#### **REVIEW**

# **Cerebral Mechanisms of Hypoxic/Ischemic Postconditioning**

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**Abstract**—This review analyzes recent data on mechanisms of cerebral hypoxia and the protective methods of hypoxic and ischemic postconditioning, as well as their interrelationship with the key mechanisms responsible for neuroprotection and neuroplasticity. Upregulation of expression of antiapoptotic factors and neurotrophins and modulation of activity of sever al protein kinases and transcription factors such as hypoxia-inducible factor-1 (HIF-1) are considered as the most impor tant aspects in the neuroprotective potential of postconditioning. The presented information indicates substantial transfor mative promise of the noninvasive techniques of hypoxic postconditioning as well as significant similarity between the adap tive pathways activated by various postconditioning methods, which are far from being fully understood.

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Hypoxia is one of the most damaging factors during adverse external and internal influences. It is well known that acute forms of hypoxia suppress neuroplasticity, cause disruptions in cognition and memory, as well as exert a disruptive effect on the neurons of the most sus ceptible brain structures such as the hippocampus. On the other hand, moderate forms of hypoxia exhibit the oppo site effect, and this phenomenon has been used for the development of preventive measures for increasing resist ance of the brain to hypoxia (intermittent hypoxic train ing, hypoxic and ischemic preconditioning) or rehabilita tion after damaging action (early, delayed, and distant ischemic postconditioning (IPostC), normobaric and hypobaric hypoxic postconditioning (NBHPostC and HBHPostC, respectively). Currently available informa tion on the cerebral mechanisms of severe brain hypoxia and the protective effects of various types of ischemic (IPostC) and hypoxic (HPostC) postconditioning will be briefly presented, as well as arguments indicating that the noninvasive NBHPostC and HBHPostC approaches should be considered as promising for medical practice.

# HYPOXIA AND MECHANISMS OF ITS ADVERSE EFFECT ON NEUROPLASTICITY

Hypoxia is the state of oxygen deprivation of either the entire organism or its separate organs and tissues. An organism can be found in the state of hypoxia as a result of the effects of external factors, such as for example, being at high altitude, in outer space, or in an inade quately ventilated facility. Hypoxia can also develop as a result of internal unfavorable condition such as clog ging/ischemia of blood vessels or anemia. Hypoxia is an important component of pathogenesis of many dis eases.

The following types of hypoxia are recognized [1]:

1. Hypoxic hypoxia (hypoxemia). The main symp tom – low partial pressure of oxygen in arterial blood and, as a consequence, low oxygen content in the entire organism.

2. Anemic hypoxia (hematic) – hypoxia developing on the deficiency of hemoglobin and/or erythrocytes and normal oxygen pressure in arterial blood.

3. Stagnant hypoxia (circulatory), which develops when enough hemoglobin and normal oxygen pressure are maintained in blood, but the amount of blood reach ing tissues does not correspond to the oxygen demand due to, for example, vessel clogging or rupture.

4. Histotoxic hypoxia (hypoxidosis) – develops as a result of disruption of functioning of respiratory chain enzymes and, hence, the oxygen supplied to the tissues cannot be used in oxidative processes.

*Abbreviations*: BBB, blood–brain barrier; CNS, central nervous system; HBHPostC, hypobaric hypoxic postconditioning; HIF-1, hypoxia-inducible factor 1; HPostC, hypoxic (non ischemic) postconditioning; IPostC, ischemic postcondition ing; LPO, lipid peroxidation; NBHPostC, normobaric hypoxic postconditioning; NMDA-receptor, N-methyl-D-aspartate binding receptor; PostС, all types of postconditioning; SH, severe (damaging) hypobaric hypoxia.

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All types of hypoxia have one common feature – deficiency of oxygen supply causing the development of irreversible changes in vital organs. Ischemic heart dis ease and stroke are the most widespread diseases associat ed with hypoxic state, and these occupy the third place in the list of leading causes of death among all diseases after myocardial infarction and oncological diseases. Ischemia is a form of the circulatory hypoxia in which blood supply to the certain tissue or organ is partially or completely blocked. The essential difference in the response of an organism to non-ischemic and ischemic hypoxia lies in the fact that in the first case the blood flow is maintained or significantly increased as well as supply of tissues with substrates and removal or products of oxidative metabo lism. The decline of high-energy compounds, acidosis, and other metabolic disruptions occurring during non ischemic hypoxia intensify not so drastically as in the case of ischemia. Disruption of motor, sensory, and visual reflexes is observed in patients with stroke caused by brain hypoxia/ischemia as well as aphasia and apathia. Neurophysiological disorders can emerge following ischemia (during the reoxygenation period) such as intel lectual decline, apraxia, spatial orientation deficit, and memory loss [2].

Hypoxia/ischemia of the brain can be caused not only by stroke, but can also develop due to the other adverse effects such as cardiac arrest, brain embolism, or acute hypotension, especially during surgery. Furthermore, the state of brain hypoxia can emerge because of external adverse factors such as hypobaric hypoxia (plane decompression at high altitude).

Independent of the cause of cerebral hypoxia, the "ischemic cascade" chain of pathobiochemical changes resulting in damage to nerve tissue and brain neuron death via either necrosis or apoptosis, permanent disruption of neuronal plasticity such as, for example, suppression of activity of the adaptive transcription factors and as a con sequence, disruption in the expression of protective pro teins always develops [3, 4]. The initial step in the induc tion of a hypoxic/ischemic cascade is decrease in oxygen supply to the brain, which inevitably leads to deficit of macroergic compounds. The decease in aerobic oxidation rate in mitochondria results in a decrease in ATP amount and increase in ADP and AMP content. The enzyme phosphofructokinase is activated at low ATP/(ADP + AMP) ratio, which allows sharp increase in the rate of anaerobic glycolysis reactions that have low energy effi ciency and occur with accumulation of lactate [3]. In the early stages, adaptation to hypoxia and stabilization of energy exchange is still possible, because lactate is formed due to NADH-dependent pyruvate reduction, and removal of the excess of NADH facilitates maintenance of glycolysis processivity. Moreover, it has been shown that lactate plays an important role in providing protection from glutamate excitotoxicity via modulation of activity of ionotropic glutamate receptors selectively binding N-

methyl-D-aspartate (NMDA-receptors) [5]. However, such stabilization is usually short-lived and is accompa nied by rather fast depletion of glycogen stores [3, 6].

Due to the excess of mitochondrial NADH related to the absence of terminal oxidizer capable of transferring electrons for generation of proton gradient, inhibition of the activity of citric acid cycle enzymes occurs together with redirection of alpha-ketoglutarate to the synthesis of glutamate – the major excitatory mediator of the central nervous system (CNS). The increase in the amount of lac tate produced as a result of glycolysis initiates intracellular acidosis. The progressing acidification denatures some proteins. In this stage of hypoxia real deficit of ATP is formed, as the aerobic mechanism does not work due to oxygen deficit, and anaerobic – due to acidosis [3]. The ATP deficit is inevitably reflected on the most energy-con suming enzyme in neurons  $- Na^{+}/K^{+}$ -ATPase  $-$  resulting in decrease in its activity, which, in turn is manifested by the loss of the ability to maintain potassium and sodium gradients across neuronal cell membranes. The most important direct consequence of the decrease in  $Na^{+}/K^{+}$ pump activity is penetration of excess sodium into cells, which causes hyperhydration and cerebral edema. The excessive influx of sodium into neurons also leads to depo larization of membranes, influx of  $Ca^{2+}$ , and  $Ca^{2+}$ -dependent release of glutamate from presynapse with the develop ment of glutamate excitotoxicity in the postsynapse.

Glutamate is the main excitatory mediator of the CNS; it participates in realization of cognitive functions, sustains the level of wakefulness together with acetyl choline, but at high concentrations is toxic to neurons. The excitotoxic effect of glutamate is due not only to sharp release of glutamate, but also due to the disruption of mechanisms of its reverse capture [7]. If reperfusion is conducted in early stages of ischemia, the glutamate con centration reverses to normal values. Glutamate realizes its effect via the group of ionotropic membrane receptors: NMDA, α-amino-3-hydroxy-5-methyl-4-isoxazolepro pionic acid receptors (AMPA-receptors), kainate recep tors, as well as via metabotropic receptors (mGluR). Excitation of the glutamate NMDA-receptors on the background of depolarization of cell membranes develop ing due to the ATP deficit causes enhanced influx of extracellular  $Ca^{2+}$ , and activation of mGluR cause release of intracellular  $Ca^{2+}$  from endoplasmic reticulum (ER) [8].  $Ca^{2+}$  at moderate concentrations plays a role as the key intracellular modulator of synaptic plasticity, in par ticular through change in the number of receptors on the membranes, activation of kinases controlling growth of axons, and formation of new synapses; however, its exces sive accumulation in cytosol modifies the activity of a number of enzymes (proteinases, phospholipases, NO synthases, and others), which eventually mediate damage to membranes, nuclei, and other cellular organelles [9]. This stage of the "ischemic" cascade is accompanied by the generation of reactive oxygen species. The brain is the

organ most sensitive and susceptible to the induction of free radical processes especially on the background of ischemia. This is related to the high content of polyunsat urated fatty acids in brain tissues and with low amounts of vitamin A, extremely low activity of glutathione peroxi dase, practically complete absence of catalase, high con tent of bivalent iron ions, and low level of transferrin and ceruloplasmin [10]. This stage is already irreversible and cannot be corrected by the restoration of oxygen supply.

Next, the "delayed" consequences of hypoxia emerge such as local inflammation reactions, microvas cular disorders, deterioration of the blood–brain barrier (BBB), and others. The signs of degradation of the BBB are present from the first minutes of acute focal ischemia, although they are most manifested after several hours. Inflammation reactions are characterized by migration of neutrophils from blood vessels to tissue, microglia activa tion, and secretion of potentially cytotoxic compounds such as antiinflammatory cytokines interleukin 1 beta (IL-1β) and tumor necrosis factor (TNF). Introduction of antiinflammatory cytokines or compounds decreasing the synthesis of antiinflammatory cytokines significantly decreases the degree of neuron damage following ischemia [11]. Hence, inflammatory reactions are involved in the pathogenesis of post-hypoxic states and facilitate neuronal death.

Induction of a program of delayed apoptotic cell death is a characteristic consequence of acute forms of brain hypoxia. Apoptotic changes can be observed in the CA1-field of the hippocampus and neocortex 3-4 days after transient global cerebral ischemia [12]. Application of protein synthesis inhibitors or growth factors counter acting apoptosis and facilitating normalization of the adequate signal transduction system in brain decreases ischemic damage [13].

Hence, the molecular-biological disorders caused by brain hypoxia/ischemia affect organism very unfavorably. Although the introduction of antioxidants or preparations stimulating the expression of growth factors that stabilize ion gradients can to a certain degree mitigate the conse quences of hypoxia, it is still insufficient to ensure com plex neuroprotection from hypoxia and ischemia during acute stroke, because they affect only one particular stage of the pathological cascade [14].

#### HYPOXIC/ISCHEMIC POSTCONDITIONING

Hypoxic/ischemic postconditioning (PostC) is a promising procedure for structural–functional brain rehabilitation following the damaging action of factors of hypoxic nature. The protective effect of PostC on heart was first demonstrated in 2003. It was shown by a group of American authors that conducting three intermittent 30-s sessions of coronary occlusion with 30-s intervals, when the coronary blood flow was restored, in the reperfusion

period following 60-min coronary occlusion in dogs helped to decrease the degree of myocardial injury. The phenomenon was termed ischemic PostC (IPostC) [15]. In a wide sense, PostC is a procedure of exposure to dam aging factors of moderate intensity following an acute damaging action with the objective to induce endogenous mechanisms facilitating correction of damages. In their pioneer study, Zhao et al. compared the protective effect of post- and pre-conditioning. The size of the infarction zone was analyzed 3 h after reperfusion. The area of injury decreased by 44% when IPostC was used in com parison with 60-min control ischemia, which was compa rable with the protective effect of pre-conditioning.

The IPostC effect that corrected negative conse quences of myocardial ischemia was later confirmed by many research groups in experiments with various ani mals *in vivo* [9, 16-19], as well as *in vitro* in cell cultures [20]. Clinical testing of IPostC in patients after myocar dial infarction also demonstrated positive results [21]. The cardioprotective effect of IPostC is currently actively investigated, data on the efficiency of PostC methods in different organs are being accumulated, and intensive studies the molecular-cellular mechanisms underlying the protective action realized with this method are being conducted [15, 22-25].

Several years after the discovery of the protective effect of IPostC on myocardium, this phenomenon was demonstrated for brain. The first study on brain PostC was published in 2006 by a group of American neurosur geons lead by H. Zhao. Stroke was modeled in rats by per manent occlusion of the middle cerebral artery (MCA) and transient bilateral occlusion of the common carotid artery, and IPostC was performed 2 min after reperfusion using three cycles of 10-s occlusion and 30-s reperfusion of the common carotid artery. The size of the infarction area was evaluated 2 days after the MCA ligation. IPostC ensured 80% decrease in the necrosis area in comparison with the control acute ischemia and facilitated decrease in apoptosis intensity [26]. The results of this study have been reproduced by many scientific groups worldwide that used various regimes of acute ischemia and IPostC [27-30]. The effect of PostC was also demonstrated in *in vitro* experiments using hippocampal slice culture [31] and primary neuronal culture [32].

# NEUROPROTECTIVE MECHANISMS ACTIVATED BY ISCHEMIC POSTCONDITIONING

The mechanisms of neuroprotective action of IPostC are under active investigation. It was shown that IPostC suppressed the generation of free radicals and initiation of apoptosis during the reperfusion period, and it decreased the level of malondialdehyde (MDA) and of oxidative protein modifications with simultaneous increase in the activity of superoxide dismutase and catalase in brain tis-

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sues [26, 27, 30, 33, 34]. This suggests high antioxidant effect of IPostC.

By analogy with heart, it was shown that IPostC of brain mediated an increase in the amount of phosphory lated protein kinase Cε (PKCε) with simultaneous decrease in the amount of phosphorylated protein kinase Cδ (PKCδ), which according to the present notion facili tated cell death [35]. The c-Jun N-terminal kinase (JNK), which activates c-jun and p53 transcription factors, plays an important role in neuronal death and in pathogenesis of neurodegenerative disorders [36]. The researchers from California established that IPostC caused a decrease in the amount of phosphorylated JNK in brain [35].

Protein kinase Akt plays a special role in the sup pression of apoptosis-induced signals, thus ensuring nor mal functioning of cells and intercellular interactions [37, 38]. It was found in experiments with hippocampal slices that the selective inhibition of the Akt activator alleviated the neuroprotective effect of IPostC [31]. The American physiologists also showed in *in vivo* experiments that brain IPostC resulted in the phosphorylation of Akt and, as a consequence, increase in activity of this enzyme [35]. Moreover, a decrease in the size of the infarction area was not observed under conditions of PI3-kinase blockade; however, there was a decrease in the expression of ER chaperones, and induction of the ER-dependent apopto sis involving caspase 12 was observed [39]. In addition, the role of Akt in realization of the IPostC effect via acti vation of mTOR kinase was demonstrated. Interestingly, anesthetics mimicking the PostC phenomenon (propofol and isoflurane) also activate Akt and PI3-kinase [40, 41].

ERK phosphorylated by MEK is another kinase ensuring survival of cells under adverse conditions [42, 43]. In 2009, Jiang et al. [44] established that IPostC of the rabbit spinal cord (*in vivo* model) resulted in phos phorylation of ERK, and the neuroprotective effect did manifest itself under conditions of MEK blocking. The involvement of ERK in the mechanisms of the brain tol erance to hypoxic conditions was acknowledged during investigation of the older phenomenon of hypoxic pre conditioning [45] of rat brain in the model of hypobaric hypoxia. However, the data of Jiang et al. contradict the results American and Chinese specialists [35, 46], who observed a decrease in the amount of phosphorylated ERK following ischemic and normobaric hypoxic PostC; moreover, the latter demonstrated the role of the MEK/ERK signaling pathway inhibition in compensa tion of the consequences of transient global cerebral ischemia [46]. The cause of this discrepancy remains unclear. The role of each separate kinase and peculiarities of their interactions during the PostC procedure is still poorly explored as well as the final effects of IPostC.

The enzymes of antioxidant cell protection [26, 27, 30], chaperones [34, 39], as well as factors controlling mitochondria-mediated induction of apoptosis [34, 47] are suggested as key effectors in IPostC. One of the bestinvestigated proteins suppressing this apoptotic pathway is the Bcl-2 (B-cell lymphoma 2) protein. Bcl-2 expres sion in brain is determined by the efficiency of signal transduction between neurons, and it reflects the bal anced operation of the signaling pathways involved in ensuring neuroplasticity [48]. Xing et al. found that IPostC facilitated an increase in Bcl-2 level, decrease in proapoptotic Bax protein, and prevented the increase in cytoplasmic cytochrome *c* level in the neuron [34]. The same group of authors reported an antiinflammatory effect of IPostC that was manifested by a decrease in myeloperoxidase activity, decrease in the amount of mal ondialdehyde (marker for lipid peroxidation, LPO), as well as by suppression of the synthesis of proinflammato ry cytokines TNFα, IL-1β, and of intercellular adhesion molecules (ICAM) – markers for inflammation [49]. The signaling pathway causing attenuation of autophagy is considered as a no less interesting IPostC-mediated stim ulation; however, the amount of data on the long-term effects of such rearrangements of the cellular molecular machinery on further neuron functioning is still insuffi cient to make any firm conclusions [50].

The work of Zhang et al. demonstrating the contri bution of IPostC to redistribution of glutamate in rat brain due to enhanced activity of glutamine synthase and upregulated expression of neuronal glutamate trans porters (GLT1) leading to decrease in excitotoxicity and, respectively, decrease in post-ischemic injuries, is worth notice [51, 52].

Recently the possibility of inclusion of processes of neuronal cell proliferation as a pathway to compensate post-stroke neuronal death attracted considerable atten tion of scientists investigating IPostC. Experimental cor roboration of this hypothesis was demonstrated by Esposito et al. [53].

For the most part, the literature accumulated so far are related to models of the so-called rapid IPostC, when reperfusion interruption is conducted in early stages (sec onds and minutes) after ischemia; however, in addition to early IPostC, "delayed" [27, 29, 32, 54, 55] and distant [56, 57] IPostC of brain are also recognized. Furthermore, combination of different variants of IPostC and their application together with the previously described phe nomenon of ischemic preconditioning have been also used, which has been reviewed in detail in several publica tions [57-61]. Successful attempts to model the effect of IPostC by so-called pharmacological PostC using anes thetic inhalation were also reported [41, 62, 63].

## NONINVASIVE METHODS OF POSTCONDITIONING

Despite the well-documented neuroprotective effi ciency of IPostC, the transformative potential of this approach to brain rehabilitation remains questionable

due to several serious drawbacks. In particular, the way of conducting IPostC by intermittent occlusion of the carotid and cerebral arteries is an invasive method requir ing surgical intervention, which poses significant risks and limitations, especially for patients who survived a severe episode for which exactly the application of PostC is required. Moreover, according to the abovementioned literature data, IPostC must be conducted within a rather narrow therapeutic window (from several minutes to 1- 2 h after the ischemic stroke), which also presents limita tions for practical application. That is why the search for noninvasive approaches for PostC and elucidation of the mechanisms of its action hold significant promise.

A novel, noninvasive method of PostC using moder ate hypobaric hypoxia (hypobaric hypoxic postcondition ing, HBHPostC) was developed recently in the laborato ry of regulation of neuron function at the Pavlov Institute of Physiology, Russian Academy of Sciences, which employed a procedure of exposing the organism to low atmospheric pressure causing decrease in oxygen supply (method for hypoxic postconditioning, Patent of the Russian Federation No. 2437164, 20.12.2011) [4, 64-69]. Simultaneously, our Chinese colleagues published the results of a series of studies confirming the efficiency of normobaric hypoxic postconditioning (NBHPostC) for correction of the consequences of transient global cere bral ischemia [46, 70, 71]. The efficiency of NBHPostC for compensation of the consequences of 10-min tran sient global brain ischemia was demonstrated. The NBHPostC involved 2-h exposure to an atmosphere comprising a mixture of 8% oxygen and 92% nitrogen in a chamber one day after the ischemia imposed by occlu sion of the two vertebral and two carotid arteries.

#### HYPOBARIC HYPOXIC POSTCONDITIONING

The efficiency of the HBHPostC method for correc tion of the consequences of severe hypoxia was demon strated in experiments with rats. Severe hypobaric hypoxia (SH) was created by 3-h exposure to the pressure of 180 mm Hg (which is equivalent to 5% of  $O_2$  and corresponds to the altitude of 11 km above sea level) in a flow through-type barochamber. This model is well studied. It is known that approximately 50% of rats do not survive under the conditions of SH, and the remaining manifest signs of brain swelling, deterioration of the BBB, massive cell death in the hippocampus and neocortex, abnormal ities in behavior and cognition, and memory disorders. The postconditioning procedure in this model involved three 2-h sessions of moderate hypobaric hypoxia (360 mm Hg, which corresponds to 10% of normobaric oxygen and is equivalent to the altitude of 5 km above sea level) with 24-h intermissions starting 24 h after SH.

As earlier, the animals that survived SH exhibited clearly pronounced behavioral abnormalities including mobility retardation, increased anxiety, disruption of stereotypic activities, and other signs of depressive-like conditions. The basal level of corticosterone in the blood plasma of these animals was 6-fold lower than in the con trol group, while the stress-reactivity of the hypophysial– adrenocortical system to minor (non-pathogenic) stress increased 5-fold. Application of HBHPostC according to the abovementioned scheme alleviated these disorders [66].

Histological analysis of brain slices stained using the Nissl procedure showed that the loss of ∼30% of pyrami dal neurons occurred in susceptible brain areas such as the hippocampal CA1-field by the 7th day after SH, but HBHPostC almost completely prevented this neuronal death [66]. As shown earlier, massive induction of reactive oxygen species (ROS) and intensification of LPO occurred in the early stages of re-oxygenation after SH in the brain of rats [72, 73], which could be the reason for the neuronal death mentioned above. Based on data on antioxidant action of IPostC [27, 30, 33, 34], we suggest that the HBHPostC-mediated prevention of cell death is related to normalization of the functioning of intracellu lar signaling pathways supporting antioxidant protection. The combination of these and other facts led us to con clude that HBHPostC exhibited high neuroprotective effect.

# MECHANISMS OF NEUROPROTECTIVE EFFECTS OF HYPOXIC POSTCONDITIONING

Unlike in the case of ischemic postconditioning, the mechanisms of neuroprotective action of hypoxic post conditioning (HPostC) are still poorly understood. Two parallel lines of enquiry must be noted that are based on application of different models – normobaric PostC (NBHPostC) and hypobaric PostC (HBHPostC). It was shown that HBHPostC stimulated the expression of anti apoptotic protein Bcl-2 and neurotrophin BDNF in the rat hippocampus [68]. Similar data were obtained in the model of early IPostC [34]. The stimulation of signaling pathways regulating the expression of neurotrophins and antiapoptotic protein is characteristic for many forms of neuroplasticity, and it likely plays a key role in the protec tive function of PostC preventing cellular death, normal izing LPO as well as behavioral characteristics of rats [68]. Moreover**,** the key transcription regulator inducing adap tation to conditions of oxygen deficiency (hypoxia inducible factor-1, HIF-1) and the target of its transcrip tional activity – protective cytokine erythropoietin (Epo), which as demonstrated is an important regulator of the synapto- and neurogenesis – clearly provide a sig nificant contribution to these processes. It was found that SH suppressed the expression of the regulatory alpha subunit of HIF-1 (HIF-1 $\alpha$ ) and, as a result, led to a decrease in the Epo level, which was in agreement with

literature data on HIF-1 regulation pathways [74] and peculiarities of the activity of protein kinases during reoxygenation periods in the ischemic models [31, 35, 39, 42, 43]. At the same time, HBHPostC facilitates an increase in the amount of HIF-1 $\alpha$  and Epo, which supposedly also contributes to the neuroprotective effect of HBHPostC [67].

The group of Chinese researchers under the leader ship of En Xu was very successful in investigation of mod ulation of activity of protein kinases controlling the func tioning of HIF-1 under conditions of pathological and adaptive hypoxia using NBHPostC to compensate for the consequences of 10-min transient global brain ischemia. The NBHPostC in this case involved 2-h exposure to a mixture of 8% oxygen and 92% nitrogen one day after an ischemia episode caused by the occlusion of the two ver tebral and two carotid arteries.

In the first experiments, the authors showed that NBHPostC resulted in an increase in phosphorylation of the Akt kinase and neuroprotective transcription factor FoxO, which correlated with a decrease in infarct size and prevention of cell death. Similarly to the case of IPostC [35, 39], the use of the PI3K inhibitor alleviates the neuroprotective effect and is accompanied by a decrease in Akt and FoxO phosphorylation, as well as by intensifica tion of cell death. At the same time, NBHPostC mediates a decrease in phosphorylation of the MEK and ERK kinases that are activated in the model of transient global cerebral ischemia, and the MEK inhibitor facilitates a decrease in neuronal loss following the acute ischemia [46].

In the next stage, the same group of authors demon strated the role of neuronal (but not glial) kinase p38 in the neuroprotective effect of NBHPostC via phosphory lation of MSK kinase, which activated the c-Rel subunit of a transcription factor of the NF-κB family [70]. In addition to the direct protective action known in the CNS, NF- $\kappa$ B plays a role of coactivator of the  $\alpha$ -subunit of the hypoxia-inducible factor  $(HIF-I\alpha)$ , which makes the data on the involvement of this hypoxia adaptation factor into providing neuroprotection during the NBHPostC very noteworthy. Indeed, NBHPostC increases the expression of  $HIF-1\alpha$ , as well as the expression of the products of its gene targets such as vascular endothelial growth factor (VEGF), thus mediating the prevention of cell death and decrease in the level of active caspase-3 in the hippocampus. The inhibitors of both the p38 kinase and the Akt kinase cause complete blocking of this signaling pathway in the process, and, on the con trary, the inhibitor of MEK/ERK-signaling stimulates the HIF-1α-dependent neuroprotective pathway in response to global cerebral ischemia in the absence of NBHPostC [71].

In conclusion, accumulated data indicate significant similarity between the neuroprotective pathways of ischemic and hypoxic (either hypobaric or normobaric) postconditioning, as well as their significant overlapping with mechanisms of brain neuroplasticity. However, these issues require further investigation, in particular to under stand the role of neuron–glial interactions in providing plasticity to the brain exposed to hypoxia.

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### REFERENCES

- 1. Van Liere, E. J., and Stickney, J. C. (1963) *Hypoxia*, The University of Chicago Press, Chicago.
- 2. Bokura, H., and Robinson, R. (1997) Long-term cognitive impairment associated with caudate stroke, *Stroke*, **28**, 970-975.
- 3. Fedin, A., and Rumyantseva, S. (2004) *Intensive Therapy of Ischemic Stroke* [in Russian], Meditsinskaya Kniga, Moscow.
- 4. Rybnikova, E., and Samoilov, M. (2015) Current insights into the molecular mechanisms of hypoxic pre- and post conditioning using hypobaric hypoxia, *Front. Neurosci*., **9**, doi: 10.3389/fnins.2015.00388.
- 5. Jourdain, P., Allaman, I., Rothenfusser, K., Fiumelli, H., Marquet, P., and Magistretti, P. J. (2016) L-Lactate pro tects neurons against excitotoxicity: implication of an ATP mediated signaling cascade, *Sci. Rep*., **6**, doi: 10.1038/ srep21250.
- 6. Gusev, E. I., Skvortsova, V. I., Izikenova, G. A., Alekseev, A. A., and Dambinova, S. A. (1996) Investigation of the level of autoimmune antibodies against glutamate receptors in blood serum of patients during acute ischemic stroke, *Zh. Nevrol. Psikhiatr*., **106**, 30-34.
- 7. Benveniste, H., Drejer, J., Schousboe, A., and Diemer, N. (1984) Elevation of the extracellular concentrations of glu tamate and aspartate in rat hippocampus during transient cerebral ischemia monitored by intracerebral microdialysis, *J. Neurochem*., **43**, 1369-1374.
- 8. Hartley, D., Kurth, M., Bjerkness, L., Weiss, J., and Choi, D. (1993) Glutamate receptor-induced  $Ca^{2+}$  accumulation in cortical cell culture correlates with subsequent neuronal degeneration, *J. Neurosci*., **13**, 1993-2000.
- 9. Kristian, T., and Siesjo, E. (1989) Calcium in ischemic cell death, *Stroke*, **29**, 705-718.
- 10. Siesjo, E., Agadh, C., and Bengtsson, F. (1989) Free radi cals and brain damage, *Brain Metab. Rev*., **1**, 165-211.
- 11. Block, F., and Schwarz, M. (1996) Dextromethorphan reduces functional deficits and neuronal damage after glob al ischemia in rats, *Brain Res*., **741**, 153-159.
- 12. Nitatori, T., Sato, N., and Waguri, S. (1995) Delayed neu ronal death in the CA1 pyramidal cell layer of the gerbil hippocampus following transient ischemia is apoptosis, *J. Neurosci*., **15**, 1001-1011.
- 13. Goto, K., Ishige, A., Sekiguchi, K., Lizuka, S., Sugimoto, A., Yuzurihara, M., Aburada, M., Hosoya, E., and Kogure,

K. (1990) Effects of cycloheximide on delayed neuronal death in rat hippocampus, *Brain Res*., **534**, 299-302.

- 14. Fisher, M., and Ratan, R. (2003) New perspectives on developing acute stroke therapy, *Ann. Neurol*., **53**, 10- 20.
- 15. Zhao, Z.-Q., Corvera, J., Halkos, M., Kerendi, F., Wang, N.-P., Guyton, R., and Vinten-Johansen, J. (2003) Inhibition of myocardial injury by ischemic postcondition ing during reperfusion: comparison with ischemic precon ditioning, *Am. J. Physiol. Heart Circ. Physiol.*, **285**, 579- 588.
- 16. Iliodromitis, E., Georgiadis, M., Cohen, M., Downey, J., Dimitrios, E., and Kremastinos, T. (2006) Protection from postconditioning depends on the number of short ischemic insults in anesthetized pigs, *Basic Res. Cardiol*., **101**, 502- 507.
- 17. Kin, H., Zatta, A., Lofye, M., Amerson, B., Halkos, M., Kerendi, F., Zhao, Z.-Q., Guyton, R., Headrick, J., and Vinten-Johansen, J. (2005) Postconditioning reduces infarct size via adenosine receptor activation by endoge nous adenosine, *Cardiovasc. Res*., **67**, 124-133.
- 18. Krolikowski, J., Weihrauch, D., Bienengraeber, M., Kersten, J., Warltier, D., and Pagel, P. (2006) Role of Erk1/2, p70s6K, and eNOS in isoflurane-induced cardio protection during early reperfusion *in vivo*, *Can. J. Anaesth*., **53**, 174-182.
- 19. Obal, D., Dettwiler, S., Favoccia, C., Scharbatke, H., Preckel, B., and Schlack, W. (2005) The influence of mito chondrial  $K_{ATP}$ -channels in the cardioprotection of preconditioning and postconditioning by sevoflurane in the rat *in vivo*, *Anesth. Analg*., **101**, 1252-1260.
- 20. Dosenko, V., Nagibin, V., Tumanovskaya, L., and Vaage, J. (2006) Proteasome inhibitors eliminate protective effect of postconditioning in cultured neonatal cardiomyocytes, *Fiziol. Zh*., **52**, 15-24.
- 21. Staat, P., Rioufol, G., Piot, C., Cottin, Y., Cung, T., L'Huillier, I., Aupetit, J.-F., Bonnefoy, E., Finet, G., Andre-Fouet, X., and Ovize, M. (2005) Postconditioning the human heart, *Circulation*, **112**, 2143-2148.
- 22. Maslov, L., Krig, T., and Diwan, V. (2009) Postconditioning is a universal protective phenomenon, *Patol. Fiziol. Eksp. Ter*., **3**, 2-6.
- 23. Skyschally, A., Van Caster, P., Boengler, K., Gres, P., Musiolik, J., Schilawa, D., Schulz, R., and Heusch, G. (2009) Ischemic postconditioning in pigs: no causal role for risk activation, *Circ. Res*., **104**, 15-18.
- 24. Zhao, H.-X., Wang, X.-L., Wang, Y.-H., Wu, Y., Li, X.-Y., Lv, X.-P., Zhao, Z.-Q., Zhao, R.-R., and Liu, H.-R. (2010) Attenuation of myocardial injury by postconditioning: role of hypoxia inducible factor-1α, *Basic Res. Cardiol*., **105**, 109-118.
- 25. Zhao, Z.-Q., and Vinten-Johansen, J. (2006) Postconditio ning: reduction of reperfusion-induced injury, *Cardiovasc. Res*., **70**, 200-211.
- 26. Zhao, H., Sapolsky, R., and Steinberg, G. (2006) Interrupting reperfusion as a stroke therapy: ischemic post conditioning reduces infarct size after focal ischemia in rats, *J. Cereb. Blood Flow Metab*., **26**, 1114-1121.
- 27. Danielisova, V., Nemethova, M., Gottlieb, M., and Burda, J. (2006) The changes in endogenous antioxidant enzyme activity after postconditioning, *Cell. Mol. Neurobiol*., **26**, 1181-1191.
- 28. Gao, X., Ren, C., and Zhao, H. (2008) Protective effects of ischemic postconditioning compared with gradual re perfusion or preconditioning, *J. Neurosci. Res*., **86**, 2505- 2511.
- 29. Ren, C., Gao, X., Niu, G., Yan, Z., Chen, X., and Zhao, H. (2008) Delayed postconditioning protects against focal ischemic brain injury in rats, *PLoS One*, **3**, e3851.
- 30. Song, W., Dong, H., Cheng, O., Lu, Z., Wang, H., Meng, J., and Xiong, L. (2008) Ischemic postconditioning induced neuroprotection via up-regulation of endogenus antioxidant enzyme activities: experiment with rabbits, *Zhonghua Yi Xue Za Zhi*, **88**, 2355-2359.
- 31. Scartabelli, T., Gerace, E., Landucci, E., Moroni, F., and Pellegrini-Giampietro, D. (2008) Neuroprotection by group I mGlu receptors in a rat hippocampal slice model of cerebral ischemia is associated with the PI3K-Akt signaling path way: a novel postconditioning strategy? *Neuropharmacology*, **55**, 509-516.
- 32. Leconte, C., Tixier, E., Freret, T., Toutain, J., Saulnier, R., Boulouard, M., Roussel, S., Schumann-Bard, P., and Bernaudin, M. (2009) Delayed hypoxic postconditioning protects against cerebral ischemia in the mouse, *Stroke*, **40**, 3349-3355.
- 33. Li, Z.-Y., Liu, B., Yu, J., Yang, F.-W., Luo, Y.-N., and Ge, P.-F. (2012) Ischaemic postconditioning rescues brain injury caused by focal ischemia/reperfusion via attenuation of protein oxidization, *J. Int. Med. Res*., **40**, 954-966.
- 34. Xing, B., Chen, H., Zhang, M., Zhao, D., Jiang, R., Liu, X., and Zhang, S. (2008) Ischemic postconditioning inhibits apoptosis after focal cerebral ischemia/reperfusion injury in the rat, *Stroke*, **39**, 2362-2369.
- 35. Gao, X., Zhang, H., Takahashi, T., Hsieh, J., Liao, J., Steinberg, G. K., and Zhao, H. (2008) The Akt signaling pathway contributes to postconditioning protection against stroke; the protection is associated with the MAPK and PKC pathways, *J. Neurochem*., **105**, 943-955.
- 36. Wang, L., Besirli, C., and Johnson, E. (2004) Mixed-line age kinases: a target for the prevention of neurodegenera tion, *Ann. Rev. Pharmacol. Toxicol*., **44**, 451-474.
- 37. Alessi, D., James, S., Downes, C., Holmes, A., Gaffney, P., Reese, C., and Cohen, P. (1997) Characterization of a 3 phosphoinositide-dependent protein kinase which phos phorylates and activates protein kinase B, *Curr. Biol*., **7**, 261-269.
- 38. Jonassen, A., Sack, M., Mjos, O., and Yellon, D. (2001) Myocardial protection by insulin at reperfusion requires early administration and is mediated via Akt and p70s6 kinase cell-survival signaling, *Circ. Res*., **89**, 1191- 1198.
- 39. Yuan, Y., Guo, Q., Ye, Z., Pingping, X., Wang, N., and Song, Z. (2011) Ischemic postconditioning protects brain from ischemia/reperfusion injury by attenuating endoplas mic reticulum stress-induced apoptosis through PI3K-Akt pathway, *Brain Res*., **1367**, 85-93.
- 40. Aronowski, J., Strong, R., and Grotta, J. (1997) Reperfusion injury: demonstration of brain damage pro duced by reperfusion after transient focal ischemia in rats, *J. Cereb. Blood Flow Metab*., **17**, 1048-1056.
- 41. Wang, H., Wang, G., Yu, Y., and Wang, Y. (2009) The role of phosphoinositide-3-kinase/Akt pathway in propofol induced postconditioning against focal cerebral ischemia reperfusion injury in rats, *Brain Res*., **1297**, 177-184.

BIOCHEMISTRY (Moscow) Vol. 82 No. 3 2017

- 42. Grewal, S., York, R., and Stork, P. (1999) Extracellular signal-regulated kinase signaling in neurons, *Curr. Opin. Neurobiol*., **9**, 544-553.
- 43. McCubrey, J., Milella, M., Tafuri, A., Martelli, A. M., Lunghi, P., Bonati, A., Cervello, M., Lee, J. T., and Steelman, L. S. (2008) Targeting the Raf/MEK/ERK path way with small-molecule inhibitors, *Curr. Opin. Invest. Drugs*, **9**, 614-630.
- 44. Jiang, X., Ai, C., Shi, E., Nakajima, Y., and Ma, H. (2009) Neuroprotection against spinal cord ischemia-reperfusion injury induced by different ischemic-postconditioning methods: roles of phosphoinositide-3-kinase-Akt and ERK, *Anesthesiology*, **111**, 1197-1205.
- 45. Samoilov, M. O., Rybnikova, E. A., Sitnik, N. A., Glushchenko, T. S., Tyul'kova, E. I., and Grinkevich, L. N. (2007) Preconditioning modifies the activities of mitogen activated protein kinases and c-jun transcription factor in the rat hippocampus after severe hypobaric hypoxia, *Neurochem. J.*, **1**, 219-226.
- 46. Zhan, L., Li, D., Liang, D., Wu, B., Zhu, P., Wang, Y., Sun, W., and Xu, E. (2012) Activation of Akt/FoxO and inactivation of MEK/ERK pathways contribute to induc tion of neuroprotection against transient global cerebral ischemia by delayed hypoxic postconditioning in adult rats, *Neuropharmacology*, **63**, 873e88.
- 47. Halestrap, A. (2010) A pore way to die: the role of mito chondria in reperfusion injury and cardioprotection, *Biochem. Soc. Trans*., **38**, 841-860.
- 48. Schabitz, W., Sommer, C., Zoder, W., Kiessling, M., Schwaninger, M., and Schwab, S. (2000) Intravenous brain-derived neurotrophic factor reduces infarct size and counter-regulates Bax and Bcl-2 expression after tempo rary focal cerebral ischemia, *Stroke*, **31**, 2212-2217.
- 49. Xing, B., Chen, H., Zhang, M., Zhao, D., Jiang, R., Liu, X., and Zhang, S. (2008) Ischemic post-conditioning pro tects brain and reduces inflammation in a rat model of focal cerebral ischemia/reperfusion, *J. Neurochem*., **105**, 1737- 1745.
- 50. Gao, L., Jiang, T., Guo, J., Liu, Y., Cui, G., Gu, L., Su, L., and Zhang, Y. (2012) Inhibition of autophagy contributes to ischemic postconditioning-induced neuroprotection against focal cerebral ischemia in rats, *PLoS One*, **7**, e46092.
- 51. Zhang, W., Miao, Y., Zhou, S., Wang, B., Luo, Q., and Qiu, Y. (2010) Involvement of glutamate transporter-1 in neuroprotection against global brain ischemia-reperfusion injury induced by postconditioning in rats, *Int. J. Mol. Sci*., **11**, 4407-4416.
- 52. Zhang, W., Miao, Y., Zhou, S., Jiang, J., Luo, Q., and Qiu, Y. (2011) Neuroprotective effects of ischemic postcondi tioning on global brain ischemia in rats through upregula tion of hippocampal glutamine synthetase, *J. Clin. Neurosci*., **18**, 685-689.
- 53. Esposito, E., Hayakawa, K., Maki, T., Arai, K., and Eng, H. (2015) Effects of postconditioning on neurogenesis and angiogenesis during the recovery phase after focal cerebral ischemia, *Stroke*, **46**, 2691-2694.
- 54. Nemethova, M., Danielisova, V., Gottlieb, M., Kravcukova, P., and Burda, J. (2010) Ischemic postcondi tioning in the rat hippocampus: mapping of proteins involved in reversal of delayed neuronal death, *Arch. Ital. Biol*., **148**, 23-32.

BIOCHEMISTRY (Moscow) Vol. 82 No. 3 2017

- 55. Ren, C., Yan, Z., Wei, D., Gao, X., Chen, X., and Zhao, H. (2009) Limb remote ischemic postconditioning protects against focal ischemia in rats, *Brain Res*., **1288**, 88-94.
- 56. Peng, B., Guo, Q., He, Z., Ye, Z., Yuan, Y., Wang, N., and Zhou, J. (2012) Remote ischemic postconditioning pro tects the brain from global cerebral ischemia/reper fusion injury by up-regulating endothelial nitric oxide syn thase through the PI3K/Akt pathway, *Brain Res*., **1445**, 92- 102.
- 57. Zhou, C., Tu, J., Zhang, Q., Lua, D., Zhua, Y., Zhanga, W., Yanga, F., Brannb, D. W., and Wang, R. (2011) Delayed ischemic postconditioning protects hippocampal CA1 neu rons by preserving mitochondrial integrity via Akt/GSK3beta signaling, *Neurochem. Int*., **59**, 749-758.
- 58. Maslov, L. N., and Lishmanov, Yu. B. (2012) Neuropro tective effect of ischemic postconditioning and distant postconditioning. The prospects of clinical application, *Angiol. Vasc. Surg.*, **18**, 27-34.
- 59. Ma, X. D., Song, J. N., Zhang, M., An, J. Y., Zhao, Y. L., and Zhang, B. (2014) Advances in research of the neuro protective mechanisms of cerebral ischemic postcondition ing, *Int. J Neurosci*., 161-169.
- 60. Zhao, H. (2009) Ischemic postconditioning as a novel avenue to protect against brain injury after stroke, *J. Cereb. Blood Flow Metab*., **29**, 873-885.
- 61. Zhao, H., Ren, C., Chen, X., and Shen, J. (2012) From rapid to delayed and remote postconditioning: the evolving concept of ischemic postconditioning in brain ischemia, *Curr. Drug Targets*, **13**, 173-187.
- 62. Adamczyk, S., Robin, E., Simerabet, M., Kipnis, E., Tavernier, B., Vallet, B., Bordet, R., and Lebuffe, G. (2010) Sevoflurane pre- and post-conditioning protect the brain via the mitochondrial K<sub>ATP</sub> channel, *Br. J. Anaesth.*, 104, 191-200.
- 63. McMurtrey, R., and Zuo, Z. (2010) Isoflurane precondi tioning and postconditioning in rat hippocampal neurons, *Brain Res*., **1358**, 184-190.
- 64. Zen'ko, M. Yu., Rybnikova, E. A., and Glushchenko, T. S. (2014) Expression of BDNF neurotrophin in hippocampus and neocortex of rats during formation of post-stress anxi ety state and its correction by hypoxic postconditioning, *Neurosci. Behav. Physiol*., **45**, 869-872.
- 65. Gamdzyk, M., Makarewicz, D., Sіomka, M., Ziembowicz, A., and Salinska, E. (2014) Hypobaric hypoxia postcondi tioning reduces brain damage and improves antioxidative defense in the model of birth asphyxia in 7-day-old rats, *Neurochem. Res*., **39**, 68-75.
- 66. Rybnikova, E., Vorobyev, M., Pivina, S., and Samoilov, M. (2012) Postconditioning by mild hypoxic exposures reduces rat brain injury caused by severe hypoxia, *Neurosci. Lett*., **513**, 100-105.
- 67. Vetrovoy, O., Rybnikova, E., Glushchenko, T., Baranova, K., and Samoilov, M. (2014) Mild hypobaric hypoxic post conditioning increases the expression of  $HIF1\alpha$  and erythropoietin in the CA1 field of the hippocampus of rats that survive after severe hypoxia, *Neurochem. J*., **8**, 103-108.
- 68. Vetrovoi, O., Rybnikova, E., Glushchenko, T., and Samoilov, M. (2015) Effects of hypoxic postconditioning on the expression of antiapoptotic protein Bcl-2 and neu rotrophin BDNF in hippocampal field CA1 in rats subject ed to severe hypoxia, *Neurosci. Behav. Physiol*., **45**, 367- 370.
- 69. Vetrovoi, O., Rybnikova, E., Glushchenko, T., and Samoilov, M. (2016) Effects of hypobaric hypoxia in vari ous modes on expression of neurogenesis marker NeuroD2 in the dentate gyrus of rats hippocampus, *Bull. Exp. Biol. Med*., **160**, 510-513.
- 70. Zhu, P., Zhan, L., Zhu, T., Liang, D., Hu, J., Sun, W., Hou, Q., Zhou, H., Wu, B., Wang, Y., and Xu, E. (2013) The roles of p38 MAPK/MSK1 signaling pathway in the neuroprotection of hypoxic postconditioning against tran sient global cerebral ischemia in adult rats, *Mol. Neurobiol*., doi: 10.1007/s12035-013-8611-7.
- 71. Zhu, T., Zhan, L., Liang, D., Hu, J., Lu, Z., Zhu, X., Sun, W., Liu, L., and Xu, E. (2014) Hypoxia-inducible factor 1a mediates neuroprotection of hypoxic postconditioning

against global cerebral ischemia, *J. Neuropathol. Exp. Neurol*., **73**, 975-986.

- 72. Kislin, M., Tulkova, E., and Samoilov, M. (2009) Changes in lipid peroxidation in the hippocampus and neocortex after severe hypobaric hypoxia, *Neurochem. J*., **3**, 184- 190.
- 73. Kislin, M., Tulkova, E., and Samoilov, M. (2011) Dynamics of lipid peroxidation of membranes in cells and mitochon drial fraction of neocortex in non- and preconditioned rats after severe hypobaric hypoxia, *J. Evol. Physiol. Biochem*., **47**, 187-192.
- 74. Dengler, V., Galbraith, M., and Espinosa, J. (2014) Transcriptional regulation by hypoxia inducible factors, *Crit. Rev. Biochem. Mol. Biol*., **49**, 1-15.