REVIEW

Changes in Retinal Glial Cells with Age and during Development of Age-Related Macular Degeneration

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> Received March 27, 2018 Revision received May 8, 2018

Abstract—Age is the major risk factor in the age-related macular degeneration (AMD) which is a complex multifactor neu rodegenerative disease of the retina and the main cause of irreversible vision loss in people over 60 years old. The major role in AMD pathogenesis belongs to structure-functional changes in the retinal pigment epithelium cells, while the onset and progression of AMD are commonly believed to be caused by the immune system dysfunctions. The role of retinal glial cells (Muller cells, astrocytes, and microglia) in AMD pathogenesis is studied much less. These cells maintain neurons and reti nal vessels through the synthesis of neurotrophic and angiogenic factors, as well as perform supporting, separating, trophic, secretory, and immune functions. It is known that retinal glia experiences morphological and functional changes with age. Age-related impairments in the functional activity of glial cells are closely related to the changes in the expression of troph ic factors that affect the status of all cell types in the retina. In this review, we summarized available literature data on the role of retinal macro- and microglia and on the contribution of these cells to AMD pathogenesis.

DOI: 10.1134/S000629791809002X

Keywords: aging, retina, age-related macular degeneration, astrocytes, Muller cells, microglia

Age-related macular degeneration (AMD) is a mul tifactor neurodegenerative disease of the retina that is the major cause of irreversible vision loss in people over 60 years old. Although age is the main risk factor of AMD, its incidence is also affected by genetic and ecological factors. According to clinical manifestations, AMD can occur in the dry or wet form. Dry AMD $(\sim)0\%$ of all cases) is characterized by the formation of drusen on the retina beneath the macula, pigment redistribution, and defects of the retinal pigment epithelium (RPE) and choroid capillaries. Wet (exudative) AMD develops in 10% patients and is characterized by neovascularization and growing of newly formed vessels under the RPE through the Bruch's membrane defects. Pathological per meability of newly formed vessels leads to macular edema, exudate secretion, bleeding into the vitreous body and retina and eventually results in vision loss [1, 2].

The incidence of AMD increases with the increasing population age. There is no efficient AMD treatment, partially because of insignificant knowledge of its patho genesis. Structure-functional changes of the retina in normal aging (loss of melanin granules by the RPE cells, impairments in folding and atrophy of microvilli, increase in the number and density of lipofuscin granules, drusen formation, compaction of the Bruch's membrane, sclero sis of choroid capillary walls, and narrowing of capillary lumen) are similar to changes observed at the early stages of AMD. These changes underlie the AMD pathogenesis but do not always lead to the disease development [3, 4]. The development of AMD and other neurodegenerative diseases is essentially contributed by age-related impair ments in the immune system functions and associated changes in homeostasis. AMD-affected retina is charac terized by an increased immune reactivity (i.e., tissue ability to develop immune response to any antigenic stim ulus) due to the formation of drusen – amorphous deposits between the RPE cells and external collagen zone of the Bruch's membrane mainly consisting of lipo fuscin [5]. Pathological changes in the RPE and choroid result in the death of retinal neurons that, in its turn, trig gers neuroinflammation. The integrity and homeostasis

Abbreviations: AMD, age-related macular degeneration; CX3CR1, C-X3-C-motif of chemokine receptor 1; GFAP, glial fibrillar acidic protein; IL, interleukin; RPE, retinal pigment epithelium; TNF, tumor necrosis factor.

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of neurons in the retina, as well in the entire central nerv ous system, are maintained in part by the glial cells [6]. Glial cells comprise only a small portion of the retina, but their influence on neurons, vascular cells, and other types of cells is very important. Using experimental models of eye diseases, it has been shown that experimentally induced or pathology-related dysfunctions of the glia lead to homeostasis disturbance and subsequent neuronal damage [7]. Glial cells are activated when the retina is damaged and releases various biologically active mole cules aimed at the tissue repair. Chronic activation of the glia is accompanied by an increased release of proinflam matory factors. Retinal glial cells are usually subdivided into macroglia (Muller cells and astrocytes) and microglia with specific morphological, physiological, and antigenic characteristics.

ASTROCYTES

Astrocytes play an important role in the development and functioning of the retina: they perform the neu rotrophic function, provide a mechanical support for damaged neurons, regulate the microenvironment, ensure optimal functioning of retinal neurons and vessels, and participate in the formation and maintenance of the blood-retinal barrier [7]. Astrocytes express on their plas ma membrane a large number of potassium channels thereby contributing to the removal of excessive extracel lular K^+ during active synaptic transmission and preventing depolarization and overexcitation of neurons. Moreover, astrocytes modulate synaptic plasticity of neu rons through regulating extracellular concentrations of neurotransmitters [8].

The bodies of astrocytes are located within the layer of nervous fibers, where they perform supplementary functions required for the development of the retinal vas cular network (figure, panels (a) and (b), II) [9]. During the embryonic development of the retina, astrocytes migrate from the optic nerve to the growing blood vessels. During the retina maturation in the postnatal period, there is a correlation between vascularization and spatial distribution of astrocytes, which are absent from the ves sel-free zones of the retina [10]. Astrocytes and Muller cells are involved in the regulation of local blood flow, i.e., its response to changes in the activity of neurons [11].

Astrogliosis (reactive astrocytosis) is an evolutionary conserved defense mechanism that involves activation of thousands of genes in a stress stimulus-dependent man ner. Activation of astrocytes and astrogliosis accompany the majority of neurodegenerative diseases of the brain and retina. The typical signs of astrogliosis are sharp increase of the astrocyte content (hyperplasia/prolifera tion), increase in the number and length of astroglial cell processes, cell body hypertrophy, migration and upregu lated expression of cytoskeleton components, such as

glial fibrillar acidic protein (GFAP), vimentin, and nestin [12]. Trauma or disease induce secretion by astrocytes of proteins capable of disturbing the integrity of the retinal brain barrier. These proteins upregulate expression of var ious genes encoding cytokines, chemokines, and ele ments of the complement cascade, thereby promoting degeneration of the retina [10, 13]. Activated retinal astrocytes express increased levels of matrix metallopro teinase 9 (MMP9) and urokinase type plasminogen acti vator (uPA). This has a negative effect on the extracellu lar matrix associated with the internal limiting membrane and promotes the loss of the retinal ganglion cells through apoptosis [14]. In wet AMD, diabetic retinopathy, and retinopathy of prematurity, astrocytes and Muller cells in the mature retina are involved in pathological neovascu larization by expressing proangiogenic factors in response to pathogenic stimuli [7]. Analysis of postmortem retinal samples from AMD patients by electron microscopy showed a large number of reactive and hypertrophic astrocytes that phagocytized dead cells in the ganglion cell layer. Together with Muller cells, these astrocytes formed "glial membranes" situated between the vitreous body and the internal limiting membrane [15].

MULLER CELLS

Muller cells are radial glial cells of the retina and have a bipolar morphology characteristic of the radial glia. Muller cells comprise 90% of all retinal glial cells. Muller cells originate from pluripotent precursors [6]. Although this process is studied insufficiently, it is known to involve activation of the Notch, Rax, and Janus-acti vated kinase (Jak) signaling pathways [16-19]. Muller cells are the only type of glial cells whose bodies are locat ed within the inner nuclear layer. Their processes pene trate all layers of the retina thereby promoting contacts between the neighboring neurons and participating in the formation of the external and internal limiting mem branes (figure, panels (a) and (b), II) [7]. It was shown that apoptosis of neurons and degeneration of the retina during the early postnatal development in mice occur only at the sites where the death of Muller cells was induced experimentally [20]. Unlike the retinas of bird and amphibians capable of photoreceptor regeneration, the retina of adult mammals has a limited ability for regeneration and is nearly incapable of *de novo* neuroge nesis [21]. A growing body of evidence obtained during the last few years showed that Muller cells are a hidden population of retinal stem cells [22]. A few first attempts have been made to pharmacologically stimulate regener ation and reprogramming of Muller cells into precursor cells capable of differentiating to the neuronal cells of the retina [23].

Muller cells maintain the viability of photoreceptors and neurons. They direct light onto photoreceptors, pro-

a) Changes in the state of retinal glial cell during AMD development. Central panel, healthy retina (II); left panel, classical variant of AMD development (I) including glia activation and migration into the inner retinal layers; right panel, AMD development on the background of immune system failure (III) with pronounced gliosis of Muller cells and astrocytes in the absence of adequate migration of macrophage cells. b) Changes in the retinal glia during AMD development in OXYS rats on the background of immune system failure. Immunohistochemical staining of the microglia (Iba1⁺ cells) (I); astrocytes and Muller cells (GFAP⁺ and Vimentin⁺ cells, respectively) (II) in the retina of Wistar rats without pathological changes; activated microglia (CD68⁺ cells) (III); migration of CD68⁺ macrophage cells (IV); gliosis of astrocytes (V) and Muller cells (VI) in the retina of OXYS rats during development of retinopathy (according to Telegina et al. [39], unpublished data). Scale, 50 μm; RPE, retinal pigment epithelium; GL, ganglion cell layer; IPL, inner plexiform layer; INL, inner nuclear layer; OPL, outer plexiform layer; ONL, outer nuclear layer; PhL, photosensory layer

vide structural stabilization of the retina, and modulate immune and inflammatory responses [24]. They perform a trophic function by supplying neurons and photorecep tors with nutrients, such as glucose, pyruvate, lactate, and amino acids [6]. Directly or indirectly, Muller cells main tain homeostasis of neurons by promoting production of trophic factors, antioxidants, cytokines, and growth fac tors, such as brain-derived neurotrophic factor (BDNF), basic fibroblast growth factor (bFGF), ciliary neu rotrophic factor (CNTF), insulin-like growth factor 1 (IGF1), glial neurotrophic factor (GDNF), leukemia-

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inhibiting factor (LIF), neurotrophin 3 (NT3), and pig ment epithelium-derived factor (PEDF) [25, 26]. Muller cells also express transporters and enzymes that capture and utilize neurotransmitters [27]; they can protect neu rons against oxidative stress via increasing secretion of antioxidants (glutathione, ferroxidase, and heme oxyge nase) [28]. Muller cells maintain the homeostasis of reti na microenvironment by regulating the extracellular medium ionic composition. Highly active neurons of the retina secret K^+ and water into the extracellular space, thereby promoting the influx of Na^+ and Ca^{2+} into the cells. Muller cells restore the physiological values of pH and ion concentrations by expressing on the plasma membrane potassium ion channels (in particular, Kir4.1) that deport K^+ ions from the regions with high K^+ concentration into the regions with low or stable K^+ concentration, such as subretinal space, vitreous body, and blood vessels [24]. Aquaporins (e.g., aquaporin 4) regulating water content in the retina function in a similar manner [29].

Activated Muller cells can either exhibit a neuropro tective effect or contribute to degenerative changes, thus inhibiting repair of the retinal neurons [12, 30].

GLIOSIS

Almost all pathogenic stimuli are able to activate retinal macroglia (astrocytes and Muller cells) and induce gliosis, i.e., reactive changes aimed at protection of the retina against further damages and maintenance of homeo stasis by increasing expression of cytoprotective factors, limiting retina remodeling, and restoring the balance between neuromediators, ions, and water [24]. Gliosis can be induced in the retina by degenerative processes, mechanical injury, inflammation, and/or aging [31]. The degree of gliosis-specific physiological, biochemical, and morphological changes depends on the damage severity. Unlike reactive gliosis, proliferative gliosis can accelerate neurodegeneration in chronic diseases of the retina, such as AMD, glaucoma, diabetic retinopathy, and pigmented retinitis, via direct and indirectly damage of neurons and vessels [4].

The specific feature of reactive gliosis is upregulated expression of intermediate filaments, such as vimentin, nestin, and GFAP [7]. The regulation of GFAP expres sion is so sensitive that it could be used as an index of reti nal stress and damage, as well as activation of Muller cells [32]. Among the retinal cells, Muller cells are the main subjects of gliosis that develops through the following stages: cell hypertrophy, loss of functionality, neuropro tection, inflammation, proliferation, and remodeling [33]. The majority of retinal diseases are associated with gliosis of Muller cells and astrocytes [3]. Gliosis of Muller cells becomes massive or proliferative with the damage of the blood-retinal barrier [28]. During proliferative gliosis, Muller cells can contribute to the death of neurons by promoting the synthesis and secretion of tumor necrosis factor (TNF), monocytic chemotactic type 1 protein (MCP-1 or Ccl-2), interleukins, interferon, intercellular adhesion molecule 1 (CD54), and nitric oxide (NO). Excessive synthesis of NO leads to the activation of free radical processes and protein nitrosylation, which are toxic for neurons [30].

Changes in the astrocyte morphology and biochem istry in the retina during reactive gliosis are similar to those in Muller cells and include increase in the expres-

sion of intermediate filament GFAP and vimentin (fig ure, panels (a) and (b), V and VI) [32, 34]. Similarly to Muller cells undergoing gliosis, reactive astroglia can expresses neurotrophic factors supporting cell viability or molecules inhibiting axon regeneration and repair via promoting neurotoxicity or secondary damage of adja cent neurons and glial cells [35]. AMD is characterized by a large amount of hypertrophic reactive astrocytes phago cytizing dead ganglion cells resulted from necrosis or apoptosis [36]. On the other hand, reactive gliosis is asso ciated with the regulation of enzymatic and non-enzy matic antioxidant defense systems that can increase the ability of astrocytes to protect neurons against free radi cals [37].

Studies in animal models of retinal diseases allowed to reveal changes in the glia at the early disease stages and to estimate their contribution to the development of neu rodegeneration. Thus, the retina detachment in GFAP and vimentin-deficient mice is characterized by lower levels of gliosis and monocyte migration leading to decreased cell death and prevention of photoreceptor degeneration [35]. Chemical inhibition of gliosis can pro tect ganglion cells from excitotoxicity [38]. In rds (retinal degradation slow) mice, an increase in the GFAP expres sion precedes neuronal apoptosis, whereas in rd (retinal degradation) mice, apoptosis of neurons was observed earlier than upregulation of expression of intermediate filaments [39]. Our studies showed that active AMD pro gression in senescence-accelerated OXYS rats was char acterized by increased GFAP levels in the retina and glio sis of Muller cells (figure, panels (a) and (b), V and VI) [40] but also by impairments in autophagy, activated pro grammed neurosis, and only a few cases of neuronal apoptosis [41]. At the same time, the number of TUNEL^+ cells at the preclinical symptom-free stage of the disease was higher and the level of the GFAP protein was lower in OXYS rats than in the control Wistar rats [40, 42].

Hence, retinal macroglia performs numerous func tions to provide and maintain normal homeostasis of neu rons and neural tissue as a whole. Any impairments in the morphology and functions of Muller cells and astrocytes increase the susceptibility of neurons to stress stimuli causing and/or aggravating the loss of retinal neurons and, resulting, as a consequence, in neurodegeneration development. Gliosis in the retina, on the one hand, is aimed at maintaining the viability of neurons; on the other hand, it can accelerate degenerative changes of neurons and AMD progression.

MICROGLIA

Microglia is the third type of retinal glial cells that includes residential macrophages of the central nervous system expressing many typical macrophage markers, such as CD11b, CD14, CD68, and EMR [43], as well as the

calcium-binding protein Iba1, a specific marker of the microglia [44]. The major function of the retinal microglia is participation in the immunity-mediated protective mechanisms. Microglial cells act as phagocytes and, together with perivascular cells, form a network of immune effector cells in the central nervous system [4]. Microglia activation is a common pathophysiological mechanism in the development of various degenerative diseases of the retina and often happens in parallel or before active death of neuronal cells [45]. Like other macrophages, brain and retinal microglial cells can have different phenotypes. Under physiological conditions, they are inactive and have a small body with branched processes. In the retina, microglial cells are located in the plexiform layers. They have a branched structure with small body and long processes capable of penetrating into the nuclear layers. Ramified microglia actively participate in the cell–cell interactions with neurons and macroglia, as well in phago cytosis and homeostasis maintenance in the retina [46]. When responding to injury, microglial cells acquire an ameba-like shape, migrate into the damaged region, and accumulate there. Activated microglial cells have a high ability for phagocytosis and express a number of pro- and anti-inflammatory molecules [47]. It has been shown that activated microglia can be neurotoxic and lead to the degeneration of photoreceptors thereby contributing (in combination with choroid macrophages) to chronic inflammation typical of some age-related diseases, includ ing AMD (figure, panels (a) and (b), III and IV) [48].

Microglia cells sense the environment through sur face receptors, including receptors of cytokines, chemokines, complement components, antibodies, etc. (for details, see review [49]). Normally, the activity of microglial cells is also regulated by a number of inhibito ry mechanisms. Thus, CX3CL1 chemokine, lectin CD22, and other membrane proteins, including CD200, CD47, and neuronal cell adhesion molecules, attenuate microglia activation [50]. An important role in this process belong to RPE cells, because transforming grow ing factor beta (TGFβ) secreted by these cells induces release of interleukin 10 (IL-10) from the microglia. IL- 10 downregulates expression of antigen-presenting pro teins CD80 and CD86 of the main histocompatibility complex II (MHCII) [49]. Analysis of the expression pro file of microglial genes revealed that TGFβ could con tribute to microglia transition to the anti-inflammatory phenotype [51].

To ensure optimal homeostasis in healthy retina, it is necessary to maintain constant level of neurotrophic fac tors responsible for the interaction between the microglia and Muller cells. Microglial cells are able to directly acti vate secretion by Muller cells of some neurotrophic fac tors, such as glia-derived neurotrophic factor (GDNF), leukemia-inhibiting factor (LIF), ciliary neurotrophic factor (CNTF), nerve growth factor (NGF), neu rotrophin 3 (NT-3), and basic fibroblast growth factor

(bFGF). These factors provide the preservation of pho toreceptors under stress and promote their recovery in the case of damage [49]. Activated Muller cells also secrete diazepam-binding inhibitor (DBI), a ligand of the 18-kDa translocator protein (TSPO). TSPO is expressed in the activated microglia and modulates microglial inflamma tion and phagocytosis via suppressing excessive activation of the microglia [52, 53]. Moreover, microglial cells acti vated as a result of retinal damage synthesize and secrete immature forms of neurotrophins (proBDNF and proNGF) that bind to the complex of $p75^{NTR}$ -receptor and sortilin and trigger cell apoptosis [54].

Microglia activation is also controlled by another important mechanism – direct physical interactions between the microglia and other retinal cells [10]. The CX3CL1 protein (fractalkine) is constitutively secreted by healthy retinal neurons and endothelial cells. It binds to the fractalkine receptor, the C-X3-C-motif of the chemokine receptor 1 (CX3CR1), on the microglia cells and thus prevents the neurotoxicity [55]. The CX3CR1 signaling pathway plays an important role in the regula tion, activation, and migration of microglial cells [56]. Under physiological conditions, fractalkine inhibits excessive activation of the microglia; however, during inflammation, fractalkine promotes activation of the microglia and astrocytes thereby promoting both neuro protective and neurotoxic effects of the microglia [57]. The CX3CR1 deficit inhibits the response of the microglia leading to neurotoxicity and degeneration [4].

Recent data show that age-associated alterations in the microglia include changes in the number and location of microglial cells in the retina, morphology of individual cells and their motility (decrease in the migration rate), and expression levels of some genes. The density of microglial cells in the outer and inner plexiform layers slightly increases (by \sim 19-21%) with aging (figure, panel (b), IV). Note that microglial cells are able to migrate from the inner to the outer plexiform layer and to accu mulate in the subretinal space [58]. Such redistribution results in the microglia contact with photoreceptors and RPE cells that induces in them secretion of immune fac tors specific for pathological AMD phenotypes (figure, panel (a)) [59, 60].

In addition to changes in cell morphology, the activ ity and reactivity of microglial changes also change with age. Many researchers have shown that during normal aging, microglial cells permanently exist in a state of moderate activation with upregulated expression of the main histocompatibility complex II (MHCII) compo nent CD11b and inflammatory cytokines (IL-1β, TNF, IL-6). This indicates that age-related specific changes in the microglia can promote para-inflammation [61] which increases the probability of neurodegenerative diseases characterized by chronic neuroinflammation.

Activation of microglial cells is a common feature of many retinal diseases, because microglial cells promptly respond to various triggers associated with apoptosis and neuronal degeneration. Although an exact stimulus responsible for the activation of microglial cells in neu rodegenerative diseases remains unknown, it has been shown that microglia activation and secretion of chemokines and TNF are preceded by astrogliosis, changes in the physiology of neurons, apoptosis of pho toreceptors, and degeneration of the retina [62]. At the first stages of neurodegeneration, the microglia trigger the repair mechanisms, for instance, formation of glial scar [3]. However, excessive or prolonged activation of the microglia can promote chronic inflammation with severe pathological side effects that can lead to the irreversible loss of neurons [45]. Activated microglial cells express a complex of neurotoxic and neurotrophic mediators. It is possible that changes in neurons and/or glial cells enhance the response of microglia that eventually influ ences the survival of neurons. On the other hand, dys function of the microglia and the loss of its protective functions (secretion of trophic factors, antioxidants, and cytokines, as well as phagocytosis) also can lead to neu ronal death [63].

Microglia activation in AMD is associated with changes in RPE cells and formation of drusen and amy loid plaques. If at the early AMD stages this activation is aimed at the removal of cell debris, including extracellu lar deposits of β-amyloid and drusen, pro-inflammatory cytokines induced by the microglia at the later stages con tribute to the development of neurodegeneration [3]. Patients with geographic atrophy of the retina have in the inner and outer nuclear layers ameba-like microglial cells that actively phagocytize dead photoreceptor cells [64].

It has been shown in many mouse AMD models that an increase in the number of microglial cells in the sub retinal space activates degenerative processes in the retina [65]. For instance, drusen-like deposits were observed in the RPE in CEP mice (mice immunized with car boxyethyl pyrrole (CEP)-conjugated albumin; model of geographic atrophy) [67]. In these animals, proinflam matory macrophages and the microglia are located between the RPE and external segments of photorecep tors. It was shown that the C–C-receptor of chemokine 2 (CCR2) is necessary for migration of immunocompetent cells and associated with the retinal damage, since no retinal damage development was observed in CEP-immu nized *CCR2*–/– mice that also contained no reactive phagocytes [68]. The number of macrophages and microglial cells is increased in the subretinal space of transgenic mice expressing regulator of the complement alternative pathway – complement factor H (CFH) with a single Y402H polymorphism. Mice of this strain display abnormalities similar to those observed in humans with AMD [69]. Accumulation of microglia in the subretinal space is observed in *Cx3cr1*–/– mice already at the age of 12 months [70]. In *Ccl2*-deficient mice, activated macrophages are accumulated in the subretinal space

[71]. Double knockout *Ccl2*–/–*Cx3cr1*–/– mice demon strate thickening of the Bruch's membrane, increase in the A2E levels, microglia infiltration, and atrophy of pho toreceptor cells [72]. These data show that chemokine deficiency leads to the development of AMD-like symp toms and suggest that dysfunction of phagocytes can play a key role in AMD pathogenesis [73].

Light-induced injury of the retina imitates photoox idative damages in AMD. Light promotes the develop ment of many diseases of the retina, as it has been con firmed by studies in animal models, in which retinal degeneration was accelerated by an increase in the inten sity of illumination, since the light-induced damage caused apoptosis more quickly than it occurred in AMD [62]. Despite the differences in the light intensity and duration of exposure, the injury of the retina is accompa nied by the loss of photoreceptors and increase in the reactivity and motility of microglial cells. Thus, it was shown that immediately after the exposure to light, the ameba-like microglial cells of the microglia rapidly translocate to the outer nuclear layer and the subretinal space. The density of phagocytizing microglia increased within two day after the exposure and then decreased to the initial level [74]. Interesting, even that ameba-like microglial cells acquired the branched shape 10 days later, they still exhibited the hidden immunophenotypical pro file of activation [74].

Therefore, microglial cells rapidly respond to various types of damage and degeneration of the retinal neurons by migrating into the nuclear layers and subretinal space that separates photoreceptors and RPE and lacks innate and adaptive immune cells under physiological condi tions [48]. Such changes in the distribution of immune cells in the external retina can lead to pathological changes in the immune environment of photoreceptors and RPE cells. As a result, degenerative changes in RPE cells cause a vicious circle promoting chronic inflamma tion in the retina and choroid. Age-related changes in the immune system contribute to this destructive process by changing the functions of immune cells [75, 76]. It should be noted that accumulation of microglia and macrophages in the subretinal space has been detected in histological retinal specimens from both AMD patients and elderly people without AMD [59].

Activation of retinal microglial cells, i.e., their tran sition into the ameba-like state and increased accumula tion in the subretinal space, is considered an important component of AMD pathogenesis. However, in the stud ies on OXYS rats, we found that the symptoms of AMD development are associated with a decrease in the phago cytic function (elimination of dead cells) of the microglia. We compared age-associated changes in the distribution of activated macrophages, microglia, and macroglia in different retinal layers in OXYS and Wistar (control) rats and found no migration of activated macrophages and microglial cells into the photoreceptor-containing layer

during manifestation of AMD-like clinical signs of retinopathy or during disease progression in OXYS rats (figure, panel (a)) [40]. Moreover, at the preclinical stage of the disease (at the age of 20 days), the number of acti vated macrophages in the retina of OXYS rats was lower than in the retina of control Wistar rats. Migration of acti vated macrophages and microglia into the inner layers of the neuroretina increased in the rats of both strains, but only in the OXYS rats, the levels of these cells were ele vated in the ganglion and inner nuclear layers and lowered in the plexiform layers. Based on the data obtained, we assumed that the retinopathy in OXYS rats develops in parallel to phagocytosis dysfunction and decreased elimi nation of dead cells [40]. We believe that these processes are determined by the imbalance in the immune respons es specific for OXYS rats, including inflammation, on the background of accelerated thymus involution and decrease in the activity of the T cell components of the immune system [77]. Indeed, an increasing number of studies indicate that genetically determined dysfunctions of the immune system and the associated changes in the inflammatory homeostasis can contribute to the AMD development and determine specific features of disease progression and organism's response to therapy [77-80].

Although the paradigm on the secondary role of glia is outdated, the contribution of changes in the glial cells to AMD pathogenesis is still insufficiently studied. However, the functions of these cells determine the inter actions of retinal neurons and adequate immune respons es to external factors, stress, and aging. Impairments in the phagocytic functions of the microglia lead to accu mulation of damaged cells, whereas gliosis of Muller cells and astrocytes decreases the bioavailability of neu rotrophic factors. The understanding of age-related changes in the state and reactivity of retinal glial cells is necessary for revealing causes and mechanisms of devel opment of age-related neurodegenerative diseases, including AMD. Moreover, recent publications suggest that glia can be a target for prophylaxis of degenerative changes in the retina and for recovery of retina functions in AMD.

Funding

The work was supported by the Russian Foundation for Basic Research (project no. 18-315-00216) and the Government of the Russian Federation (project no. 2017- 220-06-735576001).

REFERENCES

1. Shao, J., Choudhary, M. M., and Schachat, A. P. (2016) Neovascular age-related macular degeneration, in *Retinal Pharmacotherapeutics*, Karger Publishers, Vol. 55, pp. 125-136.

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- 2. Telegina, D. V., Kozhevnikova, O. S., and Kolosova, N. G. (2017) Molecular mechanisms of cell death in retina during development of age-related macular degeneration, *Adv. Gerontol*., **7**, 17-24.
- 3. Ardeljan, D., and Chan, C. C. (2013) Aging is not a disease: distinguishing age-related macular degeneration from aging, *Prog. Retin. Eye Res*., **37**, 68-89.
- 4. Cuenca, N., Fernandez-Sanchez, L., Campello, L., Maneu, V., De la Villa, P., Lax, P., and Pinilla, I. (2014) Cellular responses following retinal injuries and therapeutic approaches for neurodegenerative diseases, *Prog. Retin. Eye Res.*, **43**, 17-75.
- 5. Bora, N. S., Matta, B., Lyzogubov, V. V., and Bora, P. S. (2015) Relationship between the complement system, risk factors and prediction models in age-related macular degeneration, *Mol. Immunol.*, **63**, 176-183.
- 6. Goldman, D. (2014) Muller glial cell reprogramming and retina regeneration, *Nat. Rev. Neurosci.*, **15**, 431-442.
- 7. Coorey, N. J., Shen, W., Chung, S. H., Zhu, L., and Gillies, M. C. (2012) The role of glia in retinal vascular dis ease, *Clin. Exp. Optometry*, **95**, 266-281.
- 8. Rossi, D. (2015) Astrocyte physiopathology: at the cross roads of intercellular networking, inflammation and cell death, *Prog. Neurobiol.*, **130**, 86-120.
- 9. Kur, J., Newman, E. A., and Chan-Ling, T. (2012) Cellular and physiological mechanisms underlying blood flow regu lation in the retina and choroid in health and disease, *Prog. Retin. Eye Res.*, **31**, 377-406.
- 10. Vecino, E., Rodriguez, F. D., Ruzafa, N., Pereiro, X., and Sharma, S. C. (2016) Glia–neuron interactions in the mammalian retina, *Prog. Retin. Eye Res.*, **51**, 1-40.
- 11. Newman, E. A. (2015) Glial cell regulation of neuronal activity and blood flow in the retina by release of gliotrans mitters, *Phil. Trans. R. Soc. B*, **370**, 20140195.
- 12. De Hoz, R., Rojas, B., Ramirez, A. I., Salazar, J. J., Gallego, B. I., Trivino, A., and Ramirez, J. M. (2016) Retinal macroglial responses in health and disease, *BioMed. Res. Int.*, **2016**, 2954721.
- 13. Kim, J. H., Kim, J. H., Park, J., Lee, S. W., Kim, W. J., Yu, Y. S., and Kim, K. W. (2006) Blood–neural barrier: inter cellular communication at glio-vascular interface, *J. Biochem. Mol. Biol.*, **39**, 339-345.
- 14. Zhang, X., Cheng, M., and Chintala, S. K. (2004) Kainic acid-mediated upregulation of matrix metalloproteinase-9 promotes retinal degeneration, *Invest. Ophthalm. Vis. Sci.*, **45**, 2374-2383.
- 15. Ramirez, J. M., Ramirez, A. I., Salazar, J. J., de Hoz, R., and Trivino, A. (2001) Changes of astrocytes in retinal age ing and age-related macular degeneration, *Exp. Eye Res.*, **73**, 601-615.
- 16. Jadhav, A. P., Cho, S. H., and Cepko, C. L. (2006) Notch activity permits retinal cells to progress through multiple progenitor states and acquire a stem cell property, *Proc. Natl. Acad. Sci. USA*, **103**, 18998-19003.
- 17. Jadhav, A. P., Roesch, K., and Cepko, C. L. (2009) Development and neurogenic potential of Muller glial cells in the vertebrate retina, *Prog. Retin. Eye Res.*, **28**, 249- 262.
- 18. Goureau, O., Do Rhee, K., and Yang, X. J. (2004) Ciliary neurotrophic factor promotes Muller glia differentiation from the postnatal retinal progenitor pool, *Dev. Neurosci.*, **26**, 359-370.
- 19. Bhattacharya, S., Das, A. V., Mallya, K. B., and Ahmad, I. (2008) Ciliary neurotrophic factor-mediated signaling regu lates neuronal versus glial differentiation of retinal stem cell/progenitors by concentration-dependent recruitment of mitogens-activated protein kinase and Janus kinase-signal transducer and activator of transcription pathways in con junction with Notch signaling, *Stem Cells*, **26**, 2611-2624.
- 20. Dubois-Dauphin, M., Poitry-Yamate, C., De Bilbao, F., Julliard, A. K., Jourdan, F., and Donati, G. (1999) Early postnatal Muller cell death leads to retinal but not optic nerve degeneration in NSE-Hu-Bcl-2 transgenic mice, *Neuroscience*, **95**, 9-21.
- 21. Xia, X., and Ahmad, I. (2016) Unlocking the neurogenic potential of mammalian Muller glia, *Int. J. Stem Cells*, **9**, 169-175.
- 22. Hamon, A., Roger, J. E., Yang, X. J., and Perron, M. (2016) Muller glial cell-dependent regeneration of the neu ral retina: an overview across vertebrate model systems, *Develop. Dynam*., **245**, 727-738.
- 23. Webster, M. K., Cooley-Themm, C., Barnett, J. D., Bach, H. B., Vainner, J. M., Webster, S. E., and Linn, C. L. (2017) Evidence of BrdU-positive retinal neurons after application of an Alpha7 nicotinic acetylcholine receptor agonist, *Neuroscience*, **346**, 437-446.
- 24. Bringmann, A., and Wiedemann, P. (2012) Muller glial cells in retinal disease, *Ophthalmologica*, **227**, 1-19.
- 25. Gallina, D., Todd, L., and Fischer, A. J. (2014) A compar ative analysis of Muller glia-mediated regeneration in the vertebrate retina, *Exp. Eye Res.*, **123**, 121-130.
- 26. Kolomeyer, A. M., and Zarbin, M. A. (2014) Trophic fac tors in the pathogenesis and therapy for retinal degenerative diseases, *Survey Ophthalmol.*, **59**, 134-165.
- 27. Hurley, J. B., Chertov, A. O., Lindsay, K., Giamarco, M., Cleghorn, W., Du, J., and Brockerhoff, S. (2014) Energy metabolism in the vertebrate retina, in *Vertebrate Photoreceptors*, Springer, Japan, pp*.* 91-137.
- 28. Reichenbach, A., and Bringmann, A. (2013) New func tions of Muller cells, *Glia*, **61**, 651-678.
- 29. Schey, K. L., Wang, Z., Wenke, J. L., and Qi, Y. (2014) Aquaporins in the eye: expression, function, and roles in ocular disease, *Biochim. Biophys. Acta*, **1840**, 1513-1523.
- 30. Hippert, C., Graca, A. B., Barber, A. C., West, E. L., Smith, A. J., Ali, R. R., and Pearson, R. A. (2015) Muller glia acti vation in response to inherited retinal degeneration is high ly varied and disease-specific, *PLoS One*, **10**, e0120415.
- 31. Luna, G., Lewis, G. P., Banna, C. D., Skalli, O., and Fisher, S. K. (2010) Expression profiles of nestin and syne min in reactive astrocytes and Muller cells following retinal injury: a comparison with glial fibrillar acidic protein and vimentin, *Mol. Vis.*, **16**, 2511-2523.
- 32. Belecky-Adams, T. L., Chernoff, E. C., Wilson, J. M., and Dharmarajan, S. (2013) Reactive Muller glia as potential retinal progenitors, in *Neural Stem Cells* – *New Perspectives*, InTech.
- 33. Hol, E. M., and Pekny, M. (2015) Glial fibrillary acidic protein (GFAP) and the astrocyte intermediate filament system in diseases of the central nervous system, *Curr. Opin. Cell Biol.*, **32**, 121-130.
- 34. Nakazawa, T., Takeda, M., Lewis, G. P., Cho, K. S., Jiao, J., Wilhelmsson, U., Fisher, S. K., Pekny, M., Chen, D. F., and Miller, J. W. (2007) Attenuated glial reactions and pho toreceptor degeneration after retinal detachment in mice

deficient in glial fibrillary acidic protein and vimentin, *Invest. Ophthalmol. Vis. Sci.*, **48**, 2760-2768.

- 35. Edwards, M. M., McLeod, D. S., Bhutto, I. A., Villalonga, M. B., Seddon, J. M., and Lutty, G. A. (2016) Idiopathic preretinal glia in aging and age-related macular degenera tion, *Exp. Eye Res.*, **150**, 44-61.
- 36. Verkhratsky, A., Rodriguez, J. J., and Parpura, V. (2014) Neuroglia in ageing and disease, *Cell Tissue Res.*, **357**, 493-503.
- 37. Ganesh, B. S., and Chintala, S. K. (2011) Inhibition of reactive gliosis attenuates excitotoxicity-mediated death of retinal ganglion cells, *PloS One*, **6**, e18305.
- 38. Kalloniatis, M., Nivison-Smith, L., Chua, J., Acosta, M. L., and Fletcher, E. L. (2016) Using the rd1 mouse to understand functional and anatomical retinal remodelling and treatment implications in retinitis pigmentosa: a review, *Exp. Eye Res.*, **150**, 106-121.
- 39. Telegina, D. V., Kozhevnikova, O. S., Bayborodin, S. I., and Kolosova, N. G. (2017) Contributions of age-related alterations of the retinal pigment epithelium and of glia to the AMD-like pathology in OXYS rats, *Sci. Rep.*, **7**, 41533.
- 40. Kozhevnikova, O. S., Telegina, D. V., Devyatkin, V. A., and Kolosova, N. G. (2018) Involvement of the autophagic path way in the progression of AMD-like retinopathy in senes cence-accelerated OXYS rats, *Biogerontology*, **19**, 223-235.
- 41. Telegina, D. V., Korbolina, E. E., Ershov, N. I., Kolosova, N. G., and Kozhevnikova, O. S. (2015) Identification of functional networks associated with cell death in the retina of OXYS rats during the development of retinopathy, *Cell Cycle*, **14**, 3544-3556.
- 42. Kettenmann, H., Hanisch, U. K., Noda, M., and Verkhratsky, A. (2011) Physiology of microglia, *Physiol. Rev.*, **91**, 461-553.
- 43. Ohsawa, K., Imai, Y., Sasaki, Y., and Kohsaka, S. (2004) Microglia/macrophage-specific protein Iba1 binds to fibrin and enhances its actin-binding activity, *J. Neurochem.*, **88**, 844-856.
- 44. Langmann, T. (2007) Microglia activation in retinal degen eration, *J. Leukoc. Biol.*, **81**, 1345-1351.
- 45. Nimmerjahn, A., Kirchhoff, F., and Helmchen, F. (2005) Resting microglial cells are highly dynamic surveillants of brain parenchyma *in vivo*, *Science*, **308**, 1314-1318.
- 46. Fu, R., Shen, Q., Xu, P., Luo, J. J., and Tang, Y. (2014) Phagocytosis of microglia in the central nervous system dis eases, *Mol. Neurobiol.*, **49**, 1422-1434.
- 47. Xu, H., Chen, M., and Forrester, J. V. (2009) Para-inflamma tion in the aging retina, *Progr. Retin. Eye Res.*, **28**, 348-368.
- 48. Karlstetter, M., Scholz, R., Rutar, M., Wong, W. T., Provis, J. M., and Langmann, T. (2015) Retinal microglia: just bystander or target for therapy? *Prog. Retin. Eye Res.*, **45**, 30-57.
- 49. Perry, V. H., and Teeling, J. (2013) Microglia and macrophages of the central nervous system: the contribution of microglia priming and systemic inflammation to chronic neurodegeneration, *Semin. Immunopathol*., **35**, 601-612.
- 50. Paglinawan, R., Malipiero, U., Schlapbach, R., Frei, K., Reith, W., and Fontana, A. (2003) TGFβ directs gene expression of activated microglia to an anti-inflammatory phenotype strongly focusing on chemokines genes and cell migratory genes, *Glia*, **44**, 219-231.
- 51. Karlstetter, M., Nothdurfter, C., Aslanidis, A., Moeller, K., Horn, F., Scholz, R., Neumann, H., Weber, B. H., Rupprecht, R., and Langmann, T. (2014) Translocator pro tein (18 kDa) (TSPO) is expressed in reactive retinal

BIOCHEMISTRY (Moscow) Vol. 83 No. 9 2018

microglia and modulates microglial inflammation and phagocytosis, *J. Neuroinflamm.*, **11**, 3.

- 52. Wang, M., Wang, X., Zhao, L., Ma, W., Rodriguez, I. R., Fariss, R. N., and Wong, W. T. (2014) Macroglia-microglia interactions via TSPO signaling regulates microglial activa tion in the mouse retina, *J. Neurosci.*, **34**, 3793-3806.
- 53. Kimura, A., Namekata, K., Guo, X., Harada, C., and Harada, T. (2016) Neuroprotection, growth factors and BDNF-TrkB signalling in retinal degeneration, *Int. J. Mol. Sci.*, **17**, E1584.
- 54. Cardona, A. E., Pioro, E. P., Sasse, M. E., Kostenko, V., Cardona, S. M., Dijkstra, I. M., Huang, D., Kidd, G., Dombrowski, S., Dutta, R., Lee, J. C., Cook, D. N., Jung, S., Lira, S. A., Littman, D. R., and Ransohoff, R. M. (2006) Control of microglial neurotoxicity by the fractalkine receptor, *Nat. Neurosci.*, **9**, 917-924.
- 55. Liang, K. J., Lee, J. E., Wang, Y. D., Ma, W., Fontainhas, A. M., Fariss, R. N., and Wong, W. T. (2009) Regulation of dynamic behavior of retinal microglia by CX3CR1 signal ing, *Invest. Ophthalmol. Vis. Sci.*, **50**, 4444-4451.
- 56. Sheridan, G. K., and Murphy, K. J. (2013) Neuron–glia crosstalk in health and disease: fractalkine and CX3CR1 take centre stage, *Open Biol.*, **3**, 130181.
- 57. Damani, M. R., Zhao, L., Fontainhas, A. M., Amaral, J., Fariss, R. N., and Wong, W. T. (2011) Age-related alterations in the dynamic behavior of microglia, *Aging Cells*, **10**, 263-276.
- 58. Ma, W., Coon, S., Zhao, L., Fariss, R. N., and Wong, W. T. (2013) A2E accumulation influences retinal microglial activation and complement regulation, *Neurobiol. Aging*, **34**, 943-960.
- 59. Ma, W., and Wong, W. T. (2016) Aging changes in retinal microglia and their relevance to age-related retinal disease, *Adv. Exp. Med. Biol.*, **854**, 73-78.
- 60. Medzhitov, R. (2008) Origin and physiological roles of inflammation, *Nature*, **454**, 428-435.
- 61. Karlstetter, M., Ebert, S., and Langmann, T. (2010) Microglia in the healthy and degenerating retina: insights from novel mouse models, *Immunobiology*, **215**, 685-691.
- 62. Polazzi, E., and Monti, B. (2010) Microglia and neuropro tection: from *in vitro* studies to therapeutic applications, *Progr. Neurobiol.*, **92**, 293-315.
- 63. Gupta, N., Brown, K. E., and Milam, A. H. (2003) Activated microglia in human retinitis pigmentosa, late onset retinal degeneration, and age-related macular degen eration, *Exp. Eye Res.*, **76**, 463-471.
- 64. Luhmann, U. F., Lange, C. A., Robbie, S., Munro, P. M., Cowing, J. A., Armer, H. E., Luong, V., Carvalho, L. S., MacLaren, R. E., Fitzke, F. W., Bainbridge, J. W., and Ali, R. R. (2012) Differential modulation of retinal degeneration by Ccl2 and Cx3cr1 chemokine signaling, *PLoS One*, **7**, e35551.
- 65. Hollyfield, J. G., Bonilha, V. L., Rayborn, M. E., Yang, X., Shadrach, K. G., Lu, L., Ufret, R. L., Salomon, R. G., and Perez, V. L. (2008) Oxidative damage-induced inflamma tion initiates age-related macular degeneration, *Nat. Med.*, **14**, 194-198.
- 66. Hollyfield, J. G., Perez, V. L., and Salomon, R. G. (2010) A hapten generated from an oxidation fragment of docosa hexaenoic acid is sufficient to initiate age-related macular degeneration, *Mol. Neurobiol.*, **41**, 290-298.
- 67. Cruz-Guilloty, F., Saeed, A. M., Echegaray, J. J., Duffort, S., Ballmick, A., Tan, Y., Betancourt, M., Viteri, E.,

BIOCHEMISTRY (Moscow) Vol. 83 No. 9 2018

Ramkhellawan, G. C., Ewald, E., Feuer, W., Huang, D., Wen, R., Hong, L., Wang, H., Laird, J. M., Sene, A., Apte, R. S., Salomon, R. G., Hollyfield, J. G., and Perez, V. L. (2013) Infiltration of proinflammatory m1 macrophages into the external retina precedes damage in a mouse model of age-related macular degeneration, *Int. J. Inflamm.*, **2013**, 503725.

- 68. Ufret-Vincenty, R. L., Aredo, B., Liu, X., McMahon, A., Chen, P. W., Sun, H., Niederkorn, J. Y., and Kedzierski, W. (2010) Transgenic mice expressing variants of complement factor H develop AMD-like retinal findings, *Invest. Ophthalmol. Vis. Sci.*, **51**, 5878-5887.
- 69. Combadiere, C., Feumi, C., Raoul, W., Keller, N., Rodero, M., Pezard, A., Lavalette, S., Houssier, M., Jonet, L., Picard, E., Debre, P., Sirinyan, M., Deterre, P., Ferroukhi, T., Cohen, S. Y., Chauvaud, D., Jeanny, J. C., Chemtob, S., Behar-Cohen, F., and Sennlaub, F. (2007) CX3CR1 dependent subretinal microglia cell accumulation is associ ated with cardinal features of age-related macular degener ation, *J. Clin. Invest.*, **117**, 2920-2928.
- 70. Luhmann, U. F., Robbie, S., Munro, P. M., Barker, S. E., Duran, Y., Luong, V., Fitzke, F. W., Bainbridge, J. W., Ali, R. R., and MacLaren, R. E. (2009) The drusen-like pheno type in aging Ccl2-knockout mice is caused by an acceler ated accumulation of swollen autofluorescent subretinal macrophages, *Invest. Ophthalmol. Vis. Sci.*, **50**, 5934-5943.
- 71. Chan, C. C., Ross, R. J., Shen, D., Ding, X., Majumdar, Z., Bojanowski, C. M., Zhou, M., Salem, N., Jr., Bonner, R., and Tuo, J. (2008) Ccl2/Cx3cr1-deficient mice: an ani mal model for age-related macular degeneration, *Ophthalm. Res.*, **40**, 124-128.
- 72. Pennesi, M. E., Neuringer, M., and Courtney, R. J. (2012) Animal models of age-related macular degeneration, *Mol. Aspects Med.*, **33**, 487-509.
- 73. Santos, A. M., Martin-Oliva, D., Ferrer-Martin, R. M., Tassi, M., Calvente, R., Sierra, A., Carrasco, M. C., Marin-Teva, J. L., Navascues, J., and Cuadros, M. A. (2010) Microglial response to light-induced photoprotector degeneration in the mouse retina, *J. Comp. Neurol.*, **518**, 477-492.
- 74. Ma, W., Zhao, L., Fontainhas, A. M., Fariss, R. N., and Wong, W. T. (2009) Microglia in the mouse retina alter the structure and function of retinal pigmented epithelial cells: a potential cellular interaction relevant to AMD, *PloS One*, **4**, e7945.
- 75. Ma, W., Zhao, L., and Wong, W. T. (2012) Microglia in the outer retina and their relevance to pathogenesis of age related macular degeneration, *Adv. Exp. Med. Biol*., **732**, 37-42.
- 76. Kozhevnikova, O. S., Korbolina, E. E., Ershov, N. I., and Kolosova, N. G. (2013) Rat retinal transcriptome: effects of aging and AMD-like retinopathy, *Cell Cycle*, **12**, 1745- 1761.
- 77. Perez, V. L., and Caspi, R. R. (2015) Immune mechanisms in inflammatory and degenerative eye disease, *Trends Immunol.*, **36**, 354-363.
- 78. Shaw, P. X., Stiles, T., Douglas, C., Ho, D., Fan, W., Du, H., and Xiao, X. (2016) Oxidative stress, innate immunity, and age-related macular degeneration, *AIMS Mol. Sci.*, **3**, 196-221.
- 79. Xu, H., and Chen, M. (2016) Targeting the complement system for the management of retinal inflammatory and degenerative diseases, *Eur. J. Pharmacol.*, **787**, 94-104.