REVIEWS AND THEORETICAL ARTICLES

Epigenetic Effects in Livestock Breeding

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Abstract—Epigenetic effects are considered as a mechanism of the emergence of new inherited traits with their transmission between generations through meiosis. Modern genomic evaluation does not explain the entire phenotypic variance of traits. It is quite obvious that a significant part of the unaccounted dispersion reflects epigenetic effects carried out through DNA methylation, histone and chromatin modifications, and activity of noncoding types of RNA. Epigenetic effects could potentially be used in breeding programs. The obtained data testify to the significant role of epigenetic factors in the expression of imprinting genes, cellular processes, development of muscle tissue, and fat metabolism in animals. The ability of various additives in the diet to induce epigenetic modifications with phenotypic variability has been convincingly proven. However, there are still many contradictions and limitations in the justification of the hereditary component of epigenetics for introduction into animal breeding. Development of modern technologies, such as chromatin immunoprecipitation with microchips of DNA (ChIP-Chip), next-generation sequencing (ChIP-Seq), and epigenetic editing based on CRISPR-Cas9, gives grounds for optimism in solving problems of introducing epigenetic phenomena in livestock breeding.

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INTRODUCTION

The term epigenetics, proposed by C. Waddington [1], reflects one of the mechanisms of phenotypic expression of genes. In the process of revealing new genetic data, this term began to reflect the possibility of the emergence of new inherited traits with their transmission between generations through meiosis without changes in nucleotide sequences of DNA, where the expression of the genotype and the formation of phenotypic traits may depend on internal and external factors. Despite the fact that the genome of the cell is quite stable, the epigenome is very dynamic throughout life and is determined by the complex interaction of genetic and environmental factors [2]. The main epigenetic mechanisms change the expression of genes, which includes DNA methylation processes, histone modification, activity of noncoding RNA (ncRNA), and chromatin remodeling [3], contributing to new phenotypic manifestations in the formation of productivity [4], reproduction [5], aging [6], and diseases [7, 8]. The development and introduction of genomic selection provided an opportunity to significantly enhance genetic progress in livestock breeding [9, 10]. However, epigenetic traits remain unaccounted in indices of pedigree value of animals. Inclusion of the results of the analysis of these traits in the breeding process would be useful in improving the accuracy of the pedigree value of animals. It is generally believed that genes from both parents are equally involved in the development of the embryo. However, these views contradict the obvious genomic imprinting and the manifestation of the differential activity of the maternal and paternal alleles [11]. In mammals, more than 100 genes with monoallelic expression are known [12]. Epigenetic mechanisms regulate a wide range of biological processes, including fetal growth and development, metabolism, and behavior [13]. It is important that epigenetic information is preserved and transmitted to the next generation [14]. The mechanisms of epigenetic heredity probably evolved in the course of evolution [15]. The cognition and use of epigenetic mechanisms can have serious consequences for the formation and analysis of quantitative traits [16] and will ensure that information on epigenetic processes is included in the overall assessment of breeding value, followed by an increase in animal productivity and resistance to diseases.

DNA METHYLATION

Methylation of the fifth carbon of cytosine (5-methylcytosine) is currently the best studied epigenetic modification of DNA. DNA methylation is carried out through enzymatic activity of DNA methyltransferases and is essential for genomic stability and maintenance of development. For example, DNA(cytosine-5)-methyltransferase 1 (DNMT1) maintains mainly DNA methylation during replication, while DNMT3a and DNMT3b are responsible for *de novo* methylation of unmodified DNA. DNA methylation occurs, as a rule, on cytosine-phosphate-guanosine dinucleotides (CpG) [17] and is associated with transcriptional repression, while hypomethylation, conversely, is associated with transcriptional activation.

In fertilization, the paternal genome is actively demethylated, whereas in the maternal genome this process occurs passively. Most of the blastocyst genome is demethylated during implantation. After implantation, the genome of the developing zygote undergoes de novo methylation. After the formation of primary germ cells, the genome is demethylated. This marks the start of the second wave of reprogramming. After sex determination and gametogenesis, the gene is remethylated at different times, depending on the sex. After implantation, the developing zygote undergoes *de novo* methylation and completes the first wave of reprogramming [18]. In the process of gametogenesis, the genome is remethylated for each sex at different times and in varying degrees. The second wave of reprogramming continues during the growth of the oocyte [19]. In embryos of cattle, demethylation occurs in the stage from eight to 16 cells [20]. As noted, the paternal DNA is more demethylated than the maternal DNA [21, 22]. The difference in methylation levels between male and female pronuclei was observed as early as 8 h after copulation in mice and after 10 h in rats [23]. Thus, the maternal genome is more resistant to the effects of active demethylation, but loses most of the methyl groups of DNA during development [24]. There are significant differences in the features of DNA methylation in sperm and oocytes [25–27]. There are specific differences in the course of these processes. Thus, it should be noted that no evidence of a global demethylation of the DNA of the paternal pronucleus in the horse during the first cell cycle was found [28]. Both parental genomes demonstrated stable and similar levels of methylation and hydroxymethylation during the pronuclear process. However, there are reasons to assume that the properties of the differentially methylated regions of the maternal and paternal genomes differ [29–31]. During prenatal development, primordial germ cells, which are gamete precursors, undergo epigenetic reprogramming with DNA demethylation, followed by sexual specific de novo methylation with specificity for a particular sex [32, 33].

The best evidence of transgenerative epigenetic inheritance in mice is the study of epialelles, such as viable yellow and axin agouti, in which the levels of DNA methylation of retrotransposons control the expression of a neighboring gene [34]. DNA methylation of the intracisternal part of the retrotransposon is inserted in front of the agouti gene and controls the expression of this gene. This particle belongs to a small group of long repetition terminals that appear to form germline resistance to reprogramming in sex cells and early embryos of agouti mice leading to a range of coat colors from yellow, yellow and brown spots to brown color depending on the degree of DNA methylation [35]. Methylation of the genome can be controlled by hormonal signals and modifications involving damage to alkylated nucleic acid elements that are closely related to certain diseases [36]. Methylation persists throughout both the preimplantation period and at later stages of development [37, 38].

The change in the status of DNA methylation can have a profound effect on the expression of genes of cloned animals and the induction of epigenetic disorders. Although most offspring from somatic cloning are normal [39], some somatic cloning protocols are associated with pathological side effects, probably as a result of nonstandard epigenetic reprogramming [40]. It is important to note that the kinetics of DNA methvlation and demethylation in cloned embryos after nuclear transfer differs from that in normally fertilized embryos [41]. Consequently, assisted reproductive technologies may be responsible for some of the epigenetic disorders in the development process [5, 42]. Epigenetic reprogramming occurs aberrantly in most cloned embryos, and incomplete reprogramming may contribute to low cloning efficiency [43]. Compared to normal fetuses, cloned embryos in the middle of pregnancy have subtle abnormalities of DNA methylation [44]. Data are presented showing the association between DNA methylation features and perinatal mortality in cattle, together with the contribution of epigenetic factors to phenotypic variability [45].

Considering epigenetic processes, one should note that methylation is not the result of specific target mechanisms, and it is better to consider it as part of the general methylation processes in female and male gamete lines. Only a fraction of these gametes survive in the early stages of development; that is, there is selection of preimplantation embryos.

Obtaining the profiled DNA methylation maps of the entire genome makes it possible to identify the nature of changes in DNA methylation that occur during growth and development [46, 47]. Analysis of genomic DNA methylation profiles makes it possible to identify specific CpG sites associated with the phenotype. Observation data on the features of DNA methylation made it possible to develop a classifier for predicting DNA methylation levels provided high resolution of the CpG site [48]. Undoubtedly, these approaches help determine genomic mechanisms that interact with DNA methylation and suggest processes involved in the modification of DNA methylation and regulation of the connections with various epigenetic processes. Epigenetic markers can change over time owing to the cellular type of specificity [49]. Therefore, it is not surprising that epigenetic effects play an important role in the differentiation of cells [50],

pathological processes [51], and, in particular, carcinogenesis [52].

In epigenetic studies of agricultural animals, nextgeneration sequencing techniques based on DNA methylation were used to study the contribution of methylation to the phenotypic variability of the corresponding economically important features. DNA methylation responds to feeding and environmental conditions leading to a change in the phenotype associated with changes in productivity and the risk of animal disease [53]. Thus, the use of this method made it possible to reveal the number and localization of regions of differential methylation of DNA when omega-3 fatty acids are added to the diet of sows, which affect growth and inflammatory processes [54]. A comparison of DNA methylation profiles between fast and slow growing broilers was carried out [55]. The study revealed candidate genes, including several known growth factor genes (IGF1R, FGF12, FGF14, FGF18, FGFR2, and FGFR3) with differential methylation, which potentially regulate muscle development at the age of seven weeks. A total of 13294 methylated genes were detected, including 4085 differentially methylated genes, of which 132 were differentially methylated genes associated with growth and metabolism.

POST-TRANSLATIONAL HISTONE MODIFICATIONS

The structure of chromatin is under the control of several mechanisms involving the modification of histones and total chromatin, which play an important role in transcription processes [56]. As is known, eukaryotic DNA is tightly packed with the formation of nucleosomes, which are formed into an octamer of four histones (H2A, H2B, H3, and H4). In such a structure, histones are susceptible to many post-translational modifications that form a potential for encoding epigenetic information. Structural changes in chromatin resulting from histone modification tend to result in a set of effector proteins, such as transcription factors, which modulate gene expression [2, 57]. In addition to acetylation, histones undergo various reversible post-translational modifications, including methylation, ubiquitination, ADP ribosylation, and phosphorylation [58]. Ubiquitination can carry both inhibitory [59] and activating [60] effects, depending on the modification of specific sites. The modification of histones and DNA methylation pathways depend on each other, and the amplifying effects occur as a result of the interaction of the corresponding enzymes and the factors associated with them [61, 62]. Chromatin modifications function in two mutually exclusive ways [63] and can directly affect the chromatin structure or can provide dynamic binding of platforms for proteins with specific domains. An example of the first method is acetylation of histones, which neutralizes the positive charge of lysine and thereby disrupts the electrostatic interaction. This contributes to the formation of chromatin with a reduced compact state [64]. In addition, histone acetyltransferases function as a transcriptional coactivator, and deacetylases function as repressors. These transformations provide a stepwise model for the formation of transcriptionalrepressive heterochromatin [65]. Maternal chromatin is organized in such a way that DNA methylation and chromatin modifications often take place during fertilization [66]. They include both nucleohistone modifications and chromatin proteins associated with active and repressive configurations [67, 68]. As a result of the development of the paradigm of allosteric control of mammalian DNA methyltransferases, two new regulatory principles were discovered for them. Thus, it has been shown that their catalytic activity is under the allosteric control of N-terminal domains with autoinhibitory functions in a number of domains [69]. In addition, the activity of DNA methyltransferases has been found, which should be regulated in concert with interaction with post-translational modifications. Here, the state of enzymes DNMT1 and DNMT3, their binding to DNA, and the catalytic mechanism, as well as multimerization (the combination of protein subunits in one complex) and the processes controlling their stability in cells, play a big role. Of particular interest is the reprogramming of histone modification when manipulating cloned embryos.

Methylation of lysine-4 of histone H3 is mainly associated with transcriptionally active chromatin, and methylation of lysine-9 of histone H3 is associated with repressive chromatin [70]. The disruption of reprogramming in cloned mice correlates with their poor potential for early development. Similar data were obtained on cloned embryos of cattle [71]. These results indicate a link between methylation of DNA and histones in mammalian embryos with the identification of associations between epigenetic markers and the potential for the development of cloned embryos. Similar disorders occurred in the differential acetylation of histone H4 in blastomeres of cloned cattle [72]. During the embryonic development of mammals, along with DNA methylation, histone modification plays an important role in regulating gene expression and epigenetic reprogramming. In cloned and in vitro fertilized preimplantation embryos of cows in the stage before eight cells, the levels of acetylation and methylation of histories of H3K9as, H3K18ac. H4K5ac, H4K8ac, H3K4me3, and H3K9me2 were abnormally high [73]. These results indicate that somatic cells in the recipient oocytes produce aberrant histone modifications in several places before the donor genome of the cell is activated. Chromatin research involves repositioning or restructuring of nucleosomes inside the chromatin to facilitate or inhibit access to nearby DNA. This is mainly carried out by ATP-dependent remodeling of chromatin complexes and nucleosomes [74, 75]. Nevertheless, the dynamics in the organization of chromatin in the development process is not a unique system for all vertebrates; for example, it has its specific features in mammals (mice) and in birds (chickens) [76].

NONCODING RNA

Although DNA methylation and histone modifications are the most studied epigenetic mechanisms, there are epigenetic processes that play an important role in the regulation of gene function. A striking demonstration is the role of noncoding RNAs, which can indirectly affect the regulation of gene expression and chromatin modification. There are several known types of noncoding RNA involved in epigenetic heredity: long noncoding RNAs (lncRNA), small interfering RNAs (siRNA), RNAs of 26-32 bp (piwi RNA or piRNA), and microRNAs of 18–25 bp (miRNA), which are involved in the regulation of transcription, translation, and stability of mRNA [77]. Maternal reserves of information RNAs can also be attributed to these categories of RNA with epigenetic properties [78]. Noncoding microRNAs of approximately 22 bp are capable of controlling the activity of about 60% of all protein-encoding genes and are involved in the regulation of almost every studied cellular process in mammals [79, 80]. The number of microRNAs found in various species of farm animals varies considerably from a few dozen to several hundred [81]. The role of microRNA in the formation of the productivity of agricultural animals is associated with many processes [82], including lactation and milk production [4, 83], lipogenesis [84], and morbidity [7]. Long noncoding RNAs (lncRNA) are a heterogeneous group of transcripts longer than 200 bp, which play a noticeable regulatory role in many biological processes [85, 86]. This type of RNA is the largest part of the noncoding transcriptome of mammals [87]. LncRNA have common pathways of biogenesis with other classes of noncoding RNA. Most lncRNAs result from the activity of RNA polymerase II. Many lncRNAs function as epigenetic modulators by binding to chromatin, emergence of catalytic activity on specific sites in the genome, and influence on gene expression [88]. Thousands of genes encoding the lncRNA have been identified in mammalian genomes [89]. The regulating properties of lncRNA associated with pathological states and development of the immune system and aging [90, 91], as well as with the regulation of skin pigmentation processes in cows [92], are noted. Other studies [93] provided characteristics of 584 lncRNA in muscles of cows, some of which were localized within the loci of quantitative traits and correlated with traits of meat productivity. There were 4227 lncRNAs identified in the mammary gland, including 26 lncRNAs that differentially respond to a diet high in alpha-linolenic acid, which suggests their role in the synthesis of fatty acids and lipid metabolism [94]. It was shown that microRNA-103, which consists of 20-24 nucleotides, participates in various biological processes, including brain development, lipid metabolism, adipocyte differentiation, hematopoiesis, and immunity, and also participates in the differentiation of porcine preadipocytes [95]. In cattle, microRNAs are regulated by DNA methyltransferase 1 and are involved in the development of the mammary gland and lactation [96]. Evidence of the effect of stimulation by endogenous miRNA-143 on the differentiation of intramuscular fat of cows through regulated expression of adipocyte genes has been obtained [97].

Several studies of lncRNA in chickens have been carried out to improve understanding of the biology and differentiation of preadipocytes [98, 99]. Comprehensive analysis facilitated the detection of lncRNAs and target genes that can contribute to the regulation of various stages of skeletal muscle development. Cisand trans-regulation of target genes by differentially expressed lncRNAs were determined and lncRNA and gene interaction networks were constructed [100]. The available data suggest that the lncRNA function contains a hidden layer of regulatory information that not only acts as a mediator between DNA and protein but also plays a role in genome organization and gene expression [101]. Specific roles of lncRNA in the development of various organs and tissue types have been determined. For example, in neonatal cardiomyocytes, the knockdown of specific lncRNA alters gene expression and inhibits the normal development of maturation of cardiomyocytes in mice [102] and can lead to embryonic death [103]. Using the constructed chicken ncRNA library, 125 ncRNAs were isolated, which play an important role in the development and differentiation of tissues during evolution [104]. Expression profiles of lncRNA and mRNA in three different stages of skeletal muscle development in chickens were determined [100]. Differentially expressed lncRNAs were analyzed in cis- and transinteraction and used to construct correlation networks of lncRNA genes. An important role of lncRNA in the regulation of transcription [86, 105], in epigenetic modification [106], and in organogenesis and development [107, 108] was noted. The lncRNA group functions as regulators of gene expression and also participates in development and in a number of physiological processes [109]. It should be emphasized once again that lncRNA is the largest part of the noncoding mammalian transcriptome [110]. By pairing bases with mRNA, microRNAs regulate gene expression in animals through inhibition of translation initiation, elongation, and other mechanisms [111]. Many lncRNAs affect the expression of genes by binding to chromatin-modifying proteins, altering the catalytic effect on certain sites in the genome [112].

Analysis of the results of sequencing in chickens identified the localization of nucleotide sequences of lncRNA: 1493 as intergenic and 177 as intragenic, that is, located within the protein-encoding genes [113]. There were also differences in the types of level of mRNA expression in the mammary glands between lactating and nonlactating cows [114, 115]. These manifestations are associated with the action of genes through a variety of biological pathways [116]. In the mammary glands of goats, differential expression of microRNA was detected between lactation and dryperiod peaks and between early and late lactation [117, 118]. It was shown that microRNA-15a inhibits the expression of casein and the number of epithelial cells, as well as the expression of the growth hormone receptor gene, both through mRNA and via protein [119]. MicroRNA-103 of the mammary gland was involved in the control of fat content in milk during lactation of goats [83]. It was revealed that endogenous microRNA-143 plays a role in the differentiation and proliferation of intramuscular fat cells in cows [97]. These data on the participation of epigenetic markers in the regulation of the synthesis of milk lipids and muscles provide the basis for developing ways to manage the quality of milk and meat through the optimization of the concentration of fatty acids. The relationship of the state of microRNAs of ovarian tissues in chickens with low and high levels of productivity was analyzed [120]. Eleven known and six new microRNAs were detected. All 11 known microRNAs were involved mainly in the regulation of reproduction, such as steroid hormone biosynthesis and dopaminergic synapses. Some of the six miRNAs (for example, gga-miR-34b, gga-miR-34c, and gga-miR-216b) regulate proliferation, cell cycle, apoptosis, and metastasis. Thus, in recent years, there has been an explosion in understanding of the previously hidden role of RNA regulation [101]. Currently, there are a large number of databases of expression and analysis of the microRNA sequences [121]. Although many microRNAs have been identified, it is obvious that there may be many more still undetected. The use of genomic techniques with deep sequencing has shown that there are tens of thousands of loci in mammals that express large transcripts that do not encode proteins but have intergenic and intron localization, where many of these transcripts play a functional role [122]. In accordance with their role in differentiation and development, a number of genetic and biochemical data indicate that one of the main functions of lncRNAs and many small RNAs is the regulation of epigenetic processes [123]. Indeed, it seems that noncoding RNAs are an important component of cellular biology, developmental biology, brain function, and, possibly, even evolution [124]. The complexity and interconnection of these systems is a motivation for studying the vast and largely unknown area of regulating biological processes through RNA. It is possible to annotate transcriptional units and identify functional SNPs through the use of epigenetic maps that delineate thousands of lncRNA genes and hundreds of thousands of *cis*-regulating elements [125, 126].

INFLUENCE OF THE ENVIRONMENT

It is known that the effects of chemicals [127], nutritional supplements and nutrient availability [128,

129], maternal behavior [130], pathogenic microorganisms [131], and temperature [132] cause changes in gene expression [130, 133]. An example is the aforementioned change in the phenotype of agouti coloring in mice under the influence of a maternal diet, including folate and betaine [134]. The animal ration can manifest itself as a source of epigenetic modifications [135], including the expression of microRNAs in the subcutaneous and visceral adipose tissue of cattle [84, 136, 137]. The introduction of methylating compounds into the ration of pigs showed significant differences in DNA methylation and gene expression between groups in the generation F2 [138]. Differentially methylated regions in gene promoters were significantly associated with inhibition of expression of obesity-related genes [139]. In cattle, the expression of tissue-specific adipogenic and lipogenic genes of the longest muscle in the back is regulated by the status of DNA methylation [140]. The DNA methylation levels of all five sites of the CpG promoter of the FABP4 gene were lower (p < 0.001) in intermuscular fat than in the muscular part. The mRNA levels of the PPARG1 and *FABP4* genes were negatively correlated with the level of DNA methylation in the regulatory CpG regions of the corresponding genes. Adding maize concentrate to the diet of dairy cows leads to a change in the state of methylation of specific genes involved in fat formation and protein synthesis. Fatty acids can contribute to changes in expression of specific genes during critical growth periods [141]. Lipids and lipoprotein components interact directly with the structure of chromatin with subsequent influence on gene expression [142, 143]. Adding fodder rich in unsaturated fatty acids to the diet of dairy cows produces significant changes in the expression of two histone acetyltransferases [144]. There are various points of potential interaction between nutrition and epigenetics, including the effect of supplements or the deficiency of macroelements as well as a number of secondary plant metabolites on methylation [145, 146]. Targeted dietary supplements with folic acid, choline, and betaine may increase DNA methylation [147–149]. There is evidence of an increase in DNA methylation levels in the regulatory regions of the ZIP4 gene [150] in connection with the use of zinc-containing supplements in the diet.

The change in feeding during late pregnancy can affect the reproductive capacity of daughters [151]. In heifers born to mothers who received a diet high in protein in the second trimester of pregnancy, a decrease in the number of antral follicles was observed [152]. The restriction or excess of protein in the pig's diet during pregnancy influenced the expression of key metabolic genes in the offspring [153, 154]. A lowprotein maternal diet during pregnancy and lactation affects the hepatic cholesterol metabolism of offspring by modifying the epigenetic regulation of the 3hydroxy-3-methylglutaryl coenzyme A reductase and cholesterol-alpha-hydroxylase genes, which suggest possible long-term effects in cholesterol homeostasis later in adulthood [155].

Features of feeding affect the expression of microRNA in subcutaneous and visceral adipose tissue of cattle [136]. A higher amount of microRNAs was found in animals treated with high-fat fodder [156–158]. Epigenetic modifications unlock the loci of the milk protein gene during the development and differentiation of the mammary gland of the mouse. The role of epigenetic factors in unlocking the milk protein loci in the development and differentiation of the mammary gland was demonstrated [159]. In addition, there is increasing evidence that epigenetic factors regulate milk production in cows [4]. There are reports of differences in mRNA expression levels in the mammary glands between lactating and nonlactating cows [114]. It is obvious that epigenetic markers regulate the synthesis of lipids and the production of milk. It remains to be determined how epigenetic factors can be controlled to improve the milk productivity of cows.

There are a number of studies on the relationship of the epigenetic state and pathological processes in animals [160, 161]. DNA methylation around the STAT5binding enhancer in the α S1-casein promoter is associated with a sharp discontinuation of the synthesis of α S1-casein during acute mastitis. Experimental infection of the mammary gland with pathogenic *E. coli* revealed the involvement of epigenetic factors in the synthesis of α S1-casein and the occurrence of mastitis in cattle [162] through hypomethylation of the upper promoter region of the alpha-casein S1 gene. It was shown that the presence of bacteria changed the status of methylation of the CD4 differentiation cluster promoter in cows with mastitis [161].

With the help of the latest generation of deep sequencing technologies, the involvement of microRNAs in the development of viral and bacterial infections in cattle was shown [163]. A number of studies have shown that microRNAs are expressed in a wide range of cattle tissues, including those associated with immunity [164, 165]. It is becoming increasingly evident that microRNAs play a significant role in the formation of cow immunity. Castration in pigs can significantly affect the model of expression of microRNAs involved in lipogenesis. Differentiated expressed miRNAs can play an important role in fat deposition after castration [166, 167].

Thus, a number of external factors affect the formation of phenotypic features, which may have a direct relationship to the breeding and productive qualities of animals.

EVALUATION OF PROSPECTS OF INTRODUCTION OF EPIGENETIC EFFECTS IN LIVESTOCK BREEDING

Epigenomic mutations, as in the case with DNA mutations, can carry positive, neutral, and harmful effects with different potentials of adaptation to environmental signals. Unfortunately, the main limitation of studying the epigenomic effects in livestock breeding is the insufficient recognition of the importance of the contribution of these effects to the formation of economically significant phenotypes, including the risk of diseases. There is much evidence of the involvement of the epigenome, in particular, microRNAs, in many aspects of the formation of the productive qualities of farm animals, such as milk production [168], fat formation [84, 169, 170], early embryonic development [82], and animal morbidity [8]. Usually, two types of epigenetic inheritance are employed: inheritance in the somatic cell line and preservation of changes during mitosis [171], that is, transgeneration epigenetic inheritance through the germline, which controls patterns of gene expression with transmission from one generation to the next [172]. Some authors suggest models for the quantitative evaluation of epigenetic dispersion inherited in populations [173, 174]. However, it should be borne in mind that epigenetic markers are inherited in the germline, at least in mammals, in no more than three generations [175], since epigenetic traits are usually removed during meiosis and are not transmitted to the offspring unless the fourth generation is subject to such influence. Some epigenetic markers avoid this stage of cleansing. Therefore, the majority of hereditary epigenetic dispersions do not cover any broad populations of animals [176]. In addition, there is the problem of differentiation of the additive epigenetic effect from the effect of epistasis, dominance, and other effects on genetic dispersion [177], although epigenetic markers are established in the early stages of development, and adaptation takes place throughout life in response to internal and external stimuli and leads to phenotypic manifestation of a trait in the late stage of an animal's life [178].

It should be noted that the introduction of improved breeding programs, where imprinting is taken into account, will require making changes in the existing standard breeding programs. This will require the inclusion of variable figures for the breeding value of producers and females, as well as an assessment of the influence of dominance and additive genetic deviations. It is necessary to reveal the details of the transmission of hereditary information through epigenetic processes and to more deeply assess the changes in the epigenome that occur during the formation of germ cells and the early ontogeny, and also to have more evidence about the degree of influence of epigenetic factors on the formation of economic characteristics, for example, on the synthesis of milk in cows [4, 168]. With the development of DNA sequencing technologies, the genomic part of the variability of animal traits is currently assessed at a faster rate. Therefore, ways should be developed to take into account the epigenetic contribution to the true value of the breeding value of the individual. This is a new field of activity in animal breeding studies, in which the heterogeneity of epigenetic markers and the differences in their tissue affiliation and developmental stages complicate the task. Transgenerative epigenetic inheritance is a field for discussion in the scientific community, since it includes radically new biological phenomena, affecting even the inheritance of acquired characteristics. There is much uncertainty in the problem of including transgenerative inheritance of epimutations in breeding programs. New research approaches need to be developed to determine quantitative information on phenotypic variability caused by epimutations [179]. The development of new technologies (immunoprecipitation of chromatin with microchips of DNA (called ChIP-Chip), next-generation sequencing (ChIP-Seq), editing of the epigenome based on CRISPR-Cas9, and others) gives hope for solving the problems of introduction of epigenetic effects in livestock breeding [180].

REFERENCES

- 1. Waddington, C.H., The epigenotype, *Endeavour*, 1942, vol. 1, pp. 18–20. doi 10.1093/ije/dyr184
- Bernstein, B.E., Meissner, A., and Lander, E.S., The mammalian epigenome, *Cell*, 2007, vol. 128, pp. 669– 681. doi 10.1016/j.cell.2007.01.033
- Ibeagha-Awemu Eveline, M. and Xin Zhao, Epigenetic marks: regulators of livestock phenotypes and conceivable sources of missing variation in livestock improvement programs, *Front. Genet.*, 2015, vol. 6, pp. 302–312. doi 10.3389/fgene.2015.00302
- Singh, K., Molenaar, A.J., Swanson, K.M., et al., Epigenetics: a possible role in acute and transgenerational regulation of dairy cow milk production, *Animal*, 2012, vol. 6, pp. 375–381. doi 10.1017/S1751731111002564
- Urrego, R., Rodriguez-Osorio, N., and Niemann, H., Epigenetic disorders and altered gene expression after use of assisted reproductive technologies in domestic cattle, *Epigenetics*, 2014, vol. 9, pp. 803–815. doi 10.4161/epi.28711
- Ashapkin, V.V., Kutueva, L.I., and Vanyushin, B.F., Aging epigenetics: accumulation of errors or realization of a specific program?, Biochemistry (Moscow), 2015, vol. 80, no. 11, pp. 1406–1417. doi 10.1134/S0006297915030062
- Karrow, N., Sharma, B., Fisher, R., and Mallard, B., Epigenetics and animal health, in *Comprehensive Biotechnology*, Khatib, H., Ed., Amsterdam: Elsevier, 2011, pp. 381–394.
- Luo, J., Yu, Y., and Song, J. Epigenetics and animal health, in *Livestock Epigenetics*, Khatib, H., Ed., Hoboken, N.J.: Wiley–Blackwell, 2011, pp. 131–145.
- 9. König, S., Simianer, H., and Willam, A., Economic evaluation of genomic breeding programs, *J. Dairy*

Sci., 2009, vol. 92, no. 1, pp. 382–391. doi 10.3168/jds.2008-1310

- Bouquet, A. and Juga, J., Integrating genomic selection into dairy cattle breeding programmes: a review, *Animal*, 2013, vol. 7, no. 5, pp. 705–713. doi 10.1017/S1751731112002248
- Ferguson-Smith, A.C., Genomic imprinting: the emergence of an epigenetic paradigm, *Nat. Rev. Genet.*, 2011, vol. 12, no. 8, pp. 565–575. doi 10.1038/nrg3032
- Henckel, A. and Arnaud, P., Genome-wide identification of new imprinted genes, *Brief Funct. Genomics*, 2010, vol. 9, no. 4, pp. 304–314. https:// doi.org/10.1093/bfgp/elq 016.
- Lambertini, L., Marsi, T.C.J., Sharma, P., et al., Imprinted gene expression in fetal growth and development, *Placenta*, 2012, vol. 33, pp. 480–486. doi 10.1016/j.placenta.2012.03.001
- Murdoch, B.M., Gordon, K., Murdoch, G.K., et al., Nutritional influence on epigenetic marks and effect on livestock, *Front. Genet.*, 2016, vol. 7, p. 182. doi 10.3389/fgene.2016.00182
- Lim, J.P. and Brunet, A., Bridging the transgenerational gap with epigenetic memory, *Trends Genet.*, 2013, vol. 29, no. 3, pp. 176–186. doi 10.1016/j.tig.2012.12.008
- Jammes, H., Junien, C., and Chavatte-Palmer, P., Epigenetic control of development and expression of quantitative traits, *Reprod. Fertil. Dev.*, 2010, vol. 23, pp. 64–74. doi 10.1071/RD10259
- Ziller, M.J., Müller, F., Liao, J., et al., Genomic distribution and inter-sample variation of non-CpG methylation across human cell types, *PLoS Genet.*, 2011. 7:e1002389. doi 10.1371/journal.pgen.1002389
- Santos, F., Hendrich, B., Reik, W., and Dean, W., Dynamic reprogramming of DNA methylation in the early mouse embryo, *Dev. Biol.*, 2002, vol. 241, no. 1, pp. 172–182. doi 10.1006/dbio.2001.0501.14
- Park, J.S., Jeong, Y.S., Shin, S.T., et al., Dynamic DNA methylation reprogramming: active demethylation and immediate remethylation in the male pronucleus of bovine zygotes, *Dev. Dyn.*, 2007. V. 236, no. 9, pp. 2523–2533. doi 10.1002/dvdy.21278
- Triantaphyllopoulos, K.A., Ikonomopoulos, I., and Bannister, A.J., Epigenetics and inheritance of phenotype variation in livestock, *Epigenet. Chromatin*, 2016, vol. 9, p. 31. doi 10.1186/s13072-016-0081-5
- Ci, W. and Liu, J., Programming and inheritance of parental DNA methylomes in vertebrates, *Physiology* (Bethesda), 2015, vol. 30, no. 1, pp. 63–68. doi 0.1152/physiol.00037.2014
- 22. Smith, Z.D., Chan, M.M., Mikkelsen, T.S., et al., Unique regulatory phase of DNA methylation in the early mammalian embryo, *Nature*, 2012, vol. 484, pp. 339–344. doi 10.1038/nature10960
- Zaitseva, I., Zaitsev, S., Alenina, N., et al., Dynamics of DNA-demethylation in early mouse and rat embryos developed in vivo and in vitro, *Mol. Reprod. Dev.*, 2007, vol. 74, pp. 1255–1261. doi 10.1002/mrd.20704

- Reik, W., Stability and flexibility of epigenetic gene regulation in mammalian development, *Nature*, 2007, vol. 447, pp. 425–432. doi 10.1038/nature05918
- 25. Kobayashi, H., Suda, C., Abe, T., et al., Bisulfite sequencing and dinucleotide content analysis of 15 imprinted mouse differentially methylated regions (DMRs): paternally methylated DMRs contain less CpGs than maternally methylated DMRs, *Cytogenet. Genome Res.*, 2007, vol. 113, pp. 130–137. doi 10.1159/000090824
- Kobayashi, H., Sakurai, T., Sato, S., et al., Imprinted DNA methylation reprogramming during early mouse embryogenesis at the Gpr1-Zdbf2 locus is linked to long cis-intergenic transcription, *FEBS Lett.*, 2012, vol. 23, pp. 827–833. doi 10.1016/j.febslet.2012.01.059
- Kobayashi, H., Sakurai, T., Sato, S., et al., Contribution of intragenic DNA methylation in mouse gametic DNA methylomes to establish oocyte-specific heritable marks, *PLoS Genet.*, 2012. 8, e1002440. doi 10.1371/journal.pgen.1002440
- Heras, S., Smits, K., De Schauwer, C., and Van Soom, A., Dynamics of 5-methylcytosine and 5-hydroxymethylcytosine during pronuclear development in equine zygotes produced by ICSI, *Epigenet. Chromatin*, 2017. doi 10.1186/s13072-017-0120-x
- Biliya, S. and Bulla, L.A., Jr., Genomic imprinting: the influence of differential methylation in the two sexes, *Exp. Biol. Med.* (Maywood), 2010, vol. 235, no. 2, pp. 139–147. doi 10.1258/ebm.2009.009251
- Weimin, C. and Jiang, L., Programming and inheritance of parental DNA methylomes in vertebrates, *Physiol. Publ.*, 2015, vol. 30, no. 1, pp. 63–68. doi 10.1152/physiol.00037.2014
- Wu, H. and Zhang, Y., Reversing DNA methylation: mechanisms, genomics, and biological functions, *Cell*, 2014, vol. 156, pp. 45–68.
- 32. Cantão, I.H., Renato Borges Tesser, R.B., and Stumpp, T., An initial investigation of an alternative model to study rat primordial germ cell, *Epigenet. Reprogram. Biol. Proced. Online*, 2017, vol. 19. doi 10.1186/s12575-017-0058-1
- 33. Hyldig, S.M.W., Croxall, N., Contreras, D.A., et al., Epigenetic reprogramming in the porcine germ line *BMC, Dev. Biol.*, 2011, vol. 11. doi 10.1186/1471-213X-11-11
- Morgan, H.D., Sutherland, H.G., Martin, D.I., and Whitelaw, E., Epigenetic inheritance at the agouti locus in the mouse, *Nat. Genet.*, 1999, vol. 23, pp. 314–335.
- Rakyan, V.K., Chong, S., Champ, M.E., et al., Transgenerational inheritance of epigenetic states at the murine Axin(Fu) allele occurs after maternal and paternal transmission, *Proc. Natl. Acad. Sci. U.S.A.*, 2003, vol. 100, pp. 2538–2543. doi 10.1073/pnas.0436776100
- Vanyushin, B.F., Epigenetics today and tomorrow, *Russ. J. Genet.: Appl. Res.*, 2014, vol. 4, no. 3, pp. 168– 188.
- 37. Lees-Murdock, D.J. and Walsh, C.P., DNA methylation reprogramming in the germ line, *Epigenetics*, 2008, vol. 3, pp. 5–13.

- Sasaki, H. and Matsui, Y., Epigenetic events in mammalian germ-cell development: reprogramming and beyond, *Nat. Rev. Genet.*, 2008, vol. 9, pp. 129–140. doi 10.1038/nrg2295
- 39. Lanza, R.P., Cibelli, J.B., Faber, D., et al., Cloned cattle can be healthy and normal, *Science*, 2001, vol. 294, pp. 1893–1894. doi 10.1126/science.1063440
- 40. Palmieri, C., Loi, P., Ptak, G., and Della Salda, L., Review paper: a review of the pathology of abnormal placentae of somatic cell nuclear transfer clone pregnancies in cattle, sheep, and mice, *Vet. Pathol.*, 2008, vol. 45, pp. 865–880. doi 10.1354/vp.45-6-865
- 41. Law, J.A. and Jacobsen, S.E., Establishing, maintaining and modifying DNA methylation patterns in plants and animals, *Nat. Rev. Genet.*, 2010. vol. 11, pp. 204– 220. doi 10.1038/nrg2719
- 42. Meirelles, F.V., Bressan, F.F., Smith, L.C., et al., Cytoplasmatic inheritance, epigenetics and reprogramming DNA as tools in animal breeding, *Livest. Sci.*, 2014, vol. 166, pp. 199–205. doi 10.1016/j.livsci. 2014.05.024
- 43. Dean, W., Santos, F., Stojkovic, M., et al., Conservation of methylation reprogramming in mammalian development: aberrant reprogramming in cloned embryos, *Proc. Natl. Acad. Sci. U.S.A.*, 2001, vol. 98, no. 24, pp. 13734–13738. doi 10.1073/pnas.241522698
- 44. Couldrey, C. and Lee, R.S., DNA methylation patterns in tissues from mid-gestation bovine foetuses produced by somatic cell nuclear transfer show subtle abnormalities in nuclear reprogramming, *BMC Dev. Biol.*, 2010, Mar 7, pp. 10–27. doi 10.1186/1471-213X-10-27
- 45. Kiefer, H., Jouneau, L., Campion, E., et al., Altered DNA methylation associated with an abnormal liver phenotype in a cattle model with a high incidence of perinatal pathologies, *Sci. Rep.*, 2016, vol. 6, article number 38869. doi 10.1038/srep38869
- Li, Q., Li, N., Hu, X., et al., Genome-wide mapping of DNA methylation in chicken, *PLoS One*, 2011, vol. 6, no. 5. doi 10.1371/journal.pone.0019428
- 47. Hu, Y., Xu, H., Li, Z., et al., Comparison of the genome-wide DNA methylation profiles between fast-growing and slow-growing broilers, *PLoS One*, 2013, vol. 8, no. 2. doi 10.1371/journal.pone.0056411
- Zhang, W., Spector, T.D., Deloukas, P., et al., Predicting genome-wide DNA methylation using methylation marks, genomic position, and DNA regulatory elements, *Genome Biol.*, 2015, vol. 16, no. 1. doi 10.1186/s13059-015-0581-9
- Barrero, M.J., Boué, S., and Izpisúa Belmonte, J.C., Epigenetic mechanisms that regulate cell identity, *Cell Stem. Cell*, 2010, vol. 7, no. 5, pp. 565–570. doi 10.1016/j.stem.2010.10.009
- 50. Kiefer, J.C., Epigenetics in development, *Dev. Dyn.*, 2007, no. 236, pp. 1144–1156. doi 10.1002/dvdy. 21094
- Tost, J., DNA methylation: an introduction to the biology and the disease-associated changes of a promising biomarker, *Mol. Biotechnol.*, 2010, vol. 44, pp. 71–81. doi 10.1007/s12033-009-9216-2
- 52. Chikunov I.E. and Naboka, A.V., The role of DNA methylation in regulation of microRNAs differential

expression in cancerogenesis, *Nauka Yuga Ross.* (Vestn. Yuzhn. Nauchn. Tsentra), 2016, vol. 12, no. 4, pp. 50–56.

- Jang, H. and Serra, C., Nutrition, epigenetics, and diseases, *Clin. Nutr. Res.*, 2014, vol. 3, pp. 1–8. doi 10.7762/cnr.2014.3.1.1
- 54. Boddicker, R.L., Koltes, J.E., Fritz-Waters, E.R., et al., Genome-wide methylation profile following prenatal and postnatal dietary omega-3 fatty acid supplementation in pigs, *Anim. Genet.*, 2016. vol. 47, no. 6, pp. 658–671. doi 10.1111/age.12468
- 55. Hu, Y., Xu, H., Li, Z., et al., Comparison of the genome-wide DNA methylation profiles between fastgrowing and slow-growing broilers, *PLoS One*, 2013, vol. 8, no. 2. doi 10.1371/journal.pone.0056411
- Li, B., Carey, M., and Workman, J.L., The role of chromatin during transcription, *Cell*, 2007, vol. 128, pp. 707–719. doi 10.1016/j.cell.2007.01.015
- Zentner, G.E. and Henikoff, S., Regulation of nucleosome dynamics by histone modifications, *Nat. Struct. Mol. Biol.*, 2013, vol. 20, pp. 259–266. doi 10.1038/nbt.3248
- Kouzarides, T., Chromatin modifications and their function, *Cell*, 2007, vol. 128, pp. 693–705. doi 10.1016/j.cell.2007.02.005
- 59. Wang, H., Wang, L., Erdjument-Bromage, H., et al., Role of histone H2A ubiquitination in Polycomb silencing, *Nature*, 2004, vol. 431, pp. 873–878. doi 10.1038/nature02985
- Kao, C.F., Hillyer, C., Tsukuda, T., et al., Rad6 plays a role in transcriptional activation through ubiquitylation of histone H2B, *Genes Dev.*, 2004, vol. 18, pp. 184–195. doi 10.1101/gad.1149604
- Bannister, A.J. and Kouzarides, T., Regulation of chromatin by histone modifications, *Cell Res.*, 2011, vol. 21, pp. 381–395. doi 10.1038/cr.2011.22
- Cedar, H. and Bergman, Y., Linking DNA methylation and histone modification: patterns and paradigms, *Nat. Rev. Genet.*, 2009, vol. 10, pp. 295–304. doi 10.1038/nrg2540
- Bannister, A.J., Zegerman, P., Partridge, J.F., et al., Selective recognition of methylated lysine 9 on histone H3 by the HP1 chromo domain, *Nature*, 2001, vol. 410, pp. 120–124. doi 10.1038/35065138
- Morgan, H.D., Santos, F., Green, K., et al., Epigenetic reprogramming in mammals, *Hum. Mol. Genet.*, 2005, vol. 14, review issue 1, pp. R47–R58. doi 10.1093/hmg/ddi114
- 65. Cowell, I.G., Aucott, R., Mahadevaiah, S.K., et al., Heterochromatin, HP1 and methylation at lysine 9 of histone H3 in animals, *Chromosoma*, 2002, no. 111, pp. 22—36. doi 10.1007/s00412-002-0182-8
- 66. Lepikhov, K. and Walter, J., Differential dynamics of histone H3 methylation at positions K4 and K9 in the mouse zygote, *BMC Dev. Biol.*, 2004, vol. 4, no. 12. doi 10.1186/1471-213X-4-12
- Kourmouli, N., Jeppesen, P., Mahadevhaiah, S., et al., Heterochromatin and tri-methylated lysine 20 of histone H4 in animals, *J. Cell Sci.*, 2004, vol. 117, pp. 2491–2501. doi 10.1242/jcs.01238
- 68. Arney, K.L., Bao, S., Bannister, A.J., et al., Histone methylation defines epigenetic asymmetry in the

mouse zygote, Int. J. Dev. Biol., 2002, vol. 46, pp. 317-320.

- Jeltsch, A. and Jurkowska, R.J., Allosteric control of mammalian DNA methyltransferases—a new regulatory paradigm, *Nucleic Acids Res.*, 2016, vol. 44, no. 18, pp. 8556—8575. https://doi.org/10.1093/nar/ gkw723.
- Matoba, S., Liu, Y., Lu, F., et al., Embryonic development following somatic cell nuclear transfer impeded by persisting histone methylation, *Cell*, 2014, vol. 159, no. 4, pp. 884–895. doi 10.1016/j.cell.2014.09.055
- Santos, F., Zakhartchenko, V., Stojkovic, M., et al., Epigenetic marking correlates with developmental potential in cloned bovine preimplantation embryos, *Curr. Biol.*, 2003, vol. 13, pp. 1116–1121. https://doi.org/10.1016/S0960-9822(03)00419-6.
- 72. Maalouf, W.E., Alberio, R., and Campbell, K.H., Differential acetylation of histone H4 lysine during development of in vitro fertilized, cloned and parthenogenetically activated bovine embryos, *Epigenetics*, 2008, vol. 3, pp. 199–209. doi 10.4161/epi.3.4.6497
- Wu, X., Li, Y., Xue, L., et al., Multiple histone site epigenetic modifications in nuclear transfer and in vitro fertilized bovine embryos, *Zygote*, 2011, vol. 19, no. 1, pp. 31–45. doi 10.1017/S0967199410000328
- 74. Ho, L. and Crabtree, G.R., Chromatin remodelling during development, *Nature*, 2010, vol. 463, pp. 474– 484. doi 10.1038/nature08911
- Martin, D.M., Chromatin remodeling in development and disease: focus on CHD7, *PLoS Genet.*, 2010, vol. 6, no. 7. doi 10.1371/journal.pgen.1001010.0
- 76. Kress, C., Montillet, G., Jean, C., et al., Chicken embryonic stem cells and primordial germ cells display different heterochromatic histone marks than their mammalian counterparts, *Epigenet. Chromatin*, 2016, vol. 9. doi 10.1186/s13072-016-0056-6
- 77. Mercer, T.R. and Mattick, J.S., Structure and function of long noncoding RNAs in epigenetic regulation, *Nat. Struct. Mol. Biol.*, 2013, vol. 20, pp. 300–307. doi 10.1038/nsmb.2480
- Heard, E. and Martienssen, R., Transgenerational epigenetic inheritance: myths and mechanisms, *Cell*, 2014, vol. 157, pp. 95–109. doi 10.1016/j.cell. 2014.02.045
- Bartel, P.D., MicroRNAs: target recognition and regulatory functions, *Cell*, 2009, vol. 136, pp. 215–233. doi 10.1016/j.cell.2009.01.002
- Friedman, R.C., Farh, K.K.-H., Burge, C.B., and Bartel, D.P., Most mammalian mRNAs are conserved targets of microRNAs, *Genome Res.*, 2009, vol. 19, pp. 92–105. doi 10.1101/gr.082701.108
- Muers, M., RNA: genome-wide views of long noncoding RNAs, *Nat. Rev. Genet.*, 2011, vol. 12, pp. 742– 743. doi 10.1038/nrg3088
- Wang, X., Gu, Z., and Jiang, H., MicroRNAs in farm animals, *Animal*, 2013, vol. 7, pp. 1567–1575. doi 10.1017/S1751731113001183
- Lin, X., Luo, J., Zhang, L., et al., MiR-103 controls milk fat accumulation in goat (*Capra hircus*) mammary gland during lactation, *PLoS One*, 2013. doi 10.1371/journal.pone.0079258

- Romao, J., Jin, W., He, M., et al., MicroRNAs in bovine adipogenesis: genomic context, expression and function, *BMC Genomics*, 2014, vol. 15:137. doi 10.1186/1471-2164-15-137
- Sun, X., Haider Ali, M.S.S., and Moran, M., The role of interactions of long non-coding RNAs and heterogeneous nuclear ribonucleoproteins in regulating cellular functions, *Biochem. J.*, 2017, vol. 474, no. 17, pp. 2925–2935. doi 10.1042/BCJ20170280
- Vance, K.W. and Ponting, C.P., Transcriptional regulatory functions of nuclear long noncoding RNAs, *Trends Genet.*, 2014, vol. 30, pp. 348–355. doi 10.1016/j.tig.2014.06.001
- Mercer, T.R., Dinger, M.E., and Mattick, J.S., Long non-coding RNAs: insights into functions, *Nat. Rev. Genet.*, 2009, vol. 10, pp. 155–159. doi 10.1038/nrg2521
- Li, R., Zhu, H., and Luo, Y., Understanding the functions of long non-coding RNAs through their higherorder structures, *Int. J. Mol. Sci.*, 2016. doi 10.3390/ijms17050702
- Ulitsky, I. and Bartel, D.P., lincRNAs: genomics, evolution, and mechanisms, *Cell*, 2013, vol. 154, pp. 26– 46. doi 10.1016/j.cell.2013.06.020
- Kapusta, A. and Feschotte, C., Volatile evolution of long noncoding RNA repertoires: mechanisms and biological implications, *Trends Genet.*, 2014, vol. 30, pp. 439–452. doi 10.1016/j.tig.2014.08.004
- Devaux, Y., Zangrando, J., Schroen, B., et al., Long noncoding RNAs in cardiac development and ageing, *Nat. Rev. Cardiol.*, 2015, vol. 12, pp. 415–425. doi 10.1038/nrcardio.2015.55
- 92. Weikard, R., Hadlich, F., and Kuehn, C., Identification of novel transcripts and noncoding RNAs in bovine skin by deep next generation sequencing, *BMC Genomics*, 2013, vol. 14. doi 10.1186/1471-2164-14-789
- Billerey, C., Boussaha, M., Esquerre, D., et al., Identification of large intergenic non-coding RNAs in bovine muscle using next-generation transcriptomic sequencing, *BMC Genomics*, 2014, vol. 15:499. doi 10.1186/1471-2164-15-499
- 94. Ibeagha-Awemu, E.M., Li, R., and Dudemaine, P.-L., The long non-coding RNA transcriptome of the bovine mammary gland and potential regulatory roles in fatty acid synthesis, *Proceedings of the 6th International Symposium on Animal Functional Genomics*, Piacenza, 2015, p. 91. http://www.isafg2015.it/ISAFG2015_ PROCEEDINGS.pdf.
- 95. Li, G., Wu, Z., Li, X., et al., Biological role of microRNA-103 based on expression profile and target genes analysis in pigs, *Mol. Biol. Rep.*, 2011, vol. 38, no. 7, pp. 4777–4786. doi 10.1007/s11033-010-0615-z
- 96. Wang, J., Bian, Y., Wang, Z., et al., MicroRNA-152 regulates DNA methyltransferase 1 and is involved in the development and lactation of mammary glands in dairy cows, *PLoS One*, 2014, vol. 9. doi 10.1371/journal.pone.0101358
- Li, H., Zhang, Z., Zhou, X., et al., Effects of microRNA-143 in the differentiation and proliferation of bovine intramuscular preadipocytes, *Mol. Biol.*

Rep., 2011, vol. 38, pp. 4273–4280. doi 10.1007/s11033-010-0550-z

- Zhang, T., Zhang, X., Han, K., et al., Analysis of long noncoding RNA and mRNA using RNA sequencing during the differentiation of intramuscular preadipocytes in chicken, *PLoS One*, 2017, vol. 15. doi 10.1371/journal.pone.0172389
- 99. Zhang, T., Zhang, X., Han, K., et al., Genome-wide analysis of lncRNA and mRNA expression during differentiation of abdominal preadipocytes in the chicken, *G3* (Bethesda), 2017, vol. 7, no. 3, pp. 953– 966. doi 10.1534/g3.116.037069
- 100. Li, Z., Ouyang, H., Zheng, M., et al., Integrated analysis of long non-coding RNAs (LncRNAs) and mRNA expression profiles reveals the potential role of LncRNAs in skeletal muscle development of the chicken, *Front. Physiol.*, 2017, vol. 9. doi 10.3389/fphys.2016.00687
- 101. Morris, K.V. and Mattick, J.S., The rise of regulatory RNA, *Nat. Rev. Genet.*, 2014, vol. 15, pp. 423–437. doi 10.1038/nrg3722
- 102. Klattenhoff, C.A., Scheuermann, J.C., Surface, L.E., et al., Braveheart, a long noncoding RNA required for cardiovascular lineage commitment, *Cell*, 2013, vol. 152, pp. 570–583. doi 10.1016/j.cell.2013.01.003
- 103. Grote, P., Wittler, L., Hendrix, D., et al., The tissuespecific lncRNA Fendrr is an essential regulator of heart and body wall development in the mouse, *Dev. Cell.*, 2013, vol. 24, pp. 206–214. doi 10.1016/j.devcel.2012.12.012
- 104. Zhang, Y., Wang, J., Huang, S., et al., Systematic identification and characterization of chicken (*Gallus* gallus) ncRNAs, Nucleic Acids Res., 2009, vol. 37, pp. 6562–6574. doi 10.1093/nar/gkp704
- 105. Chen, Y.-C.A. and Aravin, A.A., Non-coding RNAs in transcriptional regulation, *Curr. Mol. Biol. Rep.*, 2015, vol. 1, pp. 10–18. doi 10.1007/s40610-015-0002-6
- 106. Morlando, M., Ballarino, M., Fatica, A., and Bozzoni, I., The role of long noncoding RNAs in the epigenetic control of gene expression, *Chem. Med. Chem.*, 2014, vol. 9, no. 3, pp. 505–510. doi 10.1002/cmdc.201300569
- 107. Grote, P. and Herrmann, B.G., Long noncoding RNAs in organogenesis: making the difference, *Trends Genet.*, 2015, vol. 31, no. 6, pp. 329–335. doi 10.1016/j.tig.2015.02.002
- 108. Fatica, A. and Bozzoni, I., Long non-coding RNAs: new players in cell differentiation and development, *Nat. Rev. Genet.*, 2014, vol. 15, no. 1, pp. 7–21. doi 10.1038/nrg3606
- 109. Knoll, M., Lodish, H.F., and Sun, L., Long non-coding RNAs as regulators of the endocrine system, *Nat. Rev. Endocrinol.*, 2015, vol. 11, no. 3, pp. 151–160. doi 10.1038/nrendo.2014.229
- 110. Engreitz, J.M., Ollikainen, N., and Guttman, M., Long non-coding RNAs: spatial amplifiers that control nuclear structure and gene expression, *Nat. Rev. Mol. Cell Biol.*, 2016, vol. 17, no. 12, pp. 756–770. doi 10.1038/nrm.2016.126
- 111. Huntzinger, E. and Izaurralde, E., Gene silencing by microRNAs: contributions of translational repression

and mRNA decay, *Nat. Rev. Genet.*, 2011, no. 12, pp. 99–110. doi 10.1038/nrg2936

- 112. Bergmann, J.H. and Spector, D.L., Long non-coding RNAs: modulators of nuclear structure and function, *Curr. Opin. Cell Biol.*, 2014, vol. 26, pp. 10–18. doi 10.1016/j.ceb.2013.08.005
- Muret, K., Klopp, C., Wucher, V., et al., Long noncoding RNA repertoire in chicken liver and adipose tissue, *Genet. Sel. Evol.*, 2017, vol. 10. doi 10.1186/s12711-016-0275-0
- 114. Li, Z., Liu, H., Jin, X., et al., Expression profiles of microRNAs from lactating and non-lactating bovine mammary glands and identification of miRNA related to lactation, *BMC Genomics*, 2012, vol. 27, no. 13. doi 10.1186/1471-2164-13-731
- 115. Alsaweed, M., Lai, C.T., Hartmann, P.E., et al., Human milk miRNAs primarily originate from the mammary gland resulting in unique miRNA profiles of fractionated milk, *Sci. Rep.*, 2016, vol. 6. doi 10.1038/srep20680
- 116. Wang, M., Moisá, S., Khan, M.J., et al., MicroRNA expression patterns in the bovine mammary gland are affected by stage of lactation, *J. Dairy Sci.*, 2012, vol. 95, pp. 6529–6535. doi 10.3168/jds.2012-574
- 117. Ji, Z., Wang, G., Xie, Z., et al., Identification of novel and differentially expressed microRNAs of dairy goat mammary gland tissues using Solexa sequencing and bioinformatics, *PLoS One*, 2012, vol. 7, no. 11. doi 10.1371/journal.pone.0049463
- 118. Li, Z., Lan, X., Guo, W., et al., Comparative transcriptome profiling of dairy goat microRNAs from dry period and peak lactation mammary gland tissues, *PLoS One*, 2012, vol. 7, no. 12. doi 10.1371/journal.pone.0052388
- 119. Li, H.M., Wang, C.M., Li, Q.Z., and Gao, X.J., MiR-15a decreases bovine mammary epithelial cell viability and lactation and regulates growth hormone receptor expression, *Molecules*, 2012, vol. 17, no. 10, pp. 12037—12048. doi 10.3390/molecules171012037
- 120. Wu, N., Gaur, U., Zhu, Q., et al., Expressed microRNA associated with high rate of egg production in chicken ovarian follicles, *Anim. Genet.*, vol. 48, pp. 205–216. doi 10.1111/age.12516
- 121. Kaya, K.D., Karakulah, G., Yakicier, C.M., et al., mESAdb: microRNA expression and sequence analysis database, *Nucleic Acids Res.*, 2011, vol. 39. doi 10.1093/nar/gkq1256
- 122. Mattick, J.S., The genetic signatures of noncoding RNAs, *PLoS Genet.*, 2009, vol. 5. doi 10.1371/journal.pgen.1000459
- 123. Imamura, T., Yamamoto, S., Ohgane, J., et al., Noncoding RNA directed DNA demethylation of *Sphk1* CpG island, *Biochem. Biophys. Res. Commun.*, 2004, vol. 322, pp. 593–600. doi 10.1016/j.bbrc.2004.07.159
- 124. Memczak, S., Jens, M., Elefsinioti, A., et al., Circular RNAs are a large class of animal RNAs with regulatory potency, *Nature*, 2013, vol. 495, pp. 333–338. doi 10.1038/nature11928
- 125. Bernstein, B.E., Birney, E., Dunham, I., et al., An integrated encyclopedia of DNA elements in the human genome, *Nature*, 2012, vol. 489, pp. 57–74. doi 10.1038/nature11247

- 126. Zhu, J., Adli, M., Zou, J.Y., et al., Genome-wide chromatin state transitions associated with developmental and environmental cues, *Cell*, 2013, vol. 152, pp. 642–654. doi 10.1016/j.cell.2012.12.033
- 127. Vandegehuchte, M.B., Lemiere, F., Vanhaecke, L., et al., Direct and transgenerational impact on *Daphnia magna* of chemicals with a known effect on DNA methylation, *Comp. Biochem. Physiol., Part C: Pharmacol., Toxicol. Endocrinol.*, 2009, vol. 151, pp. 278– 285. doi 10.1016/j.cbpc.2009.11.007
- 128. Kaminen-Ahola, N., Ahola, A., Maga, M., et al., Maternal ethanol consumption alters the epigenotype and the phenotype of offspring in a mouse model, *PLoS Genet.*, 2010, vol. 6, no. 1. doi 10.1371/journal.pgen.1000811
- 129. Fisinin, V.I., Shatskikh, E.V., Latypova, E.N., and Surai, P.F., Maternal effect in poultry farming—from vitamins to vitagens and epigenetics, *Ptitsa Ptitseprod.*, 2016, no. 1, pp. 29–33.
- Weaver, I.C., Cervoni, N., Champagne, F.A., et al., Epigenetic programming by maternal behavior, *Nat. Neurosci.*, 2004, vol. 7, no. 8, pp. 847–854. doi 10.1038/nn1276
- 131. Boyko, A., Kathiria, P., Zemp, F.J., et al., Transgenerational changes in the genome stability and methylation in pathogen-infected plants (virus-induced plant genome instability), *Nucleic Acids Res.*, 2007, vol. 35, pp. 1714–1725. doi 10.1093/nar/gkm029
- 132. Lang-Mladek, C., Popova, O., Kiok, K., et al., Transgenerational inheritance and resetting of stressinduced loss of epigenetic gene silencing in *Arabidopsis, Mol. Plant*, 2010, vol. 3, pp. 594–602. doi 10.1093/mp/ssq014
- 133. Anway, M.D., Cupp, A.S., Uzumcu, M., and Skinner, M.K., Epigenetic transgenerational actions of endocrine disruptors and male fertility, *Science*, 2005, vol. 308, pp. 1466–1469. doi 10.1126/science.1108190
- 134. Waterland, R.A. and Jirtle, R.L., Early nutrition, epigenetic changes at transposons and imprinted genes, and enhanced susceptibility to adult chronic diseases, *Nutrition*, 2004, vol. 20, pp. 63–68. doi 10.1016/j.nut.2003.09.011
- 135. Choi, S.W. and Friso, S., Epigenetics: a new bridge between nutrition and health, *Adv. Nutr.*, 2010, vol. 1, pp. 8–16. doi 10.3945/an.110.1004
- 136. Romao, J.M., Jin, W., He, M., et al., Altered microRNA expression in bovine subcutaneous and visceral adipose tissues from cattle under different diet, *PLoS One*, 2012, vol. 7. doi 10.1371/journal.pone.0040605
- 137. Li, R., Beaudoin, F., Zhao, X., et al., MicroRNAs are involved in bovine mammary gland response to dietary supplementation with sunflower oil, *Proceedings of 10th World Congress on Genetics Applied to Livestock Production*, Vancouva, BC, 2014, p. 36.
- 138. Braunschweig, M., Jagannathan, V., Gutzwiller, A., and Bee, G., Investigations on transgenerational epigenetic response down the male line in F2 pigs, *PLoS One*, 2012, vol. 7. doi 10.1371/journal.pone.0030583
- 139. Li, M., Wu, H., Luo, Z., et al., An atlas of DNA methylomes in porcine adipose and muscle tissues, *Nat. Commun.*, 2012, vol. 3. doi 10.1038/ncomms1854.3850

- 140. Baik, M., Vu, T.T.T., Piao, M.Y., and Kang, H.J., Association of DNA methylation levels with tissuespecific expression of adipogenic and lipogenic genes in longissimus dorsi muscle of Korean cattle Asian-Australas, J. Anim. Sci., 2014, vol. 27, pp. 1493–1498. doi 10.5713/ajas.2014.14283
- 141. Waterland, R.A. and Rached, M.-T., Developmental establishment of epigenotype: a role for dietary fatty acids?, *Scand. J. Food Nutr.*, 2006, vol. 50, pp. 21–26. doi 10.1080/17482970601066488
- 142. Davie, J.R., Inhibition of histone deacetylase activity by butyrate, *J. Nutr.*, 2003, vol. 133, suppl. 7, pp. 2485S-2493S.
- 143. Zaina, S., Dossing, K.B., Lindholm, M.W., and Lund, G., Chromatin modification by lipids and lipoprotein components: an initiating event in atherogenesis?, *Curr. Opin. Lipidol.*, 2005, vol. 16, pp. 549–553. doi 10.1097/01.mol.0000180165.70077.ee
- 144. Li, R., Beaudoin, F., Zhao, X., and Ibeagha-Awemu, E.M., Possible involvement of epigenetic modifying enzymes in the regulation of nutrient effect on bovine milk fat synthesis, *Proceedings of the CSAS-CMSA: Joint Annual Meeting*, Banff, Alberta, A.B., 2013, pp. 346–351.
- 145. Widiker, S., Karst, S., Wagener, A., and Brockmann, G.A.J., High-fat diet leads to a decreased methylation of the Mc4r gene in the obese BFMI and the lean B6 mouse lines, *Appl. Genet.*, 2010, vol. 51, no. 2, pp. 193–197. doi 10.1007/BF03195727
- 146. Shankar, S., Kumar, D., and Srivastava, R.K., Epigenetic modifications by dietary phytochemicals: implications for personalized nutrition, *Pharmacol. Ther.*, 2013, vol. 138, pp. 1–17. doi 10.1016/j. pharmthera.2012.11.002
- 147. Cai, D., Jia, Y., Song, H., et al., Betaine supplementation in maternal diet modulates the epigenetic regulation of hepatic gluconeogenic genes in neonatal piglets, *PLoS One*, 2014, vol. 9, no. 8. doi 10.1371/journal.pone.0105504
- 148. Anderson, O.S., Sant, K.E., and Dolinoy, D.C., Nutrition and epigenetics: an interplay of dietary methyl donors, one-carbon metabolism and DNA methylation, *J. Nutr. Biochem.*, 2012, vol. 23, pp. 853– 859. doi 10.1016/j.jnutbio.2012.03.003
- 149. Crider, K.S., Yang, T.P., Berry, R.J., and Bailey, L.B., Folate and DNA methylation: a review of molecular mechanisms and the evidence for folate's role, *Adv. Nutr.*, 2012, vol. 3, pp. 21–38. doi 10.3945/an. 111.000992
- 150. Karweina, D., Kreuzer-Redmer, S., Müller, U., et al., The zinc concentration in the diet and the length of the feeding period affect the methylation status of the ZIP4 zinc transporter gene in piglets, *PLoS One*, 2015. doi 10.1371/journal.pone.0143098
- 151. Martin, J.L., Vonnahme, K.A., Adams, et al., Effects of dam nutrition on growth and reproductive performance of heifer calves, *J. Anim. Sci.*, 2007, vol. 85, pp. 841–847. doi 10.2527/jas.2006-337
- 152. Sullivan, T.M., Micke, G.C., Greer, R.M., et al., Dietary manipulation of *Bos indicus* × *heifers* during gestation affects the reproductive development of their

heifer calves, *Reprod. Fertil. Dev.*, 2009, vol. 21, pp. 773–784. doi 10.1071/RD09004

- 153. Altmann, S., Murani, E., Schwerin, M., et al., Maternal dietary protein restriction and excess affects offspring gene expression and methylation of non-SMC subunits of condensin in liver and skeletal muscle, *Epigenetics*, 2012, vol. 7, pp. 239–252. doi 10.4161/epi. 7.3.19183
- 154. Altmann, S., Murani, E., Schwerin, M., et al., Dietary protein restriction and excess of pregnant German Landrace sows induce changes in hepatic gene expression and promoter methylation of key metabolic genes in the offspring, *J. Nutr. Biochem.*, 2013, vol. 24, pp. 484–495. doi 10.1016/j.jnutbio.2012.01.011
- 155. Cong, R., Jia, Y., Li, R., et al., Maternal low-protein diet causes epigenetic deregulation of HMGCR and CYP7α1 in the liver of weaning piglets, *J. Nutr. Biochem.*, 2012, vol. 23, pp. 1647–1654. doi 10.1016/j.jnutbio.2011.11.007
- 156. Huang, W., Zhang, X., Li, A., and Miao, X., Identification of differentially expressed genes between subcutaneous and intramuscular adipose tissue of Large White pig using RNA-seq, *Hereditas* (Beijing), 2017, vol. 39, no. 6, pp. 501–511. doi 10.16288/j.yczz.17-038
- 157. Basu, U., Romao, J.M., and Guan, L.L., Adipogenic transcriptome profiling using high throughput technologies, *J. Genomics*, 2013, vol. 1, pp. 22–28. doi 10.7150/jgen.3781
- 158. Li, R., Beaudoin, F., Zhao, X., et al., Effect of dietary supplementation with linseed oil on the miRNome profile of the bovine mammary gland, *J. Anim. Sci.*, 2014, vol. 92, suppl. 2, p. 490.
- 159. Rijnkels, M., Freeman-Zadrowski, C., Hernandez, J., et al., Epigenetic modifications unlock the milk protein gene loci during mouse mammary gland development and differentiation, *PLoS One*, 2013, vol. 8, no. 1. doi 10.1371/journal.pone.0053270
- 160. Jirtle, R.L. and Skinner, M.K., Environmental epigenomics and disease susceptibility, *Nat. Rev. Genet.*, 2007, vol. 8, pp. 253–262. doi 10.1038/nrg2045
- 161. Wang, X.S., Zhang, Y., He, Y.H., et al., Aberrant promoter methylation of the CD4 gene in peripheral blood cells of mastitic dairy cows, *Genet. Mol. Res.*, 2013, vol. 12, pp. 6228–6239. http://dx.doi.org/ 10.4238/2013.10.4238/2013.
- 162. Vanselow, J., Yang, W., Herrmann, J., et al., DNAremethylation around a STAT5-binding enhancer in the α S1-casein promoter is associated with abrupt shutdown of a α S1-casein synthesis during acute mastitis, *J. Mol. Endocrinol.*, 2006, vol. 37, pp. 463–477. doi 10.1677/jme.1.02131
- 163. Jin, W., Ibeagha-Awemu, E.M., Liang, G., et al., Transcriptome microRNA profiling of bovine mammary epithelial cells challenged with *Escherichia coli*, or *Staphylococcus aureus*, bacteria reveals pathogen directed microRNA expression profiles, *BMC Genomics*, 2014, vol. 15:181. doi 10.1186/1471-2164-247
- 164. Hata, T., Murakami, K., Nakatani, H., et al., Isolation of bovine milk-derived microvesicles carrying mRNAs and microRNAs, *Biochem. Biophys. Res. Commun.*,

2010, vol. 396, pp. 528–533. doi 10.1016/j.bbrc. 2010.04.135

- 165. Vegh, P., Foroushani, A.B.K., Magee, D.A., et al., Profiling microRNA expression in bovine alveolar macrophages using RNA-seq, *Vet. Immunol. Immunopathol.*, 2013, vol. 155, pp. 238–244. doi 10.1016/j.vetimm.2013.08.004
- 166. Bai, Y., Huang, J.-M., Liu, G., et al., A comprehensive microRNA expression profile of the backfat tissue from castrated and intact full-sib pair male pigs, *BMC Genomics*, 2014, vol. 15:47. doi 10.1186/1471-2164-15-47
- 167. Cai, Z., Zhang, L., Chen, M., et al., Castrationinduced changes in microRNA expression profiles in subcutaneous adipose tissue of male pigs, *J. Appl. Genet.*, 2014, vol. 55, pp. 259–266. doi 10.1007/s13353-014-0194-0
- 168. Singh, K., Erdman, R.A., Swanson, K.M., et al., Epigenetic regulation of milk production in dairy cows, J. Mammary Gland Biol. Neoplasia, 2010, vol. 5, pp. 101–112. doi 10.1007/s10911-010-9164-2
- 169. Lin, Q., Gao, Z., Alarcon, R.M., et al., A role of miR-27 in the regulation of adipogenesis, *FEBS J.*, 2009, vol. 276, pp. 2348–2358. doi 10.1111/j.1742-4658.2009.06967.x
- 170. Kim, S.Y., Kim, A.Y., Lee, H.W., et al., miR-27a is a negative regulator of adipocyte differentiation via suppressing PPAR [gamma] expression, *Biochem. Biophys. Res. Commun.*, 2010, vol. 392, pp. 323–328. doi 10.1016/j.bbrc.2010.01.012
- 171. Jablonka, E. and Raz, G., Transgenerational epigenetic inheritance: prevalence, mechanisms, and implications for the study of heredity and evolution, *Q. Rev. Biol.*, 2009, vol. 84, pp. 131–176. doi 10.1086/598822
- 172. Daxinger, L. and Whitelaw, E., Understanding transgenerational epigenetic inheritance via the gametes in

mammals, Nat. Rev. Genet., 2012, vol. 13, pp. 153-162. doi 10.1038/nrg3188

- 173. Slatkin, M., Epigenetic inheritance and the missing heritability problem, *Genetics*, 2009, vol. 182, pp. 845–850. doi 10.1534/genetics.109.102798
- 174. Varona, L., Munilla, S., Mouresan, E.F., et al., A Bayesian model for the analysis of transgenerational epigenetic variation, *G3* (Bethesda), 2015, vol. 5, pp. 477–485. doi 10.1534/g3.115.016725
- 175. Trerotola, M., Relli, V., Simeone, P., and Alberti, S., Epigenetic inheritance and the missing heritability, *Hum. Genomics*, 2015, vol. 28, pp. 9–17. doi 10.1186/s40246-015-0041-3
- 176. Guerrero-Bosagna, C. and Skinner, M.K., Environmentally induced epigenetic transgenerational inheritance of phenotype and disease, *Mol. Cell Endocrinol.*, 2012, vol. 6, pp. 3–8. doi 10.1016/j.mce.2011.10.004
- 177. Hill, W.G., Goddard, M.E., and Visscher, P.M., Data and theory point to mainly additive genetic variance for complex traits, *PLoS Genet.*, 2008, vol. 4, no. 2. doi 10.1371/journal.pgen.1000008
- 178. Gonzalez-Recio, O., Toro, M.A., and Bach, A., Past, present, and future of epigenetics applied to livestock breeding, *Front. Genet.*, 2015. doi 10.3389/fgene.2015.00305
- 179. Wang, X., Lan, X., Radunz, A.E., and Khatib, H., Maternal nutrition during pregnancy is associated with differential expression of imprinted genes and DNA methyltransferases in muscle of beef cattle offspring, J. Anim. Sci., 2015, vol. 93, pp. 35–40. doi 10.2527/jas.2014-8148
- 180. Zentner, G.E. and Henikoff, S., Epigenome editing made easy, *Nat. Biotechnol.*, 2015, vol. 33, pp. 606– 607. doi 10.1038/nbt.3248

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