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Cohort study of the association of antibody levels to AMA1, MSP1₁₉, MSP3 and GLURP with protection from clinical malaria in Ghanaian children

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Published: 29 July 2008

Received: 12 March 2008

Malaria Journal 2008, **7**:142 doi:10.1186/1475-2875-7-142

Accepted: 29 July 2008

This article is available from: <http://www.malariajournal.com/content/7/1/142>

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Abstract

Background: Antigen-specific antibody-mediated immune responses play an important role in natural protection against clinical malaria, but conflicting estimates of this association have emerged from immuno-epidemiological studies in different geographical settings. This study was aimed at assessing in a standardized manner the relationship between the antibody responses to four malaria vaccine candidate antigens and protection from clinical malaria, in a cohort of Ghanaian children.

Methods: Standardized ELISA protocols were used to measure isotype and IgG subclass levels to Apical Membrane Antigen 1 (AMA1), Merozoite Surface Protein 1–19 (MSP1₁₉), Merozoite Surface Protein 3 (MSP3) and Glutamate Rich Protein (GLURP) antigens in plasma samples from 352 Ghanaian children, aged three to 10 years with subsequent malaria surveillance for nine months. This is one of a series of studies in different epidemiological settings using the same standardized ELISA protocols to permit comparisons of results from different laboratories.

Results: The incidence rate of malaria was 0.35 episodes per child per year. Isotype and IgG subclasses for all antigens investigated increased with age, while the risk of malaria decreased with age. After adjusting for age, higher levels of IgG to GLURP, MSP1₁₉, MSP3 and IgM to MSP1₁₉, MSP3 and AMA1 were associated with decreased malaria incidence. Of the IgG subclasses, only IgG1 to MSP1₁₉ was associated with reduced incidence of clinical malaria. A previous study in the same location failed to find an association of antibodies to MSP1₁₉ with clinical malaria. The disagreement may be due to differences in reagents, ELISA and analytical procedures used in the two studies.

When IgG, IgM and IgG subclass levels for all four antigens were included in a combined model, only IgG1 [(0.80 (0.67–0.97), $p = 0.018$)] and IgM [(0.48 (0.32–0.72), $p < 0.001$)] to MSP1₁₉ were independently associated with protection from malaria.

Conclusion: Using standardized procedures, the study has confirmed the importance of antibodies to MSP1₁₉ in reducing the risk of clinical malaria in Ghanaian children, thus substantiating its potential as a malaria vaccine candidate.

Background

Malaria remains one of the most important causes of morbidity and mortality in the world. Current methods of control are only partially effective and, therefore, the development of a vaccine which can provide a high degree of protection is a priority. Antibody-mediated immune responses to malaria antigens are known to be involved in protecting against disease [1-4], but the antigens that induce protective antibodies have not been conclusively identified. Immuno-epidemiological studies from different laboratories have sometimes yielded conflicting results [5-8]. This may be partly due to differences in malaria endemicity and the use of different study designs, reagents, assay protocols and statistical methodologies. In an attempt to make such studies more comparable, the Afro-Immuno Assay (AIA) network project was initiated. The network includes six African Institutions in Gabon, Ghana, Burkina Faso, Senegal, Tanzania, and Zimbabwe and three European Institutions from Denmark, The Netherlands and France. The Afro-Immuno Assay network has developed standardized enzyme immuno assays [9-11] that ensure the use of the same reagents, protocols and statistical methods to assess the association between acquisition of malaria specific antibody responses to four potential malaria vaccine candidate antigens and possible protection from clinical malaria. Samples for the AIA multi-center project were retrospectively obtained from cohort studies in six different geographical and epidemiological settings, comprising low endemic to holoendemic areas. These antigens include the Glutamate Rich Protein (GLURP), the Merozoite Surface Protein 3 (MSP3) [12], the 19-kilo Dalton region of the Merozoite Surface Protein 1 (MSP1₁₉) [13] and the Apical Membrane Antigen 1 (AMA1) [14], which are all thought to induce protective antibody responses through various mechanisms [15-18]. Vaccines incorporating these antigens are currently in clinical trials and are described in detail elsewhere [7,19-26]. It is likely that a future malaria vaccine will comprise multiple rather than single antigens and it is, therefore, useful to study natural immune responses to multiple malaria antigens in relation to incidence of malaria in a more standardized way. In this study, the standardized AIA ELISA procedures [9-11], were used to assess the relationship between incidence of clinical malaria and naturally acquired isotype and IgG subclass responses to these four leading malaria vaccine candidate antigens, AMA1,

MSP1₁₉, MSP3 and GLURP in Ghanaian children from three to 10 years of age.

Materials and methods

Study area, study population and morbidity surveillance

Samples used in this study were obtained in March 2002 from a longitudinal study conducted in Dodowa, in which 352 children aged three to 10 years (in the active phase of acquiring immunity to malaria), were enrolled in a study, whose original aim had been to assess the role of cytokine regulation and immunity to malaria. Dodowa is a semi-rural town in the Dangme West District of the Greater Accra Region of Ghana, about 50 km from the capital Accra and is an area of moderate and stable malaria transmission with a seasonal peak. Bed net coverage in this area was low, about 10% [27]. The study was approved by the Noguchi Memorial Institute's Ethical Review Board. After obtaining consent from parents, blood was obtained from each child and plasma stored at -20°C until use. Malaria episodes were detected using both active and passive surveillance implemented over a period of nine-months, spanning the entire malaria transmission season. Clinical and parasitological information was captured using a standard questionnaire. Each child was visited once a week and the child and the parents or guardians were asked about symptoms of malaria since the last visit and whether she/he had received any anti-malarial treatment. The child was then given a physical examination and the body temperature measured. Children with a history of fever within 48 hours and/or axillary temperature equal to or above 37.5°C had a rapid test for malaria parasitaemia using OptiMAL™ (DiaMed, FLOW Inc. Portland, Oregon) and then thick and thin blood films were prepared for microscopy for estimating the parasite density which was used later in the more specific malaria case definition employed for data analysis. Only children with measured or reported fever, and with a positive rapid test were treated with chloroquine (in accordance with the then prevailing national malaria treatment policy) and in the case of severe symptoms, the child was referred to the hospital. At monthly intervals, blood smears from finger pricks were obtained from all children irrespective of symptoms to estimate the prevalence of asymptomatic malaria infections. The parents of the children were instructed to report to the field team if the child had any symptoms of disease at any time. The

qualifying case definition for malaria in the data analysis was reported fever and/or a measured temperature equal to or above 37.5°C, with parasitaemia $\geq 5,000$ parasites/ μl of blood; this case definition has been found to have 90% sensitivity and specificity in the study area [28-30]. Positive and negative control plasma used in ELISA measurements were obtained from adult Liberians and Danes respectively.

Malaria antigens

AMA1 was from the *Pichia pastoris* expressed ectodomain of *Plasmodium falciparum* FVO strain comprising amino acids 25–545 [31] (Donated by A Thomas, Biomedical Primate Research Centre). GLURP was an *Escherichia coli* recombinant protein containing the conserved non-repeat N-terminal region (amino acids 25–514) called R0 [32] (Donated by M. Theisen, Statens Serum Institut). MSP1₁₉ was a *Baculovirus* antigen of the C-terminal region of the merozoite protein surface 1, produced in insect cells infected with a recombinant *Baculovirus* containing a synthetic G-C enriched PfMSP1 gene (Palo Alto allele), coding for 43 N-terminal MSP1 precursor residues and 16 amino acid residues upstream of the "classical" MSP-1₁₉ (NIS--FCS) [33] (Donated by S. Longacre, Institut Pasteur). The MSP3 antigen used in this study was a long synthetic peptide called LR55 (amino acids 181 – 276) of the merozoite surface protein 3 [34] (Donated by M. Theisen, Statens Serum Institut). All the antigens were provided through the AIA Project.

Enzyme-linked immunosorbent assay (ELISA)

Specific isotype and IgG subclass levels against GLURP, MSP1₁₉, MSP3 and AMA1 were measured using indirect ELISA according to the AIA standard ELISA protocols [9-11]. All antigens tested were optimized and shown to be stable for at least three weeks, when antigen-coated plates and serum/plasma dilutions are refrigerated. The subclass specific reagents used were selected on the basis of low cross reactivities among themselves. To control for inter-assay and day-to-day variations in the standardized ELISA procedure, three-fold serial dilutions of reference standard reagents (IgG, IgM and IgG1 to IgG4) were directly coated on each ELISA plate (Maxisorp Nunc, Denmark) at a start concentration of 1,000 ng/ml (100 μl /well). OD values for the test samples were converted into antibody units with the standard reference curves generated for each ELISA plate using a four parameter curve-fit Microsoft Excel-based application. Samples were re-tested if the coefficient of variation between duplicate absorbance values were higher than 15% and plates were also re-tested if the R-square value of the standard curve was less than 97%. The reference standards, PBS buffer blank, positive and negative control plasma pools that were included in each ELISA test plate allow for the determination of failed

assay runs. The AIA ELISA procedure used in this study is described in detail elsewhere [10].

Statistical analysis

Clinical data were double entered using Microsoft Fox Pro and immunological data using Excel. STATA version 9.2 (Statcorp, Texas) was used for statistical analysis. Children were considered to have a clinical malaria episode if they had parasitaemia of $\geq 5,000$ parasites/ μl , with a measured temperature $\geq 37.5^\circ\text{C}$ or a history of fever in the last 48 hours [6,35,36].

For each antigen, Poisson regression was used to investigate the association between the levels of antibody measured at baseline and the incidence rate of the first (or only) episode of clinical malaria. The level of total IgG, IgM and each IgG subclass, were analysed for each antigen in turn. Antibody values were transformed to log base 2, so that the rate ratio represents the ratio of malaria incidence corresponding to a doubling of antibody level. To investigate whether the relationship between malaria incidence and antibody level was nonlinear, a likelihood ratio test was used to compare the fit of the model when antibody level was included as a categorical or a continuous variable. When there were zero antibody values, indicating levels below the detection limit, the zero (left censored) values were assigned a nominal value equal to half the smallest measured value for that variable. If the proportion of zero values was large, the variable was treated as categorical with the reference category containing the zero values and the positive values divided into three equal groups. A likelihood ratio test was used to determine the P-value for the association with malaria incidence. Age at enrolment was considered to be an important potential confounder, and was included in the regressions as a factor with categories defined by quintiles. To model seasonality in malaria incidence, the calendar month of surveillance was included in the models as a factor. To construct a parsimonious model using all the immunological variables, firstly a model was produced for each antigen; in this model each IgG subclass, total IgG and IgM were candidates for inclusion provided the P-value for association with malaria incidence was 0.1 or less when considered individually. Variables were then removed from the model if the P-value for the likelihood ratio test was more than 0.1, provided removal did not change coefficients of variables in the model by more than 10%. In a second stage, the variables included in these models were candidates for inclusion in a final model derived in a similar way. Baseline parasitaemia was not considered as a potential confounder, but the interaction between each immunological variable and the presence of parasitaemia at baseline in their effects on malaria incidence was examined. A consequence of using a more specific case definition in the analysis than was used to decide

treatment during the study is that children could have received anti-malarial treatments during the period they are considered at risk in the analysis. To explore the impact of these drug treatments, a time dependent variable was defined, to allow for a reduced risk of malaria for a period of 28 days after each drug treatment, which was included as a covariate in the Poisson regression model. LOESS smoothing was applied in R software to plot antibody levels in relation to age. Spearman's rank correlation test was used to assess associations between antibody levels and age.

Results

Pattern of *P. falciparum* infections and malaria in the study cohort

Of the 352 children recruited for the original study, eight were lost to follow-up immediately after the baseline blood sampling. Of the 344 children followed up, sixty four (19%) had at least one episode of malaria (53 children had one episode, nine had two and two had three episodes). The incidence rate of malaria in the study cohort was 0.35 attacks per child per year (Table 1). The risk of clinical malaria decreased with increasing age [37]. Sixty-six percent of the children had asymptomatic parasitaemia at baseline. Parasites were predominantly *P. falciparum* (95%) and the prevalence of parasitaemia measured each month was roughly constant (ranging from 50% to 65%). The incidence of clinical malaria, however, varied during the survey period, rising gradually from March to May, peaking in July and then decreasing until November (Figure 1).

Relationship between age and antibody levels

Several studies in malaria endemic regions have shown increasing antibody levels with age. This pattern is more pronounced the greater the intensity and duration of malaria transmission. In this study, the relationship between antibody levels and age were assessed for antibodies against MSP1₁₉, AMA1, GLURP and MSP3. The levels of IgG and IgM to MSP1₁₉, MSP3, AMA1 and GLURP increased with age (Spearman correlation coefficient (r_s) 0.21 – 0.45; $p < 0.001$, Figure 2). IgG levels to AMA1 however, gradually increased until six years of age,

and then leveled off. For the four antigens tested, IgG1, IgG2 and IgG3 significantly increased with age (r_s , 0.12 – 0.36; $p < 0.03$) with the exception of IgG2 to AMA1. There was no evidence that the level of IgG4 was associated with age for any of the antigens. Like IgG to AMA1, IgG3 to MSP3 steadily increased with age until seven years of age, then leveled off (Figure 3).

The levels of both IgG and IgG1 were highest for AMA1 and lowest for MSP3, whereas IgG levels to MSP1₁₉ and GLURP were comparable (Figures 2 and 3). The levels of IgG3 were higher in AMA1 than the comparable levels in GLURP, MSP1₁₉ and MSP3 (Figure 3). In general, the levels of cytophilic IgG1 and IgG3 were higher than those of non-cytophilic IgG2 and IgG4, IgG4 levels being the lowest.

Antibody levels in relation to protection from clinical malaria

Total IgG to MSP3, MSP1₁₉, and GLURP, and IgM to all four antigens tested (MSP3, MSP1₁₉, GLURP, and AMA1) were associated with reduced malaria incidence in crude analyses (Table 2). The incidence of both clinical malaria and antibody levels were associated with age, age is therefore a potential confounder, and it is important to adjust for its effects. After adjusting for the effect of age, there was evidence of a significant association between total IgG to MSP3, MSP1₁₉, GLURP and reduced risk of malaria (IgG to MSP3: rate ratio 0.69 (95%CI 0.53, 0.90) $P = 0.01$; IgG to MSP1₁₉: 0.75 (0.61, 0.92) $P = 0.01$; IgG to GLURP: 0.79 (0.64, 0.98) $P = 0.04$); and IgM levels to AMA1, MSP3, MSP1₁₉ were also significantly associated with reduced risk of malaria (Table 2).

A large proportion of the measurements of IgG4 to MSP3 and AMA1 and IgG2 to AMA1 were zero (left-censored) values and so these variables were treated as categorical variables (Table 3). In the crude analysis, IgG1, IgG2 and IgG3 to MSP3, MSP1₁₉, GLURP and also IgG4 to MSP3, were associated with a reduced risk of clinical malaria (Table 3, 4), but after adjustment for age, only one of these variables, IgG1 to MSP1₁₉, remained significantly

Table 1: Malaria incidence by age group

Age in yrs:	No. of children	Cum. incidence of malaria	no episodes of malaria (years at risk)	Incidence rate per child year (95%CI)
3	32	12/32 (38%)	15 (19.52)	0.79 (0.48–1.31)
4–5	86	24/86 (28%)	28 (54.75)	0.51 (0.35–0.74)
6–7	79	11/79 (14%)	13 (51.38)	0.25 (0.15–0.44)
8	48	6/48 (13%)	7 (31.11)	0.22 (0.11–0.47)
9–10	99	11/99 (11%)	14 (61.22)	0.23 (0.14–0.39)
Total	344	64/344 (19%)	77 (217.48)	0.35 (0.28–0.44)

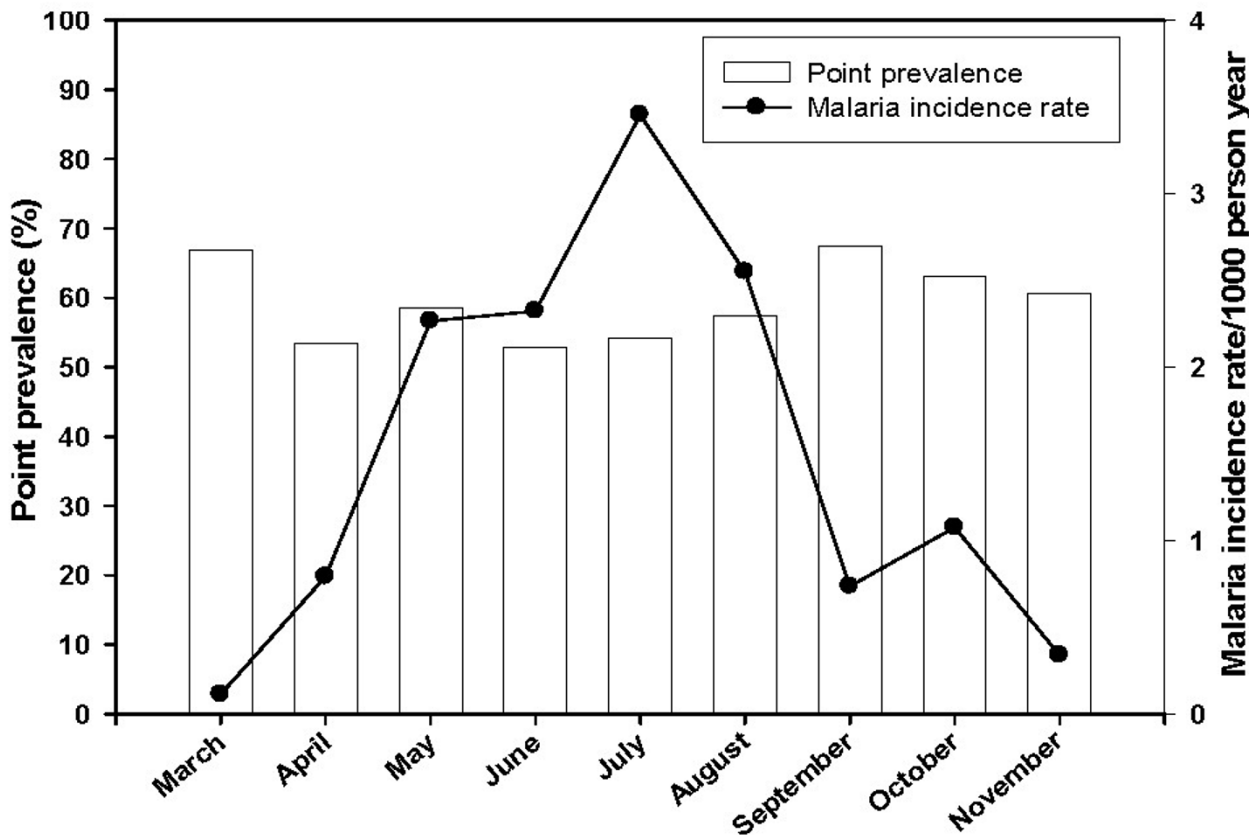


Figure 1
The monthly point prevalence of asymptomatic malaria parasitaemia and incident rate of clinical malaria. The point prevalence of asymptomatic parasitaemia for each month is represented as a bar graph and the pattern of clinical malaria (incident rate) is shown as a line graph.

associated with malaria incidence (rate ratio 0.89 (95%CI 0.80, 0.99), P = 0.04, Table 4).

When all immunological variables were considered simultaneously, only two variables were independently associated with reduced malaria incidence, IgG1 [(0.80 (0.66–0.96), p = 0.018)] to MSP1₁₉, and IgM [(0.48 (0.32–0.72), p < 0.001)] to MSP1₁₉ (Table 5).

The estimated rate ratio for the effect of antimalarial treatments given to children with parasitaemia < 5000/uL was 0.25 (95%CI 0.06–1.05), indicating these children had a substantially reduced risk of being found positive with parasitaemia ≥ 5000/ul during the 28 days following the treatment. But the estimates of rate ratios for other variables in the model were unchanged when this variable was included in the model suggesting treatment effects did not

cause a bias in the estimation of the effects of immunological variables.

Discussion

This study in Ghanaian children is one of a series of studies designed to assess, using standardized methods, the association of antibody levels to four leading asexual blood-stage malaria antigens (MSP1₁₉, MSP3, AMA1 and GLURP) with the incidence of clinical malaria in different epidemiological settings. Previous results have been difficult to interpret due to different study protocols and analytical methods having been used [5-7,9,14,29,38-40]. In this study, the prevalence of asymptomatic malaria parasitaemia was relatively high and stable, while incidence of clinical malaria fluctuated in parallel with the intensity of transmission and seasonal rainfall pattern. These patterns are typical of this area and have been reported in previous

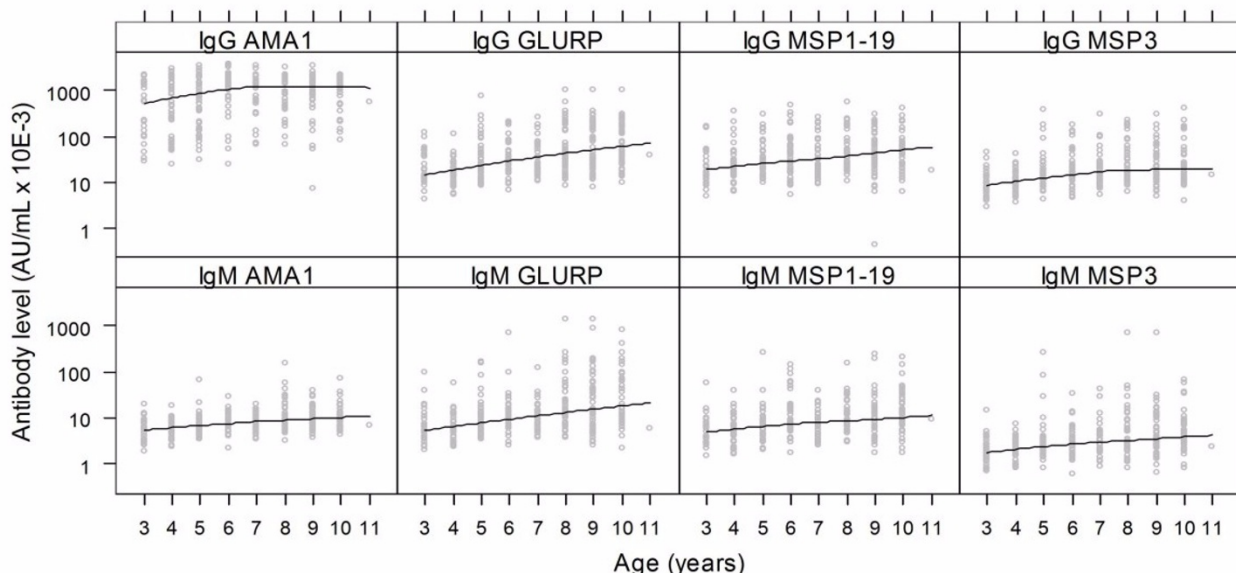


Figure 2
Baseline IgG and IgM levels in relation to age. The top and bottom panels represents total IgG or IgM levels to AMA1, GLURP, MSP1₁₉ and MSP3 in relation to age of Ghanaian children, respectively. The line shows the LOESS smoothed estimate of the geometric mean.

studies [6,41-43]. The variation in the incidence of clinical malaria during the study period may be due to the introduction of new parasites with different antigenic presentation into the population leading to clinical malaria in susceptible individuals. The risk of malaria decreased with age, while isotype and IgG subclass levels to the four antigens generally increased with age. This is consistent with the hypothesis that immunity to malaria is largely effected through antibody-mediated mechanisms and that protective antibody levels to relevant antigens increase with age-related exposure to the parasites [44]. Increasing IgG and IgM levels with age may reflect greater cumulative exposure of older children but may also be due to older children having a more mature immune system [45]. The association of IgM responses with reduced malaria incidence indicates a possible role in immunity in Ghanaian children. Although much emphasis has been placed on IgG as the important isotype in immunity against malaria, IgM, which has lower affinity but is multivalent, may afford protection via other mechanisms such as the blocking of merozoite invasion of erythrocytes, complement activation, agglutination of merozoites [46].

The association with malaria incidence of IgG responses to MSP3, MSP1₁₉ and GLURP is consistent with data from several other immuno-epidemiological studies [5,7-9,12,29,30,38,47-49] indicating that these antigens may be targets of protective antibodies [1,2,4]. There was no evidence that IgG levels to AMA1 were associated with

malaria incidence, and there was no evidence of an interaction with baseline parasitaemia in contrast with similar studies conducted in Kenya [14]. As shown in other studies, the cytophilic antibody levels to the four antigens tested in this study were higher than the non-cytophilic ones, emphasizing their importance in anti-malaria immunity [9,29,38,50,51]. IgG and cytophilic antibody levels were highest for AMA1, while the levels were relatively low for MSP3. These differences in specific antibody levels may be related to the number of immunogenic B-Cell epitopes exposed to the immune system and could also be related to the structure, location and function of the particular antigen(s). With the exception of IgG2 levels to AMA1, IgG levels to GLURP, MSP1₁₉ and MSP3 increased with age which may suggest that IgG2 is involved in immunity against malaria. In recent studies, malaria antigen specific IgG2 have been shown to bind with high affinity to mutant Fcγ RII H131->R receptors [52] on monocytes, granulocytes and B cells, thus affording protection against malaria through monocytes and or neutrophil mediated mechanisms in subjects expressing the mutant CD32 form [53]. There was however, no evidence found that IgG2 was associated with malaria incidence for any of the four antigens tested.

The IgG subclasses, IgG1, IgG2 and IgG3 for MSP3, MSP1₁₉, GLURP and IgG4 to MSP3 were associated with a reduced risk of malaria in un-adjusted analysis but of these only IgG1 to MSP1₁₉ was independently associated

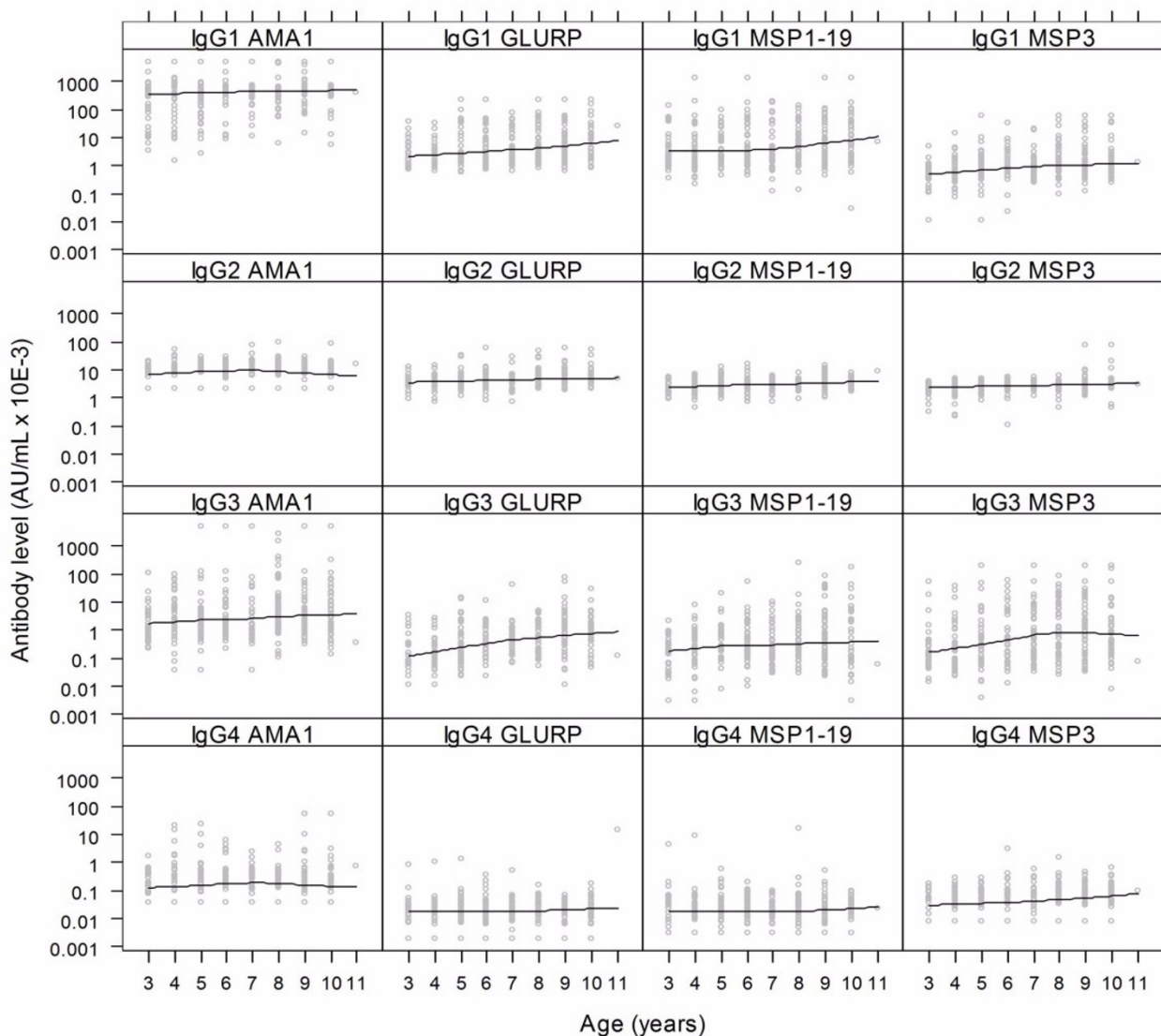


Figure 3
Baseline IgG subclass levels in relation to age. The top, 2nd, 3rd and 4th panels represent IgG subclasses 1 to 4 against AMA1, GLURP, MSP1₁₉ and MSP3 in relation to age of Ghanaian children, respectively. The line shows the LOESS smoothed estimate of the geometric mean.

with malaria incidence after adjustment for age. Other studies have shown the importance of IgG1 in clearing parasitaemia in children [5,47,54]. In a previous cohort study conducted in the same area, there was no association between antibody levels to MSP1₁₉ and malaria incidence [6]. This may be due to differences in antigen and antibody reagents used in the two studies; the MSP1₁₉ used in this study was a Baculovirus product that included a synthetic G-C enriched PfMSP1 gene that coded for the 43 N-terminal MSP1 precursor residues and 16 amino

acid residues upstream of the "classical" MSP-1₁₉ (NIS---FCS) [33] compared to the one produced in *E. coli*, which had been used in the previous study. Although antibodies to MSP1₁₉ have been shown to be associated with both exposure and protection from disease, the fine specificities of such responses may contribute to protection [40]. The antigen used in this study may have assessed antibodies of fine specificities that are protective [40,55,56], whereas the antigen used in the previous study did not. It may, therefore, be important to assess in a standardized

Table 2: Association of total IgG and IgM with malaria incidence

IgG	Crude IRR (95%CI)	P-value	IRR adjusted for age (95%CI)	P-value
MSP3	0.59 (0.45, 0.76)	< 0.0001	0.69 (0.53, 0.90)	0.01
MSP1 ₁₉	0.74 (0.63, 0.88)	< 0.0001	0.75 (0.61, 0.92)	0.01
GLURP	0.68 (0.55, 0.82)	< 0.0001	0.79 (0.64, 0.98)	0.04
AMA1	0.96 (0.84, 1.09)	0.50	1.03 (0.89, 1.20)	0.67
IgM:				
MSP3	0.62 (0.49, 0.80)	< 0.0001	0.76 (0.59, 0.97)	0.03
MSP1	0.59 (0.47, 0.75)	< 0.0001	0.68 (0.53, 0.88)	< 0.01
GLURP	0.71 (0.58, 0.87)	< 0.01	0.84 (0.68, 1.03)	0.09
AMA1	0.48 (0.34, 0.67)	< 0.0001	0.63 (0.44, 0.91)	0.01

Malaria incidence rate ratios (IRR) with 95% confidence interval, corresponding to a doubling of baseline antibody level, before and after adjustment for the effect of age.

Table 3: Association of IgG2 and IgG4 with malaria incidence.

Antigen	Antibody	Crude IRR (95%CI)	P-value	Age-adjusted IRR	P-value
MSP3					
	IgG2* (coded as log(2))	0.74 (0.57, 0.98)	0.04	0.85 (0.63, 1.15)	0.29
	IgG4 < detection limit		0.01		0.20
	< median	0.45 (0.21, 0.95)		0.56 (0.26, 1.19)	
	≥ median	0.47 (0.28, 0.79)		0.68 (0.40, 1.15)	
MSP1₁₉					
	IgG2* (coded as log(2))	0.62 (0.45, 0.86)	< 0.01	0.836 (0.606, 1.157)	0.275
	IgG4* (coded as log(2))	1.07 (0.95, 1.21)	0.25	0.838 (0.608, 1.159)	0.278
GLURP					
	IgG2* (coded as log(2))	0.65 (0.49, 0.86)	< 0.01	0.79 (0.58, 1.08)	0.13
	IgG4* (coded as log(2))	0.94 (0.81, 1.08)	0.37	1.01 (0.87, 1.16)	0.94
AMA1					
	IgG2 < detection limit		0.18		0.54
	< median	1.80 (0.86, 3.78)			
	≥ median	1.78 (0.90, 3.51)			
	IgG4 < detection limit		0.19		0.54
	< median	0.96 (0.47, 1.93)			
	≥ median	0.62 (0.36, 1.07)			

For IgG 2 to AMA1 and for IgG4 to AMA1 and MSP3, antibody level was categorized.

*For IgG2 to MSP3, MSP1₁₉, and GLURP, the association is expressed as the incidence rate ratio (IRR) corresponding to a doubling of baseline antibody level.

Table 4: Association of IgG1 and IgG3 with malaria incidence

IgG1:	Crude IRR (95%CI)	P-value	IRR adjusted for age (95%CI)	P-value
MSP3	0.82 (0.72, 0.94)	< 0.01	0.88 (0.76, 1.03)	0.11
MSP1 ₁₉	0.86 (0.77, 0.97)	0.01	0.89 (0.80, 0.99)	0.04
GLURP	0.85 (0.74, 0.98)	0.02	0.93 (0.81, 1.08)	0.35
AMA1	0.95 (0.86, 1.06)	0.36	1.01 (0.90, 1.14)	0.83
IgG3:				
MSP3	0.90 (0.82, 0.98)	0.01	0.94 (0.86, 1.03)	0.17
MSP1 ₁₉	0.90 (0.82, 0.99)	0.03	0.93 (0.84, 1.03)	0.18
GLURP	0.83 (0.74, 0.93)	< 0.01	0.91 (0.80, 1.03)	0.14
AMA1	0.97 (0.89, 1.06)	0.48	0.98 (0.90, 1.07)	0.73

The incidence rate ratio (IRR) corresponds to a doubling of baseline antibody level, before and after adjustment for effects of age

Table 5: Adjusted rate ratios for immunological variables independently associated with malaria risk in the final model.

	Crude IRR (95%CI)	P-value	IRR adjusted for age (95%CI)	P-value	IRR adjusted for effects of age and treatment (95%CI)
IgM to MSP1 ₁₉	0.44 (0.30–0.64)	P < 0.001	0.49 (0.33–0.73)	P < 0.001	0.48 (0.32–0.72)
IgG1 to MSP1 ₁₉	0.78 (0.65–0.93)	P = 0.005	0.80 (0.66–0.96)	P = 0.018	0.80 (0.67–0.97)
Treatment effect	0.28 (0.07–1.15)	P = 0.076	-	-	0.25 (0.06–1.05)

way the various MSP1₁₉, and other antigens, that are produced in different expression systems, in order to select the most appropriate antigen/expression system for malaria vaccine development. In another study in this series, in Burkina Faso, the same antigens, reagents, ELISA procedure and analytical methods were used; none of the isotypes and subclasses to MSP1₁₉ was associated with the incidence of clinical malaria. Since the same laboratory methods were used, the different outcomes of these two studies may be attributed to differences in malaria transmission or to the age of the children [44]. In Burkina Faso, the malaria transmission season is much shorter, which may influence the induction of differing antibody types for controlling malaria as shown in recent studies conducted in areas with different malaria endemicities in Tanzania [9,44]. Although total IgG to GLURP and MSP3 were associated with the risk of malaria, none of the constituent subclasses was identified to be associated with protection. When the effects of all the immunological variables were considered simultaneously, only IgG1 and IgM to MSP1₁₉ were independently associated with the incidence of clinical malaria, which may indicate the importance of MSP1₁₉ in malaria vaccine development. Parasite growth inhibition assays would be required to confirm if this association reflects a functional role of MSP1₁₉ in immunity.

Conclusion

In conclusion, using standardized AIA ELISA, anti-MSP1₁₉ antibodies (IgG1 and IgM) have been shown to be the most strongly correlated with reduced risk of clinical malaria among the four malaria vaccine candidates tested. The standardized AIA ELISA developed for this project could be used to validate malaria vaccine candidate antigens, provide useful baseline information for clinical trials, and contribute to quality assured laboratory capacity in Africa.

Authors' contributions

DD carried out field studies, developed assays and drafted the manuscript. AA performed the ELISA, compiled data and assisted in the manuscript writing. KAK and HL assisted with the ELISA. MT assisted with the assay development and manuscript writing, while PM, SB and ER wrote the analysis plan, performed the data analysis and

together with RC, BDA and YDO contributed to the writing of the manuscript. All authors read and approved the final manuscript.

Acknowledgements

The authors are grateful to the children in Dodowa community for participation in this study. Dr. Soren Jepsen is acknowledged for supporting and encouraging the Network. Dr. Ramadhani Abdalla Noor is also acknowledged for the support and management contribution.

This investigation received support of AMANET and Netherlands Ministry of Foreign Affairs (DGIS). The Wellcome Trust (grant 057736) is also acknowledged for funding the prospective study that generated plasma samples and morbidity survey data.

References

- Cohen S, McGREGOR IA, CARRINGTON S: **Gamma-globulin and acquired immunity to human malaria.** *Nature* 1961, **192**:733-737 [<http://>].
- Cohen S, Butcher GA: **Serum antibody in acquired malarial immunity.** *Trans R Soc Trop Med Hyg* 1971, **65**:125-135.
- Good MF: **Towards the development of the ideal malaria vaccine. A decade of progress in a difficult field.** *Med J Aust* 1991, **154**:284-289.
- Sabchareon A, Burnouf T, Ouattara D, Attanath P, Bouharoun-Tayoun H, Chantavanich P, Foucault C, Chongsuphaisiddhi T, Druilhe P: **Parasitologic and clinical human response to immunoglobulin administration in falciparum malaria.** *Am J Trop Med Hyg* 1991, **45**:297-308.
- Braga EM, Barros RM, Reis TA, Fontes CJ, Morais CG, Martins MS, Krettli AU: **Association of the IgG response to Plasmodium falciparum merozoite protein (C-terminal 19 kD) with clinical immunity to malaria in the Brazilian Amazon region.** *Am J Trop Med Hyg* 2002, **66**:461-466.
- Dodoo D, Theander TG, Kurtzhals JA, Koram K, Riley E, Akanmori BD, Nkrumah FK, Hviid L: **Levels of antibody to conserved parts of Plasmodium falciparum merozoite surface protein 1 in Ghanaian children are not associated with protection from clinical malaria.** *Infect Immun* 1999, **67**:2131-2137.
- Egan AF, Morris J, Barnish G, Allen S, Greenwood BM, Kaslow DC, Holder AA, Riley EM: **Clinical immunity to Plasmodium falciparum malaria is associated with serum antibodies to the 19-kDa C-terminal fragment of the merozoite surface antigen, PfMSP-1.** *J Infect Dis* 1996, **173**:765-769.
- Soe S, Theisen M, Roussilhon C, Aye KS, Druilhe P: **Association between protection against clinical malaria and antibodies to merozoite surface antigens in an area of hyperendemicity in Myanmar: complementarity between responses to merozoite surface protein 3 and the 220-kilodalton glutamate-rich protein.** *Infect Immun* 2004, **72**:247-252.
- Lusingu JP, Vestergaard LS, Alifrangis M, Mmbando BP, Theisen M, Kitua AY, Lemnge MM, Theander TG: **Cytophilic antibodies to Plasmodium falciparum glutamate rich protein are associated with malaria protection in an area of holoendemic transmission.** *Malar J* 2005, **4**:48.
- Nebie I, Diarra A, Ouedraogo A, Soulama I, Bougouma EC, Tiono AB, Konate AT, Chilengi R, Theisen M, Dodoo D, Remarque E, Bosomprah S, Milligan P, Sirima SB: **Humoral responses to Plasmodium**

- falciparum blood-stage antigens and association with incidence of clinical malaria in children living in an area of seasonal malaria transmission in Burkina Faso, West Africa.** *Infect Immun* 2008, **76**:759-766.
11. Sirima SB, Nebie I, Ouedraogo A, Tiono AB, Konate AT, Gansane A, Derme AI, Diarra A, Ouedraogo A, Soulama I, Cuzzin-Ouattara N, Cousens S, Leroy O: **Safety and immunogenicity of the *Plasmodium falciparum* merozoite surface protein-3 long synthetic peptide (MSP3-LSP) malaria vaccine in healthy, semi-immune adult males in Burkina Faso, West Africa.** *Vaccine* 2007, **25**:2723-2732.
 12. Meraldi V, Nebie I, Tiono AB, Diallo D, Sanogo E, Theisen M, Druilhe P, Corradin G, Moret R, Sirima BS: **Natural antibody response to *Plasmodium falciparum* Exp-1, MSP-3 and GLURP long synthetic peptides and association with protection.** *Parasite Immunol* 2004, **26**:265-272.
 13. Hui G, Hashimoto C: ***Plasmodium falciparum* anti-MSP1-19 antibodies induced by MSP1-42 and MSP1-19 based vaccines differed in specificity and parasite growth inhibition in terms of recognition of conserved versus variant epitopes.** *Vaccine* 2007, **25**:948-956.
 14. Polley SD, Mwangi T, Kocken CH, Thomas AW, Dutta S, Lanar DE, Remarque E, Ross A, Williams TN, Mwangingu G, Lowe B, Conway DJ, Marsh K: **Human antibodies to recombinant protein constructs of *Plasmodium falciparum* Apical Membrane Antigen I (AMA1) and their associations with protection from malaria.** *Vaccine* 2004, **23**:718-728.
 15. Bouharoun-Tayoun H, Oeuvray C, Lunel F, Druilhe P: **Mechanisms underlying the monocyte-mediated antibody-dependent killing of *Plasmodium falciparum* asexual blood stages.** *J Exp Med* 1995, **182**:409-418.
 16. Hermsen CC, Verhage DF, Telgt DS, Teelen K, Bousema JT, Roestenberg M, Bolad A, Berzins K, Corradin G, Leroy O, Theisen M, Sauerwein RW: **Glutamate-rich protein (GLURP) induces antibodies that inhibit in vitro growth of *Plasmodium falciparum* in a phase I malaria vaccine trial.** *Vaccine* 2007, **25**:2930-2940.
 17. Miura K, Zhou H, Muratova OV, Orcutt AC, Giersing B, Miller LH, Long CA: **In immunization with *Plasmodium falciparum* apical membrane antigen 1, the specificity of antibodies depends on the species immunized.** *Infect Immun* 2007, **75**:5827-5836.
 18. Theisen M, Soe S, Oeuvray C, Thomas AW, Vuust J, Danielsen S, Jepsen S, Druilhe P: **The glutamate-rich protein (GLURP) of *Plasmodium falciparum* is a target for antibody-dependent monocyte-mediated inhibition of parasite growth in vitro.** *Infect Immun* 1998, **66**:11-17.
 19. Blackman MJ, Holder AA: **Secondary processing of the *Plasmodium falciparum* merozoite surface protein-1 (MSP1) by a calcium-dependent membrane-bound serine protease: shedding of MSP133 as a noncovalently associated complex with other fragments of the MSP1.** *Mol Biochem Parasitol* 1992, **50**:307-315.
 20. Borre MB, Dziegiel M, Høgh B, Petersen E, Rieneck K, Riley E, Meis JF, Aikawa M, Nakamura K, Harada M, etc: **Primary structure and localization of a conserved immunogenic *Plasmodium falciparum* glutamate rich protein (GLURP) expressed in both the preerythrocytic and erythrocytic stages of the vertebrate lifecycle.** *Mol Biochem Parasitol* 1991, **49**:119-132.
 21. Holder AA, Blackman MJ, Burghaus PA, Chappel JA, Ling IT, McCallum-Deighton N, Shai S: **A malaria merozoite surface protein (MSP1)-structure, processing and function.** *Mem Inst Oswaldo Cruz* 1992, **87 Suppl** 3:37-42.
 22. Lunel F, Druilhe P: **Effector cells involved in nonspecific and antibody-dependent mechanisms directed against *Plasmodium falciparum* blood stages in vitro.** *Infect Immun* 1989, **57**:2043-2049.
 23. Mitchell GH, Thomas AW, Margos G, Dluzewski AR, Bannister LH: **Apical membrane antigen 1, a major malaria vaccine candidate, mediates the close attachment of invasive merozoites to host red blood cells.** *Infect Immun* 2004, **72**:154-158.
 24. Narum DL, Thomas AW: **Differential localization of full-length and processed forms of PF83/AMA-1 an apical membrane antigen of *Plasmodium falciparum* merozoites.** *Mol Biochem Parasitol* 1994, **67**:59-68.
 25. O'Donnell RA, Saul A, Cowman AF, Crabb BS: **Functional conservation of the malaria vaccine antigen MSP-119 across distantly related *Plasmodium* species.** *Nat Med* 2000, **6**:91-95.
 26. Oeuvray C, Bouharoun-Tayoun H, Gras-Masse H, Bottius E, Kaidoh T, Aikawa M, Filgueira MC, Tartar A, Druilhe P: **Merozoite surface protein-3: a malaria protein inducing antibodies that promote *Plasmodium falciparum* killing by cooperation with blood monocytes.** *Blood* 1994, **84**:1594-1602.
 27. Ahorlu CK, Dunyo SK, Afari EA, Koram KA, Nkrumah FK: **Malaria-related beliefs and behaviour in southern Ghana: implications for treatment, prevention and control.** *Trop Med Int Health* 1997, **2**:488-499.
 28. Cavanagh DR, Dodoo D, Hviid L, Kurtzhals JA, Theander TG, Akanmori BD, Polley S, Conway DJ, Koram K, McBride JS: **Antibodies to the N-terminal block 2 of *Plasmodium falciparum* merozoite surface protein 1 are associated with protection against clinical malaria.** *Infect Immun* 2004, **72**:6492-6502.
 29. Dodoo D, Theisen M, Kurtzhals JA, Akanmori BD, Koram KA, Jepsen S, Nkrumah FK, Theander TG, Hviid L: **Naturally acquired antibodies to the glutamate-rich protein are associated with protection against *Plasmodium falciparum* malaria.** *J Infect Dis* 2000, **181**:1202-1205.
 30. Theisen M, Dodoo D, Toure-Balde A, Soe S, Corradin G, Koram KK, Kurtzhals JA, Hviid L, Theander T, Akanmori B, Ndiaye M, Druilhe P: **Selection of glutamate-rich protein long synthetic peptides for vaccine development: antigenicity and relationship with clinical protection and immunogenicity.** *Infect Immun* 2001, **69**:5223-5229.
 31. Kocken CH, Withers-Martinez C, Dubbeld MA, van der WA, Hackett F, Valderrama A, Blackman MJ, Thomas AW: **High-level expression of the malaria blood-stage vaccine candidate *Plasmodium falciparum* apical membrane antigen 1 and induction of antibodies that inhibit erythrocyte invasion.** *Infect Immun* 2002, **70**:4471-4476.
 32. Theisen M, Vuust J, Gottschau A, Jepsen S, Hogh B: **Antigenicity and immunogenicity of recombinant glutamate-rich protein of *Plasmodium falciparum* expressed in *Escherichia coli*.** *Clin Diagn Lab Immunol* 1995, **2**:30-34.
 33. Bonnet S, Petres S, Holm I, Fontaine T, Rosario S, Roth C, Longacre S: **Soluble and glyco-lipid modified baculovirus *Plasmodium falciparum* C-terminal merozoite surface protein 1, two forms of a leading malaria vaccine candidate.** *Vaccine* 2006, **24**:5997-6008.
 34. Audran R, Cachat M, Lurati F, Soe S, Leroy O, Corradin G, Druilhe P, Spertini F: **Phase I malaria vaccine trial with a long synthetic peptide derived from the merozoite surface protein 3 antigen.** *Infect Immun* 2005, **73**:8017-8026.
 35. Schellenberg JR, Smith T, Alonso PL, Hayes RJ: **What is clinical malaria? Finding case definitions for field research in highly endemic areas.** *Parasitol Today* 1994, **10**:439-442.
 36. Smith T, Schellenberg JA, Hayes R: **Attributable fraction estimates and case definitions for malaria in endemic areas.** *Stat Med* 1994, **13**:2345-2358.
 37. Cavanagh DR, Elhassan IM, Roper C, Robinson VJ, Giha H, Holder AA, Hviid L, Theander TG, Arnot DE, McBride JS: **A longitudinal study of type-specific antibody responses to *Plasmodium falciparum* merozoite surface protein-1 in an area of unstable malaria in Sudan.** *J Immunol* 1998, **161**:347-359.
 38. Oeuvray C, Theisen M, Rogier C, Trape JF, Jepsen S, Druilhe P: **Cytophilic immunoglobulin responses to *Plasmodium falciparum* glutamate-rich protein are correlated with protection against clinical malaria in Dielmo, Senegal.** *Infect Immun* 2000, **68**:2617-2620.
 39. Okech B, Mujuzi G, Ogwal A, Shirai H, Horii T, Egwang TG: **High titers of IgG antibodies against *Plasmodium falciparum* serine repeat antigen 5 (SERA5) are associated with protection against severe malaria in Ugandan children.** *Am J Trop Med Hyg* 2006, **74**:191-197.
 40. Okech BA, Corran PH, Todd J, Joynson-Hicks A, Uthaihibull C, Egwang TG, Holder AA, Riley EM: **Fine specificity of serum antibodies to *Plasmodium falciparum* merozoite surface protein, PfMSP-1 (19), predicts protection from malaria infection and high-density parasitemia.** *Infect Immun* 2004, **72**:1557-1567.
 41. Afari EA, Nakano T, Binka F, Owusu-Agyei S, Asigbee J: **Seasonal characteristics of malaria infection in under-five children of**

- a rural community in southern Ghana. *West Afr J Med* 1993, **12**:39-42.
42. Afari EA, Appawu M, Dunyo S, Baffoe-Wilmot A, Nkrumah FK: **Malaria infection, morbidity and transmission in two ecological zones Southern Ghana.** *Afr J Health Sci* 1995, **2**:312-315.
 43. Doodoo D, Staalsoe T, Giha H, Kurtzhals JA, Akanmori BD, Koram K, Dunyo S, Nkrumah FK, Hviid L, Theander TG: **Antibodies to variant antigens on the surfaces of infected erythrocytes are associated with protection from malaria in Ghanaian children.** *Infect Immun* 2001, **69**:3713-3718.
 44. Tongren JE, Drakeley CJ, McDonald SL, Reyburn HG, Manjurano A, Nkya WM, Lemnge MM, Gowda CD, Todd JE, Corran PH, Riley EM: **Target antigen, age, and duration of antigen exposure independently regulate immunoglobulin G subclass switching in malaria.** *Infect Immun* 2006, **74**:257-264.
 45. Baird JK: **Host age as a determinant of naturally acquired immunity to Plasmodium falciparum.** *Parasitol Today* 1995, **11**:105-111.
 46. Boudin C, Chumpitazi B, Dziegiel M, Peyron F, Picot S, Hogh B, Ambruisse-Thomas P: **Possible role of specific immunoglobulin M antibodies to Plasmodium falciparum antigens in immunoprotection of humans living in a hyperendemic area, Burkina Faso.** *J Clin Microbiol* 1993, **31**:636-641.
 47. Branch OH, Udhayakumar V, Hightower AW, Oloo AJ, Hawley WA, Nahlen BL, Bloland PB, Kaslow DC, Lal AA: **A longitudinal investigation of IgG and IgM antibody responses to the merozoite surface protein-1 19-kiloDalton domain of Plasmodium falciparum in pregnant women and infants: associations with febrile illness, parasitemia, and anemia.** *Am J Trop Med Hyg* 1998, **58**:211-219.
 48. Branch OH, Oloo AJ, Nahlen BL, Kaslow D, Lal AA: **Anti-merozoite surface protein-1 19-kDa IgG in mother-infant pairs naturally exposed to Plasmodium falciparum: subclass analysis with age, exposure to asexual parasitemia, and protection against malaria. V. The Asembo Bay Cohort Project.** *J Infect Dis* 2000, **181**:1746-1752.
 49. Morais CG, Soares IS, Carvalho LH, Fontes CJ, Krettli AU, Braga EM: **IgG isotype to C-terminal 19 kDa of Plasmodium vivax merozoite surface protein I among subjects with different levels of exposure to malaria in Brazil.** *Parasitol Res* 2005, **95**:420-426.
 50. Bouharoun-Tayoun H, Druilhe P: **Plasmodium falciparum malaria: evidence for an isotype imbalance which may be responsible for delayed acquisition of protective immunity.** *Infect Immun* 1992, **60**:1473-1481.
 51. Bouharoun-Tayoun H, Druilhe P: **Antibodies in falciparum malaria: what matters most, quantity or quality?** *Mem Inst Oswaldo Cruz* 1992, **87 Suppl 3**:229-234.
 52. Nasr A, Iriemenam NC, Troye-Blomberg M, Giha HA, Balogun HA, Osman OF, Montgomery SM, ElGhazali G, Berzins K: **Fc gamma receptor IIa (CD32) polymorphism and antibody responses to asexual blood-stage antigens of Plasmodium falciparum malaria in Sudanese patients.** *Scand J Immunol* 2007, **66**:87-96.
 53. Braga EM, Scopel KKG, Komatsu NT, daSilva-Nunes M, Ferreira MU: **Polymorphism of the Fc gamma receptor IIA and malaria morbidity.** *Journal of Molecular and Genetic Medicine* 2005, **1**:5-10.
 54. Aubouy A, Migot-Nabias F, Deloron P: **Correlations between treatment outcome and both anti-MSP119 antibody response and erythrocyte-related genetic factors in Plasmodium falciparum malaria.** *Infect Genet Evol* 2007, **7**:147-154.
 55. Corran PH, O'Donnell RA, Todd J, Uthaiyibull C, Holder AA, Crabb BS, Riley EM: **The fine specificity, but not the invasion inhibitory activity, of 19-kilodalton merozoite surface protein I-specific antibodies is associated with resistance to malarial parasitemia in a cross-sectional survey in The Gambia.** *Infect Immun* 2004, **72**:6185-6189.
 56. Uthaiyibull C, Aufiero B, Syed SE, Hansen B, Guevara Patino JA, Angov E, Ling IT, Fegeding K, Morgan WD, Ockenhouse C, Birdsall B, Feeney J, Lyon JA, Holder AA: **Inhibitory and blocking monoclonal antibody epitopes on merozoite surface protein I of the malaria parasite Plasmodium falciparum.** *J Mol Biol* 2001, **307**:1381-1394.

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