= EXPERIMENTAL ARTICLES ===

Successional Changes in the Microbial Community of the Alkaline Lake Khilganta during the Dry Season

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Abstract—Microbial processes in a shallow, saline, alkaline Lake Khilganta (Southern Siberia) were studied during the dry season. During the drought, a crust was formed on the lake surface, where low rates of production processes were observed, with predominance of anoxygenic photosynthesis at 2.3 mg C/(dm³ day). The rates of microbial processes increased after short-term rains. During this period, a thin cyanobacterial mat was formed on the bottom, in which filamentous cyanbacteria *Geitlerinema* spp. predominated and the rate of oxygenic photosynthesis was up to 18 mg C/(dm³ day). Subsequent water evaporation and salinity increase resulted in altered community types and their activity. Red spots emerged on the mat surface, where anoxygenic prototrophic members of the genus *Ectothiorhodospira* predominated. Anoxygenic photosynthesis became the main production process in microbial mats, with the rate of 60 mg C/(dm³ day). At salinity increase to 200 g/L, the water remained in small depressions on the bottom, where extremophilic green algae *Dunaliella* sp. predominated, and the rate of oxygenic photosynthesis was 0.877 mg C/(dm³ day). These changes in the type and activity of microbial communities is an example of succession of microbial communities in Southern Siberia saline lakes during drought.

Keywords: saline lakes, dry period, community succession, rates of microbial processes, photosynthesis, sulfate reduction, methanogenesis

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Succession is one of the basic ecological concepts, implying a process of change in the species structure at a certain site resulting from the action of environmental factors. In the course of succession, apart from sequential change of species and species associations, the abiotic components also change, affecting organic matter accumulation, mineral transformation, and modification of environmental parameters (Zavarzin, 2007). Research on succession processes in microbial communities is presently concentrated on the species composition of these communities and relationships between community composition and environmental factors, while investigation of the functional activity in the course of succession is relatively uncommon (Dupraz and Visscher, 2005; Navarro et al., 2009; Podell et al., 2014).

The microbial community in a saline, alkaline Lake Khilganta (Southern Siberia) was investigated. Extensive research of this lake was carried out during the moist period, when its parameters were relatively stable (Zavarzin, 1993; Gorlenko et al., 1993; Gerasimenko et al., 2003; Kompantseva et al., 2005; Namsaraev, B.B. and Namsaraev, Z.B., 2007; Tsyrenova et al., 2011a, 2011b). Since 2000, droughts

occurred in the area of the lake location, resulting in its frequent drying up and the basin being filled only during short-termed rains. After the rain period, the basin became dry, with salinity increasing from 40 to at least 260 g/L in the course of several days (Namsaraev et al., 2010). Drastic changes of the water parameters in the lake enabled us to carry out research impossible at the stable state of the lake ecosystem and to investigate successional changes in the community. The goal of the present work was to investigate the functional activity of the Lake Khilganta microbial community in the course of succession processes caused by the alterations in lake water salinity.

MATERIALS AND METHODS

Subject of research. Lake Khilganta ($50^{\circ}42.535'$ N, $115^{\circ}06.086'$ E) is located in the steppe zone in the southeastern Transbaikal area, at the divide of Onon and Aga rivers, 668 m above the sea level, and 76 km south to the Aginskoe settlement. The lake is drainless, round, with inclined shores. The maximal water area during the moist period is up to 0.3 km², the greatest

water depth is 64 cm. The basin dries completely during the dry period.

Research techniques. The temperature, pH, and redox potential (Eh) were determined at the sampling site, as well as carbonate concentration (by titration) and salinity (refractometrically). The concentrations of chlorophyll a and bacteriochlorophyll a were determined by spectrophotometry (Namsaraev, 2009). The mineralogy of the bottom sediments was investigated using light microscopy (Namsaraev et al., 2010). The data on the weather and precipitation were obtained from the World Meteorological Organization database, Aginskoe weather station, WMOID 30859 (https://www.ncdc.noaa.gov/dataaccess/land-based-station-data/land-based-datasets). A moisture of the sediments was determined by drying at 105°C and weighing before and after drying (GOST 28268-89, 2009). The rates of photosynthesis and dark CO₂ fixation were determined in the samples supplemented with ¹⁴C-bicarbonate. The rates of sulfate reduction were measured using ³⁵S-labeled sulfate (Kuznetsov and Dubinina, 1989; Gorlenko et al., 1999), and the rate of lithotrophic methanogenesis was measured using ¹⁴C-labeled bicarbonate (Namsaraev et al., 1999). Carbon consumption for sulfate reduction and methanogenesis was calculated according to the known equations (Belvaev et al., 1981). The samples of water, microbial mats, crust, and bottom sediments in 20-mL glass vials were exposed for 12 h under in situ conditions of temperature and illumination. To determine the rate of photosynthesis, the samples were incubated under light, while for determination of the rates of dark CO₂ fixation, sulfate reduction, and methanogenesis they were incubated in the dark. To determine the rate of anoxygenic photosynthesis, the sample was supplemented with diuron. For calculation of production according to the known formula (Kuznetsov and Dubinina, 1989), only the carbon of dissolved bicarbonate was considered. This assumption was based on bicarbonate concentration exceeding that of carbonate and on expected slow redistribution of labeled bicarbonate between the natural and introduced bicarbonate. For determination of the rates of microbial processes in cvanobacterial mats and in the samples of green microalga Dunaliella sp., lake water collected at the time of sampling was added. The effect of salinity on the rates of production processes was studied in freshly collected mat samples, which were supplemented with lake water diluted with freshwater to desired concentration. The vials with crust samples were filled with freshwater in order to imitate rain precipitation, stored for 60 min until salt dissolution (final pH 8.18, salinity 50 g/L), supplemented with the radioisotope label, and incubated upon the lake bottom. For determination of the species composition of phototrophs, the samples were fixed with 4% formalin. For microscopy of the dry crust, the sample was homogenized on the slide, supplemented with 5% HCl for carbonate removal, and examined under a light microscope at ×400-1000 magnification. The following manuals were used for identification of cyanobacteria, green algae, and anoxygenic phototrophic bacteria, respectively: Komárek and Anagnostidis (2007), Dedusenko-Shchegoleva et al. (1959), and *Bergey's Manual of Systematic Bacteriology* (2005).

RESULTS

Climatic conditions and hydrochemistry of Lake Khilganta. The chemical composition, pH, and salinity of Lake Khilganta water depend on climatic conditions of the southeastern Transbaikal area. During moist years (before 1999), at the annual rainfall of up to 550 mm, the lake was filled with water of ~40-50 g/L salinity and pH 9.5–9.9, which belonged to the chloride-sulfate-sodium type. During dry years, including the sampling periods (259 and 152 mm in 2006 and 2007, respectively), the lake was usually completely dry. Water of saturated green color remained in small depressions at the lake bottom. In these depressions, salinity was 200–260 g/L, pH 7.2– 8.1, and the water was of the chloride-sulfate-sodium type. During dry years, water occurred in the lake during short-term rains and was evaporated after several days due to high temperature. This was observed in detail in August 2007, when water salinity increased from 43 to 260 g/L, while pH decreased from 9.5 to 7.99 during water evaporation, and bicarbonate concentration increased from 0.14 to 0.21 g/L. The results of hydrochemical analysis were published previously (Namsaraev et al., 2010).

Bottom sediments during the dry period. During the dry period, the lake was covered with a crust ~0.5 cm thick. At the crust surface, bloedite $(Na_2Mg(SO_4)_2 \cdot 4H_2O)$ and halite (NaCl) crystals precipitated. Humidity of the crust was 4.7% (2007). The crust prevented water evaporation and drying of the bottom sediments. Water content in the sediments increased to 26.66% immediately below the crust (0.5–2 cm) and decreased to 11.83% in the 55–70-cm horizon. The pH of the water extract from the sediments decreased with depth from 8.6 (0–3 cm) to 8.13 (70 cm). In 2006–2007, excavation revealed pore water at the depth of 35–80 cm. Pore water salinity was 128–155 g/L, pH was 7.1–7.54.

The phototrophic community during the dry period. Microscopy of the crust at the bottom of dry lake revealed the high diversity of the cyanobacterial morphotypes (*Chroococcus minutus, Leptolyngbya tenuis, Leptolyngbya woronichinii, Coleofasciculus chthonoplastes,* and *Phormidium molle*) and green algae (*Oocistis* sp. and *Dunaliella salina*) in the upper, most dry layer of the crust (up to 1 mm thick). The subsurface crust layer (3–4 mm) was softer and often had purple coloration. In this layer, the number of anoxygenic phototrophic bacteria increased, which belonged to the families *Ectothiorhodospiraceae (Ectothiorhodospira* sp.), *Chromatiaceae (Marichromatium* sp., *Allochromatium* sp., *Marichromatium* sp.) and to the purple nonsulfur bacteria (*Rhodobacter* sp. and *Rhod-*

Sample	Oxygenic photosynthesis	Anoxygenic photosynthesis	Dark CO ₂ fixation	Sulfate reduction	Lithotrophic methanogenesis
	mg C /(dm ³ day)			mg S/(dm ³ day)	$\mu L CH_4/(dm^3 day)$
Crust above bottom sediments (2006)	1.5	2.3	0.68	15.93	230
Bottom sediments below the crust (2006)	N.D.	N.D.	0.24-1.082	0.4–2.2	20-379.5
Sample with <i>Dunaliella</i> sp. (2006)	0.877	0.029	0.034	N.D.	N.D.
Lake water (2007, S* 50 g/L)	3.28	0.27	N.D.	N.D.	N.D.
Cyanobacterial mat (2007, S 100 g/L)	18	16.6	8.2	2.87	29.3
Purple mat (2007, S 170 g/L)	13.1	60.6	59.8	N.D.	N.D.
Bottom sediments below thin mats at the rain period (2007, S 50 g/L)	N.D.	N.D.	0.7	48.05	576
Cyanobacterial mats, moist period (1999)	57.6	328.4	2.6	N.D.	N.D.
Bottom sediments below the cyanobacterial mat, moist period (1999)	N.D.	N.D.	N.D.	1.05-8.9	0.37-1.53

 Table 1. Rates of production and terminal degradation processes in Lake Khilganta

* S indicates total salinity. N.D. - no data.

ovulum sp.). The concentrations of chlorophyll *a* and bacteriochlorophyll *a* in the crust were 448 and 106 mg/m², respectively. Microscopy of the upper crust layer revealed the ratio close to $\sim 1 : 1$ for the numbers of empty sheaths of filamentous cyanobacteria to trichome-containing ones. The share of cyanobacterial sheaths with the cells decreased significantly with depth (to $\sim 1 : 10$ in the subsurface crust layer). This was probably an indication of more pronounced decomposition of cyanobacterial cells in deeper, more humid layers than at the surface.

During short-term rains of the dry period, the lake basin was filled with water for a short time. A thin film of a cyanobacterial mat developed on the lake bottom during 2-3 days, detached from the bottom, and floated to the surface due to oxygen bubbles forming inside the mat. The predominant cyanobacterial genus in the mat was *Geitlerinema*, while members of the genera Nodularia and Oscillatoria were also present. While the concentration of chlorophyll a in cyanobacterial mats was $61-89 \text{ mg/m}^2$, bacteriochlorophyll a content was below detection limit. After water evaporation for several days, salinity in the lake reached ~ 100 g/L, and cyanobacterial mats precipitated to the bottom and gradually decomposed. Within the salinity range of ~100-150 g/L, spots of purple bacteria emerged on the mat surface, in which Ectothiorhodospira sp. predominated. Both bacteriochlorophyll a

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(443 mg/m²) and chlorophyll *a* (568 mg/m²) were detected. At the final stage of drying (salinity of 200–260 g/L), pools remained in some depressions of the lake bottom, where green alga *Dunaliella salina* predominated, which is incapable of forming structured mats.

Rates of production processes. The rate of oxygenic photosynthesis in the mat crust (2006) was 1.5 mg C/(dm³ day), while the rate of anoxygenic photosynthesis was 2.3 mg C/(dm³ day). The rate of dark CO₂ fixation in the dry crust was 0.68 mg C/(dm³ day).

The rate of oxygenic photosynthesis in the brine (2006), carried out mainly by *Dunaliella* sp., which predominated after evaporation of most water, was 0.877 mg C/(dm³ day), while the rate of anoxygenic photosynthesis was lower, 0.029 mg C/(dm³ day), and the rate of dark CO₂ fixation was 0.034 mg C/(dm³ day).

During short-term rains of the dry period (2007), the lake basin was filled with water for sometime. In the water of the lake (2007, salinity 50 g/L, pH 9.03), the rates of oxygenic and anoxygenic photosynthesis were 3.28 and 0.27 mg C/(dm³ day), respectively. Further water evaporation resulted in salinity reaching 100 g/L. The rates of oxygenic photosynthesis in cyanobacterial mats, anoxygenic photosynthesis, and



Fig. 1. Rates of production processes depending on salinity: oxygenic photosynthesis (1), anoxygenic photosynthesis (2), and dark CO_2 fixation (3). Cyanobacterial mat (graph with salinity values of 20-40-80-60-100 g/L) and purple mat (graphs with salinity values 90-110-130-150-170 g/L).

dark CO₂ fixation were 18, 16.6, and 8.2 mg C/(dm³ day), respectively. When salinity increased to 170 g/L, the rate of oxygenic photosynthesis decreased to 13.1 mg C/(dm³ day), while the rates of anoxygenic photosynthesis and dark CO₂ fixation increased sharply to 60.6 and 59.8 mg C/(dm³ day), respectively. The rate of dark CO₂ fixation in the silt below the mat was 0.76 mg C/(dm³ day).

The samples of cyanobacterial and purple mats collected at salinity of 100 and 170 g/L were incubated under conditions of different salinity. For this purpose, the brine from the lake was diluted with freshwater to desired salinity levels. Oxygenic photosynthesis was found to have a peak at 20–60 g/L salinity (46.6–50.4 mg C/(dm³ day)). Salinity increase to 80 and 170 g/L resulted in the rate of oxygenic photosynthesis decreasing to 28 and 13.1 mg C/(dm³ day), respectively. The trends for anoxygenic photosynthesis and dark CO₂ fixation were opposite. The rates of these processes were only 4.3–16 mg C/(dm³ day) at 20–80 g/L, while at salinity of 170 g/L these processes became predominant with the rates of 60.6 and 59.8 mg C/(dm³ day), respectively (Fig. 1).

Rates of the terminal decomposition processes. Microbial sulfate reduction was the dominant terminal decomposition process in the bottom sediments. The highest sulfate reduction rate was found in the crust, to 15.93 mg S/(dm³ day), which corresponded to organic matter consumption of 11.95 mg C/(dm³ day). In the crust, organic matter consumption via sulfate reduction exceeded the value for lithotrophic methanogenesis by two orders of magnitude.

Sulfate reduction was also the predominant decomposition process in the thin cyanobacterial mats developing during short-term rains. The rates of sulfate reduction and lithotrophic methanogenesis in the mat were 2.87 mg S/(dm³ day) and 29.3 μ L CH₄/(dm³ day), respectively (salinity at the sampling time was 100 g/L). In the mats, organic matter consumption via sulfate reduction was two orders of magnitude higher than via methanogenesis, 2.11 and 0.06 mg $C/(dm^3 day)$, respectively. The rates of the terminal processes of sulfate reduction and methanogenesis in the upper sediment layers below the cyanobacterial mat were 48.05 mg S/(dm³ day) and 576 μ L CH₄/(dm³ day), respectively. Carbon consumption via sulfate reduction and methanogenesis was 36 and 1.2 mg $C/(dm^3 day)$, respectively.

DISCUSSION

Our results made it possible to reconstruct successional changes of the functional activity of Lake Khilganta microbial community during the dry period. Evaporative concentration and increased water salinity after short-term rains resulted in changes of the types of microbial communities and the dominant pathways of carbon assimilation. After short-term rains in the dry period, the lake basin was filled with water with salinity \sim 30–50 g/L and pH \sim 9–9.5, and a thin cyanobacterial mat developed, in which filamentous cyanobacteria of the genus Geitlerinema predominated. The water gradually evaporated in the course of approximately 10 days, which resulted in its increasing salinity (Namsaraev et al., 2010). The highest rate of oxygenic photosynthesis, 18 mg C/($dm^3 day$), was observed at 100 g/L salinity. Further increase in salinity resulted in development of red spots on the surface of the cyanobacterial mat; anoxygenic phototrophic bacteria of the genus Ectothiorhodospira predominated in these areas. The rate of oxygenic photosynthesis decreased, and anoxygenic photosynthesis and dark CO_2 fixation became predominant at the rates of 60.6 and 59.8 mg C/($dm^3 day$), respectively. Their highest rates were observed at salinity ~ 170 g/L. Shifts in the types of photosynthetic dominant processes were accompanied by the changes in the ratio of chlorophyll a and bacteriochlorophyll a in microbial mats. Chlorophyll *a* predominated at 100 g/L salinity, while bacteriochlorophyll a, at 170 g/L. At ~200 g/L salinity the lake dried almost completely, with the water remaining the the surface only in isolated depressions, where salinity is 200-260 g/L and extremely halophilic eukaryotic microalga Dunaliella sp. predominated. Oxygenic photosynthesis was the dominant process at this stage, although its rate was relatively low, $0.877 \text{ mg C/(dm^3 day)}$. Further evaporation resulted in formation of a dry crust, in which anoxygenic photosynthesis predominated at the rate of 2.3 mg C/($dm^3 day$), which was somewhat higher than the rate of oxygenic photosynthesis.

Increased rates of microbial processes after shortterm rains plays an important role in the functioning of the community, since significant amounts of organic matter are produced in the lake basin during this period. Oxygenic photosynthesis was the dominant process in the thin cyanobacterial mats formed after the rains, and the total production at this period exceeded the rates of terminal decomposition processes. Total production was 42.8 mg C/($dm^3 day$), while carbon consumption via sulfate reduction and methanogenesis was 2.1 mg C/(dm^3 day). In the absence of rains, production processes were detected in the crust at the surface of the lake bottom, although their rates were an order of magnitude lower than the values for the rain period. Anoxygenic photosynthesis was the dominant process, as was indicated by the visually discernible purple layer below the crust. Total production was 4.5 mg C/(dm^3 day), while carbon consumption via the degradation processes was 12 mg $C/(dm^3 day)$, indicating predominance of the degradation processes in the intervals between rainfalls.

The highest rates of photosynthetic production in Lake Khilganta during the moist period, 386 mg C/(dm³ day) or 3.86 g C/(m² day) were two to three times lower than the values of 5-12 g C/(m² day) for Solar Lake in Egypt or those of up to 11 g C/(m² day) for the soda lakes of the East African rift zone (Melack and Kilham, 1974; Krumbein et al., 1977; Namsaraev et al., 2015). However, the rate of light-dependent CO₂ assimilation in Lake Khilngata mats and crusts was comparable to the rates of 0.15-0.29 g C/(m² day) reported for cyanobacterial and lichen crusts developing on soil in the Namib Desert and of up to 1 g C/(m² day) reported for the Central European steppe zone (Lange et al., 1994; Evans and Lange, 2001).

Sulfate reduction rate in Lake Khilganta was 0.4– 48.05 mg S/(dm³ day), which is comparable to the values of 0.128–13.5 mg S/(dm³ day) reported for the bottom sediments of the Kulunda Steppe hypersaline alkaline lakes and is considerably higher than the maximal sulfate reduction rate of 0.073 mg S/(dm³ day) revealed in the water column of the hypersaline alkaline Lake Mono (Oremland et al., 2004; Foti et al., 2007). The rate of lithotrophic methanogenesis in Lake Khilganta water and sediment samples was 0.37– 576 μ L CH₄/(dm³ day), which was also comparable to the rates of 0.3–75.6 μ L CH₄/(dm³ day) reported for other saline and soda lakes in Mongolia and Transbaikalia (Namsaraev et al., 2015).

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REFERENCES

Belyaev, S.S., Lein, A.Yu., and Ivanov, M.V., Role of methanogenic and sulfate-reducing bacteria in organic matter decomposition, *Geokhimiya*, 1981, no. 3, pp. 437–445.

Bergey's Manual of Systematic Bacteriology, vol. 2: The Proteobacteria, George, M.G., Ed., Berlin: Springer, 2005.

Dedusenko-Shchegoleva, N.T., Matvienko, A.M., and Shkorobatov, L.A., *Opredelitel' presnovodnykh vodoroslei SSSR. Vyp. 8. Zelenye vodorosli. Klass vol'voksovye* (Identification Manual of Freshwater Algae of the USSR, no. 8. Green Algae. Class Volvocaceae), Moscow: AN SSSR, 1959.

Dupraz, C. and Visscher, P. T., Microbial lithification in marine stromatolites and hypersaline mat, *Trends Microbiol.*, 2005, vol. 13, no. 9, pp. 429–438.

Evans, R.D. and Lange, O.L., Biological soil crusts and ecosystem nitrogen and carbon dynamics, in *Biological Soil Crusts: Structure, Function, and Management*, Belnap, J. and Lange, O.L., Eds., Berlin: Springer, 2001, pp. 263–279.

Foti, M., Sorokin, D.Y., Lomans, B., Mussman, M., Zacharova, E.E., Pimenov, N.V., Kuenen, J.G., and Muyzer, G., Diversity, activity, and abundance of sulfate-reducing bacteria in saline and hypersaline soda lakes, *Appl. Environ. Microbiol.*, 2007, vol. 73, no. 7, pp. 2093–2100.

Gerasimenko, L.M., Mityushina, L.L., and Namsaraev, B.B., *Microcoleus* mats from alkaliphilic and halophilic communities, *Microbiology* (Moscow), 2003, vol. 72, no. 1, pp. 71–79.

Gorlenko, V.M., Zhilina, T.N., Namsaraev, B.B., Kulyrova, A.V., and Zavarzina, D.G., The activity of sulfatereducing bacteria in bottom sediments of soda lakes of the Southeastern Transbaikal region, *Microbiology* (Moscow), 1999, vol. 68, no. 5, pp. 580–585.

GOST (State Standard) 28268-89: Soils. Methods for Determination of Humidity, Maximal Hygroscopic Humidity, and Humidity of Stable Plant Wilting, 2006.

Komàrek, J. and Anagnostidis, K., Cyanoprokariota 2. Teil: Oscillatoriales, in *Süsswasserflora von Mitteleuropa*, Büdel, B., Gärtner, G., Krienitz, L., and Schagerl, M., Eds., 2007. Bd. 19/2.

Kompantseva, E.I., Sorokin, D.Yu., Gorlenko, V.M., and Namsaraev, B.B., The phototrophic community found in Lake Khilganta (an alkaline saline lake located in the Southeastern Transbaikal Region), *Microbiology* (Moscow), 2005, vol. 74, no. 3, pp. 352–361.

Krumbein, W.E., Cohen, Y., and Shilo, M., Solar Lake (Sinai). 4. Stromatolitic cyanobacterial mats, *Limnol. Oceanogr.*, 1977, vol. 22, no. 4, pp. 635–656.

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Kuznetsov, S.I. and Dubinina, G.A., *Metody izucheniya* vodnykh mikroorganizmov (Methods for Investigation of Aquatic Microorganisms), Moscow: Nauka, 1989.

Lange, O.L., Meyer, A., Zellner, H., and Heber, U., Photosynthesis and water relations of lichen soil crusts: field measurements in the coastal fog zone of the Namib Desert, *Funct. Ecol.*, 1994, vol. 8, no. 2, pp. 253–264.

Melack, J.M. and Kilham, P., Photosynthetic rates of phytoplankton in East African alkaline, saline lakes, *Limnol. Oceanogr.*, 1974, vol. 19, no. 5, pp. 743–755.

Namsaraev, B.B. and Namsaraev, Z.B., Microbial processes of the carbon cycle and environmental conditions in alkaline lakes of Transbaikalia and Mongolia, *Proc. Winogradsky Inst. Microbiol.*, Moscow: Nauka, 2007, no. 14, pp. 299–322.

Namsaraev, B.B., Zhilina, T.N., Gorlenko, V.M., and Kulyrova A.V., Bacterial methanogenesis in soda lakes of the Southeastern Transbaikal Region, *Microbiology* (Moscow), 1999, vol. 68, no. 5, pp. 586–591.

Namsaraev, Z.B., Application of extinction coefficients for quantification of chlorophylls and bacteriochlorophylls, *Microbiology* (Moscow), 2009, vol. 78, no. 6, pp. 794–797.

Namsaraev, Z.B., Gorlenko, V.M., Dulov, L.E., Sorokin, V.V., Buryukhaev, S.P., Barkhutova, D.D., Dambaev, V.B., and Namsaraev, B.B., Water regime and variations in hydrochemical characteristics of the soda salt Lake Khilganta (Southeastern Transbaikalia), *Water Res.*, 2010, no. 4, pp. 513–519.

Namsaraev, Z.B., Zaitseva, S.V., Gorlenko, V.M., Kozyreva, L.P., and Namsaraev, B.B., Microbial processes and factors controlling their activities in alkaline lakes of the Mongolian plateau, *Chinese J. Oceanol. Limnol.*, 2015, vol. 33, no. 6, pp. 1391–1401.

Navarro, J.B., Moser, D.P., Flores, A., Ross, C., Rosen, M.R., Dong, H., Zhang, G., and Hedlund, B.P., Bacterial succession within an ephemeral hypereutrophic Mojave Desert Playa Lake, *Microb. Ecol.*, 2009, vol. 57, no. 2, pp. 307–320.

Oremland, R.S., Stolz, J.F., and Hollibaugh, J.T., Microbial arsenic cycle in Mono Lake, California, *FEMS Microbiol. Ecol.*, 2004, vol. 48, pp. 15–27.

Podell, S., Emerson, J.B., Jones, C.M., Ugalde, J.A., Welch, S., Heidelberg, K.B., Banfield, J.F., and Allen, E.E., Seasonal fluctuations in ionic concentrations drive microbial succession in a hypersaline lake community, *The ISME J.*, 2014, vol. 8, no. 5, pp. 979–990.

Tsyrenova, D.D., Bryanskaya, A.V., Namsaraev, Z.B., and Akimov, V.N., Taxonomic and ecological characterization of cyanobacteria from some brackish and saline lakes of Southern Transbaikal Region, *Microbiology* (Moscow), 2011, vol. 80, no. 2, pp. 216–227.

Tsyrenova, D.D., Kozyreva, L.P., Namsaraev, B.B., Bryanskaya, A.V., and Namsaraev, Z.B., Structure and formation properties of the haloalkaliphilic community of Lake Khilganta, *Microbiology* (Moscow), 2011, vol. 80, no. 2, pp. 237–243.

Zavarzin, G.A., Genesis and development: evolution, succession, and haecceitas, *Herald Russ. Acad. Sci.*, 2007, vol. 77, no. 2, pp. 131–136.

Zavarzin, G.A., Epicontinental soda lakes as supposed relic biotopes for formation of terrestrial biota, *Mikrobiologiya*, 1993, vol. 62, pp. 789–800.

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