

RESEARCH ARTICLE

Open Access

Antibacterial constituents of three Cameroonian medicinal plants: *Garcinia nobilis*, *Oricia suaveolens* and *Balsamocitrus camerunensis*

Hugues Fouotsa^{1,4}, Armelle T Mbaveng^{2,3}, Celine D Mbazona¹, Augustin E Nkengfack¹, Shaheen Farzana⁴, Choudhary M Iqbal⁴, Jacobus J Marion Meyer⁵, Namrita Lall⁵ and Victor Kuete^{2,5*}

Abstract

Background: Multidrug resistance is a worrying cause of treatment failure in bacterial infections. The search of bioactive constituents from medicinal plants against multidrug resistant (MDR) bacteria has significantly evolved in the two last decades. In the present study, twenty-two compounds (three terpenoids, eleven phenolics and eight alkaloids) isolated from three Cameroonian medicinal plants, namely *Garcinia nobilis*, *Oricia suaveolens* and *Balsamocitrus camerunensis*, as well as the crude extracts were tested for their antibacterial activities against *Mycobacterium tuberculosis* and Gram-negative bacteria amongst which were MDR active efflux pumps expressing phenotypes.

Methods: The microplate alamar blue assay (MABA) and the broth microdilution methods were used to determine the minimal inhibitory concentrations (MIC) and minimal bactericidal concentrations (MBC) of the studied samples.

Results: The results of the MIC determinations indicate that, the best crude extract was that from *G. nobilis* (GNB), its inhibitory effects being noted against 12 of the 14 tested bacteria. The extract of GNB also exhibited better anti-tuberculosis (MIC of 128 µg/ml *M. tuberculosis* against ATCC 27294 strain) and antibacterial (MIC of 64 µg/ml against *Escherichia coli* ATCC10536) activities compared to the extracts of *O. suaveolens* and *B. camerunensis*. Interestingly, 4-prenyl-2-(3,7-dimethyl-2,6-octadienyl)-1,3,5,8-tetrahydroxyxanthone (**2**), isolated from the most active extract GNB, also showed the best activity amongst compounds, inhibiting the growth of all the fourteen tested microorganisms. The lowest MIC value obtained with compound **2** was 8 µg/ml against *M. tuberculosis* ATCC 27294 and *M. tuberculosis* clinical MTCS2 strains. Other compounds showed selective activities with 11 of the 14 tested bacteria being sensitive to the xanthone, morusignin I (**5**) and the alkaloid, kokusaginine (**13**).

Conclusions: The results of the present investigation provide evidence that the crude extract from *G. nobilis*, *O. suaveolens* and *B. camerunensis* as well as some of their compounds, and mostly compound **2** (isolated from *G. nobilis*), could be considered as interesting natural antibacterial products.

Keywords: Antimycobacterial, Antibacterial, Cameroon, Compounds, *Garcinia nobilis*, *Oricia suaveolens*, *Balsamocitrus camerunensis*

* Correspondence: kuetevictor@yahoo.fr

²Department of Biochemistry, Faculty of Science, University of Dschang, Dschang, Cameroon

⁵Department of Plant Science, Faculty of Agricultural and Biological Science, Pretoria 0002, South Africa

Full list of author information is available at the end of the article

Background

The continuous emergence of multidrug-resistant (MDR) bacteria drastically reduced the efficacy of our antibiotic armory and, consequently, increases the frequency of therapeutic failure. Drug resistance is a consequence of the worldwide use of antibiotics, and the acute challenge for health care is to find measures to efficiently combat resistant organisms [1]. Several natural compounds have successfully been tested for their abilities to prevent the growth of MDR bacteria [2]. In our continuous search of antibacterial drugs from natural source, we targeted three Cameroonian medicinal plants, *Garcinia nobilis* Engl. (*Clusiaceae*), *Oricia suaveolens* Engl. [Commonly known in West Africa as Abe iolo or Kru-bete parihi (Ivory Coast) or Mende jagbole (Sierra Leone)] and *Balsamocitrus camerunensis* Letouzey, R. (*Rutaceae*). Plants of the genus *Garcinia*, widely distributed in the tropical Africa, Asia, New Caledonia, and Polynesia, have yielded many biologically active and structurally intriguing natural products [3]. *Garcinia* species are known to contain a wide variety of oxygenated and prenylated xanthenes, as well as polyisoprenylated benzophenones such as the guttiferones [4]. Previous studies of the chemistry of the genus *Oricia*, including *O. suaveolens* revealed the presence of alkaloids and triterpenes [5,6]. *B. camerunensis* is a new species recently found in Batouri (Cameroon) and Boukoko (Central African Republic) [7]. Plants of genus *Balsamocitrus* (including the decoction of the bark of *B. camerunensis*) are used in traditional African medicine to treat malaria, hypertension, infertility, and influenza [8,9]. Previous phytochemical investigations of this genus revealed the presence of coumarins, quinoline alkaloids, free aliphatic acids and steroids, some of these compounds exhibiting potent antibacterial, fungicidal, and algicidal properties [8]. The aqueous decoctions of the bark of *G. nobilis* as well as that of the roots of *O. suaveolens* are used in Cameroon to treat gastro-intestinal infections [Personal communication]. The combination of the three plants (*G. nobilis*, *O. suaveolens* and *B. camerunensis*) is also used locally to treat stomachache and diarrheal infections [Personal communication]. The present study was therefore designed to evaluate the antibacterial activities of the naturally occurring compounds from *G. nobilis*, *O. suaveolens* and *B. camerunensis* with emphasis on MDR Gram-negative bacteria and *Mycobacterium tuberculosis*.

Methods

General experimental procedure

^1H and ^{13}C NMR spectra were recorded in chloroform on Digital NMR BRUCKER AVANCE 400 and 500 MHz. EIMS were obtained on Joel the MS route JMS.600H and HREIMS were performed on a thermo finnigan Mat 95 XP. Thin layer chromatography (TLC) and pre-coated TLC were performed on silica gel GF₂₅₄ (Merck). Column

chromatography (CC) was performed on silica gel (Merck) type 100(70–230 Mesh ASTM) eluted either with gradient system (Hex-Ethyl Acetate-MeOH; Hex-CH₂Cl₂-MeOH and Hex-CH₂Cl₂-Acetone). All the solvents used were distilled commercial grade. The isolated compounds were crystallized from the same solvent and their purity was checked by TLC. Pre-coated plates of silica gel GF₂₅₄ (Merck) were used for this purpose; the spots were detected with UV lamp at 254 and 366 nm and by spraying with 50% H₂SO₄ or ceric sulfate following by heating.

Plant material

The stem bark of the *G. nobilis* was collected in Okola-Yaounde (Center Region of Cameroon) in April, 2010 whilst *O. suaveolens* (woods and stem bark), and *B. camerunensis* (stem bark) were collected in Nkobi village (Batouri, East region of Cameroon), in August 2007. The plants were identified by Mr. Victor Nana of the Cameroon National Herbarium (Yaoundé) where voucher specimen (50779/HNC/Cam for *G. nobilis*, 6161/SRF/Cam for *O. suaveolens* and 3785/SRFK for *B. camerunensis*) were deposited.

Extraction

The air-dried and powdered samples from each plant were macerated in either 7.0 L methanol (MeOH) for the stem bark of *G. nobilis* (2 kg) or in 10 L MeOH/dichloromethane (CH₂Cl₂) mixture for the wood (4 kg) and roots (3 kg) of *O. suaveolens*, and the stem bark (3.8 kg) of *B. camerunensis*. The extraction was done at room temperature for two days. The evaporation under reduced pressure yielded the crude extracts from the stem bark of *G. nobilis* (GNB; 100 g), wood (OSW; 173 g) and roots (OSR; 145 g) from *O. suaveolens*, and from the stem bark *B. camerunensis* (BCB; 128 g).

Isolation and identification of compounds from *garcinia nobilis*

The compounds from GNB tested herein, caroxanthone (1), 3-dimethyl-2-geranyl-4-prenylbellidifolin (2), smeath xanthone A (3), 8-hydroxycudraxanthone G (4) and morusignin I (5) (Figure 1) were obtained directly from our chemical bank. We previously reported their isolation and identification from GNB [10].

Isolation and identification of compounds from *oricia suaveolens*

One hundred grams of OSW were submitted to vacuum liquid chromatography (VLC; 25–40 μm , 8 cm \times 60 cm, 400 g), eluting with *n*-hexane-Ethyl acetate (EtOAc) of increasing polarity. Fractions of 500 mL each were collected and subsequently pooled on the basis of their thin layer chromatography (TLC) profile into five main fractions

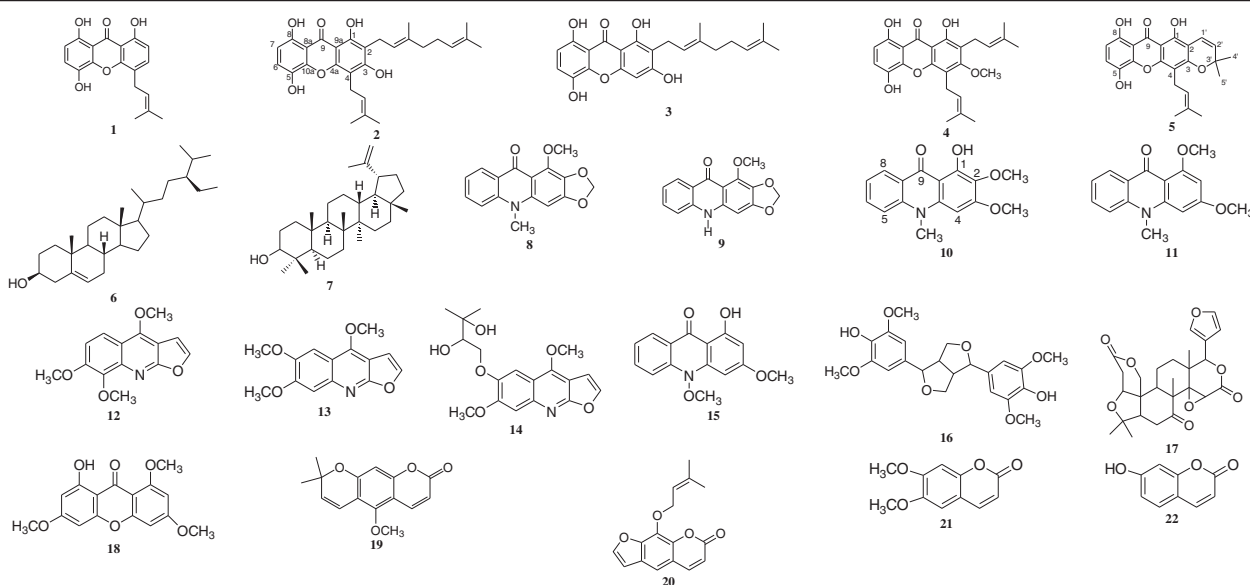


Figure 1 Chemical structures of compounds isolated from the stem bark of *Garcinia nobilis* (1-5), *Oricia suaveolens* woods (6-11) and roots (12-17), and the stem back of *Balsmocitrus camerunensis* (18-22). Caroxanthone (1), 4-prenyl-2-(3,7-dimethyl-2,6-octadienyl)-1,3,5,8-tetrahydroxanthone (2), smeathxanthone A (3), 8-hydroxycudraxanthone G (4), morusignin I (5), stigma-5-en-3-ol (6), lupol (7), evoxanthine (8), norevioxanthine (9), 1-hydroxy-2,3-dimethoxy-10-methylacridone (10), 1,3-dimethoxy-10-methylacridone (11), skimmianine (12), kokusaginine (13), monrifoline (14), 1-hydroxy-4-methoxy-10-methylacridone (15), syringaresinol (16), limonin (17), 1-hydroxy-3,6-dimethoxy-8-methylxanthone (18), xanthoxyletin (19), imperatorin (20), 6,8-dimethoxycoumarin (21), umbelliferone (22).

[*n*-hexane- EtOAc 8:2 (2 L, 15 g, fraction 1); 6:4 (2 L, 30 g, fraction 2); 4:6 (1.5 L, 12 g, fraction 3); 2:8 (1.5 L, 15 g, fraction 4); 0:10 (1 L, 10.5 g, fraction 5)]. Fraction 2 (30 g) was then subjected to silica gel 60 column chromatography (25–40 μ m, 6 cm \times 60 cm, 100 g), eluting with *n*-hexane-EtOAc mixture of increasing polarity. Sub-fractions (sub-fr) of 100 mL each were collected, then pooled into nine main sub-frs obtained as follows: *n*-hexane- EtOAc 10:0 (1 L, sub-fr A); 9:1 (1.5 L, sub-fr B); 8:2 (1.5 L, sub-fr C); 7.5:2.5 (1 L, sub-fr D); 7:3 (750 mL, sub-fr E); 6:4 (900 mL, sub-fr F); 5:5 (500 mL, sub-fr G); 3:7 (500 mL, sub-fr H) and pure EtOAc (1 L, sub-fr I). Stigma-5-en-3-ol C₂₉H₅₀O (6; 88.3 mg; *m/z*: 414; mp: 250°C) [11] and lupeol C₃₀H₅₀O (7; 57.1 mg; *m/z*: 426; mp: 215–216°C) [12] were obtained from sub-frs A (4.2 g) and B (6.0 g) respectively by recrystallization. Sub-frs A and B residues were then combined based on their similar TLC profile to obtain a new sub-fr named AB (7.5 g). The sub-fr AB was purified on silica gel column (25–40 μ m, 4 cm \times 60 cm, 80 g) using *n*-hexane-EtOAc with increasing polarity. Based on their TLC profiles, 15 new sub-frs of 100 mL each were obtained; evoxanthine C₁₆H₁₃NO₄ (8; 30.0 mg; *m/z*: 283; mp:218–218°C) [13] was obtained by recrystallisation from new sub-frs 3–6 (*n*-hexane-EtOAc 8:2; 7.5:2.5; 7:3; 6.5:3.5) whilst sub-frs 9–12 (*n*-hexane- EtOAc: 5:5; 4.5:5.5; 4:6; 3:7) yielded evoxanthidine C₁₅H₁₁NO₄ (9; 18.1 mg; *m/z*: 269; mp:310–312°C) by recrystallisation [14].

Sub-frs C-D (7 g) were combined on the basis of their TLC profiles, then subjected to column chromatography over silica gel (25–40 μ m 3 cm \times 6 cm, 70 g) and eluted with the increasing polarity *n*-hexane- EtOAc to afford ten other sub-frs. Sub-frs 2–4 (obtained with *n*-hexane-EtOAc 7:3; 6:4; 5:5, 2.6 g) yielded 1-hydroxy-2,3-dimethoxy-10-methylacridone or arborinine C₁₆H₁₅NO₄ (10; 12.1 mg; *m/z*: 285; mp:175–176°C) [15] and subsequent sub-frs 6–8 (obtained *n*-hexane- EtOAc 3:7; 2.5:7.5; 2:8) yielded 1,3-dimethoxy-10-methylacridone C₁₆H₁₃NO₃ (11; 8.0 mg; *m/z*:269; mp:165°C) [14].

One hundred grams of OSR were partitioned in the water to obtain sequentially hexane fraction (20.1 g), CH₂Cl₂ fraction (24.7 g), acetone fraction (17.0 g) and the MeOH fraction (32.5 g). The CH₂Cl₂ crude extract (24.7 g) was then submitted to VLC on silica gel (25–40 μ m, 8 cm \times 60 cm, 100 g) and eluted with hexane-ethyl acetate gradients: (8:2, 1.5 L), (6:4, 1.5 L), (5:5, 1.25 L), (4:6, 1.25 L), (2:8, 1 L) and finally with pure EtOAc (500 mL) to give 40 fractions of 250 ml each. These fractions were pooled on the basis of their TLC profiles. Fractions 1–17 (10 g), obtained with 80% hexane–EtOAc were subjected to column chromatography over silica gel (25–40 μ m, 4 cm \times 60 cm, 65 g) eluting with hexane-CH₂Cl₂ with a continuous gradient (95:5 to 7:3) to yield skimmianine C₁₄H₁₃NO₄ (12; 2.1 mg; *m/z*: 259; mp:176–177°C) [16] and kokusaginine C₁₄H₁₃NO₄ (13; 7.5 mg; *m/z*: 259; mp:171°C) [16]. A continuous

elution by increasing the solvent polarity (6:4 to pure CH₂Cl₂) yielded four mixtures sub-frs (A-D). Sub-fr B (2.7 g), showed after examination on TLC precoated plate a mixture of two compounds. This fraction was further purified on silica gel column chromatography (25–40 μm; 3 × 15 cm, 15 g) eluting with hexane-CH₂Cl₂ with a continuous gradient (6:4 to 3:7) to yield monrifoline C₁₈H₂₁NO₆ (**14**; 2.7 mg; *m/z*: 347; mp:1190-192°C) [17] and 1-hydroxy-3-methoxy-10-methylacridone C₁₅H₁₃NO₃ (**15**; 15.0 mg; *m/z*: 255; mp:164–165; 174-176°C) [18]. Sub-fr C (2.55 g) was then subjected to silica gel column chromatography eluting with hexane-CH₂Cl₂ with a continuous gradient (6:4 to pure CH₂Cl₂) to yield (+/-) syringaresinol C₂₂H₂₆O₈ (**16**; 9.2 mg; *m/z*: 418; mp:185-186°C; mp:1175-176°C) [19] whilst combined sub-frs 21–40 (25 g) obtained from the (1:1) to (1:3) hexane-EtOAc mixtures and pure EtOAc were subjected successively to silica gel column chromatography and preparative TLC, eluting with solvent mixtures (hexane-CH₂Cl₂-EtOAc) of increasing polarity to yield limonin C₂₆H₃₀O₈ (**17**; 12.0 mg; *m/z*: 470; mp:298°C) [20].

Isolation and identification of compounds from *balsamocitrus camerunensis*

One hundred and twenty grams of BCB were partitioned by dissolving in CH₂Cl₂ to give a soluble (42.0 g) and insoluble (76.5 g) fractions. The CH₂Cl₂ soluble fraction was subjected to silica gel VLC (25–40 μm, 8 cm × 60 cm, 120 g) and eluted with the increasing polarity of *n*-hexane-EtOAc and MeOH to afford 160 sub-frs of 250 mL each. These sub-frs were pooled on the basis of TLC analysis into four main fractions (A-D).

Fraction A (12.5 g, combined from sub-frs 1–40) was column chromatographed over silica gel (25–40 μm, 3.5 - cm × 50.0 cm) with *n*-hexane-CH₂Cl₂ gradient. A total of 40 new sub-fractions of 100 mL each were collected and pooled on the basis of the TLC analysis. Sub-fr 5–15 were further column chromatographed over silica gel (25–40 μm, 2.5 cm × 30.0 cm), using *n*-hexane-CH₂Cl₂ (4:1) mixture as eluent to afford compound **7** (99 mg). Sub-frs 16–40 were further chromatographed over silica gel column (25–40 μm, 2.5 cm × 30.0 cm), eluting with *n*-hexane-CH₂Cl₂ (1:1) mixture to yield 1-hydroxy-3,6-dimethoxy-8-methylxanthone or lichexanthone C₁₇H₁₆O₄ (**18**; 5.2 mg; *m/z*: 284; mp:187-188°C) [21] and xanthoxyletin C₁₅H₁₄O₄ (**19**; 7.0 mg; *m/z*: 258; mp:133°C) [22]. Fraction B (10.0 g, combined from the sub-frs 41–79) was purified over silica gel column (25–40 μm, 3.5 × 50.0 cm) with *n*-hexane-CH₂Cl₂ gradient. A total of 50 sub-frs of 100 mL each were collected and pooled on the basis of TLC analysis. Sub-frs 10 to 40 were further purified over silica gel column (25–40 μm, 2.5 cm × 30.0 cm) with a mixture of *n*-hexane-CH₂Cl₂ (9 : 1) to yield imperatorin C₁₆H₁₄O₄ (**20**; 12.0 mg, *m/z*: 270; mp:102°C

[23]. Fraction C (16.5 g, combined from sub-frs 80–110) was purified over silica gel column (25-40 μm, 3.5 cm × 50.0 cm) with a Hexane- CH₂Cl₂ gradient. A total of 45 fractions of 100 mL each were collected and combined on the basis of their TLC profile. Fractions 10–25 were further purified over silica gel column (25–40 μm, 2.5 cm × 30.0 cm) with CH₂Cl₂-acetone (4: 1) mixture to afford scoparone C₁₁H₁₀O₄ (**21**; 8.7 mg; *m/z*: 206; mp: 144°C) [24]. Fraction D was further chromatographed on silica gel column, using CH₂Cl₂-acetone mixture as eluent, then the recrystallization in CH₂Cl₂-acetone (6:4) gave umbelliferone C₉H₆O₃ (**22**; 11.0 mg; *m/z*: 162; mp:230-232°C) [25].

Chemicals for antibacterial assay

Chloramphenicol (Sigma-Aldrich, St. Quentin Fallavier, France) was used as reference antibiotics (RA) against Gram-negative bacteria. *p*-Iodonitrotetrazolium chloride (INT, Sigma-Aldrich) was used as microbial growth indicator [26,27]. Isoniazid (INH) (Sigma) was used as RA against *M. tuberculosis*.

Antibacterial assays

Microbial strains and culture media

The studied microorganisms included reference and clinical strains (Tables 1 and 2) of *Escherichia coli*, *Enterobacter aerogenes*, *Klebsiella pneumoniae*, *Providencia stuartii*, *Pseudomonas aeruginosa*, a drug-susceptible strain of *M. tuberculosis* H37Rv obtained from the American Type Culture Collection, and two clinical strains of *M. tuberculosis* MTCS1, MTCS2. *M. tuberculosis* was plated on Löwenstein-Jensen medium and allowed to grow for 3–4 weeks at 37°C. Middlebrook 7H9 broth supplemented with 0.2% glycerol and 10% Oleic Acid–Albumin–Dextrose–Catalase (OADC) (Sigma) was used to determine the MIC and MBC values of the test samples on *M. tuberculosis*. We previously reported the features [28] of all the tested Gram-negative bacteria. Nutrient agar was used for the activation of bacteria other than *M. tuberculosis* strains [28]. The clinical strains used in this work are our laboratory collection previously obtained from Yaoundé General Hospital (Cameroon), and from the Mediterranean University (Marseille, France).

INT colorimetric assay for MIC and MBC determinations

The MIC determinations on Gram-negative bacteria were conducted using rapid INT colorimetric assay according to previously described methods [26,27] with some modifications. The test samples and RA were first of all dissolved in DMSO/Mueller Hinton Broth (MHB) or DMSO/7H9 broth. The final concentration of DMSO was lower than 2.5% and does not affect the microbial growth [29]. The solution obtained was then added to MHB, and serially diluted two fold (in a 96- wells microplate). Then, 100 μl of inoculum 1.5 × 10⁶ CFU/ml

Table 1 MICs of the extracts, compounds from *Garcinia nobilis*, *Orcia suaveolens*, *Balsmocitrus camerunensis* and reference drugs on documented strains and clinical MDR isolates

Tested samples ^a	Microorganisms, strains and MIC (µg/ml) ^b														
	<i>E. coli</i>			<i>E. aerogenes</i>			<i>K. pneumoniae</i>		<i>P. stuartii</i>		<i>P. aeruginosa</i>		<i>M. tuberculosis</i>		
	ATCC 10536	AG 100	AG 102	ATCC 13048	CM 64	EA 27	ATCC 11296	KP55	ATCC 29916	NAE 16	PA 01	PA 124	ATCC 27294	MTCS1	MTCS2
Crude extracts															
GNB	64	256	256	256	512	256	128	512	512	-	512	-	128	256	256
OSR	128	512	-	512	-	-	256	256	512	256	-	-	512	-	512
OSW	128	512	-	-	-	-	128	128	-	128	-	-	256	-	512
BCB	512	-	-	256	-	-	512	512	-	-	-	-	-	-	-
Compounds															
1	256	128	-	-	-	-	-	-	-	-	-	-	-	-	-
2	64	64	128	128	256	64	512	512	256	64	64	128	8	32	8
3	128	128	64	256	-	256	-	-	-	-	-	-	64	-	128
4	-	64	128	-	256	128	-	-	-	-	256	-	512	-	512
5	64	128	128	-	-	512	512	-	512	-	64	128	32	64	64
8	512	128	256	-	512	128	512	-	-	-	-	-	128	512	256
9	-	128	256	-	256	256	-	-	-	-	-	-	-	-	-
10	64	128	256	-	-	128	-	-	512	-	-	-	128	-	512
11	128	256	128	-	256	512	512	-	-	-	-	512	-	-	-
13	512	64	256	-	512	128	256	-	-	-	128	512	256	512	512
15	128	256	256	-	-	256	-	-	-	-	-	-	-	-	-
16	128	128	-	-	-	512	-	-	512	-	-	-	512	-	-
17	-	128	-	-	-	-	-	-	-	-	-	-	-	-	-
18	-	256	-	-	-	256	-	-	-	256	-	-	-	-	-
19	-	256	-	-	-	-	-	-	-	-	-	-	-	-	-
20	-	512	-	-	-	-	-	-	-	-	-	-	-	-	-
21	512	256	-	-	-	256	256	-	-	-	-	-	-	-	-
22	512	256	256	512	128	-	-	512	-	-	-	-	-	-	-
RA	<4	16	512	64	-	64	4	32	-	64	128	128	1	128	4

^aTested samples [methanol-dichloromethane 1:1 extract of the stem bark of *Garcinia nobilis* (GNB), the wood (OSW)/roots (OSR) of *Orcia suaveolens*, and stem bark of *Balsmocitrus camerunensis* (BCB); compounds isolated from the stem bark of *Garcinia nobilis* (1–5), *Orcia suaveolens* woods (6–11) and roots (12–17), and the stem bark of *Balsmocitrus camerunensis* (18–22); caroxanthone (1), 4-prenyl-2-(3,7-dimethyl-2,6-octadienyl)-1,3,5,8-tetrahydroxanthone (2), smeathxanthone A (3), 8-hydroxycudraxanthone G (4), morusignin I (5), stigma-5-en-3-ol (6), 20(29)lupen-2-ol (7), evoxanthine (8), norevioxanthine (9), 1-hydroxy-2,3-dimethoxy-10-methylacridone (10), 1,3-dimethoxy-10-methylacridone (11), kokusaginine (13), 1-hydroxy-4-methoxy-10-methylacridone (15), syringaresinol (16), limonin (17), 1-hydroxy-3,6-dimethoxy-8-methylxanthone (18), xanthoxyletin (19), imperatorin (20), 6,8-dimethoxycoumarin (21), umbelliferone (22)]; RA: reference antibiotics were chloramphenicol for bacteria, isoniazid for *M. tuberculosis*; ^b(-): MIC > 512 µg/mL.

Table 2 MBCs of the extracts, compounds from *Garcinia nobilis*, *Orcia suaveolens*, *Balsmocitrus camerunensis* and reference drugs on documented strains and clinical MDR isolates

Tested samples ^a	Microorganisms, strains and MBC (µg/ml) ^b														
	<i>E. coli</i>			<i>E. aerogenes</i>			<i>K. pneumoniae</i>		<i>P. stuartii</i>		<i>P. aeruginosa</i>		<i>M. tuberculosis</i>		
	ATCC 10536	AG 100	AG 102	ATCC 13048	CM 64	EA 27	ATCC 11296	KP55	ATCC 29916	NAE 16	PA 01	PA 124	ATCC 27294	MTCS1	MTCS2
Crude extracts															
GNB	256	512	>512	>512	>512	>512	256	>512	>512	-	>512	-	256	>512	512
OSR	256	>512	-	>512	-	-	512	512	>512	512	-	-	>512	-	>512
OSW	512	>512	-	-	-	-	256	512	-	512	-	-	512	-	>512
BCB	>512	-	-	512	-	-	>512	>512	-	-	-	-	-	-	-
Compounds															
1	>512	>512	-	-	-	-	-	-	-	-	-	-	-	-	-
2	128	256	256	256	>512	128	>512	>512	512	128	128	256	16	64	32
3	256	256	128	512	-	512	-	-	-	-	-	-	128	-	256
4	-	128	256	-	512	256	-	-	-	-	512	-	>512	-	>512
5	128	256	256	-	-	>512	>512	-	>512	-	128	256	64	128	128
8	>512	256	>512	-	>512	256	>512	-	-	-	-	-	256	>512	512
9	-	256	>512	-	512	>512	256	-	-	-	-	-	-	-	-
10	256	256	512	-	-	-	-	-	>512	-	-	-	512	-	>512
11	>512	-	-	-	-	>512	-	-	-	-	-	-	-	-	-
13	>512	128	512	-	>512	256	512	-	-	-	256	>512	512	>512	>512
15	256	512	>512	-	-	512	-	-	-	-	-	-	-	-	-
16	256	256	-	-	-	>512	-	-	>512	-	-	-	>512	-	-
17	-	256	-	-	-	-	-	-	-	-	-	-	-	-	-
18	-	512	-	-	-	512	-	-	-	512	-	-	-	-	-
19	-	512	-	-	-	-	-	-	-	-	-	-	-	-	-
20	-	>512	-	-	-	-	-	-	-	-	-	-	-	-	-
21	>512	512	-	-	-	512	512	-	-	-	-	-	-	-	-
22	>512	512	512	>512	256	-	-	>512	-	-	-	-	-	-	-
RA	<4	32	>512	128	-	128	8	64	-	128	256	256	2	256	16

^aTested samples [methanol-dichloromethane 1:1 extract of the stem bark of *Garcinia nobilis* (GNB), the wood (OSW)/roots (OSR) of *Orcia suaveolens*, and stem bark of *Balsmocitrus camerunensis* (BCB); compounds isolated from the stem bark of *Garcinia nobilis* (**1–5**), *Orcia suaveolens* woods (**6–11**) and roots (**12–17**), and the stem bark of *Balsmocitrus camerunensis* (**18–22**); caroxanthone (**1**), 4-prenyl-2-(3,7-dimethyl-2,6-octadienyl)-1,3,5,8-tetrahydroxanthone (**2**), smeathxanthone A (**3**), 8-hydroxycudraxanthone G (**4**), morusignin I (**5**), stigma-5-en-3-ol (**6**), 20(29)lupen-2-ol (**7**), evoxanthine (**8**), norevioxanthine (**9**), 1-hydroxy-2,3-dimethoxy-10-methylacridone (**10**), 1,3-dimethoxy-10-methylacridone (**11**), kokusaginine (**13**), 1-hydroxy-4-methoxy-10-methylacridone (**15**), syringaresinol (**16**), limonin (**17**), 1-hydroxy-3,6-dimethoxy-8-methylxanthone (**18**), xanthoxyletin (**19**), imperatorin (**20**), 6,8-dimethoxycoumarin (**21**), umbelliferone (**22**)]; RA: reference antibiotics were chloramphenicol for bacteria, isoniazid for *M. tuberculosis*; ^b(–): MBC not determined as the MIC was above 512 µg/ml.

prepared in appropriate broth was added [30]. The plates were covered with a sterile plate sealer, then agitated to mix the contents of the wells using a plate shaker and incubated at 37°C for 18 h. The assay was repeated thrice. Wells containing adequate broth, 100 µl of inoculum and DMSO to a final concentration of 2.5% served as negative control. The MIC of samples was detected after 18 h incubation at 37°C, following addition (40 µl) of 0.2 mg/ml INT and incubation at 37°C for 30 minutes. Viable bacteria reduced the yellow dye to a pink. MIC was defined as the sample concentration that prevented this change and exhibited complete inhibition of microbial growth. The MBC was determined by adding 50 µl aliquots of the preparations, which did not show any growth after incubation during MIC assays, to 150 µl of adequate broth. These preparations were incubated at 37°C for 48 h. The MBC was regarded as the lowest concentration of extract, which did not produce a color change after addition of INT as mentioned above [30,31].

Microplate Alamar blue assay (MABA) against *M. tuberculosis*

The activity of all samples against *M. tuberculosis* strains was tested using the MABA [32]. Briefly, each of the above *M. tuberculosis* strains was cultured at 37°C in Middlebrook 7H9 broth supplemented with 0.2% glycerol and 10% OADC (oleic acid–albumin–dextrose–catalase; Sigma) until logarithmic growth was reached. The homogenous culture was obtained using sterile glass beads and vortex. About 6×10^6 CFU/ml inoculum (100 µl) of *M. tuberculosis* was then added to the two fold serially diluted samples (100 µl). The final concentration of DMSO in all assays was 2.5% or less and this dilution also served as solvent control. The samples were assayed in triplicate. All tests were carried out in sterile flat-bottomed 96-well microplates. Each microplate was incubated for 5 days at 37°C in a sealed plastic CO₂-permeable bag. After 5 days of incubation, 32 µl of a mixture of freshly prepared Alamar Blue solution and 20% sterile Tween-80 (Sigma) 1:1 v/v were added to one growth-control well. The microplates were incubated again at 37°C for 24 h. If a color shift from blue to pink was observed in the growth-control sample, 32 µl of alamar blue solution was added to each of the remaining wells, and the microplate was further incubated for 24 h. A well-defined pink color was interpreted as positive bacterial growth, whereas a blue color indicated an absence of growth. The MIC corresponded to the greatest dilution of sample extract in which the color shift from blue to pink was not observed.

Samples with recorded MIC values following MABA were assayed for their mycobactericidal effect [32]. Briefly, 5 µl of the undeveloped mycobacterial suspensions were

transferred from the former to a new microplate containing 195 µl of fresh culture medium per well. Three wells were inoculated with 100 µl of fresh inoculum as for MABA and three more wells were incubated with 200 µl of culture medium only, as negative controls. The microplates were incubated and developed with alamar blue as for MABA. The MBC corresponded to the minimum sample concentration that did not cause a color shift in cultures that were re-incubated in fresh medium.

Results and discussion

The chemical structures of the compounds isolated from *G. nobilis*, *O. suaveolens* and *B. camerunensis* (Figure 1) were established by spectroscopic methods. The compounds were isolated from the stem bark of *G. nobilis* (1–5), the woods of *O. suaveolens* (6–11), the roots (12–17) and the stem bark of *B. camerunensis* (7, 18–22). The twenty two isolated compounds were identified as caroxanthone (1), 4-prenyl-2-(3,7-dimethyl-2,6-octadienyl)-1,3,5,8-tetrahydroxyxanthone (2), smeathxanthone A (3), 8-hydroxycudraxanthone G (4), morusignin I (5), stigma-5-en-3-ol (6), lupeol (7), evoxanthine (8), norevioxanthine (9), 1-hydroxy-2,3-dimethoxy-10-methylacridone (10), 1,3-dimethoxy-10-methylacridone (11), skimmianine (12), koku saginine (13), monrifoline (14), 1-hydroxy-4-methoxy-10-methylacridone (15), syringaresinol (16), limonin (17), 1-hydroxy-3,6-dimethoxy-8-methylxanthone (18), xantho xyletin (19), imperatorin (20), scoparone (21) and umbelliferone (22) [11-25]. Amongst the twenty-two compounds were three terpenoids (6, 7 and 17), eleven phenolic compounds (1–5, 16, 18–22), and eight alkaloids (10–17). The isolated terpenoids were steroid (6), triterpenoid (7), and limonoid (17) whilst the alkaloids included five acridones (8–11, 15) and three furanoquinolines (12–14). The phenolics obtained herein were six xanthonones (1–5, 18), one lignan (16) and four coumarins (19–22). The isolation and identification of compounds 1–5 from *G. nobilis* was previously reported [10]. The occurrence of alkaloids and terpenoids from *O. suaveolens* was reported [5,6], and their isolation in the present study is in consistence with previous reports. The occurrence of coumarins, quinoline alkaloids, and free aliphatic acids was also reported in *B. camerunensis* [9]. However, in this study, only coumarins were isolated. The crude extracts as well as the isolated compounds [excluding compounds 6, 7, 12 (known to have low or no antimicrobial activity), and 14 (isolated in very low quantities)] were tested for the antibacterial activities against Gram-negative bacteria and *M. tuberculosis* and the results are summarized in Tables 1 and 2.

The results of the MIC determinations (Table 1) showed that the crude extract from *G. nobilis* (GNB) was the most active amongst the studied extracts, its inhibitory effects being noted on 12 of the 14 tested bacteria. GNB also

exhibited the best activity against *M. tuberculosis* ATCC 27294 (MIC of 128 µg/ml) and *E. coli* ATCC10536 (MIC of 64 µg/ml) than OSR, OSW and BCB. The inhibitory effects of the extracts OSR and OSW from *O. suaveolens* were noted on 9/14 and 7/14 studied bacteria respectively, meanwhile that of the extract BCB of *B. camerunensis* was observed on 4/14 pathogens tested. Interestingly, compound **2** isolated from the most active extract GNB, also exhibited the best activity, preventing the growth of all the fourteen tested microorganisms. The lowest MIC obtained with compound **2** was 8 µg/ml against *M. tuberculosis* ATCC 27294 and the clinical MTCS2 strains. It is noteworthy that compound **2** was more active than chloramphenicol on two Gram-negative MDR bacteria, namely *E. coli* AG102 and *E. aerogenes* CM64 (Table 1). Other compounds showed selective activities, their effects being noted on 1/14 tested bacteria for compounds **16**, **17**, **19** and **20**; 2/14 for **1**; 3/14 for **18**; 4/14 for **9** and **21**; 5/14 for **22**; 7/14 for **3**, **4**, **10** and **11**; 9/14 for **8**; and 11/14 for **5** and **13**. The results of the MBC determinations (Table 2) also showed the activities of the studied samples on some of the tested microorganisms. As observed with MIC data (Table 1), the lowest MBC value (Table 2) was recorded with compound **2** (16 µg/ml) against *M. tuberculosis* ATCC 27294.

Phytochemicals are routinely classified as antimicrobials on the basis of susceptibility tests that produce MIC in the range of 100 to 1000 µg/ml [33]. The activity is considered to be significant if MIC values are below 100 µg/ml for crude extract and moderate when the MICs vary from 100 to 625 µg/ml [34,35]. Also, the activity of compounds is considered to be significant when the MIC is below 10 µg/ml, and moderate when such values vary between 10 and 100 µg/ml [34,35]. On the basis of such criteria, the activity of the studied crude extracts can mostly be considered as moderate, though a significant effect was observed with GNB on *E. coli* ATCC strains. Compound **2** was significantly active against *M. tuberculosis* ATCC 27294 and MTCS2 strains. However its activities can also be considered as moderate against the majority of the bacteria tested. All the tested compounds were active on at least one of the studied microorganisms, and their presence can explain the activity of the crude extracts. Nonetheless, it can be observed that the activity of GNB was not detected on both *P. stuartii* ATCC NAE16 and *P. aeruginosa* PA124, while the extract yielded at least one active compound on these bacteria (Table 1). This can be explained by the fact that the activity of the crude extract does not only depend on the presence of the active compounds, but is also influenced by the quantity and/or possible interaction with other constituents of the plant. This observation can also be applied when carefully analysing the activities of the crude extracts from *O. suaveolens* and *B.*

camerunensis and their constituents (Table 1). The activity of the crude extracts and compounds studied herein (and mostly compound **2**) can be considered interesting when regarding the medical importance of the studied bacteria. In fact, the clinical MDR Gram-negative bacteria tested express active efflux pumps and are involved in many therapeutic failures [28]. To the best of our knowledge, the antibacterial activities of the extracts of *G. nobilis*, *O. suaveolens* and *B. camerunensis* as well as those of most of the studied compounds are being reported for the first time. Nevertheless, some of the isolated compounds were reported for their antibacterial properties. In effect, lupeol (**7**) is known to have low antibacterial activities, the lowest MIC value obtained against *Enterococcus faecalis* ATCC 29212 being 63 µg/ml [36]. The activity of compound **7** was also not detected against the sensitive *Mycobacterium smegmatis* when it was tested at a concentration up to 312 µg/ml [37] and consequently, this compound was not tested again in this work. Previously we also reported the moderate activity of the alkaloid **13** on some sensitive Gram-negative bacteria as well as its low effect against *M. smegmatis* [37]. This compound was not more active against MDR bacteria as observed in the present work, confirming its low antibacterial potential. It has also been demonstrated that compounds **3** (active only on two of 21 tested microorganisms) [38] and **16** (inhibition zone diameters for 10 µl discs at 10⁴ ppm reported as 0.0; 0.2; 0.4 and 0.5 mm against *K. pneumoniae*, *S. aureus*, *Pseudomonas syringae* and *Bacillus subtilis* respectively) [39] have low antibacterial activities. Such reports are in accordance with the results obtained in the present study.

Though compounds **17** (limonin) and **20** (imperatorin) showed poor activities as reported herein, their antilisterial inhibitory effects were found moderate, the MIC values obtained against five *Listeria monocytogenes* ATCC strains varying between 15.62-31.25 µg/ml [40].

Compounds **19** (xanthoxyletin) and **22** (umbelliferone) were also found inactive against *M. tuberculosis* and poorly active against Gram-negative bacteria, consolidating the previously reported data [41,42]. It is noteworthy that compound **12** was reported not to have any bacterial activity [43] and was not tested in the present work.

When regarding the structure-activity relationship, it appears that coumarins had the lowest activities, none of them been active against *M. tuberculosis*. The tested tetranortriterpenoid, compound **17** also showed very weak antibacterial activities. These results are in consistency with previous studies, showing the low antibacterial activities of terpenoids and coumarins against bacteria expressing MDR phenotype [2]. Alkaloids also showed low antibacterial activities. However moderate inhibitory effects were noted with **10** (1-hydroxy-2,3-dimethoxy-10-methylacridone) and **13** (kokusaginine) respectively

against *E. coli* ATTC and AG100 strains (Table 1). Amongst alkaloids, compound **13**, one of the three isolated furanoquinolines, showed the best spectrum of activity (contrary to the five acridone alkaloids) and was found active on both *M. tuberculosis* and Gram-negative bacteria. Within the acridone alkaloids, and when comparing the effects of compounds **10** and **15** [antibacterial spectra (7/14 of the tested bacteria for **10** and 4/14 for **15**); lowest MIC value (64 µg/ml for **10** and 128 µg/ml for **15**)], it appears that the presence of methoxyl- (–OCH₃) group in C2 (compound **10**) increases the bacterial susceptibility (Figure 1, Table 1). A comparison of the activities of compounds **8** and **9** [the presence of –OCH₃ group in C1 (compound **8**) instead of –OH group (compound **9**)] on one hand, and those of **11** and **15** [the presence of –OCH₃ group in C1 (compound **11**) instead of the hydroxyl (–OH) group (compound **15**)] on another hand seems to confirm the fact that the natural substitution of –OCH₃ by –OH in the studied alkaloids increases their activities (Figure 1, Table 1). The best antibacterial activities were recorded with xanthonenes. Within the studied xanthonenes and when regarding the activities of compounds **2** and **3**, it appears that the presence of additional prenyl- group in C4 significantly increases the antibacterial activity (Figure 1, Table 1). Between compounds **4** and **5**, it can also be deduced that the cyclisation increase the activity (Figure 1, Table 1).

Conclusion

The results of the present investigation are important, in regards to the medical importance of the studied microorganisms. Hence, these data provide evidence that some of the constituents of *G. nobilis*, *O. suaveolens* and *B. camerunensis* and mostly compound **2** (isolated from *G. nobilis*), as well as some of the crude extracts from the three plants could be potential antimicrobial products to fight MDR bacteria. The combination of the three plants as used locally in the treatment of infections will further be investigated to provide better understanding to the traditional use.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

HF, ATM and VK carried out the study; HF and VK wrote the manuscript; VK, AEN, CDM, SF, MIC, JJMM and NL supervised the work. All authors read and approved the final manuscript.

Acknowledgements

Authors are thankful to the Cameroon National Herbarium (Yaoundé) for the plant identification. Authors are also grateful to the International Foundation for Science (IFS-Grant F/4579-2 to VK), TWAS for financial support for ICCBS-TWAS fellowship at the H.E.J. Research Institute of Chemistry, International Center of Chemical Sciences, University of Karachi, Pakistan on year 2010 (HF). Authors are also thankful to UMR-MD1 (Mediterranean University, Marseille, France) for providing some clinical bacteria.

Author details

¹Department of Organic Chemistry, Faculty of Science, University of Yaoundé 1, P.o.box 812, Yaoundé, Cameroon. ²Department of Biochemistry, Faculty of Science, University of Dschang, Dschang, Cameroon. ³Department of Organic Chemistry, Higher Teachers Training College, University of Maroua, P.O.Box 46, Maroua, Cameroon. ⁴H. E. J. Research Institute of Chemistry, International Center for Chemical and Biological Sciences, University of Karachi, Karachi 75270, Pakistan. ⁵Department of Plant Science, Faculty of Agricultural and Biological Science, Pretoria 0002, South Africa.

Received: 16 August 2012 Accepted: 8 April 2013

Published: 10 April 2013

References

1. Kuete V, Ngameni B, Tangmouo JG, Bolla JM, Alibert-Franco S, Ngadjui BT, Pagès JM: **Efflux pumps are involved in the defense of Gram-negative bacteria against the natural products isobavachalcone and diospyrone.** *Antimicrob Agents Chemother* 2010, **54**:1749–1752.
2. Kuete V, Alibert-Franco S, Eyong KO, Ngameni B, Folefoc GN, Nguemaving JR, Tangmouo JG, Fotso GW, Komguem J, Ouahouo BM, Bolla JM, Chevalier J, Ngadjui BT, Nkengfack AE, Pagès JM: **Antibacterial activity of some natural products against bacteria expressing a multidrug-resistant phenotype.** *Int J Antimicrob Agents* 2011, **37**:156–161.
3. Ampofo AS, Waterman GP: **Xanthonenes from three *Garcinia* species.** *Phytochemistry* 1986, **25**:2351–2355.
4. Nguyen LH, Venkatraman G, Sim KY, Harrison LJ: **Xanthonenes and benzophenones from *Garcinia griffithii* and *Garcinia mangostana*.** *Phytochemistry* 2005, **66**:1718–1723.
5. Abe MO: **The occurrence of the alkaloid oricine in the wood of *Orcia suaveolens*.** *Phytochemistry* 1971, **10**:3328–3330.
6. Abe MO, Taylor DAH: **A quinolone alkaloid from *Orcia suaveolens*.** *Phytochemistry* 1971, **10**:1167–1169.
7. Letouzey R: *Flore du Cameroun No. 1. Rutacées, Zygophyllacées, Balanitacées.* Paris: Museum National d'Histoire Naturelle; 1963.
8. Tsassi VB, Hidayat H, Meffo BY, Kouam SF, Dongo E, Schulz B, Green IR, Krohn K: **Anti-microbial coumarins from the stem bark of *Afraegle paniculata*.** *Nat Prod Commun* 2010, **5**:559–561.
9. Asase A, Oteng-Yeboah AA, Odamtten GT, Simmonds MSJ: **Ethnobotanical study of some Ghanaian anti-malarial plants.** *J Ethnopharmacol* 2005, **99**:273–279.
10. Fouotsa H, Meli Lannang L, Djama Mbazono C, Rasheed S, Marasini BP, Ali Z, Devkota KP, Kengfack AE, Shaheen F, Choudhary MI, Sewald N: **Xanthonenes inhibitors of α-glucosidase and glycation from *Garcinia nobilis*.** *Phytochem Lett* 2012, **5**:236–239.
11. Gella MD, Monaco P, Previtera L: **Stigmasterols from *Typha latifolia*.** *J Nat Prod* 1990, **53**:1430–1435.
12. Sholichin M, Yamasaki K, Kasai R, Tanaka O: **¹³C Nuclear Resonance of lupane-type triterpenes, lupeol, betulin and betulinic acid.** *Chem Pharm Bull* 1980, **28**:1006–1008.
13. Govindachari TR, Pai BR, Subramaniam PS, Subramanyam V: **A synthesis of evoxanthine.** *Tetrahedron* 1967, **23**:1827–1829.
14. Ermias D, Abiy Y, Peter GW, Alexander IG: **The chemical systematics of the Rutaceae, subfamily Toddaliodeae, in Africa.** *Biochem Syst Ecol* 1988, **16**:179–188.
15. Vaquette J: **Alkaloids from leave of *Teclea boiviniana*.** *Planta Med* 1978, **60**:78–84.
16. Al-Rehailly AJ, Ahmad MS, Muhammad I, Al-Thukair AA, Perzanowski HP: **Furoquinoline alkaloids from *Teclea nobilis*.** *Phytochemistry* 2003, **64**:1405–1411.
17. Wondimu A, Dagne E, Waterman PG: **Quinoline alkaloids from the leaves of *Teclea simplicifolia*.** *Phytochemistry* 1988, **27**:959–960.
18. Kuzovkina I, Al'terman I, Schneider B: **Specific accumulation and revised structures of acridone alkaloid glucosides in the tips of transformed roots of *Ruta graveolens*.** *Phytochemistry* 2004, **65**:1095–1100.
19. Semwal S, Sharma RK: **A new lignan rhamnoside from *Bauhinia retusa* seed pods (Caesalpinaceae).** *Chinese Chem Lett* 2011, **22**:1081–1083.
20. Barton DHR, Pradhan SK, Sternhell S, Templeton JF: **Part XXV: The constitutions of limonin and related bitter principles.** *J Chem Soc* 1961:255–275.
21. Chaurasia N, Wichtl M: **Sterols and Steryl Glycosides from *Urtica dioica*.** *J Nat Prod* 1987, **50**:881–885.

22. Lin LC, Yang LL, Chou CJ: Cytotoxic naphthoquinones and plumbagic acid glucosides from *Plumbago zeylanica*. *Phytochemistry* 2003, **62**:619–622.
23. Renmin L, Aifeng L, Ailing S: Preparative isolation and purification of coumarins from *Angelica dahurica* (Fisch. ex Hoffm) Benth, et Hook. f (Chinese traditional medicinal herb) by high-speed counter-current chromatography. *J Chromatograph A* 2004, **1052**:223–227.
24. Ulubelen A, Oksuz S, Aynehchi Y: Salehi Sormaghi MH, Souri A, Mabry TM: Capillarins and Scaporone from *Artemisia lamprocaulos*. *J Nat Prod* 1984, **47**:170–171.
25. Shouce ZYC: *The Committee of Chinese Herb Information*. Beijing: Renmin Weisheng Publisher; 1986.
26. Eloff JN: A sensitive and quick microplate method to determine the minimal inhibitory concentration of plant extracts for bacteria. *Planta Med* 1998, **64**:711–713.
27. Mativandlela SPN, Lall N, Meyer JJM: Antibacterial, antifungal and antitubercular activity of (the roots of) *Pelargonium reniforme* (CURT) and *Pelargonium sidoides* (DC) (Geraniaceae) root. *S Afr J Bot* 2006, **72**:232–237.
28. Lacmata ST, Kuete V, Dzoyem JP, Tankeo SB, Teke GN, Kuate JR, Pages JM: Antibacterial Activities of Selected Cameroonian Plants and Their Synergistic Effects with Antibiotics against Bacteria Expressing MDR Phenotypes. *Evid Based Complement Alternat Med* 2012, **2012**:623723.
29. Kuete V, Ngameni B, Fotso Simo CC, Kengap Tankeu R, Tchaleu Ngadjui B, Meyer JJM, Lall N, Kuate JR: Antimicrobial activity of the crude extracts and compounds from *Ficus chlamydocarpa* and *Ficus cordata* (Moraceae). *J Ethnopharmacol* 2008, **120**:17–24.
30. Tereschuk ML, Riera MVQ, Castro GR, Abdala LR: Antimicrobial activity of flavonoid from leaves of *Tagetes minuta*. *J Ethnopharmacol* 1997, **56**:227–232.
31. Zgoda JR, Porter JR: A convenient microdilution method screening natural products against bacteria and fungi. *Pharmaceut Biol* 2001, **39**:221–225.
32. Jimenez-Arellanes A, Meckes M, Ramirez R, Torres J, Luna-Herrera J: Activity against multidrug-resistant *Mycobacterium tuberculosis* in Mexican plants used to treat respiratory diseases. *Phytother Res* 2003, **17**:903–908.
33. Simões M, Bennett RN, Rosa EA: Understanding antimicrobial activities of phytochemicals against multidrug resistant bacteria and biofilms. *Nat Prod Rep* 2009, **26**:746–757.
34. Kuete V: Potential of Cameroonian plants and derived products against microbial infections: a review. *Planta Med* 2010, **76**:1479–1491.
35. Kuete V, Efferth T: Cameroonian medicinal plants: pharmacology and derived natural products. *Front Pharmacol* 2010, **1**:123.
36. Gallo MBC, Sarachine MJ: Biological activities of lupeol. *Intl J Biomed Pharma Sci* 2009, **3**:46–66.
37. Kuete V, Wansi JD, Mbaveng AT, Kana Sop MM, Tcho Tadjong A, Penlap Beng V, Etoa FX, Wandji J, Marion Meyer JJ, Lall N: Antimicrobial activity of the methanolic extract and compounds from *Teclea afzelii* (Rutaceae). *S Afr J Bot* 2008, **74**:572–576.
38. Kuete V, Komguem J, Penlap Beng V, Meli AL, Tangmouo JG, Etoa FX, Lontsi D: Antimicrobial components of the methanolic extract from the stem bark of *Garcinia smeathmannii* Oliver (Clusiaceae). *S Afr J Bot* 2007, **73**:347–354.
39. Kim Y-G, Watanabe N, Sano Y, Uraki Y, Sano Y: Extractives of *kitakobusi Magnolia kobus* DC. var. *borealis* Sarg. III: Antibacterial and Antifungal Activity of Extractives. *Res Bull Hokkaido Univ For* 1998, **55**:63–73.
40. Rahman A, Na M, Kang SC: Antilisterial potential of imperatorin and limonin. *J Food Biochem* 2012, **36**:217–223.
41. Sunthitikawinsakul A, Kongkathip N, Kongkathip B, Phonnakhu S, Daly JW, Spande TF, Nimit Y, Rochanaruangrai S: Coumarins and carbazoles from *Clausena excavata* exhibited antimycobacterial and antifungal activities. *Planta Med* 2003, **69**:155–157.
42. Kuete V, Metuno R, Ngameni B, Tsafack AM, Ngandeu F, Fotso GW, Bezabih M, Etoa FX, Ngadjui BT, Abegaz BM, Beng VP: Antimicrobial activity of the methanolic extracts and compounds from *Treculia obovoidea* (Moraceae). *J Ethnopharmacol* 2007, **112**:531–536.
43. Olila D, Opuda-Asibo J: Olwa-Odyek: Bioassay-guided studies on the cytotoxic and *in vitro* trypanocidal activities of a sesquiterpene (Muzigadial) derived from a Ugandan medicinal plant (*Warburgia ugandensis*). *Afr Health Sci* 2001, **1**:12–15.

doi:10.1186/1472-6882-13-81

Cite this article as: Fouotsa et al.: Antibacterial constituents of three Cameroonian medicinal plants: *Garcinia nobilis*, *Orcia suaveolens* and *Balsamocitrus camerunensis*. *BMC Complementary and Alternative Medicine* 2013 **13**:81.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

