scientific reports

Check for updates

Efectiveness of local isolates OPEN of *Trichoderma* **spp. in imparting drought tolerance in rice,** *Oryza sativa*

Mohammad Haniph Shah  & Mujeebur Rahman Khan*

Rice is a crop that requires high amount of water, and the drought is a major constraint in paddy cultivation. Water stress condition frequently prevails due to shortage of rain which results in signifcantly reduced plant growth and yield of rice. In the present study capability of *Trichoderma* **spp. in imparting drought tolerance to rice,** *Oryza sativa* **was explored. Eleven local strains of** *Trichoderma* **spp. were applied to rice cv. Swarna Sub-1 through soil application (2 g/kg soil) and seed treatment (20 g/kg seed) under 0, 25, 50 and 75% less watering of the recommended amount. The soil application of** *T. harzianum* **AMUTHZ84 signifcantly promoted the shoot and root length (23.6 and 21.3%) followed by seed treatment (19.7 and 18.2%) under recommended level of irrigation condition (100% irrigation). Next in efectiveness was** *T. viride* **AMUTVR73 (21.5 and 18.1%) over untreated control. However, under 75% water availability, soil application with** *T. harzianum* **AMUTHZ82 was found superior over other isolates in enhancing shoot and root length (17.7 and 16.4%). The same isolate was also recorded to be superior under 50% (12.4 and 10.1%) and 25% water availability (9.3 and 8.1%) in enhancing the plant growth and biomass of rice cv. Swarna Sub-1. The isolate also signifcantly enhanced the leaf pigments, and photosynthesis in the rice plants grown under 25–75% water stress condition. In general, soil application of** *Trichoderma* **isolates was found more efective than seed treatment, and the** *T. harzianum* **AMUTHZ82 provided 8–17% enhancement in the plant growth, biomass, leaf pigments and photosynthesis of rice cv. Swarna Sub-1 grown under 25–75% water stress condition.**

Rice is the primary and fundamental food source providing 20% of the global calories consumed by over half the world's population and fulfls the dietary requirements of a large proportion of the human population especially in Asia and Africa¹. It is cultivated on 165.25 mh with an annual global production of around 787.29 mt 3 . India has the largest land area under rice cultivation, but occupies second place in the tonnage globally afer China. Tis is largely due to lower productivity rate of rice in India (4.21 tons/ha) over China (7.11 tons/ha) and the rest of the world 4.76 tons/ha^{[2](#page-10-2)}. In India, rice is grown in all states. West Bengal is the highest rice-producing state (16.72 mt), while Punjab has highest productivity rate (5.10 ton/ha). Uttar Pradesh is the second largest producer of rice, where around 5.60 mha of land is used to grow rice, with a total production of around 15.2 $\mathrm{m}44$.

Despite of being highest in rice acreage in the world, the annual gross productivity of rice in India is 40% less than China^{[2](#page-10-2)}. The significantly lower rice productivity rate in India is due to regular occurrence of biotic and abiotic stresses in rice cultivation^{[5](#page-10-4)}. Drought or lesser availability of water to paddy crop is a major and challenging abiotic stress to rice crop^{[5](#page-10-4)}. Rice plants require saturated or semi-saturated condition to maintain proper growth and development for better productivity. The crop yield is hampered severely when there is a limitation in the water availability^{5[,6](#page-10-5)}. The moderate drought can potentially reduce rice crop yields by up to [9](#page-10-7)–10% globally⁷⁻⁹.

Trichoderma spp. are a group of flamentous fungi that colonize the rhizosphere in all natural habitates, and may also form symbiotic relationships with plants^{[7](#page-10-6),[11](#page-10-8)}. They are renowned for their multifunctional capabilities, including suppression of plant pathogens^{2,[12](#page-11-0)}, promotion of plant growth, and modification of plant responses to abiotic stresse[s13.](#page-11-1) Several studies have demonstrated the positive impact of *Trichoderma* spp. in enhancing drought tolerance in rice^{14,15}. Some species of *Trichoderma*, for example *T. harzianum* is an efficient mineral solu-bilizer and pathogen suppressor that can colonize the roots of crop plants and act as an efficient mycoparasite^{16-[18](#page-11-5)}.

Department of Plant Protection, Faculty of Agricultural Sciences, Aligarh Muslim University, Aligarh 202002, UP, India. [⊠]email: mrkhan777in@yahoo.co.in

The biopriming with *Trichoderma* spp. can enhance nutrient absorption and improves water-holding capacity¹⁹, leading to gradual improvement in the plant growth and development under water stress conditions^{20–22}. Besides the plant breeding for drought tolerance, no other reliable and efective technique has been developed to protect rice from water stress due to low rainfalls. In recent years potential of microorganisms in imparting tolerance against water stress has been evaluated^{[23](#page-11-9)}. *Trichoderma* spp. have demonstrated promising potential that can be exploited commercially to protect plants under moderate level of drought[s24](#page-11-10)[,25.](#page-11-11) *Trichoderma* spp. have been reported in reducing the impact of biotic and abiotic stress on plants^{26,27}. The present study was carried out to examine and ascertain the role of *Trichoderma* spp. in protecting rice plants from the impact of water stress.

Results

Isolation and characterization of *Trichoderma* **species**

The DNA of *Trichoderma* species was sequenced by Macrogen[®] Incorporation, South Korea using universal primer (ex, 16S V3-V4, ITS, 18S) and Sanger sequencing method. The 16S and 18S rDNA gene sequences showed 99% similarity of the isolates with *Trichoderma* spp. (Fig. [1\)](#page-1-0). The isolate showed 99 homologies with the ITS region of *Trichoderma* spp. Tus, the isolates were identifed as *Trichoderma harzianum, T. viride, T. virens, T. hamatum* and *T. koningii.* The 16S and 18S rDNA gene sequences were deposited in NCBI gene bank under the accession number listed in Table [1](#page-2-0). However, to avoid error in the grouping by ITS region, it is recommended to perform sequencing of the conserved region genes such as the TEF gene, Beta-tubulin gene, and Histone H3 gene for species-level identifcation in fungal isolates, especially, the TEF gene which shows a high level of sequence polymorphism among closely related species, even in comparison to the intron-rich portions of protein-coding genes such as the calmodulin gene, beta-tubulin gene, and histone H3 gene.

Plant growth

The rice cv. Swarna Sub-1 grown under water deficiency condition (75, 50 and 25% water availability of the recommended amount) showed the symptoms of water stress, exhibiting reduced plant growth, with shorter stems and smaller leaves in comparison to plants received the recommended level of irrigation. The symptoms varied with the level of water stress. In general, the rice cv. Swarna Sub-1 was found highly sensitive to drought (water stress). However, the soil application of *T. harzianum* AMUTHZ82 subsided the water stress symptoms and promoted shoot and root length by 19.7 and 16.8% with 75% watering, 17.4 and 14.1% with 50% watering and 12.3 and 11.2% with 25% watering over respective controls (Tables [2](#page-2-1), [3\)](#page-3-0). Next in efectiveness was *T. viride* AMUTVR73, other isolates did not produce the significant effect. The seed treatment also increased the length

2

Table 2. Efects of *Trichoderma* spp. against diferent water stress levels on the plant growth (shoot length (cm)) of rice cultivar Swarna sub-1. Values are mean of 5 replicates. Values in the parenthesis are percent increase over control. Values (means) followed by diferent alphabets within a column are signifcantly diferent at P≤0.05 according two-factor ANOVA and Tukey HSD test.

of shoot and root i.e., 16.1 and 13.4% (*T. harzianum* AMUTHZ82) and 10.6 and 8.2% (*T. viride* AMUTVR73) under 50% drought condition. Next in efectiveness were *T. viride* AMUTVR72, *T. hamatum* AMUTHAM_71, *T. harzianum* AMUTHZ81 and *T. viride* AMUTVR72 which signifcantly increased shoot and root length under drought conditions (*P*≤0.05; Tables [2,](#page-2-1) [3\)](#page-3-0). However, under normal irrigation (100% watering) highest promotion in the plant growth of rice was recorded, 21.3–23.6% (soil application) and 18–19.2% (seed treatment with *T. harzianum* AMUTHZ84. Next in efectiveness was *T. viride* AMUTVR73 (19.8 and 17.5%) over untreated control (Tables [2](#page-2-1), [3\)](#page-3-0).

Table 3. Efects of *Trichoderma* spp. against diferent water stress levels on the plant growth (root length (cm)) of rice cultivar Swarna sub-1. Values are mean of 5 replicates. Values in the parenthesis are percent increase over control. Values (means) followed by diferent alphabets within a column are signifcantly diferent at P≤0.05 according two-factor ANOVA and Tukey HSD test.

Plant biomass production

The biomass production of rice plants grown under water stress conditions (75, 50 and 25% water availability of the recommended amount) was adversely affected. The fresh weight of shoot and root decreased by 21-26%, 36–45% and 52–62% at 75, 50, 25% watering, respectively over 100% watering (control). Te soil application of *T. harzianum* AMUTHZ82 signifcantly promoted the plant biomass production under 75 and 50% water availability with a maximum increase 20.6 and 17.6% in shoot fresh weight and 18.5 and 15.3% in root fresh weight respectively, over control (*P* ≤ 0.05). Next in effectiveness was *T. viride* AMUTVR73, followed by *T. harzianum* AMUTHZ84 and *T. viride* AMUTVR71 which also signifcantly promoted the fresh weight of shoot and root (*P*≤0.05; Tables [4](#page-4-0), [5](#page-5-0)) over control. Under 25% water availability, *T. harzianum* AMUTHZ82 scored highest increase in fresh weight of shoot and root (13.8 and 11.3% respectively; Tables [4,](#page-4-0) [5](#page-5-0)), followed by *T. viride* AMUTVR73 (11.4 and 10.4%), and *T. hamatum* AMUTHAM_71 (8.2 and 8.1%) over control (*P*≤0.05).

However, in 100% watered plants, maximum increase in biomass production was recorded with the soil application of *T. harzianum* AMUTHZ84 (22.6% and 21.4% respectively; Tables [4](#page-4-0), [5\)](#page-5-0) followed by seed treatment (22.1 and 20.7%). Next in efectiveness were *T. viride* AMUTVR73 (20.4 and 20.9%), *T. harzianum* AMUTHZ83 (19.3 and 19.3%) and *T. viride* AMUTVR71 (11.8 and 18.4%) over respective control. Other strains viz., *T. koningii* AMUTKNG_70 and *T. virens* AMUTVIRENS_71 did not signifcantly infuence the biomass production of rice over respective control (*P*≤0.05; Tables [4](#page-4-0), [5](#page-5-0)). Overall, the soil application of *Trichoderma* AMUTHZ82 isolates induced 7–11% greater enhancement in the biomass production of rice in comparison to seed treatment.

Grain yield

The water stress caused significant decline (12–51%) in per plant grain yield production of rice cv. Swarna Sub-1 over normal irrigated control (Table [6](#page-6-0)). However, soil application of *Trichoderma* spp. improved the grain yield in both normal and water stress conditions. In normal irrigated condition, maximum increase in production of grain yield was recorded with the treatment of *T. harzianum* AMUTHZ84 (24.68%), followed by *T. viride* AMUTVR73 and *T. harzianum* AMUTHZ82 (22.08 and 20.13%, respectively; Table [6\)](#page-6-0). Under 75 and 50% of water availability, *T. harzianum* AMUTHZ82 was found superior among all isolates in increasing grain yield by22.79 and 19.63%, respectively, over control (Table [6](#page-6-0)). Next in efectiveness were *T. viride* AMUTVR73, *T. harzianum* AMUTHZ84 and *T. hamatum* AMUTHM_71 which signifcantly increased grain yield of rice under drought conditions over control. Under 25% water availability of recommended amount, none of the isolates of *Trichoderma* were found efective in signifcantly enhancing the grain yield of rice plants over control. Overall, the soil application of *Trichoderma* AMUTHZ82 isolates induced 10–22% greater grain yield production of rice in comparison to seed treatment (Table [6](#page-6-0)).

4

Table 4. Efects of *Trichoderma* spp. against diferent water stress levels on the plant biomass (Shoot fresh weight (gm)) of rice cultivar Swarna sub-1. Values are mean of 5 replicates. Values in the parenthesis are percent increase over control. Values (means) followed by diferent alphabets within a column are signifcantly diferent at P≤0.05 according two-factor ANOVA and Tukey HSD test.

Photosynthesis and transpiration rate

Rice plants grown under drought conditions experienced decline in the photosynthesis rate and transpiration rate (Figs. [2,](#page-6-1) [3\)](#page-7-0). This resulted in reduced plant growth, and quality biomass production. The photosynthesis rate was found maximum in the rice plants grown under normal condition (100% water availability) with the soil application of *T. harzianum* AMUTHZ82 (7.12 µmol m−2 s−1), followed by *T. viride* AMUTVR72, *T. harzianum* AMUTHZ84, *T. harzianum* AMUTHZ83 and *T. hamatum* AMUTHAM_71. Lowest photosynthesis rate was recorded under 25% of water availability in comparison to 100% water availability. Soil application with *T. harzianum* AMUTHZ82 signifcantly enhanced the photosynthesis rate of rice plants growing under 75% (37% increase), 50% (19% increase) and 25% water availabilities (9% increase) over respective control. The seed treatment with *Trichoderma* spp. also minimized the negative efect of water stress on photosynthesis, but less than the soil application.

The transpiration rate was also influenced under low water availability with the treatment of *Trichoderma* spp. The plants treated with *T. harzianum* AMUTHZ82 showed minimum transpiration rate (2.36 µmol m⁻² s⁻¹), followed by *T. viride* AMUTVR72, *T. hamatum* AMUTHAM_71 and *T. harzianum* AMUTHZ84 under normal irrigated condition. The reduction in transpiration rate was observed lesser under 50% and 25% drought condition with the treatment of *Trichoderma* spp. over normal irrigated plants. Whereas, maximum transpiration rate was observed in untreated plant receiving recommended level of irrigation (Fig. [2](#page-6-1), [3\)](#page-7-0).

Leaf pigments

Total chlorophyll content (Chlorophyll A and B) in the leaves of rice plants under water stress conditions (75, 50 and 25% water availability of recommended amount) was recorded with a 20, 37 and 49% decline, respectively over control (Fig. [4\)](#page-7-1). Under regular irrigation (100% watering), soil application of *T. harzianum* AMUTHZ84 resulted a maximum increase (21.3%) in chlorophyll content, followed by *T. viride* AMUTVR73 (17%) in the leaves of rice plants. Whereas, under 75 and 50% watering regime, *T. harzianum* AMUTHZ82 increased the total chlorophyll content (16.6 and 13.1%) over respective control (Figs. [4](#page-7-1), [5](#page-8-0)). *T. harzianum* AMUTHZ83 and *T. viride* AMUTVR73 were found next in increasing chlorophyll content under 25% and 50% water stress conditions.

Table 5. Efects of *Trichoderma* spp. against diferent water stress levels on the plant biomass (root fresh weight (gm)) of rice cultivar Swarna sub-1. Values are mean of 5 replicates. Values in the parenthesis are percent increase over control. Values (means) followed by diferent alphabets within a column are signifcantly diferent at P≤0.05 according two-factor ANOVA and Tukey HSD test.

The plants treated through seed treatment of *Trichoderma* isolates under water stress had significantly greater content of leaf pigment, but less than soil application. The plants receiving 25% water of the recommended amount recorded with 2–7% increase in total chlorophyll content with the soil application or seed treatment with *Trichoderma* isolates (Figs. [4](#page-7-1), [5\)](#page-8-0).

Discussion

The rice cv. Swarna Sub-1 was found sensitive to drought (water stress), and exhibited significant reduction in the plant growth, biomass production, leaf pigments, photosynthesis etc. grown under water stress conditions (75, 50 and 25% water of the recommended amount) in comparison to plants received normal irrigation (100% water availability of the recommended level). Adequate supply of water to a plant is essential for carrying out various physiological, biological and biochemical activities of the plan[t28](#page-11-14)[,29](#page-11-15). A decrease in the water supply infuences the physiological activities especially the photosynthesis^{[30](#page-11-16),[31](#page-11-17)}, and subsequently the plant growth and development³², which directly affect the biomass production^{[33,](#page-11-19)34} as observed in the present study.

Te soil application of *Trichoderma* spp. especially *T. harzianum* AMUTHZ82 and *T. viride* AMUTVR73 subsided the negative efect of water stress on rice plants, and promoted the shoot and root growth, and biomass production of rice cv. Swarna Sub-1. *Trichoderma* spp. are universally occurring benefcial microbes in varied kind of soils¹³, and are generally known to suppress plant pathogenic fungi^{17,28}, by parasitizing them or by producing metabolites, toxins, enzymes, etc. toxic to pathogens[28,](#page-11-14)[35.](#page-11-22) *Trichoderma* spp. also solubilize minerals present in soil, and produce hormones^{[36](#page-11-23),[37](#page-11-24)}, which may ultimately lead to plant growth promotion^{14[,38](#page-11-25)[,39](#page-11-26)}. Some strains of *Trichoderma* spp. form endophytic relationship with roots, and develop symbiotic association with plants^{13[,40](#page-11-27)}. With such association, plants are able to efficiently use the water and nutrients in soil⁴¹, and are able to survive under various stress conditions such as nutrition deficiency, water stress, etc. $42,43$.

Khan et al[.44](#page-11-31) conducted a study in which the application of *T. harzianum* signifcantly enhanced plant height, root length, and shoot biomass of rice cv. Swarna Sub-1, under water-defcient conditions. Similarly, Haque et al.⁴⁵ demonstrated that *T. harzianum* treated rice plants exhibited improved water-use efficiency, enhanced

Table 6. Efects of *Trichoderma* spp. against diferent water stress levels on the grain yield (gm/plant) of rice cultivar Swarna sub-1. Values are mean of 5 replicates. Values in the parenthesis are percent increase over control. Values (means) followed by diferent alphabets within a column are signifcantly diferent at P≤0.05 according two-factor ANOVA and Tukey HSD test.

7

Figure 3. The transpiration rate in the leaves of rice plants under different water stress levels. Error bars show standard error.

photosynthetic activity, and increased grain yield compared to non-treated plants during drought stress. *Trichoderma* spp. enhance plant health and stimulate the synthesis of plant growth-promoting substances such as auxins and cytokinins²⁸, which play crucial roles in root development, nutrient uptake, and overall plant growth^{46[,47](#page-11-34)}. Additionally, *T. harzianum* triggers the expression of stress-responsive genes in rice, activating defence mechanisms and antioxidant systems that counteract the detrimental effects of water stress. The present study has demonstrated that soil application of *T. harzianum* AMUTHZ82 or *T. viride* AMUTVR73 may efficiently subside the impact of water stress at least up to 50% water availability levels and may enhance the plant growth and biomass production by 17–21%.

Figure 5. Efects of seed treatment of *Trichoderma* isolates on the total chlorophyll (chlorophyll A and B) content in the leaves of rice plants under diferent water stress levels. Error bars show standard error. Where CH=Chlorophyll.

Conclusion

The rice cv. Swarna Sub-1 was found sensitive to drought (water stress), and exhibited significant reductions in the plant growth, biomass production, leaf pigments, photosynthesis etc. grown under water stress conditions (75, 50 and 25% water of the recommended amount) in comparison to plants received normal irrigation (100% water availability of the recommended level). The soil application of *Trichoderma* spp. especially *T. harzianum* AMUTHZ82 and *T. viride* AMUTVR73 subsided the negative efect of water stress on rice plants, and promoted the shoot and root growth and biomass production of rice. The present study has demonstrated that soil application of *T. harzianum* AMUTHZ82 or *T. viride* AMUTVR73 may efficiently reduce the impact of water stress at least up to 50% water availability levels and may enhance the plant growth and biomass production by 17–21%.

Materials and methods

Seeds of rice cv. Swarna Sub-1 were procured from Regional Centre, International Rice Research Institute, New Delhi, India.

Isolation and characterization of *Trichoderma* **species**

The local isolates of different *Trichoderma* spp. were isolated from rice fields using the serial dilution method on selective medium²¹. The corresponding GPS locations (latitude and longitude) of sampling sites were given in Table [7](#page-9-0). The plates after inoculation were kept in an incubator at 25 ± 2 °C for 10 days. However, the fungus almost fully colonized the medium in 7 days. Afer incubation the *Trichoderma* colonies were pure cultured using the same culture medium and conditions. Tereafer, morphological characters of the *Trichoderma* isolates were studied to identify the species, followed by molecular characterization to confirm the identity⁴⁸. The DNA of *Trichoderma* species was sequenced by Macrogen® Incorporation, South Korea, and the homology search was carried out using BLAST on the 18s rRNA-ITS region gene sequences to fnd and obtain the closest neighbour sequences from the NCBI database^{[49](#page-12-1)}. The MEGA-X software was used to access the Clustal-W alignment tool, and used to align all of the sequences. Simple comparison with the database of known species/isolates of *Trichoderma harzianum*, *T. viride, T. virens, T. hamatum* and *T. konigii* allowed the identifcation of the species. Te UPGMA algorithm included in MEGA-X was used to create the phylogenetic tree (Fig. [1](#page-1-0)). The sequences have been deposited to the NCBI database and GenBank accession numbers listed in Table [1](#page-2-0).

Mass multiplication of *Trichoderma* **spp.**

Eleven isolates of *Trichoderma* spp. as enlisted in Table [1](#page-2-0) were mass cultured on sorghum grains in conical fasks. The seeds were soaked for 12 h. in distilled water containing sucrose (5%) and chloramphenicol (0.0003%). The seeds were kept in 500 ml capacity conical flasks and sterilized at 15 kg/cm² steam pressure at 121 °C for 20 min. Next day the flasks were inoculated with the fungus, and incubated in an incubator for 10 days at 25 ± 2 °C. The sorghum seeds colonized by *Trichoderma* spp. were crushed with distilled water in an electrical grinder. The fungal suspension containing 10 g *Trichoderma* colonized seeds was applied to the soil (1 kg) in each pot a day

before the planting of rice seedlings. For seed treatment, paste of sorghum seeds was applied to the rice seeds (@10 g/kg seeds), 3 h before their sowing in the nursery bed.

Treatments and plant cultures

The seedlings of rice cultivar Swarna Sub-1 were transplanted in the two sets of pots (1 seedling/pot) filled with sterilized soil and FYM (4:1). In one set of pots *Trichoderma* isolates were applied as soil application (@10 g/ kg soil), whereas, in other set rice seedlings raised from *Trichoderma* treated seeds were planted. Four watering treatments viz., 100, 75, 50 and 25% of recommended irrigation were maintained for each isolate of *Trichoderma* spp. with fve replicate/pots. Under 100% watering, the pots were watered regularly on alternate days. For 75% watering pots were watered every 4th day, and for 50% and 25% watering every 6th and 8th day, respectively. Similar treatments for drought have also been maintained in other researches⁵⁰. Further, the soil moisture percentage is an important matrix for determining the drought level, but it is very difficult to use it as a treatment because of it changes with the progress of time. Since, there were 11 *Trichoderma* isolates, a total of 480 Pots were maintained, which were arranged in a complete randomized block design on the roof top receiving uniform sunlight. At maturity, 3 months afer transplanting, the plants were harvested to measure plant length (shoot and root) and fresh weight per plant. The treatments for each *Trichoderma* isolate for soil application or seed treatment were as follows.

- $T_1 =$ Plant + 100% watering (control)
- T_2 =Plant + 75% watering
- T_3 =Plant + 50% watering
- \overline{T}_4 =Plant + 25% watering
- **T5**=Plant+100% watering+*Trichoderma* isolate
- **T6**=Plant+75% watering+*Trichoderma* isolate
- T_7 =Plant + 50% watering + *Trichoderma* isolate
- T_8 =Plant + 25% watering + Trichoderma isolate

Statistical analysis

The data obtained from the five replicates were used to calculate the means. However, replicate data were used to analyze least significant difference (LSD at *P*≤0.05) with the help of Agricolae Package of R software^{[51](#page-12-3)}. Tukey HSD test was employed to mark signifcantly diferent treatments in the Tables. Two factor ANOVA was used to evaluate the data on plant growth variables.

Ethical statements

Both the authors have approved the content and authorship of the submitted manuscript, and all prevailing local, national, and international regulations and conventions, and scientifc and ethical practices have been respected.

Plant compliance statement

The studies in this research article (plant field studies and experimental research, as well as the collection of plant material) were undertaken following laws and regulations pertinent to the institution.

Data availability

Both authors shall provide the datasets utilized and/or analyzed throughout the current work upon reasonable request.

Received: 26 January 2024; Accepted: 18 July 2024 Published online: 30 July 2024

References

- 1. Khan, M.R., Ahmad, I., Shah, M.H. and Ansari, M.S.A. Root-knot nematodes in cereal and pulse crops, and their management by novel biological and biotechnological approaches. In M. R. Khan (eds). *Novel Biological and Biotechnological Applications in Plant Nematode Management*, pp. 299–328 (Springer Nature, Singapore, 2023). ISBN: 978-981-99-2892-7.
- 2. Khan, M.R., Shah, M.H., & Ahamad, F. Relative efectiveness of eight *Trichoderma* species against white mould of pea caused by *Sclerotinia sclerotiorum*. *Indian Phytopathol.* 1–8 (2022).
- 3. FAOSTAT.<https://www.fao.org/faostat/en/#compare>(2022).
- 4. INDIASTAT.<https://www.indiastat.com/table/agriculture/state-season-wise-area-production-productivity-ric/1440252>(2023).
- 5. Khan, M. R., Haque, Z. & Kausar, N. Management of the root-knot nematode *Meloidogyne graminicola* infesting rice in the nursery and crop feld by integrating seed priming and soil application treatments of pesticides. *Crop. Prot.* **63**, 15–25 (2014).
- 6. Al-Turki, A., Murali, M., Omar, A. F., Rehan, M. & Sayyed, R. Z. Exploring the recent advances in PGPR mediated resilience towards interactive efects of drought and salt stress in plants. *Front. Microbiol.* **14**, 1214845. [https://doi.org/10.3389/fmicb.2023.](https://doi.org/10.3389/fmicb.2023.1214845) [1214845](https://doi.org/10.3389/fmicb.2023.1214845) (2023).
- 7. Venuprasad, R., Laftte, H. R. & Atlin, G. N. Response to direct selection for grain yield under drought stress in rice. *Crop Sci.* **47**(1), 285–293 (2007).
- 8. Chepsergon, J., Mwamburi, L. & Kassim, M. K. Mechanism of drought tolerance in plants using *Trichoderma* spp. *Int. J. Sci. Res.* **3**(11), 1592–1595 (2014).
- 9. Shah, S., Wang, D., Shah, F., Alharby, H., Bamagoos, A.A., Almjrashi, A., Alabdallah, A.M., Alzahrani, S.S., AbdElgawad, H., Adnan, M., Sayyed, R.Z., & Shah, H. Comprehensive impacts of climate change on rice production and adaptive strategies in China. *Front Microbiol*. **13**.<https://doi.org/10.3389/fmicb.2022.926059>(2022).
- 10. Lesk, C., Rowhani, P. & Ramankutty, N. Infuence of extreme weather disasters on global crop production. *Nature* **529**(7584), 84–87 (2016).
- 11. Khan, M. R., Majid, S., Mohidin, F. A. & Khan, N. A new bioprocess to produce low cost powder formulations of biocontrol bacteria and fungi to control fusarial wilt and root-knot nematode of pulses. *Biol. Control* **59**(2), 130–140 (2011).
- 12. Mohiddin, F. A., Khan, M. R. & Khan, S. M. Why *Trichoderma* is considered super hero (super fungus) against the evil parasites?. *Plant Pathol. J.* **9**(3), 92–102 (2010).
- 13. Shoresh, M. & Harman, G. E. The relationship between increased growth and resistance induced in plants by root colonizing microbes. *Plant Signal Behav.* **3**(9), 737–739 (2008).
- 14. Harman, G. E., Howell, C. R., Viterbo, A., Chet, I. & Lorito, M. *Trichoderma* species—opportunistic, avirulent plant symbionts. *Nat. Rev. Microbiol.* **2**(1), 43–56 (2004).
- 15. Brotman, Y. *et al. Trichoderma*-plant root colonization: Escaping early plant defense responses and activation of the antioxidant machinery for saline stress tolerance. *PLOS Pathogens* **9**(4), e1003221.<https://doi.org/10.1371/journal.ppat.1003221> (2013).
- 16. Schuster, A. & Schmoll, M. Biology and biotechnology of *Trichoderma*. *Appl. Microbiol. Biotechnol.* **87**, 787–799 (2010).
- 17. Khan, M. R. Nematode biocontrol agents: Diversity and efectiveness against phytonematodes in sustainable crop protection. *Indian Phytopathol.* **69**(4s), 453–463 (2016).
- 18. Afrouz, M. *et al.* Seed bio-priming with benefcial *Trichoderma harzianum* alleviates cold stress in maize. *PeerJ* **1**, e15644. [https://](https://doi.org/10.7717/peerj.15644) doi.org/10.7717/peerj.15644 (2023).
- 19. Doni, F., Isahak, A., Che Mohd Zain, C.R., Mohd Arifn, S., Wan Mohamad, W.N.A., & Wan Yusof, W.M. Formulation of *Trichoderma* sp. SL2 inoculants using diferent carriers for soil treatment in rice seedling growth. *Springerplus* **3**, 1–5 (2014).
- 20. Bae, H. et al. The beneficial endophyte *Trichoderma hamatum* isolate DIS 219b promotes growth and delays the onset of the drought response in *Teobroma cacao*. *J. Exp. Bot.* **60**(11), 3279–3295 (2009).
- 21. Mohammed, R. K. A. & Khan, M. R. Management of root-knot nematode in cucumber through seed treatment with multifarious benefcial microbes under protected cultivation. *Indian Phytopathol.* **74**(4), 1035–1043 (2021).
- 22. Gowtham, H. G. *et al.* Insight into recent progress and perspectives in improvement of antioxidant machinery upon PGPR augmentation in plants under drought stress: *A Review*. *Antioxidants* **11**(9), 1763. <https://doi.org/10.3390/antiox11091763>(2022).
- 23. Marulanda, A., Barea, J. M. & Azcón, R. Stimulation of plant growth and drought tolerance by native microorganisms (AM fungi and bacteria) from dry environments: Mechanisms related to bacterial efectiveness. *J. Plant Growth Regul.* **28**, 115–124 (2009).
- 24. Bernier, J., Atlin, G. N., Serraj, R., Kumar, A. & Spaner, D. Breeding upland rice for drought resistance. *J. Sci. Food Agric.* **88**(6), 927–939 (2008).
- 25. Sudha, A. *et al.* Volatilome profling and evaluation of plant growth stimulation and Anti-ergot activity of VOCs produced by *Trichoderma asperelloides*. *Sydowia* **76**, 93–109. <https://doi.org/10.12905/0380.sydowia76-2024-0093> (2023).
- 26. Contreras-Cornejo, H. A., Macías-Rodríguez, L., Beltrán-Peña, E., Herrera-Estrella, A. & López-Bucio, J. *Trichoderma*-induced plant immunity likely involves both hormonal-and camalexin-dependent mechanisms in *Arabidopsis thaliana* and confers resistance against necrotrophic fungi Botrytis cinerea. *Plant Signal Behav.* **6**(10), 1554–1563 (2011).
- 27. Khan, M. R. & Mohiddin, F. A. *Trichoderma*: its multifarious utility in crop improvement. In *New and Future Developments in Microbial Biotechnology and Bioengineering: Crop improvement through microbial biotechnology* (eds Prasad, R. *et al.*) 263–291 (Elsevier publications, 2018).
- 28. Mir, M. I. *et al.* Multifarious indigenous diazotrophic rhizobacteria of rice (Oryza sativa L.) rhizosphere and their efect on plant growth promotion. *Front. Nutri.* **8**, 781764.<https://doi.org/10.3389/fnut.2021.781764>(2022).
- 29. Sagar, A., Sayyed, R.Z., Ramteke, P.W., Sharma, S., Marraiki, N., Elgorban, A.M., & Syed, A. ACC deaminase and antioxidant enzymes producing halophilic *Enterobacter* sp. PR14 promotes the growth of rice and millets under salinity stress. *Physiol. Mol. Biol. Plants* **26**, 1847–54. <https://doi.org/10.1007/s12298-020-00852-9>(2020).
- 30. Chaves, M. M., Flexas, J. & Pinheiro, C. Photosynthesis under drought and salt stress: Regulation mechanisms from whole plant to cell. *Ann. Bot.* **103**(4), 551–560 (2009).
- 31. Khan, M. R. & Rizvi, T. F. Effect of elevated levels of CO₂ on powdery mildew development in five cucurbit species. *Sci. Rep.* 10, 4986 (2020).
- 32. Tardieu, F., & Simonneau, T. Variability among species of stomatal control under fuctuating soil water status and evaporative demand: Modelling isohydric and anisohydric behaviours. *J. Exp. Bot*. 419–432 (1998).
- 33. Farooq, M., Wahid, A., Kobayashi, N.S.M.A., Fujita, D.B.S.M.A., & Basra, S.M.A. Plant drought stress: efects, mechanisms and management. *Sustain. Agric.* 153–188. (2009).
- 34. Rizvi, T. F. & Khan, M. R. Biological and physiological responses of root-knot disease development on fve cucurbits exposed to diferent concentrations of sulfur dioxide. *Toxics* **11**, 334 (2023).
- 35. Rubio, M. B., Dominguez, S., Monte, E. & Hermosa, R. Comparative study of *Trichoderma* gene expression in interactions with tomato plants using high-density oligonucleotide microarrays. *Microbiology* **158**(1), 119–128 (2012).
- 36. Suriani, N.L., Suprapta, D., Novizar, N., Parwanayoni, N., Darmadi, A., Dewi, D., Sudatri, N., Ahmad, F., Sayyed, R.Z., Syed, A., Elgorban, A.M., Bahkali, A.H., Enshasy, H.E., & Dalin, D.J. A mixture of piper leaves extracts and rhizobacteria for sustainable plant growth promotion and biocontrol of blast pathogen of organic bali rice. *Sustainability*. 12, 8490. [https://www.mdpi.com/](https://www.mdpi.com/2071-1050/12/20/8490) [2071-1050/12/20/8490](https://www.mdpi.com/2071-1050/12/20/8490) (2020).
- 37. Khan, M.R., & Rizvi, T.F. Nanotechnology, a tool for reducing pesticide input in plant protection. In *Bio-intensive approaches: application and efectiveness in plant diseases management*, pp. 225–242 (Today and Tomorrow Publishes, New Delhi, 2018).
- 38. Basu, A., Prasad, P., Subha, N.D., Sadaf, K., Sayyed, R.Z., Reddy, M.S., & Hesham, E.E. Plant growth promoting rhizobacteria (PGPR) as green bioinoculants: recent developments, constraints, and prospects. *Sustainability* **13**, 1140. [https://www.mdpi.com/](https://www.mdpi.com/2071-1050/13/3/1140) [2071-1050/13/3/1140](https://www.mdpi.com/2071-1050/13/3/1140) (2021).
- 39. Khan, M.R., & Mohiddin, F.A. Biocontrol strategies for nematode management, an overview. In M. R. Khan (eds). *Novel Biological and Biotechnological Applications in Plant Nematode Management,* pp 116–141 (Springer Nature, Singapore, 2023). ISBN: 978-981-99-2892-7.
- 40. Enshasy, H.E.A., Ambehati, K.K. Ashraf, F., Ramchuran, S. Sayyed, R.Z. Amalin, D. Dailin, D.J., & Hanapi, S.Z. *Trichoderma*: Biocontrol agents for promoting plant growth and soil health. In Agriculturally important fungi for sustainable agriculture: Vol 2: *Functional annotation for crop protection*, pp. 239–259. (2020).
- 41. Mastouri, F., Björkman, T. & Harman, G. E. Seed treatment with *Trichoderma harzianum* alleviates biotic, abiotic, and physiological stresses in germinating seeds and seedlings. *Phytopathol* **100**(11), 1213–1221 (2010).
- 42. Contreras-Cornejo, H. A., Macías-Rodríguez, L., Cortés-Penagos, C. & López-Bucio, J. *Trichoderma virens*, a plant benefcial fungus, enhances biomass production and promotes lateral root growth through an auxin-dependent mechanism in Arabidopsis. *Plant Physiol.* **149**(3), 1579–1592 (2009).
- 43. Contreras-Cornejo, H. A., Macías-Rodríguez, L. & Cortés-Penagos, C. Mechanism involved in *Trichoderma harzianum*-mediated maize drought tolerance. *Physiol. Mol. Plant Pathol.* **88**, 41–53 (2014).
- 44. Khan, M. R. *et al.* Management of root-rot disease complex of mungbean caused by *Macrophomina phaseolina* and *Rhizoctonia solani* through soil application of *Trichoderma* spp. *Crop. Prot.* **119**, 24–29 (2019).
- 45. Haque, M. Z., Anwar, M., Zaman, M. & Hasanuzzaman, M. *Trichoderma harzianum* enhances drought tolerance in rice by modulating physiological and biochemical attributes. *J. Plant Growth Regul.* **40**, 764–778 (2021).
- 46. Vos, C. M., Yang, Y. & Harris, A. R. Exploring the impact of *Trichoderma* on nutrient acquisition by plants. *Front. Microbiol.* **12**, 649403 (2021).
- 47. Khan, M.R., Ahamad, F., Rizvi, T. F., & Akram, M. Novel nano-materials and nano-formulations for nematode management. novel biological and biotechnological applications in plant nematode management. In M. R. Khan (eds). *Springer Nature*, Singapore. (2023). Pp. 256–279. ISBN: 978-981-99-2892-7
- 48. Vinale, F. *et al. Trichoderma* plant pathogen interactions. *Soil Biol. Biochem.* **40**, 1–10 (2008).
- 49. El-Bondkly, A.M. Sequence analysis of industrially important genes from *Trichoderma*. In Biotechnology and biology of *Trichoderma* (pp. 377–392). *Elsevier*. (2014).
- 50. Yang, X., Wang, B., Chen, L., Li, P. & Cao, C. Te diferent infuences of drought stress at the fowering stage on rice physiological traits, grain yield, and quality. *Sci. Rep.* **9**(1), 3742 (2019).
- 51. De Mendiburu, F., Simon, R. Agricolae-ten years of an open source statistical tool for experiments in breeding, agriculture and biology (No. e1748) *Peer J.* (2015).

Author contributions

M.R.K. conceived the idea and critically checked the MS; M.H.S. executed the experiments, analyze the data and prepared the manuscript. Both authors reviewed the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

Correspondence and requests for materials should be addressed to M.R.K.

Reprints and permissions information is available at [www.nature.com/reprints.](www.nature.com/reprints)

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional afliations.

Open Access Tis article is licensed under a Creative Commons Attribution-NonCommercial- \odot 1 NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modifed the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by-nc-nd/4.0/>.

 $© The Author(s) 2024$