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SYSTEMATIC REVIEW Immune cell composition in unipolar depression: a comprehensive systematic review and meta-analysis

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Depression has been associated with inflammatory pathophysiological mechanisms, including alterations in amount of circulating immune cells. However, no meta-analysis within the past 20 years have reevaluated the circulating immune cells in blood and cerebrospinal fluid (CSF) from patients with depression compared to healthy controls. The aim of this study was to systematically evaluate the circulating immune cells in blood and CSF from patients with unipolar depression compared to healthy controls. Databases were searched up until February 12, 2021. Data-extraction was performed by two independent reviewers. 104 studies were included in the meta-analysis using fixed and random-effects models. Patients with depression had a significantly higher overall leukocyte count (35 studies; SMD, 0.46; 95% CI: 0.31–0.60, $l^2 = 68\%$), higher neutrophil count (24 studies; SMD, 0.52; 95% CI: 0.33–0.71, $l^2 = 77\%$) and higher monocyte count (27 studies; SMD, 0.32; 95% CI: 0.11–0.53, $l^2 = 77\%$) compared to healthy controls. Leukocyte counts were higher in inpatients, indicating a relation to depression severity. Furthermore, there were significant alterations in several lymphocyte subsets, including higher natural killer cells and T cell subsets. Higher neutrophil/lymphocyte ratio (11 studies; SMD = 0.24; 95% CI: 0.06–0.42, $l^2 = 73\%$), CD4/CD8 cell-ratio (26 studies; SMD = 0.14; 95% CI: 0.01–0.28, $l^2 = 42\%$) and T helper 17/T regulatory ratio (2 studies; SMD = 1.05; 95% CI: 0.15–1.95, $l^2 = 86\%$) were found in patients compared to healthy controls. CSF white cell count was higher in patients compared to controls (3 studies; SMD = 0.20; 95% CI: 0.01–0.38, $l^2 = 0\%$). There were no data for CSF cell subsets. This study suggests that there are several blood immune cell alterations in patients with unipolar depression compared to healthy controls, both in major leukocyte subsets and more specialized immune cell subsets.

Molecular Psychiatry (2023) 28:391-401; https://doi.org/10.1038/s41380-022-01905-z

INTRODUCTION

Depression is estimated to affect more than 260 million people on a global scale [1]. Despite the immense impact of depression, our understanding of the etiology and pathophysiology remains inadequate - emphasized by the fact that approximately a third of patients with depression do not respond adequately to antidepressant treatment [2–4]. The past decades, immunological alterations have increasingly been associated with depression. Epidemiological studies have demonstrated that immune exposures, such as infections and autoimmune disorders, are associated with an increased risk of subsequent depression [5]. Moreover, studies have found that therapeutic treatment with pro-inflammatory medicine can induce depressive symptomatology [6], and anti-inflammatory treatment has shown to reduce depressive symptoms [7].

Several meta-analyses have shown alterations of inflammatory biomarkers in the blood and cerebrospinal fluid (CSF) from patients with depression [8–10]; however, no meta-analysis within the past 20 years have been conducted regarding subsets of circulating immune cells in blood or CSF from patients with unipolar depression. The most recent meta-analysis from 2018

focused exclusively on ratios between different immune cells in blood from patients with mood disorders compared to healthy controls and revealed an increased neutrophil-lymphocyte ratio among patients with major depressive disorder [11]. Two metaanalyses, published in 1993 and 2001, reported leukocytosis in blood from patients with depression compared to healthy controls [12, 13]. In 1993, a significant increase in the absolute neutrophil count and decreased absolute lymphocyte count were reported [12], whereas the meta-analysis from 2001 only reported significant changes in the relative proportions of these cells [13]. In regard to other immune cell subsets the results are also conflicting [11, 14]. Since the latest meta-analysis of individual immune cell subsets from 2001, numerous studies on the relationship between depression and circulating immune cell subsets have been published - both regarding major leukocyte subsets [14-18] and lymphocyte subsets [19-23]. However, no systematic review and meta-analysis has recently reevaluated the amount of individual circulating immune cell subsets in patients diagnosed with depression. In addition, no meta-analysis has previously been conducted regarding circulating immune cells in CSF from patients with depression.

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Received: 31 May 2022 Revised: 21 November 2022 Accepted: 28 November 2022 Published online: 14 December 2022

The objective of this systematic review and meta-analysis was to comprehensively investigate differences in the composition of circulating immune cells in blood and/or CSF from patients with depression compared to healthy controls. Furthermore, our aim was to explore whether there was an association between the amounts of immune cells and clinical features of the depression (e.g., severity of depression and antidepressant medication status).

MATERIALS AND METHODS

The study protocol was registered on PROSPERO (ID: CRD42021237884) and is available online. Meta-analysis of Observational Studies in Epidemiology (MOOSE) and The Preferred Reporting Items for Systematic Reviews and Meta-analyses (PRISMA) guidelines were followed (Supplementary Tables 1 and 2).

Study selection and search method

We included clinical studies which met the following criteria:

- 1. Investigation of circulating immune cells in blood and/or CSF obtained from living, human study participants.
- Inclusion of patients with a primary diagnosis of depression according to a diagnostic system (Research Diagnostic Criteria (RDC), International Classification of Diseases (ICD), Diagnostic and Statistical Manual of Mental Disorders (DSM), or similar classifications), of both sexes and all races, aged 18 or older.
- 3. Comparison to healthy controls.
- 4. Publications in peer-reviewed journals.
- 5. Publications in English.

We excluded studies on the following criteria:

- 1. Contained previously published data. If study samples overlapped across studies, the smaller study group was excluded.
- 2. Investigated depression in study participants with a diagnosis of psychotic or bipolar disorder (ICD-10: F20-F31.9).
- Investigated depression in participants with detected severe medical condition (including neurologic, vascular, autoimmune, malignant, endocrine, or infectious diseases with possible impact on immune cells which may disturb the interpretation of the immunological assessments).
- 4. Investigated pregnant or lactating women.
- 5. Had a control group admitted to hospital.
- Used other methods than direct measurements of immune cell composition.

We searched PubMed, EMBASE, Cochrane Library, Web of Science, Clinicaltrials.gov and PsycINFO from inception to the 12th of February 2021 using the following Medical Subject Headings (or similar headings) or text word terms:

(Depressive Disorder OR Depression OR Depressive) AND (Leukocyte* OR Leukocyte count OR White blood cell* OR WBC OR Complete blood count* OR CBC OR Peripheral blood mononuclear cell* OR PBMC OR Neutrophil* OR Basophil* OR Eosinophil* OR Monocyte* OR Lymphocyte* OR T-lymphocyte* OR T-cell* OR T helper OR T regulatory OR T cytotoxic OR Th1 OR Th2 OR Th17 OR Blymphocyte* OR B-cell* OR Natural Killer OR CD3 OR CD4 OR CD8 OR CD14 OR CD16 OR CD19 OR CD20 OR CD25 OR CD56) AND (Blood OR Plasma OR Serum OR Spinal puncture OR Spinal tap OR Lumbar puncture OR Cerebrospinal fluid OR CSF).

Animal studies, case studies, review articles and conference abstracts were excluded. The search string was decided in cooperation with an information specialist from the National Library of Denmark. Reference lists of relevant reviews were searched for additional studies. One investigator (BHF) examined titles and abstracts, and two investigators (BHF and SOW) examined the remaining full text reports to determine the study's fulfillment of the inclusion criteria. When the two investigators reached different decisions in the study selection, a third author (MEB) was consulted, and a consensus was reached.

Outcomes

The primary outcome was to investigate the amount (as counts or percentages) of circulating immune cells measured in blood and/ or CSF of patients with depression compared to healthy controls.

Secondary outcomes included: associations between immune cell count and clinical features of the depression (e.g., severity of the depression and antidepressant medication status).

Data extraction and bias assessment

One author (BHF) was responsible for data extraction, while SOW/ NVS validated, and quality checked all extracted data. Data extraction included bibliographic data and characteristics of the participants including antidepressant medication status. In case of missing details, authors were contacted by email. In case of no response, a reminder was sent. Risk of bias was assessed by two authors (BHF and SOW/NVS) according to the Newcastle-Ottawa Scale for case-control studies [24]. The risk of publication bias was assessed with funnel plots for cell counts, percentages and ratios investigated in 10 or more studies. Furthermore, subgroup analyses comparing a total score of ≥ 4 vs <4 on the Newcastle-Ottawa Scale and publication before vs after the year 2000 were conducted. In addition, meta-regressions associating size of studies and publication year with standardized mean differences (SMDs) were performed for leukocyte, neutrophil, monocyte, lymphocyte, natural killer cell, T cell, T helper cell, T cytotoxic cell and B cell counts.

Statistical analysis

Meta-analyses were performed using standardized mean differences (SMDs). Results from the random-effects and fixed-effects models are presented. In random-effects meta analyses the DerSimonian-Laird estimator [25] was used to estimate the between study variance, τ , [26] with the inverse-variance weighting method. Bias-correction of SMD estimates were done by using Hedges' g [27, 28]. χ^2 tests for heterogeneity were performed and l^2 calculated to assess heterogeneity [29]. Possible publication bias was assessed by Egger's test [30]. Medians, quartiles, and ranges were used for imputation of means and standard deviations, when necessary. To associate effect sizes with the average number of participants (harmonic mean), metaregression was used. To investigate associations between depression severity and immune cell count, percentages, and ratios, subgroup analyses of inpatient versus outpatient status were performed, when information hereof was available in ten or more studies. Furthermore, meta-regression analyses were performed to investigate associations between Hamilton Depression Rating Scale (HAMD) score and SMDs for leukocyte, neutrophil, monocyte, lymphocyte, natural killer cell, T cell, T helper cell, T cytotoxic cell and B cell counts. All statistical analyses were conducted in R version 4.02 or later, using the meta-package [31]. All *p*-values were two-sided and *p*-values < 0.05 were considered significant.

Deviations from the protocol

Measurements on the function of immune cells were not included due to a too large variability in methods for assessing the amount of circulating immune cells in blood. Correlations between the immune cell count and prognosis, and associations between immune cell measures in CSF and in blood were not assessed due to lack of sufficient data.

RESULTS

A total of 20.162 studies were identified. After removal of duplicates [32] and title/abstract screening, 659 studies remained for full text screening. A total of 104 studies had data required for a quantitative meta-analysis (Supplementary Fig. 1). Detailed study characteristics are presented in Supplementary Table 3 and a detailed overview of cell surface markers used for categorization of included blood immune cells is found in Fig. 1 and Supplementary Tables 4–8.

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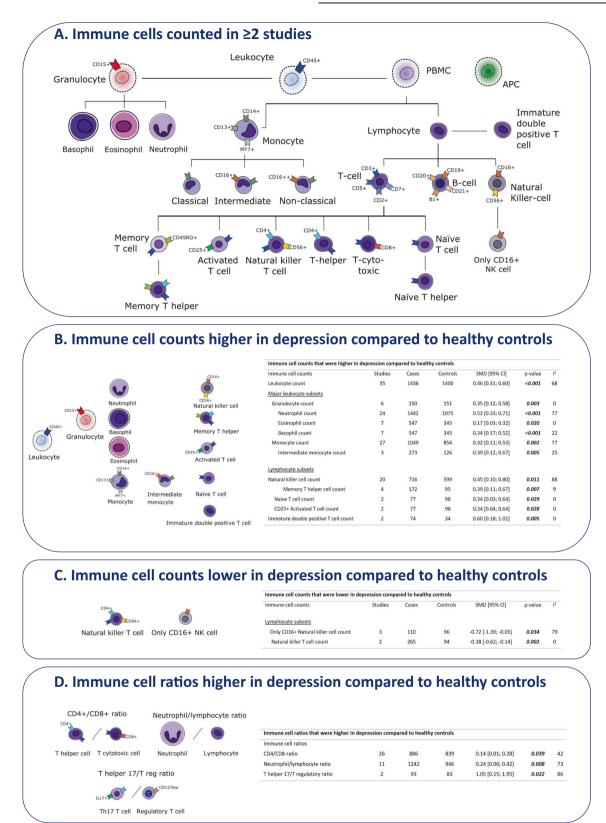


Fig. 1 Graphical abstract of main findings. A An overview of immune cells counted in ≥ 2 studies included in the meta-analysis. B Immune cells counted in ≥ 2 studies and higher in depression compared to healthy controls. C Immune cells counted in ≥ 2 studies and lower in depression compared to healthy controls. D Immune cell ratios altered in depression compared to healthy controls.

	Patients	s with de	pression		Healthy	controls		Standa	rdised I	Mean			
Study	Total	Mean	SD	Total	Mean	SD		Di	fference)	SMD	95%-CI	Weight
Leukocyte count													
Cattaneo 2020	130	6.76	2.01	40	5.90	1.40				_	0.45	[0.10; 0.81]	3.6%
Grudet 2020	48	5.50	1.40	54	5.60	1.50		-			-0.07	[-0.46; 0.32]	3.5%
Tunc 2019	32	7109.38	2522.72	34	7059.71	1962.31		-	<u> </u>		0.02	[-0.46; 0.50]	3.1%
dePunder 2018	40	7.27	2.42	40	6.49	1.90				_	0.35	[-0.09; 0.79]	3.3%
Cai 2017	103	6.11	1.12	106	5.67	0.98				-	0.42	[0.14; 0.69]	4.0%
Euteneuer 2017	98	6780.00	1889.00	30	6740.00	2209.00					0.02	[-0.39; 0.43]	3.4%
Peng 2016	167	6.10	1.46	180	6.00	1.51			- 		0.07	[-0.14; 0.28]	4.2%
Rybka 2016	15	6.90	1.05	19	6.40	2.75		-	1	_	0.22	[-0.45; 0.90]	2.4%
Demir 2015	41	7.80	1.70	47	6.90	1.30				-	0.59	[0.17; 1.02]	3.3%
Garcia-Rizo 2013	15	6.40	1.30	70	7.10	1.80					-0.40	[-0.96; 0.16]	2.8%
Moreno 2013	30	6.60	2.40	30	6.40	1.10					0.11	[-0.40; 0.61]	3.0%
Miller 2005	36	6.17	1.26	36	6.22	1.68		-	- <u>1</u>		-0.03	[-0.50; 0.43]	3.2%
Schlatter 2004b		6791.30		15	6933.30	1659.00				-	-0.05	[-0.85; 0.75]	2.0%
Atanackovic 2004	10	7000.00	1617.00	11	5836.00	1487.00			+	1	0.72	[-0.17; 1.61]	1.7%
Bauer 2002	36		18.26	31	56.71	18.54				4	0.84	[0.33; 1.34]	3.0%
Jung 1999	107	7.34	2.19	138	5.72	1.20					0.94	[0.68; 1.21]	4.0%
Miller 1999		6570.00			6160.00					-	0.24	[-0.25; 0.74]	3.0%
Natelson 1999	14	7222.79	2811.55	73	6276.34	1588.20					0.51	[-0.06; 1.09]	2.7%
Vandoolaeghe 1999	47	7.52	2.17	32	6.43	2.03					0.51	[0.05; 0.97]	3.2%
Maes 1997	32		1.50	14	7.10	1.80						[-0.20; 1.07]	2.5%
Schleifer 1996/1999	21	9.18	2.73	21	7.64						0.65	[0.03; 1.27]	2.6%
Seidel 1996	33	6.90	1.78	44	5.12							[0.71; 1.70]	3.1%
Sluzewska 1996	46		0.30	20	6.00	0.30					→ 1.98	[1.35; 2.61]	2.5%
Reynaert 1995		6670.60			5777.80							[-0.12; 1.04]	2.7%
Maes 1994b	29		2.10	13	5.17	0.97				+		[0.06; 1.41]	2.4%
Maes 1994a	59		2.13	17	4.96	1.24			-	1	0.97	[0.41; 1.53]	2.8%
Anesi 1994		7120.00			6985.00			-	1			[-0.57; 0.67]	2.6%
McAdams 1993	20		6.26	30	5.70	3.83			-			[-0.12; 1.03]	2.7%
Marazziti 1992	10		2.39	12		1.03			-	_		[-0.89; 0.79]	1.9%
Caldwell 1991	10		2.90	10	4.90	0.80				1		[0.32; 2.29]	1.5%
Darko 1989		7865.00			6050.00					1		[0.16; 1.45]	2.5%
Kronfol 1989		8565.00			6787.00					+		[0.24; 1.16]	3.2%
Targum 1989	21	6.92	2.47	77	6.77	1.84						[-0.41; 0.56]	3.1%
Darko 1988b		7422.00			6322.00					_		[-0.04; 1.30]	2.4%
Kronfol 1984		8161.00	3412.00		5779.00	1978.00				1		[-0.01; 1.64]	1.9%
Random effects mod				1400							0.46	[0.31; 0.60]	100.0%
Heterogeneity: $I^2 = 68\%$	$t_{0}, \tau^{2} = 0.12$	258, p < 0.	01										
							I	I	I	I	I		
						-	2	-1	0	1	2		

Fig. 2 Blood leukocyte counts in patients with unipolar depression compared to healthy controls. With fixed and random effects models.

Forest plots of significant results on immune cells counted in ≥ 2 studies are found in Figs. 2–5 and forest plots of immune cell ratios are found in Fig. 6. An overview of results for immune cell counts is provided in Supplementary Table 9 and for percentages in Supplementary Table 10. Forest plots presenting fixed effects models of significant results by random effects models are found in Supplementary Figs. 2–6. Forest plots of immune cell counts, percentages and ratios including cerebrospinal fluid counts are found in Supplementary Figs. 7–45 Results from the sensitivity analyses related to antidepressant treatment and depression severity are provided in Supplementary Figs. 46–76 and Supplementary Figs. 100–165. Leave-one-out analyses are presented in Supplementary Figs. 166–174.

Leukocyte count and major leukocyte subsets in blood

Leukocyte count. The leukocyte count was significantly higher among patients compared to healthy controls (35 studies [1436 patients]; SMD = 0.46; 95% CI: 0.31–0.60, $l^2 = 68\%$) (Fig. 2 and Supplementary Figs. 2 and 7).

Granulocytes. The granulocyte count (6 studies [150 patients]; SMD = 0.35; 95% CI: 0.12–0.58, $l^2 = 0\%$) were significantly higher in patients compared to healthy controls. Patients with depression also had a higher neutrophil count (24 studies [1442 patients]; SMD = 0.52; 95% CI: 0.33–0.71, $l^2 = 77\%$) and a higher neutrophil percentage (15 studies [568 patients]; SMD = 0.44; 95% CI: 0.27–0.60, $l^2 = 36\%$) compared to healthy controls. Basophil count

(7 studies [547 patients]; SMD = 0.34; 95% CI: 0.17–0.52, $l^2 = 22\%$) and eosinophil count (7 studies [547 patients]; SMD = 0.17; 95% CI: 0.03–0.32, $l^2 = 0\%$) were higher in patients as well. However, the overall granulocyte, eosinophil and basophil percentages did not differ between groups. Results are shown in Fig. 3 and Supplementary Figs. 3 and 8–12.

Monocytes. Overall monocyte count was significantly higher in patients with depression compared to healthy controls (27 studies [1049 patients]; SMD = 0.32; 95% Cl: 0.11–0.53, $l^2 = 77\%$) (Fig. 4), however, with no significant difference in the monocyte percentages (17 studies [1559 patients]; SMD = -0.10; 95% Cl: -0.29 to 0.08, $l^2 = 74\%$) (Supplementary Fig. 12). In the analysis of monocytes subsets, the intermediate monocyte count was higher in patients (3 studies [273 patients]; SMD = 0.39; 95% Cl: 0.12–0.67, $l^2 = 25\%$) (Fig. 4), whereas there were no significant group differences regarding other monocyte subset amounts (Supplementary Figs. 4 and 13–16).

Lymphocytes. The lymphocyte count (44 studies [1857 patients]; SMD = 0.03; 95% CI: -0.11 to 0.16, $l^2 = 71\%$) did not differ significantly between groups, whereas the percentage of lymphocytes was lower among patients with depression (22 studies [821 patients]; SMD = -0.40; 95% CI: -0.56 to -0.23, $l^2 = 60\%$) (Supplementary Fig. 17).

Lymphocyte subsets in blood

Natural killer cells. The natural killer cell count was significantly higher in patients compared to healthy controls (20 studies [716

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Study	Total	with dep Mean		Total	Mean	controls SD	Standardised Mean Difference	SMD	95%-CI	Weight
•							1			J
Granulocyte count		0.00			0.00	0.00	_			00 70/
dePunder 2018	36	0.03	0.04	39	0.02		<u></u>		[-0.15; 0.76]	20.7%
Patas 2018	20	3.95	1.43	20	3.94				[-0.62; 0.62]	15.7%
Miller 1999		4200.00		31	3800.00				[-0.21; 0.79]	19.2%
Griffiths 1997	18	3.46	1.01	30	3.15	1.04			[-0.29; 0.89]	16.5%
Schleifer 1996/1999	21	5.75	1.92	21	4.47	1.16			[0.16; 1.42]	15.4%
Maes 1992a		4092.00	1327.77		3309.00	1239.61			[-0.17; 1.34]	12.5%
Random effects mod Heterogeneity: $I^2 = 0\%$,		- 0.61		151			\diamond	0.35	[0.12; 0.58] <i>^</i>	100.0%
Neutrophil count										
Cattaneo 2020	130	4.09	1.68	40	3.51	1.17		0.36	[0.01; 0.72]	5.0%
Grudet 2020		3366.02			3458.40				[-0.47; 0.32]	4.7%
Lynall 2020	206	3.92	1.51	77	3.42	1.13			[0.09; 0.62]	5.7%
Sahin 2020	57	6.22	2.28	59	3.77	0.98			[0.99; 1.80]	4.6%
Tunc 2019		4132.53				1574.98			[-0.61; 0.36]	4.1%
Öztürk 2019		4537.00				1472.00			[-0.27; 0.53]	4.7%
dePunder 2018	38	4.44	2.10	39	3.80				[-0.11; 0.79]	4.3%
Hasselmann 2018	35	3.32	1.54	35	3.63				[-0.66; 0.28]	4.2%
Cai 2017	103	3.85	0.77	106	3.27	0.82			[0.45; 1.01]	5.6%
Euteneuer 2017		4099.00			3520.00				[0.02; 0.84]	4.6%
Peng 2016	167	3.60	1.10	180	3.40	1.11	-		[-0.03; 0.39]	6.1%
Demir 2015	41	4.80	1.40	47	4.10	1.00			[0.15; 1.00]	4.5%
Garcia-Rizo 2013	15	3.90	0.80	70	4.10	1.80			[-0.68; 0.44]	3.6%
Miller 2005	36	3.68	1.08	36	3.70	1.56			[-0.48; 0.45]	4.2%
Bauer 2002	36	50.75	15.34	31	35.35	16.98			[0.44; 1.45]	3.9%
Vandoolaeghe 1999	47	4.39	1.81	32	3.39	1.36			[0.14; 1.06]	4.3%
Maes 1997	32	4.37	1.30	14	3.96	1.37			[-0.33; 0.94]	3.2%
Seidel 1996	33	4.59	1.50	44	3.19	1.40			[0.48; 1.44]	4.1%
Sluzewska 1996	46	3.90	0.40	20	3.20	0.20			[1.33; 2.59]	3.2%
Maes 1994b	29	3.82	1.71	13	2.74	0.69			[0.04; 1.39]	3.0%
Maes 1994a	59	4.88	1.73	17	3.13	1.03			[0.51; 1.65]	3.5%
Maes 1992b	79	4.00	1.72	22	2.82	0.70			[0.27; 1.23]	4.1%
Caldwell 1991	10	4.80	2.30	10	2.70	0.70		→ 1.18	[0.22; 2.15]	1.8%
Darko 1988b	18	4562.00	1495.00	18	3536.00	1176.00			[0.07; 1.42]	2.9%
Random effects mod	el 1442			1075			\diamond	0.52	[0.33; 0.71]	100.0%
Heterogeneity: $I^2 = 77\%$, τ ² = 0.15	76, p < 0.	01							
Basophil count										
Cattaneo 2020	130	0.03	0.02	40	0.02	0.02		0.61	[0.25; 0.97]	15.7%
Grudet 2020	46	21.80	26.48	53	21.81	28.37		-0.00	[-0.40; 0.39]	14.9%
Lynall 2020	206	0.03	0.01	77	0.02	0.01		0.39	[0.13; 0.66]	18.0%
dePunder 2018	38	0.06	0.03	39	0.04	0.02		0.42	[-0.03; 0.87]	13.6%
Garcia-Rizo 2013	15	0.07	0.12	70	0.02	0.12		0.40	[-0.16; 0.96]	11.3%
Seidel 1996	33	0.05	0.02	44	0.04	0.02		0.49	[0.04; 0.95]	13.4%
Maes 1992b	79	0.04	0.02	22	0.04	0.02		0.00	[-0.47; 0.47]	13.1%
Random effects mod				345			\diamond	0.34	[0.17; 0.52]	100.0%
Heterogeneity: $I^2 = 22\%$, τ ² = 0.01	17, <i>p</i> = 0.3	26							
Eosinophil count										
Cattaneo 2020	130	0.21	0.17	40	0.15		-		[-0.01; 0.70]	15.7%
Grudet 2020	46	156.28	90.42	53	136.68	107.94			[-0.20; 0.59]	14.8%
Lynall 2020	206	0.16	0.12	77	0.14		+		[-0.02; 0.50]	17.9%
dePunder 2018	38	0.15	0.10	39	0.15				[-0.50; 0.40]	13.6%
Garcia-Rizo 2013	15	0.26	0.20	70	0.23	0.20			[-0.41; 0.71]	11.3%
Seidel 1996	33	0.14	0.08	44	0.13				[-0.34; 0.57]	13.5%
Maes 1992b	79	0.15	0.08	22	0.15	0.09			[-0.51; 0.44]	13.1%
Random effects mod				345			♦	0.17	[0.03; 0.32]	100.0%
Heterogeneity: $I^2 = 0\%$,	τ ⁻ = 0, p =	= 0.81				r		-		
						1		' 2		
						-2	2 -1 0 1	2		

Fig. 3 Blood granulocyte counts in patients with unipolar depression compared to healthy controls. With fixed and random effects models.

patients]; SMD = 0.45; 95% CI: 0.10–0.80, $l^2 = 88\%$), whereas the percentages did not differ between groups. The "Only CD16 + natural killer cell count" (3 studies [110 patients]; SMD = -0.72; 95% CI: -1.39 to -0.05, $l^2 = 79\%$) and CD16low natural killer cells percentages (3 studies [109 patients]; SMD = -0.31; 95% CI: -0.58 to -0.05, $l^2 = 0\%$) was significantly lower in patients compared to healthy controls. Results are found in Fig. 5, Supplementary Figs. 5 and 18–21.

percentage (26 studies [1730 patients]; SMD = -0.06; 95% Cl: -0.19 to 0.07, $l^2 = 52\%$) (Supplementary Fig. 22). Overall T helper cell count tended to be higher in patients with depression (31 studies [1265 patients]; SMD = 0.12; 95% Cl: 0.00-0.24, $l^2 = 44\%$) compared to healthy controls, whereas there was no difference in T helper cell percentage (29 studies [997 patients]; SMD = -0.00; 95% Cl: -0.16 to 0.15, $l^2 = 61\%$) (Supplementary Fig. 23).

T cells and T helper cells. No significant differences were found regarding the overall T cell count (28 studies [938 patients]; SMD = -0.04; 95% Cl: -0.19 to 0.10, $l^2 = 52\%$) or overall T cell

T helper cell subsets. The memory T helper cell count (4 studies [172 patients]; SMD = 0.39; 95% CI: 0.11–0.67, $l^2 = 9\%$) (Fig. 5) as well as the percentage of memory T helper cells (5 studies [281

Pati		vith dep Mean			ealthy c Mean	ontrols SD	Standardised Mean Difference	SMD	95%-CI	Weight
Monocyte count							1			
Cattaneo 2020	130	0.41	0.14	40	0.40	0.14		0.10	[-0.26; 0.45]	4.5%
Grudet 2020		345.26			363.68	104.19	<u> </u>		[-0.56; 0.23]	4.3%
Sahin 2020	57	0.77	0.30	59	0.54	0.15			[0.58; 1.35]	4.3%
Hellmann-Regen 2019	47	0.50	0.14	47	0.48	0.14		0.13	[-0.27; 0.54]	4.3%
dePunder 2018	38	0.53	0.16	39	0.52	0.16			[-0.38; 0.52]	4.1%
Hasselmann 2018	35	0.49	0.17	35	0.48	0.15		0.09	[-0.37; 0.56]	4.0%
Krause 2018	39	457.00	169.00	51	458.00	209.00		-0.01	[-0.42; 0.41]	4.2%
Patas 2018	20	0.47	0.26	20	0.38	0.19		0.41	[-0.22; 1.03]	3.4%
Euteneuer 2017	98	403.00	134.00	30	310.00	153.00		0.67	[0.25; 1.08]	4.2%
Euteneuer 2014	38	463.24	221.22	47	465.37	197.30		-0.01	[-0.44; 0.42]	4.2%
Garcia-Rizo 2013	15	0.38	0.20	70	0.41	0.20		-0.15	[-0.71; 0.41]	3.7%
Lisi 2013	10	0.36	0.28	10	0.46	0.47		-0.24	[-1.13; 0.64]	2.5%
Miller 2005	36	0.37	0.12	36	0.39	0.14		-0.15	[-0.61; 0.31]	4.0%
Schlatter 2004b	10	470.30	172.10	15	405.00	75.20		0.52	[-0.30; 1.33]	2.7%
Bauer 2002	36	4.17	1.36	31	3.45	1.39		0.51	[0.03; 1.00]	3.9%
Rothermundt 2001b	43	484.00	192.00	43	452.00	125.00		0.20	[-0.23; 0.62]	4.2%
Lanquillon 2000	24	259.00	178.00	15	207.00	125.00		0.32	[-0.33; 0.97]	3.3%
Miller 1999	31	480.00	179.84	31	450.00			0.20	[-0.30; 0.70]	3.9%
Vandoolaeghe 1999	47		0.17	32		0.19			[-0.39; 0.50]	4.1%
Castilla-Cortázar 1998		549.00	38.00		382.00	49.00			[2.95; 4.72]	2.5%
Griffiths 1997	18		0.09	30	0.36	0.11			[-0.71; 0.46]	3.6%
Maes 1997	32		0.22	14	0.66	0.18		0.14	[-0.49; 0.77]	3.4%
Schleifer 1996/1999	21	0.42	0.23	21	0.43	0.18			[-0.65; 0.56]	3.5%
Sluzewska 1996	46		0.15	20	0.28	0.05			[0.23; 1.31]	3.7%
Maes 1994b	29		0.22	13	0.37	0.14			[-0.28; 1.04]	3.3%
Maes 1994a	59	0.45	0.19	17	0.24	0.12			[0.62; 1.76]	3.6%
Caldwell 1991	10	0.40	0.10	10	0.40	0.20			[-0.88; 0.88]	2.5%
Random effects model				854			\diamond	0.32	[0.11; 0.53]	100.0%
Heterogeneity: $I^2 = 77\%$, τ^2	2 = 0.22	257, p < 0	0.01							
Intermediate monocyte										
Lynall 2020	206		0.01	77	0.01	0.01		0.24	[-0.03; 0.50]	39.6%
Nowak 2019	32		12.48	14	6.13	2.59			[0.08; 1.38]	27.5%
Hasselmann 2018	35		0.02	35	0.02	0.01			[0.06; 1.02]	33.0%
Random effects model				126			\diamond	0.39	[0.12; 0.67]	100.0%
Heterogeneity: $I^2 = 25\%$, τ^2	² = 0.01	68, p = 0	0.26							
						1		I		
						-2	2 -1 0 1	2		

Fig. 4 Blood monocyte counts in patients with unipolar depression compared to healthy controls. Total monocyte count and significantly altered monocyte subsets measured in ≥ 2 studies. Fixed and random effects models.

patients]; SMD = 0.40; 95% CI: 0.22–0.58, $l^2 = 0\%$) (Supplementary Fig. 29) were significantly higher in patients compared to healthy controls. There were no significant differences in the percentages of Th1, Th2 or Th17 cells when comparing patients to healthy controls (Supplementary Figs. 24–26), neither differed the percentage of T regulatory cells (Supplementary Fig. 27). No significant differences were found concerning the amount of naïve T helper cells or the percentage of activated T helper cells nor effector memory T helper cell percentages (Supplementary Figs. 28–31).

T cytotoxic cells and *T* cytotoxic cell subsets. No significant differences were found between patients with depression and healthy controls regarding the T cytotoxic cell count (31 studies [1265 patients]; SMD = -0.05; 95% CI: -0.18 to 0.07, $l^2 = 47\%$) or percentages of T cytotoxic cells and subsets (Supplementary Figs. 32–36).

Activated, naïve and memory T cells. A higher amount of several T cell subsets was found in patients, including a higher naïve T cell count (2 studies [77 patients]; SMD = 0.34; 95% CI: 0.03–0.64, $l^2 = 0\%$) and a higher amount of activated T cells: Both CD25 + activated T cell count (2 studies [77 patients]; SMD = 0.34; 95% CI: 0.04–0.64, $l^2 = 0\%$) (Fig. 5) and percentage of HLA-DR + activated T cells (4 studies [128 patients]; SMD = 0.50; 95% CI: 0.19–0.81, $l^2 = 0\%$) were higher in patients (Fig. 5 and Supplementary Figs. 37 and 38). No significant differences were found regarding memory T cells when comparing patients and healthy controls.

Natural killer T cells and immature double positive T cells. The natural killer T cell count was lower in patients compared to healthy controls (2 studies [265 patients]; SMD = -0.38; 95% Cl: -0.62 to -0.14, $l^2 = 0\%$) (Fig. 5). No difference was found regarding the percentage of natural killer T cells (Supplementary Fig. 39). There was a higher immature double positive T cell count in patients with depression compared to healthy controls (2 studies [74 patients]; SMD = 0.60; 95% Cl: 0.18-1.01, $l^2 = 0\%$) (Fig. 5 and Supplementary Fig. 40).

B cells, unspecified antigen-presenting cells and activated lymphocytes

The amount of B cells, activated lymphocytes, and antigenpresenting cells were not significantly different when comparing patients with depression to healthy controls (Supplementary Figs. 41–43). However, interestingly the leave-one-out analysis of B cells revealed that exclusion of the study by Pavon et al. [33], altered the result to a significantly higher B cell count in patients compared to healthy controls (30 studies; SMD = 0.14; 95% CI: 0.01–0.26) (Supplementary Fig. 174).

Cell ratios

The CD4/CD8-ratio (T helper cell/T cytotoxic cell ratio) (26 studies [886 patients]; SMD = 0.14; 95% Cl: 0.01–0.28, $l^2 = 42\%$), neutro-phil/lymphocyte ratio (11 studies [1242 patients]; SMD = 0.24; 95% Cl: 0.06–0.42, $l^2 = 73\%$), and T helper 17/T regulatory ratio (2 studies [93 patients]; SMD = 1.05; 95% Cl: 0.15–1.95, $l^2 = 86\%$) were significantly higher in patients compared to healthy controls,

Church	Patients				-	ontrols	s		ised Mean			•		M/a:
Study	Total	Mean	50	Total	Mean	SD		Diffe	rence		SMD	9	5%-01	Weight
Natural killer cell c			~~~ ~~											
Krause 2018	39					129.00			-			[-0.20;	-	5.5%
Euteneuer 2017 Euteneuer 2014	98 38					128.00 119.26						[-0.57;		5.5% 5.4%
Hernandez 2010	30					30.00						[-0.91; [2.04;	-	4.3%
Hosseini 2007	27					105.40	-		-			[-1.23;		4.8%
Pavón 2006	33				173.10							[2.18;		4.6%
Schlatter 2004a	42	1.47	2.47	20	0.40	0.30						[-0.03;		5.1%
Atanackovic 2004	10	303.00	98.00	11	271.00	132.00					0.26	[-0.60;	1.12]	4.1%
Bauer 2002	36		1.25	31	2.22	1.28		_			0.19	[-0.29;	0.67]	5.3%
Rothermundt 2001a					360.80							[0.14;		5.4%
Miller 1999	31				147.40			_	-			[-0.27;	-	5.2%
Natelson 1999 Ravindran 1999	14 26		0.07	73 26	201.64 0.08	93.78 0.05			·			[-0.38; [0.32;		5.0% 5.0%
Ravindran 1998	69		0.07	44	0.08	0.05						[0.32;		5.6%
Griffiths 1997	18		0.07	30	0.08							[0.15;		4.9%
Schleifer 1996/1999			0.05	21	0.09	0.10			 			[-0.98;	-	4.9%
Ravindran 1996	18		0.05	23	0.08	0.04			_			[1.05;	-	4.5%
Reynaert 1995	34		82.20	18	134.60				+			[-0.85;		5.0%
Maes 1994b	29		0.10	13	0.28	0.22	-		t			[-1.22;	-	4.7%
Maes 1994a	59		141.03		194.00	131.94			<u> </u>			[-0.50;		5.1%
Random effects m				599					\diamond		0.45	[0.10;	0.80]	100.0%
Heterogeneity: $I^2 = 88$	3%, τ ² = 0.55	503, p < 0.0	01											
Only CD16+ Natura														
Castilla-Cortázar 19					285.00		< 1	_				[-2.16;	-	
Reynaert 1995	34				178.30				+			[-0.96;		
Andreoli 1993	53	0.25	0.16	53	0.31	0.16			1			[-0.76;	-	
Random effects me Heterogeneity: $I^2 = 79$		720. p < 0.0	01	96			V			-	0.72	[-1.39;	-0.05]	100.0%
Memory T helper c		50.07	45.54	00	40 70	40 70					~ 4 4	r o 40.	0.001	05 40/
Schlatter 2004a Seidel 1996	42 33		15.54 5.20	20 44	49.70 26.10	10.70 6.30						[-0.10;	-	25.4% 26.7%
Maes 1992d	73		0.23	21	0.45	0.30						[-0.38; [0.05;	-	
Maes 1992a		448.50				151.79								21.8%
Random effects m			221.02	95	200.00	101.10			\diamond				-	100.0%
Heterogeneity: $I^2 = 9$ %	$\%, \tau^2 = 0.007$	76, p = 0.3	5									•		
Naive T cell count														
Krause 2018	39	1114.00	504.00	51	914.00	466.00					0.41	[-0.01;	0.831	50.1%
Euteneuer 2014		1024.38			914.22			_						49.9%
Random effects me				98					\diamond				-	100.0%
Heterogeneity: $I^2 = 0$ %	$\%, \tau^2 = 0, p =$	= 0.62										• •	-	
CD25+ Activated T	cell count	ł												
Krause 2018	39	55.00	55.00	51	43.00	49.00		_			0.23	[-0.19;	0.65]	50.2%
Euteneuer 2014	38	50.18	50.86	47	31.18	31.95					0.45	[0.02;	0.89]	49.8%
Random effects me				98					\diamond		0.34	[0.04;	0.64]	100.0%
Heterogeneity: $I^2 = 0$ %	$\%, \tau^2 = 0, p =$	= 0.47												
Natural killer T cell	count													
Lynall 2020	206		0.04	77	0.06	0.05				-1	0.37	[-0.63;	-0.10]	53.4%
Maes 1994a	59		122.57		173.00	103.08			+				-	46.6%
Random effects me				94				\diamond		-	0.38	[-0.62;	-0.14]	100.0%
Heterogeneity: $I^2 = 0$ %	$\%, \tau^{-} = 0, p =$	= 0.80												
Immature double p				~~~	0.00	0.00			_			10.00	4 4 67	E4 50/
Schlatter 2004a	42		0.63	20	0.30									51.5%
Maes 1997 Random effects me	32 odel 74		0.01	14 34	0.01	0.00		-						48.5% 100.0%
Heterogeneity: $I^2 = 0$ %				34							0.00	[0.10;	1.01]	100.0%
insterogeneity. 7 = 0)	ο, τ = 0, <i>μ</i> -	5.01						1						
						-	2	-1 (0 1	2				

Fig. 5 Blood natural killer cell counts and T cell subset counts in patients with unipolar depression compared to healthy controls. Total natural killer cell count and significantly altered natural killer cell/T cell subset counts measured in \geq 2 studies. Fixed and random effects models.

whereas there was no difference in platelet/lymphocyte ratio (Fig. 6).

White cell count in cerebrospinal fluid

When including the CSF studies stating to have healthy controls, no significant difference in overall CSF white cell count (4 studies

[290 patients]; SMD = 0.11; 95% CI: -0.05 to 0.28, $l^2 = 41\%$) was found (Supplementary Fig. 44A). However, in one of the studies the healthy controls had values clearly exceeding the cut-off for neuroinflammation [34], and when excluding this study and thus only including the studies with a healthy control group, the CSF white cell count was higher among the patients

F	Patients w Tota	vith depre		Hea Total	althy co Mean	ntrols SD		ised Mean rence	SMD	95%-CI	Weight
-								1			
CD4/CD8-ratio	0	0 1 50	0 74	00	1 50	0.00			0.00	[0 07: 0 40]	c 00/
Ford 2020	9			98	1.56	0.83				[-0.37; 0.19]	6.2%
Krause 2018	3			51	2.01	0.70	_	*		[-0.30; 0.53]	5.0%
Basterzi 2010	6		0.40	36	1.15	0.40		<u> </u>		[-0.38; 0.43]	5.1%
Hernandez 2010	3			22	2.00	0.17				[-0.69; 0.41]	4.0%
Schlatter 2004a	4				1.80		-			[-0.14; 0.94]	4.0%
Atanackovic 2004	1			11	1.34					[-0.61; 1.11]	2.3%
Ravindran 1999	2			26	1.91	0.62	-			[-0.15; 0.94]	4.0%
Ravindran 1998	6		0.76	44	2.24		-+	+		[-0.59; 0.17]	5.3%
Maes 1997	3		1.36		1.78	0.79				[0.06; 1.35]	3.3%
Perini 1995	2				1.70		-			[-0.15; 1.06]	3.6%
Ravindran 1995	1	8 2.08	0.55	17	2.16	0.74			-0.12	[-0.78; 0.54]	3.2%
Anesi 1994	2	0 1.90	0.60	20	2.00	0.50	+	<u> </u>	-0.18	[-0.80; 0.44]	3.5%
Zisook 1994		6 2.70	2.30	21	2.00	1.30			0.44	[-0.48; 1.35]	2.1%
Andreoli 1993	5	3 1.84	0.69	53	1.67	0.65	-		0.25	[-0.13; 0.63]	5.3%
Maes 1992d	7	3 2.47	1.28	21	1.88	1.10			0.47	[-0.02; 0.96]	4.4%
Maes 1992a	2	4 2.03	1.06	10	1.52	0.60	-			[-0.23; 1.27]	2.8%
Bartoloni 1991		8 1.70			1.50	0.60				[-0.56; 1.13]	2.4%
Wilson 1990	1		0.45		1.98	0.80		+		[-1.35; 0.08]	2.9%
Kronfol 1989	4		0.84	37	1.63	0.48	_			[-0.22; 0.68]	4.7%
Schleifer 1989	. 9		1.63		1.65	1.73	_	<u> </u>		[-0.30; 0.28]	6.1%
Targum 1989	2		0.78		2.92			_		[-1.01; -0.03]	4.4%
Darko 1988a	2		0.81	20	1.73			.		[0.07; 1.35]	3.4%
Tondo 1988	2				1.51					[0.23; 1.34]	3.9%
Calabrese 1986	1			11	0.51	0.23				[-0.93; 0.79]	2.3%
	1				1.50	0.20					3.3%
Syvalathi 1985 Kronfol 1984	1				1.42					[0.01; 1.32]	2.6%
Random effects m			0.60	839	1.42	0.40		\diamond		[-1.13; 0.45] [0.01; 0.28]	
Heterogeneity: $I^2 = 42$			0.01	000				Ť	0.14	[0.01, 0.20]	100.070
N											
Neutrophil/lympho	•				0.04				o 07		0.00/
Grudet 2020	4		0.89		2.31	1.18	_			[-0.46; 0.33]	8.3%
Sahin 2020	5		1.10		1.90	0.62				[0.40; 1.16]	8.6%
Yu 2020	8		1.18		1.78	0.55				[-0.04; 0.53]	10.0%
Tunc 2019	3		1.05	34	2.00	0.80		T. C.		[-0.43; 0.54]	7.2%
Öztürk 2019	4		1.02		2.09	0.95	-	- x		[-0.20; 0.60]	8.3%
Arabska 2018	46		1.31	219	1.95			+		[-0.31; 0.01]	11.5%
Cai 2017	10		0.62		2.00	0.73				[0.14; 0.69]	10.1%
Ekinci 2017	10				1.81	0.33	_	<u> </u>		[-0.28; 0.40]	9.1%
Euteneuer 2017	9	8 2.00	0.70	30	1.60	0.60			0.59	[0.17; 1.00]	8.1%
Peng 2016	16	7 1.90	0.73	180	1.70	0.62			0.30	[0.08; 0.51]	10.9%
Demir 2015	4	1 2.30	0.90	47	2.00	0.60			0.39	[-0.03; 0.82]	7.9%
Random effects me				946				\diamond	0.24	[0.06; 0.42]	100.0%
Heterogeneity: $I^2 = 73$	$3\%, \tau^2 = 0.0$	0624, <i>p</i> < 0	0.01								
Platelet/lymphocyt	e ratio										
Sahin 2020		7 122.67	43.93	59	141.11	48.32		-	-0.40	[-0.76; -0.03]	22.0%
Yu 2020		2 134.28			113.12					[0.32; 0.90]	
Cai 2017		3 128.69			118.79					[0.04; 0.59]	
Peng 2016		7 125.30			124.40		-			[-0.19; 0.23]	
Random effects m			51.55	465	12-1.40	55.15	<			[-0.23; 0.23]	
Heterogeneity: $I^2 = 86$			0.01	403					0.15	[-0.22, 0.31]	100.070
Th17/T regulators	colle ratio										
Th17/T regulatory			10 04	E 0	0.60	0 00		_	0.64	10 22. 4 001	56 E0/
Ghosh 2020	5		10.84	53	0.69					[0.22; 1.00]	
Chen 2011	4		23.18		2.41	2.63				[0.99; 2.07]	
Random effects m	odel 9			83					- 1.05	[0.15; 1.95]	100.0%
Heterogeneity: $I^2 = 86$	6%, τ ² = 0.3	3651, p < 0	0.01				· · · · ·				
							· · ·		1 2		
							2 -1	0 1	2		

Fig. 6 Blood immune cell ratios in patients with unipolar depression compared to healthy controls. With fixed and random effects models.

with depression than the healthy controls (3 studies [224 patients]; SMD = 0.20; 95% CI: 0.01–0.38, $I^2 = 0\%$) (Supplementary Fig. 44B). No other immune cell subsets in the CSF were investigated.

Sensitivity analyses on antidepressants

Studies on clearly defined antidepressant-naïve patients were sparse. However, there appeared to be a subgroup difference related to the length of discontinuation of antidepressants for leukocyte count (p < 0.01) and a near significant difference for

lymphocyte count (p = 0.05) (Supplementary Figs. 46–58). Leukocyte counts were higher among patients ongoingly treated with antidepressants (6 studies [369 patients]; SMD = 0.53; 95% CI: 0.17–0.89, $l^2 = 74\%$) compared to antidepressant-naïve patients (1 study [15 patients]; SMD = -0.40; 95% CI: -0.96 to 0.16). The meta-regression analyses indicated the same association between duration of discontinuation of antidepressants for leukocytes (Supplementary Figs. 59–67). Meta-regression analyses on patients treated with antidepressants did not reveal any associations (Supplementary Figs. 68–76).

Sensitivity analyses on depression severity

The subgroup analysis of inpatient vs outpatient status revealed leukocyte counts to be higher in inpatients with depression, but not significant in outpatients or mixed groups with a significant subgroup difference (p = 0.04). Likewise, neutrophil counts were only higher in inpatients with significant subgroup differences (p < 0.01) (Supplementary Figs. 77–90). Meta-regression analyses revealed no significant associations to depression severity measured by HAM-D (Supplementary Figs. 91–99).

Bias assessment and related sensitivity analyses

The mean score according to the Newcastle-Ottawa Quality Assessment Scale across the 104 included studies was 3.89 points out of 8 possible points (Supplementary Tables 11 and 12). No studies received the maximum of eight stars. Most of the included studies had risk of bias regarding adequate case definition (82/ 104) and representativeness of cases (79/104). Moreover, approximately half of the studies had risk of bias concerning selection of controls (50/104), definition of controls (46/104) and comparability between cases and controls (38/104). Risk of bias was further assessed by subgroup analyses of Newcastle-Ottawa Scale score <4 versus ≥4, revealing significant subgroup differences in neutrophil percentage (p = 0.04) and T cell percentage (p = 0.03) (Supplementary Figs. 100–118). Subgroup analyses of studies published before year 2000 versus in year 2000 or later revealed several significant subgroup differences (Supplementary Figs. 119–137). Meta-regressions on size of studies and publication year showed significant associations between leukocyte and neutrophil count and publication year (p < 0.001), whereas there were no significant associations to size of studies (Supplementary Figs. 138-156). Funnel plots did not indicate publication bias (Supplementary Figs. 157-165). Leave-one-out-analyses did not alter results significantly despite as described for the overall B cell count (Supplementary Figs. 166-174).

DISCUSSION

This study is the largest and most comprehensive meta-analysis on the composition of circulating immune cells in unipolar depression to date. The meta-analysis is comprised of 104 studies in which 100 studies evaluated immune cells in blood, whereas only four studies evaluated immune cells in CSF. Our results indicate broad immune cell activation in unipolar depression. When evaluating absolute cell counts, a total of 12 subsets showed significant increases compared to healthy controls, whereas only CD16+ natural killer cells and natural killer T cell counts were lower in patients. Furthermore, several immune cell ratios were altered in patients with unipolar depression compared to healthy controls. These findings are consistent with previous published meta-analyses, showing leukocytosis, higher amounts of neutrophils, lower amounts of lymphocytes and subsequently an increased neutrophil/lymphocyte ratio [11-13]. However, our updated meta-analysis includes considerably more studies than the previous meta-analyses (e.g., 14 more studies of leukocyte count as compared to the most recent of the previous metaanalyses [13]). This enables us, as the first, to explore and identify differences in more specialized leukocyte subsets, including our findings of higher intermediate monocyte count and higher memory T helper cell count among the patients with depression. Furthermore, we also explore the CSF white cell count, evaluate the individual studies' risk of bias and the effect of antidepressant medication status on immune cell subsets.

Our results suggest a generalized increased immune activation in patients with unipolar depression, since the immune cell contrasts are not limited to one specific immune cell subset or lineage, but affect both the myeloid linage (e.g., monocytes and granulocytes) and the lymphoid linage (e.g., T cells and natural killer cells) indicating a generalized upregulation of the bone marrow activity. A recent study has shown that patients with unipolar depression can be differentiated into subgroups with either a myeloid-biased or lymphoid-biased immune cell profile [14]. Since unipolar depression has a heterogeneous appearance [35], it could be hypothesized that changes in specific cell linages might characterize specific subgroups of depression. Furthermore, previous studies have indicated a more pronounced inflammatory response to be related to more severe depression [36, 37]. This corroborates with the subgroup analyses of our study showing that higher levels of overall leukocytes, driven by higher levels of neutrophils, were associated with inpatient status and thus indicating that increased peripheral inflammation is associated with more severe depression. This could be explored further in new studies, as identification of subtypes of depression with specific immune cell profiles might pave the way for new immunotherapeutic strategies in the treatment of depression.

Experimental studies indicate that induced pro-inflammation increase depressive-like behavior in animals [38] and depressivelike symptoms in humans [39]. Childhood trauma [40] and environmental psychological stressors [41] have been associated with both increased peripheral inflammation and depression. Several mechanisms have been suggested to underlie this association, including alterations of the hypothalamic-pituitaryadrenal (HPA) axis through the impact on the kynurenine pathway and hereby the monoamine availability [42]. However, two recent meta-analyses evaluating blood and CSF monoamine metabolite levels found no significant differences on a group level, which question the current theory of lower brain serotonin levels as causative to depression in a majority of cases [9, 43]. Thus, the mechanisms underlying depression are not yet fully understood and likely heterogeneous, however, could perhaps be driven by brain alterations caused by systemic inflammation in subgroups. Interestingly, a recent animal study suggested that transplantation of bone marrow mononuclear cells can prevent depressive-like behavior in rats [44], indicating that more profound alterations in the peripheral immune system might be implicated in the pathogenesis of depression.

Systemic inflammation has been shown to impair the bloodbrain barrier [45], hereby potentially increasing the risk of peripheral immune cells entering the brain. When including the CSF studies stating to have healthy controls we did not find the CSF WCC to be significantly increased; however, one of the studies reported a range of CSF WCC up to 14 cells in one of their healthy control groups [34]. As this is markedly above the normal upper range of CSF WCC maximum of 5 cells, this is concerning since such high CSF WCC in the control group potentially masks group differences. When excluding this study and only including the CSF studies with healthy controls, CSF WCC was indeed increased among the patients with depression compared to healthy controls. Moreover, only one of the CSF studies had a primary aim of evaluating CSF WCC [37] and, thus, in the remaining three papers there are no details on information relevant to measurement of low-grade neuroinflammation by CSF WCC. This includes lack of information regarding correction for blood contamination and time from sample collection until laboratory analyses. The one study that corrected data for blood contamination found a CSF WCC to be higher among patients with depression and that the CSF WCC count was increased the most among patients with severe depression [37]. This dose-response relationship with severity of depression has only been evaluated in this one CSF WCC study [37]. Thus, more high-quality large-scale studies evaluating this direct marker of neuroinflammation is highly needed to explore inflammation in the brain related to depression.

Interestingly, we found a tendency towards long discontinuation of antidepressants (>14 days) to be associated with lower overall leukocyte count. As (some) antidepressants are thought to yield immunosuppressive effects [46] this result is somewhat surprising. However, as we also found inpatient-status to be associated with higher leukocyte count, the effect of antidepressants could be confounded by the most severely ill patients not having their medication discontinued. The tendency could also be due to different patterns in use of specific antidepressants prior to and after year 2000 as our sensitivity analysis hereof on leukocytes also revealed a subgroup difference.

Identification of subtypes of depression with specific immune cell profiles might pave the way for new immunotherapeutic strategies in the treatment of depression. Personalized immunotherapeutic treatment is widely used in cancer treatment - one example is adoptive cell therapy, where T-cells harvested from the patient are genetically modified ex-vivo to specifically target the tumor cells when reintroduced to the patient by infusion. Development of similar personalized treatment modalities for depression might help resolve the immune cell disruptions in this patient group.

Strengths and limitations

Despite being the most comprehensive meta-analysis on circulating immune cells in unipolar depression, several limitations should be considered. The laboratory methods used across the included studies were highly diverse and included (among others) microscopy, hematology analyzers and flow cytometric analyses. Especially the field of flow cytometric analysis and cell surface markers has evolved rapidly during the past forty years, thus compromising the comparability of studies on immune cells from the 1980–1990's with more recent literature [22]. Furthermore, international consensus on cell surface markers for immunophenotyping is needed. Only peer-reviewed studies were included and therefore, conference abstracts and other types of gray literature were not eligible for this analysis. Additionally, most of the studies included in this meta-analysis are comprised of heterogenous patient groups regarding the characteristics of the depression (subtypes, severity, and duration), and therefore it was not possible to investigate associations between immune cells and different characteristics of the disease.

Most of the included studies showed bias regarding adequate definition and representativeness of cases. In addition, around half of the included studies did not match or adjust for important confounding factors such as age and sex and even fewer studies matched for body-mass index and smoking, which is known to influence the immune system [47–49]. In addition, the degree of heterogeneity was high for many of the reported outcomes indicating a high variation among the included studies.

Conclusion and perspectives

This meta-analysis indicates immune activation in unipolar depression with alterations in both the myeloid and lymphoid immune cell linages. This activation appears to be most pronounced in inpatients, indicating a relation to depression severity. However, many of the included studies had risk of bias and did not match for important factors such as age, sex, bodymass index, and smoking. Future studies are needed to unravel whether the immune activation is restricted to a specific subtype of depression, while adjusting for important confounding factors. Moreover, further studies on immune cells in CSF are needed in order to explore whether cellular neuroinflammation is part of the pathogenesis of depression. Identification of the mechanism underlying the potential association between increased peripheral immune activity and depression would also enhance the understanding of the disorder and could lead to new immunotherapeutic treatment strategies in depression.

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AUTHOR CONTRIBUTIONS

MEB, SOW and BHF designed the study and systematic literature search. BHF conducted the screening of titles and abstracts. BHF and SOW did full text screening independently. BHF extracted all data. Data was validated by either NVS or SOW. TBB and NØ helped categorize immune cells. RHC made the statistical analysis plan and conducted the statistical analyses. All authors contributed to data interpretation. RHC, NVS and BHF conducted figures and tables. NVS drafted the manuscript including preparation of the supplementary. All authors contributed to the editing of the manuscript and all authors have read and accepted the final version of the manuscript.

FUNDING

This study was funded by grants from The Lundbeck Foundation (grant number R268–2016–3925 and R277–2018–1411). The funding agencies had no impact on the study design, data collection, data analysis, data interpretation, or writing of the report.

COMPETING INTERESTS

The authors declare no competing interests.

ADDITIONAL INFORMATION

Supplementary information The online version contains supplementary material available at https://doi.org/10.1038/s41380-022-01905-z.

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