

RESEARCH

Open Access

# *In vitro* and *in vivo* anti-malarial activity of *Boerhavia elegans* and *Solanum surattense*

Ali Ramazani<sup>1</sup>, Sedigheh Zakeri\*<sup>1</sup>, Soroush Sardari<sup>2</sup>, Nastaran Khodakarim<sup>1</sup> and Navid Dinparas Djadid<sup>1</sup>

## Abstract

**Background:** There is an urgent need to identify new anti-malarial drug targets for both prophylaxis and chemotherapy, due to the increasing problem of drug resistance to malaria parasites. In the present study, the aim was to discover novel, effective plant-based extracts for the activity against malaria.

**Methods:** Ten plants found in Iran were selected by ethnobotanical survey of medicinal plants. The crude ethanolic extracts were tested for *in vitro* anti-plasmodial activity against two strains of *Plasmodium falciparum*: K1 (chloroquine-resistant strain) and CY27 (chloroquine-sensitive strain), using the parasite lactate dehydrogenase (pLDH) assay. The anti-plasmodial activity of the extracts was also assessed in the 4-day suppressive anti-malarial assay in mice inoculated with *Plasmodium berghei* (ANKA strain). Crude ethanolic extracts showed good anti-plasmodial activity were further fractionated by partitioning in water and dichloromethane.

**Results:** Of 10 plant species assayed, three species: *Boerhavia elegans* (Choisy), *Solanum surattense* (Burm.f.) and *Prosopis juliflora* (Sw.) showed promising anti-plasmodial activity *in vitro* ( $IC_{50} \leq 50 \mu\text{g/ml}$ ) and *in vivo* with no toxicity. The dichloromethane fraction of three extracts revealed stronger anti-plasmodial activity than the total extracts.

**Conclusion:** Anti-plasmodial activities of extracts of *B. elegans* and *S. surattense* are reported for the first time.

## Background

Malaria is one of the oldest recorded diseases in the world. Each year 300 to 500 million new cases are diagnosed and approximately 1.5 million people die of the disease; the majority of them are children [1]. The re-emerging of malaria in many parts of the world is due to the rapid increase of resistance to most of the available anti-malarial drugs, as well as resistance of vectors to insecticides [2,3]. Drug resistant strains of *P. falciparum* have been found in many endemic areas of the world and many of conventional anti-malarial drugs have been associated with treatment failure. Furthermore, the difficulty of creating efficient vaccines and also adverse side-effects of the existing anti-malarial drugs highlight the urgent need for novel, well-tolerated anti-malarial drugs [2] for both prophylaxis and treatment of malaria.

History reveals that plants have always been considered as an important source of medicine against malaria: both

quinine and artemisinin have been derived from traditional medicine and plant extracts. Artemisinin derivatives are now recommended by the World Health Organization worldwide [4,5], in combination with other drugs, such as lumefantrine, amodiaquine, mefloquine, sulphadoxine-pyrimethamine (SP), as the first-line treatment of malaria. This fact has encouraged the continuing search for new natural product-derived anti-malarial drugs. In malaria-endemic countries, several plants are utilized in traditional medicine for the treatment of malaria and/or fever. Furthermore, several studies have been undertaken to evaluate not only the inhibitory effects of various plant extracts on *P. falciparum* [6,7] using *in vitro* culture, but also *in vivo* anti-malarial properties on *Plasmodium berghei*-infected mice [8,9].

Malaria is endemic in Iran with 16,000 cases in 2008 (Center for Diseases Management and Control, Ministry of Health and Medical Education, unpublished). Despite intensive efforts to control malaria, the disease continues to be one of the greatest health problems in the south-eastern part of the country. In Iran, *P. falciparum* resistance to CQ has been reported since 1983 [10]. As the

\* Correspondence: zakeris@yahoo.com

<sup>1</sup> Malaria and Vector Research Group (MVRG), Biotechnology Research Center, Institut Pasteur of Iran, Tehran, Iran, Pasteur Avenue, P.O. BOX 1316943551, Tehran, Iran

Full list of author information is available at the end of the article

intensity of CQ resistance increased, the country implemented a change of first-line anti-malarial treatment in 2007 to a combination of SP-artesunate with artemether-lumefantrine (Coartem<sup>®</sup>), as the second-line drug [11]. Considering the great potential of Iranian plant biodiversity and based on ancient Iranian traditional physician's books [12-15], 10 different species of Iranian medicinal plants were selected in this investigation. The aim of this study was to discover novel, effective plant-based medicines for the treatment of malaria. In light of this, *in vitro* screening the anti-plasmodial properties of these plant species and their possible cytotoxic activities were also determined using the brine shrimp assay [16]. Subsequently, based on *in vitro* results, the active extracts were followed-up in a murine model of malaria.

## Methods

### Selection and collection of plant materials

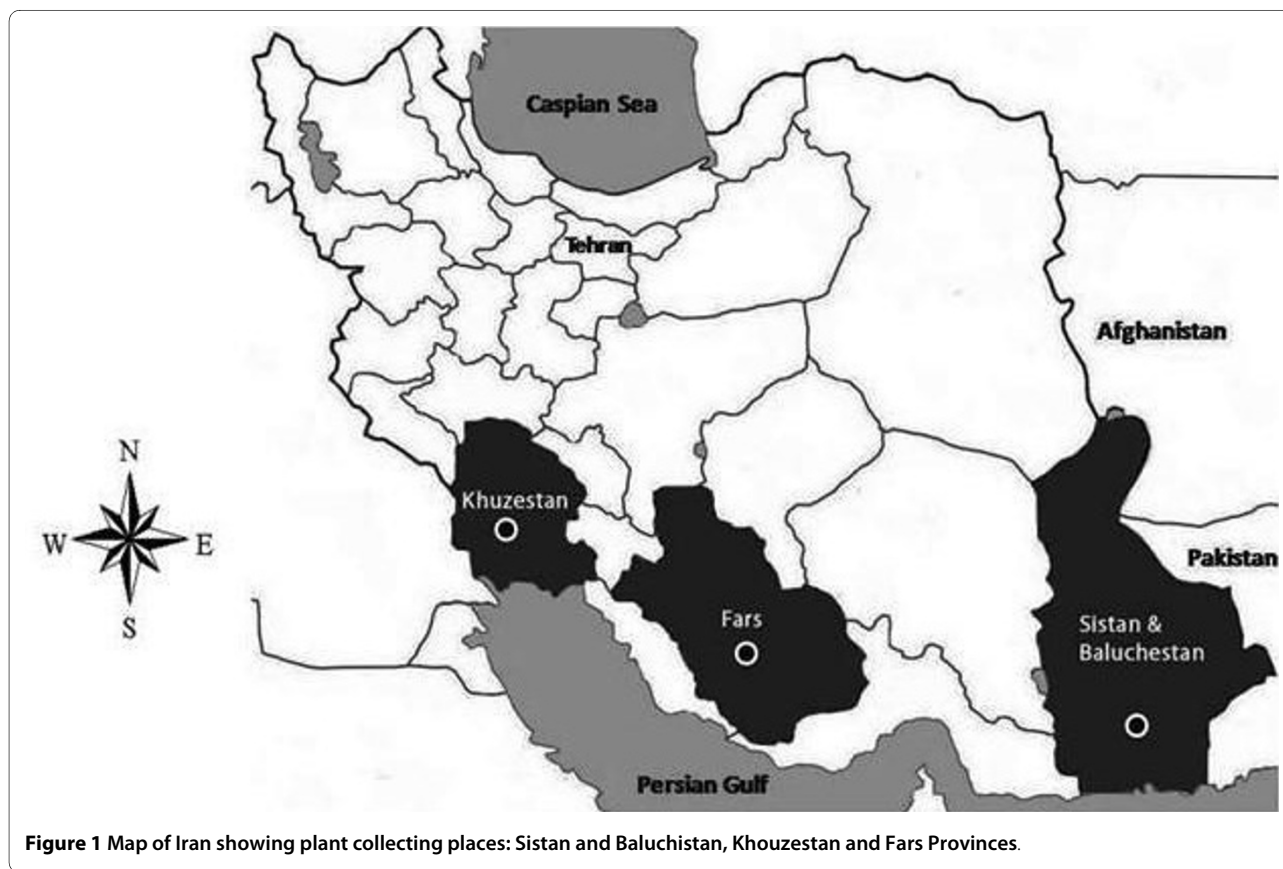
Based on literature and interviews with traditional health practitioners, plants representing five families, seven genera, and 10 species that used against fever, inflammation and microbial infection have been selected. The plants were collected from different parts of Iran, Sistan and Baluchistan, Khuzestan and Fars provinces, from March 2005 to October 2007 (Figure 1). Collections were undertaken from areas endemic for malaria at the present (Sis-

tan and Baluchistan) and that had been endemic in the past (Khuzestan and Fars).

The specimens were identified by Dr. S. Sardari and voucher specimens were deposited at Drug Discovery Group of Biotechnology Research Center at Institut Pasteur of Iran (Table 1). After identification, aerial parts of each specimen were air-dried in shadow at room temperature. The plants were crushed into fine powder using an electric grinder. The powdered samples were stored in appropriate containers and were kept at a cold room (4°C). The studied plant species were indicated in Table 1.

### Extraction and fractionation

For each plant, 100 g of powdered materials was extracted by percolation three times using 80% ethanol at room temperature. The ethanol extracts were filtered, pooled and dried at 40°C or below using a rotary evaporator. All the extracts were kept in airtight containers and were stored at 4°C for use in anti-plasmodial bioassay and general toxicity tests. Crude ethanolic extracts that showed good anti-plasmodial activity against *P. falciparum* strains, were further fractionated by partitioning in water and dichloromethane. The organic and aqueous phases were concentrated and dried by rotary evaporator and were dissolved in dimethylsulphoxide (DMSO) and



**Figure 1** Map of Iran showing plant collecting places: Sistan and Baluchistan, Khuzestan and Fars Provinces.

**Table 1: Names and origins of the selected plants of Iran**

Family	Species	Locality (province)	Date of collection	Some traditional uses (Jorjani, 1992; Khorasani, 1992; Avicenna, 2004)	Voucher no.
Nyctaginaceae	<i>Boerhavia elegans</i>	S & B	March, 2005	Dysmenorrhea, urinary tract and intestinal infections, inflammation	CH-3
Solanaceae	<i>Solanum surattense</i>	S & B	March, 2005	Antipyretic, inflammation	CH-45
Solanaceae	<i>Solanum alatum</i>	Khouzestan	October, 2007	Headache, infection, wound healing, dysentery	74-10
Fabaceae	<i>Prosopis juliflora</i>	Khouzestan	October, 2007	Fever, inflammation	74-32
Moraceae	<i>Ficus bengalensis</i>	Khouzestan	May, 2007	Wound healing, sedative, anti-rheumatism, inflammation	74-31
Moraceae	<i>Ficus carica</i>	Khouzestan	May, 2007	Ascites, a Anemia, inflammation	74-46
Rutaceae	<i>Citrus limon</i>	Khouzestan	May, 2007	disinfectant, rheumatism, inflammation	74-35
Moraceae	<i>Morus alba</i>	Khouzestan	May, 2007	Antipyretic, diuretic, laxative	74-38
Fabaceae	<i>Acacia farnesiana</i>	Khouzestan	May, 2007	wound healing, analgesic	74-37
Rutaceae	<i>Citrus aurantifolia</i> Swingle	Fars	September, 2007	Disinfectant, sedative, cold, appetite,	78-3

S & B = Sistan and Baluchistan

double distilled water, respectively, and then stored at -20°C for use in anti-plasmodial assay.

#### Plasmodium falciparum strains and in vitro culture

Laboratory-adapted *P. falciparum* K1 (chloroquine-resistant) and CY27 (chloroquine-sensitive) strains, both originally obtained from Thai patients, were continuously cultured based on a modified method previously described [17]. Briefly, parasites were maintained in continuous culture on human erythrocytes (blood group O<sup>+</sup> obtained from the Blood Transfusion Organization, Tehran, Iran), in RPMI 1640 medium supplemented with 10% human AB<sup>+</sup> serum, 25 mM N-2-hydroxyethylpiperazine-N-2-ethanesulfonic acid (HEPES), 25 mM NaHCO<sub>3</sub>, and 60 µg/ml gentamicin sulfate, at pH 7.2. The cultures were incubated at 37°C in an atmosphere of 91% N<sub>2</sub>, 6% CO<sub>2</sub> and 3% O<sub>2</sub>. Parasite cultures were synchronized to the ring stage by treatment with 5% D-sorbitol [18].

#### In vitro anti-plasmodial assay

Plant extracts were assessed for anti-plasmodial activity *in vitro* using modified parasite lactate dehydrogenase (pLDH) method as described previously [19,20]. Crude plant extracts were first dissolved in DMSO at concentration of 50 mg/ml, sonicated for 10 min and then diluted in malaria culture medium to prepare a 2 mg/ml solution. The highest concentration of solvent that the parasites were exposed to was < 1%, which was shown to have no

measurable effect on parasite viability. Microtitration techniques were used to measure the activity of samples over a wide range of concentrations (ranging from 200-1.56 µg/ml). Chloroquine diphosphate and artemisinin (both from Sigma Chemical, USA) were dissolved in double distilled water (1 mg/ml) and DMSO (1 mg/ml), respectively and served as controls in all experiments. All tests were performed in triplicate. Synchronous cultures with parasitaemia of 1% and a final haematocrit of 1.5% were aliquoted into the plates and incubated at 37°C for 72 h. After incubation period, the plates were frozen at -20°C overnight, followed by thawing at room temperature to haemolyze the red blood cells. Parasite growth was determined spectrophotometrically at 650 nm, by measuring the activity of the pLDH in control and drug-treated cultures, using a microplate reader (PowerWave 340, BioTek, USA). At the end of incubation, the cultures were resuspended, and aliquots of 20 µl were removed and added to 100 µl of the Malstat reagent [19,20] in a 96-well microtiter plate. The spectrophotometric assessment of pLDH activity was obtained by adding 25 µl of a solution of 1.9 µM NBT (Nitro Blue Tetrazolium) and 0.24 µM PES (Phenazine Ethosulphate) to the Malstat reagent. The anti-malarial activity of the test compound was expressed as IC<sub>50</sub> (mean ± S.D. of the least three separate experiments performed in triplicate). The OD values from control wells devoid of plant extracts or drug were referred to as having 100% pLDH activity. The inhibition

of each extract or drug concentration was calculated as compared to the untreated control to obtain the  $IC_{50}$  values. These values were then expressed as a percentage of 100% growth value and plotted against corresponding concentrations of the drug, using Gene5 microplate data collection and analysis software (BioTek, USA) to generate log dose-response curves.

#### Brine shrimp toxicity assay

Inhibition of the growth of brine shrimps (*Artemia salina*) was used as a measure of the toxicity of the extracts. The general toxicity in brine shrimp lethality test was carried out according to the methodology described before [16] by using brine shrimp eggs, obtained from Artemia and Aquatic Animals Research Center (Urmia University, Urmia, Iran). The eggs were hatched in artificial sea water (38 g/lit saline) in a vessel described previously [21]. After a 28-30 h hatching period at room temperature, the nauplii were ready for the experiment.

Extracts were prepared by dissolving 50 mg/ml in DMSO and then serially diluted in artificial sea water to the desired concentrations. The final DMSO concentration did not exceed 1%, which was shown not to have any harmful effects on the larvae. Finally, ten shrimps in 100  $\mu$ l of artificial sea water were added to each vial to give a final volume of 2.5 ml. After 24 h, the survivors were counted under microscope and recorded. Gallic acid ( $LC_{50} = 20 \mu$ g/ml) was used as a positive control, and vials containing shrimps, but without any extracts were included in each test. The toxicity of each extract was determined from the 50% lethality dose ( $LC_{50}$ ) by Finney's Probit analysis [22] of the data created with SPSS 15.0 for windows (SPSS Inc., USA). The general toxicity activity was considered weak when the  $LC_{50}$  was between 500 and 1000  $\mu$ g/ml, moderate when the  $LC_{50}$  was between 100 and 500  $\mu$ g/ml, and strong when the  $LC_{50}$  ranged from  $\leq 100 \mu$ g/ml [23].

#### In vivo anti-plasmodial assay

The anti-plasmodial activity of the *Prosopis juliflora* (Sw.), *Boerhavia elegans* (Choisy), and *Solanum surattense* (Burm.f.) extracts were assessed by the classical 4-day suppressive test [24,25]. Female BALB/C mice, weight 18-20 g were infected by interaperitoneal (i.p.) inoculation of  $10^7$  infected erythrocytes with CQ-sensitive *P. berghei* (ANKA strain) in a saline suspension of 0.2 ml on the first day (D0) of the experiment.

Plant extracts were solubilized in 18% DMSO or in PBS (water-soluble extract) and administrated within 3 h post-inoculation of mice with the parasite (D0) at different concentrations in a dose volume of 0.2 ml. Groups of five mice were dosed daily by i.p. injection for 4 consecu-

tive days. On day 4, tail blood smear were taken, stained with 10% Giemsa in phosphate buffer, pH 7.2 for 15 min and examined under microscope at 100  $\times$ . The percentage parasitaemia was determined by counting the parasitized red blood cells on at least 3,000 red blood cells. The % suppression of parasitaemia was calculated for each extract by comparing the parasitaemia present in infected controls with those of test mice. Chloroquine diphosphate (25 mg/kg), PBS and DMSO (18%) were used as positive and negative controls, respectively. For all the groups survival time in days was recorded and the mean for each group calculated. The results were analysed statistically using one-way ANOVA and two-tailed Student's *t*-test (SPSS 15.0 Inc., USA) to identify the differences between treated group and controls. The data were considered significant at  $P < 0.05$ .

## Results

#### In vitro anti-plasmodial assay

*In vitro* activity of 10 plant extracts against CQ-sensitive (CY27) and -resistant (K1) *P. falciparum* strains are summarized in Table 2. Of 10 extracts tested, three showed significant anti-plasmodial activity at a concentration of 4.68 to 50  $\mu$ g/ml. Three hydro-alcoholic extracts of *Pr. juliflora*, *B. elegans* and *S. surattense*. showed significant anti-plasmodial activity against both chloroquine-resistant (K1) ( $IC_{50}$ : 14.78, 15.33 and 50  $\mu$ g/ml, respectively) and chloroquine-sensitive (CY27) ( $IC_{50}$ : 4.68, 11.97 and 40.88  $\mu$ g/ml, respectively). The rest of the other plant extracts showed weak or no activity ( $IC_{50} > 200 \mu$ g/ml) against the studied isolates. The selectivity index (SI) is defined as the ratio of the brine shrimp toxicity to the anti-plasmodial activity and is determined by dividing the  $LC_{50}$  values for the brine shrimp nauplii by the  $IC_{50}$  value for *P. falciparum*. The extract with higher selectivity (high SI value) indicate potentially safer therapy.

The crude extract of three active extracts (*Pr. juliflora*, *B. elegans* and *S. surattense*) were further fractionated in water and dichloromethane mixture solution and the anti-plasmodial activity of organic and aqueous phases was evaluated against CQ-resistant and -sensitive *P. falciparum* strains. The dichloromethane fraction of the three extracts also showed stronger anti-plasmodial activity than the total extracts (Table 3).

#### Brine shrimp toxicity test

All the ethanolic active extracts were screened against brine shrimp nauplii and the results showed no significant or toxicity activity ( $LC_{50} > 1,000 \mu$ g/ml) for *Pr. juliflora*, *B. elegans*, *S. surattense*, *Solanum alatum* and *Ficus bengalensis*. However, the rest of the extracts showed mild toxicity ( $LC_{50}$  100-500  $\mu$ g/ml) (Table 2).

**Table 2: In vitro bioassays on *P. falciparum* CY27 and K1 strains and toxicity assay of the selected plants**

No.	Scientific name		<i>Artemia salina</i> assay, LC50 (µg/ml)	Anti-plasmodial activity, IC50 (µg/ml) ± SD		SI	
	Family	Species		K1	CY27	K1	CY27
1	Nyctaginaceae	<i>Boerhavia elegans</i>	1020	15.33 ± 0.07	11.97 ± 0.05	18.19	23.3
2	Solanaceae	<i>Solanum surattense</i>	2507	50 ± 0.09	40.88 ± 0.84	61.32	48.86
3	Solanaceae	<i>Solanum alatum</i>	1050	>200	>200	NA	NA
4	Fabaceae	<i>Prosopis juliflora</i>	1636.68	14.78 ± 0.08	4.68 ± 0.03	110.73	349.71
5	Moraceae	<i>Ficus bengalensis</i>	1050	>200	>200	NA	NA
6	Moraceae	<i>Ficus carica</i>	130.16	>200	>200	NA	NA
7	Rutaceae	<i>Citrus limon</i>	471.39	>200	>200	NA	NA
8	Moraceae	<i>Morus alba</i>	256.42	>200	>200	NA	NA
9	Fabaceae	<i>Acacia farnesiana</i>	460.12	>200	>200	NA	NA
10	Rutaceae	<i>Citrus aurantifolia</i> Swingle	270.17	>200	>200	NA	NA
		Gallic acid	20	ND	ND	NA	NA
Controls		Chloroquine	ND	1.25 ± 0.08	0.4 ± 0.01	NA	NA
		Artemisinin	ND	0.002 ± 0.008	0.007 ± 0.0005	NA	NA

**In vivo anti-malarial activity of *Prosopis juliflora*, *Boerhavia elegans* and *Solanum surattense* on *Plasmodium berghei***  
 Table 4 shows a summary of parasitaemia suppression (%) for mice on day 4. Three extracts from three plant species showed a significant suppression of parasitaemia ( $P < 0.05$ ) ranging from 44.1 to 66.2%. In contrast, results with the water-soluble extracts of *Pr. juliflora* were non-significant ( $P > 0.05$ ) (Table 4). Based on day 10 post-infection, *S. surattense* extract gave 80%, but *Pr. juliflora* and *B. elegans* gave 100% mouse survival.

## Discussion

In this investigation, anti-plasmodial activity of 10 Iranian traditional selected plants, representing five families was studied. The anti-plasmodial activities of the extracts were qualified as "active" when  $IC_{50}$  is  $\leq 50$  µg/ml. Extracts having activity beyond this range were considered inactive. The selectivity index (SI) is defined as the ratio of the cytotoxicity on the brine shrimp to the anti-plasmodial activity. Those that showed high SI ( $>10$ ) should offer the potential for safer therapy.

**Table 3: Anti-plasmodial activity (IC<sub>50</sub>) of water-soluble and dichloromethane fractions of three active plants against *P. falciparum* strains**

Plant species	IC50 (µg/ml) ± *SD			
	K1		CY27	
	Aqueous	Dichloromethane	Aqueous	Dichloromethane
<i>B. elegans</i>	>200	15 ± 0.38	>200	9.37 ± 0.01
<i>S. surattense</i>	>200	40 ± 0.16	>200	18.75 ± 0.4
<i>Pr. juliflora</i>	32.43 ± 0.2	9.95 ± 0.06	18 ± 0.16	1.4 ± 0.02

\*SD = Standard deviation

Among 10 plants, three species, *Pr. juliflora*, *B. elegans* and *S. surattense* showed promising anti-plasmodial activity with IC<sub>50</sub> values of ≤ 50 µg/ml and SI ranging between 18.19 and 349.71. From these three species, two (*Pr. juliflora* and *B. elegans*) were found to be highly active with IC<sub>50</sub> values of < 20 µg/ml. The crude extract of these three active herbs was further fractionated with a mixture of water and dichloromethane. The dichloromethane fraction of three active plants showed anti-plasmodial activity, but only the aqueous phase of *Pr. juliflora* showed activity against two strains of *P. falciparum*. Fortunately, these three active plants had low toxicity indicating the possibility of using them as lead structures in further studies.

*Prosopis juliflora* (IC<sub>50</sub> = 14.78 µg/ml on K1 strain, SI = 110.73 and IC<sub>50</sub> = 4.68 µg/ml on CY27 strain, SI = 349.71) is a tree native to Mexico, South America and Caribbean

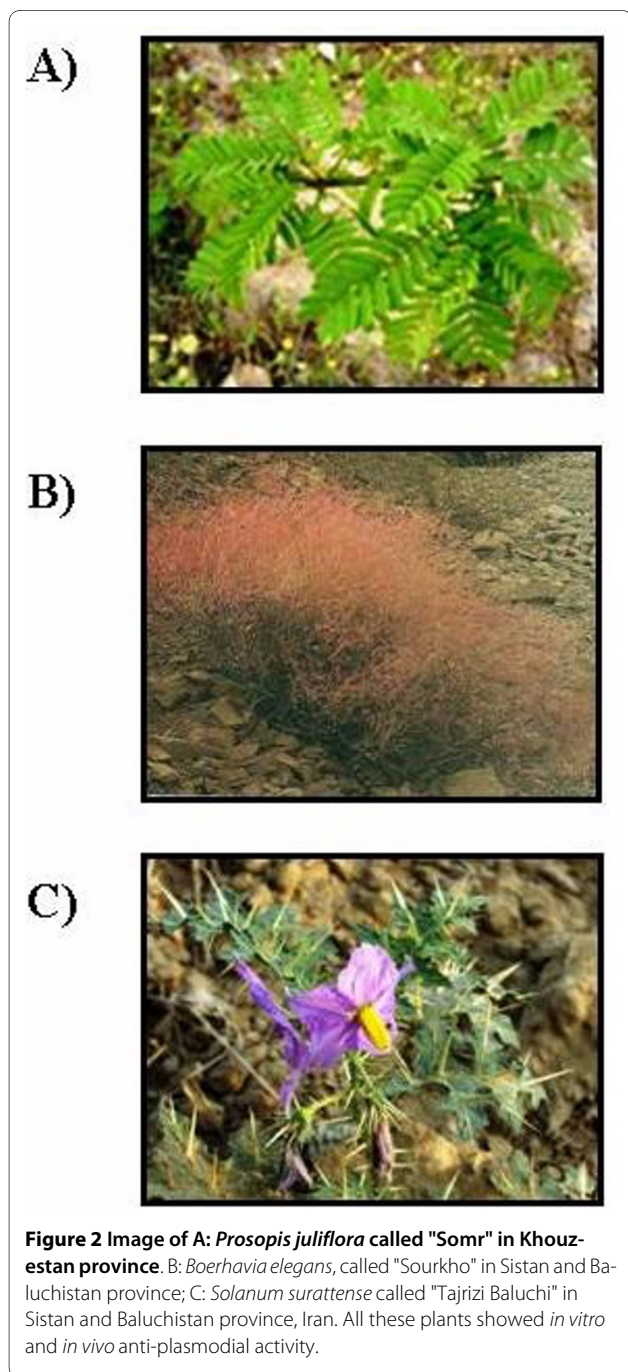
that later become established as a weed in Asia, Australia and elsewhere (Figure 2A). *Pr. juliflora* has been used in traditional medicine for fever, inflammation, scurvy, as a disinfectant, healing of wounds and skin problems [26]. The anti-malarial activity of this plant against CQ-susceptible strain of *P. falciparum* (3D7) was previously reported [27] and the present result correlates well with that and our study confirmed this observation with higher activity than previous study against both CQ-resistant (K1) and -sensitive (CY27) strains. Furthermore, treatment with the dichloromethane extract of *Pr. juliflora* significantly inhibited parasitaemia of *P. berghei* infection in BALB/c mice compared to no treatment, implying direct parasiticidal activity. The water-soluble extract of *Pr. juliflora* was active in *in vitro* against both sensitive and resistance *P. falciparum* strains, but had no *in vivo* activity against *P. berghei*. The results for *in vivo* anti-malarial activity do not necessarily correlate with

**Table 4: *In vivo* activities of plant extracts against *P. berghei***

Plant species	Extract/drug	Dose (mg/kg/day)	Suppression of parasitaemia at day 4 (SP ± SD*)	Survival (%) on day 10	P value (P < 0.05)
<i>S. surattense</i>	Dichloromethane	100	58.11 ± 4.3	80	0.035
<i>Pr. juliflora</i>	Dichloromethane	100	44.13 ± 1.79	100	0.008
	Water-soluble extract	50	2.59 ± 1.54	100	0.598
<i>B. elegans</i>	Dichloromethane	300	66.18 ± 2.11	100	0.007
	PBS	-	0	100	
	DMSO	18%	0	100	
	CQ	25	100	100	

\*SD = Standard deviation





those for *in vitro* anti-malarial activity as reported before [28] and this may be due to poor bioavailability of the active compounds *in vivo*.

*Boerhavia elegans* is an erect herb up to 1 m high, with a stout rootstock. The stems fleshy become woody towards the base, which is green, often flushed with red, glabrescent to pubescent, branching mainly from the base with the nodes swollen. Its leaves broadly ovate to lanceolate, green above, grayish-white beneath, sometimes tinged purple (Figure 2B). It showed promising anti-plasmodial activity with an  $IC_{50} = 15.33 \mu\text{g/ml}$  on K1 strain

( $SI = 18.19$ ) and  $IC_{50} = 11.97 \mu\text{g/ml}$  on CY27 strain ( $SI = 23.3$ ). The dichloromethane extract of *B. elegans* had the highest suppressive activity on *P. berghei* (66.18%) at a concentration of 300 mg/kg/per day. The result has shown promising suppressive and curative anti-malarial activities in *P. berghei* infected mice.

*Solanum surattense* is a suffrutescent perennial under-shrub plant, which grows in dry situations throughout India. The zigzag branches spread close to the ground covered with strong, broad and sharp yellowish white prickles. Its leaves armed with sharp prickles, blue flowers, fruit berries, yellow or white (Figure 2C). This species also showed anti-plasmodial activity with an  $IC_{50} = 50 \mu\text{g/ml}$  on K1 strain ( $SI = 61.32$ ) and  $IC_{50} = 40.88 \mu\text{g/ml}$  on CY27 strain ( $SI = 48.86$ ) and *in vivo* anti-plasmodial activities with 58.11% suppression.

*B. elegans* and *S. surattense* are growing in the south-eastern of Iran, where malaria is endemic. In Sistan and Baluchistan province, people use the extract of *S. surattense* against fever and infections. Also, this plant and *B. elegans* are used for inflammation in folk medicine [29]. No biological properties have been reported in the literature for these species, and the anti-plasmodial activity of these two plants is reported for the first time in the present study. It is interesting to note that the majority of the aqueous extracts did not show any activity. However, the dichloromethane fraction of three extracts showed better anti-plasmodial activity than the total extracts.

In this study, a number of selected plants did not display *in vitro* anti-plasmodial activity. Among them, *Acacia farnesiana* was collected from Khuzestan province in Iran and showed no anti-plasmodial activity. However, an ethanolic extract of this plant from Colombia had shown anti-plasmodial activity against *P. falciparum* [30]. A possible explanation could be that factors such as chemotypes, environmental parameters, harvesting and storage conditions that could collectively influence the plant metabolites prior to and following harvestings, which in turn, would be reflected in the bioactivity.

## Conclusion

Overall, increasing the global spread of multi-drug resistant malaria parasite showed that there is a need for new chemotherapeutic agents to combat malaria. In this study, aiming to search for new anti-malarial drugs, we found, for the first time, *in vitro* and *in vivo* anti-plasmodial activities of the plants *B. elegans* and *S. surattense*. The plant *Pr. juliflora* has already been described in the literature for anti-malarial activity against CQ-sensitive *P. falciparum* strain and the present study also confirmed these observations against CQ-resistant *P. falciparum* strain as well as on *P. berghei*. The present finding is only preliminary, the next step will be to isolate and identify the active compound of both *B. elegans* and *S. surattense*.

### Competing interests

The authors declare that they have no competing interests.

### Authors' contributions

AR (PhD student) contributed in the laboratory work, analysis of the data and drafted the paper. SZ designed the work and supervised the *in vitro* and *in vivo* studies, analysed the data and wrote the manuscript. SS contributed in making plant target list, collection and identification of plants, extraction and toxicity design. NK contributed in plant data gathering. NDD helped with the preliminary analysis of the data and also critical reading of the manuscript. All authors contributed to the critical review of the manuscript and agree to submission.

### Acknowledgements

We are grateful for the hospitality and generous collaboration of Zahedan University of Medical Sciences, and the staff of the Public Health Department, Sistan and Baluchistan province, Chabahar district (Dr. Nazari, Mr. Gorgij and Mr. Jadjal) for their assistance in collecting plant samples from the field. The authors are grateful to Professor M. Hommel (Liverpool School of Tropical Medicine) and Dr. H. Baseri (Department of Medical Entomology, School of Public Health, Tehran University) for providing K1, CY27 strains of *P. falciparum* and *P. berghei* (ANKA), respectively. We are grateful to the staff at the Blood Transfusion Organization, Tehran, Iran for supplying serum and blood products. This project (no. 289) received financial support from Institut Pasteur of Iran.

### Author Details

<sup>1</sup>Malaria and Vector Research Group (MVRG), Biotechnology Research Center, Institut Pasteur of Iran, Tehran, Iran, Pasteur Avenue, P.O. BOX 1316943551, Tehran, Iran and <sup>2</sup>Drug Discovery and Bioinformatics Group, Biotechnology Research Center of Institut Pasteur of Iran, Tehran, Iran

Received: 22 December 2009 Accepted: 12 May 2010

Published: 12 May 2010

### References

- Greenwood BM, Bojang K, Whitty CJ, Targett GA: **Malaria.** *Lancet* 2005, **365**:1487-1498.
- Ridley RG: **Medical need, scientific opportunity and the drive for antimalarial drugs.** *Nature* 2002, **415**:686-693.
- Zirihi GN, Mambu L, Guédé-Guina F, Bodo B, Grellier P: **In vitro antiparasitodal activity and cytotoxicity of 33 West African plants used for treatment of malaria.** *J Ethnopharmacol* 2005, **98**:281-285.
- WHO: **The use of antimalarial drugs, part II: Artemisinin and its derivatives.** [<http://www.rbm.who.int/>].
- Mutabingwa TK: **Artemisinin-based combination therapies (ACTs): best hope for malaria treatment but inaccessible to the needy!** *Acta Trop* 2005, **95**:305-315.
- Tran QL, Tezuka Y, Ueda JY, Nguyen NT, Maruyama Y, Begum K, Kim HS, Wataya Y, Tran QK, Kadota S: **In vitro antiparasitodal activity of antimalarial medicinal plants used in Vietnamese traditional medicine.** *J Ethnopharmacol* 2003, **86**:249-252.
- Wanyoike GN, Chhabra SC, Lang'at-Thoruwa CC, Omar SA: **Brine shrimp toxicity and antiparasitodal activity of five Kenyan medicinal plants.** *J Ethnopharmacol* 2004, **90**:129-133.
- Andrade-Neto VF, Brandão MG, Stehmann JR, Oliveira LA, Krettli AU: **Antimalarial activity of Cinchona-like plants used to treat fever and malaria in Brazil.** *J Ethnopharmacol* 2003, **87**:253-256.
- Sudhanshu Saxena NP, Jain DC, Bhakuni RS: **Antimalarial agents from plant sources.** *Current Science* 2003, **85**:1314-1329.
- Edrissian GH, Shahabi S, Pishva E, Hajseyed-Javadi J, Khaleghian B, Ghorbani M, Emadi AM, Afshar A, Saghari H: **Imported cases of chloroquine-resistant falciparum malaria in Iran.** *Bull Soc Pathol Exot* 1986, **79**:217-221.
- Zakeri S, Afsharpad M, Kazemzadeh T, Mehdizadeh K, Shabani A, Djadjid ND: **Association of *pfcr* but not *pfmdr1* alleles with chloroquine resistance in Iranian isolates of *Plasmodium falciparum*.** *Am J Trop Med Hyg* 2008, **78**:633-640.
- Jorjani L: **Zakhireh Kharazmshahi.** Tehran: Bonyade Farhang Iran; 1992.
- Khorasani A: **Makhzan ol-Advieh.** Tehran: Enghelabe e Eslami Publishing and Educational Organization; 1992.
- Avicenna H: **Qanoon.** In *Lithography* Tehran: Reprinted by Institute of Medical History Study, Islamic and Complementary Medicine; 2004.
- Emami SA, Ardakani Shams MR, Mehregan I: **Atlas of Medicinal Plants.** In *Traditional Medicine and Materia Medica Research Center (TMRC)* Shahid Beheshti University of Medical Sciences, Tehran, Iran; 2004.
- McLaughlin JLR, Rogers LL: **The use of biological assays to evaluate botanicals.** *Drug Inf J* 1998, **32**:513-524.
- Trager W, Jensen JB: **Human malaria parasites in continuous culture.** *Science* 1976, **193**:673-675.
- Lambros C, Vanderberg JP: **Synchronization of *Plasmodium falciparum* erythrocytic stages in culture.** *J Parasitol* 1979, **65**:418-420.
- Makler MT, Hinrichs DJ: **Measurement of the lactate dehydrogenase activity of *Plasmodium falciparum* as an assessment of parasitemia.** *Am J Trop Med Hyg* 1993, **48**:205-210.
- Makler MT, Ries JM, Williams JA, Bancroft JE, Piper RC, Gibbins BL, Hinrichs DJ: **Parasite Lactate Dehydrogenase as an Assay for *Plasmodium falciparum* Drug Sensitivity.** *Am J Trop Med Hyg* 1993, **48**:739-741.
- Meyer BN, Ferrigni NR, Putnam JE, Jacobsen LB, Nichols DE, McLaughlin JL: **Brine shrimp: A convenient general bioassay for active plant constituents.** *Planta Med* 1982, **45**:31-34.
- Finney DJ: **The adjustment for a natural response rate in probit analysis.** *Ann Appl Biol* 1949, **36**:187-195.
- Padmaja R, Arun PC, Prashanth D, Deepak M, Amit A, Anjana M: **Brine shrimp lethality bioassay of selected Indian medicinal plants.** *Fitoterapia* 2002, **73**:508-510.
- Peters W: **The chemotherapy of rodent malaria, XXII. The value of drug-resistant strains of *P. berghei* in screening for blood schizontocidal activity.** *Ann Trop Med Parasitol* 1975, **69**:155-171.
- Peters W, Robinson BL: **The chemotherapy of rodent malaria. XLVII. Studies on pyronaridine and other Mannich base antimalarials.** *Ann Trop Med Parasitol* 1992, **86**:455-465.
- de Albuquerque UP, Muniz de Medeiros P, de Almeida AL, Monteiro JM, Machado de Freitas Lins Neto E, Gomes de Melo J, dos Santos JP: **Medicinal plants of the caatinga (semi-arid) vegetation of NE Brazil: a quantitative approach.** *J Ethnopharmacol* 2007, **114**:325-354.
- Simonsen HT, Nordskjold JB, Smitt UW, Nyman U, Palpu P, Joshi P, Varughese G: **In vitro screening of Indian medicinal plants for antiplasmodial activity.** *J Ethnopharmacol* 2001, **74**:195-204.
- Gessler MC, Tanner M, Chollet J, Nkunya MHH, Heinrich M: **Tanzanian medicinal plants used traditionally for the treatment of Malaria: in vivo antimalarial and in vitro cytotoxic activities.** *Phytother Res* 1995, **9**:504-508.
- Zargari A: **Iranian medicinal plants.** In Tehran University Publications; 1987.
- Garavito G, Rincon J, Arteaga L, Hata Y, Bourdy G, Gimenez A, Pinzon R, Deharo E: **Antimalarial activity of some Colombian medicinal plants.** *J Ethnopharmacol* 2006, **107**:460-462.

doi: 10.1186/1475-2875-9-124

**Cite this article as:** Ramazani et al., In vitro and in vivo anti-malarial activity of *Boerhavia elegans* and *Solanum surattense* *Malaria Journal* 2010, **9**:124

**Submit your next manuscript to BioMed Central and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
[www.biomedcentral.com/submit](http://www.biomedcentral.com/submit)

