onset of spontaneous activity in uninjured C-fiber nociceptors after injury to neighboring nerve fibers.** *J Neurosci* 2001, **21**:RC140.
62. Ossipov MH, Porreca F: **Neuropathic pain: basic mechanisms (animal).** In *Science of Pain*. firstth edition Edited by Basbaum AI, Bushnell MC. Oxford, UK: Elsevier; 2009:833–855.
63. Cendan CM, Pujalte JM, Portillo-Salido E, Montoliu L, Baeyens JM: **Formalin-induced pain is reduced in sigma(1) receptor knockout mice.** *Eur J Pharmacol* 2005, **511**:73–74.
64. Entrena JM, Cobos EJ, Nieto FR, Cendan CM, Gris G, Del Pozo E, Zamanillo D, Baeyens JM: **Sigma-1 receptors are essential for capsaicin-induced mechanical hypersensitivity: studies with selective sigma-1 ligands and sigma-1 knockout mice.** *Pain* 2009, **143**:252–261.

65. de la Puente B, Nadal X, Portillo-Salido E, Sanchez-Arroyos R, Ovalle S, Palacios G, Muro A, Romero L, Entrena JM, Baeyens JM, *et al*: **Sigma-1 receptors regulate activity-induced spinal sensitization and neuropathic pain after peripheral nerve injury.** *Pain* 2009, **145**:294–303.
66. Kibaly C, Meyer L, Patte-Mensah C, Mensah-Nyagan AG: **Biochemical and functional evidence for the control of pain mechanisms by dehydroepiandrosterone endogenously synthesized in the spinal cord.** *FASEB J* 2008, **22**:93–104.
67. Penas C, Pascual-Font A, Mancuso R, Fores J, Casas C, Navarro X: **Sigma receptor agonist 2-(4-morpholinethyl)1 phenylcyclohexanecarboxylate (Pre084) increases GDNF and BiP expression and promotes neuroprotection after root avulsion injury.** *J Neurotrauma* 2011, **28**:831–840.
68. Ruscher K, Shamloo M, Rickhag M, Ladunga I, Soriano L, Gisselsson L, Toresson H, Ruslim-Litrus L, Oksenberg D, Urfer R, *et al*: **The sigma-1 receptor enhances brain plasticity and functional recovery after experimental stroke.** *Brain* 2011, **134**:732–746.
69. Hall AK, Landis SC: **Division and migration of satellite glia in the embryonic rat superior cervical ganglion.** *J Neurocytol* 1992, **21**:635–647.
70. Shen YC, Wang YH, Chou YC, Liou KT, Yen JC, Wang WY, Liao JF: **Dimemorfan protects rats against ischemic stroke through activation of sigma-1 receptor-mediated mechanisms by decreasing glutamate accumulation.** *J Neurochem* 2008, **104**:558–572.
71. Behensky AA, Cortes-Salva M, Seminerio MJ, Matsumoto RR, Antilla JC, Cuevas J: **In vitro evaluation of guanidine analogs as sigma receptor ligands for potential anti-stroke therapeutics.** *J Pharmacol Exp Ther* 2013, **344**:155–166.
72. Moon JY, Roh DH, Yoon SY, Kang SY, Choi SR, Kwon SG, Choi HS, Han HJ, Beitz AJ, Lee JH: **Sigma-1 receptor-mediated increase in spinal p38 MAPK phosphorylation leads to the induction of mechanical allodynia in mice and neuropathic rats.** *Exp Neurol* 2013, **247**:383–391.
73. Kim SH, Chung JM: **An experimental model for peripheral neuropathy produced by segmental spinal nerve ligation in the rat.** *Pain* 1992, **50**:355–363.
74. Hogan Q, Sapunar D, Modric-Jednacak K, McCallum JB: **Detection of neuropathic pain in a rat model of peripheral nerve injury.** *Anesthesiology* 2004, **101**:476–487.
75. Wu HE, Gemes G, Zoga V, Kawano T, Hogan QH: **Learned avoidance from noxious mechanical stimulation but not threshold semmes weinstein filament stimulation after nerve injury in rats.** *J Pain* 2010, **11**:280–286.
76. Bangaru ML, Park F, Hudmon A, McCallum JB, Hogan QH: **Quantification of gene expression after painful nerve injury: validation of optimal reference genes.** *J Mol Neurosci* 2012, **46**:497–504.
77. Hayashi T, Lewis A, Hayashi E, Betenbaugh MJ, Su TP: **Antigen retrieval to improve the immunocytochemistry detection of sigma-1 receptors and ER chaperones.** *Histochem Cell Biol* 2011, **135**:627–637.
78. Stemkowski PL, Smith PA: **Long-term IL-1beta exposure causes subpopulation-dependent alterations in rat dorsal root ganglion neuron excitability.** *J Neurophysiol* 2012, **107**:1586–1597.
79. Obata K, Katsura H, Mizushima T, Sakurai J, Kobayashi K, Yamanaka H, Dai Y, Fukuoka T, Noguchi K: **Roles of extracellular signal-regulated protein kinases 5 in spinal microglia and primary sensory neurons for neuropathic pain.** *J Neurochem* 2007, **102**:1569–1584.

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