# **Temperature Sensitivity (Q10) of Soil Basal Respiration as a Function of Available Carbon Substrate, Temperature, and Moisture**

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**Abstract**—Basal respiration is one of the key indicators of soil C mineralization. Temperature sensitivity  $(Q_{10})$ of basal respiration is important for predicting changes in C mineralization due to warming. A modified methodology of  $Q_{10}$  determination is proposed. Soil samples were incubated at 25 $^{\circ}$ C with periodic short-term (2 h) decline of temperature to 15<sup>o</sup>C and high-frequency measurements of  $CO<sub>2</sub>$  production rates. The temperature sensitivity is estimated as the average rate of  $CO<sub>2</sub>$  production at 25 $^{\circ}$ C (before and after temperature decline) divided by the rate of  $CO_2$  production at 15<sup>o</sup>C. With this method we demonstrated that glucose addition most strongly affects the  $Q_{10}$  values at low temperature ranges (20–10°C), while temperature range affects  $Q_{10}$  stronger than the glucose additions. The negative effect of soil moisture on  $Q_{10}$  of basal respiration was demonstrated: the  $Q_{10}$  values decreased with increasing soil moisture.

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## INTRODUCTION

An increase in the activity of soil organic matter mineralization and  $CO<sub>2</sub>$  emission from soil into the atmosphere as a result of warming is the key mechanism of positive feedback of ecosystems to global climate change [8–14]. It is called positive because warming results in the increasing emission of  $CO<sub>2</sub>$  the main greenhouse gas—which additionally accelerates warming [17–19]. Most models of global carbon cycle take into account this relationship, taking that the rate of carbon mineralization increases twofold with the temperature rise by every 10°C ( $Q_{10} = 2$ ) [12, 14]. Because soils are the main source of  $CO<sub>2</sub>$  in the atmosphere [24, 26, 27], high variability of  $Q_{10}$  values in different soil types can be one of the causes of ambiguities in models of global carbon cycle.

In spite of a great number of works on temperature sensitivity, the mechanisms of response and the factors that control it remain insufficiently studied [20, 21]. Most researchers measure the rate of  $CO<sub>2</sub>$  emission under the temperatures, which do not allow direct calculation of  $Q_{10}$ . The obtained data are approximated by different models (Arrhenius [7], van't Hoff [29], Lloyd–Taylor [22], and others), on the basis of which  $Q_{10}$  and other parameters of temperature sensitivity are calculated. Inequality of models and mathematical errors are the source of high variation in  $Q_{10}$  values obtained by different authors [11, 12, 14, 18, 28, 31, 32].

In this study, we suggest an improved method of estimating  $Q_{10}$  of basal respiration—one of the main indicators of carbon mineralization in soils—by means of automatic regular measurements of temperature [16]. The modification consists of in additional automatic variation in the temperature of soil samples with regular short-term (for 2 h) temperature decrease by 10°С. The short time of temperature decrease is important in order to prevent changes in the species composition of microbial community. On the contrary, many researchers apply temperature rise for determining  $Q_{10}$ . In this case, the rate of mineralization increases, and this results in a quick depletion of substrate available for microorganisms [12, 18]. The substrate deficiency decreases the rate of  $CO_2$  emission and, hence, the  $Q_{10}$  of soil organic matter mineralization.

Our work was aimed to testing the modified method of assessing the temperature sensitivity of С mineralization and to determining the influence of (1) added of easily decomposable organic substrate (glucose) in a wide range of concentrations, (2) temperature range, and (3) soil moisture (and a combination of these factors) on  $Q_{10}$  of C mineralization in the gray forest soil.





**Fig. 1.** (a) Temperature schedule for cycles 1 and 3, (b) the rate of  $CO<sub>2</sub>$  production during incubation of three soil samples (variant without glucose; the calculation of  $Q_{10}$  is illustrated for one of the replicates), and (c) the dynamics of calculated  $Q_{10}$  values.

# *Object of Study*

The samples of the upper  $(0-10 \text{ cm})$  mineral horizon of gray forest soil (А horizon) were taken under three herbaceous birch groves in the area of Akademgorodok (city of Krasnoyarsk). The 80-yearold birch groves are typical ecosystems of Krasnoyarsk forest-steppe. The samples taken under different birch groves were combined into one sample. The obtained sample was sieved through a 1 mm sieve to remove coarse roots and homogenize the soil mass. The time from soil sampling to the beginning of measuring  $CO<sub>2</sub>$ flux was less than 2 h, and this allows us to consider the dynamics of mineralization of fresh organic matter.

## *Measurement of the Rate of CO<sub>2</sub> Production, Calculation of Q10, and Variants of Treatment*

The 100-g portions of soil were placed into 16 glass vessels of 0.5 L in volume, and 5 mL of water solution of glucose in concentrations of 0 (control), 25, 125, 250, 500, 1250, 2500, and 5000 μg С/g were added into every two vessels. The vessels were placed into an incubator with a Peltier cooler and programmed temperature regime (Memmert, Germany). Every vessel was connected via the 16-port multiplexer Li-Cor 8150 with an IR  $CO<sub>2</sub>$  analyzer Li-Cor 8100 (Li-Cor Incorporated, Lincoln, Nebraska, USA). The multiplexer closed by turn each of 16 vessels into a hermetical chain for 1.5 min, and the rate of  $CO<sub>2</sub>$  production by the soil sample was calculated from the rate of  $CO<sub>2</sub>$ accumulation in a vessel. While the measurement was carried out in one vessel, other vessels were vented with room air in order to prevent the accumulation of  $CO<sub>2</sub>$ , the excessive concentration of which limits the rate of  $CO<sub>2</sub>$  production.

Overall, three cycles of measurements were performed, and each cycle continued for three days. Incubation was carried out at 25°С with regular (every 8 h) short-term (2 h) lowering of temperature to 15°С in cycles 1 (low moisture) and 3 (increased moisture). The diagram of temperature regime during incubation is presented in Fig. 1a. In cycle 3, the soil water content was increased from 30 to 100% of the field capacity along with the added glucose. In cycle 2, the range of temperatures (20–10 $^{\circ}$ C) was 5 $^{\circ}$ C lower than in cycles 1 and 3. The rate of  $CO<sub>2</sub>$  production was measured every 2 h; four measurements were performed at temperatures 20 or 25°C, and one measurement characterized low-temperature (10 or 15°C) periods (Fig. 1c). The  $Q_{10}$  coefficient was calculated for every cycle of lowering/rising temperature according to the following equation:  $Q_{10} = (F_1 + F_2)/(2F_3)$ , where  $F_1$  and  $F_2$ are the activities of  $CO_2$  emission at 25°C before and after temperature lowering, and  $F_3$  is the activity of  $CO<sub>2</sub>$  emission at 15°C.

## *Statistical Analysis of Data*

The rate of  $CO_2$  production and calculated  $Q_{10}$  values were tested for normality of distribution (Kolmogorov–Smirnov test) and uniformity of the sample (Levin test). Then, the two-way ANOVA was performed for repeated measurements of the rate of  $CO<sub>2</sub>$ production and  $Q_{10}$  values. The two major factors were the concentration of glucose (eight levels, including the control) and the cycle (three levels, cycles  $1-3$ ). Forty time points for the rate of  $CO<sub>2</sub>$  production and eight points for  $Q_{10}$  were the repeated measurements. The analysis demonstrated the absence of the influence of time on  $Q_{10}$ , i.e., its values did not change with time (Fig. 1c). So, average values were calculated for every variant, and the two-way ANOVA was performed again as described above, but without repeated measurements. All effects were significant at  $P \le 0.050$ .

# RESULTS AND DISCUSSION

The temperature coefficient  $Q_{10}$  for the rate of reaction was determined as a measure of its change with an increase in temperature by  $10^{\circ}$ C [7, 29]. The temperature sensitivity  $(Q_{10})$  of carbon mineralization and basal respiration has been widely studied all over the world [1, 5, 31]. The method suggested by us has certain advantages: (1) it is based on regular short-time lowering of temperature, which allows one to take into account limitation of the rate of reaction by the substrate; (2) the calculation of  $Q_{10}$  is performed without fitting of exponential models, and this excludes mathematical errors and reduces labor expanses; (3) automation increases the accuracy and efficiency of measurements and ensures standardization of the time under particular temperature [16]. Automation of measurements and their high efficiency make it possible to test many hypotheses about the impact of particular substrates or environmental factors on the rate of the  $CO<sub>2</sub>$  production. While automated measurements was previously reported [16], we advanced the methodology by automated temperature variation and the way of  $Q_{10}$  estimation accounting for a decline of  $CO<sub>2</sub>$  respiration rates with time. In this study, we tested the influence of glucose added in different concentrations, soil water content, and soil temperature range on  $Q_{10}$  of basal respiration. The temperature regime during incubation in cycles 1 and 3 is shown in Fig. 1a; it was characterized by periodical lowering of temperature from 25 to 15 $^{\circ}$ C. The rate of CO<sub>2</sub> production gradually decreased in the course of the entire incubation period and during each cycle of temperature lowering (Fig. 1b). This is explained by the loss of easily available carbon and gradual drying of the sample because of ventilation of the vessels [16]. However, calculated values of  $Q_{10}$  were about 2 and did not change with time (Fig. 1c); they were close in different replicates of the experiment, i.e., the applied method ensured stable results.

As expected, the rate of  $CO<sub>2</sub>$  emission was lower at  $20^{\circ}$ C than at  $25^{\circ}$ C (Fig. 2). A sharp increase in the rate of  $CO<sub>2</sub>$  emission was observed, when the samples were moistened; the initial fresh samples were relatively dry, limiting the activity of microorganisms. An increase in the rate of  $CO<sub>2</sub>$  production was also observed with the rise in the concentration of glucose, because the rate of the reaction depends on the amount of the substrate.



**Fig. 2.** The rate of  $CO<sub>2</sub>$  production under different concentrations of glucose added at maximum temperatures in three cycles of measurements at (*1*) low moisture (cycle 1); (*2*) low temperature (cycle 2), and (*3*) high moisture (cycle 3).



**Fig. 3.** The relationship between  $Q_{10}$  and concentration of glucose added at each of three cycles: (*1*) low moisture (cycle 1); (*2*) low temperature (cycle 2), and (*3*) high moisture (cycle 3).

The temperature range had a maximum effect on  $Q_{10}$ , i.e., the  $Q_{10}$  value was significantly greater (Fig. 3) under low temperatures ( $10-20$ °C) than under higher temperatures (15–25°С). The decrease in temperature sensitivity with an increase in temperatures was noted

**Table 1.** The results of the two-way ANOVA (*Р* values). The influence of the time factor (duration of incubation) was estimated the first analysis of variance. The influences of all other factors were obtained from the second analysis of variance, in which time was not taken into account, and time-averaged values were calculated

Factor	P value	
	rate of $CO2$ emission	Q10
Glucose concentration	<0.001	0.235
Incubation cycle	<0.001	< 0.001
Time	0.045	0.187
Concentration $\times$ cycle	0.160	< 0.001

by Arrhenius [7, 12]. The reason for this phenomenon is that the reaction rate increases with the rise in temperature, but, at the same time, the availability (concentration) of the substrate decreases, and this decrease is more pronounced.

The effect of concentration of added С was negative and was only observed in the low temperature range (cycle 2). The addition of glucose in different concentrations had no influence on  $Q_{10}$  of basal respiration in the range of higher temperatures both under low and high moisture content values.

Two concepts were suggested earlier to explain the influence of temperature on the rate of carbon mineralization in relation to the quality and availability of organic matter [12]. The concept of quality (the kinetic theory of Arrhenius [7]) stipulated that soil organic matter is a mixture of different substances, each of which has its own energy of activation, i.e., their mineralization requires some minimal temperature. Most molecules of organic matter are stable, i.e., their energy of activation is high, so that they begin to decompose under high temperatures [12]. On the contrary, glucose is characterized by the low energy of activation and can be mineralized under lower temperatures, and this is the cause of much quicker decomposition of added glucose than soil organic matter by microorganisms [12, 14]. Thus, the emission of  $CO<sub>2</sub>$  and temperature sensitivity of C mineralization in the case of added glucose are mainly determined by glucose mineralization [10, 12, 20, 25, 28]. Hence, the addition of glucose introduction should decrease  $Q_{10}$ of С mineralization. The availability of soil organic matter for microorganisms has an opposite effect on  $Q_{10}$ . In fact, organic substrate should be available in loci of enzyme activity. In other words, there may a lot of organic matter in the soil sample, but its concentration in the reaction loci may be low. In this case, the diffusion transport that depends both on moisture and temperature takes place. In general, the decrease in concentration of available substrate results in the underestimation of  $Q_{10}$ , and this explains the decrease of  $Q_{10}$  in the range of higher temperatures. The kinetic theory of Arrhenius was confirmed experimentally in our study, and the increase on  $Q_{10}$  values was observed in the range of lower temperatures (cycle 3) in comparison with the range of higher temperatures (Fig. 3). However, neither the kinetic theory of Arrhenius nor the concept of availability explain the absence of the effects of glucose and its concentrations on  $Q_{10}$  of basal respiration in cycles 2 and 3. Recent studies suggest the significant influence of priming effect—shortterm change in the rate of C mineralization in the case of carbon or nitrogen addition—on  $Q_{10}$  of C mineralization [2, 32]. We suggest that the decrease in  $Q_{10}$  of С mineralization in response to an increase in glucose concentration, which was observed in the range of lower temperatures, could take place because of priming effect. It is likely that priming effect also existed in the range of higher temperatures. However, the value of  $Q_{10}$  of basal respiration became higher under lower temperatures, and this made the participation of new fractions of organic matter in mineralization processes more pronounced according to  $Q_{10}$  values.

If the  $Q_{10}$  index of glucose mineralization differed significantly from  $Q_{10}$  of soil organic matter mineralization, then the effect of added glucose adding would also be observed in the range of higher temperatures. According to the kinetic theory of Arrhenius, the fraction of mineralizable organic matter decreases in the range of lower temperatures. At the same time,  $Q_{10}$  of С mineralization increased significantly (Fig. 3). We suppose that the addition of glucose caused limitation of the development of microorganisms by nitrogen. In order to get nitrogen, microorganisms began to mineralize soil organic matter, and this process took place at higher temperatures and stopped at lower temperatures. Mineralization of this fraction of organic matter under the influence of priming effect in the range of lower temperatures brought  $Q_{10}$  to the values observed in range of higher temperatures. This assumption is confirmed by the fact that  $Q_{10}$  of basal respiration decreased, when concentrations of added glucose were in the range of  $25-1250 \mu g C/g$ , under which the maximum priming effect is observed [15].

It is also important that we confirmed the decrease in  $Q_{10}$  values with an increase in soil moisture (Fig. 3). The most plausible explanation of this is the increase in the rate of diffusion of organic substances to the loci of enzymatic reactions [3, 4, 6, 11, 23, 30]. This confirms the influence of organic matter availability on  $Q_{10}$  of carbon mineralization.

Thus, environmental factors have different effects on  $Q_{10}$  of soil organic matter mineralization. Prevalent theories—the kinetic theory by Arrhenius and the theory of organic matter availability—do not explain the entire diversity of effects. However, the addition of the theory of priming effect and its temperature sensitivity can improve significantly our understanding of the behavior of  $Q_{10}$  of C mineralization under changing environmental factors.

## **CONCLUSIONS**

(1) It was experimentally confirmed that  $Q_{10}$  of basal soil respiration is higher at the lower temperature range and decreases at the higher temperatures range, which is in agreement with the Arrhenius kinetic theory.

(2) The addition of glucose led to a decrease of  $Q_{10}$ , and this effect was more pronounced in the range of lower temperatures. We argue that priming effect is responsible for the influence of added glucose on  $Q_{10}$ of С mineralization at lower temperatures.

(3) The influence of soil moisture on  $Q_{10}$  was confirmed: with the rise in the soil water content, the  $Q_{10}$ values decreased because of an increase in the diffusion of organic matter to the loci of enzymatic reactions thus confirming the concept about limitation of C mineralization by the availability of the substrate.

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## CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

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