Mixed Ligand Complexes Derived from Semicarbazone Schiff Base and Heterocyclic Ligands: Structure and Antimicrobial Activity¹

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Abstract—A series of M(II) ternary complexes $[M(L)Phen]$ and $[M(L)Oxine]$, where $M = Co(II)$, $Ni(II)$, $Cu(II)$, and $Zn(II)$, $L = ((2-hydroxynaphthalen-1-vl)methylene) -4-nitrobenzohydrazide, Phen = 1,10-phenan$ throline and Oxine = 8-hydroxy chloroquinoline, were synthesized and characterized by UV-Vis, ESR, IR, mass spectra, and TG-DTA data. Morphology and crystallinity of the complex [Ni(L)Phen] were studied by scanning electron microscopy. The crystal systems of Cu(II) complexes were determined by powder X-ray diffraction. Biological activity of the ligand and its metal complexes has been screened against four bacterial strains.

Keywords: ternary metal(II) complex, powder XRD, docking, fluorescence, antimicrobial **DOI:** 10.1134/S1070363218050274

INTRODUCTION

Transition metal complexes of Schiff bases are among the most adaptable and thoroughly studied systems up to date due to their numerous clinical, analytical and industrial applications [1]. The Schiff base ligand family of hydrazide and hydrazones is of certain interest due to chelating ability, structural flexibility, interesting electrical and magnetic properties of their complexes with transition metals [2,3]. Such complexes demonstrate some potential in treatment of cancer, tuberculosis [4], leprosy, and mental disorders [5]. 1,10-phenanthroline (phen) molecule, which contains nitrogen as a hetero atom, and its derivatives exhibited antiviral and antifungal activities [6]. The stable phenanthroline complexes inhibited DNA or RNA polymerase and induced strand scission of DNA in the presence of reducing agents [7, 8]. Derivatives of 8-hydroxyquinoline (oxine) and their metal complexes have been used as antibacterial and antifungal agents. One of the oxine derivatives, clioquinol (5-chloro-7 iodo-8-hydroxyquinoline), and its copper complex were used as antifungal, antiprotozoal, antialzheimer, and anticancer drugs [9].

Based on these observations and considering the pharmacological significance of metal complexes of hydrazones and hetero cyclic ligands (1,10-phenanthroline and 5-chloro-8-hydroxy quinoline), here we report synthesis, characterization and antimicrobial studies of some transition metal(II) ternary complexes containing hydrazone and heterocyclic ligands [10]. Our attention was directed to the study of crystal and complex structures.

RESULTS AND DISCUSSION

The ligand was synthesized according to the earlier reported method. Synthetic approach to ternary metal(II) complexes is presented in Scheme 1 and Table 1. Struc-tures of the complexes were established by IR, UV-Vis, ESR, and Mass spectra, TG-DTA, powder X-ray diffraction, and scanning electron microscopy (SEM) analysis. All the complexes were colored, non-hygroscopic and stable at room temperature, insoluble in water and alcohol, but soluble in DMF and DMSO.

IR spectra. IR bands recorded at 3418, 3214, 1245, 1625, and 1580 cm^{-1} for the free ligand were attributed to $v(OH)$, $v(N-H)$, $v(C=O)$, $v(C=O)$, and $v(C=N)$, respectively [11]. The band attributed to the azo-

 $¹$ The text was submitted by the authors in English.</sup>

Scheme 1. Synthetic route to Cu(II), Zn(II), Ni(II), and Co(II) ternary complexes.

 $M = Cu(II), Zn(II), Ni(II)$ and $Co(II)$.

methine group of the free ligand was shifted in complexes to $1572-1624$ cm⁻¹ which confirmed binding of azomethine nitrogen to the metal ion [12]. In the spectra of [M(L)Phen] complexes bands of free phenolic–enolic hydroxide were not recorded due to deprotonation, and the ν(C−O) stretching band was shifted to the range of $1193-1244$ cm⁻¹, indicating coordination of oxygen to the metal ion. The IR spectrum of free 1,10-phenanthroline exhibited the intense bands at 738 and 853 cm^{-1} attributed to aromatic ring vibrations and the band at 1421 cm^{-1} of the pyridyl vibrations [13]. The above bands were shifted to 720–834 and 1061–1108 cm^{-1} , respectively, in their metal complexes spectra [14].

In the spectrum of [M(L)Oxine] complexes the band recorded in the range of $2924-3217$ cm⁻¹ was

assigned to the shift of $v(N-H)$ band typical for the free ligand. Repositioning of the $v(C=O)$ band to the range of $1616-1651$ cm⁻¹ was the result of coordination between oxygen atom of the amide carbonyl and the metal ion [13]. The band $v(C=N)$ of pyridine ring recorded at 1580 cm^{-1} was shifted to the lower frequency region $1462-1497$ cm⁻¹ in the spectra of its metal complexes. The ν(C–O) band of free oxine, 1120 cm⁻¹, was observed in the range of 962–1088 cm⁻¹ in the spectra of its metal complexes [15]. The new bands recorded in the range of 513–581 and 448– 484 cm^{-1} for the oxine complex spectrum indicated coordination between the metal and oxygen, and the metal and nitrogen atoms, respectively [15].

ESR spectra. ESR spectra of [Cu(L)Phen] and [Cu(L)Oxine] were recorded in DMSO solution at

	Color	mp, °C	Found, $%$				Calculated, %			
Compound			C	H	N	M	\mathcal{C}	Н	N	M
[Cu(L)Phen]	Black	280	62.22	3.66	12.09	10.97	62.44	3.32	12.14	11.01
[Co(L)Phen]	Brown	307	62.90	3.46	12.28	10.12	62.95	3.35	12.23	10.30
[Ni(L)Phen]	Brick red	280	62.75	3.12	12.33	10.22	62.97	3.35	12.24	10.26
[Zn(L)Phen]	Orange	310	62.03	3.64	12.06	11.22	62.24	3.31	12.10	11.30
[Cu(L)Oxine]	Light green	340	56.29	2.29	9.72	11.30	56.26	2.27	9.72	11.02
[Co(L)Oxine]	Black	382	56.75	3.12	9.85	10.35	56.71	3.00	9.80	10.31
[Ni(L)Oxine]	Gray	314	56.70	3.31	9.84	10.22	56.73	3.00	9.80	10.27
[Zn(L)Oxine]	Light orange	398	56.08	2.93	9.72	11.31	56.08	2.96	9.69	11.31

Table 1. Elemental analysis and physical data for metal(II) ternary complexes

room temperature (Table 2). Spectra of Cu(II) complexes demonstrated one intense peak in the high field region. The spectrum of $[Cu(L)Phen]$ demonstrated a typical axial peak with well-defined g_{\parallel} and g_{\perp} values at 2.2910 and 2.0832 , and $\lceil Cu(L)Ox \rceil$ complex demonstrated the similar values at 2.2582 and 2.0962 respectively. The geometric parameter G, which measures the exchange interaction between copper centers in the polycrystalline compound, was calculated using the equation:

$G = (g_{||} - 2.0023)/(g_{\perp} - 2.0023).$

The *G* values for the complexes [Cu(L)Phen] and $[Cu(L)Oxine]$ were 3.5686 and 2.7252, respectively, which indicated the considerable exchange interaction [11]. In both Cu(II) complexes the values $g_{\parallel} > g_{\perp} >$ 2.0023 indicated the presence of an unpaired electron in d_x^2 _{-v}2 orbital [12–16]. The g_\perp values (2.0832 and 2.0962) were almost the same for both Cu(II) complexes indicating the similar bonding nature [17, 18]. The *g||* values lower than 2.3 and the *G* values for Cu(II) complexes lower than 4 [19–21] indicated that the metal ligand bonding was of significant covalent nature. The order of values $g_{\parallel} \geq g_{\perp}$ suggested the square pyramidal geometry of five coordinated complexes.

Electronic spectra. The electronic spectrum of free Schiff base demonstrated three bands at 265, 320, and

Table 2. ESR data for Cu(II) ternary complexes

Complex	g_{\parallel}	g⊥	
[Cu(L)Phen]	2.2910	2.0832 3.5686	
[Cu(L)Oxine]	2.2582	2.0962 2.7252	

377 nm [10] that indicated the $\pi-\pi^*$ transition in aromatic benzene ring, the $\pi-\pi^*$ transition in azomethine and the $n-\pi^*$ transition of carbonyl and nitro chromophores [22], accordingly. In the spectra of metal(II) complexes the second and third bands were shifted to a longer wavelength [23] due to donation of electrons lone pairs of nitrogen and oxygen of the Schiff base to the metal ion and the broad band in the range 385–471 nm could be assigned to LMCT. The broad band in the range of 688–723 nm was due to the *d–d* transition [24]. The expected weak *d–d* transition in the visible region for the paramagnetic nickel complexes could not be detected even for concentrated solutions, probably due to its overlap with the low energy tail of the charge transfer transition [25]. The Zn(II) complexes did not exhibit any *d–d* transitions typical for diamagnetic d^{10} configuration.

Mass spectra. Mass spectra data of the complexes supported their compositions and indicated that the complexes were monomeric with the metal to ligand ratio 1 : 1 : 1.

Thermal analysis. The complex [Ni(L)Phen] exhibited thermal stability up to 200°C which confirmed that the complex was free from coordinated water molecules. Decomposition proceeded in three steps. In the first step the nitro group of the Schiff base ligand was given up at the range of 200–300°C. In the second step the observed fulminant loss of coordinated phenanthroline ring took place in the range of 400– 600°C. In third step the Schiff base ligand gradual decomposition was observed at 600–1000°C, and the leftover non-volatile component was metal oxide [24]. The entire decomposition process of the complex [Ni(L)Phen] was endothermic.

Fig. 1. Optimized geometries of (a) [Cu(L)Phen] and (b) [Cu(L)Oxine].

X-Ray diffraction. The powder X-ray diffraction data for the complexes $[Cu(L)Phen]$ and $[Cu(L)Oxine]$ are presented in Table 3. The indexed results [25, 26] yielded the monoclinic unit cell for both complexes. The figures of merit were sufficiently acceptable to support the obtained indexing results $M(20) = 35.6$ for $[Cu(L)Phen]$ and $M(20) = 24.8$ for $[Cu(L)Oxine]$. The best estimated space group was *P*12/*m*1 in the monoclinic system for both complexes.

Morphological analysis (SEM and EDX). SEM image of the free ligand revealed its powdery and soft morphological nature [10]. Upon chelation of the ligand with a metal(II) ion substantial changes in morphology of the compound were observed. From the SEM image of [Ni(L)Phen], sharp nanorod structures with various crystal sizes were observed, whereas agglomeration of [Cu(L)Phen] molecules in nano size poly crystalline particles was determined.

Molecular modeling. According to the computation data for [Cu(L)Oxine] complex, bond length and bond energies of $Cu^{17}-O^{28}$ were 2.049 Å and 244.563 kcal/mol, respectively. Those were different from the other Cu–L bond lengths and energies, indicating that oxygen occupied the axial

Crystal data	[Cu(L)Phen]	[Cu(L)Oxine]	Crystal data	[Cu(L)Phen]	[Cu(L)Oxine]
Empirical formula		$C_{30}H_{19}CuN_5O_4$ $C_{27}H_{16}ClCuN_4O_5$	α , deg	90	90
$M_{\rm W}$	578.1	576.4	β , deg	115.469	124.9
Content of volume	649.16	617.7	γ , deg	90	90
Volume per atom	16.917	15.576	V, \mathring{A}^3	676.683	591.87
Crystal size, nm	34.33	20.4	Ζ	2	2
Lattice strain	0.0043	0.0082	2θ range	$10 - 80$	$10 - 70$
Space group	P12/m1	P12/m1	D_{calc} , g/cm ³	1.419	1.617
Crystal system	Monoclinic	Monoclinic	$\mu(CuK_{\alpha})$, cm ⁻¹	1.543	1.543
Secific surface area, m^2/g	12.317	18.18	Number of peaks indexed	16	13
a, \AA	22.952	21.347	ρ_{max}	0.17409	0.14021
b, \AA	2.5605	2.856	$\rho_{\rm min}$	0.0032	θ
c, \AA	12.7538	11.838	d (resolution)	1.19834	1.3353
			Reflection no.	1547	2687

Table 3. Crystallographic data for complexes [Cu(L)Phen] and [Cu(L)Oxine]

Fig. 2. Interaction of (a) Ligand-4YLD (dotted lines represents the H-bond), (b) [Co(L)Phen]- 4YLD, and (c) [Ni(L)Oxine]-2VAM.

position. The remaining Cu–L fragments were planar (Fig. 1a). In case of [Cu(L)Phen] all five Cu–L coordinated bond lengths and bond angles were similar (Fig. 1b).

Docking studies. In the present study the target microbial proteins data were obtained from protein data bank (PDB). The possible docking modes between the compounds and the target microbial proteins [*B. subtilis* (PDB: 2VAM) and *S. rolfsii* (PDB: 4YLD)] were studied (Fig. 2). Binding energy and shape based orientation energies for each docking were calculated (Table 4)*.* The energy values of metal complexes were lower than those of ligand against the bacterial protein *B. subtilis.* This indicated that the binding affinity to *B. subtilis* protein was higher for metal complexes, whereas binding affinity to *S. rolfsii* protein lower, except [Co(L)Oxine]. The results of docking matched well the experimental results of antimicrobial activity. The binding modes and geometrical orientations of all the compounds were

almost identical, suggesting that all inhibitors occupied the similar cavity in the receptor but the binding affinities were variable.

Fluorescence emission spectra. Schiff base exhibited a strong fluorescence emission at 513 nm with excitation at 450 nm. In contrast with fluorescence spectra of the Schiff base, partial fluorescence quenching phenomenon was observed in its metal complexes demonstrating emission peaks at 545, 565, 553, and 544 nm for Co(II), Ni(II), Cu(II), and Zn(II) complexes, respectively. The metal complexes of [M(L)oxine] demonstrated weak fluorescence emission peaks at 513, 531, 546, and 541 nm for Co(II), Ni(II), Cu(II), and Zn(II) complexes, respectively. The fluorescence spectral data indicated that fluorescence emission intensity of Schiff base was decreased dramatically upon its formation of complexes with transition metal ions. The decrease in emission intensity of [M(L)Phen] complexes was in the order of

	Bacillus subtilis (PDB: 2VAM)		Sclerotium rolfsii(PDB: 4YLD)			
Compound	E_{tot}^a , kJ/mol	$E_{\text{shape}}^{\text{b}}$, kJ/mol	$E_{\text{tot}}^{\text{a}}$, kJ/mol	$E_{\text{shape}}^{\text{b}}$, kJ/mol		
Ligand (L)	-312.60	-312.60	-326.4	-326.4		
[Cu(L)Phen]	-315.82	-315.82	-321.44	-325.66		
[Co(L)Phen]	-356.05	-356.05	-311.20	-321.16		
[Ni(L)Phen]	-321.88	-321.75	-255.20	-253.16		
[Zn(L)Phen]	-318.28	-316.24	-253.89	-253.21		
[Cu(L)Oxine]	-338.59	-338.59	-303.89	-295.75		
[Co(L)Oxine]	-335.78	-331.41	-335.61	-333.61		
[Ni(L)Oxine]	-361.32	-361.32	-319.53	-315.08		
[Zn(L)Oxine]	-335.73	-335.73	-268.65	-275.59		

Table 4. Docking data for the ligand and its metal complexes against *Bacillus subtilis* and *Sclerotium rolfsii* proteins

^a Total binding energy (score). ^b Shape based orientation energy.

 $L > Ni(II) > Zn(II) > Cu(II) > Co(II)$ and in the case of [M(L)Oxine] complexes it in the order of $L > Co(II)$ > $Ni(II)$ $> Cu(II)$ $> Zn(II)$.

Antimicrobial activity. The synthesized Schiff base and its metal complexes were screened for their antimicrobial activity by the well diffusion method (Kirby-Bauer method). The bacterial strains of *E. coli, S. aureus, B. subtilis and P. putida* and fungal strains of *S. rolfsii and M. phaseolina* and standard antibacterial and antifungal drugs, streptomycin and endofill, accordingly, were incubated for 24 h at 37°C. Activity of the compounds was tested at different minimum inhibition concentrations (MIC). At the MIC value of 200 µg/mL, the *B. subtilis* and *S. aureus* demonstrated high inhibition, whereas *E. coli* and *P. putida* did not exhibit significant inhibition in the presence of metal complexes (Table 5). Antifungal activity was exhibited by the compounds only at concentration of 1 mg/mL and at lower concentration no activity was detected (Table 6). The metal complexes were more potent than the Schiff base.

EXPERIMENTAL

Metal salts, $[CoCl_2·6H_2O]$, $[CuCl_2·2H_2O]$, $[Zn(CH_3COO)_2·6H_2O]$, [NiCl₂·2H₂O], and hydrazine hydrochloride were purchased from Merck. Nitro benzoic acid, 2-hydroxy naphthaldehyde, 5-chloro quinolin-8-ol and 1,10-phenanthroline were purchased from Sigma Aldrich. The metal ratio for the complexes was determined on a Perkinelmer-500 atomic absorption spectrophotometer. Melting points of the compounds were measured on a Polmon instrument (Model no. MP-96). IR spectra were recorded on a Schimadzu FT-IR 8400S spectrophotometer using KBr pellets. UV-Vis spectra were recorded on a Shimadzu UV-Vis 2600 in DMSO at room temperature in the wavelength range of 200–800 nm. Fluorescence study was carried out on a Shimadzu spectrofluorometer at room temperature. ESR spectra were recorded by ESR Spectrometer (JEOL-Japan). CHN Analysis was carried out on a Vario macro elemental analyzer (USA). Powder X-ray diffraction data were collected at room temperature on a Shimadzu diffractometer using monochromatic Cu K_{α} radiation (λ = 1.54060 Å). The patterns were scanned over the angular range $1.64^{\circ} - 70^{\circ}$ and $10^{\circ} - 80^{\circ}$ for $\lceil Cu(L)Ox \rceil$ and $\lceil Cu(L) \rceil$ Phen], respectively. Indexing of the powder diffraction pattern was carried out with the expo-2014 software program. The pattern was pretreated by calculating and subtracting the back-ground then smoothing before stripping $K_{\alpha 2}$ radiation by using the Pearson method. Diffraction peaks were searched automatically based on the Cheby-shave method with the number of coefficients 20. Indexing has been carried out using peak positions read from the powder diffraction profiles by DICVOL6 and NTREOR9 methods. Morphology of compounds was studied on a ZEISS SEM-EDX packed scanning elec-tron microscope. Thermal degradation of compound was studied on a Mettler TG-DTA analyzer. Docking study of compounds was carried out by using Hex 8.0 software. ESP mapping was done by Argus Lab 4.0 software.

	Sensitivity				Sensitivity						
Compound	MIC concentration, μ g/mL	Bacillus subtilis	Staphylococcus aureus	Escherichia coli	Pseudomonas putida	Compound	MIC concentration, μ g/mL	Bacillus subtilis	Staphylococcus aureus	Escherichia coli	Pseudomonas putida
Ligand (L)	50				$\overline{}$	[Co(L)Phen]	50	$^{+}$	$^{+}$	$^{+}$	
	100	$^+$	$^{+}$	$+$	$^{+}$		100	$^{+}$	$^{+}$	$^{+}$	
	200	$^{+}$	$++$	$\overline{}$	$\qquad \qquad -$		200	$^{+}$	$++$	$^{+}$	
[Co(L)Oxine]	50	$^{+}$	$+$		$\overline{}$	[Ni(L)Phen]	50	—	$^+$		
	100	$+++$	$++$	—	—		100	$^{+}$	$++$	$^{+}$	
	200	$^{+++}$	$++$	$+$	$\overline{}$		200	$^{+}$	$^{+++}$	$^{+}$	
[Cu(L)Oxine]	50	$^{+}$		$\overline{}$	$\overline{}$	[Zn(L)Phen]	50	$^{+}$	$++$	$\overline{}$	
	100	$++$	$^{+}$	$+$	$\qquad \qquad -$		100	$^{++}$	$^{++}$	$^{+}$	
	200	$^{+++}$	$++$	$+$	$\qquad \qquad -$		200	$^{+++}$	$^{+++}$	$^{+}$	
[Zn(L)Oxine]	50	$+$		$\qquad \qquad -$		[Cu(L)Phen]	50		$^{+}$	—	
	100	$+++$	$++$				100	$++$	$++$	$+$	
	200	$++$	$^{+++}$		$\overline{}$		200	$^{++}$	$^{+++}$	$^{+}$	
[Ni(L)Oxine]	50	—	$++$	$+$	$\overline{}$	Streptomycin	100	$+++$	$^{+++}$	$++++$	$++++$
	100	$^{+}$	$^{+++}$	$^{+}$	$\qquad \qquad -$						
	200	$++$	$^{+++}$	$\! + \!\!\!\!$							

Table 5. Antibacterial activity of the ligand and its metal complexes

Table 6. Antifungal activity of the ligand and its metal complexes at 1 mg/mL

	Inhibition zone, mm						
Compound	Sclerotium rolfsii	Macrophomina phaseolina					
Ligand (L)	2						
[Co(L)Oxine]	4	6					
[Cu(L)Oxine]		3					
[Ni(L)Oxine]	3	1					
[Zn(L)Oxine]		4					
[Co(L)Phen]		5					
[Cu(L)Phen]		1					
[Ni(L)Phen]	2						
[Zn(L)Phen]	4	1					
Endofill							

Molecular modeling of metal complexes was carried out using the Argus Lab 4.0 software program. Geometry optimization was carried out using the Universal force field (UFF). The selected bond distances, bond angles and bond energies of [Cu(L) Phen] and [Cu(L)Oxine] can be presented by the authors. The Hex 8.0 program was applied for protein docking using spherical polar fourier correlations. Parameters used for docking included: correlation type shape only, FFT mode 3D, grid dimension 0.6, receptor range 180, ligand range 180, twist range 360, distance range 40, and solutions 2000, and search program included: SPF transform, FFT steric scanning, FFT final search, energy refinement, and total docking. Fluorescence emission spectra of the Schiff base (L) and its metal ternary complexes [M(L)Phen] and [M(L)Oxine], concentration 0.03 mM, were recorded in DMSO at room temperature with the measured emission wave length range from 200 to 800 nm.

Synthesis of ternary metal(II) complexes. Hot methanol solution of the ligand [(2-Hydroxynaphthalene-1-yl)methylene-4-nitrobenzohydrazide] (1.0 mmol), hot methanol solution of a metal salt, $[CoCl_2·6H_2O]$, $[CuCl₂·2H₂O]$, $[Zn(CH₃COO)₂·6H₂O]$, or $[NiCl₂·$ $2H₂O$ (1.0 mmol), and methanol solution of 1,10phenanthrolin/5-chloro-8-hydroxyquinoline (1.0 mmol) were mixed together upon constant stirring. The mixture was refluxed for 4–8 h. The mixture was cooled down. The precipitate was filtered off, washed, recrystallized, and dried in a desiccators over P_4O_{10} .

 $[Cu(L)Phen]$. Yield 78%. IR spectrum, v, cm⁻¹: 1244 (C–Ophen), 1580 (C=N), 1600 (N=C–O), 1101 $(C=N_{\text{phen}}), 794, 774 (C=C_{\text{phen}}), 484 (Cu-N), 536 (Cu-O).$ UV-Vis, spectrum, λ*,* nm: 270, 323, 391, 713. MS: m/z : 577 [$M + 1$]. C₃₀H₁₉CuN₅O₄.

 $[Co(L)Phen]$. Yield 76%. IR spectrum, v, cm⁻¹: 1222 (C–Ophen), 1596 (N=C–O), 1586 (C=N), 1106 $(C=N_{\text{phen}}), 773 (C=C_{\text{phen}}), 473 (Co-N), 513 (Co-O).$ UV-Vis spectrum, λ*,* nm: 275, 358, 471, 688. MS: *m/z*: 572 [$M + 1$]. C₃₀H₁₉CoN₅O₄.

 $[Ni(L)Phen]$. Yield 80%. IR spectrum, v, cm⁻¹: 1220 (C–Ophen), 1596 (N=C–O), 1586 (C=N), 1108 $(C=N_{\text{phen}})$, 775, 834 $(C=C_{\text{phen}})$, 470 (Ni–N), 581 (Ni–O). UV-Vis spectrum, λ*,* nm: 264, 323, 377, 458. MS: *m/z*: 572 $[M+1]$. C₃₀H₁₉NiN₅O₄.

[Zn(L)Phen]. Yield 84%. IR spectrum, v , cm⁻¹: 1193 (C–Ophen), 1607 (N=C–O), 1619 (C=N), 1061 $(C=N_{phen})$, 720, 788 $(C=C_{phen})$, 477 (Zn–N), 571 (Zn–O). UV-Vis spectrum, λ*,* nm: 272, 325, 385. MS: *m/z*: 578 $[M+1]$. C₃₀H₁₉ZnN₅O₄.

[Cu(L)Oxine]. Yield 75%. IR spectrum, ν, cm–1: 3217 (N–H), 1243 (C–Ophen), 1651 (C=O), 1624 (C=N), 1462 (C=N_{oxine}), 1088, 748 (C=C_{phen}), 448 (Cu–N), 552 (Cu–O). UV-Vis spectrum, λ*,* nm: 265, 325, 439, 723. MS: m/z : 577 $[M+1]$. C₂₇H₁₇ClCuN₄O₅.

 $[Co(L)Oxine]$. Yield 88%. IR spectrum, v, cm⁻¹: 2924 (N–H), 1277 (C–Ophen), 1623 (C=O), 1572 (C=N), 1497 (C=N_{oxine}), 1088, 833 (C=C_{phen}), 485 (Co–N), 552 (Co–O). UV-Vis, λ*,* nm: 236, 263, 336, 414, 695. MS: m/z : 569.7 (M-1). C₂₇H₁₇ClCoN₄O₅.

[Ni(L)Oxine]. Yield 74%. IR spectrum, ν, cm⁻¹: 2926 (N–H), 1239 (C–Ophen), 1616 (C=O), 1578 (C=N), 1464 (C=N_{oxine}), 1084, 712 (C=C_{phen}), 481 (Ni–N), 530 (Ni–O). UV-Vis spectrum, λ*,* nm: 281, 328, 408. MS: *m/z*: 569 (M-1). C₂₇H₁₇ClNiN₄O₅.

[Zn(L)Oxine]. Yield 78%. IR spectrum, v , cm⁻¹: 2926 (N–H), 1242 (C–Ophen), 1618 (C=O), 1577 (C=N), 1496 (C=N_{oxine}), 962, 711 (C=C_{phen}), 451 (Zn–N), 542 (Zn–O). UV-Vis spectrum, λ*,* nm: 275, 332, 439. MS: m/z : 578 (M+2). C₂₇H₁₇ClZnN₄O₅.

CONCLUSIONS

A number of bivalent ternary metal complexes has been synthesized and characterized by IR, ESR, UV-Vis, and Mass spectrometry. The experimental results matched the theoretical molecular modeling. The latter supported the square pyramidal geometry of the metal complexes. Powder X-ray crystallography revealed the monoclinic crystal structure for [Cu(L)Oxine] and [Cu(L)Phen] complexes. The morphological study of [Ni(L)Phen] and [Cu(L)Phen] supported the polycrystalline nature of the products. According to antimicrobial tests the complexes demonstrated clear antibacterial activity.

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REFERENCES

- 1. Sujarani, S. and Ramu, A., *J. Mol. Struct*., 2014, vol. 1059, p. 299. doi 10.1016/j.molstruc.2013.11.038
- 2. Abou-Melha, K.S. and Faruk, H., *J. Coord. Chem*., 2008, vol. 61, no. 12, p. 1862. doi 10.1080/ 00958970701768455
- 3. Walcourt, A., Loyevsky, M., Lovejoy, D.B., Gordeuk, V.R., and Richardson, D.R., *Int. J. Biochem. Cell Biol*., 2004, vol. 36, no. 3, p. 401. doi 10.1016/S1357-2725(03) 00248-6
- 4. Richardson, D.R. and Bernhardt, P.V., *J. Biol. Inorg. Chem*., 1999, vol. 4, no. 3, p. 266. doi 10.1007/ s007750050312
- 5. Yadawe, M.S. and Patil, S.A., *Transition Met. Chem*., 1997, vol. 22, no. 3, p. 220. doi 10.1023/ A:1018400121316
- 6. Zoroddu, M.A., Zanetti, S., Pogni, R., and Basosi, R., *J. Inorg. Biochem.*, 1996, vol. 63, no. 4, p. 291. doi 10.1016/0162-0134(96)00015-3
- 7. Chikira, M., Tomizawa, Y., Fukita, D., Sugizaki, T., Sugawara, N., Yamazaki, T., Sasano, A., Shindo, H.,

 Palaniandavar, M., and Antholine, W.E., *J. Inorg. Biochem*., 2002, vol. 89, no. 3-4, p. 163. doi 10.1016/ S0162-0134(02)00378-1

- 8. Tardito, S. and Marchio, L., *Curr. Med. Chem*., 2009, vol. 16, no. 11, p. 1325. doi 10.2174/ 092986709787846532
- 9. Pushie, M.J., Nienaber, K.H., Summers, K.L., Cotelesage, J.J.H, Ponomarenko, O., Nichol, H.K., Pickering, I.J., and George, G.N., *J. Inorg. Biochem*., 2014, vol. 133, p. 50. doi 10.1016/ j.jinorgbio.2014.01.003
- 10. Edward, J. T., Gauthier, M., Chub, F.L., and Ponka, P., *J. Chem .Eng. Data,* 1988, vol. 33, no. 4, p. 538. doi 10.1021/je00054a044
- 11. *Essays in Chemistry*, Hathway, B.J., Bardley, G.N., and Gillard, R.D., Eds., New York: Academic Press, 1971.
- 12. Tharmaraj, P., Kodimunthiri, D., Sheela, C.D., and Priya C.S.S., *J. Serb. Chem Soc*., 2009, vol. 74, no. 8-9, p. 927. doi 10.2298/JSC0909927T
- 13. Bindu, P. and Kurup, M.R.P., *Transition Met. Chem*., 1997, vol. 22, no. 6, p. 578. doi 10.1023/ A:1018512708055
- 14. Stamate, D., Ferbinteanu, M., Opreal, C.I., Panait, P., Cimpoesu, F., and Girţu, M.A., Abstracts of Papers, *1 Int. Electronic Conf. on Material*, 2014.
- 15. Malghe, Y.S., Prabhu, R.C., and Raut, R.W., *Acta Pol. Pharm*., 2009, vol. 66, no. 1, p. 45.
- 16. Procter, I.M., Hathaway, B.J., Billing, D.E., and Nicholls, P., *J. Chem. Soc. A,* 1968, p. 1678. doi 10.1039/

J19680001678

- 17. Seena, E.B. and Kurup, M.R.P., *Polyhedron*, 2007, vol. 26, no. 4, p. 829. doi 10.1016/j.poly.2006.09.040
- 18. Joseph, M. Suni, V., Kurup, M.R.P., Nethaji, M., Kishore, A., and Bhat, S.G., *Polyhedron*, 2004, vol. 23, no. 18, p. 3069. doi 10.1016/j.poly.2004.09.026.
- 19. Maki, A.H. and McGarvey, B.R., *J. Chem. Phys*., 1958, vol. 35, no. 29, p. 2935. doi 10.1063/1.1744457
- 20. Chandra, S. and Gupta, L.K., *Spectrochim. Acta, Part A,* 2004, vol. 60, no. 13, p. 3079. doi 10.1016/ j.saa.2004.01.030
- 21. Chityala, V.K., Kumar, K.S., Macha, R., Tigulla, P., and Shivaraj, *Bioinorg. Chem. Appl.*, 2014. doi 10.1155/2014/691260
- 22. Bessy Raj, B.N., Kurup, M.R.P., and Suresh, E., *Spectrochim. Acta, Part A,* 2008, vol. 71, no. 4, p. 1253. doi 10.1016/j.saa.2008.03.025
- 23. Kumar, M.P., Vamsikrishna, N., Ramesh, G., Subhashini, N.J.P., Nanubolu, J.B., and Shivaraj, *J. Coord. Chem*., 2017, vol. 70, no. 8, p. 1368. doi 10.1080/00958972.2017.1292503
- 24. Arish, D. and Nair, M.S., *J. Mol. Struct*., 2010, vol. 983, no. 1, p. 112. doi 10.1016/j.molstruc.2010.08.040
- 25. Han, J., Xing, Y., Wang, C., Hou, P., Bai, F., Zeng, X., Zhang, X., and Ge, M., *J. Coord. Chem*., 2009, vol. 62, no. 5, p. 745. doi 10.1080/00958970802326161
- 26. Ahamed, M.A.R., Azarudeen, R.S., and Kani, N.M., *Bioinorg. Chem. Appl*., 2014, doi 10.1155/2014/764085