

OPEN

Comparative genomics of *Salmonella enterica* serovar Enteritidis ST-11 isolated in Uruguay reveals lineages associated with particular epidemiological traits

Bruno D'Alessandro¹, Victoria Pérez Escanda¹, Lucía Balestrazzi¹, Florencia Grattarola², Andrés Iriarte¹, Derek Pickard³, Lucía Yim¹, José Alejandro Chabalgoity¹ & Laura Betancor^{1,2*}

Salmonella enterica serovar Enteritidis is a major cause of foodborne disease in Uruguay since 1995. We used a genomic approach to study a set of isolates from different sources and years. Whole genome phylogeny showed that most of the strains are distributed in two major lineages (E1 and E2), both belonging to MLST sequence type 11 the major ST among serovar Enteritidis. Strikingly, E2 isolates are over-represented in periods of outbreak abundance in Uruguay, while E1 span all epidemic periods. Both lineages circulate in neighbor countries at the same timescale as in Uruguay, and are present in minor numbers in distant countries. We identified allelic variants associated with each lineage. Three genes, *ycdX*, *pduD* and *hsdM*, have distinctive variants in E1 that may result in defective products. Another four genes (*ybiO*, *yiaN*, *aas*, *aceA*) present variants specific for the E2 lineage. Overall this work shows that *S. enterica* serovar Enteritidis strains circulating in Uruguay have the same phylogenetic profile than strains circulating in the region, as well as in more distant countries. Based on these results we hypothesize that the E2 lineage, which is more prevalent during epidemics, exhibits a combination of allelic variants that could be associated with its epidemic ability.

Salmonella is a major cause of human foodborne disease worldwide. A singular epidemiological feature of human salmonellosis is that one particular serovar can become prevalent over the others, but the prevalent serovar may change over time. Previous to the 1980's, *Salmonella enterica* serovar Typhimurium was the most commonly isolated serovar worldwide, but then at the beginning of the 1990's *S. enterica* serovar Enteritidis emerged as the most common cause of human salmonellosis, first in Europe and then in many other countries around the world¹⁻⁶. The reasons for this serovar shift are still not fully understood.

Several studies have addressed the phylogenetic diversity within *S. enterica* serovar Enteritidis, and suggested that diversification events could be related to its epidemiological features⁷⁻¹⁰. The work of Allard *et al.* first reported the use of whole genome sequencing (WGS) and single nucleotide polymorphisms (SNP) analysis to address molecular epidemiology of a set of isolates previously indistinguishable by other techniques. Deng *et al.* suggested that serovar Enteritidis diversified from a few major lineages spread worldwide, and associated some lineages with geographic and epidemiological characteristics. Feasey *et al.* found a strong correlation between prophage content and accessory genome features with the establishment of a new epidemiological course. In our studies, we found that the acquisition of a new prophage was probably determinant for the onset of a particular lineage⁸. The vast majority of *S. enterica* serovar Enteritidis isolates worldwide belong to the eBurst Group 4 (EBG4). Achtman *et al.* and more recently our group described that among EBG4, multi locus sequence type

¹Departamento de Desarrollo Biotecnológico, Instituto de Higiene, Facultad de Medicina, Universidad de la República, Av. Alfredo Navarro 3051, CP, 11600, Montevideo, Uruguay. ²Departamento de Bacteriología y Virología, Instituto de Higiene, Facultad de Medicina, Universidad de la República, Av. Alfredo Navarro 3051, CP, 11600, Montevideo, Uruguay. ³Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, Hinxton, Cambridge, UK. *email: laurabet@higiene.edu.uy

(MLST) ST11 is the major ST and constitutes a polyphyletic lineage from which several other minor lineages emerged^{8,11,12}.

In Uruguay, until 1994 *S. enterica* serovar Enteritidis was only sporadically isolated, but in 1995 an important outbreak of human salmonellosis associated to this serovar occurred. Since then it became the most prevalent serovar in the country despite some periodical declining¹. Five different epidemiological periods can be distinguished, as related to the prevalence of serovar Enteritidis in the country: a pre-epidemic period, the first epidemic (1995–2004), a declining (2005–2008), a second epidemic period (2009–2013) and further declining afterward (see Fig. 1(a)).

In previous studies, we found that the two oldest pre-epidemic isolates (31/88 and 8/89) belong to a unique genetic type by RAPD, PFGE, and MLST and have impaired behavior in different models of infection^{1,8,13}. In spite of these observations, we could not find a correlation between genetic features and different pathogenic or epidemiological abilities among the isolates. In the present study, we analyze the genomic sequences of 61 *S. enterica* serovar Enteritidis strains isolated in Uruguay between 1988 and 2017 from different sources by comparative genomics and SNP based phylogenetic analysis. We extended the study to include strains from Argentina and Brazil. An analysis of the genomic differences between the lineages that have been circulating since the beginning of the *S. enterica* serovar Enteritidis epidemic period is presented.

Methods

Bacterial strains, culture and DNA extraction. A total of 61 *S. enterica* serovar Enteritidis strains isolated in Uruguay from different sources and periods were selected from the collection of the National Salmonella Center (Instituto de Higiene, Universidad de la República). All strains were previously characterized by our group using different genetic and phenotypic methods^{13,14}. Strain names, source and date of isolation are shown in Supplementary Table 1. The strains were recovered from freeze stocks (kept at -80°C in LB broth with 16% glycerol) by culture in LB agar (Sigma-Miller). Liquid cultures used for DNA extraction were performed in LB broth at 37°C with shaking. DNA extraction and purification were performed using commercial purification kits (DNeasy Blood & Tissue Kit, Qiagen).

Genome sequencing and assembly. Genome sequencing was performed using Illumina technology, and the sequences were deposited in the NCBI database. The total number of produced paired-end reads for each strain ranged from 0.66×10^6 to 7.9×10^6 , with a read size of 100, 277 and 76 for platforms HiSeq. 2000, MySeq v3, and Illumina Genome Analyzer II, respectively. Generated reads were trimmed using sickle (available at github.com/najoshi/sickle), with a phred score threshold of 30 (Q30). Quality was finally checked using FastQC (www.bioinformatics.babraham.ac.uk/projects/fastqc/). For platform and sequencing details see Supplementary Table 1. *De novo* assembly was performed with Spades (version 3.6.1)¹⁵ using a pre-assembly approach with Velvet (version 1.2.10)¹⁶. The first assembly was done using Velvet with default parameters. The final assembly was performed in Spades using a range of k-mer sizes between 29 and the maximum read size minus one. Velvet-generated contigs were used as a reference with the “--untrusted-contigs” option. On average 97.7% of the generated reads were mapped and the mean observed nucleotide coverage ranged from 27 to $254 \times$. N50 contig size has an average size of 447,312. Assembly stats are summarized in Supplementary Table 1. All the Uruguayan genomes are available in the NCBI BioProject database with accession numbers PRJEB2130 and PRJEB4649 (Supplementary Table 1).

Comparative genomics. The genome of phage type 4 strain P125109, (GenBank ID AM933172.1, NCBI refseq accession NC_011294.1), was used as reference when needed. All the genomic sequences were annotated using RASTtk¹⁷ and verified with EnteroBase annotation pipeline. Comparative SNP analysis was performed using NUCmer (NUCleotide MUMmer version 3.1) and local blast (version 2.2.30)¹⁸. NUCmer was run with the parameters “-maxmatch” and the minimum length of a single exact match set to 12 (-l parameter). SNPs were obtained using the show-snps module with the -C option, so excluding all SNPs from repeats.

Phylogeny figures were generated using iTOL¹⁹. Figures showing the alignment between variants of genes were generated through BLASTn alignments using EasyFig²⁰, exported to svg images and edited using Inkscape software (inkscape.org).

For phylogenetic analysis all fastq files were submitted to EnteroBase which automatically performs a pipeline of assembly, annotation and some *in silico* typing schemes including Achtman 7 gene MLST and ribosomal MLST (rMLST) (<http://enterobase.warwick.ac.uk>)¹². EnteroBase tools were used to perform a whole genome SNP phylogeny, using the genome of strain P125109 as reference. The EnteroBase algorithm performs *de novo* assembly from reads and uses LAST software to call the SNPs from the assembled genomes, filters out SNPs from repetitive regions and low quality variants, and builds a SNP matrix which is used as input to perform a RAxML maximum likelihood phylogeny²¹. For the SNP tree we considered only the sites present in 100% of the genomes.

Genomes from strains isolated in Argentina and Brazil that were included in phylogenetic analysis are shown in Supplementary Table 2 and were selected from the available genomes in EnteroBase.

Results and Discussion

Comparative genomics. First we compared the 61 Uruguayan isolates using SNP analysis with the P125109 genome as reference. This analysis showed that the two oldest isolates have more than 600 SNPs of difference compared to the reference genome (607 and 652 for 31/88 and 08/89 respectively), whereas all other 59 isolates have less than two hundred SNPs (from 60 to 166) (Supplementary Table 1). On the other hand, all Uruguayan genomes share 17 SNPs of difference with P125109 genome.

A phylogenetic analysis based on SNPs revealed the existence of two clades that we named E1 and E2 (Fig. 1(b)) that comprise 57 out of the 61 strains. These 57 strains belong to the major *S. enterica* serovar

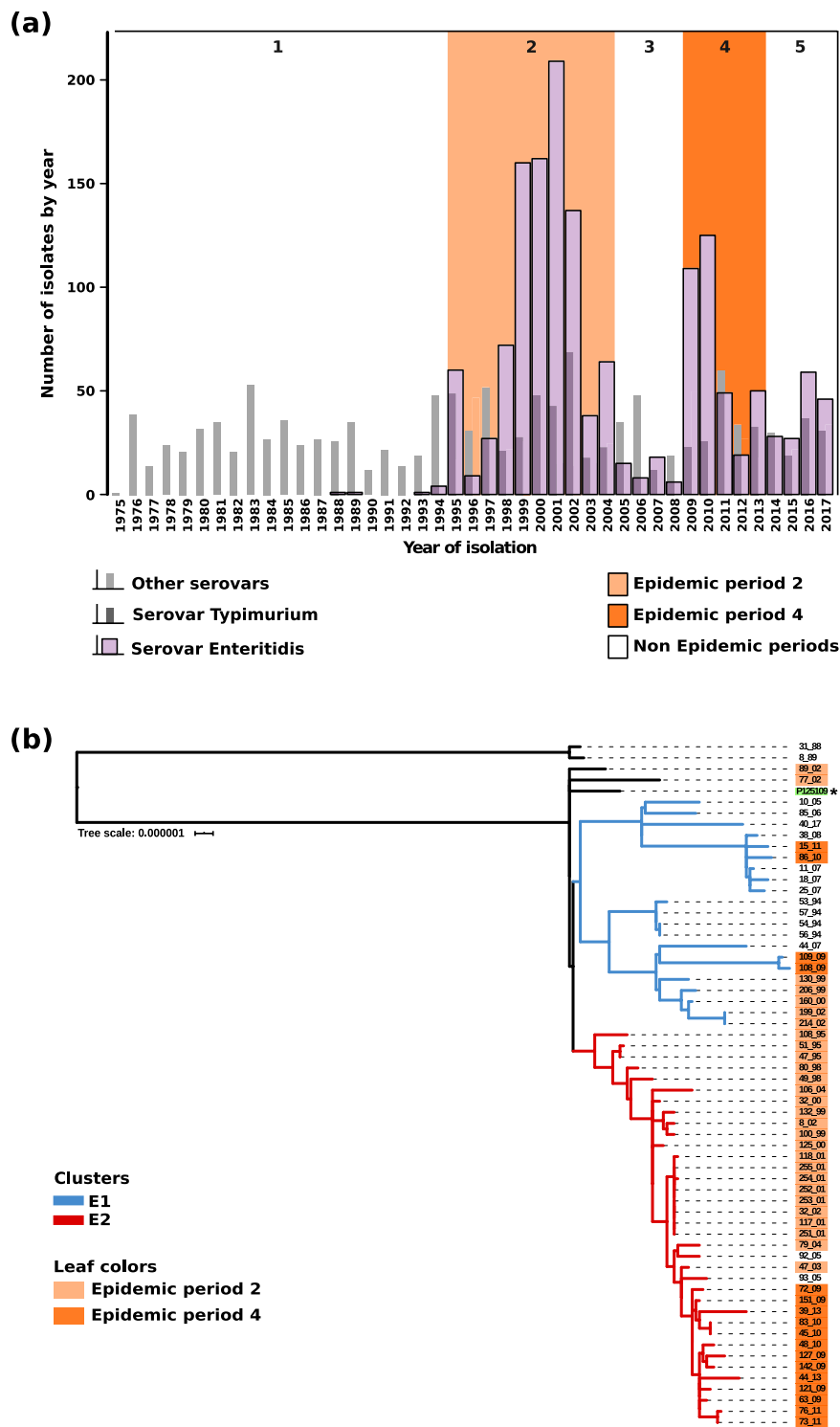


Figure 1. (a) Distribution of *S. enterica* serovars received at the National *Salmonella* Center over the time (1975 to 2017). The 5 epidemiological periods of isolation are indicated with numbers. In white: non epidemic periods (1, 3 and 5). In orange: epidemic periods (2 and 4). (b) Phylogenetic tree showing the two clusters among Uruguayan *S. enterica* serovar Enteritidis genomes (E1 in blue, E2 in red and other strains in black). Highlight over the strain designation indicate the period of isolation as explained above. The genome of reference (P125109) is highlighted in green and marked with an asterisk. The isolates are named using a number followed by an underscore and the last two digits of the year of isolation. Scale bar units represents changes per variant site.

Enteritidis MLST sequence type 11 (ST-11) and have been co-circulating in the country since 1994. The remaining four isolates (31/88, 8/89, 77/02 and 89/02) did not group with any of the two clades (Fig. 1(b)). We previously reported that the two oldest isolates (31/88 and 8/89) belong to the MLST sequence type 1974 (ST1974) and that ST1974 originated from ST11 after the acquisition of a characteristic prophage⁸. The other two isolates, 77/02 and 89/02, belong to ST11 as the majority of the strains.

The E1 clade comprises 21 strains that span all five epidemiological periods, and were isolated from food, animal or human infections. Instead, E2 clade is represented by 36 isolates of which 34 were isolated in the periods associated to epidemics (Fig. 1(a,b)). Hence, it is reasonable to hypothesize that E2 may constitute a lineage that was responsible for the *S. enterica* serovar Enteritidis epidemics in the country.

Previously we reported a phylogenetic analysis of 203 strains, representing intra-serovar diversity among *S. enterica* serovar Enteritidis EBG4 isolates worldwide, that included 6 of the 61 Uruguayan isolates used in the present study⁸. As we found now, half of these 6 strains are E1 and half are E2 (E1: 53/94, 206/99 and 214/02; E2: 8/02, 251/01 and 253/01), and each of these two groups is closely related with strains from Europe and North America. This relation is showed in Supplementary Fig. S1, containing the previously reported phylogeny with a zoom on the clade formed by these strains. This suggests that E1 and E2 may have a broad circulation pattern.

In order to further support this assumption we performed a new phylogenetic analysis, this time with focus in strains isolated in the region, including 12 from Argentina, 122 from Brazil and the same 61 genomes from Uruguay (Fig. 2(a)). Interestingly we found that all these strains clustered in a highly similar way of what we found among Uruguayan isolates, i.e. E1 cluster now contains strains from Argentina and Brazil whereas E2 cluster contains strains from Brazil (103 in E1, and 62 in E2). Given the low number of genomes from Argentina available in Enterobase, we can not exclude the possibility that lineage E2 was circulating in this country. Only 30 strains from all three countries grouped outside E1 and E2, of which 24 were isolated before 1994 (Fig. 2(a), Supplementary Table 2). All in all our results strongly suggest that E1 and E2 strains were introduced in the region around 1994 and have been circulating ever since in Uruguay, Brazil, Argentina, Europe and North America, supporting the idea that both constitute lineages of the serovar Enteritidis.

We then performed a comparative genomics analysis aimed to identify genetic differences associated with lineages E1 and E2, including 165 genomes: 57 from Uruguay, 11 from Argentina and 97 from Brazil (Fig. 2(a)).

Allelic variants among E1 and E2 lineages. As we did not find differences in accessory genome between E1 and E2 lineages, we looked for SNP differences among the 165 selected strains. This analysis showed that the main differences between E1 and E2 are located in three genes: *ycdX*, *pduD* and *hsdM*. These genes are interrupted in E1 genomes when compared to their counterparts in the E2 genomes. In addition, we found allelic variants in other four genes, *ybiO*, *yiaN*, *aas* and *aceA*, that are specific for each lineage (Fig. 2(a,b); Table 1).

All the 103 E1 strains (11 from Argentina, 71 from Brazil and 21 from Uruguay) show a 4 base pair insertion located in gene *ycdX* (SEN1912) as compared to the reference and to all E2 strains (Table 1). This insertion introduces a premature stop codon, which may produce a pseudo-gene or a truncated version of the protein (Fig. 2(b) and Table 1). The *ycdX* gene encodes an hydrolase that belongs to the superfamily of PHP proteins, that contain a NH2-terminal php domain (polymerase histidinol phosphatase). This domain is characteristic of the DNA polymerase III alpha subunit of bacteria and is involved in the proofreading function²². It has been suggested that YcdX is involved in the DNA repair pathways in *E. coli* and could act as a nuclease or a phosphatase in these pathways²³. Recently, Wang *et al.* found two allelic variants for this gene (non synonymous SNP) between *S. enterica* serovar Enteritidis isolates with markedly different abilities to survive in egg white²⁴. Inoue *et al.* showed that mutation in *ycdY* (the adjacent gene in the *ycdWXYZ* operon) caused repression of swarming in *E. coli*²⁵.

All the strains from the E2 lineage have the same allelic variant for *pduD* gene (SEN2039) while 102 out of the 103 strains from E1 lineage show a base substitution that introduce a stop codon, similar to the one observed for *ycdX* (Table 1). The remaining strain (i.e. 11/07 from Uruguay) completely lacks the *pduD* gene (Fig. 2(b) and Table 1). *pduD* encodes for a propanediol dehydratase, part of the *pdu* operon which encodes all the required proteins for the 1,2-propanediol catabolism. This operon that was acquired by lateral gene transfer in *Salmonella*^{26,27} is disrupted in different serovars that cause invasive extra-intestinal infections in human such as Typhi and Paratyphi A amongst others²⁸. Faber *et al.* reported that the functionality of this operon can be important to compete with intestinal microbiota, as 1,2 propanediol is a metabolic product in the anaerobic gut environment that can be used by *Salmonella* when coupled to the tetrathionate anaerobic respiration²⁹. It is proposed that ability to use 1,2 propanediol allow the expansion of the pathogen population in the gut increasing the excretion in feces, which is an important factor for epidemic dissemination²⁹.

Furthermore, all E2 strains have the same allelic variant for the *hsdM* gene (SEN4290) but among the E1 strains there are several variants for this gene (Fig. 2(b) and Table 1). Most of the E1 strains contain variants of *hsdM* that would produce either a truncated version (4 out of the 6 variants) or a non synonymous substitution for the protein. Only three strains in the E1 (including the Uruguayan 44/07) have the same variant as E2, suggesting that only in E1 strains *hsdM* is prone to accumulate mutations (Fig. 2(b)). The *hsdM* gene encodes a DNA methylase involved in type I restriction-modification system (RM) in enterobacteria^{30,31}. Previous reports linked restriction modification systems type 2 with pathogenicity in *Salmonella* due to epigenetic effects on the virulence gene expression³². Silva *et al.* showed that mutations in the RM genes produce an impaired phenotype in the mouse model of invasive salmonellosis³³. Type 1 RM systems were also described as involved in pathogenicity of *Yersinia pseudotuberculosis*³⁴.

Given that E2 but not E1 have intact copies of *ycdX*, *pduD* and *hsdM*, and considering that these genes have been previously implicated in *Salmonella* pathogenicity, it can be taken to suggest that these three genes may be relevant for the epidemic ability of the E2 strains. We also looked for the allelic variants in the global phylogeny representing the intra-serovar diversity (Supplementary Fig. 1 and ref. ⁸) and found that the predominant allele is the E2 allele in all cases, whereas the disrupted variants are specific for E1 (data not shown).

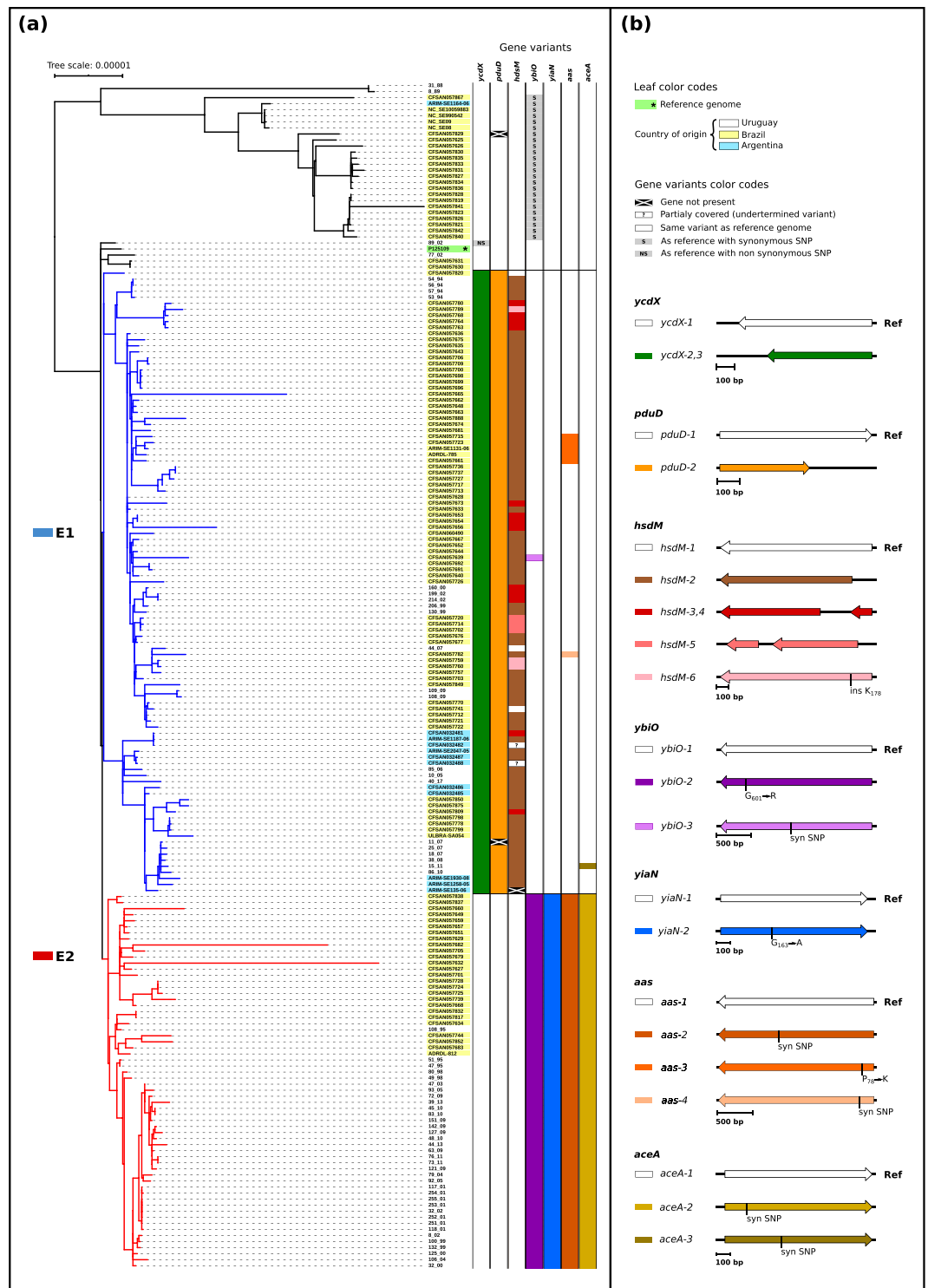


Figure 2. (a) Phylogenetic tree of the 195 genomes including 61 from Uruguay (white leaves), 12 from Argentina (light blue leaves) and 122 from Brazil (yellow leaves). Branches of E1 lineage are colored in blue. Branches of E2 lineage are indicated in red. Lanes at right represent with different colors the allelic variants for *ycdX*, *pduD*, *hsdM*, *ybiO*, *yiaN*, *aas* and *aceA* respectively. See colour legends inside the (b). Scale bar units represents changes per variant site. (b) Gene alignments for the different allelic variants of *ycdX*, *pduD*, *hsdM*, *ybiO*, *yiaN*, *aas* and *aceA* respectively. Genes are depicted as annotated in the genome containing each variant and aligned to the reference P125109. The scale of the alignment in base pairs (bp) is indicated by horizontal bars. SNPs associated to each variant respect to the reference genome are detailed in Table 1. SNPs causing non synonymous variants are marked as X → Y (in single letter aminoacid code) relative to its position in the gene. Synonymous SNPs are only marked relative to its position in the gene (syn SNP).

Gene	Variant name	SNP	E1	E2	Observations
<i>ycdX</i>	ycdX-1	—	0%	100%	
	ycdX-2	(- → AGCC) ₅₅₂	99%	0%	Frameshift ₁₈₇ ; stop ₁₉₃
	ycdX-3	(- → CCAG) ₅₅₂	1%	0%	Frameshift ₁₈₇ ; stop ₁₉₃
<i>pduD</i>	pduD-1	—	0%	100%	
	pduD-2	(C → T) ₃₉₆	99%	0%	Stop ₁₃₃
<i>hsdM</i>	<i>hsdM</i> -1	—	3%	100%	
	<i>hsdM</i> -2	(- → T) ₁₇₁	77%	0%	Frameshift ₆₀ ; stop ₆₄ ; late start CDS.
	<i>hsdM</i> -3	(- → TT) ₁₇₁	13%	0%	Frameshift ₆₀ ; stop ₆₀ and CDS in 2nd frame
	<i>hsdM</i> -4	(- → T) ₁₇₈ ; (G → T) ₇₉₇	1%	0%	Frameshift ₆₀ ; stop ₆₄ and CDS in 2nd frame
	<i>hsdM</i> -5	(- → T) ₁₇₈ ; (T → -) ₈₆₇	3%	0%	Frameshift ₆₀ ; stop ₆₄ ; late start and truncated CDS.
	<i>hsdM</i> -6	(- → TTT) ₁₇₈	3%	0%	insert Lys ₅₉
<i>ybiO</i>	ybiO-1	—	98%	0%	
	ybiO-2	(T → G) ₁₈₀₃	0%	100%	Gly ₆₀₁ → Arg
	ybiO-3	(G → A) ₁₁₂₅	2%	0%	Synonymous
<i>yiaN</i>	yiaN-1	—	100%	0%	
	yiaN-2	(G → C) ₄₈₉	0%	100%	Gly ₁₆₃ → Ala
<i>aas</i>	aas-1	—	94%	0%	
	aas-2	(T → C) ₁₃₀₁	0%	100%	Synonymous
	aas-3	(C → T) ₂₃₃	5%	0%	(Pro ₇₈ → Lys)
	aas-4	(G → A) ₂₇₀	1%	0%	Synonymous
<i>aceA</i>	aceA-1	—	99%	0%	
	aceA-2	(C → T) ₁₉₈	0%	100%	Synonymous
	aceA-3	(G → A) ₅₄₃	1%	0%	Synonymous

Table 1. SNPs and gene variants of the 7 gene markers for the lineages E1 and E2. The variant names correspond to those showed in Fig. 2(b). SNP column show the nucleotidic change respect to the reference genome in brackets (bases as in the leading strand of the reference genome P125109) and its relative position to the gene start in subscript. Columns E1 and E2 indicate the percentage of genomes that present the corresponding variant in each lineage. Note that the total number of E1 genomes is 103 and the total number of E2 genomes is 62.

As mentioned above, another four genes *ybiO*, *yiaN*, *aas* and *aceA* were found with minor sequence differences between both lineages. These four genes present a single variant in E2 strains whereas E1 are mostly identical to P125109 reference strain.

All E2 strains present a particular allelic variant of the *ybiO* gene (SEN0772) due to a substitution that produce a Gly(601) → Arg in the protein (Fig. 2(a,b), Table 1). The *ybiO* gene encode for a putative mechanosensitive channel protein involved in osmotic adaptation that is activated by hypo-osmotic shock in *E. coli*³⁵. It was previously reported that there are two allelic variants for this gene among *S. enterica* serovar Enteritidis isolates that have differing ability to survive in egg white²⁴. For *yiaN* (SEN3494) (L-dehydroascorbate transporter large permease subunit, a putative membrane protein) all E2 genomes present the same allele that differs in Gly(163) → Ala respect to E1. Similarly, all E2 share the same variant for *aas* (2-acylglycerophosphoethanolamine acyltransferase/acyl-ACP synthetase, SEN2853) and *aceA* (isocitrate lyase, SEN3966) genes. In E1, most of the strains have an *aas* gene identical to the reference, and a few strains present either a non synonymous substitution (5 strains) or a synonymous substitution (1 strain) (Fig. 2(a), Table 1). Similarly, the *aceA* gene is identical to the reference in all but one of the E1 genomes.

In summary, we found that the E2 alleles for *ybiO*, *yiaN*, *aas* and *aceA* are specific for this lineage.

Conclusions

Two different lineages have been circulating in Uruguay after the start of the epidemic period. These same two lineages also circulated in Brazil in similar periods of time as well as in Europe and North America. Our results are in agreement with those recently published by Campioni *et al.* that report the existence of a shift in the genomic profile of *S. enterica* serovar Enteritidis circulating in South America after 1994³⁶. Based on all these results, we hypothesize that around 1994 phage type 4 strains (likely from E1 and E2 lineages) were introduced in the region and successfully spread causing epidemics. These introduced strains belong to one of the sub-clades described by Feasey as part of the global epidemic *S. enterica* serovar Enteritidis clade¹⁰.

Further, as reported here, at least in Uruguay, the E2 lineage is clearly associated with epidemics. This lineage presents a combination of allelic variants that could be associated with its epidemic ability. The fact that genes *ycdX*, *pduD* and *hsdM* are disrupted in E1 strains that come from different regions and periods of isolation is suggestive of a biological relevance for conserving this disruption. The disrupted version of these genes is a genetic marker for the E1 lineage, and we propose that it might be related to impaired epidemic ability. On the other hand, the allelic variants of another four genes (*ybiO*, *yiaN*, *aas*, *aceA*) are specific markers for the E2 lineage, and could be useful for tracking the presence of this lineage in further epidemic events.

Data availability

All the Uruguayan genomes are available in the NCBI BioProject database with accession numbers PRJEB2130 and PRJEB4649, with the specific accession for each sample showed in Supplementary Table 1. The accession numbers for all the genomes from regional strains are showed in Supplementary Table 2.

Received: 11 April 2019; Accepted: 7 February 2020;

Published online: 27 February 2020

References

1. Betancor, L. *et al.* Prevalence of *Salmonella enterica* in poultry and eggs in Uruguay during an epidemic due to *Salmonella enterica* serovar Enteritidis. *J. Clin. Microbiol.* **48**, 2413–23 (2010).
2. Cogan, T. A. & Humphrey, T. J. The rise and fall of *Salmonella* Enteritidis in the UK. *J. Appl. Microbiol.* **94**, 114–119 (2004).
3. Herikstad, H., Motarjemi, Y. & Tauxe, R. V. *Salmonella* surveillance: A global survey of public health serotyping. *Epidemiol. Infect.* **129**, 1–8 (2002).
4. Rodrigue, D. *et al.* Outbreaks of *Salmonella enteritidis* Infections in the United States, 1985–1991. *J. Infect. Dis.* **169**, 547–552 (2011).
5. Poppe, C. In *Salmonella enterica serovar Enteritidis in human and animals* (eds. Saeed, A. M., Gast, R. K., Potter, M. E. & Wall, P. G.) 3–18 (Iowa State University Press, 1999).
6. Schroeder, C. M. *et al.* Estimate of illnesses from *Salmonella* Enteritidis in eggs, United States, 2000. *Emerg. Infect. Dis.* **11**, 113–115 (2005).
7. Allard, M. W. *et al.* On the Evolutionary History, Population Genetics and Diversity among Isolates of *Salmonella* Enteritidis PFGE Pattern JEGX01.0004. *PLoS One* **8** (2013).
8. D'Alessandro, B. *et al.* A novel prophage identified in strains from *Salmonella enterica* serovar Enteritidis is a phylogenetic signature of the lineage ST-1974. *Microb. Genomics* **4** (2018).
9. Deng, X. *et al.* Genomic epidemiology of *Salmonella enterica* serotype Enteritidis based on population structure of prevalent lineages. *Emerg. Infect. Dis.* **20**, 1481–1489 (2014).
10. Feasey, N. A. *et al.* Distinct *Salmonella* Enteritidis lineages associated with enterocolitis in high-income settings and invasive disease in low-income settings. *Nat. Genet.* **48**, 1211–1217 (2016).
11. Achtman, M. *et al.* Multilocus Sequence Typing as a Replacement for Serotyping in *Salmonella enterica*. *PLoS Pathog.* **8**, e1002776 (2012).
12. Alikhan, N.-F., Zhou, Z., Sergeant, M. J. & Achtman, M. A genomic overview of the population structure of *Salmonella*. *PLOS Genet.* **14**, e1007261 (2018).
13. Yim, L. *et al.* Differential Phenotypic Diversity among Epidemic-Spanning *Salmonella enterica* Serovar Enteritidis Isolates from Humans or Animals. *Appl. Environ. Microbiol.* **76**, 6812–6820 (2010).
14. Betancor, L. *et al.* Genomic and phenotypic variation in epidemic-spanning *Salmonella enterica* serovar Enteritidis isolates. *BMC Microbiol.* **9**, 237 (2009).
15. Bankevich, A. *et al.* SPAdes: A New Genome Assembly Algorithm and Its Applications to Single-Cell Sequencing. *J. Comput. Biol.* **19**, 455–477 (2012).
16. Zerbino, D. R. & Birney, E. Velvet: Algorithms for de novo short read assembly using de Bruijn graphs. *Genome Res.* **18**, 821–829 (2008).
17. Brettin, T. *et al.* RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. *Sci. Rep.* **5**, 8365 (2015).
18. Kurtz, S. *et al.* Versatile and open software for comparing large genomes. *Genome Biol.* **5**, R12 (2004).
19. Letunic, I. & Bork, P. Interactive tree of life (iTOL) v3: an online tool for the display and annotation of phylogenetic and other trees. *Nucleic Acids Res.* **44**, W242–W245 (2016).
20. Sullivan, M. J., Petty, N. K. & Beatson, S. A. Easyfig: a genome comparison visualizer. *Bioinformatics* **27**, 1009–1010 (2011).
21. Zhou, Z. *et al.* Pan-genome Analysis of Ancient and Modern *Salmonella enterica* Demonstrates Genomic Stability of the Invasive Para C Lineage for Millennia. *Curr. Biol.* **28**, 2420–2428.e10 (2018).
22. Wiczorek, A. & McHenry, C. S. The NH 2-terminal ϕ Domain of the α Subunit of the *Escherichia coli* Replicase Binds the ϵ Proofreading Subunit. *J. Biol. Chem.* **281**, 12561–12567 (2006).
23. Teplyakov, A. *et al.* Crystal structure of the *Escherichia coli* YcdX protein reveals a trinuclear zinc active site. *Proteins Struct. Funct. Genet.* **51**, 315–318 (2003).
24. Wang, Y. *et al.* Comparative Genomic Analysis and Characterization of Two *Salmonella enterica* Serovar Enteritidis Isolates From Poultry With Notably Different Survival Abilities in Egg Whites. *Front. Microbiol.* **9**, 1–14 (2018).
25. Inoue, T. *et al.* Genome-Wide Screening of Genes Required for Swarming Motility in *Escherichia coli* K-12. *J. Bacteriol.* **189**, 950–957 (2007).
26. Bobik, T. A., Xu, Y., Jeter, R. M., Otto, K. E. & Roth, J. R. Propanediol utilization genes (pdu) of *Salmonella typhimurium*: three genes for the propanediol dehydratase. *J. Bacteriol.* **179**, 6633–9 (1997).
27. Bobik, T. A., Havemann, G. D., Busch, R. J., Williams, D. S. & Aldrich, H. C. The propanediol utilization (pdu) operon of *Salmonella enterica* serovar Typhimurium LT2 includes genes necessary for formation of polyhedral organelles involved in coenzyme B(12)-dependent 1, 2-propanediol degradation. *J. Bacteriol.* **181**, 5967–75 (1999).
28. Nuccio, S.-P. & Bäuml, A. J. Comparative Analysis of *Salmonella* Genomes Identifies a Metabolic Network for Escalating Growth in the Inflamed Gut. *MBio* **5**, e00929–14 (2014).
29. Faber, F. *et al.* Respiration of Microbiota-Derived 1,2-propanediol Drives *Salmonella* Expansion during Colitis. *PLOS Pathog.* **13**, e1006129 (2017).
30. Sharp, P. M., Kelleher, J. E., Daniel, A. S., Cowan, G. M. & Murray, N. E. Roles of selection and recombination in the evolution of type I restriction-modification systems in enterobacteria. *Proc. Natl. Acad. Sci.* **89**, 9836 LP–9840 (1992).
31. Taylor, J. E., Swiderska, A. & Geoff Kneale, G. A rapid purification procedure for the HsdM protein of EcoR124I and biophysical characterization of the purified protein. *Protein Expr. Purif.* **87**, 136–140 (2013).
32. Heusipp, G., Falker, S. & Alexandersmidt, M. DNA adenine methylation and bacterial pathogenesis. *Int. J. Med. Microbiol.* **297**, 1–7 (2007).
33. Silva, C. A. *et al.* Infection of Mice by *Salmonella enterica* Serovar Enteritidis Involves Additional Genes That Are Absent in the Genome of Serovar Typhimurium. *Infect. Immun.* **80**, 839–849 (2012).
34. Pouillot, F., Fayolle, C. & Carniel, E. A putative DNA adenine methyltransferase is involved in *Yersinia pseudotuberculosis* pathogenicity. *Microbiology* **153**, 2426–2434 (2007).
35. Edwards, M. D. *et al.* Characterization of three novel mechanosensitive channel activities in *Escherichia coli*. *Channels* **6**, 272–281 (2012).
36. Campioni, F. *et al.* Changing of the Genomic Pattern of *Salmonella* Enteritidis Strains Isolated in Brazil Over a 48 year-period revealed by Whole Genome SNP Analyses. *Sci. Rep.* **8**, 10478 (2018).

Acknowledgements

We thanks to Aracé Martínez for her technical assistance with the collection of isolates. We thanks to Professor Gordon Dougan and Professor Nicholas Thomson for their collaboration with the sequencing of genomes. This work was supported by the University Research Council (Comisión Sectorial de Investigación Científica, CSIC), Universidad de la República, Uruguay (Research groups program PI Laura Betancor and Lucía Yim).

Author contributions

B.D., A.I., L.B., V.P.E. and F.G. performed all the bioinformatic analysis. L.B., L.Y. and D.P. participate on strain collection, conservation and sequencing. B.D., L.B., A.I., L.Y. and J.A.C. participate in the interpretation of results, discussion and conclusions. B.D. and L.B. wrote the manuscript. All authors reviewed the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary information is available for this paper at <https://doi.org/10.1038/s41598-020-60502-8>.

Correspondence and requests for materials should be addressed to L.B.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2020