

Structure and catalytic mechanism of a human triacylglycerol-synthesis enzyme

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Triacylglycerols store metabolic energy in organisms and have industrial uses as foods and fuels. Excessive accumulation of triacylglycerols in humans causes obesity and is associated with metabolic diseases¹. Triacylglycerol synthesis is catalysed by acyl-CoA diacylglycerol acyltransferase (DGAT) enzymes^{2–4}, the structures and catalytic mechanisms of which remain unknown. Here we determined the structure of dimeric human DGAT1, a member of the membrane-bound *O*-acyltransferase (MBOAT) family, by cryo-electron microscopy at approximately 3.0 Å resolution. DGAT1 forms a homodimer through N-terminal segments and a hydrophobic interface, with putative active sites within the membrane region. A structure obtained with oleoyl-CoA substrate resolved at approximately 3.2 Å shows that the CoA moiety binds DGAT1 on the cytosolic side and the acyl group lies deep within a hydrophobic channel, positioning the acyl-CoA thioester bond near an invariant catalytic histidine residue. The reaction centre is located inside a large cavity, which opens laterally to the membrane bilayer, providing lipid access to the active site. A lipid-like density—possibly representing an acyl-acceptor molecule—is located within the reaction centre, orthogonal to acyl-CoA. Insights provided by the DGAT1 structures, together with mutagenesis and functional studies, provide the basis for a model of the catalysis of triacylglycerol synthesis by DGAT.

The triacylglycerol biosynthesis pathway was elucidated nearly 60 years ago⁵, and the DGAT enzymes were identified more than 20 years ago^{2–4}. Mammalian triacylglycerol synthesis is catalysed by DGAT1 (Extended Data Fig. 1a) and DGAT2 enzymes, which belong to different protein families with distinct predicted membrane topologies⁶. Both enzymes utilize fatty acyl-CoA and diacylglycerol (DAG) substrates, although DGAT1 has a broader acyl-acceptor substrate specificity⁷ (Extended Data Fig. 1a). DGAT1 and DGAT2 have distinct roles in cells⁸ and physiology^{9,10}, but together account for most mammalian triacylglycerol synthesis¹¹. Despite these insights, the enzymatic mechanism of triacylglycerol synthesis has remained unknown.

We investigated DGAT1, a member of the MBOAT family, which includes enzymes that acylate proteins (for example, ghrelin, Wnt and hedgehog) or lipids (for example, DAG and sterols)¹². DGAT1 is a polytopic endoplasmic reticulum membrane protein^{2,13} with a conserved histidine residue (His415 in human DGAT1) that is probably involved in catalysis^{13–16}. A crystal structure of DltB, a prokaryotic MBOAT, was reported recently¹⁷. However, DltB catalyses the transfer of an alanyl moiety to teichoic acid, and therefore its structure provides limited insights into the mechanism of lipid-acylating MBOATs.

Structure determination of human DGAT1

We determined the structure of DGAT1 by cryo-electron microscopy (cryo-EM). We purified human DGAT1 in digitonin, and

gel-filtration chromatography revealed two DGAT1-containing peaks, presumably representing different oligomeric states (Extended Data Fig. 1b, c). Initial studies of the major peak (peak 2 in Extended Data Fig. 1b) failed to yield high-resolution structures. We therefore reconstituted DGAT1 from the two peak fractions (Extended Data Fig. 1b, c) into PMAL-C8 amphipol (Extended Data Fig. 1d–f). Gel-filtration analysis revealed that the reconstituted DGAT1 also displayed two peaks (red and blue arrows, Extended Data Fig. 1d). The later elution peak was found to contain DGAT1 dimers; the earlier peak probably represented DGAT1 tetramers (Extended Data Fig. 1g), which have been observed previously^{13,14}. These two DGAT1 populations showed comparable activities (Extended Data Fig. 1f).

Reconstitution with amphipol improved homogeneity of DGAT1 particles (Extended Data Fig. 1g), enabling us to use DGAT1 in the later elution peak (blue arrow, Extended Data Fig. 1d) to generate a cryo-EM three-dimensional (3D) reconstruction at approximately 3.0 Å overall resolution (Extended Data Figs. 2, 3, Extended Data Table 1). This cryo-EM map showed well-defined side-chain densities for most amino acid residues, enabling unambiguous model building (Extended Data Fig. 3d). The first 65 amino acids and a loop region (amino acids 229–238) were invisible, probably owing to flexibility in these regions. A belt-shaped density of PMAL-C8 surrounded most of DGAT1 (Fig. 1a, b, Extended Data Fig. 2d), consistent with DGAT1 being mostly embedded in the membrane¹³.

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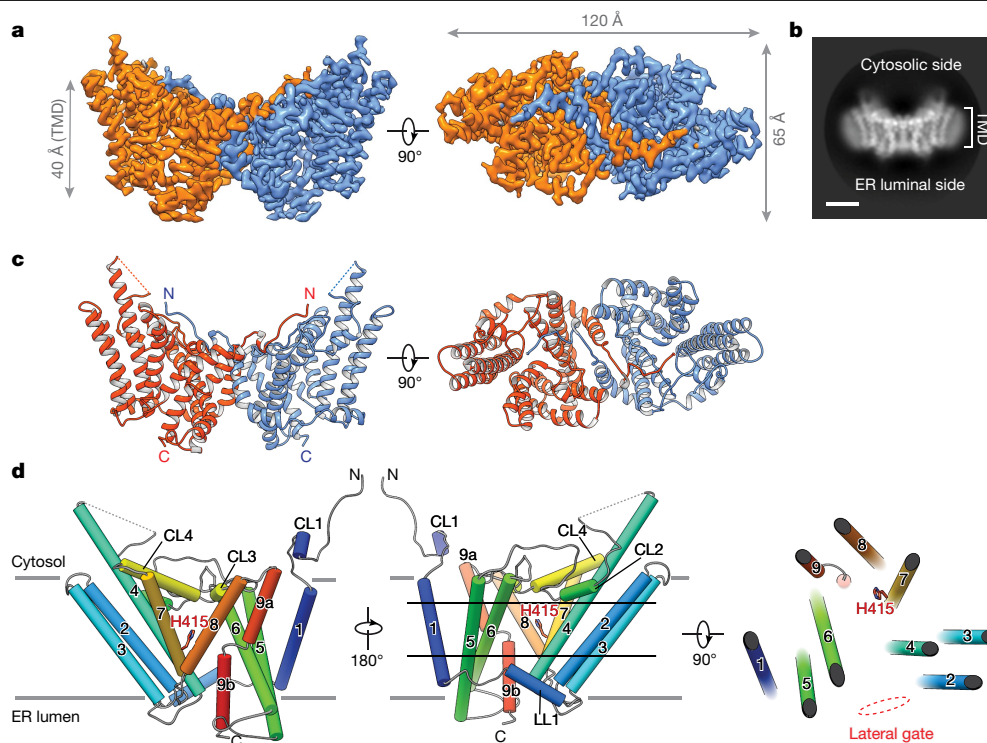


Fig. 1 | Cryo-EM structure of human DGAT1. **a**, Cryo-EM map of human DGAT1, coloured by monomer, viewed along the membrane plane or from the cytosolic side. Map is contoured at 6σ . TMD, transmembrane domain. **b**, Representative two-dimensional (2D) average of DGAT1 cryo-EM images. A micelle-like density surrounds the major portion of DGAT1. Scale bar, 40 Å. ER, endoplasmic

reticulum. **c**, Ribbon representation of the human DGAT1 dimer. The dashed line indicates a disordered segment (residues 229–238) not resolved in the cryo-EM map. **d**, Topology of DGAT1 monomer. The conserved His415 residue is shown. The red dashed oval (right) indicates the membrane-embedded lateral gate to the central cavity.

Overall architecture of human DGAT1

The cryo-EM map reveals that DGAT1 forms a dimer (Fig. 1a–c). Each DGAT1 subunit has nine transmembrane helices, with N and C termini located on the cytosolic and luminal sides of the endoplasmic reticulum membrane, respectively (Fig. 1c, d). Short helices in cytosolic loop (CL) and luminal loop (LL) regions, including CL2–CL4 and LL1, orient in parallel to the membrane surface (Fig. 1d). The DGAT1 dimer forms through extensive hydrogen-bonding interactions of the first 20 resolved residues (His69–Gly87) with a cytosolic surface groove on the opposing subunit, and through hydrophobic interactions of the transmembrane helix 1 (TM1) region of Phe82–Ile98 with the other monomer (Extended Data Fig. 4a). Additional densities, the shapes and sizes of which are consistent with four phospholipids, were present at the dimer interface and appear to contribute to the contacts between the DGAT1 monomers (Extended Data Fig. 4b). Consistently, phospholipids were identified in extracts of purified DGAT1 after amphipol reconstitution (Extended Data Fig. 4c).

To assess the functional importance of dimerization, we measured acyl-transferase activities in lysates of N-terminal truncation mutants expressed in cells lacking DGAT1 (Extended Data Fig. 4d, e). Similar to previous reports^{13,15}, deletion of the first 85 ($\Delta 85$) or 90 residues of DGAT1 led to lower protein expression levels and reduced, but still detectable, activity (Extended Data Fig. 4f–h). The purified detergent-solubilized $\Delta 85$ -mutant protein exhibited almost complete absence of enzyme activity (Extended Data Fig. 10c), indicating that the N-terminal region is required for optimal activity.

The DGAT1 active site is within the membrane

The transmembrane helices of DGAT1 form a large central cavity within the membrane that is open to the bilayer via a wide lateral gate (Fig. 2a, b)

lined by TM2, TM4, TM5, TM6 and LL1 (Figs. 1d, 2b). The conserved catalytic histidine residue (His415)¹³ is buried inside the central cavity (Figs. 1d, 2d). As well as through the lateral gate, this cavity is accessible through openings on the cytosolic and luminal sides (Fig. 2a, c).

Structural comparison of DGAT1 and DltB¹⁷ reveals an overall similar fold, in which the majority of transmembrane helices form a concave-shaped ridge on either side of the membrane and the conserved histidine residue is located in the central region embedded in the membrane (Fig. 2b, c). A DALI search with the DGAT1 structure showed that this architecture appears to be unique to DGAT1 and DltB. We therefore refer to this conserved domain structure as the ‘MBOAT core’ (Extended Data Fig. 5a–c). Within the MBOAT core, a tunnel-like region equivalent to the alanyl-donor binding pocket of DltB is also present in DGAT1 (Fig. 2b, green circle). Despite their overall similarity, the lateral gate found in DGAT1 is absent from DltB. Instead, the N-terminal transmembrane helices TM1 and TM2 of DltB are in close proximity, blocking lateral access to the central cavity from the membrane (Fig. 2b, Extended Data Figs. 5a, b). This suggests that the acyl-acceptor lipid substrates for DGAT1, such as DAG, access the active site through the lateral gate within the membrane, whereas DltB, which alanylates extracellular teichoic acid, does not require a membrane opening for substrates. This hypothesis is consistent with a computationally predicted structure of ghrelin O-acyltransferase¹⁸, an MBOAT that acylates the secreted peptide hormone ghrelin at the luminal side of the endoplasmic reticulum, in which the transmembrane region lacks an opening within the lipid bilayer.

The putative active site of DGAT1 contains both the invariant His415 and several highly conserved nearby polar residues, including Asn378, Gln437 and Gln465, with their side chains oriented towards the cavity centre (Fig. 2d, Extended Data Fig. 6a). The orientation of His415 appears to be stabilized by a hydrogen bond of its N δ_1 atom with the S δ atom of Met434 (Fig. 2d). Single alanine mutations of His415 or Asn378 abolished DGAT1 activity, whereas Gln437Ala, Met434Ala or Gln465Ala mutations

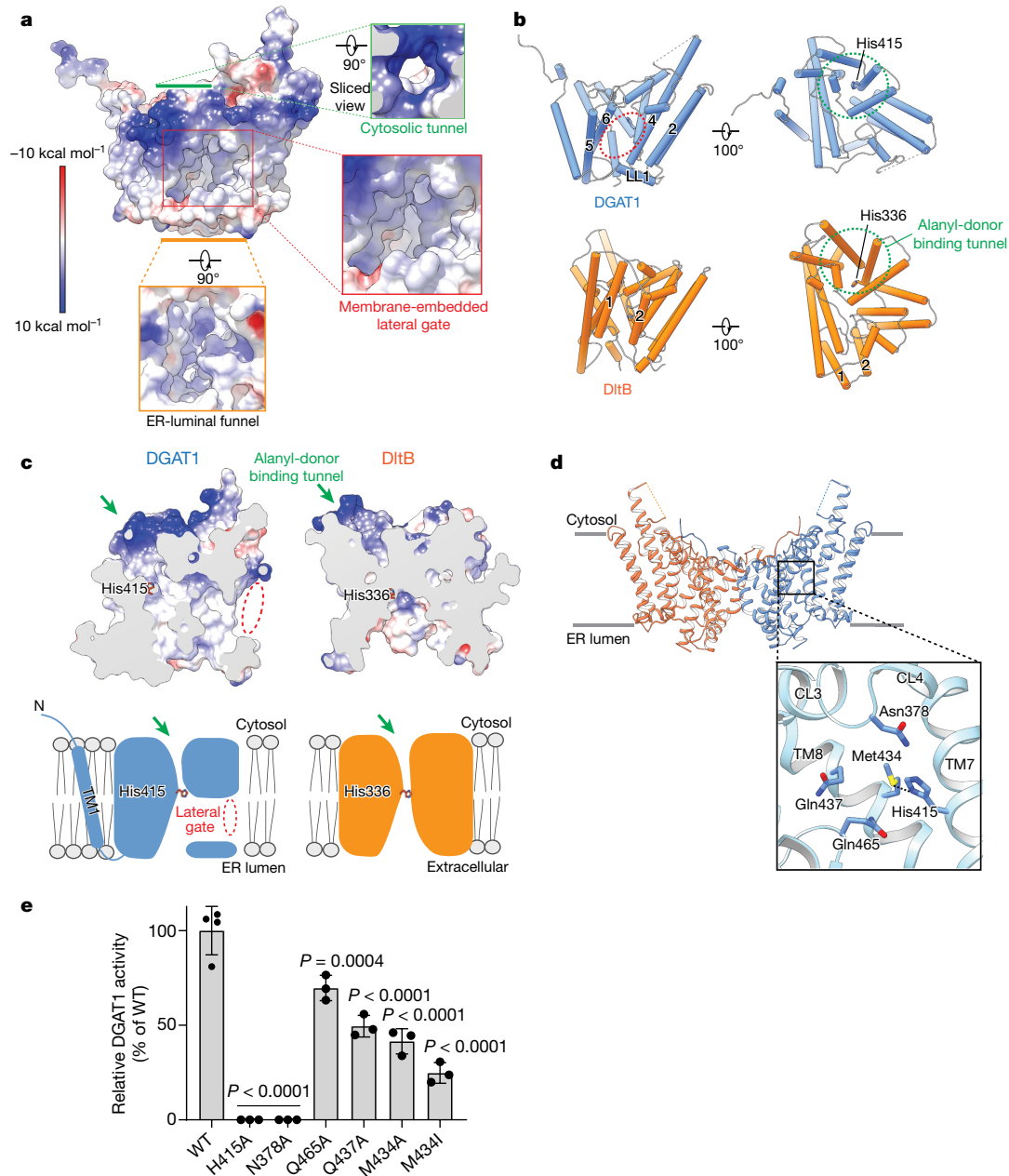


Fig. 2 | DGAT1 reaction centre and structural comparison with DltB. a, Three major openings leading to the DGAT1 putative catalytic centre include a cytosolic tunnel (green box), an ER-luminal funnel (orange box), and a membrane-embedded lateral gate (red box). The electrostatic surface is shown. **b,** DGAT1 structural comparison with DltB. DGAT1 and DltB structures are superimposed and separately shown as cylinders. The lateral gate in DGAT1 (red dashed oval) is absent in DltB owing to the presence of TM1 and TM2, which block membrane access to the catalytic centre. A cytosolic tunnel in DGAT1 (green box in **a**) is also present in DltB and functions as a binding tunnel for alanyl-donor protein DltC (green dashed circle). **c,** Sliced-surface views (top) of superimposed DGAT1 and DltB as in **b**, and cartoon representations (bottom) illustrating the common and distinct structural features. The red oval indicates

reduced activity by 50–75% (Fig. 2e). Together, these studies suggest that the central cavity of DGAT1 within the membrane is the catalytic site.

Acyl-CoA binding to DGAT1

To investigate DGAT1-mediated acyl-CoA binding and acyl-chain transfer, we characterized the structure of human DGAT1 in the presence of

the lateral gate shown in **a**, and the green arrow indicates the cytosolic tunnel shown in **a** and **b**. Note that the N terminus and TM1 of DGAT1 appear as extra structural elements compared with DltB (also see Extended Data Fig. 5a, b). **d,** Zoomed-in view of the DGAT1 active centre. The conserved His415 and highly conserved polar residues in the vicinity are shown as sticks. A hydrogen bond between His415 and Met434 (3.1 Å) is shown as a dashed line. **e,** DGAT1 activities of wild type (WT) and reaction centre mutants expressed in DGAT1-knockout cells. Activities were normalized to the amount of DGAT1 protein expressed. Data are mean \pm s.d.; $n = 4$ independent experiments for WT, $n = 3$ independent experiments for all mutants. One-way ANOVA with Dunnett's post hoc test was applied.

oleoyl-CoA, a preferred substrate³, and generated cryo-EM maps at 3.2 Å and 2.8 Å resolution (Extended Data Figs. 7, 8). These two maps are essentially identical for DGAT1 protein and show strong densities for the bound oleoyl-CoA (Fig. 3a, Extended Data Fig. 9a). In the 3.2 Å resolution map, a complete oleoyl-CoA molecule is visible (Fig. 3a), representing an unprocessed substrate. In the cryo-EM maps calculated with other particle images, the oleoyl-CoA density appears to be broken

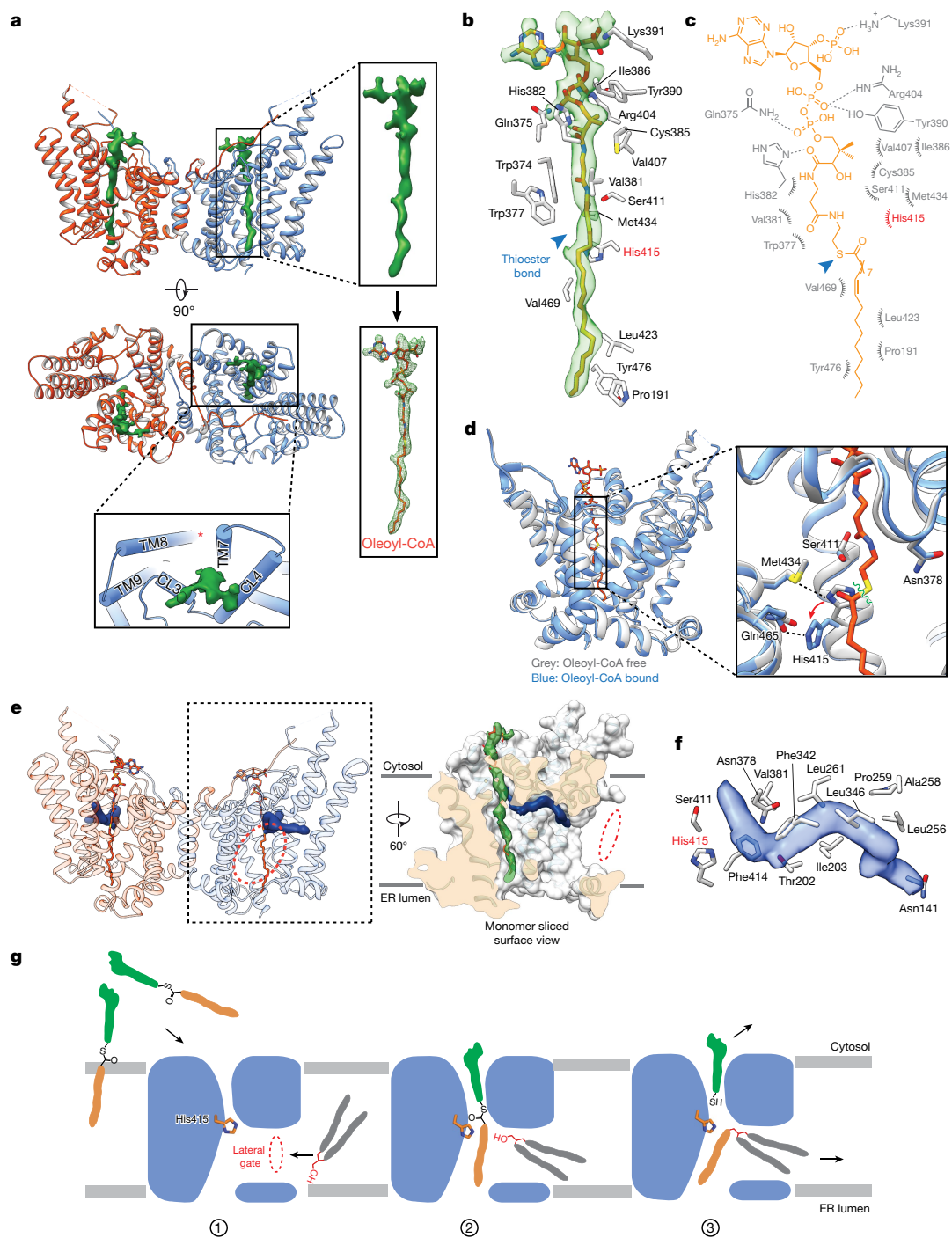


Fig. 3 | Cryo-EM structures of DGAT1 complexed with a fatty acyl-donor substrate and a lipid-like molecule resembling an acyl-acceptor substrate.

a, Cryo-EM structure of human DGAT1 bound to an intact oleoyl-CoA molecule (green density at 3.5 σ contour level). Zoomed-in view shows the modelled oleoyl-CoA molecule superimposed with the electron microscopy density. Lower panel illustrates the cytosolic tunnel of DGAT1, consisting of TM7, TM8, TM9, CL3 and CL4, which interacts with oleoyl-CoA substrate. The red asterisk indicates the gap between TM7 and TM8 in the cytosolic membrane leaflet. **b, c**, Interactions between oleoyl-CoA and DGAT1. Residues interacting with oleoyl-CoA within a 4 Å distance are shown. Detailed molecular interactions are depicted in **c**. Polar interactions (dashed lines) and interaction distances are labelled. Non-dipolar interactions are shown as spiked arcs. The blue arrowhead indicates the thioester bond in the acyl-CoA substrate. **d**, Conformational changes in the DGAT1 active centre upon oleoyl-CoA binding. The red arrow indicates the His415 conformational change before (grey) and after (blue) oleoyl-CoA binding, and dashed lines indicate the

hydrogen bonds between His415 and Met434 without oleoyl-CoA and between His415 and Gln465 (2.4 Å) upon binding to oleoyl-CoA. Asn378, which is essential for DGAT1 activity, is also shown. The green wavy line indicates the scissile bond in the thioester group during the acyl-transfer reaction. **e**, Left, unidentified lipid-shaped density (blue) residing in the DGAT1 central cavity. The lateral gate (red dashed circle) and oleoyl-CoA molecules (sticks) are shown. Right, surface representation, showing the orientation of the lipid-like density in the monomer of the DGAT1-acyl-CoA complex structure. **f**, Residues accommodating the lipid-like density (transparent blue surface) shown in the acyl-CoA free DGAT1 structure. **g**, Hypothetical model for DGAT1-catalysed triacylglycerol formation. The reaction is initiated by binding of acyl-donor (via the cytosolic channel) and acyl-acceptor (via the lateral gate) substrates to the active centre within the membrane (step 1). The acyl-transfer reaction is catalysed by the conserved His415 residue (step 2). The reaction product of free CoA-SH is released into the cytosol, and the hydrophobic triacylglycerol molecule diffuses into the lipid bilayer (step 3).

at the thioester bond (Extended Data Fig. 9a), possibly representing a state after catalytic cleavage. However, validation that the cleaved acyl-CoA is a post-reaction state requires further investigation.

Our structure of DGAT1 bound to intact oleoyl-CoA reveals that the acyl-CoA molecule occupies the cytosolic tunnel, with the CoA moiety at the cytosolic face and its acyl chain extending through the reaction centre towards the endoplasmic reticulum lumen (Fig. 3a). This orientation of the acyl-CoA is consistent with acyl-CoA molecules being synthesized on the cytosolic side of the endoplasmic reticulum membrane¹⁹. Acyl-CoA and DGAT1 interaction is mediated predominantly through the 4'-phosphopantetheine moiety in the oleoyl-CoA molecule (Fig. 3b, c). The adenosine group of CoA contributes minimally to interactions with DGAT1. The relatively weak cryo-EM density in this region suggests that the adenosine is mobile and could possibly adopt various conformations (Extended Data Fig. 9b). By contrast, the density for the 4'-phosphopantetheine moiety and surrounding residues is clearly visible (Extended Data Fig. 9b). Single alanine mutations of the residues interacting with oleoyl-CoA decreased DGAT1 activity levels, although by less than 50% (Extended Data Fig. 9c). More bulky hydrophobic side-chain mutations, intended to block the binding of acyl-CoA to the tunnel (Val381Trp, Cys385Trp, Val407Phe and Ser411Ile) decreased DGAT1 activity, and a Ser411Trp mutation, which we predicted would block the tunnel, completely inactivated DGAT1 (Extended Data Fig. 9d). The loss of activity was apparently not caused by protein misfolding (Extended Data Fig. 10a, b). Activity was also validated by using purified DGAT1 (Val381Trp and Ser411Trp) mutants (Extended Data Fig. 10c).

The oleoyl-CoA in the tunnel is surrounded by TM7, TM8, TM9, CL3 and CL4, leaving a gap between TM7 and TM8 at the level of the cytosolic membrane leaflet (Fig. 3a, asterisk). This gap might accept membrane-partitioned acyl-CoA substrates, although soluble acyl-CoA could also enter the channel from the cytoplasm. The distal end of the acyl chain of oleoyl-CoA interacts with DGAT1 deep within the hydrophobic channel, suggesting that the binding of longer acyl chains may help to accurately position the acyl-donor substrate for the reaction. Consistently, DGAT1 was more active with longer acyl-chain substrates (C16–C20) than shorter ones (C10–C12) (Extended Data Fig. 9e). The enzyme exhibited activity for a variety of saturated and unsaturated long-chain fatty acyl-CoA molecules (Extended Data Fig. 9e), as reported^{3,20}. An analysis of the conservation of DGAT1 and selected MBOAT sequences showed that the acyl-CoA binding tunnel within the MBOAT core of DGAT1 is the most conserved region among MBOAT enzymes (Extended Data Fig. 6b, c). This tunnel is analogous to the pocket in DltB for binding DltC, the alanyl-donor protein¹⁷ (Fig. 2b, c, Extended Data Fig. 5b, c).

Comparison of the structures of DGAT1 with or without bound acyl-CoA reveals no large conformational differences (Fig. 3d), similar to the minor structural changes for DltB upon DltC binding¹⁷. However, several residues near the cleavage site of acyl-CoA, including Ser411, show altered conformations upon acyl-CoA binding. Among these, His415 flips towards the endoplasmic reticulum-luminal side when acyl-CoA binds (Fig. 3d). This change involves the breaking of the hydrogen bond between His415 and Met434, which opens the tunnel to accommodate the acyl-CoA substrate and enables the formation of a hydrogen bond between the His415 N_{ε2} atom and the Gln465 O_{ε1} atom (Fig. 3d), positioning His415 near the thioester bond of the acyl-CoA. Thus, acyl-CoA binding to DGAT1 results in small but important conformational changes in the active-site region that may prime the enzyme for catalysis.

A putative acyl acceptor

The cleavage site for oleoyl-CoA is presumably within a short distance of a lipid acceptor, such as DAG. Of note, our cryo-EM maps (with or without oleoyl-CoA) revealed a strong, elongated, lipid-like density in the central cavity (Fig. 3e), orthogonal to the bound oleoyl-CoA and

in close proximity to His415 (Fig. 3e, f). The shape of this density is consistent with a DAG that was co-purified with the enzyme (Extended Data Fig. 4c). Hydrophobic residues, including Phe342, Leu261, Val381 and the highly conserved Asn378, line the region and form a channel surrounding the lipid-like density (Fig. 3f). The channel exhibits a bent, hydrophobic pathway that appears to allow the binding of hydrophobic linear or curvilinear molecules (Extended Data Fig. 9g). The bent architecture of this tunnel may explain how DGAT1 distinguishes acyl acceptors such as DAG or long-chain alcohols⁷ from more planar and rigid substrates such as cholesterol, which is acylated by acyl CoA:cholesterol acyltransferases²¹. In the cryo-EM map of DGAT1 containing a cleaved oleoyl-CoA, the cleaved acyl chain is merged onto the orthogonally orientated density, giving rise to a density with a three-way branched shape resembling triacylglycerol (Extended Data Fig. 9f). Further studies are required to determine the identity of the lipid-like density and to elucidate the basis of acyl-acceptor preferences of DGAT1.

Discussion

Our cryo-EM structures of human DGAT1, obtained in the absence and presence of bound fatty acyl-CoA substrate, reveal the detailed molecular architecture of this mammalian MBOAT and, together with extensive functional studies, provide mechanistic insight into the catalysis of lipid acylation mediated by DGAT1. Our results reveal the dimeric architecture of DGAT1, the arrangement of crucial residues within the catalytic centre, the structural basis for acyl-CoA binding and thioester cleavage, and a potential mechanism of acyl transfer to a lipid acceptor.

On the basis of our results, we propose a model for triacylglycerol synthesis mediated by DGAT1 in which an acyl-CoA substrate binds, from the cytosolic side, via a combination of electrostatic features of the cytosolic face and a deep, hydrophobic tunnel (Fig. 3b, c). When an acyl acceptor—such as DAG—binds in an orthogonal hydrophobic tunnel accessed from within the lipid bilayer, the acyl chain from acyl-CoA is transferred to the acceptor in a reaction involving the conserved His415 residue²², ultimately generating CoA-SH and triacylglycerol (see model, Fig. 3g). The crucial Asn378 residue may serve to interact with the acyl-acceptor substrate. The reaction products are released to either the cytoplasm (CoA-SH) or the membrane bilayer (triacylglycerol); within the membrane bilayer, the triacylglycerol can initiate lipid-droplet formation²³. Additional structural and biochemical studies are needed to test this model and delineate the catalytic mechanism.

Our structure shows that the N-terminal region of DGAT1 mediates dimerization, whereas the analogous region of the DltB monomer instead blocks the lateral gate to the membrane. This suggests evolutionary functional adaptations that occurred within MBOATs to accommodate different substrates. Previous studies indicated that the N-terminal region of DGAT1 mediates its oligomerization^{13,15} and may be involved in acyl-CoA binding and enzyme regulation^{15,16}. Because our structure lacks N-terminal segments, it does not provide new insights into these possibilities.

Our DGAT1 structure provides a framework for understanding mutations in human *DGAT1*, which cause congenital diarrhoea²⁴. Most of these *DGAT1* mutations result in a loss of protein expression, but as additional mutations are identified, they can be mapped onto the enzyme structure to predict the functional consequences. The DGAT1 structure may also further our understanding of the mechanisms of DGAT1 inhibitors that have been developed for treating metabolic diseases^{25,26}.

Online content

Any methods, additional references, Nature Research reporting summaries, source data, extended data, supplementary information, acknowledgements, peer review information; details of author contributions and competing interests; and statements of data and code availability are available at <https://doi.org/10.1038/s41586-020-2289-6>.

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Methods

No statistical methods were used to predetermine sample size. The experiments were not randomized. The investigators were not blinded to allocation during experiments and outcome assessment.

Protein expression and purification

DGAT1 from *Homo sapiens* (UniPort ID: O75907) was expressed in HEK293 GnTi- (ATCC) cells using the BacMam system. In brief, the DGAT1 cDNA sequence was cloned into a pFastBacMam vector constructed with the mammalian CMV promoter. The protein was fused with an N-terminal His₈ tag followed by a maltose-binding protein (MBP). A TEV cleavage site was inserted between the MBP and DGAT1 sequences. The bacmid for expressing the MBP-fusion protein was generated in DH10Bac *Escherichia coli* cells (Thermo Fisher Scientific) by transformation of the construct. Baculovirus was generated by transfecting *Spodoptera frugiperda* (Sf9) cells (Expression Systems) cultured at 27 °C in ESF921 medium (Expression Systems) with the bacmid using the BaculoPORTER transfection reagent (Genlantis). After that, the baculovirus was amplified twice in Sf9 cells to obtain sufficient virus for large-scale infection. HEK293 GnTi- cells were grown in suspension at 37 °C in FreeStyle 293 Expression Medium (Thermo Fisher Scientific) supplemented with 1% FBS and 1× Glutamax solution (Gibco). When the cells reached a density of 2.0–2.5 × 10⁶ cells per ml, baculovirus was added to the culture at 5% (v/v). After 15 h infection at 37 °C, the culture was supplemented with 10 mM sodium butyrate to boost the expression. After further growth for ~36 h, cells were collected and washed once in PBS, and cell pellets were flash frozen in liquid nitrogen and stored at –80 °C or placed on ice for immediate use.

All protein purification steps were performed at 4 °C. Cell pellets collected from 1 l cell culture were resuspended in 80 mL TSGE buffer (50 mM Tris-HCl, pH 8.0, 400 mM NaCl, 10% (v/v) glycerol and 1 mM EDTA) and supplemented with 1× complete protease inhibitor cocktail (Roche). To lyse the cells, GDN detergent (Anatrace) was added into the solution at a final concentration of 0.5% (w/v), and the mixture was generally agitated for 1 h before MgCl₂ was added to a final concentration of 5 mM. The mixture was agitated for 1 more hour. Insoluble debris was removed by centrifugation, and the supernatant was incubated with prewashed amylose resin for 2 h. The resin was carefully washed to remove contaminant proteins. The MBP-fused DGAT1 protein was eluted in TSGE buffer containing 20 mM maltose and 0.1% (w/v) digitonin (Sigma-Aldrich). The eluate containing MBP-DGAT1 was concentrated, and the MBP tag was cleaved by TEV protease. The protein was then further purified by size-exclusion chromatography on a Superose 6 column (GE Healthcare) in TSM buffer (50 mM Tris-HCl, pH 7.5, 400 mM NaCl, 10 mM MgCl₂) supplemented with 0.05% (w/v) digitonin. The peak fractions containing DGAT1 were collected and concentrated. For preparation of samples for cryo-EM analysis, the purified DGAT1 in detergent was mixed with PMAL-C8 (Anatrace) at a 1:3–1:5 (w/w) ratio, followed by gentle agitation overnight in cold room. Detergent was then removed with Bio-Beads SM-2 (Bio-Rad) for 1 h, and the beads were subsequently removed over a disposable polyprep column. The eluent was cleared by passage through a 0.22-μm filter before further purification by gel filtration with TSM buffer. The peak containing DGAT1 was collected for further use. For some assays of partially purified DGAT1 mutant proteins, recombinant proteins were transiently expressed in HEK293 cells and purified by the His₈ tag before activity measurements were performed.

Electron microscopy sample preparation and data acquisition

Negatively stained specimens were prepared using an established protocol with minor modifications²⁷. Specifically, 2.5 μl of purified DGAT1 in PMAL-C8 at 0.03–0.05 mg ml⁻¹ were applied to glow-discharged copper electron microscopy grids covered with a thin layer of continuous carbon film, and the grids were stained with 1.5% (w/v) uranyl formate for 30 s. These grids were imaged on a Tecnai T12 microscope (Thermo

Fisher Scientific) operated at 120 kV and equipped with a 4k × 4k charge-coupled device camera (UltraScan 4000; Gatan). A nominal magnification of 67,000×, corresponding to a pixel size of 1.68 Å on the specimen, and a defocus of –1.5 μm were used to record the images.

For cryo-EM samples, 2–3 μl of purified DGAT1 in PMAL-C8 was applied to a Quantifoil holey carbon grid (Cu R1.2/1.3; 400 mesh) that was glow-discharged for 30 s. Optimal particle distribution was obtained with a protein concentration of 4–5 mg ml⁻¹. After applying protein, the grids were blotted with a Whatman #1 filter paper for 5 s with ~95% humidity at 4 °C and plunge frozen in liquid ethane cooled by liquid nitrogen using a Vitrobot Mark IV system (Thermo Fisher Scientific). For the oleoyl-CoA treated sample, oleoyl-CoA freshly prepared in water was added into the concentrated DGAT1 at 5.5 mg ml⁻¹ in PMAL-C8 with a final concentration of 1 mM. The mixture was incubated on ice for 1 h before freezing the cryo-EM grids. Cryo-EM data were collected on a Titan Krios electron microscope (Thermo Fisher Scientific). Images were recorded using SerialEM²⁸. Refer to Supplementary Table 1 for the detailed data collection parameters.

Electron microscopy data processing

For negative-stain electron microscopy data, the images were binned over 2 × 2 pixels, yielding a pixel size of 3.36 Å, for further processing using Simplified Application Managing Utilities for EM labs (SAMUEL) scripts²⁹. For cryo-EM data, drift correction was performed using Miton-Cor2³⁰, and images were binned 2 × 2 by Fourier cropping to a pixel size of 3.3 Å. The defocus values were determined using CTFIND4³¹ and motion-corrected sums without dose-weighting. Motion-corrected sums with dose-weighting were used for all other steps of image processing. Particle picking was performed using a semi-automated procedure³². The 2D classification of selected particle images were performed using samclasscas.py, which uses SPIDER operations to run 10 cycles of correspondence analysis, *k*-means classification, and multireference alignment, or RELION 2D classification^{33,34}. Initial 3D models were generated with 2D averages using SPIDER 3D projection matching refinement (samrefine.py) starting from a cylindrical density that mimics the general shape and size of DGAT1. The 3D classification and refinement were performed using relion3_refine_mpi in RELION. One round of 3D classification without applying symmetry was performed to remove bad particles. Subsequently, particle images that generated 2D class averages showing clear structural features were combined for one round of 2D classification followed by 3D refinement with C2 symmetry. Refer to Extended Data Figs. 2, 7 for the detailed data processing procedure. All refinements followed the gold-standard procedure in which two half datasets are refined independently. The overall resolutions were estimated based on the gold-standard Fourier shell correlation (FSC) = 0.143 criterion. Local resolution variation of cryo-EM maps was calculated using ResMap³⁵. The amplitude information of the final maps was corrected by applying a negative B factor using relion_postprocessing with the –auto_bfac option. The details related to data processing are summarized in Extended Data Table 1.

Model building and refinement

DGAT1 density maps in MRC/CCP4 format were converted to the structure factors MTZ format with map_to_structure_factors. Then a rough initial model was obtained in by using map_to_model module on the auto-sharpened map. All these functional modules were implemented in the PHENIX suite³⁶. The modelling building was then performed manually in COOT³⁷. The DGAT1 map shows clear densities for most transmembrane helices that allowed a confident sequence registration by aromatic and other residues with bulky side chains to these α-helical regions in the density map. The linker domains that connect the transmembrane-helical regions were then built by connecting helical domains, with large and well-resolved side chains serving as markers. After building the DGAT1 monomer model, the coordinate file was docked into DGAT1 dimer map in UCSF Chimera³⁸ to generate

the dimer model. Then the dimer model was manually adjusted and refined in PHENIX real-space refinement package³⁶. The refined model was visually inspected and adjusted in COOT³⁷, and the resulting map was further put back through the real-space refinement procedure to undergo further refinement. After several cycles of manual model adjustments and computational refinement, the ligands for oleoyl-CoA and POPE lipids were placed into the residual active-site electron density. Atomic coordinates and geometric restraints for the ligands were generated using the GRADE server (Global Phasing). Then the protein coordinates with ligands were refined. This iterative refinement process was repeated until the dimer model reached optimal geometric statistics as evaluated by MolProbity³⁹. The regions showing weak signals were omitted in the final model. The protein–protein and protein–ligand interactions in the structures were analysed by LigPlot⁴⁰.

Generation of DGAT1-knockout cells

The DGAT1-knockout SUM159 cell line in wild-type background was generated by CRISPR–Cas9 gene editing⁴¹. The sequence 5′-CCACGGTAGTTGCTGAAGCCACT-3′ was used as a guide RNA (gRNA) to direct Cas9 into exon 2 of the *DGAT1* locus. Cells were selected with 1.5 μg ml⁻¹ puromycin for 48 h. Genomic DNA of clones was amplified by PCR (sense: 5′-GAGAGCTTGGCCACTGTAGG-3′, antisense: 5′-CTGGGTGACAGAGCCTGTTC-3′). PCR products were sequenced and revealed a clone (designated 2.10) with two deletion alleles, a 1-bp deletion (5′-AGTGGCTTCAGCAACTACCGTGGCATCCTGAACTGG-3′ → 5′-AGTGGCTTCAGCAACTCCGTGGCATCCTGAACTGG-3′) and predicted to result in a 14-bp deletion (5′-AGTGGCTTCAGCAACTACCGTGGCATCCTGAACTGG-3′ → 5′-AGTGGCTTCAGCAACTACCGTGGCATCCTGAACTGG-3′). These deletions (underlined bases) result in frameshifts and premature stop codons within the first 100 amino acids. The knockout was verified by immunoblotting.

DGAT1 mutagenesis and activity assay

DGAT1 mutants were generated by the QuickChange Site-Directed Mutagenesis kit (Agilent) by using the protocol provided by the manufacturer. The desired mutations were confirmed by DNA sequencing. Plasmid was amplified in Stbl2 competent cells (Thermo Fisher Scientific) before transfection. For the acyl-transferase activity, DGAT1 activity was measured *in vitro* as published⁴². In brief, *DGAT1*^{-/-} SUM159 cells, generated by the CRISPR–Cas9 system, were transfected with constructs to express wild type or mutant DGAT1 with the FuGENE HD transfection reagent (Promega) per the manufacturer's instructions. Cells were collected three days after transfection, and the cell pellet was washed with PBS before being snap-frozen in liquid nitrogen and stored in -80 °C. The cell pellet was lysed by sonication in ice-cold lysis buffer containing 250 mM sucrose, 50 mM Tris-HCl (pH 7.4) and EDTA-free protease inhibitor cocktail (Sigma-Aldrich). The unbroken debris was removed by centrifugation at 5,000 rpm, 4 °C for 5 min on a bench-top centrifuge. Each 100 μl of cell lysate (equivalent to ~5 × 10⁵ cells) was incubated with 25 μM DGAT2 inhibitor PF-06424439 (Sigma-Aldrich) on ice for 30 min. Then, 100 μl of reaction mixture was added to the cell lysate to a final concentration of 100 mM Tris-HCl (pH 7.4), 25 mM MgCl₂, 0.625 g l⁻¹ delipidated BSA, 200 μM 1,2-dioleoylglycerol and 50 μM oleoyl-CoA containing 0.2 μCi [¹⁴C]-oleoyl-CoA as a tracer (American Radiolabelled Chemicals). In some experiments, DGAT1 inhibitor PF-04620110 (Sigma-Aldrich) was added at final concentration of 10 μM into the reaction mixture. For activity assays comparing different acyl-CoA substrates, 0.2 μCi [¹⁴C]-1,2 diacylglycerol was spiked into 200 μM cold DAG substrate, and 50 μM acyl-CoA with different acyl-CoA substrate was used for individual reactions. Reactions were carried out at 37 °C with gentle agitation for 30 min or as indicated. The reactions were quenched by adding chloroform/methanol (2:1 v/v), followed by 2% phosphoric acid for phase separation. The organic phase was collected, dried, resuspended in chloroform and loaded on a silica gel thin-layer chromatography (TLC) plate (Analtech). Lipids were

resolved in a solvent system consisting of hexane, diethyl ether and acetic acid (80:20:1 v/v/v). The radioactivity of the triacylglycerol bands was revealed by Typhoon FLA 7000 phosphor imager (GE Healthcare Life Sciences) and quantified by Quantity One (V4.6.6). The final activity was normalized to the DGAT1 protein amount in the cell lysate that was measured by immunoblotting against DGAT1 or the GFP tag fused to DGAT1 N-terminal end⁴². A known amount of purified DGAT1 served as the control for quantification of DGAT1 from cell lysate.

Reporting summary

Further information on research design is available in the Nature Research Reporting Summary linked to this paper.

Data availability

The 3D cryo-EM density maps have been deposited into the Electron Microscopy Data Bank under accession numbers EMD-21461 (oleoyl-CoA free), EMD-21481 (with intact oleoyl-CoA) and EMD-21488 (with cleaved oleoyl-CoA). The coordinates have been deposited into the Protein Data Bank (PDB) with accession numbers 6VYI (oleoyl-CoA free) and 6VZI (with intact oleoyl-CoA). All other undeposited models and data are available upon request.

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Author contributions X.S., M.L., T.C.W. and R.V.F. conceived the project. X.S. performed protein expression, purification and reconstitution into amphipol, prepared cryo-EM grids, processed cryo-EM data and built the atomic models. M.L. advised on cryo-EM data processing. K.W., N.L.G. and X.S. performed the mutagenesis and activity studies. S.D.E. made genetically modified cell lines. X.S., M.L., T.C.W. and R.V.F. wrote the manuscript. All authors analysed and discussed the results and contributed to the manuscript.

Competing interests The authors declare no competing interests.

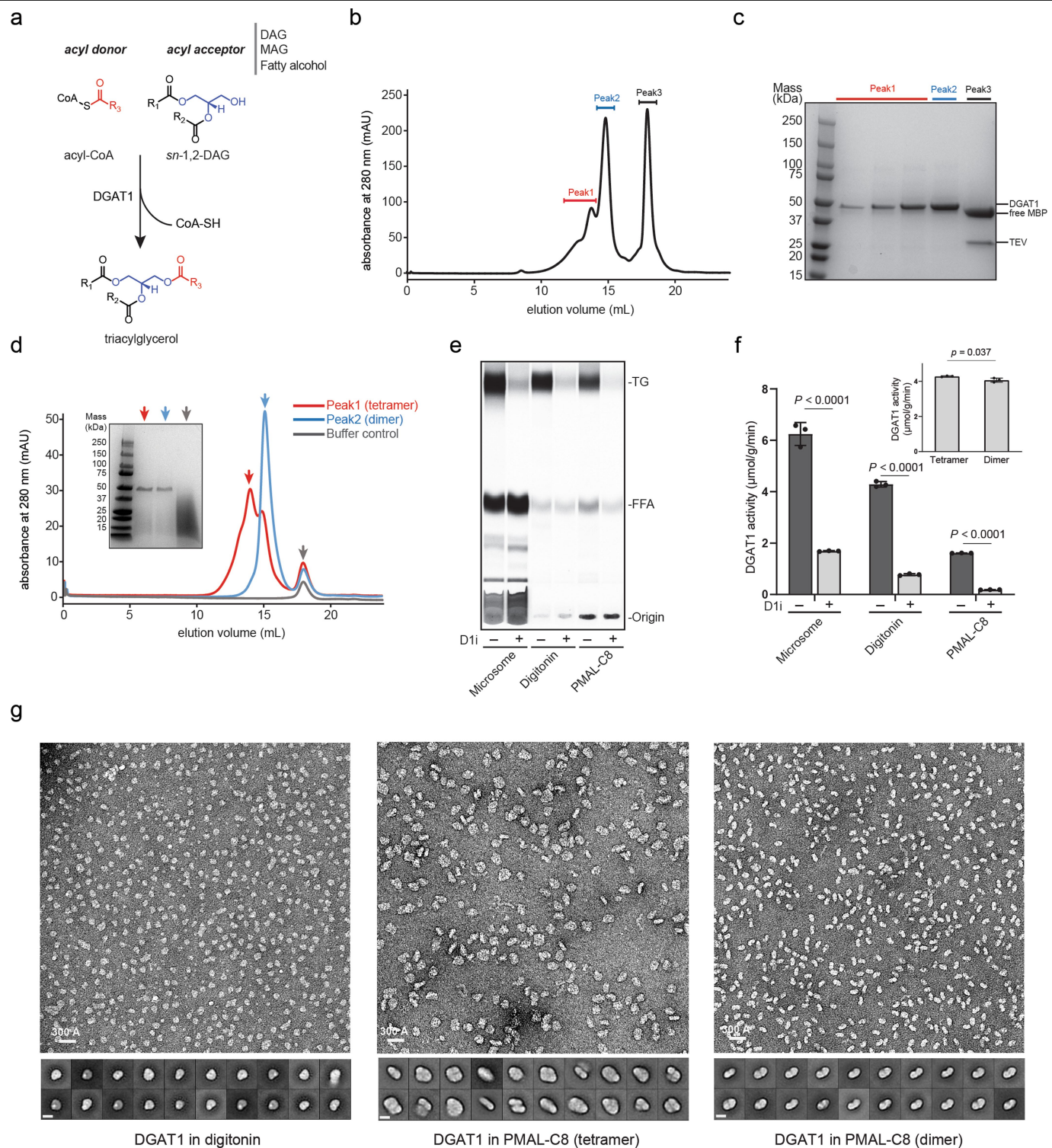
Additional information

Supplementary information is available for this paper at <https://doi.org/10.1038/s41586-020-2289-6>.

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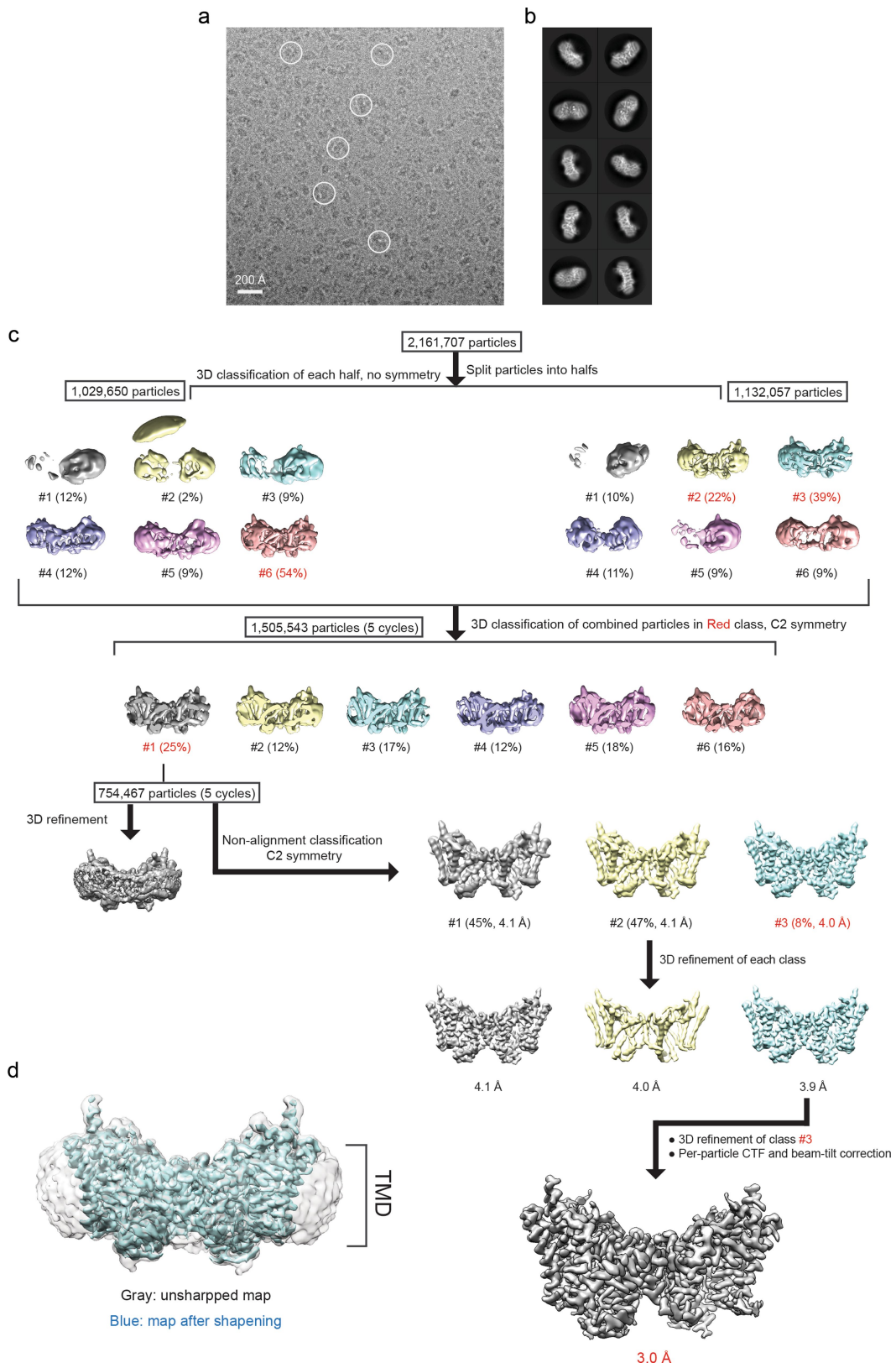
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Extended Data Fig. 1 | Purification and characterization of human DGAT1.

a The acyl-transfer reaction catalysed by DGAT1. The enzyme utilizes acyl-CoA as the sole acyl donor and recognizes different lipid molecules (that is, DAG, monoacylglycerol (MAG) and fatty alcohol) as the acyl acceptor. The panel shows the reaction with *sn*-1,2-DAG as the acyl-acceptor and triacylglycerol as the product. The acyl-group in acyl-CoA is coloured in red, and the glycerol backbone in DAG is coloured in blue. **b, c**, Gel-filtration profile (**b**) and SDS-PAGE analysis (**c**) of purified DGAT1 digitonin. Peaks 1 and 2 containing purified DGAT1 by SDS-PAGE analysis were separately collected and pooled. **d**, Gel-filtration of DGAT1 reconstituted in the amphipol PMAL-C8 that was purified from **b**. A red (tetramer) and blue (dimer) arrow denote the different oligomerization states of DGAT1. The SDS-PAGE analysis of each peak is shown in the insert. **e**, Activity analysis of

DGAT1-overexpressed microsome or purified enzyme with or without DGAT1 inhibitor (D1i). The protein from the digitonin sample of peak 2 in **b** and PMAL-C8 sample of the dimer peak in **d** were used for the assay. The reaction product of triacylglycerol was separated and analysed by TLC. FFA, free fatty acid. **f**, Quantification of triacylglycerol product shown in **e** by phosphorimaging. The insert shows the activity of DGAT1 tetramer and dimer from **d** (mean \pm s.d., $n = 3$ independent experiments). Analysis was performed using two-way ANOVA with Sidak's post hoc test. **g**, Representative negative-stain electron micrograph and 2D averages of purified DGAT1 in digitonin (peak 2 in **b**), and the tetramer and dimer species of DGAT1 in PMAL-C8 (red and blue peaks in **d**, respectively). The bar in 2D average is 100 Å. Experiments shown in **b–e** and **g** were repeated at least three times with similar results. TG, triglyceride.

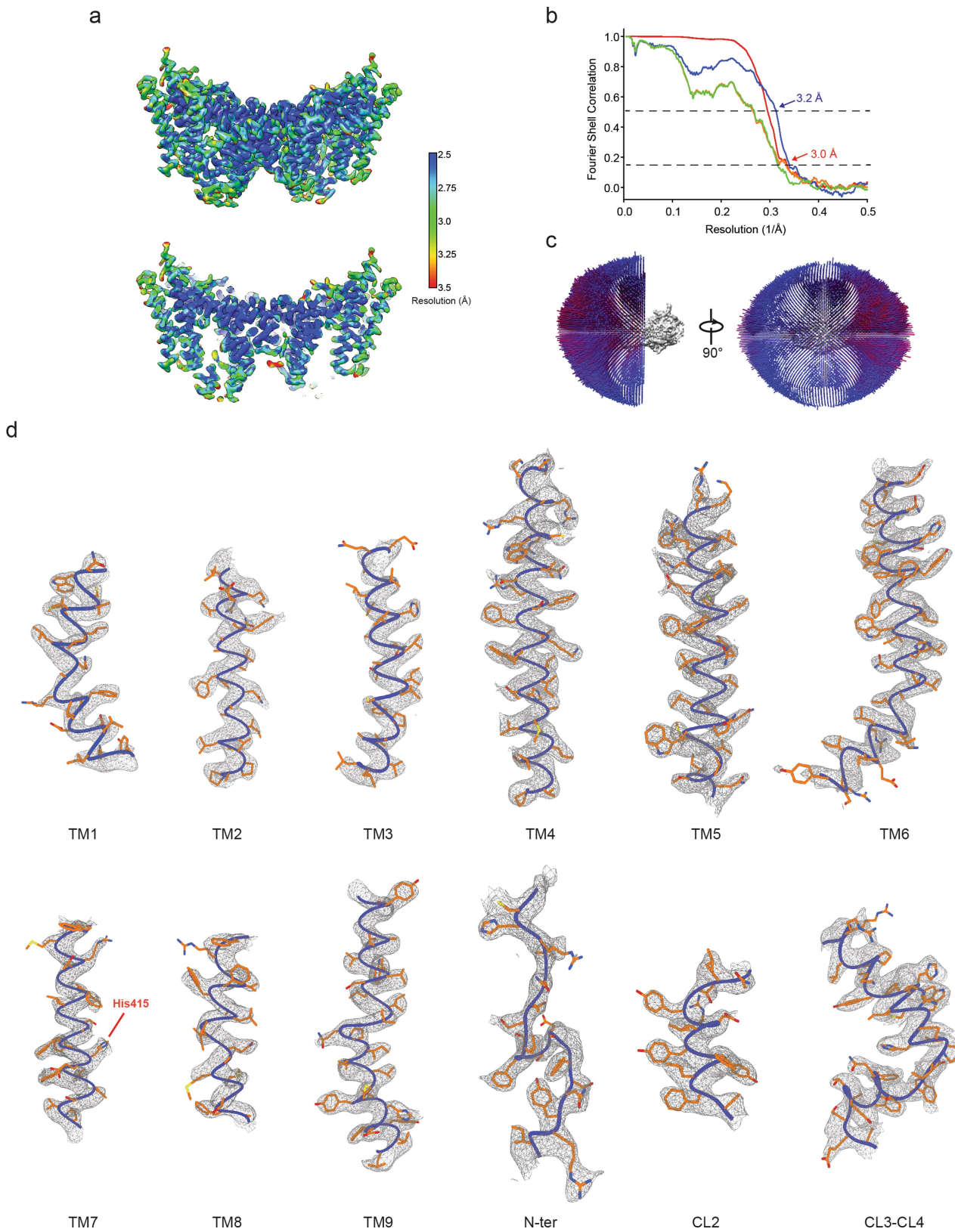


Extended Data Fig. 2 | See next page for caption.

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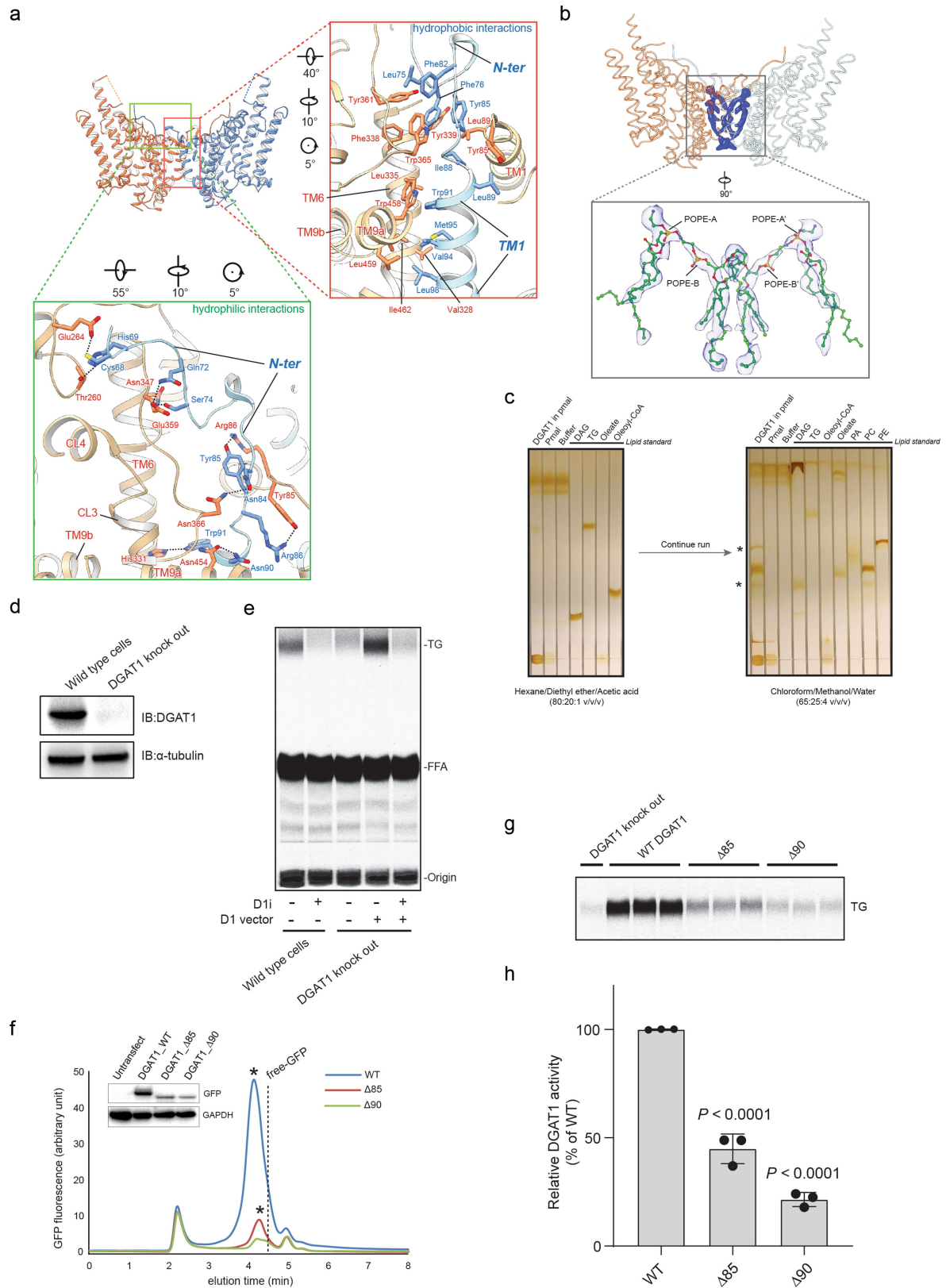
Extended Data Fig. 2 | Cryo-EM image processing of human DGAT1 in PMAL-C8. **a**, Representative cryo-EM image of DGAT1 in PMAL-C8. Some DGAT1 particles are outlined by circles. **b**, The 2D class averages of cryo-EM particle images. The box size of 2D averages is 210 Å. **c**, Three-dimensional classification and refinement of cryo-EM particles. The initial particle stack was split into two stacks due to the large number of particles for 3D classification. After the first round of classification without imposing symmetry, all of the particles classified into one best class (class 1 contains the most abundant 25% particles) in the final five iterations (indicated as '5 cycles')

were kept for further processing. This subset of particles was further classified into three classes by non-alignment classification with C2 symmetry. Afterwards, another round of refinement was performed on each individual class. Among them, class 3 exhibited the highest estimated resolution by Relion and the best side-chain signals by visual inspection and was kept for per-particle contrast transfer function (CTF) and beam-tilt corrections. The resulting cryo-EM map was used for the final cryo-EM maps, contoured at 5σ . **d**, Unsharpened map (grey) superimposed with the final DGAT1 cryo-EM map (blue) showing the detergent micelle-like signals around DGAT1.



Extended Data Fig. 3 | Single-particle cryo-EM analysis of DGAT1 reconstituted in PMAL-C8. **a**, Local resolution of the final cryo-EM map of DGAT1. A sliced view of local resolution is shown in the lower panel. **b**, FSC curves: gold-standard FSC curve between the two half maps with indicated resolution at FSC = 0.143 (red); FSC curve between the atomic model and the final map with indicated resolution at FSC = 0.5 (blue); FSC curve between half

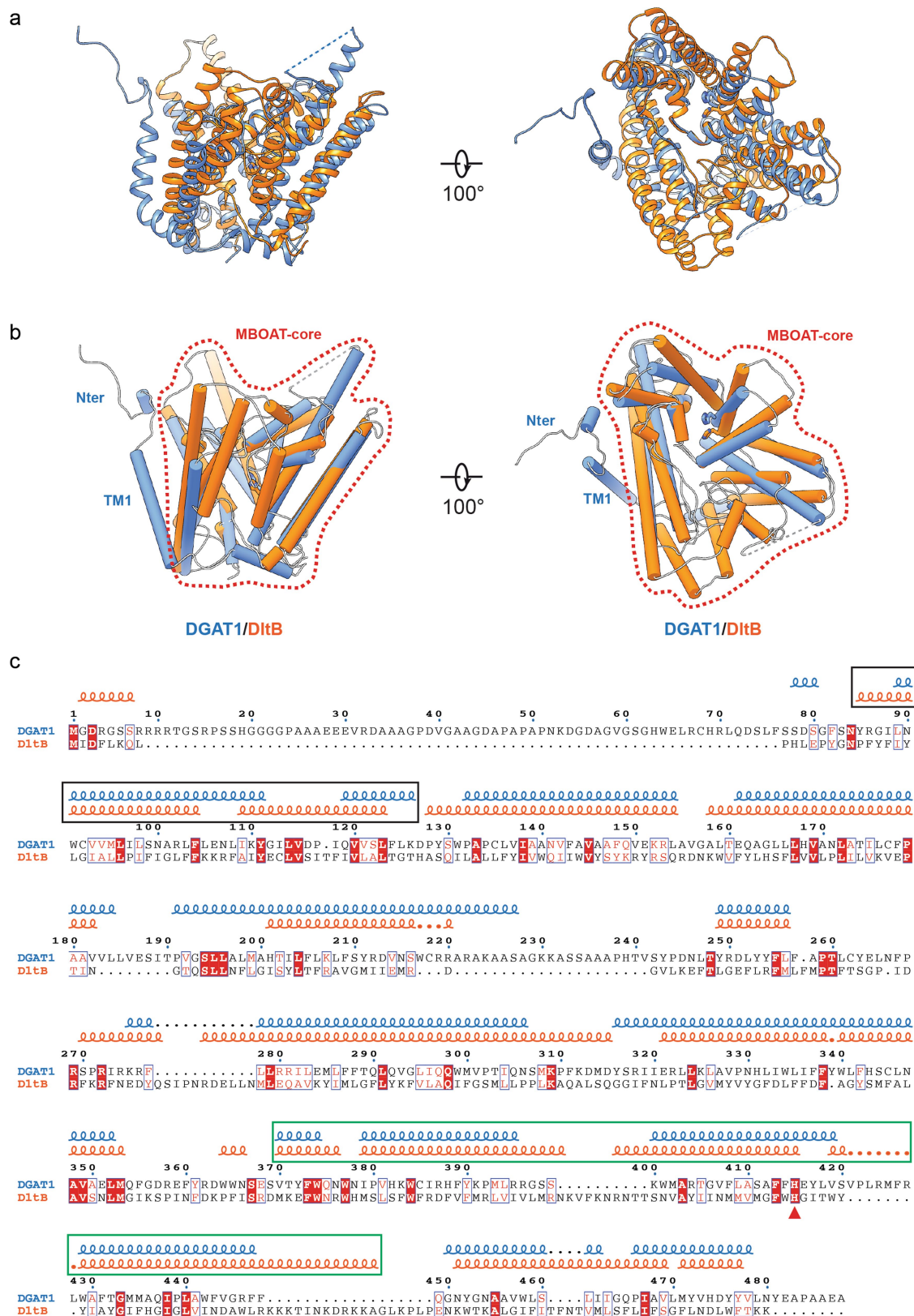
map 1 (orange) or half map 2 (green) and the atomic model refined against half map 1. **c**, Cutaway views of angular distribution of particle images included in the final 3D reconstruction. **d**, Cryo-EM densities superimposed with the atomic model for individual transmembrane helices (TM1–TM9), resolved N-terminal region (N-ter), and helices in the cytosolic loop region (CL2–CL4). The conserved His415 residue is also labelled. Maps are contoured at 4σ .



Extended Data Fig. 4 | Dimer assembly of DGAT1 and phospholipid

molecules residing at the dimer interface. **a**, Structural details of the DGAT1 dimer interaction. The resolved N-terminal domain (N-ter, amino acids 68–86) of DGAT1 forms a hydrogen-bond network with the opposing subunit depicted in a green box. Dashed lines denote hydrogen-bonding pairs of residues. Hydrophobic interactions mediating the DGAT1 dimer are shown in a red box. **b**, Densities attributed to phospholipid lipids at the DGAT1 dimer interface are shown as blue surface. The lower panel shows four 1-palmitoyl-2-oleoyl-*sn*-glycero-3-phosphoethanolamine (POPE) molecules modelled into the density; each pair of POPE densities was symmetry-related and labelled as POPE-A/B and POPE-A'/B'. Maps are contoured at 2.5σ . **c**, TLC analysis of lipids that co-purified with DGAT1 by iodine staining. The asterisks indicate the presence of phosphatidyl ethanolamine (PE) and DAG in purified DGAT1. PA, phosphatidic acid; PC, phosphatidylcholine. **d, e**, The SUM159

DGAT1-knockout cell line system analysed by western blot (**d**) and DGAT1 activity analysis by using lysates from SUM159 DGAT1-knockout cells or cells transiently overexpressing DGAT1 (**e**). **f**, Truncation of N termini reduces DGAT1 expression. Fluorescence size-exclusion chromatography (FSEC) and western blot analyses of two N-terminal truncations lacking the first 85 and 90 residues. The asterisks in the gel-filtration profiles denote the DGAT1-containing peak. The dashed line marks the peaking containing free GFP. **g, h**, Truncation of N-terminal region reduces DGAT1 activity. TLC analysis of triacylglycerol products is shown in **g**. Each assay was performed in triplicate. The final triacylglycerol products were normalized to the protein expression level with the quantification results shown in **h**. Mean \pm s.d., $n = 3$ independent experiments. Analysis was performed using one-way ANOVA with Dunnett's post hoc test. Analysis shown in **c** was performed once; experiments in **d–g** were repeated three times with similar results.

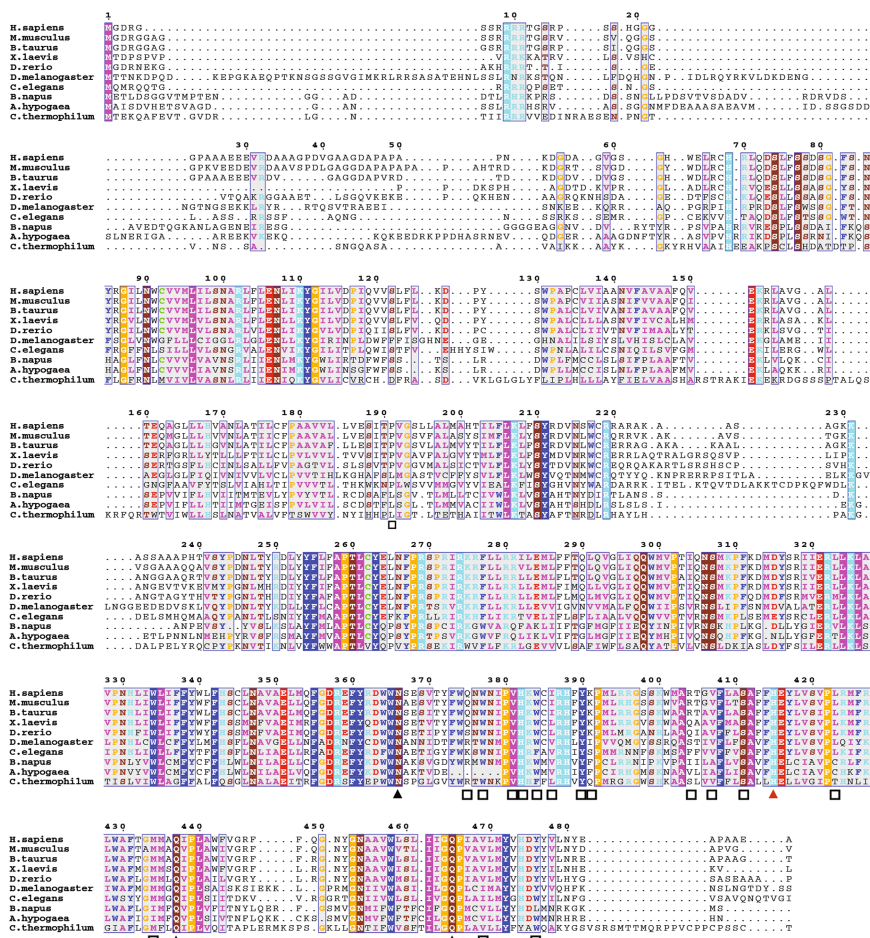


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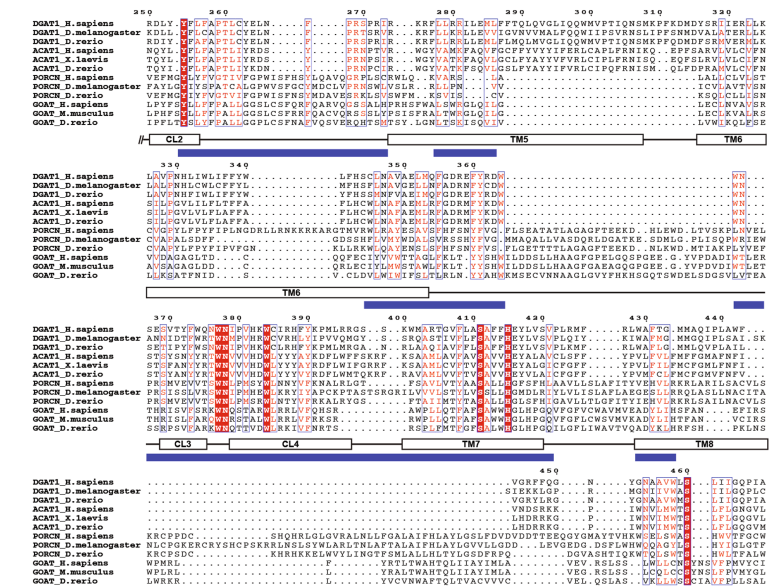
Extended Data Fig. 5 | Structural comparison of DGAT1 with DltB. **a**, Structural superposition of DGAT1 (blue) and DltB (orange, PDB ID: 6BU1)¹⁷. The two structures were superimposed with an r.m.s.d. of 1.39 Å over 360 matched C α positions. **b**, As **a**, but the protein structures are shown as cylinders with the conserved histidine residue shown as sticks. The area denoted by the red dashed line exhibits a similar overall architecture in DGAT1 and DltB (MBOAT core) that is not found in other protein structures. Note that beyond this common area, the resolved N-terminal and TM1 regions in DGAT1 appear as extra structural elements beyond the MBOAT-core region as compared to DltB.

c, Structure-based sequence alignment of DGAT1 and DltB. The squiggles on the top represent α -helices in DGAT1 (blue) and DltB (orange). Sequences in the black rectangle indicate the most structurally variable region in the two enzymes. In DGAT1, these regions are involved in dimer formation, whereas in DltB, the equivalent two helices (TM1 and TM2) seal off the lateral cavity within the membrane (see main text). Sequence in green rectangle denotes the alanyl-donor binding pocket in DltB. The red triangle denotes the conserved histidine residue. The alignment was performed with PROMALS3D⁴³, and the final alignment figure was generated with ESPript 3.0⁴⁴.

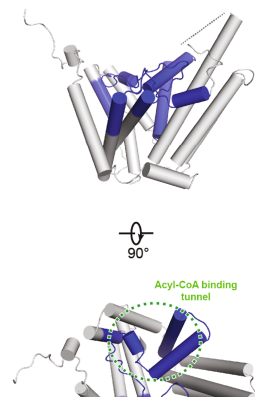
a



b



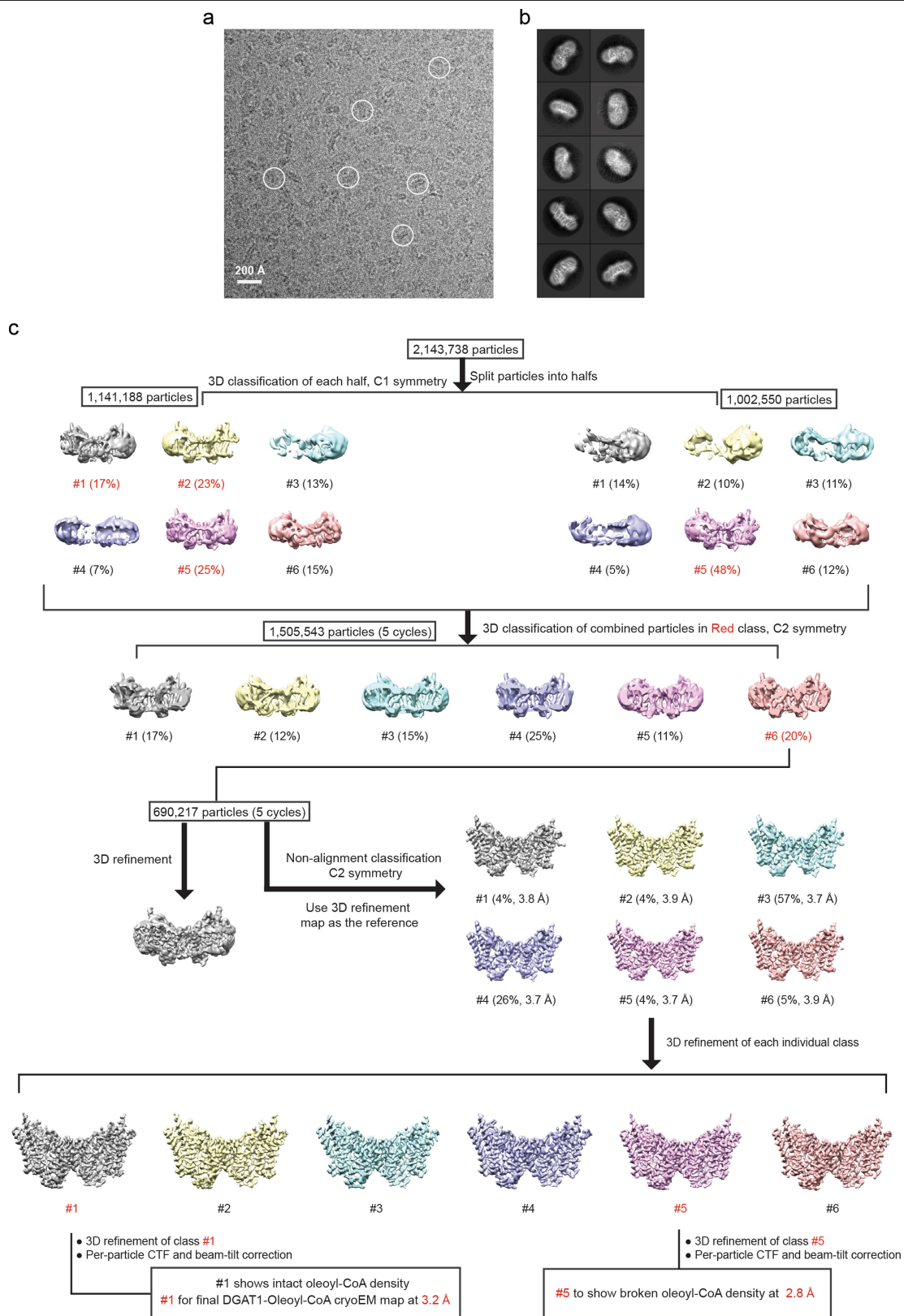
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Extended Data Fig. 6 | See next page for caption.

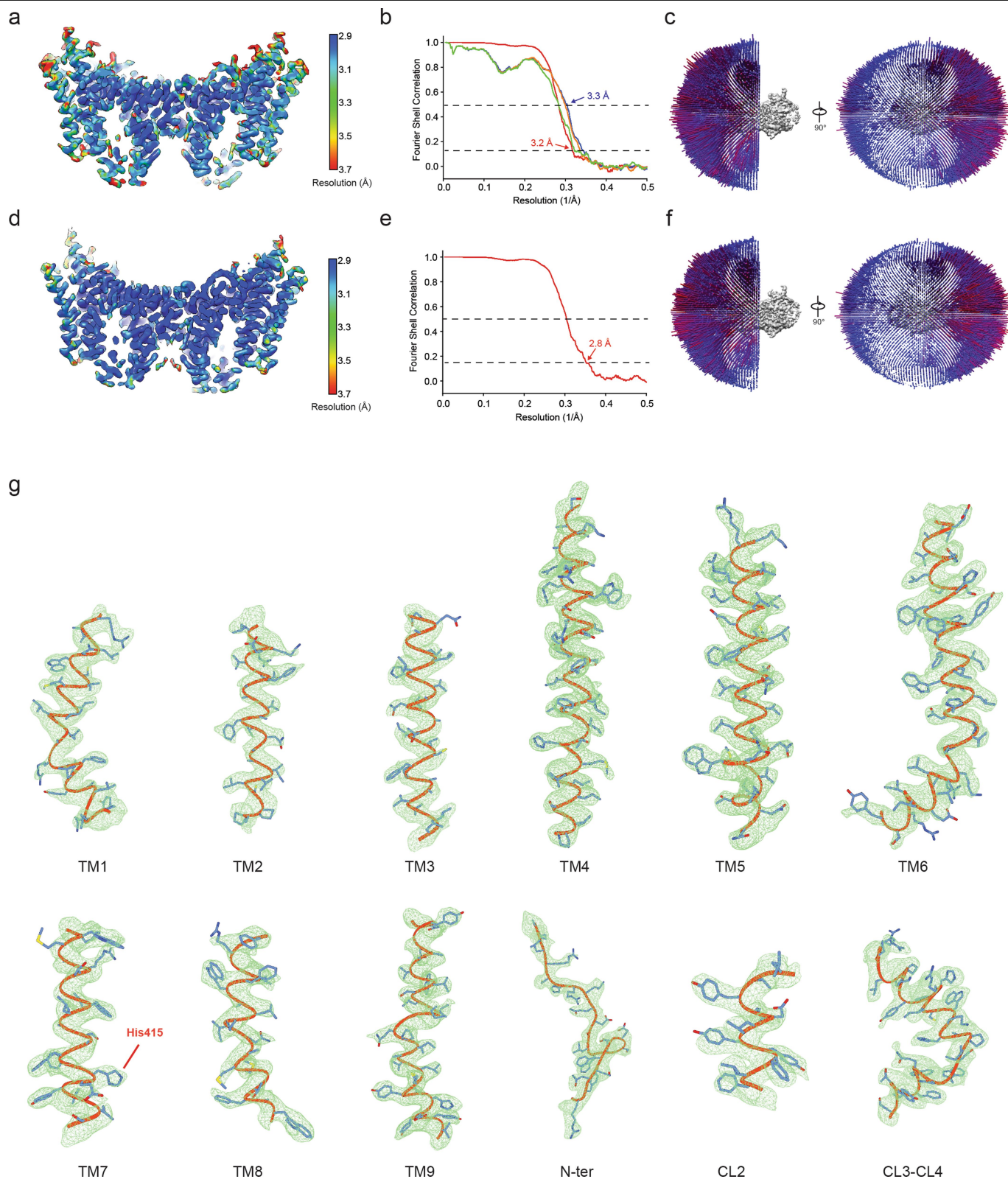
Extended Data Fig. 6 | DGAT1 sequence alignment and conservation analysis of MBOAT enzymes. **a**, Sequence alignment of DGAT1 enzymes from *Homo sapiens*, *Mus musculus*, *Bos taurus*, *Xenopus laevis*, *Danio rerio*, *Drosophila melanogaster*, *Caenorhabditis elegans*, *Brassica napus*, *Arachis hypogaea* and *Chaetomium thermophilum*. The colour scheme of amino acids is based on their physicochemical properties. A red triangle denotes the highly conserved histidine residue; black triangles denote highly conserved polar residues in the DGAT1 active centre; black squares denote residues interacting with oleoyl-CoA. The residue numbers for human DGAT1 are indicated at the top. The sequence alignment was performed with T-Coffee⁴⁵, and the final

alignment figure was generated with ESPrpt 3.0⁴⁴. **b**, Sequence alignment of MBOAT enzymes that acylate lipids (DGAT1 and ACAT1) or proteins (PORCN and GOAT). Structural information of DGAT1 was incorporated into the alignment, where the regions containing a cluster of conserved residues among MBOAT enzymes were labelled as blue boxes. Note the alignment starts at Arg250 of DGAT1. **c**, Mapping the conserved blue region shown in **b** into the DGAT1 structure. The DGAT1 structure is shown as grey cylinders. Blue areas denote the conserved region among MBOATs. Note the acyl-CoA binding tunnel in DGAT1 containing the most conserved region among MBOATs (blue area in **b**) is highlighted by the dashed circle.



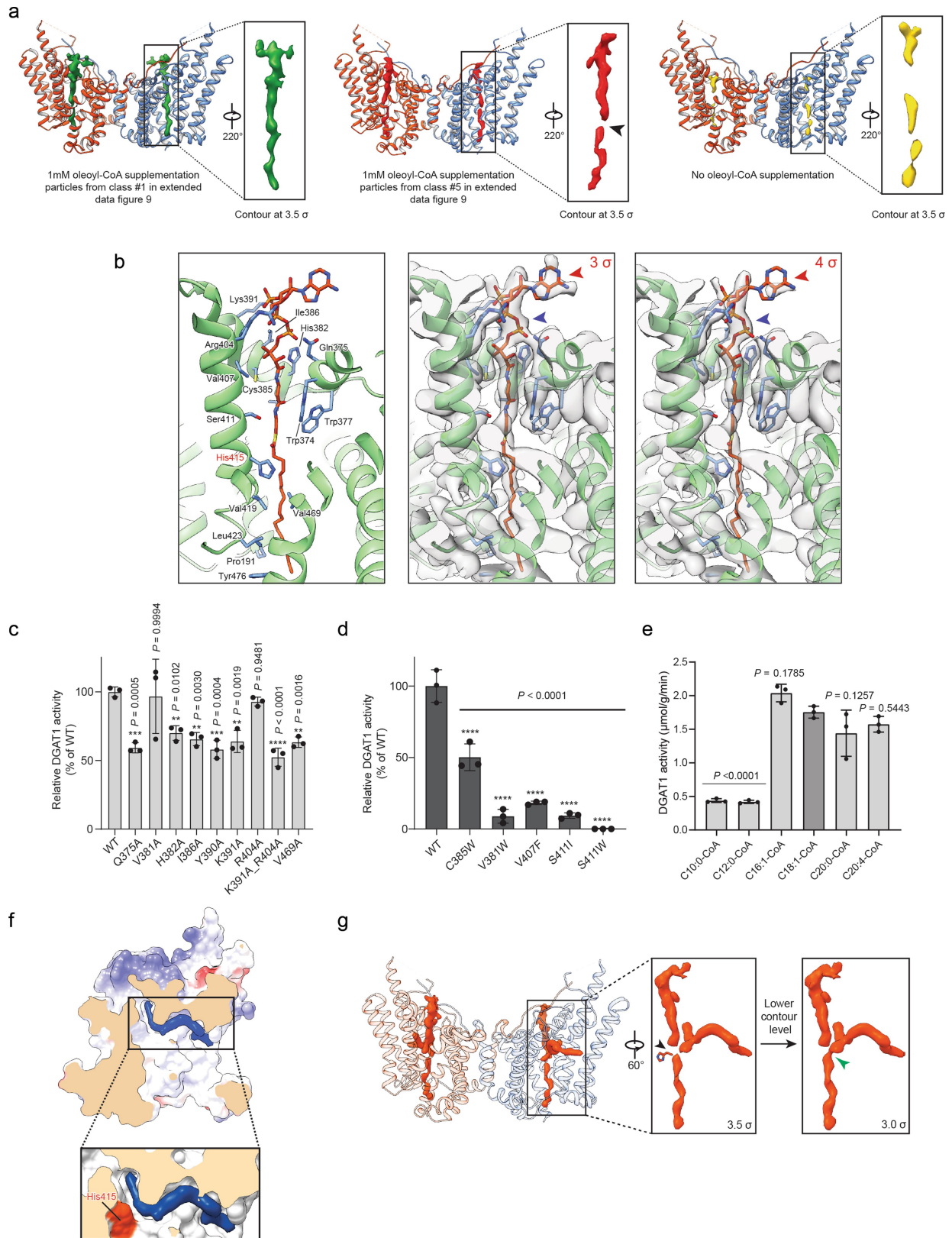
Extended Data Fig. 7 | Cryo-EM image processing of human DGAT1 supplemented with acyl-CoA substrate. **a**, Representative cryo-EM image of DGAT1 with oleoyl-CoA. Some DGAT1 particles are marked by circles. **b**, 2D class averages of cryo-EM particle images with the box size of 210 Å. **c**, Three-dimensional classification and refinement of cryo-EM particle images. The processing flow is similar to the dataset without acyl-CoA treatment (Extended Data Fig. 2). After the second round of 3D classification, all particles

classified into class 6 during the final five iterations (indicated as ‘5 cycles’) were kept for next round of non-alignment classification into six classes. Particles in each class (estimated resolution shown in parentheses) were individually refined. After per-particle CTF and beam-tilt corrections, two resulting maps were chosen to represent DGAT1 complexed with the intact and broken oleoyl-CoA molecule.



Extended Data Fig. 8 | Single-particle cryo-EM analysis of DGAT1 with acyl-CoA substrate. **a**, Local resolution of the final cryo-EM map of DGAT1 (shown as sliced view) complex with an intact oleoyl-CoA molecule. **b**, FSC curves: gold-standard FSC curve between the two half maps with indicated resolution at FSC = 0.143 (red); FSC curve between the atomic model and the final map with indicated resolution at FSC = 0.5 (blue); FSC curve between half map 1 (orange) or half map 2 (green) and the atomic model refined against half

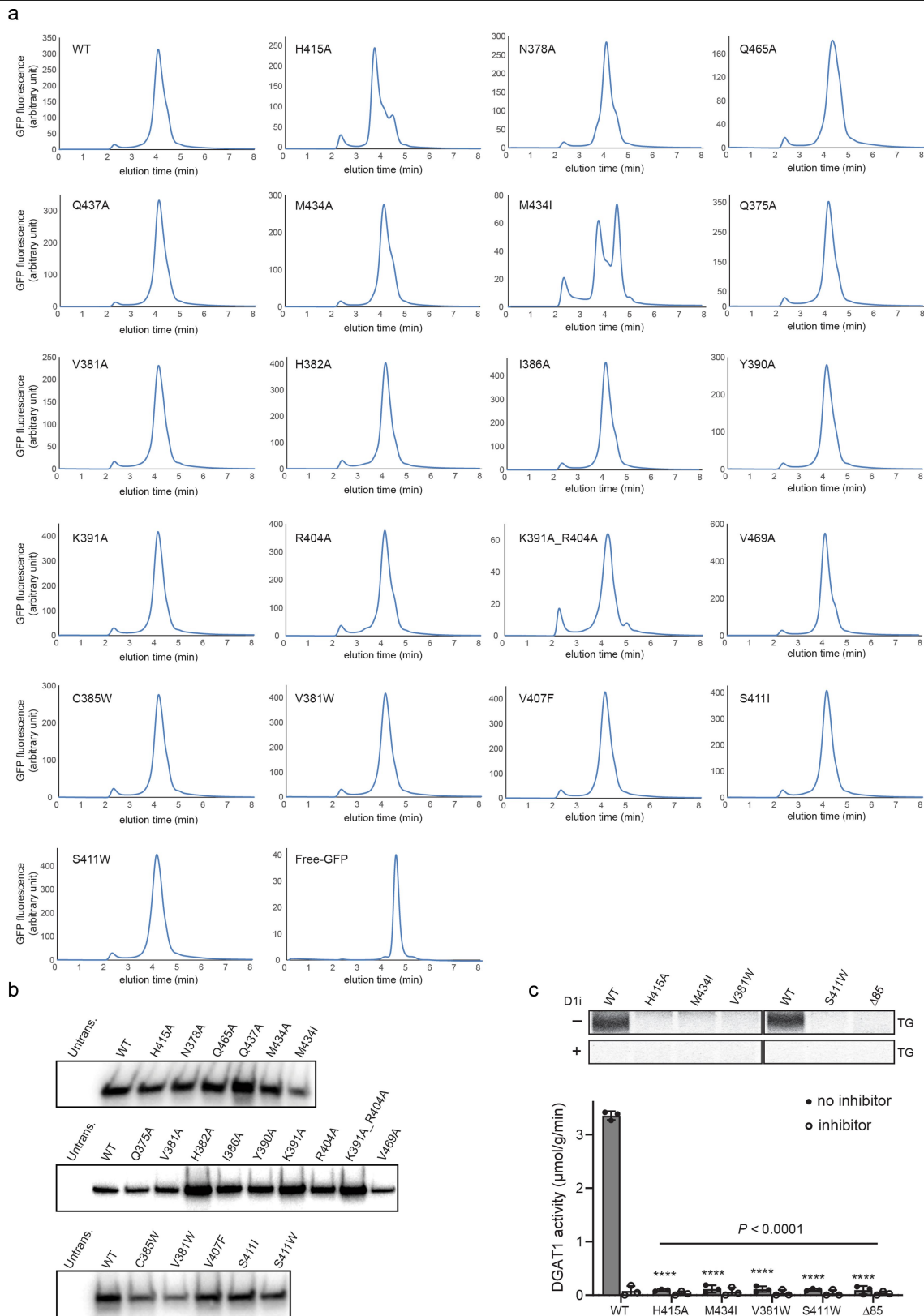
map. **c**, Cutaway views of angular distribution of particle images included in the final 3D reconstruction. **d-f**, Similar to **a-c**, but showing the cryo-EM analysis of the map with a broken oleoyl-CoA density. Note that in **e**, only the FSC curve between half maps was calculated. **g**, Cryo-EM densities with the intact oleoyl-CoA density (shown in **a-c**) superimposed with the atomic model for individual transmembrane helices similar to that shown in Extended Data Fig. 3d. Maps are contoured at 4 σ .



Extended Data Fig. 9 | See next page for caption.

Extended Data Fig. 9 | Cryo-EM density analysis of the bound acyl-CoA and lipid-like density and mutagenesis study of residues interacting with oleoyl-CoA substrate. **a**, A side-by-side density comparison of the bound oleoyl-CoA molecule, oleoyl-CoA with a broken density, and density in the acyl-CoA binding pocket without oleoyl-CoA treatment. **b**, Cryo-EM density map of the oleoyl-CoA molecule and the surrounding region. Maps are shown at different contour levels to illustrate mobility of the adenosine moiety (red arrowhead) of oleoyl-CoA. By contrast, the phosphopantetheine region (green arrowhead) exhibits a stronger density signal. **c**, Activities of acyl-CoA binding tunnel alanine mutants of DGAT1 expressed in DGAT1-knockout cells. **d**, Activities of the bulky side-chain substitutions of residues in the acyl-CoA binding tunnel. Activities in **c** and **d** were normalized to the amount of DGAT1 protein expressed (mean \pm s.d., $n = 3$ independent experiments). Analysis was

performed using one-way ANOVA with Dunnett's post hoc test. **e**, Effect of the acyl chain of the acyl-donor substrate on DGAT1 activity. The analyses used purified DGAT1 in amphipol (mean \pm s.d., $n = 3$ independent experiments). Analysis was performed using one-way ANOVA with Dunnett's post hoc test comparing C18:1-CoA to other acyl-CoA substrates (coloured in dark grey). **f**, The zoomed-in views of the lipid-like density in the oleoyl-CoA-free DGAT1 monomer structure shown as surface representation. The protein surface at His415 in the zoomed-in view is coloured in orange. The map is contoured at 3.5σ . **g**, Cryo-EM density in orange showing both the bound acyl-CoA with a broken signal and the lipid-like density. A black arrowhead denotes the disconnected acyl chain in oleoyl-CoA molecule. A cyan arrowhead indicates the connected density between the acyl chain of oleoyl-CoA and the uncharacterized lipid-like density at a lower contour level.



Extended Data Fig. 10 | FSEC, western blot and activity analyses of DGAT1 mutants. **a**, FSEC profiles of wild type and all tested mutants in this study. Wild-type DGAT1 and mutants were transiently expressed as GFP-fusion proteins in HEK293 cells. The folding of each mutant was analysed by size-exclusion chromatography monitoring GFP fluorescence. All tested mutants exhibit peaks similar to the wild-type DGAT1 protein, suggesting their correct overall protein folding. **b**, Western blot analyses of wild-type and tested

DGAT1 mutant proteins expressed in SUM159 DGAT1-knockout cells. Untrans., untransfected control. **c**, Activity of wild type and selected DGAT1 mutants purified by His-tag affinity chromatography purification. The top panel of the raw TLC plates shows the formation of triacylglycerol. The assays were performed with or without DGAT1 inhibitor (D1i) (mean \pm s.d., $n = 3$ independent experiments). One-way ANOVA with Dunnett's post hoc test was applied. Experiments shown in **a-c** were repeated twice with similar results.

Extended Data Table 1 | Cryo-EM data collection, refinement and validation statistics

	DGAT1 in PMAL-C8 (EMD-21461) (PDB 6VYI)	DGAT1 with intact oleoyl-CoA (EMDB-21481) (PDB 6VZ1)	DGAT1 with broken oleoyl-CoA density (EMDB-21488)
Data collection and processing			
Magnification	105,000	105,000	105,000
Voltage (kV)	300	300	300
Electron exposure (e-/Å ²)	52	43	43
Defocus range (µm)	1.5-3.0	1.0 - 2.5	1.0 - 2.5
Pixel size (Å)	0.825	0.83	0.83
Symmetry imposed	C2	C2	C2
Initial particle images (no.)	2,161,707	2,143,738	2,143,738
Final particle images (no.)	61,608	28,165	27,750
Map resolution (Å)	3.0	3.2	2.8
FSC threshold	0.143	0.143	0.143
Map resolution range (Å)	211.2 - 3.0	212.5 - 3.2	212.5 - 2.8
Refinement			
Initial model used (PDB code)	NA	NA	
Model resolution (Å)			
FSC threshold	0.143	0.143	
Model resolution range (Å)	211.2 - 3.0	212.5 - 3.2	
Map sharpening <i>B</i> factor (Å ²)	-120	-103	
Model composition			
Non-hydrogen atoms	6916	7112	
Protein residues	812	822	
Ligands	4	6	
<i>B</i> factors (Å ²)			
Protein	53.8	67.4	
Ligand	44.4	67.7	
R.m.s. deviations			
Bond lengths (Å)	0.01	0.01	
Bond angles (°)	0.74	0.85	
Validation			
MolProbity score	2.1	2.1	
Clashscore	9.3	9.9	
Poor rotamers (%)	0	0	
Ramachandran plot			
Favored (%)	88.8	87.9	
Allowed (%)	11.2	12.1	
Disallowed (%)	0	0	

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For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

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- The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
- A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided
Only common tests should be described solely by name; describe more complex techniques in the Methods section.
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
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- For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
Give P values as exact values whenever suitable.
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection

Serial EM version 3.6

Data analysis

Phenix (v 1.16); Coot (v 0.8.9); MotionCor2 (v 1.1.0); CTFFIND4 (v 4.1.5); SAMUEL (v 17.05); SamViewer (v 16.01); SPIDER (v 17.05); RELION 3.0; ResMap (v 1.1.4); bfactor (v 1.03); Chimera (v 1.13); LigPlot (v 2.2)

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- Accession codes, unique identifiers, or web links for publicly available datasets
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- A description of any restrictions on data availability

Three cryoEM maps have been deposited into the Electron Microscopy Data Bank under the accession code EMD-21461, EMD-21481, and EMD-21488; Two protein coordinates have been deposited in the Protein Data Bank with the accession code 6VYI and 6VZ1.

Field-specific reporting

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Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	No statistical methods were used to predetermine sample size. Sufficient EM data were collected to achieve adequate single-particle EM analysis and 3D cryo-EM reconstructions.
Data exclusions	No data were excluded from analyses.
Replication	Each experiment was repeated at least two times in independent experiments. Experimental findings were reproduced reliably.
Randomization	This is not relevant to our study, because no grouping was needed.
Blinding	Investigators were not blinded to group allocation, because no grouping was needed for this study.

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We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Involvement in the study
<input checked="" type="checkbox"/>	<input type="checkbox"/> Antibodies
<input type="checkbox"/>	<input checked="" type="checkbox"/> Eukaryotic cell lines
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<input checked="" type="checkbox"/>	<input type="checkbox"/> Human research participants
<input checked="" type="checkbox"/>	<input type="checkbox"/> Clinical data

Methods

n/a	Involvement in the study
<input checked="" type="checkbox"/>	<input type="checkbox"/> ChIP-seq
<input checked="" type="checkbox"/>	<input type="checkbox"/> Flow cytometry
<input checked="" type="checkbox"/>	<input type="checkbox"/> MRI-based neuroimaging

Eukaryotic cell lines

Policy information about [cell lines](#)

Cell line source(s)	HEK293S GnTI- cell (ATCC, CRL-3022). SUM159 CRISPR DGAT1 knock out cell (created from ATCC, CRL-2915)
Authentication	No further authentications were performed for this study.
Mycoplasma contamination	No mycoplasma contamination test was performed for this study.
Commonly misidentified lines (See ICLAC register)	None of the cell lines used are listed in the ICLAC database.