REVIEW ARTICLE

IL7RA genetic variants differentially affect IL-7Rα expression and alternative splicing: a role in autoimmune and infectious diseases?

Christia[n](http://orcid.org/0000-0002-4703-5652) Lundtoft $\bigcirc^1 \cdot$ $\bigcirc^1 \cdot$ $\bigcirc^1 \cdot$ Julia Sevfarth $\bigcirc^1 \cdot$ Marc Jacobsen \bigcirc^1

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Abstract

Interleukin-7 receptor α chain (IL-7R α) single nucleotide polymorphisms (SNPs) are associated with susceptibility to immunopathologies like autoimmune and inflammatory diseases. The current hypothesis about underlying mechanisms is based on the regulation of IL-7 availability for self-reactive T cells by influencing the generation of a soluble (s)IL-7R α variant. This assumption was mainly predicated on the well-defined IL7RA SNP rs6897932, which affects alternative splicing and causes aberrant generation of the sIL-7R α variant with potential effects on the IL-7 serum reservoir. However, more recent studies shed light on novel functions of autoimmunity risk-associated IL7RA SNPs and characterized the largely neglected effect of rs6897932 on membrane (m)IL-7Rα expression. These findings as well as a described role of impaired mIL-7R α expression and IL7RA SNP influence on chronic infectious diseases necessitates the reevaluation of previous findings on the role of IL7RA SNPs in immunopathology.

Introduction

Interleukin (IL)-7 is an important cytokine for T-cell development, homeostasis, and the generation of immunological memory. Fine balanced regulation of IL-7 availability and T-cell response to IL-7 is crucial since dysregulation may contribute to development of autoimmunity and impaired immunity against chronic infections. An important role of IL-7 receptor α-chain (IL-7Rα) variants on regulation of IL-7 availability—especially for the soluble IL-7Rα—has been assumed.

Gain-of-function and loss-of-function mutations of the IL7RA gene either cause malignancies due to uncontrolled proliferation or abrogate development of T cells. Other IL7RA polymorphisms (the best-defined example is the IL7RA exon 6 SNP rs6897932) exert more moderate effects, e.g., on alternative splicing and expression of the soluble

These authors contributed equally: Christian Lundtoft, Julia Seyfarth

 \boxtimes Marc Jacobsen marc.jacobsen@med.uni-duesseldorf.de IL-7Rα. rs6897932 and a subset of other single nucleotide polymorphisms (SNPs) (e.g., rs1494555, rs1494558) have been shown to tag haplotypes associated with susceptibility to autoimmune diseases. Besides effects on soluble IL-7Rα expression and regulation of IL-7 availability, recent findings shed light on additional SNP-mediated mechanisms affecting other IL-7R α variants and the membranous IL-7Rα. This review summarizes findings on IL7RA SNPs with moderate regulatory effects and their potential role on T-cell function. Furthermore, it is discussed how IL7RA SNPs may affect immune pathology of autoimmune and infectious diseases.

IL-7 availability for T cells and IL-7 receptor dependent regulation in immune homeostasis and pathology

IL-7 plays a crucial role for T-cell development and function. This is well established since both absence and aberrant strong signaling of IL-7 result in immune pathology. Abrogated IL-7 signaling leads to severe combined immunodeficiency demonstrating the crucial and nonredundant role during lymphocyte (especially T-cell) development [[1\]](#page-4-0). In contrast, aberrant high IL-7 signaling causes lymphoproliferation and contributes to development of T-cell acute lymphoblastic leukemia [\[2](#page-5-0), [3\]](#page-5-0). Both types of immune

¹ Department of General Pediatrics, Neonatology, and Pediatric Cardiology, Medical Faculty, University Children's Hospital, Moorenstr. 5, 40225 Duesseldorf, Germany

pathology in patients are caused by germline or somatic mutations, which predominantly affect the IL-7 receptor, either the private IL-7 receptor α (IL-7R α) or the common-γ receptor chain [[4\]](#page-5-0). These immune pathologies demonstrate that regulation of IL-7 responses on the level of IL-7R expression is central for balanced IL-7 functions. Several studies investigated the complex mechanisms how IL-7R expression is regulated to assure appropriate T-cell functions, and these have been reviewed elsewhere [[5\]](#page-5-0). These studies were the basis for immune intervention strategies using IL-7R α blocking antibodies, which strengthened the important role of $IL-7R\alpha$ signaling in pathology of autoimmune and inflammatory diseases [\[6](#page-5-0)–[10](#page-5-0)], as well as transplantation [[11\]](#page-5-0). These promising results formed the basis for first clinical trials of anti-IL-7Rα treatment in humans [\[12](#page-5-0), [13](#page-5-0)].

However, changes in IL-7 availability may also lead to immune pathology, as aberrant high IL-7 serum levels, e.g., during lymphopenia, increase the risk for T-cell mediated autoimmunity [\[14](#page-5-0)]. Such high IL-7 serum concentrations are mainly occurring as a consequence of immune suppressive treatment (e.g., bone marrow irradiation, anti-Tcell immune globulins or biologicals) as well in primary and secondary immune deficiencies [[15,](#page-5-0) [16\]](#page-5-0). As part of recovery, IL-7 induces increased proliferation of T cells and the growing T-cell population consumes more IL-7. This feedback loop mechanism leads to normalized IL-7 levels at physiological T-cell population size. Therefore, also under lymphopenic conditions and high IL-7 levels, regulation by IL-7R expressing T cells is decisive for IL-7-mediated homeostasis [[5\]](#page-5-0). There is initial evidence that certain SNPs in the IL7RA gene affect IL-7-dependent mechanisms during lymphopenia with clinically relevant consequences, i.e., recovery from lymphopenia in HIV/AIDS patients [\[17](#page-5-0)–[20](#page-5-0)], or risk for development of graft-versus-host disease (GVHD) after bone marrow transplantation $[21-23]$ $[21-23]$ $[21-23]$ $[21-23]$. These relevant SNPs were also shown to influence susceptibility to autoimmune and inflammatory diseases [\[4](#page-5-0)], and, hence, attracted strong interest of the scientific community during the past two decades. Our advanced knowledge on regulation of IL-7R variants is strongly based on studies characterizing effects mediated by the IL7RA exon 6 SNP rs6897932.

The autoimmunity-associated SNP rs6897932 affects alternative splicing and expression of the soluble IL-7Rα variant

The membrane-associated IL-7R forms as a heterodimer composed of the IL-7Rα and the common-γ receptor chain [\[24,](#page-5-0) [25\]](#page-5-0). The IL-7R α chain binds IL-7 specifically, and recruitment of the common γ-chain receptor increases

affinity and catalyzes signaling [[26](#page-5-0)]. Like for other cytokines and hormone receptors, a soluble (s)IL-7R variant is generated [[27](#page-5-0)]. Although few studies identified shedding of mIL-7R α as the underlying mechanism [\[28,](#page-5-0) [29](#page-5-0)], it is now largely accepted that alternative mRNA splicing is the dominant mechanism for $sIL-7R\alpha$ generation. Secreted sIL-7Rα monomers result from exclusion of the IL7RA exon 6, coding for the transmembrane domain of the IL-7R α chain [[30\]](#page-5-0). This causes a shift in the reading frame and generates a new stop codon early in the exon 8 [\[27,](#page-5-0) [31,](#page-5-0) [32\]](#page-5-0). A truncated IL-7R α chain lacking transmembrane and intracellular domains is translated leading to sIL-7Rα secretion [[33](#page-5-0)]. The sIL-7Rα received sudden utmost interest when Gregory et al. reported association of a nonsynonymous SNP variant (i.e., rs6897932C > T; Thr \rightarrow Ile) within the exon 6 of the IL7RA gene with susceptibility to multiple sclerosis [[34\]](#page-5-0). They found that the protective allele (i.e., rs6897932T) interfered with IL7RA exon 6 splicing [\[34](#page-5-0)], and lower sIL-7R α serum levels were found in carriers of the protective IL7RA allele [\[35](#page-5-0)–[37](#page-6-0)]. Since then, several studies demonstrated the relevance of rs6897932T for protection against autoimmune and inflammatory diseases [[4,](#page-5-0) [38](#page-6-0)–[46](#page-6-0)]. In addition, Galarza-Munoz et al. found epistatic regulation of the rs6897932 SNP by a DDX39B allele (associated with multiple sclerosis susceptibility) that markedly strengthened the effect of alternative IL7RA splicing [[47](#page-6-0)]. These studies suggested a functional role of the sIL-7R α and as shown for other soluble hormone and cytokine receptors—interference with IL-7 availability and consumption was assumed. In accordance, inhibition of IL-7 signaling by a sIL-7R-Fc chimera in vitro has been found [[48](#page-6-0), [49](#page-6-0)]. Lundstrom et al. provided a model for sIL-7Rα-mediated IL-7 regulation in vivo, showing that sIL-7R α (at a specific concentration ratio relative to IL-7) may increase bioactivity of IL-7 by stabilizing the IL-7 reservoir [[35](#page-5-0)]. The sIL-7R α binds IL-7 with moderate affinity [k_D = 6 nM], but interference with high-affinity mIL-7R complex/IL-7 can be assumed because of high molar excess of sIL-7R α in the serum (i.e., about 10³ times more than IL-7) [\[35\]](#page-5-0). A role of the sIL-7R α in ensuring IL-7 availability would strengthen the thesis that links high sIL-7 $R\alpha$ serum concentrations with increased risk for generation of IL-7 dependent self-reactive T cells in autoimmunity [[50](#page-6-0), [51](#page-6-0)]. In accordance, lower IL-7 concentrations were found in autoimmune disease patients carrying the protective rs6897932T genotype that causes decreased sIL-7Rα serum levels [[35](#page-5-0), [52](#page-6-0), [53\]](#page-6-0).

Previous studies strongly focused on rs6897932Tmediated effects on the sIL-7R α and this can be explained by the fact that expected alternative splicing effects of rs6897932 SNPs on mIL-7Rα expression of T cells were not detected [\[48](#page-6-0)] (own unpublished data). There is now

increasing evidence that the SNP-associated mechanisms affecting IL-7 levels are more complex and that sIL-7Rαindependent mechanisms of IL-7 regulation are involved.

The autoimmunity risk-associated SNP rs1494558 causes lower sIL-7Rα expression

The nonsynonymous SNP (i.e., $rs1494558C > T$; Thr \rightarrow Ile) located within the IL7RA exon 2 tags an autoimmunity riskassociated haplotype [[34](#page-5-0), [54\]](#page-6-0). This SNP is in strong linkage equilibrium with the rs1494555 SNP in exon 4 that has originally been identified [\[34\]](#page-5-0). We characterized rs1494558 effects on IL-7Rα and found decreased soluble and membranous IL-7R expression in rs1494558Ile-transfected cell lines [\[55\]](#page-6-0). Decreased sIL-7R α secretion of the rs1494558Iletagged haplotype was confirmed in T1D patients, who had lower sIL-7Rα serum concentrations when carrying rs1494558Ile [[55](#page-6-0)]. In the same study, we confirmed that also T1D patients with the rs6897932Ile haplotype had lower sIL-7Rα serum levels indicating that both autoimmunity risk- and protection-associated haplotypes are characterized by a similar pattern. Lower IL-7 serum levels of T1D patients, however, were exclusively found for the protection-associated rs6897932Ile haplotype [\[53](#page-6-0)]. These results, and the fact that no correlation was detected between sIL-7Rα and IL-7 serum levels from individual T1D patients $[53]$ $[53]$ $[53]$, argue for sIL-7R α independent mechanisms of IL7RA haplotypes affecting IL-7 availability.

Differential mIL-7Rα expression is an alternative possible explanation for contrary effects mediated by rs6897932Ile and rs1494558Ile variants. To proof this assumption, we generated a cell line-based model for concomitant assessment of risk- and protection-associated variants on sIL-7Rα and mIL-7Rα expression. As expected, both variants showed similar effects on sIL-7Rα secretion, but only rs6897932Ile led to a significant increase of mIL-7R α expression [\[55\]](#page-6-0). This was in accordance with the predicted effects of alternative splicing on full-length vs. shortened IL-7R mRNA expression. Jager et al. detected increased mIL-7Rα expression NK cells from multiple sclerosis patients carrying the protective haplotype tagged by rs6897932 [\[48\]](#page-6-0). However, no mIL-7Rα differences for T cells associated with the rs6897932-tagged haplotype were found [\[48\]](#page-6-0) (own unpublished data). Possible explanations are the high variability of mILα-7R expression due to marked differences between T-cell subpopulations as well as described regulatory mechanisms of mIL-7Rα expression and internalization [\[56](#page-6-0), [57\]](#page-6-0). These mechanisms may also restrict mIL-7Rα expression to a plateau of maximal expression potentially hiding genotype-specific effects. Initial in vitro studies to characterize genotype effects on IL-7Rα regulation of T cells, however, did not identify differences (own unpublished data). In conclusion, although previous studies suggested IL7RA haplotype effects on mIL-7R α expression, the biological relevance has to be proven.

An IL7RA haplotype found in African populations affects alternative splicing of exon 5

The vast majority of previous studies focused on SNPs associated to autoimmune diseases and potential implications on sIL-7Rα regulation. There is, however, a significant number of additional SNPs in the IL7RA locus and also additional variants, besides the full length and the $sIL-7R\alpha$, have been described [\[31\]](#page-5-0). Based on exonic SNPs with relevance for autoimmune diseases, previous studies defined four haplotypes of the IL7RA gene in Caucasian populations (Fig. [1](#page-3-0)) [\[34\]](#page-5-0). Haplotype 1 is associated with increased risk to develop autoimmune diseases (tagged by the rs1494555G/ rs1494558T alleles). Haplotype 3 is associated with a decreased risk to develop an autoimmune disease (tagged by the rs6897932T allele). Altogether these haplotypes comprise more than 99% of the Caucasian population (Fig. [1\)](#page-3-0). Own studies in Sub-Saharan Africa showed that especially the haplotypes 1 and 3 are less frequent in this African population, whereas two additional haplotypes contribute significantly to the variability of the IL7RA gene locus [\[52\]](#page-6-0). Almost 40% of individuals from Kumasi/Ghana are haplotype 5 or 6 carriers (Fig. [1\)](#page-3-0). Haplotype 6 is tagged by the rs11567764A allele and also includes the haplotype 1-tagging rs1494558T allele (Fig. [1\)](#page-3-0). In a case/control study comparing tuberculosis patients with healthy contacts, we detected increased frequencies of individuals homozygous for the rs11567764A allele in the healthy contact group [[52](#page-6-0)]. In addition, carriers of the rs1494558T SNP were more frequent in the contact group [\[52\]](#page-6-0). Contacts of tuberculosis patients are frequently infected by Mycobacterium (M.) tuberculosis without developing tuberculosis disease because of successful immune surveillance. Hence, higher frequencies of the rs11567764A/rs1494558T alleles suggested potential involvement in disease protection. Since the rs1494558T allele is found in two IL7RA haplotypes in Africans, it is not possible to clearly assign the relevant haplotype for differential susceptibility. However, we identified functional implications for both alleles, rs1494558T (described above) and rs11567764A [\[52\]](#page-6-0). rs11567764G > A is a synonymous SNP that does not affect IL7RA coded amino acids. Located within exon 5, however, rs11567764A was predicted to affect alternative splicing of the IL7RA [\[52\]](#page-6-0). T-cell analyses of tuberculosis patients and contacts confirmed significantly reduced expression of a IL-7Rα variant lacking exon 5–6 (Δexon_5/6) in rs11567764A carriers, whereas mRNA coding for the sIL-7R α (i.e., Δ exon 6) was not affected [\[52](#page-6-0)]. How differences in the expression of the Δexon_5/6 mRNA may influence T-cell

Fig. 1 IL-7R α exon composition, cellular localization, and exonic SNPs are schematically shown. Respective amino acid exchanges are indicated. The table summarizes deduced haplotypes of exonic SNPs and indicates frequencies for Caucansian and African populations.

Ile: isoleucine; Thr: threonine; Val: valine; His: histamine; Lys: lysine; TM: transmembrane; ID: identifier; EU: Caucasian population; AFR: African population.

function remains elusive. Since proper folding and secretion of this variant is unlikely, also indirect effects on, e.g., other alternatively spliced IL-7R variants would be possible. We assume that such differences can be of relevance for T-cell function against infection. In accordance with this assumption, aberrant low sIL-7Rα serum levels and low mIL-7Rα expression accompanied by impaired antimycobacterial functions were found in acute tuberculosis patients [\[58\]](#page-6-0). Future studies may address a potential role of the novel haplotypes in other infectious and autoimmune diseases in African populations.

IL7RA SNPs effects on immune pathology of autoimmune and GVHD entities

IL7RA SNP effects have been described for several diseases, and strongest evidence came from studies on multiple sclerosis [\[34,](#page-5-0) [43](#page-6-0), [44](#page-6-0), [46,](#page-6-0) [59](#page-6-0)–[66\]](#page-6-0). Also, for other diseases with autoimmune pathology (i.e., type 1 diabetes [\[39](#page-6-0), [42](#page-6-0), [67](#page-6-0), [68\]](#page-6-0), systemic lupus erythematosus [[45](#page-6-0)]) as well as inflammatory diseases (atopic dermatitis [[41\]](#page-6-0), sarcoidosis [\[40\]](#page-6-0), arthritis [\[38\]](#page-6-0)), an influence of *IL7RA* SNPs on disease susceptibility was shown. Predominantly, the rs6897932T-tagged haplotype was associated with disease susceptibility, but also other SNPs were identified [\[69](#page-6-0), [70](#page-6-0)]. Functionally, IL-7 availability and promoting effects on the generation/expansion of selfreactive T cells [[35](#page-5-0), [71\]](#page-6-0) as well as potentially inhibitory effects of IL-7 on regulatory T cells are assumed to be causative [\[72\]](#page-7-0). In addition, IL-7 effects early during T-cell development may be affected by other IL7RA SNPs. Broux et al. found that the frequency of recent thymic emigrants (RTEs) was affected in MS patients carrying the IL7RA haplotype 2 (termed "haplotype 4" in this paper, tagged by the rs11567685C allele that is in strong linkage disequilibrium

with rs3194051G) [\[69\]](#page-6-0). RTEs of these MS patients were characterized by moderately increased IL-7Rα expression levels. Additional studies are needed to elucidate the underlying mechanisms and to identify the causative SNP.

Lymphopenia-induced autoimmunity strengthened the central role of IL-7 in disease pathogenesis [\[14\]](#page-5-0). Interestingly, lymphopenia induced by bone marrow irradiation and subsequent transplantation of hematopoietic stem cells was shown to be affected by the IL7RA rs6897932T allele as well [\[21](#page-5-0)–[23](#page-5-0)]. Here, T-cell mediated GVHD is a frequent and harmful side effect correlating with high IL-7 serum concentration [\[73](#page-7-0)]. Donor cell IL7RA haplotypes were described to be associated with treatment related mortality of GVHD, whereas recipient haplotypes were not influential [\[22,](#page-5-0) [23\]](#page-5-0). rs6897932T donor T cells were more often associated with acute as well as chronic GVHD [\[22,](#page-5-0) [23](#page-5-0)]. These studies suggested that high serum IL-7/sIL-7Rα ratios in patients carrying the sIL-7R_{low} rs6897932T-tagged haplotype are causative for T-cell induced GVHD. However, one may also assume rs6897932-mediated effects on mIL-7Rα expression of donor hematopoietic stem cells, which contribute to development of GvHD. Further evidence for a role of the IL7RA in lymphopenia-induced autoimmunity comes from patients with new onset diabetes after transplantation (NODAT). Two IL7RA alleles (i.e., rs1494558T, rs2172749C) were associated with increased risk to develop NODAT [[74](#page-7-0)].

A potential role of IL7RA SNPs and IL-7 sensitivity in chronic infectious diseases

In contrast to the aforementioned immune pathologies, a role of IL7RA SNPs in infectious diseases remains enigmatic. Strongest evidence for a role of IL-7 in infections have been provided from studies on HIV/AIDS. Here, IL-7 may play a

dual role being important for immune reconstitution after CD4 $+$ T-cell depletion $[75–78]$ $[75–78]$ $[75–78]$ $[75–78]$ and to sustain effector/memory Tcell functions during antiviral host defense. Early studies demonstrated IL-7-promoted antigen-specific effector T-cell functions including enhanced antiviral cytotoxicity [\[28](#page-5-0), [79\]](#page-7-0). Impaired IL-7 response of antiviral cytotoxic T-cell response and lower IL-7Rα expression have been described for HIV/ AIDS patients [[80](#page-7-0)–[86\]](#page-7-0). Interestingly, gene association studies demonstrated improved immune recovery in treated HIV/ AIDS patients carrying the rs6897932-tagged haplotype [[17](#page-5-0)– [20\]](#page-5-0), whereas contrary results of haplotype effects for disease progression and mortality of HIV-infected individuals were found [\[87,](#page-7-0) [88](#page-7-0)]. Previously we identified rs6897932 SNP effects for tuberculosis patients, where the rs6897932T allele contributed to differential sIL-7Rα serum levels found during acute disease [[58](#page-6-0)]. Frequencies of rs6897932T allele carriers, however, were not different between tuberculosis patients and healthy contacts [[58](#page-6-0)]. Tuberculosis patients were characterized by increased IL-7 and lower sIL-7Rα serum levels. As for HIV/AIDS patients, impaired mIL-7R α expression likely affected IL-7 consumption of tuberculosis patients [\[58\]](#page-6-0). Increased IL-7 serum concentrations were also found in patients with cystic fibrosis who are highly susceptible for repeated and chronic infections [\[89\]](#page-7-0). Although we did not find a correlation with specific infectious agents, we demonstrated clinical relevance since IL-7 serum levels increased during disease course and correlated with decreased lung function of cystic fibrosis patients [[89](#page-7-0)]. Functional T-cell assays could not be performed as part of this study and, hence, it remains elusive if impaired IL-7 consumption was causative as we assume for M. tuberculosis infection.

Future approaches for elucidation of IL-7/IL-7R functions and the role of IL7RA SNPs

Previous studies strongly focused on the sIL-7Rα-mediated function for IL-7-availability. This was comprehensible given the significant effect of the rs6897932 SNP on sIL-7Rα expression and IL-7 serum levels. Novel findings for the role of the rs1494558 SNP in IL-7R α expression and potential effects of the rs6897932 SNP on mIL-7Rα expression should broaden the focus of future studies to elucidate differential effects of disease associated polymorphisms. Lymphopenia-induced autoimmunity and GVHD can be promising models to demonstrate the biological significance of IL7RA SNPs.

In addition to its role in IL-7 response, IL7RA SNPs may also affect signaling of thymic stromal lymphopoetin (TSLP). The TSLP receptor and IL-7R α form the heterodimeric receptor for TSLP, which is an important factor in allergic diseases [[90\]](#page-7-0).

To the best of our knowledge, there are no reports of IL7RA SNP effects on TSLP signaling. Although the TSLP receptor is only weakly expressed on T cells [[91\]](#page-7-0), other immune cell populations that express TSLP receptor and IL-7R α concomitantly (e.g., monocytes after activation [[92\]](#page-7-0)) may be affected by IL7RA haplotype differences. In this regard, Al-Mossawi et al. could recently show that expected rs6897932 SNP effects on differential mIL-7Rα and sIL-7Rα expression are seen for lipopolysaccharide- or TNFαactivated monocytes [\[93](#page-7-0)]. These important observations should broaden our view on the role of IL-7/mIL-7R α beyond lymphoid cell populations and consider potential effects of IL7RA SNPs on TSLP signaling.

Future studies should also investigate potential effects of IL7RA SNPs in chronic infectious diseases and impaired Tcell sensitivity to IL-7. This may affect T-cell exhaustion and memory generation with potential relevance for recurrence of chronic infections. Finally, novel IL7RA SNPs and differences between populations (e.g., between Caucasians and Africans) should be investigated in more detail. How novel IL7RA SNPs affect immune pathology may improve our general understanding of disease mechanisms.

Conclusions

IL7RA genetic variants affect the fine balanced regulation of IL-7 availability and dependent T-cell functions with potential effects on autoimmune and infectious disease pathologies. The influence of sIL-7R α on IL-7 serum level is an important factor but not the only relevant mechanism. Novel functions of IL7RA SNPs rs11567764, rs1494558, and rs6897932 may contribute to our understanding of underlying mechanisms, especially the role of membranous IL-7R expression in immune pathology.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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References

1. Puel A, Ziegler SF, Buckley RH, Leonard WJ. Defective IL7R expression in $T(-)B(+)NK(+)$ severe combined immunodeficiency. Nat Genet. 1998;20:394–7.

- 2. Shochat C, Tal N, Bandapalli OR, Palmi C, Ganmore I, te Kronnie G, et al. Gain-of-function mutations in interleukin-7 receptoralpha (IL7R) in childhood acute lymphoblastic leukemias. J Exp Med. 2011;208:901–8.
- 3. Zenatti PP, Ribeiro D, Li W, Zuurbier L, Silva MC, Paganin M, et al. Oncogenic IL7R gain-of-function mutations in childhood Tcell acute lymphoblastic leukemia. Nat Genet. 2011;43:932–9.
- 4. Mazzucchelli RI, Riva A, Durum SK. The human IL-7 receptor gene: deletions, polymorphisms and mutations. Semin Immunol. 2012;24:225–30.
- 5. Mazzucchelli R, Durum SK. Interleukin-7 receptor expression: intelligent design. Nat Rev Immunol. 2007;7:144–54.
- 6. Lee LF, Logronio K, Tu GH, Zhai W, Ni I, Mei L, et al. Anti-IL-7 receptor-alpha reverses established type 1 diabetes in nonobese diabetic mice by modulating effector T-cell function. Proc Natl Acad Sci USA. 2012;109:12674–9.
- 7. Penaranda C, Kuswanto W, Hofmann J, Kenefeck R, Narendran P, Walker LS, et al. IL-7 receptor blockade reverses autoimmune diabetes by promoting inhibition of effector/memory T cells. Proc Natl Acad Sci USA. 2012;109:12668–73.
- 8. Gonzalez-Quintial R, Lawson BR, Scatizzi JC, Craft J, Kono DH, Baccala R, et al. Systemic autoimmunity and lymphoproliferation are associated with excess IL-7 and inhibited by IL-7Ralpha blockade. PloS ONE. 2011;6:e27528.
- 9. Hartgring SA, Willis CR, Alcorn D, Nelson LJ, Bijlsma JW, Lafeber FP, et al. Blockade of the interleukin-7 receptor inhibits collagen-induced arthritis and is associated with reduction of T cell activity and proinflammatory mediators. Arthritis Rheum. 2010;62:2716–25.
- 10. Belarif L, Mary C, Jacquemont L, Mai HL, Danger R, Hervouet J, et al. IL-7 receptor blockade blunts antigen-specific memory T cell responses and chronic inflammation in primates. Nat Commun. 2018;9:4483.
- 11. Mai HL, Nguyen TVH, Branchereau J, Poirier N, Renaudin K, Mary C, et al. Interleukin-7 receptor blockade by an anti-CD127 monoclonal antibody in nonhuman primate kidney transplantation. Am J Transplant. 2020;20:101–11.
- 12. Ellis J, van Maurik A, Fortunato L, Gisbert S, Chen K, Schwartz A, et al. Anti-IL-7 receptor alpha monoclonal antibody (GSK2618960) in healthy subjects - a randomized, double-blind, placebo-controlled study. Br J Clin Pharm. 2019;85:304–15.
- 13. Herold KC, Bucktrout SL, Wang X, Bode BW, Gitelman SE, Gottlieb PA, et al. Immunomodulatory activity of humanized anti–IL-7R monoclonal antibody RN168 in subjects with type 1 diabetes. JCI Insight. 2019;4.
- 14. Calzascia T, Pellegrini M, Lin A, Garza KM, Elford AR, Shahinian A, et al. CD4 T cells, lymphopenia, and IL-7 in a multistep pathway to autoimmunity. Proc Natl Acad Sci USA. 2008;105:2999–3004.
- 15. Ponchel F, Cuthbert RJ, Goeb V. IL-7 and lymphopenia. Clin Chim Acta. 2011;412:7–16.
- 16. Giliani S, Mori L, de Saint Basile G, Le Deist F, Rodriguez-Perez C, Forino C, et al. Interleukin-7 receptor alpha (IL-7Ralpha) deficiency: cellular and molecular bases. Analysis of clinical, immunological, and molecular features in 16 novel patients. Immunol Rev. 2005;203:110–26.
- 17. Rajasuriar R, Booth DR, Gouillou M, Spelman T, James I, Solomon A, et al. The role of SNPs in the alpha-chain of the IL-7R gene in CD4+ T-cell recovery in HIV-infected African patients receiving suppressive cART. Genes Immun. 2012;13:83–93.
- 18. Hartling HJ, Thorner LW, Erikstrup C, Harritshoj LH, Kronborg G, Pedersen C, et al. Polymorphism in interleukin-7 receptor alpha gene is associated with faster CD4(+) T-cell recovery after initiation of combination antiretroviral therapy. AIDS. 2014; 28:1739–48.
- 19. Guzman-Fulgencio M, Berenguer J, Jimenez-Sousa MA, Micheloud D, Garcia-Alvarez M, Bellon JM, et al. IL7RA polymorphisms predict the CD4+ recovery in HIV patients on cART. Eur J Clin Investig. 2015;45:1192–9.
- 20. Resino S, Navarrete-Munoz MA, Blanco J, Pacheco YM, Castro I, Berenguer J, et al. IL7RA rs6897932 polymorphism is associated with better $CD4(+)$ T-cell recovery in HIV infected patients starting combination antiretroviral therapy. Biomolecules. 2019;9.
- 21. Shamim Z, Spellman S, Haagenson M, Wang T, Lee SJ, Ryder LP, et al. Polymorphism in the interleukin-7 receptor-alpha and outcome after allogeneic hematopoietic cell transplantation with matched unrelated donor. Scand J Immunol. 2013;78:214–20.
- 22. Kielsen K, Enevold C, Heilmann C, Sengelov H, Pedersen AE, Ryder LP, et al. Donor genotype in the interleukin-7 receptor alpha-chain predicts risk of graft-versus-host disease and cytomegalovirus infection after allogeneic hematopoietic stem cell transplantation. Front Immunol. 2018;9:109.
- 23. Kielsen K, Shamim Z, Thiant S, Faucher S, Decker W, Christensen IJ, et al. Soluble interleukin-7 receptor levels and risk of acute graft-versus-host disease after allogeneic haematopoietic stem cell transplantation. Clin Immunol. 2018;187:26–32.
- 24. Rochman Y, Spolski R, Leonard WJ. New insights into the regulation of T cells by gamma(c) family cytokines. Nat Rev Immunol. 2009;9:480–90.
- 25. Waickman AT, Park JY, Park JH. The common gamma-chain cytokine receptor: tricks-and-treats for T cells. Cell Mol Life Sci. 2016;73:253–69.
- 26. McElroy CA, Holland PJ, Zhao P, Lim JM, Wells L, Eisenstein E, et al. Structural reorganization of the interleukin-7 signaling complex. Proc Natl Acad Sci USA. 2012;109:2503–8.
- 27. Goodwin RG, Friend D, Ziegler SF, Jerzy R, Falk BA, Gimpel S, et al. Cloning of the human and murine interleukin-7 receptors: demonstration of a soluble form and homology to a new receptor superfamily. Cell. 1990;60:941–51.
- 28. Carini C, McLane MF, Mayer KH, Essex M. Dysregulation of interleukin-7 receptor may generate loss of cytotoxic T cell response in human immunodeficiency virus type 1 infection. Eur J Immunol. 1994;24:2927–34.
- 29. Vranjkovic A, Crawley AM, Gee K, Kumar A, Angel JB. IL-7 decreases IL-7 receptor alpha (CD127) expression and induces the shedding of CD127 by human CD8+ T cells. Int Immunol. 2007;19:1329–39.
- 30. Pleiman CM, Gimpel SD, Park LS, Harada H, Taniguchi T, Ziegler SF. Organization of the murine and human interleukin-7 receptor genes: two mRNAs generated by differential splicing and presence of a type I-interferon-inducible promoter. Mol Cell Biol. 1991;11:3052–9.
- 31. Rose T, Lambotte O, Pallier C, Delfraissy JF, Colle JH. Identification and biochemical characterization of human plasma soluble IL-7R: lower concentrations in HIV-1-infected patients. J Immunol. 2009;182:7389–97.
- 32. Rane L, Vudattu N, Bourcier K, Graniar E, Hillert J, Seyfert V, et al. Alternative splicing of interleukin-7 (IL-7) and interleukin-7 receptor alpha (IL-7Ralpha) in peripheral blood from patients with multiple sclerosis (MS). J Neuroimmunol. 2010;222:82–6.
- 33. Faucher S, Crawley AM, Decker W, Sherring A, Bogdanovic D, Ding T, et al. Development of a quantitative bead capture assay for soluble IL-7 receptor alpha in human plasma. PloS ONE. 2009;4:e6690.
- 34. Gregory SG, Schmidt S, Seth P, Oksenberg JR, Hart J, Prokop A, et al. Interleukin 7 receptor alpha chain (IL7R) shows allelic and functional association with multiple sclerosis. Nat Genet. 2007;39:1083–91.
- 35. Lundstrom W, Highfill S, Walsh ST, Beq S, Morse E, Kockum I, et al. Soluble IL7Ralpha potentiates IL-7 bioactivity and promotes autoimmunity. Proc Natl Acad Sci USA. 2013;110:E1761–70.
- 36. Monti P, Brigatti C, Krasmann M, Ziegler AG, Bonifacio E. Concentration and activity of the soluble form of the interleukin-7 receptor alpha in type 1 diabetes identifies an interplay between hyperglycemia and immune function. Diabetes. 2013;62:2500–8.
- 37. Kreft KL, Verbraak E, Wierenga-Wolf AF, van Meurs M, Oostra BA, Laman JD, et al. Decreased systemic IL-7 and soluble IL-7Ralpha in multiple sclerosis patients. Genes Immun. 2012;13:587–92.
- 38. O'Doherty C, Alloza I, Rooney M, Vandenbroeck K. IL7RA polymorphisms and chronic inflammatory arthropathies. Tissue Antigens. 2009;74:429–31.
- 39. Todd JA, Walker NM, Cooper JD, Smyth DJ, Downes K, Plagnol V, et al. Robust associations of four new chromosome regions from genome-wide analyses of type 1 diabetes. Nat Genet. 2007;39:857–64.
- 40. Heron M, Grutters JC, van Moorsel CH, Ruven HJ, Huizinga TW, van der Helm-van Mil AH, et al. Variation in IL7R predisposes to sarcoid inflammation. Genes Immun. 2009;10:647–53.
- 41. Hoffjan S, Beygo J, Akkad DA, Parwez Q, Petrasch-Parwez E, Epplen JT. Analysis of variation in the IL7RA and IL2RA genes in atopic dermatitis. J Dermatol Sci. 2009;55:138–40.
- 42. Santiago JL, Alizadeh BZ, Martinez A, Espino L, de la Calle H, Fernandez-Arquero M, et al. Study of the association between the CAPSL-IL7R locus and type 1 diabetes. Diabetologia. 2008;51:1653–8.
- 43. Kiselev I, Bashinskaya V, Baulina N, Kozin M, Popova E, Boyko A, et al. Genetic differences between primary progressive and relapsing-remitting multiple sclerosis: The impact of immunerelated genes variability. Mult Scler Relat Disord. 2019;29:130–6.
- 44. Wu S, Liu Q, Zhu JM, Wang MR, Li J, Sun MG. Association between the IL7R T244I polymorphism and multiple sclerosis risk: a meta analysis. Neurol Sci. 2016;37:1467–74.
- 45. Wang XS, Wen PF, Zhang M, Hu LF, Ni J, Qiu LJ, et al. Interleukin-7 receptor single nucleotide polymorphism rs6897932 (C/T) and the susceptibility to systemic lupus erythematosus. Inflammation. 2014;37:615–20.
- 46. Zhang R, Duan L, Jiang Y, Zhang X, Sun P, Li J, et al. Association between the IL7R T244I polymorphism and multiple sclerosis: a meta-analysis. Mol Biol Rep. 2011;38:5079–84.
- 47. Galarza-Munoz G, Briggs FB, Evsyukova I, Schott-Lerner G, Kennedy EM, Nyanhete T, et al. Human epistatic interaction controls IL7R splicing and increases multiple sclerosis risk. Cell. 2017;169:72–84 e13.
- 48. Jager J, Schulze C, Rosner S, Martin R. IL7RA haplotypeassociated alterations in cellular immune function and gene expression patterns in multiple sclerosis. Genes Immun. 2013;14:453–61.
- 49. Crawley AM, Faucher S, Angel JB. Soluble IL-7R alpha (sCD127) inhibits IL-7 activity and is increased in HIV infection. J Immunol. 2010;184:4679–87.
- 50. Dooms H. Interleukin-7: fuel for the autoimmune attack. J Autoimmun. 2013;45:40–8.
- 51. Heninger AK, Theil A, Wilhelm C, Petzold C, Huebel N, Kretschmer K, et al. IL-7 abrogates suppressive activity of human CD4+CD25+FOXP3+ regulatory T cells and allows expansion of alloreactive and autoreactive T cells. J Immunol. 2012;189:5649–58.
- 52. Lundtoft C, Awuah AA, Guler A, Harling K, Schaal H, Mayatepek E, et al. An IL7RA exon 5 polymorphism is associated with impaired IL-7Ralpha splicing and protection against tuberculosis in Ghana. Genes Immun. 2019;20:514–9.
- 53. Seyfarth J, Lundtoft C, Fortsch K, Ahlert H, Rosenbauer J, Baechle C, et al. Interleukin-7 receptor alpha-chain haplotypes differentially affect soluble IL-7 receptor and IL-7 serum concentrations in children with type 1 diabetes. Pediatr Diabetes. 2018;19:955–62.
- 54. Hoe E, McKay F, Schibeci S, Heard R, Stewart G, Booth D. Interleukin 7 receptor alpha chain haplotypes vary in their influence on multiple sclerosis susceptibility and response to interferon Beta. J Interferon Cytokine Res. 2010;30:291–8.
- 55. Lundtoft C, Seyfarth J, Oberstrass S, Rosenbauer J, Baechle C, Roden M, et al. Autoimmunity risk- and protection-associated IL7RA genetic variants differentially affect soluble and membrane IL-7Ralpha expression. J Autoimmun. 2019;97:40–7.
- 56. Henriques CM, Rino J, Nibbs RJ, Graham GJ, Barata JT. IL-7 induces rapid clathrin-mediated internalization and JAK3 dependent degradation of IL-7Ralpha in T cells. Blood. 2010;115:3269–77.
- 57. Faller EM, Ghazawi FM, Cavar M, MacPherson PA. IL-7 induces clathrin-mediated endocytosis of CD127 and subsequent degradation by the proteasome in primary human CD8 T cells. Immunol Cell Biol. 2016;94:196–207.
- 58. Lundtoft C, Afum-Adjei Awuah A, Rimpler J, Harling K, Nausch N, Kohns M, et al. Aberrant plasma IL-7 and soluble IL-7 receptor levels indicate impaired T-cell response to IL-7 in human tuberculosis. PLoS Pathog. 2017;13:e1006425.
- 59. Weber F, Fontaine B, Cournu-Rebeix I, Kroner A, Knop M, Lutz S, et al. IL2RA and IL7RA genes confer susceptibility for multiple sclerosis in two independent European populations. Genes Immun. 2008;9:259–63.
- 60. Zuvich RL, McCauley JL, Oksenberg JR, Sawcer SJ, De Jager PL, International Multiple Sclerosis Genetics C, et al. Genetic variation in the IL7RA/IL7 pathway increases multiple sclerosis susceptibility. Hum Genet. 2010;127:525–35.
- 61. Lundmark F, Duvefelt K, Iacobaeus E, Kockum I, Wallstrom E, Khademi M, et al. Variation in interleukin 7 receptor alpha chain (IL7R) influences risk of multiple sclerosis. Nat Genet. 2007;39:1108–13.
- 62. Taheri M, Sayad A. Investigating the exon 6 sequence changes of interleukin 7 receptor A (IL7RA) gene in patients with relapsingremitting multiple sclerosis. Hum Antib. 2018;26:43–8.
- 63. Akkad DA, Hoffjan S, Petrasch-Parwez E, Beygo J, Gold R, Epplen JT. Variation in the IL7RA and IL2RA genes in German multiple sclerosis patients. J Autoimmun. 2009;32:110–5.
- 64. Alcina A, Fedetz M, Ndagire D, Fernandez O, Leyva L, Guerrero M, et al. The T244I variant of the interleukin-7 receptor-alpha gene and multiple sclerosis. Tissue Antigens. 2008;72:158–61.
- 65. International Multiple Sclerosis Genetics C. Refining genetic associations in multiple sclerosis. Lancet Neurol. 2008;7:567–9.
- 66. Svejgaard A. The immunogenetics of multiple sclerosis. Immunogenetics. 2008;60:275–86.
- 67. Barrett JC, Clayton DG, Concannon P, Akolkar B, Cooper JD, Erlich HA, et al. Genome-wide association study and metaanalysis find that over 40 loci affect risk of type 1 diabetes. Nat Genet. 2009;41:703–7.
- 68. Smyth DJ, Plagnol V, Walker NM, Cooper JD, Downes K, Yang JH, et al. Shared and distinct genetic variants in type 1 diabetes and celiac disease. N. Engl J Med. 2008;359:2767–77.
- 69. Broux B, Hellings N, Venken K, Rummens JL, Hensen K, Van Wijmeersch B, et al. Haplotype 4 of the multiple sclerosisassociated interleukin-7 receptor alpha gene influences the frequency of recent thymic emigrants. Genes Immun. 2010;11:326–33.
- 70. Cortes A, Hadler J, Pointon JP, Robinson PC, Karaderi T., International Genetics of Ankylosing Spondylitis C et al. Identification of multiple risk variants for ankylosing spondylitis through high-density genotyping of immune-related loci. Nat Genet. 2013;45:730–8.
- 71. Deshpande P, Cavanagh MM, Le Saux S, Singh K, Weyand CM, Goronzy JJ. IL-7- and IL-15-mediated TCR sensitization enables T cell responses to self-antigens. J Immunol. 2013;190:1416–23.
- 72. Gupta S, Cerosaletti K, Long SA. Renegade homeostatic cytokine responses in T1D: drivers of regulatory/effector T cell imbalance. Clin Immunol. 2014;151:146–54.
- 73. Sinha ML, Fry TJ, Fowler DH, Miller G, Mackall CL. Interleukin 7 worsens graft-versus-host disease. Blood. 2002;100:2642–9.
- 74. Kim YG, Ihm CG, Lee TW, Lee SH, Jeong KH, Moon JY, et al. Association of genetic polymorphisms of interleukins with newonset diabetes after transplantation in renal transplantation. Transplantation. 2012;93:900–7.
- 75. Napolitano LA, Grant RM, Deeks SG, Schmidt D, De Rosa SC, Herzenberg LA, et al. Increased production of IL-7 accompanies HIV-1-mediated T-cell depletion: implications for T-cell homeostasis. Nat Med. 2001;7:73–9.
- 76. Sereti I, Dunham RM, Spritzler J, Aga E, Proschan MA, Medvik K, et al. IL-7 administration drives T cell-cycle entry and expansion in HIV-1 infection. Blood. 2009;113:6304–14.
- 77. Thiebaut R, Jarne A, Routy JP, Sereti I, Fischl M, Ive P, et al. Repeated cycles of recombinant human interleukin-7 in HIVinfected patients with low CD4 T cell reconstitution on antiretroviral therapy: results of two phase II multicentre studies. Clin Infect Dis. 2016;62:1178–85.
- 78. Levy Y, Lacabaratz C, Weiss L, Viard JP, Goujard C, Lelievre JD, et al. Enhanced T cell recovery in HIV-1-infected adults through IL-7 treatment. J Clin Investig. 2009;119:997–1007.
- 79. Ferrari G, King K, Rathbun K, Place CA, Packard MV, Bartlett JA, et al. IL-7 enhancement of antigen-driven activation/expansion of HIV-1-specific cytotoxic T lymphocyte precursors (CTLp). Clin Exp Immunol. 1995;101:239–48.
- 80. Kiazyk SA, Fowke KR. Loss of CD127 expression links immune activation and CD4(+) T cell loss in HIV infection. Trends Microbiol. 2008;16:567–73.
- 81. Nguyen TP, Shukla S, Asaad R, Freeman ML, Lederman MM, Harding CV, et al. Responsiveness to IL-7 but not to IFN-alpha is diminished in CD4+ T cells from treated HIV infected patients who experience poor CD4+ T-cell recovery. AIDS. 2016;30:2033–42.
- 82. Rethi B, Fluur C, Atlas A, Krzyzowska M, Mowafi F, Grutzmeier S, et al. Loss of IL-7Ralpha is associated with CD4 T-cell depletion, high interleukin-7 levels and CD28 down-regulation in HIV infected patients. AIDS. 2005;19:2077–86.
- 83. Sasson SC, Zaunders JJ, Zanetti G, King EM, Merlin KM, Smith DE, et al. Increased plasma interleukin-7 level correlates with

decreased CD127 and Increased CD132 extracellular expression on T cell subsets in patients with HIV-1 infection. J Infect Dis. 2006;193:505–14.

- 84. Shive CL, Mudd JC, Funderburg NT, Sieg SF, Kyi B, Bazdar DA, et al. Inflammatory cytokines drive CD4+ T-cell cycling and impaired responsiveness to interleukin 7: implications for immune failure in HIV disease. J Infect Dis. 2014;210:619–29.
- 85. Tanaskovic S, Fernandez S, Price P, French MA. Interleukin-7 signalling defects in naive CD4+ T cells of HIV patients with CD4+ T-cell deficiency on antiretroviral therapy are associated with T-cell activation and senescence. AIDS. 2014;28:821–30.
- 86. Young CD, Angel JB. HIV infection of thymocytes inhibits IL-7 activity without altering CD127 expression. Retrovirology. 2011;8:72.
- 87. Medrano LM, Jimenez JL, Jimenez-Sousa MA, Fernandez-Rodiguez A, Gutierrez-Rivas M, Bellon JM, et al. IL7RA polymorphisms are not associated with AIDS progression. Eur J Clin Investig. 2017;47:179–27.
- 88. Hartling HJ, Thorner LW, Erikstrup C, Zinyama R, Kallestrup P, Gomo E, et al. Polymorphisms in the interleukin-7 receptor alpha gene and mortality in untreated HIV-infected individuals. AIDS. 2013;27:1615–20.
- 89. Seyfarth J, Sivagurunathan S, Ricken S, Weinreich G, Olbrich L, Taube C, et al. Higher interleukin-7 serum concentrations in patients with cystic fibrosis correlate with impaired lung function. J Cyst Fibros. 2019;18:71–7.
- 90. Mitchell PD, O'Byrne PM. Biologics and the lung: TSLP and other epithelial cell-derived cytokines in asthma. Pharm Ther. 2017;169:104–12.
- 91. Reche PA, Soumelis V, Gorman DM, Clifford T, Liu M, Travis M, et al. Human thymic stromal lymphopoietin preferentially stimulates myeloid cells. J Immunol. 2001;167:336–43.
- 92. Borriello F, Iannone R, Di Somma S, Vastolo V, Petrosino G, Visconte F, et al. Lipopolysaccharide-elicited TSLPR expression enriches a functionally discrete subset of human CD14(+) CD1c (+) monocytes. J Immunol. 2017;198:3426–35.
- 93. Al-Mossawi H, Yager N, Taylor CA, Lau E, Danielli S, de Wit J, et al. Context-specific regulation of surface and soluble IL7R expression by an autoimmune risk allele. Nat Commun. 2019; 10:4575.