

Table 1. Polyunsaturated fatty acids and esters thereof studied in the present work

PUFAs and esters thereof	Acronym
Linoleic acid ^a	LA
Ethyl linoleate ^a	EtL
Linolenic acid ^a	LnA
Ethyl linolenate ^a	EtLn
11,11-Dideuterolinoleic acid ^b	11,11-D ₂ -LA
Ethyl 11,11-Dideuterolinoleate ^b	11,11-D ₂ -EtL
11,11,14,14-Tetradeuterolinolenic acid ^b	11,11,14,14-D ₄ -LnA
Ethyl 11,11,14,14-tetradeuterolinolenate ^b	11,11,14,14-D ₄ -EtLn

^a Purchased from "Sigma".

^b Synthesized by V. Shmanai's group (Minsk State University); the extent of deuteration not less than 98% [31].

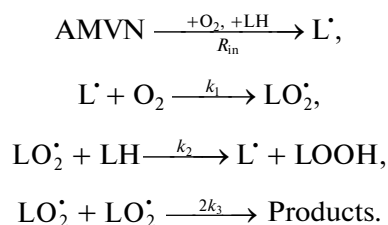
radical-initiated oxidation of PUFAs is virtually non-existent. The present work, devoted to studying the KIE in the oxidation of PUFAs in the initiated-oxidation mode, is essentially a pioneering one.

EXPERIMENTAL

Measurements were carried out in chlorobenzene in air at 310 K. In addition to the oxidation substrates, the reaction mixture contained 2,2-azobis(2,4-dimethylvaleronitrile) (AMVN, Polyscience Inc.), as the source (initiator) of active radicals, and 6-hydroxy-2,2,5,7,8-pentamethylchroman (HPMC, Sigma), as a reference antioxidant in the procedure of measuring the rate of generation of radicals R_{in} (initiation). AMVN and HPMC were introduced into the reaction mixture as reference solutions in chlorobenzene. The kinetics of O₂ uptake during the oxidation was studied using a glass capillary microvolumeter. The oxidation rate, R_{OX} , was determined from the slope of the [O₂] versus time dependence. The initiation rate was measured by means of the inhibitor method using HPMC.

RESULTS AND DISCUSSION

The kinetics of the oxidation of PUFAs was studied in the controlled chain reaction mode. This approach enables to maintain R_{OX} at a constant level during a particular kinetic experiment. The process under consideration can be described by a simple kinetic scheme [9–11]:



The rate of the chain oxidation of LH PUFA is given by

$$R_{OX} = \frac{k_2[\text{LH}]R_{in}^{1/2}}{(2k_3)^{1/2}}. \quad (1)$$

The initiation rate R_{in} was calculated from the induction period of the inhibited chain oxidation (t_{ind}) at the known concentration of HPMC reference antioxidant:

$$R_{in} = \frac{2[\text{HPMC}]}{t_{ind}}. \quad (2)$$

The fit of the experimental data to Eq. (1) is confirmed by the results of Fig. 2. The values of the oxidizability parameter $k_2/(2k_3)^{1/2}$ calculated from Eq. (1) are listed in Table 2.

The primary product of the oxidation of PUFAs is the conjugated pentadienyl radical, $\sim\text{HC}=\dot{\text{C}}\text{H}-\text{CH}=\text{CH}-\dot{\text{C}}\text{H}-\text{CH}=\text{CH}\sim$ (LO_2^{\bullet}). The structure of this radical was confirmed by its EPR spectrum [32].

Table 2. Oxidation of PUFA and esters thereof at 310 K

PUFA	$\frac{k_2}{(2k_3)^{1/2}},$ $\text{L}^{0.5} \text{mol}^{-0.5} \text{s}^{-0.5}$
Methyl oleate	0.0021 ^a
LA	0.0207 ± 0.009 (6) ^b
EtL	0.0230 ± 0.009 (3)
LnA	0.051 ± 0.004 (5)
EtLn	0.046 ± 0.003 (3)

^a From [11].

^b Figures in parentheses indicate the number of parallel runs.

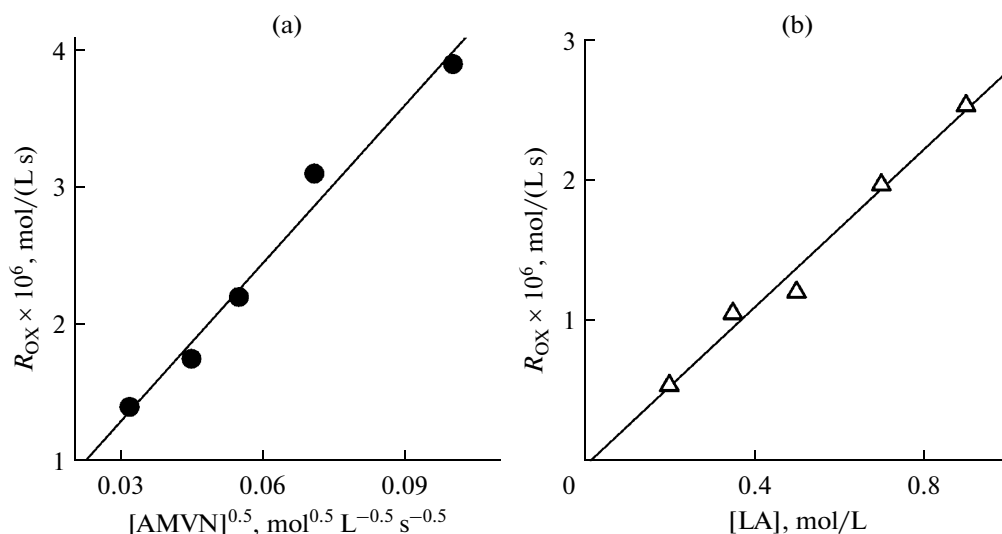


Fig. 2. Dependence of the oxidation rate of LA on (a) [AMVN] at [LA] = 1.0 mol/L and (b) [LA] at [AMVN] = 0.005 mol/L in chlorobenzene at 310 K.

The selective deuteration of the PUFAs (replacement of H atoms by deuterium atoms in the most active CH_2 groups at positions, 11, 11 in LA and 11,11,14,14 in LnA (Fig. 1) leads to a drastic reduction in R_{OX} . A typical kinetic experiment is shown in Fig. 3. It can be seen that the rate of the oxidation of uninhibited 11,11,14,14- D_4 -LnA is virtually identical to the initiation rate. A similar effect is observed for the oxidation of 11,11- D_2 -LA and the respective ethyl esters. Furthermore, the addition of HPMC, a typical antioxidant, which terminates oxidation chains in the reaction with LO_2^* , does not change R_{OX} . These observations indicate that the oxidation of the above selectively deuterated PUFAs is a nonchain process.

Kinetic Isotope Effect

The KIE can serve as a quantitative measure of the effect of deuteration on the chemical reaction kinetics. The most natural way to determine the KIE is to measure the ratio of the rates for the oxidation of nondeuterated ($R_{OX,H}$) and deuterated ($R_{OX,D}$) compounds.

Table 3. Kinetic isotope effect for some deuterated PUFAs in chlorobenzene at 310 K, [AMVN] = 0.01 mol/L

PUFA	[PUFA], mol/L	KIE
11,11- D_2 -LA	1.0	≥ 90
11,11- D_2 -EtL	1.0	≥ 120
11,11,14,14- D_4 -LnA	0.80	≥ 230
11,11,14,14- D_4 -EtLn	0.80	≥ 210

Unfortunately, this approach is not suitable for the above selectively deuterated PUFAs. The fact is that, as already mentioned, the rate of the oxidation of compounds of this kind does not differ from R_{in} . In this case, the $(R_{OX,H})/(R_{OX,D})$ ratio enables to estimate only the lower limit of the KIE (Table 3). This table shows that the estimated KIE values for the acids and the respective esters differ little from each other.

Oxidation of a Mixture of Deuterated and Nondeuterated PUFA

Figure 4 displays the dependence of the rate of the oxidation of a mixture of deuterated and nondeuterated PUFAs (a mixture of 11,11- D_2 -LA and LA) on its composition. The data presented in Fig. 4 show that the partially deuterated PUFA protects the nondeuterated analogue from chain oxidation. For example, the experimental value of the rate of oxidation of an equimolar mixture of LA and 11,11- D_2 -LA is $0.5 \times 10^{-6} \text{ mol}/(\text{L s})$, whereas the additive value of R_{OX} is $2.4 \times 10^{-6} \text{ mol}/(\text{L s})$. Thus, the addition of 50% deuterated analogue of LA is responsible for approximately a fivefold reduction in R_{OX} .

Generally, the KIE for the oxidation of PUFAs is considerably higher than for other compounds. In particular, the KIE value is 10.6–15 for the reaction styrene peroxy radicals with substituted phenols [33, 34], 24 for the reaction of ascorbate with 2,2,6,6-tetramethylpiperidine-1-oxyl [35], 18 for the vitamin E radical with ubiquinol-10 [36], 3–8 for the reaction of ascorbate with the free radical formed from α -tocopherol in micellar solutions [37]. Note that the oxidation of PUFAs is so far the only example in which the KIE value changes nonadditively with the composi-

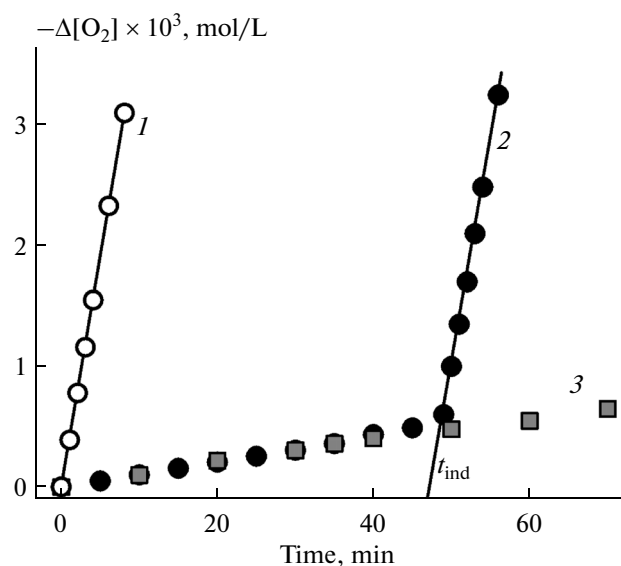


Fig. 3. Comparison of the kinetic characteristics of the oxidation of LnA and 11,11,14,14-D₄-LnA: (1) LnA oxidation (0.57 mol/L), (2) LnA oxidation (0.57 mol/L) in the presence of 1×10^{-4} mol/L HPMC, and (3) 11,11,14,14-D₄-LnA oxidation (0.57 mol/L); chlorobenzene, [AMVN] = 0.025 mol/L, 310 K.

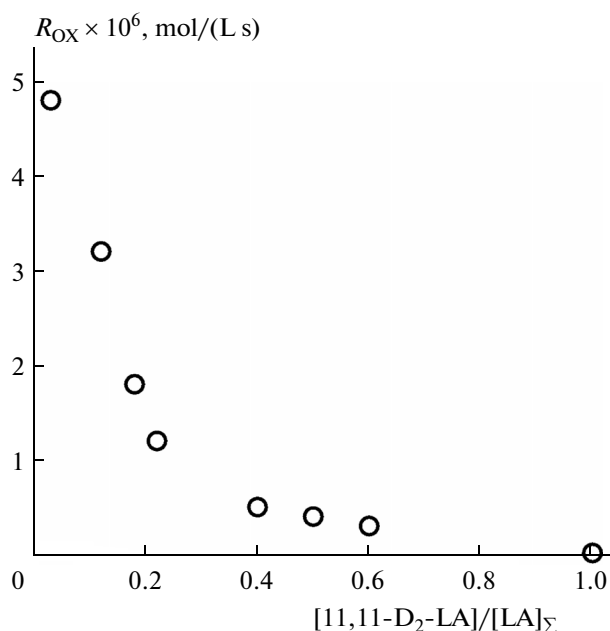


Fig. 4. Dependence of the rate of the oxidation of a mixture of LA and 11,11-D₂-LA on its composition. The total concentration of PUFA is 1.0 mol/L, [AMVN] = 0.025 mol/L, 310 K.

tion of the reaction mixture composed of nondeuterated and deuterated compounds.

Some kinetic features of the oxidation of PUFAs in which the bis-allylic CH₂ group are fully deuterated (11,11-D₂-LA, 11,11-D₂-EtL, 11,11,14,14-D₄-LnA, 11,11,14,14-D₄-EtLn) seem somewhat unexpected and have received so far no rational explanation. The matter is that, in addition to the above deuterated fragments, these compounds have potentially reactive allyl groups (~CH₂-CH=CH~). Given that the values of R_{OX} for these compounds do not differ markedly from R_{in} , it can be argued that these allyl fragments do not take part in the oxidation. This means that the deuteration of a bis-allylic CH₂ group leads to a significant reduction in the reactivity simple allylic C-H bonds.

The literature has repeatedly pointed out that many diseases are caused by an increased rate of the oxidation of PUFAs [22–30]. It is expected that the decrease of R_{OX} may lead to a reduction in the intensity of such diseases. One way to reduce R_{OX} is the deuteration of PUFAs. It is such an effect, namely the non-additive decrease of R_{OX} with increasing concentrations of the deuterated component in the mixture (mixture of 11,11-D₂-LA and LA, Fig. 4) that is demonstrated by the results of the present study. This means that a mixture of deuterated and nondeuterated PUFAs can provide the same biological effect as the deuterated PUFA. In principle, this enables to achieve the same biological effect using a smaller concentration of the deuterated PUFA, a more valuable component. Note that, in contrast to the above results, the rate of the enzymatic oxidation of such mixtures obeys

the additivity rule [22]. Therefore, it should be emphasized that the mechanism of the influence of PUFA deuteration on R_{OX} is far from having been elucidated.

ACKNOWLEDGMENTS

The author are grateful to V.V. Shmanai for kindly providing deuterated PUFAs, to M.E. Solov'ev for help with the quantum-chemical calculations, as well as to E.M. Pliss and I.V. Tikhonov for useful discussions.

The work was supported by Retrope Comp. company (USA).

REFERENCES

1. N. Erdemoglu, S. Kusmenoglu, and M. Vural, *Eur. J. Lipid Sci. Technol.* **106**, 160 (2004).
2. M. Buchgraber, F. Ulberth, H. Emons, et al., *Eur. J. Lipid Sci. Technol.* **106**, 621 (2004).
3. K. Aitzetmuller, B. Matthaus, and H. Friedrich, *Eur. J. Lipid Sci. Technol.* **105**, 92 (2003).
4. T. T. Reed, *Free Rad. Biol. Med.* **51**, 1302 (2011).
5. G. Spitteller, *Free Rad. Biol. Med.* **41**, 362 (2006).
6. G. Paradies, G. Petrosillo, V. Paradies, et al., *Free Rad. Biol. Med.* **48**, 1286 (2010).
7. M. Inoue, E. F. Sato, M. Nishikawa, et al., *Redox Rep.* **9**, 237 (2004).
8. J. Parry, L. Su, M. Luther, et al., *J. Agric. Food Chem.* **53**, 566 (2005).
9. N. A. Porter, *Acc. Chem. Res.* **19**, 262 (1986).

10. J. P. Cosgrove, D. F. Church, and W. A. Pryor, *Lipids* **22**, 299 (1987).
11. V. A. Roginsky, *Mol. Biol.* **24**, 1582 (1990).
12. A. D. French, A.-M. Kelterer, G. P. Johnson, et al., *J. Mol. Struct.* **556**, 303 (2000).
13. S. Korcek, J. H. B. Chenier, J. A. Howard, et al., *Can. J. Chem.* **50**, 2285 (1972).
14. L. R. C. Barclay and K. U. Ingold, *J. Am. Chem. Soc.* **103**, 6478 (1981).
15. L. R. C. Barclay, S. J. Locke, and J. M. MacNeil, *Can. J. Chem.* **61**, 1288 (1983).
16. L. R. C. Barclay, S. J. Locke, J. M. MacNeil, et al., *Can. J. Chem.* **63**, 2633 (1985).
17. N. A. Porter and D. G. Wujek, *J. Am. Chem. Soc.* **106**, 2626 (1984).
18. V. A. Roginsky, *Kinet. Katal.* **31**, 546 (1990).
19. V. Roginsky, *Chem. Phys. Lipids* **163**, 127 (2010).
20. S. Scheiner, *Biochim. Biophys. Acta* **1458**, 28 (2000).
21. A. Kohen, *Prog. React. Kinet. Mech.* **28**, 119 (2003).
22. M. H. Glickman, J. S. Wiseman, and J. P. Klinman, *J. Am. Chem. Soc.* **116**, 793 (1994).
23. S. Peng and W. A. van der Donk, *J. Am. Chem. Soc.* **125**, 8988 (2003).
24. D. L. Luthria and H. Sprecher, *Biochim. Biophys. Acta* **1213**, 1 (1994).
25. K. W. Rickert and J. P. Klinman, *Biochemistry* **38**, 12218 (1999).
26. C. Jacquot, A. T. Wecksler, C. M. McGinley, et al., *Biochemistry* **47**, 7295 (2008).
27. M. J. Knapp and J. P. Klinman, *Biochemistry* **42**, 11466 (2003).
28. A. Kohen and J. P. Klinman, *Acc. Chem. Res.* **31**, 397 (1998).
29. C. Jacquot, S. Peng, and W. A. van der Donk, *Bioorg. Med. Chem. Lett.* **18**, 5959 (2008).
30. M. J. Knapp and J. P. Klinman, *Eur. J. Biochem.* **269**, 3113 (2002).
31. S. Hill, K. Hirano, V. V. Shmanai, et al., *Free Rad. Biol. Med.* **50**, 130 (2010).
32. E. Bascetta, F. D. Gunstone, and J. C. Walton, *J. Chem. Soc., Perkin Trans. II*, No. 5, 603 (1983).
33. J. A. Howard and K. U. Ingold, *Can. J. Chem.* **40**, 1851 (1962).
34. J. A. Howard and K. U. Ingold, *Can. J. Chem.* **41**, 1744 (1963).
35. I. Sajenko, V. Pilepic, and C. J. Brala, *J. Phys. Chem. A* **114**, 3423 (2010).
36. A. Ouchi, S. Nagaoka, and K. Mukai, *J. Phys. Chem. B* **114**, 6601 (2010).
37. R. H. Bisby and A. W. Parker, *Arch. Biochem. Biophys.* **317**, 170 (1995).

Translated by V. Smirnov