

ORIGINAL ARTICLE

Synthesis and SARs of novel lincomycin derivatives Part 5: optimization of lincomycin analogs exhibiting potent antibacterial activities by chemical modification at the 6- and 7-positions

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In order to modify lincomycin at the C-6 and C-7 positions, we prepared target molecules, which have substituted pipercolinic acid at the 6-amino group and a *para*-substituted phenylthio group at the C-7 position, in application of palladium-catalyzed cross-coupling as a key reaction. As the result of structure-activity relationship (SAR) studies at the 6-position, analogs possessing 4'-*cis*-(cyclopropylmethyl)piperidine showed significantly strong antibacterial activities against *Streptococcus pneumoniae* and *Streptococcus pyogenes* with an *erm* gene. On the basis of SAR, we further synthesized novel analogs possessing 4'-*cis*-(cyclopropylmethyl)piperidine by transformation of a C-7 substituent. Consequently, novel derivatives possessing a *para*-heteroaromatic-phenylthio group at the C-7 position exhibited significantly strong activities against *S. pneumoniae* and *S. pyogenes* with an *erm* gene even when compared with those of telithromycin. Finally, *in vivo* efficacy of selected two derivatives was evaluated in a rat pulmonary infection model with resistant *S. pneumoniae* with *erm* + *mef* genes. One of them exhibited strong and constant *in vivo* efficacy in this model, and both compounds showed strong *in vivo* efficacy against resistant *S. pneumoniae* with a *mef* gene. *The Journal of Antibiotics* (2018) 71, 298–317; doi:10.1038/ja.2017.114; published online 1 November 2017

INTRODUCTION

Macrolide antibiotics have antibacterial activities against *S. pneumoniae*, *S. pyogenes*, *Haemophilus influenzae*, *Moraxella catarrhalis*, *Mycoplasma pneumoniae* and *Neisseria gonorrhoeae*, and have an acceptable safety profile as oral antibiotics. Consequently, macrolides have widely been used in clinical sites for bacterial respiratory infections. Recently, macrolide resistant bacteria with an *erm* gene have markedly increased.^{1–3} Clarithromycin⁴ and azithromycin^{5,6} are not effective against *S. pneumoniae* and *S. pyogenes* with an *erm* gene and have low sensitivity against *S. pneumoniae* with a *mef* gene (Figure 1 and Table 1). Although telithromycin (TEL)⁷ exhibits effective activities against *S. pneumoniae* with *erm* and/or *mef* genes, its activities are influenced by a *mef* gene. A serious liver damage^{8,9} and loss of consciousness^{10,11} were reported as side effects of TEL and medication with TEL was discontinued in Japan. Novel azalides reported by Miura *et al.*^{12,13} are also effective against the above resistant pathogens, but these analogs are still under research process. Development of an oral antibiotic possessing potent antibacterial activities and an acceptable safety profile is strongly desired in clinical sites for respiratory infections.

Lincomycin (LCM)^{14–17} and clindamycin (CLDM)¹⁸ are effective against clinically isolated pathogens with a *mef* gene, but they are not effective against resistant bacteria with an *erm* gene (Figure 1, Table 1).

As an overview, CLDM exhibits the following positive characters: (1) availability in p.o. and i.v. administrations (switch therapy is possible), (2) good distributions to the tissue and cells, (3) suppression¹⁹ of toxin production by streptococcal strains and (4) expected reasonable production cost of its derivatives compared with that of ketolides with a complex chemical structure. Thus, LCM derivatives might be clinically more valuable than ketolide antibiotics, if they are effective against Gram-positive pathogens with an *erm* gene.

Chemical modifications at the C-7 position of LCM were reported by several research groups.^{17,18,20–32} However, none of those compounds showed antibacterial activities against resistant Gram-positive pathogens with an *erm* gene. On the other hand, we reported that novel LCM derivatives modified at the C-7 position possessed antibacterial activities against resistant bacteria with an *erm* gene.^{33–43} In particular, compound 1 (Wakiyama *et al.*⁴⁰) (Figure 1) and its analogues (possessing a '1-methylpiperidin-3-yl' or '1-methyl-1,2,5,6-tetrahydropyridin-3-yl' moiety instead of a 'pyrimidin-5-yl' moiety in compound 1) had significantly potent antibacterial activities against resistant bacteria with an *erm* gene.

Chemical modification at the C-6 position of LCM and/or CLDM was also performed by several research groups.^{17,24,44–54} The C-6 side chain, originally 1'-*N*-methyl-4'-*trans*-*n*-propylproline, has the following characters: (1) diastereoisomers with 2',4'-*trans* configuration were

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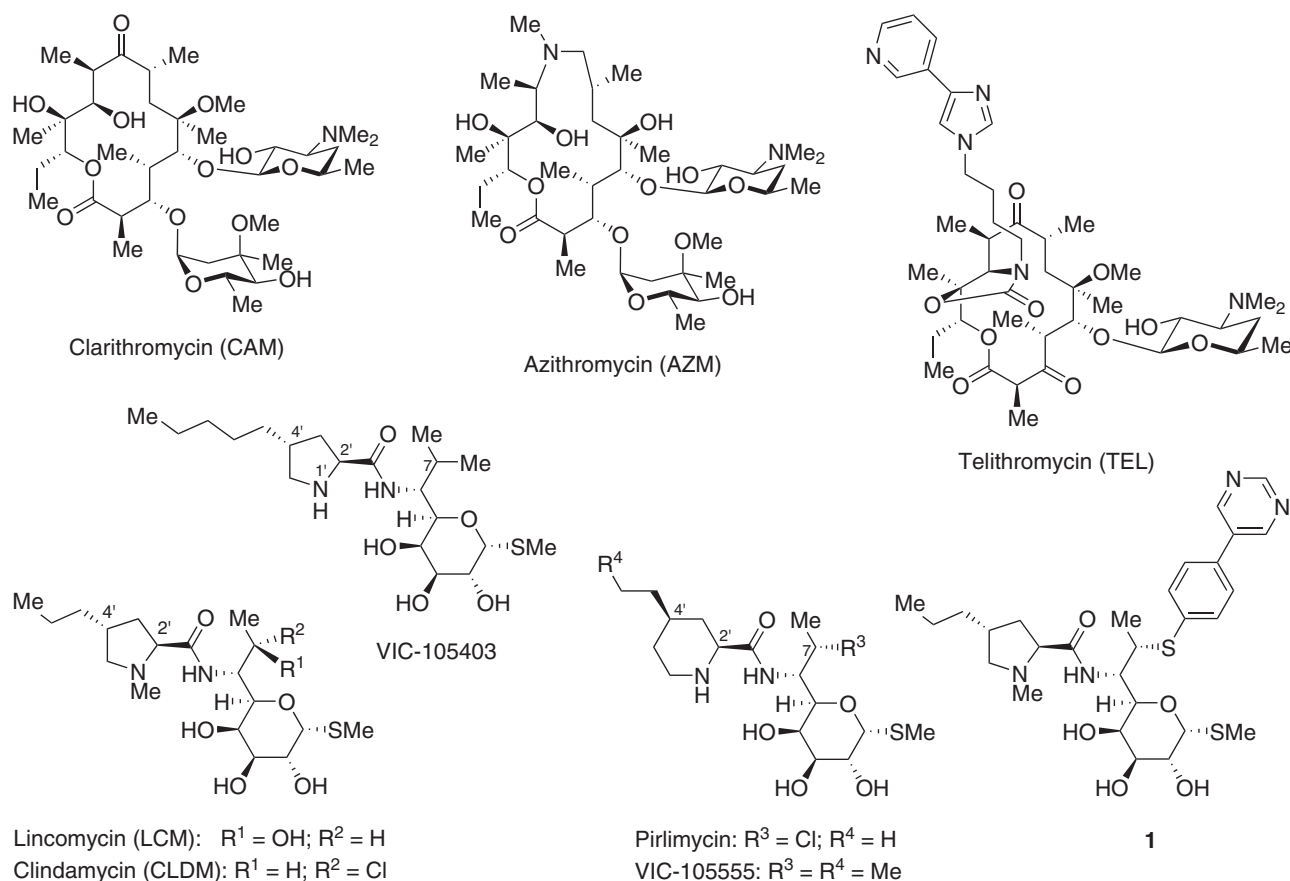


Figure 1 Chemical structures of macrolide derivatives and lincomycin/clindamycin derivatives.

Table 1 Antibacterial activities (MIC, $\mu\text{g ml}^{-1}$) of CAM, AZM, LCM, CLDM, VIC-105555, TEL and previously reported lincomycin derivative 1

Test organism ^a	Characteristics	CAM	AZM	LCM	CLDM	VIC-105555	TEL	1
<i>S. pneumoniae</i> DP1 Typel	Susceptible	0.03	0.06	1	0.06	0.03	≤ 0.008	≤ 0.008
<i>S. pneumoniae</i> -2	Susceptible	0.03	0.03	1	0.12	0.06	≤ 0.008	≤ 0.008
<i>S. pneumoniae</i> -3	Susceptible	0.015	0.03	0.25	0.06	0.03	≤ 0.008	≤ 0.008
<i>S. pneumoniae</i> -4	<i>ermAM</i> methylase (c)	> 128	> 128	> 128	> 128	> 128	0.5	0.5
<i>S. pneumoniae</i> -5	<i>ermAM</i> methylase (c)	> 128	> 128	> 128	> 128	> 128	2	1
<i>S. pneumoniae</i> -6	<i>ermAM</i> methylase (c) + <i>mefE</i>	> 128	> 128	> 128	> 128	> 128	1	2
<i>S. pneumoniae</i> -7	<i>ermAM</i> methylase (i)	> 128	> 128	128	128	128	0.03	0.25
<i>S. pneumoniae</i> -8	<i>ermAM</i> methylase (i)	> 128	> 128	128	128	128	0.03	0.25
<i>S. pneumoniae</i> -9	<i>mefE</i> efflux	0.5	0.5	1	0.12	0.015	0.06	≤ 0.008
<i>S. pyogenes</i> Cook	susceptible	0.015	0.06	0.12	0.06	0.06	≤ 0.008	≤ 0.008
<i>S. pyogenes</i> -2	<i>ermAM</i> methylase (c)	> 128	> 128	> 128	128	> 128	16	0.5
<i>S. pyogenes</i> -3	<i>mefE</i> efflux	8	8	0.25	0.12	0.06	0.25	0.015
<i>H. influenzae</i>	Susceptible	2	0.25	8	16	32	0.5	4
<i>H. influenzae</i> -2	Susceptible	4	1	16	8	16	2	2
<i>H. influenzae</i> -3	Susceptible	8	2	16	16	32	1	8
<i>H. influenzae</i> -4	Δ <i>acr</i>	0.5	0.5	4	1	2	0.25	0.06

Abbreviations: AZM, azithromycin; c, constitutive; CAM, clarithromycin; CLDM, clindamycin; 1, inducible Gray shading strains are target strains; LCM, lincomycin; TEL, telithromycin.

Gray shading strains are target strains.

^aAll strains except standard organisms were clinically isolated.

more potent than *cis* isomers; (2) 1'-*N*-demethylclindamycin was twice as active *in vitro* against *Sarcina lutea* as CLDM, but 1'-*N*-demethyl-lincomycin was about one twentieth as active as LCM;^{21,44-45} (3) as for chain length (H, Me to octyl) at the 4'-position of LCM, the hexyl analog showed maximum *in vitro* antibacterial activity;^{21,46} (4)

introduction of hetero atoms to the 4'-side chain essentially lost activity;^{17,21,47} and (5) VIC-105403 (Lewis *et al.*²⁴) (Figure 1) had potent activities compared with CLDM.

As another background information on chemical modifications at the C-6, azetidine,⁴⁸⁻⁵¹ piperidine^{48,49,52-54} and azepane analogs^{48,49}

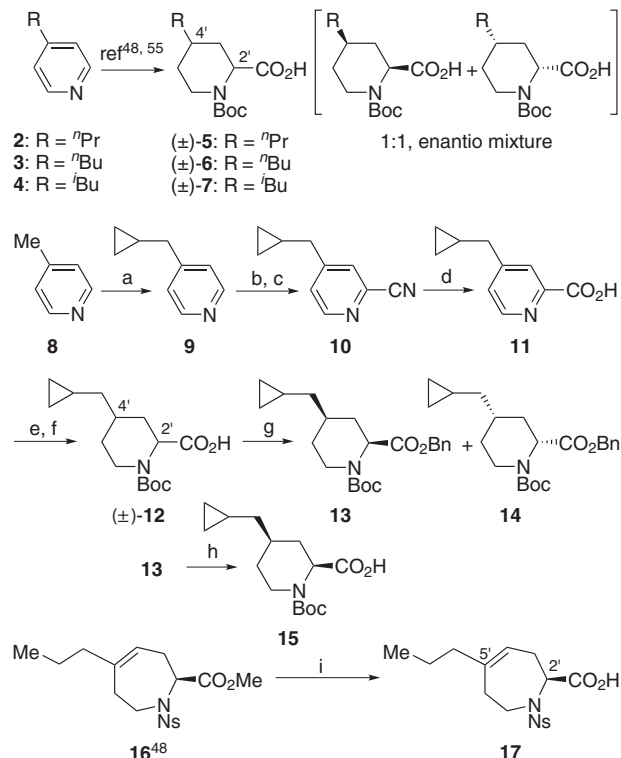
were synthesized accompanied with modifications at the C-7. Regarding azetidine derivatives, 3'-*trans*-cyclobutylethyl CLDM derivative showed significant antibacterial activities against sensitive *S. pneumoniae* compared with CLDM, but 3'-*trans*-cyclopropylmethyl, 3'-*trans*-*n*-propyl and 3'-*trans*-*n*-butyl analogs exhibited similar potency as CLDM. As a piperidine derivative, 4'-*cis*-ethyl CLDM analog, pirlimycin, is used in mastitis therapy for cattle in the European countries and United States. On the other hand, VIC-105555 (Figure 1) was selected as a candidate, which exhibited preferable pharmacokinetics and characteristic *in vitro* antibacterial activities against methicillin-resistant *Staphylococcus aureus* (MRSA) and *Enterococcus faecalis*. Furthermore, azepane-type CLDM analogs were also synthesized and 5'-(3-fluorobutyl) analog was 32 times as active (MIC: 0.25 µg ml⁻¹) against *H. influenzae* as CLDM (MIC: 8 µg ml⁻¹).⁴⁸

None of the C-6-modified compounds were disclosed to possess activities against Gram-positive-resistant bacteria with an *erm* gene. We reported that novel (7*S*)-substituted analogs⁴² modified at the N-1' and C-4' positions in a proline moiety had potent activities against Gram-positive-resistant bacteria with an *erm* gene. We further pursued modifications of LCM with a combination manner at the C-6 and C-7 positions in order to generate novel LCM derivatives exhibiting as strong antibacterial activities as TEL. Then, we synthesized novel (7*S*)-substituted analogs attached with piperidine or azepane instead of pyrrolidine (a part of proline) at the C-6 position. We have found three representative molecules so far and we chose a 'pyrimidin-5-yl'-phenyl derivative (1) (Figure 1) as a C-7 side chain for optimization of a C-6 moiety, because a '1-methylpiperidin-3-yl' moiety has a chiral center (anxiety for relatively complex production) and a '1-methyl-1,2,5,6-tetrahydropyridin-3-yl' moiety has an isolated double bond (anxiety for potential instability).

RESULTS AND DISCUSSION

Synthesis of the substituted piperidines and 2,3,6,7-tetrahydro-1*H*-azepine

Syntheses of substituted piperidines and 2,3,6,7-tetrahydro-1*H*-azepine are shown in Scheme 1. Substituted piperidines (±)-5-7 and 2,3,6,7-tetrahydro-1*H*-azepine 16 were synthesized by methods reported by Shuman *et al.*⁵⁵ and Lewis *et al.*⁴⁸ Compound (±)-12 was prepared from 4-methylpyridine (8) in improved reaction conditions based on reported methods^{48,55} shown in Scheme 1. It was reported that hydrogenation of disubstituted pyridine in the presence of PtO₂ resulted in a racemate of *cis*-products as major products by Lewis *et al.*⁴⁸ Going back in time, Shuman *et al.*⁵⁵ proved that 2,4-disubstituted piperidine prepared from disubstituted pyridine by hydrogenation had *cis*-configuration by NOE experiments. At the beginning of this research, we used (±)-*cis*-carboxylic acids 5-7 and 12, but later on we could separate (±)-12 into each *cis*-enantiomer for efficient synthetic study. Carboxylic acid (±)-12 was protected by a benzyl group for the purpose of optical resolution by HPLC and both enantiomers were purified by chiral column chromatography to obtain a desired compound 13. Stereochemistry of compounds 13 and 14 was assigned as following. Pirlimycin and VIC-105555 are reported as representative LCM derivatives possessing a substituted piperidine moiety (Figure 1). Absolute stereochemistry of pirlimycin was clarified by X-ray crystallographic studies.⁴⁹ Absolute stereochemistry of VIC-105555 was reported by Vicuron at 44th Interscience Conference on Antimicrobial Agents and Chemotherapy.⁵⁴ Both compounds have 2'-β-4'-β-configuration and they showed remarkable polarity (lower R_f value) and stronger potency compared with the corresponding 2'-α-4'-α-diastereoisomer, respectively. When we coupled a substituted piperidine with methyl α-thiolincosaminide (MTL), we assigned

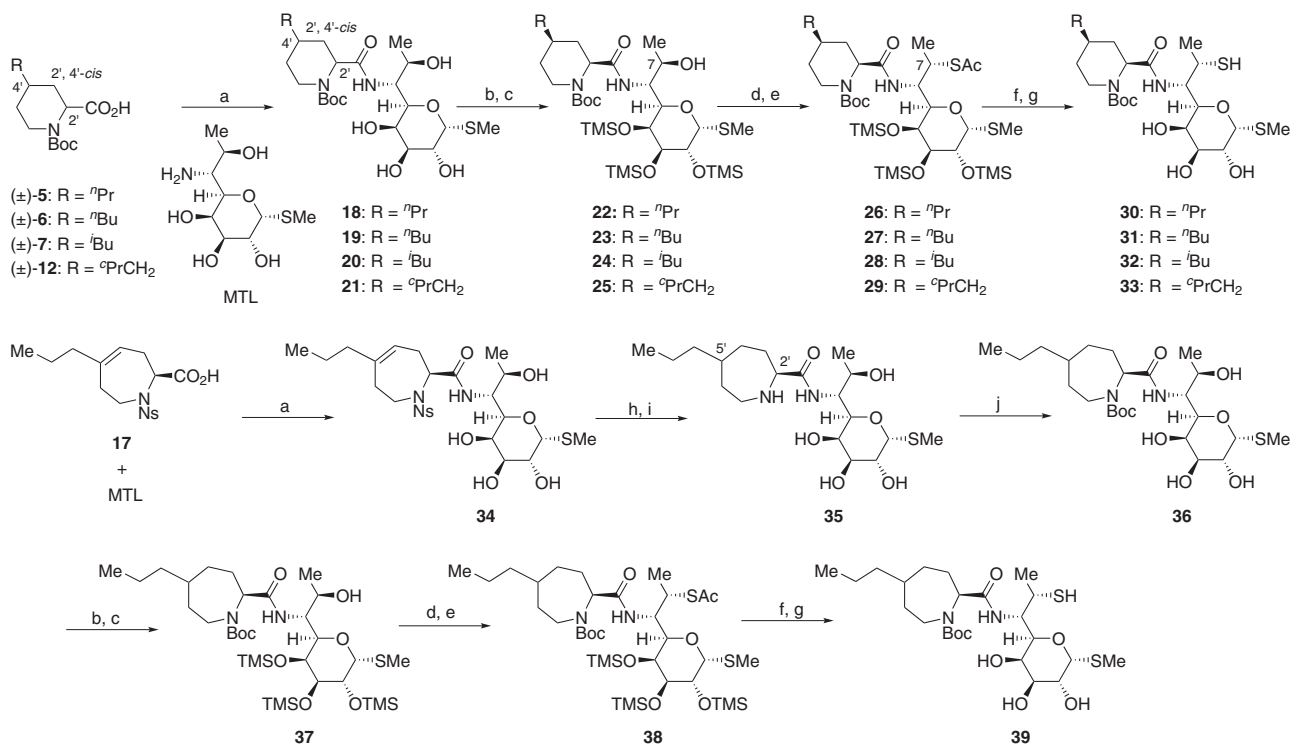


Scheme 1 Synthesis of substituted piperidines and 2,3,6,7-tetrahydro-1*H*-azepine. Conditions: (a) bromocyclopropane, lithium diisopropylamide, THF, -78 °C, 1 h; (b) *m*-chloroperoxybenzoic acid (*m*CPBA), CH₂Cl₂, 0 °C to r.t., 1 h; (c) TMSCN, Me₂NCOCI, CH₂Cl₂, 20 °C 40 min, then r.t., 17 h; (d) 5 N NaOH, MeOH, 50 °C, 8 h; (e) H₂, PtO₂, AcOH, r.t., 24 h; (f) Boc₂O, 2 N NaOH, dioxane, r.t., 15 h; (g) BnBr, ^tPr₂NEt, CH₃CN, r.t., 48 h; (h) H₂, Pd/C, MeOH, r.t., 1 h; (i) LiOH·H₂O, dioxane:H₂O = 4:1, r.t. 5 h.

2'-β-4'-β-configuration for a polar product. The benzyl group in 13 was removed by hydrogenolysis to give a key intermediate 15. A seven-membered intermediate (17) was prepared by basic hydrolysis of 16.

Synthesis of key intermediates 30–33 and 39

Syntheses of key intermediates 30–33 and 39 are shown in Scheme 2. Diastereomeric mixtures 18–21 and compound 34 were synthesized by coupling of compounds (±)-5-7, (±)-12 and 17 with MTL, respectively. MTL was prepared by a reported method.⁵⁶ Although each isomer was almost one to one mixture except 17 when the coupling reactions were completed, precipitation process gave 2'-β-4'-β-rich *cis*-isomers. As 'Experimental procedure' reported, ratio of diastereoisomeric mixture was difference for each compound. Tetra-*O*-trimethylsilylation of mixtures 18–21 and regioselective deprotection of the 7-*O*-TMS group followed by silica gel column chromatography finally gave single compounds 22–25^{48,57} as 2'-β-4'-β-pure *cis*-isomers. Methanesulfonylation of the 7-OH group and then S_N2 reaction by potassium thioacetate gave compounds 26–29. Key intermediates 30–33^{33–35,37–38,40,42} were prepared by deprotections of all TMS groups and an acetyl group. On the other hand, the Ns group of compound 34 was deprotected by 4-bromobenzenethiol under the basic condition, and then the olefin group was reduced by hydrogenation to give an azepane intermediate 35 (stereochemistry at the C-5' position is not assigned). An amino group of 35



Scheme 2 Synthesis of key intermediates **30** to **33** and **39**. Conditions: (a) *N,N*-dicyclohexylcarbodiimide (DCC) or EDC·HCl, HOBT, DMF, r.t., 6–20 h; (b) TMSCl, HMDS, pyridine, r.t., 20 min–1 h; (c) 6 N AcOH or 2 N AcOH, MeOH, r.t., 40 min–6 h; (d) MsCl, NEt₃, CH₂Cl₂, 0 °C to r.t., 1 h; (e) AcSK, DMF, 80 °C, 1.5–3 h; (f) 1 N HCl, MeOH, r.t. or 0 °C, 5–100 min; (g) NaOMe, MeOH, r.t., 15 min–3 h; (h) 4-bromobenzenethiol, Cs₂CO₃, DMF, r.t., 2 h; (i) H₂ (0.95 MPa), Pd/C, MeOH, 40 °C, 70 h; (j) Boc₂O, LiOH·H₂O, dioxane:H₂O = 1:1, r.t. 3 h.

was protected by a Boc group to give **36**, and a key intermediate **39** was synthesized from **36** by the similar procedures as described for the preparation of **30**.

Synthesis of novel (7*S*)-4-(pyrimidin-5-yl)phenylthio LCM derivatives possessing piperidine or azepane as the C-6 side chain
 Syntheses of novel (7*S*)-4-(pyrimidin-5-yl)phenylthio LCM derivatives possessing piperidine or azepane as the C-6 side chain are shown in Scheme 3. Compounds **40–43** and **48** were synthesized from key intermediates **30–33** and **39** by palladium-catalyzed cross-coupling reaction with 5-(4-bromophenyl)pyrimidine, respectively.^{33–35,38,40,42,58} The Boc group of **40–43** and **48** was finally removed with TFA to give desired compounds **44–47** and **49**.

Synthesis of divergent intermediate **54** and novel LCM derivatives possessing a 4-(2-(dimethylamino)ethyl)phenylthio group at the C-7 position

Because we had to develop a more divergent synthetic route than those exemplified in Schemes 2 and 3, we decided to apply the next key intermediate **54**. Syntheses of divergent intermediate **54** and novel LCM derivatives possessing a 4-(2-(dimethylamino)ethyl)phenylthio group at the C-7 position are shown in Scheme 4. Compound **50**⁵⁹ was synthesized by trifluoroacetylation of an amino group of MTL, and tetra-*O*-trimethylsilylation of all OH groups of **50** gave compound **51**. Divergent intermediate **54** was synthesized from **51** by the similar procedures as described for preparation of **30**. Palladium-catalyzed cross-coupling reaction of **54** with 2-(4-bromophenyl)-*N,N*-dimethylethanamine gave **55**, which was hydrolyzed in the presence of phase transfer catalyst under the basic condition to give diamine **56**. A

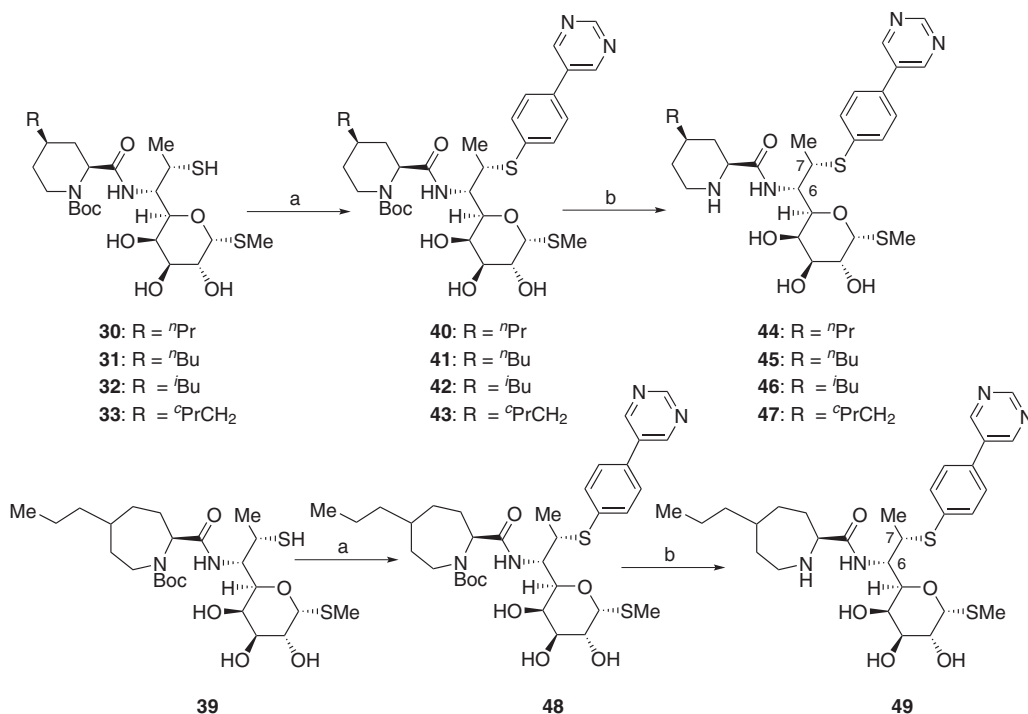
coupling reaction of **56** with enantio-pure **15** provided desired **57** with all carbon's framework. Deprotection of the Boc group finally gave **58** and its reductive *N*-methylation provided compound **59**.

Synthesis of novel 4'-*cis*-(cyclopropylmethyl)piperidine LCM derivatives possessing a 4-substituted phenylthio group at the C-7 position

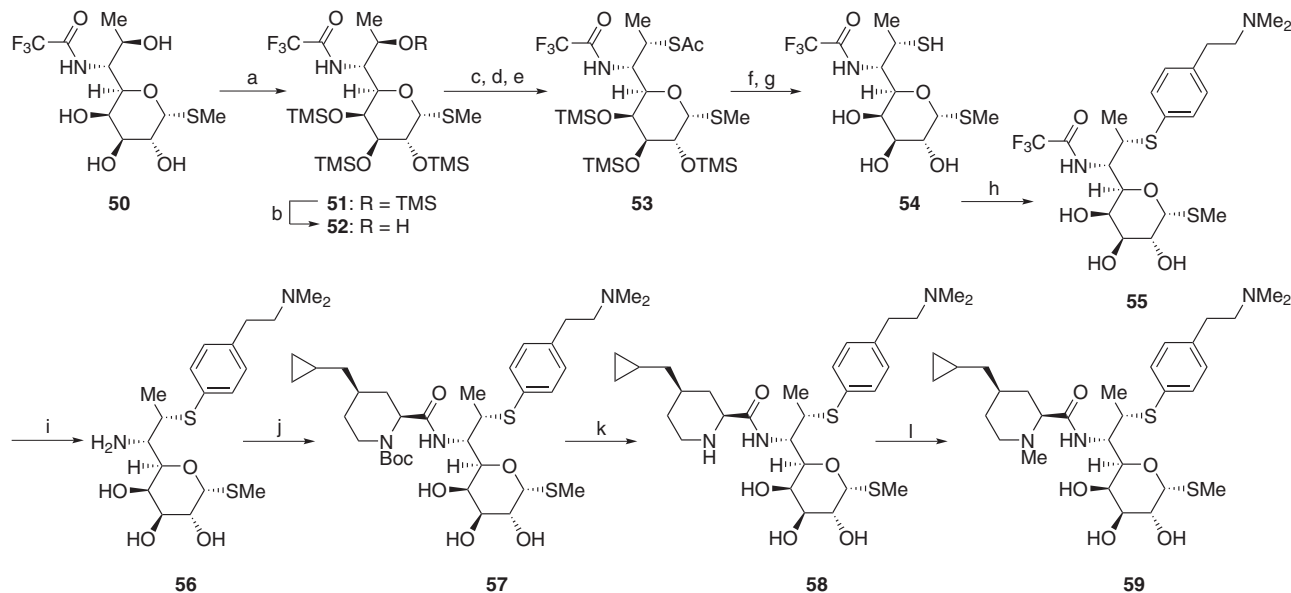
Syntheses of novel 4'-*cis*-(cyclopropylmethyl)piperidine LCM derivatives possessing a 4-substituted phenylthio group at the C-7 position are shown in Scheme 5. Compounds **60**, **61** and **63–65** were synthesized from the key intermediate **33** by palladium-catalyzed cross-coupling reaction with the corresponding 4-substituted phenyl bromides. Reduction of **61** afforded saturated *N*-methylpiperidine **62** as a mixture of diastereoisomers at an *N*-methylpiperidine ring. The first half of desired compounds **66**, **68**, **70**, **72**, **74** and **77** were prepared by deprotection of a Boc group and their free secondary amine was methylated by reductive alkylation to give the second half of desired compounds **67**, **69**, **71**, **73**, **75** and **78**, respectively. Compound **76** was also synthesized from **47** with the similar procedure. We confirmed that compound **76** derived from compound **33** had 4'-*cis*-stereochemistry by ROESY experiments. As the above, 4'-*cis*-stereochemistry of compound **12** was assigned.

SAR analysis of C-6 modified and (7*S*)-7-(4-(pyrimidin-5-yl)phenyl)thio-substituted LCM derivatives **44–47** and **49**

We reported potent antibacterial activities of **1** possessing the (7*S*)-4-(pyrimidin-5-yl)phenylthio group at the C-7 position. For the purpose of generating novel compounds possessing more potent antibacterial activities against resistant Gram-positive pathogens with



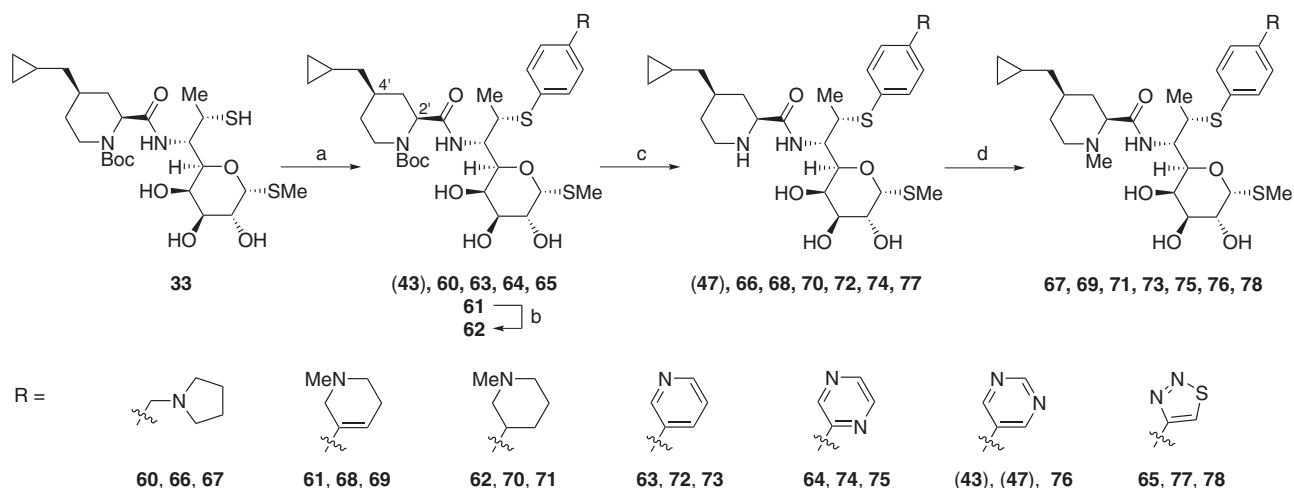
Scheme 3 Synthesis of novel (7*S*)-4-(pyrimidin-5-yl)phenylthio LCM derivatives possessing piperidine or azepane as the C-6 side chain. Conditions: (a) 5-(4-bromophenyl)pyrimidine, Pd₂(dba)₃, Xantphos, ⁱPr₂NEt, dioxane, reflux, 2–6 h; (b) TFA, CH₂Cl₂, –20 °C to r.t., 1.5–6 h.



Scheme 4 Synthesis of divergent intermediate **54** and novel LCM derivatives possessing a 4-(2-(dimethylamino)ethyl)phenylthio group at the C-7 position. Conditions: (a) TMSCl, HMDS, pyridine, r.t., 1 h; (b) 6 *N* AcOH, MeOH, r.t., 15 min; (c) MsCl, NEt₃, CHCl₃, r.t., 1 h; (d) AcSK, DMF, 80 °C, 1.5 h; (e) TMSCl, HMDS, pyridine, r.t., 3 h; (f) 1 *N* HCl, MeOH, r.t., 10 min; (g) NaOMe, MeOH, r.t., 15 min; (h) 2-(4-bromophenyl)-*N,N*-dimethylethanamine, Pd₂(dba)₃, Xantphos, ⁱPr₂NEt, dioxane, reflux, 17 h; (i) 20% aq. KOH, *N*-benzyl-*N,N,N*-triethylammonium bromide, r.t., 4 h; (j) **15**, EDC·HCl, HOBT, DMF, r.t., 5.5 h; (k) TFA, CH₂Cl₂, 0 °C, 3.5 h; (l) 36% HCHO, NaBH(OAc)₃, AcOH, MeOH, r.t., 1 h.

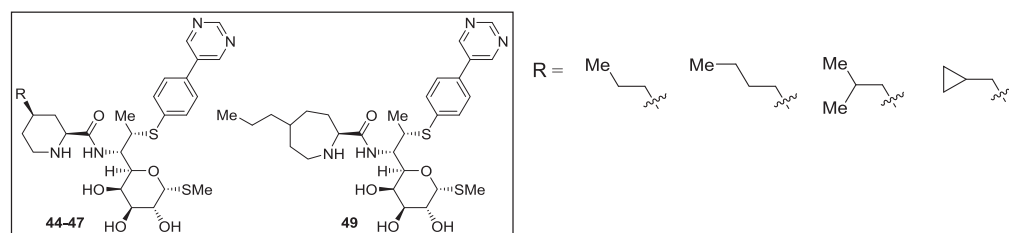
an *erm* gene, we performed an SAR analysis of C-6-modified and (7*S*)-7-(4-(pyrimidin-5-yl)phenyl)thio-substituted LCM derivatives **44–47** and **49** (Table 2). According to our reported SAR studies, (7*S*) stereochemistry was selected among all novel derivatives.⁴⁰ Compound **44**, which possesses *n*-propyl-piperidine instead of *n*-propyl-pyrrolidine as the C-6 side chain, showed stronger activities against

resistant *S. pneumoniae* with an *erm* gene than **1**. However, its antibacterial activity against resistant *S. pneumoniae* with both *erm* and *mef* genes (*S. pneumoniae*-6) was not sufficient (MIC: 1 µg ml⁻¹). Because there were a couple of reports^{21,42,46} stating that elongation of a side chain in a piperidine ring enhanced antibacterial activity, we synthesized alternative derivatives with a longer carbon chain



Scheme 5 Synthesis of novel 4'-*cis*-(cyclopropylmethyl)piperidine LCM derivatives possessing an aliphatic- or aromatic-phenylthio group at the C-7 position. Conditions: (a) the corresponding 4-substituted phenylbromides Pd₂(dba)₃, Xantphos, ^tPr₂NEt, dioxane, reflux, 4–5 h; (b) 4-methylbenzenesulfonylhydrazide, toluene, reflux, 5.5 h; (c) TFA, CH₂Cl₂, –20 °C to r.t., 0.5–5 h; (d) 36% HCHO, NaBH(OAc)₃, AcOH, MeOH, r.t., 0.5–2 h.

Table 2 Antibacterial activities (MIC, μg ml⁻¹) of novel lincomycin derivatives modified at the C-6 position



Test organism ^a	Characteristics	44	45	46	47	49	TEL
<i>Streptococcus pneumoniae</i> DPI Typel	susceptible	0.015	0.015	0.015	≤0.008	≤0.008	≤0.008
<i>S. pneumoniae</i> -2	susceptible	0.015	0.015	0.03	≤0.008	≤0.008	≤0.008
<i>S. pneumoniae</i> -3	susceptible	0.015	0.03	0.015	0.015	≤0.008	≤0.008
<i>S. pneumoniae</i> -4	<i>ermAM</i> methylase (c)	0.5	1	2	0.03	0.12	0.5
<i>S. pneumoniae</i> -5	<i>ermAM</i> methylase (c)	0.25	1	2	0.03	0.12	2
<i>S. pneumoniae</i> -6	<i>ermAM</i> methylase (c) + <i>mefE</i>	1	2	2	0.06	0.25	1
<i>S. pneumoniae</i> -7	<i>ermAM</i> methylase (i)	0.06	0.25	NT	0.015	≤0.008	0.03
<i>S. pneumoniae</i> -8	<i>ermAM</i> methylase (i)	0.03	0.12	NT	0.015	NT	0.03
<i>S. pneumoniae</i> -9	<i>mefE</i> efflux	≤0.008	≤0.008	NT	≤0.008	≤0.008	0.06
<i>Streptococcus pyogenes</i> Cook	susceptible	0.015	≤0.008	0.015	0.015	≤0.008	≤0.008
<i>S. pyogenes</i> -2	<i>ermAM</i> methylase (c)	0.25	0.5	0.06	0.03	0.03	16
<i>S. pyogenes</i> -3	<i>mefE</i> efflux	0.015	0.03	0.015	0.015	≤0.008	0.25
<i>Haemophilus influenzae</i>	susceptible	2	4	16	1	2	0.5
<i>H. influenzae</i> -2	susceptible	2	4	16	1	2	2
<i>H. influenzae</i> -3	susceptible	8	16	>64	2	4	1
<i>H. influenzae</i> -4	Δ <i>acr</i>	0.06	0.12	0.5	0.03	0.03	0.25

Abbreviations: c, constitutive; i, inducible; NT, not tested; TEL, telithromycin. Gray shading strains are target strains.

^aAll strains except standard organisms were clinically isolated.

or a branched side chain. However, antibacterial activities of compounds 45 and 46 were not improved. On the other hand, both compounds 47 and 49 possessing a 4'-*cis*-(cyclopropylmethyl)piperidine-2-carbonyl and 5'-*n*-propylazepane-2-carbonyl group at the C-6 position exhibited potent antibacterial activities against resistant *S. pneumoniae* with *erm* gene. Because the cyclopropylmethyl analog (47) especially exhibited stronger activities against Gram-positive pathogens with an *erm* gene even compared with TEL, we chose a 4'-*cis*-cyclopropylmethyl moiety as the C-6 side chain for further medicinal chemistry.

Antibacterial activities of novel LCM derivatives 58, 59 and 66–71 possessing an aliphatic amine at the para-position of phenylthio group at the C-7 position

For the purpose of accumulating detail information of SAR on (7S)-7-(4-substituted-phenylthio) LCM derivatives with a 4'-*cis*-(cyclopropylmethyl)piperidine moiety, we synthesized novel derivatives possessing various substituents at the C-7 position with a set of R² = both 'N-H' and 'N-Me' analogs (Table 3). Consequently, compounds 58, 66 and 68–71 showed potent antibacterial activities against target pathogens

Table 3 Antibacterial activities (MIC, $\mu\text{g ml}^{-1}$) of novel lincomycin derivatives modified at the C-7 position with an aliphatic moiety

Test organism ^a	Characteristics	R ¹ =									
		58	59	66	67	68	69	70	71	TEL	
<i>Streptococcus pneumoniae</i> DP1 TypeI	susceptible	0.03	≤0.008	0.03	0.015	0.015	0.06	0.06	0.015	≤0.008	
<i>S. pneumoniae</i> -2	susceptible	0.03	≤0.008	0.03	0.03	0.015	0.06	0.06	0.03	≤0.008	
<i>S. pneumoniae</i> -3	susceptible	0.06	0.03	0.06	0.06	0.015	0.06	0.06	0.03	≤0.008	
<i>S. pneumoniae</i> -4	<i>ermAM</i> methylase (c)	0.5	1	0.25	0.5	0.06	0.25	0.25	0.25	0.5	
<i>S. pneumoniae</i> -5	<i>ermAM</i> methylase (c)	0.5	0.5	0.25	0.5	0.06	0.25	0.25	0.25	2	
<i>S. pneumoniae</i> -6	<i>ermAM</i> methylase (c) + <i>mefE</i>	0.5	2	0.25	1	0.06	0.5	0.12	0.5	1	
<i>S. pneumoniae</i> -7	<i>ermAM</i> methylase (i)	0.03	0.015	0.03	0.06	0.03	0.06	0.06	0.06	0.03	
<i>S. pneumoniae</i> -8	<i>ermAM</i> methylase (i)	NT	NT	NT	NT	0.03	0.06	0.06	0.06	0.03	
<i>S. pneumoniae</i> -9	<i>mefE</i> efflux	0.015	≤0.008	0.015	≤0.008	0.015	0.03	0.03	0.015	0.06	
<i>Streptococcus pyogenes</i> Cook	susceptible	0.015	0.015	0.06	0.06	0.015	0.03	0.03	0.03	≤0.008	
<i>S. pyogenes</i> -2	<i>ermAM</i> methylase (c)	0.12	0.5	0.12	0.5	0.03	0.12	0.06	0.12	16	
<i>S. pyogenes</i> -3	<i>mefE</i> efflux	0.03	0.015	0.06	0.12	0.015	0.03	0.03	0.03	0.25	
<i>Haemophilus influenzae</i>	susceptible	2	2	1	4	0.5	2	1	2	0.5	
<i>H. influenzae</i> -2	susceptible	4	1	4	2	1	2	4	4	2	
<i>H. influenzae</i> -3	susceptible	4	4	4	8	2	4	4	8	1	
<i>H. influenzae</i> -4	Δ <i>acr</i>	0.12	0.12	0.12	0.12	0.06	0.12	0.12	0.12	0.25	

Abbreviations: c, constitutive; i, inducible; NT, not tested; TEL, telithromycin.

Gray shading strains are target strains.

^aAll strains except standard organisms were clinically isolated.

with an *erm* gene and their activities were relatively stronger even when compared with those of TEL. In addition, antibacterial activity of all compounds against *S. pyogenes* with an *erm* gene was significantly potent than that of TEL. We confirmed that combination of chemical modifications with the 4'-*cis*-(cyclopropylmethyl)piperidine group at the C-6 position and an aliphatic amine to the *para*-position of a phenylthio group at the C-7 position was important to enhance antibacterial activities against *S. pneumoniae* and *S. pyogenes* with an *erm* gene.

Antibacterial activities of novel LCM derivatives 47 and 72–78 possessing an aromatic amine at the *para*-position of a phenylthio group at the C-7 position

In order to expand possibilities of the combination modification at both the C-6 and C-7 positions, we synthesized novel derivatives possessing various aromatic amines as a substituent on the phenyl group with a set of both 'N-H' and 'N-Me' analogs (Table 4). All their antibacterial activities against target Gram-positive pathogens with an *erm* gene were also relatively stronger than those of TEL. To be more precise, compounds 47 and 72 showed potent activities in 'N-H' analogs and compound 76 exhibited the strongest activities among all 'N-Me' analogs in this article. As pharmacokinetic property must be different between 'N-H' and 'N-Me' analogs, it is important to select these two types of analogs for further development. Furthermore, antimicrobial activity of compounds 72, 47 and 77 against *H. influenzae* was relatively strong among all LCM derivatives we reported, and their potency was stronger than that of clarithromycin and catching up with that of TEL. We also investigated antibacterial activity against *M. pneumoniae* (Table 4), because resistant *M. pneumoniae* is causing problems for respiratory infections in clinical sites. All evaluated compounds including 47 and 76 had significant antibacterial activity against resistant *M. pneumoniae*, which TEL was not effective against. We could generate several novel LCM derivatives exhibiting very strong antibacterial activities against resistant Gram-positive pathogens with *erm* and/or *mef* genes by combination

modification at the C-6 position (the proline moiety) and the C-7 position. These derivatives were also effective against resistant *M. pneumoniae*.

In vitro antibacterial activity (sensitivity distribution analysis) of selected compounds against sixty clinical isolates of *S. pneumoniae*

We evaluated the antibacterial activity of compounds 47, 68, 72, 76, 77 and TEL against 60 clinical isolates of *S. pneumoniae* including susceptible strains and resistant strains with *erm* and/or *mef* genes for sensitivity distribution analysis (Figure 2). MIC₉₀ values of five novel LCM derivatives (0.06–0.125 $\mu\text{g ml}^{-1}$) were relatively smaller than that of TEL (0.25 $\mu\text{g ml}^{-1}$). Notably, 47 and 72 were significantly potent among tested compounds. These results reflect MIC values in Table 4 and it was suggested that these derivatives would also be effective against *S. pneumoniae* in clinical sites.

In vivo efficacy of 47 and 76 (subcutaneous administration) in rat pulmonary infection model with resistant *S. pneumoniae* with *erm* + *mef* genes and a *mef* gene

We finally investigated the *in vivo* efficacy of selected compounds in rat pulmonary infection model with resistant *S. pneumoniae* with *erm* + *mef* genes. Among derivatives reported in this study, *in vitro* activities of compounds 47 and 72 are rather strong (Figure 2). On the other hand, we had to clarify *in vivo* efficacy in the set of 'N-H' and 'N-Me' in the piperidine moiety (to evaluate '72 and 73' or '47 and 76'). As *in vitro* activities of 73 were slightly weaker than those of 76, we decided to select the set of compounds 47 and 76 for *in vivo* evaluation. Compound 72 had weak hemolytic activity and thus compound 72 might not be appropriate for further evaluation. Compounds 47, 76 and TEL were subcutaneously administered (10 mg kg⁻¹) to rats at 2 h after bacterial infection, and *in vivo* efficacies are shown in Figure 3a. Compound 47 exhibited strong *in vivo* efficacy (3 log reduction or more) against resistant *S. pneumoniae* with *erm* + *mef* genes and its efficacy was constant (small s.d. value) compared with that of TEL (<2 log reduction). For our

Table 4 Antibacterial activities (MIC, $\mu\text{g ml}^{-1}$) of optimized novel lincomycin derivatives modified at the C-7 position with an aromatic moiety

Test organism ^a	Characteristics	72	73	74	75	47	76	77	78	TEL
<i>Streptococcus pneumoniae</i> DP1 Type1	susceptible	0.015	0.03	≤ 0.008	0.03	≤ 0.008	≤ 0.008	≤ 0.008	≤ 0.008	≤ 0.008
<i>S. pneumoniae</i> -2	susceptible	0.015	0.03	≤ 0.008	0.03	≤ 0.008	≤ 0.008	≤ 0.008	0.015	≤ 0.008
<i>S. pneumoniae</i> -3	susceptible	0.015	0.06	0.015	0.06	0.015	0.03	0.015	0.015	≤ 0.008
<i>S. pneumoniae</i> -4	<i>ermAM</i> methylase (c)	0.03	0.5	0.06	0.5	0.03	0.25	0.06	0.5	0.5
<i>S. pneumoniae</i> -5	<i>ermAM</i> methylase (c)	0.06	0.25	0.06	0.5	0.03	0.12	0.12	0.5	2
<i>S. pneumoniae</i> -6	<i>ermAM</i> methylase (c) + <i>mefE</i>	0.06	0.5	0.12	1	0.06	0.5	0.25	0.5	1
<i>S. pneumoniae</i> -7	<i>ermAM</i> methylase (i)	0.015	0.06	0.03	0.25	0.015	0.015	0.03	0.25	0.03
<i>S. pneumoniae</i> -8	<i>ermAM</i> methylase (i)	NT	NT	0.03	0.25	0.015	0.03	0.03	0.25	0.03
<i>S. pneumoniae</i> -9	<i>mefE</i> efflux	≤ 0.008	0.015	≤ 0.008	0.03	≤ 0.008	≤ 0.008	≤ 0.008	≤ 0.008	0.06
<i>Streptococcus pyogenes</i> Cook	susceptible	≤ 0.008	0.015	≤ 0.008	0.015	0.015	0.015	≤ 0.008	≤ 0.008	≤ 0.008
<i>S. pyogenes</i> -2	<i>ermAM</i> methylase (c)	0.06	0.25	0.03	0.25	0.03	0.12	0.03	0.12	16
<i>S. pyogenes</i> -3	<i>mefE</i> efflux	0.015	0.03	0.015	0.03	0.015	≤ 0.008	0.015	0.015	0.25
<i>Haemophilus influenzae</i>	susceptible	1	4	2	4	1	4	1	4	0.5
<i>H. influenzae</i> -2	susceptible	2	4	2	4	1	2	1	2	2
<i>H. influenzae</i> -3	susceptible	2	8	4	8	2	4	2	4	1
<i>H. influenzae</i> -4	Δ acr	0.03	0.12	0.03	0.12	0.03	0.06	0.03	0.03	0.25
<i>Mycoplasma pneumoniae</i> -1	susceptible	NT	NT	≤ 0.004	NT	≤ 0.004	≤ 0.004	≤ 0.004	NT	≤ 0.004
<i>M. pneumoniae</i> -2	A2063G	NT	NT	≤ 0.03	NT	≤ 0.03	≤ 0.03	≤ 0.03	NT	64

Abbreviations: c, constitutive; i, inducible; NT, not tested; TEL, telithromycin. Gray shading strains are target strains.

^aAll strains except standard organisms were clinically isolated.

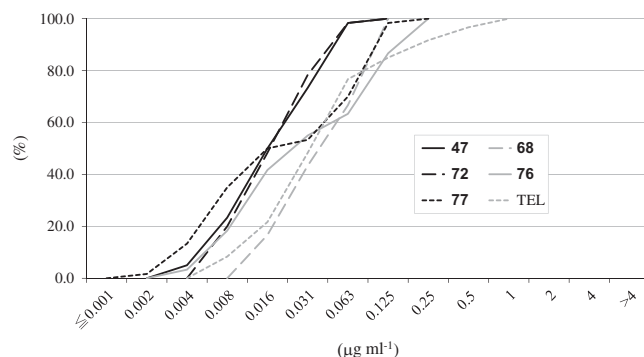


Figure 2 *In vitro* antibacterial activity (sensitivity distribution) of compounds 47, 68, 72, 76, 77 and TEL against 60 clinical isolates of *S. pneumoniae*.

references, we also evaluated *in vivo* efficacy of those by subcutaneous administration (3 mg kg^{-1}) to rats at 2 h after bacterial infection with *S. pneumoniae* with a *mef* gene, because resistant strains with a *mef* gene have increased in the US.⁶⁰ As a result shown in Figure 3b, 47 and 76 had significantly strong *in vivo* efficacy as expected on the basis of *in vitro* evaluation. *In vivo* efficacy of 76 (5 log reduction) was very constant (0 s.d. value) compared with that of TEL. Clinical efficacy of these novel LCM derivatives is expected from the above fundamental experimental data.

CONCLUSION

As the result of SAR studies at the 6-position of 44–47 and 49, compound 47 possessing 4'-*cis*-(cyclopropylmethyl)piperidine showed significantly strong antibacterial activities against *S. pneumoniae* and *S. pyogenes* with an *erm* gene. On the basis of SAR, we synthesized novel analogs possessing 4'-*cis*-(cyclopropylmethyl)piperidine by

transformation of a C-7 substituent. Consequently, compounds 47, 68, 72, 76 and 77 (Figure 4) exhibited significantly strong activities against *S. pneumoniae* and *S. pyogenes* with an *erm* gene even when compared with those of TEL. Then, the *in vitro* antibacterial activities of compounds 47, 68, 72, 76, 77 and TEL were evaluated (sensitivity distribution analysis) against 60 clinical isolates of *S. pneumoniae* containing sensitive bacteria and resistant bacteria with *erm* and/or *mef* genes. As a result, compounds 47 and 72 showed relatively strong activities than that of TEL. Finally, the *in vivo* efficacy of compound 47 and its 1'-*N*-Me-derivative 76 was evaluated in the rat pulmonary infection model (subcutaneous administration) with resistant *S. pneumoniae* with *erm* + *mef* genes. Compound 47 exhibited strong and constant *in vivo* efficacy. Moreover, compounds 47 and 76 showed strong *in vivo* efficacy against resistant *S. pneumoniae* with a *mef* gene. These two compounds are under consideration toward next developing stage.

EXPERIMENTAL PROCEDURE

General methods

¹H NMR spectra were measured with a BRUKER Ascend 400 NMR spectrometer (BRUKER Corporation, Coventry, UK) for 400 MHz, JEOL JNM-GSX 400 NMR spectrometer (JEOL Ltd, Tokyo, Japan) for 400 MHz or a Varian Gemini 300 NMR spectrometer (Varian Inc., Palo Alto, CA, USA) for 300 MHz in CDCl₃ or CD₃OD. TMS (0 p.p.m.) in CDCl₃ or CD₃OD was used as an internal reference standard. Mass spectra were obtained on a JEOL JMS-700 mass spectrometer (JEOL Ltd) or Agilent Technologies 6530-Q-TOF-LC/MS mass spectrometer (Agilent Technologies, Santa Clara, CA, USA). The optical rotations were recorded with Jasco P-2300 digital polarimeter (JASCO Corporation, Tokyo, Japan). Column chromatography was performed with silica gel (Wakogel C200, Wako Pure Chemical Industries Ltd, Osaka, Japan). Preparative TLC was performed with silica gel (Merck, Darmstadt, Germany; TLC plates Silica gel 60 F254). All organic extracts were dried over anhydrous

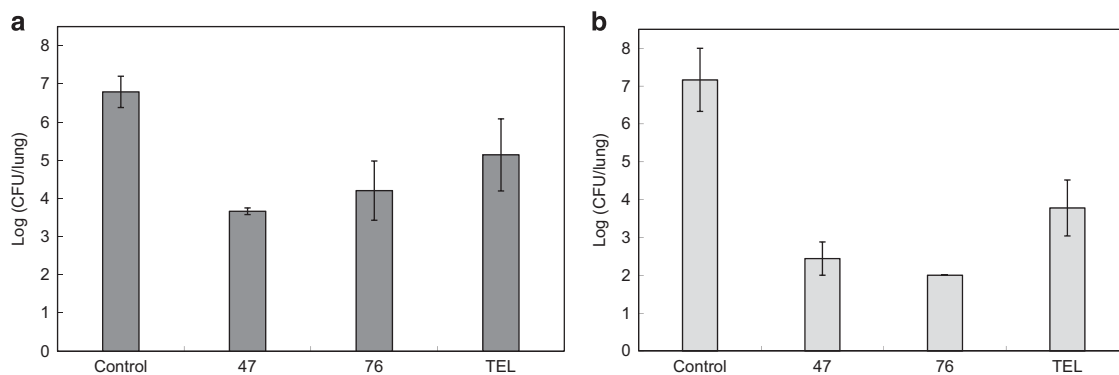


Figure 3 (a) *In vivo* efficacy of **47** and **76** in a rat pulmonary infection model with *S. pneumoniae* MSC06856 (*erm* + *mef*). (b) *In vivo* efficacy of **47** and **76** in a rat pulmonary neutropenic infection model with *S. pneumoniae* MSC06729 (*mef*). Comparison of the efficacy of novel lincomycin derivatives **47** and **76** in a rat pulmonary neutropenic infection model with *S. pneumoniae* MSC06856 (*erm* + *mef*) and *S. pneumoniae* MSC06729 (*mef*). Three rats per group were rendered neutropenic and 10^6 CFU per rat of *S. pneumoniae* MSC06856 or *S. pneumoniae* MSC06729 was injected into the lung, followed by s.c. administration of the test compounds at 2 h after infection. The mean log₁₀ CFU per lung recovered from the infected lung after 24 h is shown. Error bars represent the s.d.

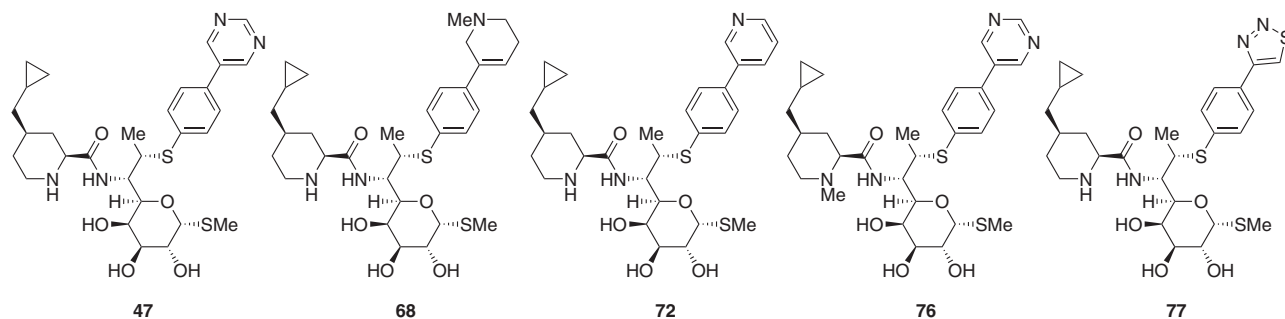


Figure 4 Structures of novel lincomycin derivatives possessing strong *in vitro* antibacterial activity.

MgSO₄ and the solvent was removed with a rotary evaporator under reduced pressure.

4-(Cyclopropylmethyl)pyridine (9)

To a solution of **8** (19.0 g, 204 mmol) in THF (136 ml) at -78°C was added 2.0 M lithium diisopropylamide in tetrahydrofuran (THF) solution (204 ml, 408 mmol) and then was stirred in argon atmosphere at -40°C for 20 min. The mixture was cooled to -78°C . Then, bromocyclopropane (16.3 ml, 204 mmol) was added with dropwise to the solution. After stirring for 1 h, the solution was poured into saturated aqueous NH₄Cl. The desired compound was extracted with ethyl acetate, was washed with brine and then the organic phase was dried over Na₂SO₄, filtrated and concentrated under reduced pressure. The resulting residue was purified by distillation under reduced pressure (84 °C/8 mm Hg) to obtain the title compound (13.8 g, 51%) as colorless oil. Fast atom bombardment (FAB)-MS *m/z* 134 (M+H)⁺ as C₉H₁₁N; ¹H NMR (400 MHz, CDCl₃) δ 0.18–0.26 (m, 2 H), 0.54–0.62 (m, 2 H), 0.93–1.05 (m, 1 H), 2.54 (d, *J* = 7.1 Hz, 2 H), 7.17–7.23 (m, 2 H), 8.47–8.53 (m, 2 H).

4-(Cyclopropylmethyl)picolinonitrile (10)

To a solution of **9** (25.5 g, 191 mmol) in CH₂Cl₂ (300 ml) at 0 °C was added *m*-chloroperoxybenzoic acid (50.8 g, 191 mmol) and stirred at room temperature for 1 h. To the mixture was added Na₂S₂O₃ solution (75 g in 150 ml of H₂O). The solution was added to mixture of saturated aqueous NaHCO₃ (500 ml), saturated aqueous K₂CO₃ (40 ml) and CHCl₃ (500 ml). The organic phase was separated and then further extracted twice with CHCl₃ (500 ml)-isopropanol (100 ml), the organic phase was dried over Na₂SO₄, filtrated and concentrated under reduced pressure to obtain 4-(cyclopropylmethyl)pyridine *N*-oxide (30.7 g as crude). ¹H NMR (400 MHz, CDCl₃) δ 0.15–0.26 (m, 2 H),

0.54–0.66 (m, 2 H), 0.90–1.02 (m, 1 H), 2.54 (d, *J* = 7.1 Hz, 2 H), 7.11–7.24 (m, 2 H), 8.08–8.18 (m, 2 H).

To a solution of 4-(cyclopropylmethyl)pyridine *N*-oxide (30.7 g) in CH₂Cl₂ (350 ml) were added trimethylsilanecarbonitrile (30.6 ml, 0.229 mmol) and dimethylcarbamic chloride (7.03 ml, 76.3 mmol) at room temperature. Then, dimethylcarbamic chloride (7.03 ml, 76.3 mmol) was added in two portions after 20 min interval to the mixture at 20 °C. The mixture was stirred at room temperature for 17 h. The solution was added to 10% aqueous K₂CO₃. The desired compound was extracted with CH₂Cl₂ and then the organic phase was dried over Na₂SO₄, filtrated and concentrated under reduced pressure. The resulting residue was purified by silica gel column chromatography (hexane to hexane/ethyl acetate = 17/3) to obtain the title compound (25.5 g, 84% in 2 steps) as a colorless oil. EI-MS *m/z* 158 (M)⁺ as C₁₀H₁₀N₂; ¹H NMR (400 MHz, CDCl₃) δ 0.22–0.28 (m, 2 H), 0.60–0.70 (m, 2 H), 0.92–1.06 (m, 1 H), 2.61 (d, *J* = 7.1 Hz, 2 H), 7.41–7.46 (m, 1 H), 7.64 (br dd, *J* = 1.7, 0.7 Hz, 1 H), 8.55–8.64 (m, 1 H).

4-(Cyclopropylmethyl)picolinic acid (11)

To a solution of **10** (25.5 g, 161 mmol) in MeOH (300 ml) was added 5 M aqueous NaOH (250 ml) and stirred at 50 °C for 8 h. The mixture was cooled down to 0 °C, added to 5 M aqueous HCl (250 ml) at 0 °C and then concentrated under reduced pressure to remove MeOH. The solution was adjusted at pH 3 by 1 M aqueous HCl, extracted with CHCl₃ (500 ml)-isopropanol (150 ml) and then the organic phase was dried over Na₂SO₄, filtrated and concentrated under reduced pressure to obtain the title compound (27.6 g, 97%) as a colorless solid. FAB-MS *m/z* 178 (M+H)⁺ as C₁₀H₁₁NO₂; ¹H NMR (400 MHz, CDCl₃) δ 0.22–0.30 (m, 2 H), 0.60–0.67 (m, 2 H), 0.95–1.11 (m, 1 H), 2.68 (d, *J* = 7.1 Hz, 2 H), 7.48–7.54 (m, 1 H), 8.15–8.20 (m, 1 H), 8.58 (d, *J* = 5.1 Hz, 1 H).

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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