

Review

Regulation of the nongenomic actions of retinoid X receptor- α by targeting the coregulator-binding sites

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Retinoid X receptor- α (RXR α), a unique member of the nuclear receptor superfamily, represents an intriguing and unusual target for pharmacologic interventions and therapeutic applications in cancer, metabolic disorders and neurodegenerative diseases. Despite the fact that the RXR-based drug Targretin (bexarotene) is currently used for treating human cutaneous T-cell lymphoma and the fact that RXR α ligands (retinoids) show beneficial effects in the treatment of cancer and diseases, the therapeutic potential of RXR α remains unexplored. In addition to its conventional transcription regulation activity in the nucleus, RXR α can act in the cytoplasm to modulate important biological processes, such as mitochondria-dependent apoptosis, inflammation, and phosphatidylinositol 3-kinase (PI3K)/AKT-mediated cell survival. Recently, new small-molecule-binding sites on the surface of RXR α have been identified, which mediate the regulation of the nongenomic actions of RXR α by a class of small molecules derived from the nonsteroidal anti-inflammatory drug (NSAID) Sulindac. This review discusses the emerging roles of the nongenomic actions of RXR α in the RXR α signaling network, and their possible implications in cancer, metabolic and neurodegenerative disorders, as well as our current understanding of RXR α regulation by targeting alternate binding sites on its surface.

Keywords: RXR α ; retinoid; RXR α modulator; nongenomic action; coregulator site; apoptosis; inflammation; PI3K; NSAID

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Introduction

Retinoid X receptor-alpha (RXR α) belongs to a unique RXR subfamily of the nuclear receptor superfamily, which is encoded by 3 distinct genes: RXR α , RXR β , and RXR γ ^[1–9]. RXRs, like other nuclear receptors, consist of 3 distinct domains: a disordered N-terminal A/B region, a DNA-binding domain, and a C-terminal ligand-binding domain (LBD). The LBD possesses a canonical ligand-binding pocket (LBP), a transactivation function domain 2, a coregulator-binding surface groove, and a dimerization surface (Figure 1A). RXRs were initially identified as heterodimeric partners of the retinoic acid receptor (RAR), thyroid hormone receptor (T3R) and vitamin D receptor (VDR). Today, about one-third of the 48 human nuclear receptor superfamily members serve as RXR heterodimerization partners, including Nur77, peroxisome proliferator-activated receptors (PPARs), liver X receptor (LXR), and farnesoid X receptor (FXR)^[1–4, 6, 7]. In addition, RXR α can form homodimers^[10] and homotetramers^[11–13]

(Figure 1B), suggesting that RXR α may control its own specific signaling pathways. Binding of RXR α by a ligand regulates the ability of the receptor to dimerize and alters the receptor's cofactor-binding surface due to the rearrangement of helices 10, 11, and 12 (Figure 1C). Aside from its role in DNA binding and transactivation, accumulating evidence indicates that RXR α also has extranuclear functions^[14–18]. RXR α resides in the cytoplasm at different stages of development^[19]. It migrates from the nucleus to the cytoplasm in response to differentiation^[16], survival^[20, 21], apoptosis^[14], and inflammation^[17, 18, 20, 21]. 9-*cis*-retinoic acid (RA) was originally identified as a natural RXR α ligand. Subsequently, several dietary fatty acids were found to bind RXR α and to act as natural RXR α ligands (Figure 2). These include docosahexaenoic acid (DHA), oleic acid, and phytanic acid. However, none of these molecules has been proved to be the *bona fide* endogenous ligand of RXR α ^[22, 23]. Numerous natural products and synthetic compounds (retinoids) have been shown to bind to RXR α and to modulate its activities^[2–4, 24–26]. Thus, the heterodimerization capacity of RXR α together with the diversity of its ligands suggests that RXR α is an important regulator of a wide range of cellular pathways.

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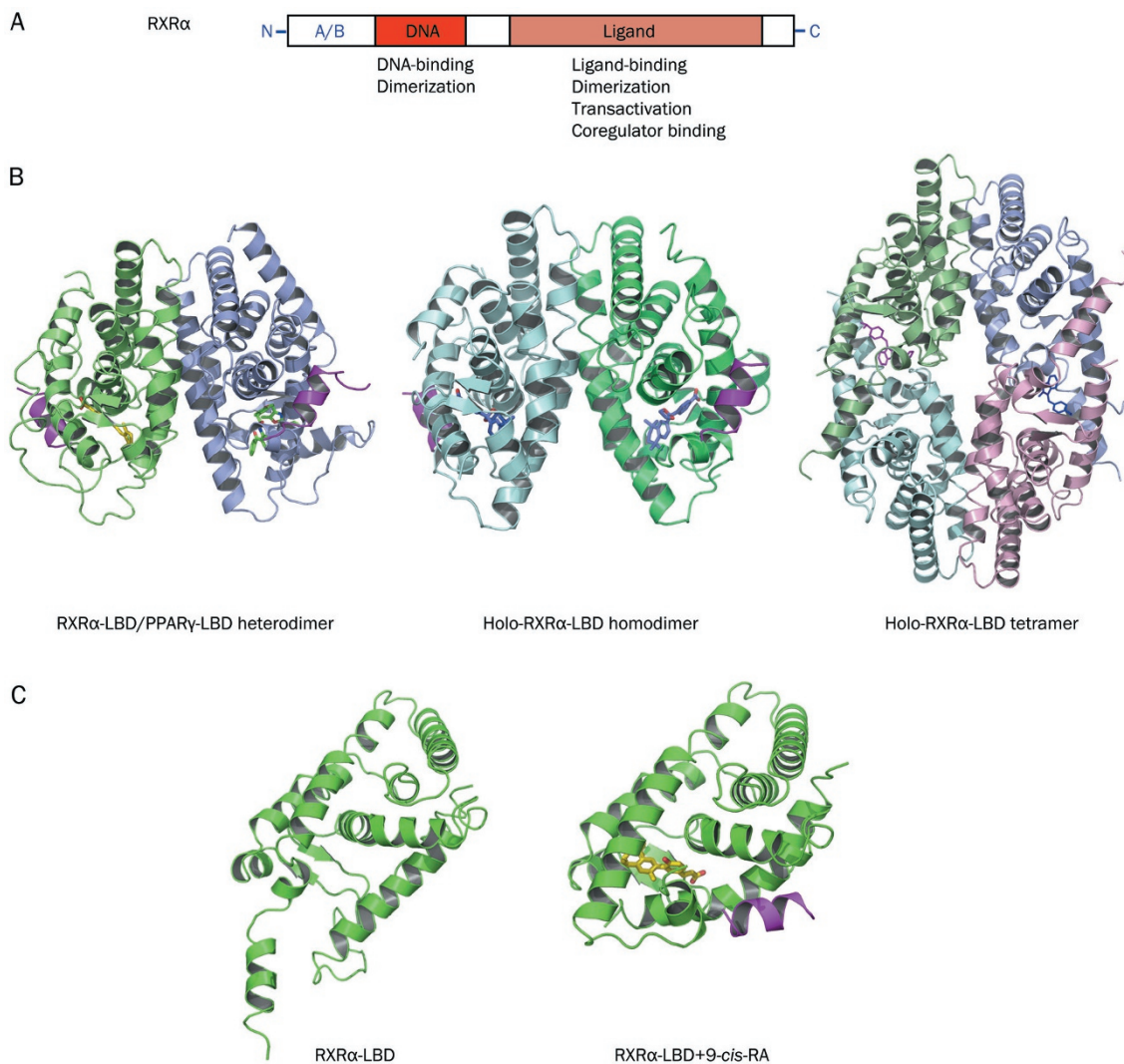
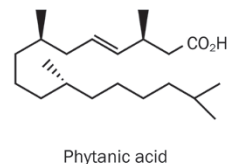
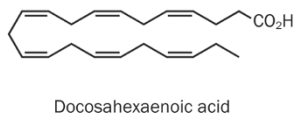
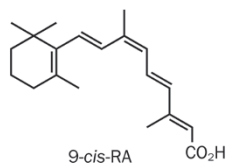
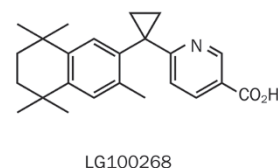
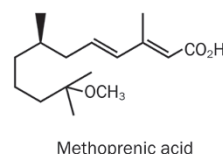
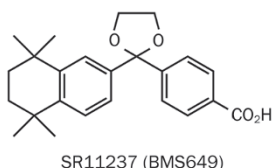
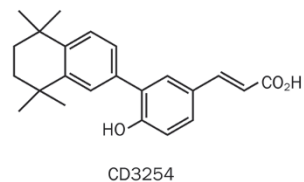
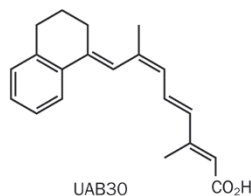
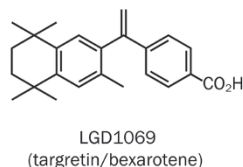
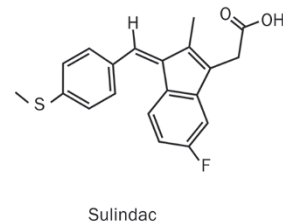
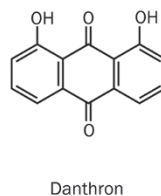
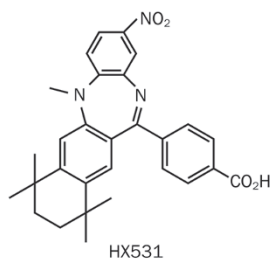
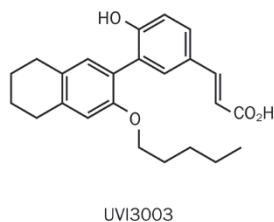


Figure 1. RXRα structure, homo- and hetero-dimerization, and effect of ligand. (A). Schematic representation of RXRα. (B). Structures of RXRα heterodimer, homodimer, and tetramer. Left, RXRα-LBD/PPARγ heterodimer, PDB code 1FM9. Middle, holo-RXRα-LBD homodimer, PDB code 1MZN. Right, holo-RXRα-LBD tetramer, PDB code 4N8R. (C). Structure of apo-RXRα and holo-RXRα. Left, monomer conformation in the apo-tetramer structure, PDB code 1G1U. Right, RXRα-LBD in complex with agonist CD3254 and coactivator GRIP1, PDB code 3FUG.

Genetic analysis demonstrated that RXRα is involved in a plethora of developmental and physiological pathways. A knockout of RXRα was embryonic lethal^[27]. Tissue-specific inactivation of RXRα in hepatocytes^[28], skin^[29], prostate^[30], or adipose tissue^[31] induces strong phenotypes, indicating a major role of RXRα in these tissues. The phenotypes observed in most RXRα-mutant mice may be related to alterations in pathways regulated by its heterodimerization partners. Structurally, RXRα homodimerization and heterodimerization can be separated by specific amino acid residues at the dimerization interfaces^[32, 33]. Ligand-activated RXRα homodimers up-regulate p21 expression through the direct binding of RXRα homodimers to the p21 promoter^[34]. Characterization of mice lacking RXRα in myeloid cells reveals an important role of RXRα homodimers in the innate immune response to inflammatory stimuli^[35]. Retinoids function as insulin sensi-

tizers and can decrease hyperglycemia and hypertriglyceridemia through an RXRα homodimer-mediated mechanism that is distinct from the one utilized by PPARγ in different mouse models^[35]. Consistent with this, a homodimer-specific RXRα agonist effectively lowers blood glucose in an animal model of insulin-resistant diabetes^[36]. Mechanistically, RXRα homodimers can selectively bind to functional PPAR response elements and induce transactivation *in vivo*^[37]. These observations underscore the importance of a very intricate RXRα signaling pathway for developing potential therapeutic uses of RXRα-specific modulators.

Altered expression and changes in the function of RXRα have been implicated in the development of a number of cancers and diseases. Although an RXRα knockout fetus dies in the embryonic stage^[27], targeted disruption of the RXRα gene leads to preneoplastic lesions in the prostate^[30], alopecia,

Natural RXR α agonistsSynthetic RXR α agonistsRXR α antagonistsFigure 2. RXR α ligands.

epidermal interfollicular hyperplasia, keratinocyte hyperproliferation and aberrant terminal differentiation in the skin^[29], the development of malignant cervical lesions^[38], alteration of fatty acid oxidation and hepatocyte lifespan in the liver^[28], and resistance to diet-induced obesity due to impaired adipocyte differentiation in adipose tissue^[31]. Diminished RXR α expression is also associated with the development of certain malignancies, which is largely attributed to proteolytic cleavage of RXR α in tumor cells^[15, 39–43]. In addition, alteration of RXR α function by phosphorylation is associated with the development of human cancer^[44]. Intriguingly, several studies have demonstrated that alteration of the subcellular localization of RXR α is implicated in the development of cancer and certain diseases. RXR α is translocated from the nucleus to the cytoplasm in response to endotoxin and other inflammatory mediators to inhibit its transactivation function^[17, 45], while an altered localization of RXR α to the splicing factor compartments occurs in highly malignant human breast cancer cells^[46]. We recently reported that an N-terminally truncated form of RXR α (tRXR α) produced in cancer cells resides in the cytoplasm to promote the growth of tumor cells^[21]. A recent finding that RXR α binding to PML/RAR α is required for the

development of acute promyelocytic leukemia in transgenic mice^[47, 48] further demonstrates the oncogenic potential of this protein when it functions inappropriately.

The pleiotropic action of RXR α under both physiological and pathophysiological conditions suggests that RXR α is an important target for pharmacologic interventions and therapeutic applications. This is highlighted by the FDA approval of the RXR-based drug Targretin (bexarotene) for treating T-cell lymphoma and its beneficial effects against other indications such as metabolic syndromes and neurodegenerative diseases. Targretin was found to induce a 50% overall inhibitory response in patients with refractory or persistent cutaneous T-cell lymphoma when administered either orally or topically^[49]. The therapeutic use of Targretin has been extended to other cancer types, including breast cancer and lung cancer^[2–4, 6]. Although a phase III clinical trial of Targretin for non-small cell lung carcinoma did not meet the end points, a subgroup of patients was shown to benefit from Targretin treatment^[50, 51]. Numerous studies have also reported the broad impact of rexinoids on metabolic regulation. Rexinoids improve insulin sensitivity, which is similar to the effect of thiazolidinedione (TZD), a PPAR γ ligand, and this is likely due to its activation of RXR α /

PPAR γ as well as a separate RXR signaling pathway. Rexinoids also provoke a very efficient inhibition of cholesterol absorption, and show beneficial effects on the development of atherosclerosis^[52]. Recently, Cramer *et al*^[53] reported that Targretin enhances apoE-dependent β -amyloid (A β) clearance from the brain and improves neural network function and reversal of behavioral deficits in mouse models of Alzheimer disease. This is exciting because there is currently no cure for Alzheimer disease. The effect of reducing soluble A β levels has been confirmed by several studies, although the reduction of A β plaques by Targretin remains controversial. Targretin also acts to prevent loss of dopaminergic neurons and restore behavioral function in rodent models of Parkinson's disease^[54], and it relieved positive symptoms of schizophrenia in a randomized, double-blind, placebo-controlled multicenter trial^[55]. Thus, RXR α -selective modulators are a class of very promising drug candidates for cancer, metabolic syndromes, and neurodegenerative disorders.

The promiscuous nature of RXR α has conferred rexinoids some unwanted side effects^[2-4, 6], which has hindered their further development. Thus, there is an urgent need to dissect RXR α signaling pathways and to identify and develop new RXR α modulators that have unique properties and improved therapeutic indexes. In this review, we focus on the nongenomic activity of RXR α and highlight recent advances in this field with an emphasis on tRXR α actions^[21] and RXR α modulation by targeting alternate binding sites on its surface^[56, 57].

Nongenomic activity of RXR and apoptosis

Apoptosis, programmed cell death, plays a central role both in development and in homeostasis, eliminating redundant cells and ensuring that cells that have migrated to their proper destinations survive^[58]. Abnormal regulation of apoptosis, as a result of either genetic anomalies and/or a persistent disease state, contributes to the establishment and progression of a number of human cancers and diseases, such as autoimmune and neurological disorders, inflammatory diseases, obesity, type 2 diabetes, and atherosclerosis. Apoptosis occurs following either the triggering of cell surface death receptors (the extrinsic pathway) or the perturbation of mitochondria (the intrinsic pathway)^[58]. The intrinsic pathway is initiated by the release of apoptogenic factors such as cytochrome *c* from mitochondria, while the extrinsic pathway involves the activation of the initiator caspase-8 through stimulation of death receptors of the tumor necrosis factor (TNF) receptor superfamily.

The role of RXR α and RXR α ligands in apoptosis was initially recognized by the finding that 9-*cis*-RA is a potent negative regulator of activation-induced T-cell apoptosis through its binding of both RXR and RAR^[59]. Subsequent studies demonstrated that rexinoids could either induce or promote apoptosis depending on the nature of the ligands and/or the cellular environment. 9-*cis*-RA inhibits activation-induced apoptosis in T-cell hybridomas and thymocytes by blocking the expression of Fas ligand following activation. RXR α has a protective role in cellular apoptosis of keratinocytes and melanocytes^[60], and RXR α antagonist HX531 inhibits the apoptotic

effect of 4-para-Nonylphenol in mouse embryonic neuronal cells through an RXR-mediated mitochondria-dependent signaling pathway^[61]. Activation of RXR α induces apoptosis in NB4 acute promyelocytic leukemia cells^[3]. Insulin-like growth factor binding protein (IGFBP)-3 binding to RXR α results in apoptosis of cancer cells^[62]. DHA induces apoptosis of colonocytes^[63] in an RXR α -dependent manner, while it promotes the survival of rat retina photoreceptors through RXR α -dependent activation of the mitogen-activated protein kinase (MAPK) signaling pathway^[64]. Targretin suppresses the progression of colonic adenomas to adenocarcinomas in animals, which is accompanied by the induction of apoptosis^[65], while R-etodolac binds to RXR α and induces RXR α -dependent apoptosis of prostate cancer cells *in vitro* and in animals^[66]. Together, the ability of rexinoids to positively or negatively regulate apoptosis likely contributes to their therapeutic effects in cancer, metabolic disorders, and neurodegenerative diseases.

One way that RXR α and its ligands regulate apoptosis is through their regulation of the Nur77-Bcl-2 apoptotic pathway through RXR α heterodimerization with Nur77^[67] (Figure 3). In response to several apoptotic stimuli, Nur77 migrates from the nucleus to the cytoplasm, where it targets mitochondria by interacting with Bcl-2^[68], leading to cytochrome *c* release and apoptosis. Nur77 mitochondrial targeting occurs not only in cancer cells, but also in other cell types such as CD4(+)CD8(+) thymocytes^[69], cardiomyocytes^[70], and cerebellar granule neurons^[71]. RXR β can cotranslocate with Nur77 from the nucleus to the cytoplasm as a heterodimer in PC12 cells in response to nerve growth factor (NGF) treatment^[16]. We reported that RXR α serves as an active partner in shuttling Nur77 from the nucleus to mitochondria in cancer cells^[14]. The shuttling of the Nur77/RXR α heterodimers between the nucleus and the cytoplasm is subject to regulation by RXR α ligands. 9-*cis*-RA suppresses apoptosis by inhibiting Nur77/RXR mitochondrial targeting^[14]. Such regulation of Nur77 activity by 9-*cis*-RA may account for its inhibitory effect on apoptosis. 9-*cis*-RA is known to potently inhibit the activation-induced apoptosis of T cells and thymocytes^[72], in which Nur77 plays a role. It is worth noting that 9-*cis*-RA is able to induce RXR-mediated nucleo-cytoplasmic shuttling of Nur77 and its translocation to mitochondria for apoptosis^[73] and it can relieve the inhibitory effect of RXR α on EGF-induced Nur77 nuclear accumulation^[74], suggesting that certain RXR α ligands may act to promote RXR nuclear export and apoptosis under certain cellular conditions.

RXR α and ligands can also modulate the extrinsic apoptotic pathway. TNF α is a multifunctional cytokine that controls diverse cellular events such as cell survival and death that control the destiny of cancer cells^[75]. The death effect of TNF α is mediated by the recruitment of TNF receptor-associated death domain (TRADD) and Fas-associated death domain (FADD) to TNF α receptor TNF-R1, which then recruits caspase-8, a key initiator of apoptosis. Although less characterized, TNF-R1 also recruits phosphatidylinositol 3-kinase (PI3K) to activate the PI3K/AKT survival pathway^[76]. We found that tRXR α

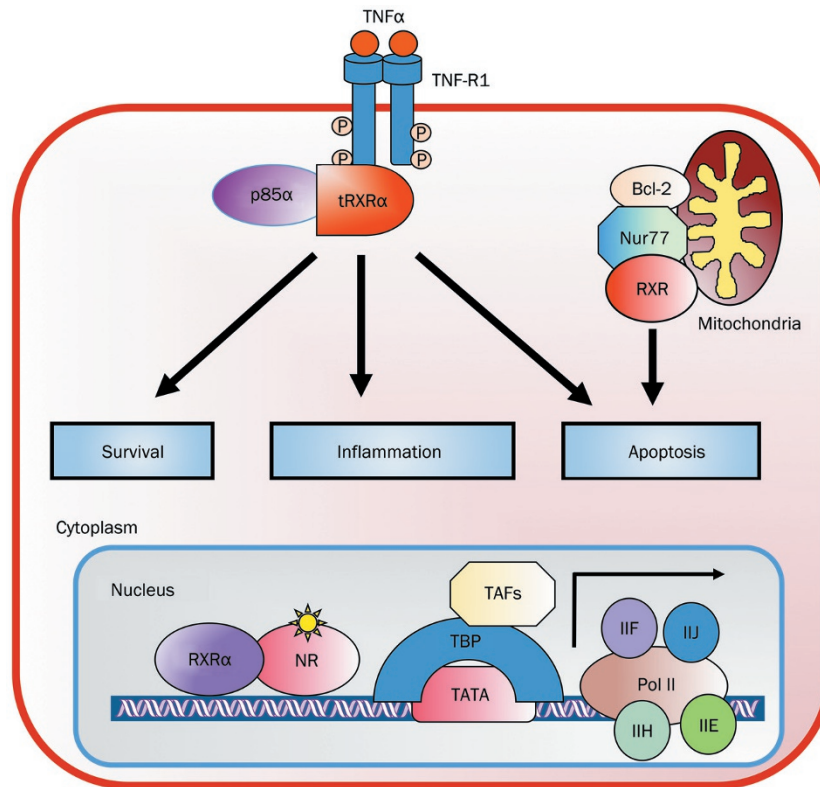


Figure 3. The nongenomic RXR α actions. The cytoplasmic tRXR α through its interaction with p85 α subunit of PI3K or undefined factors associated with TNF-R1 regulates cell survival, inflammation, and apoptosis. In addition, RXR α can target mitochondria through heterodimerization with Nur77 to modulate mitochondria-dependent apoptosis.

could bind to the p85 α regulatory subunit of PI3K in response to TNF α treatment, leading to activation of the PI3K/AKT pathway^[21]. This finding implies that rexinoids could act nongenomically to modulate the TNF α -dependent extrinsic apoptotic pathway. Indeed, inhibition of tRXR α binding to p85 α by Sulindac (also called CLINORIL[®]), a nonsteroidal anti-inflammatory drug (NSAID) currently used for treating pain and inflammation, and analogs, results in caspase-8-dependent apoptosis^[21]. Several natural products, including CF31, can activate this death pathway through direct binding to tRXR α ^[20]. Although the apoptotic effect of Sulindac and analogs can be attributed to their inhibition of tRXR α -dependent activation of PI3K/AKT, it remains to be seen whether tRXR α is directly involved in the formation of the TNF-R1-TRADD-FADD apoptosome to modulate the extrinsic apoptotic pathway.

Nongenomic action of RXR α and inflammation

Like other nuclear receptors, RXR α and its ligands regulate diverse aspects of immunity and inflammation. The Karpen laboratory showed that inflammatory mediators decrease the nuclear levels of RXR α and its transactivation in a c-Jun N-terminal kinase (JNK)-dependent manner^[17], suggesting a role for RXR α and its ligands in inflammation. Accumulating evidence has now revealed their active role in the modulation of inflammatory responses and immunity. Acute challenge with

AOM/DSS induces colitis in RXR α heterozygous mice with increased inflammatory marker expression^[77], and RXR α is highly expressed in macrophages^[7]. Consistent with this, certain anti-inflammatory agents serve as RXR α ligands, implying that RXR α may be an intracellular target that mediates the anti-inflammatory effects of these agents. DHA induces growth inhibition and apoptosis by inhibiting NF- κ B activity^[78] and suppressing cytokine production in macrophages^[79]. The NSAID R-etodolac, which induces RXR α -dependent apoptosis of tumor cells^[66], decreases constitutive and RANKL-stimulated NF- κ B activation in macrophages and suppresses TNF α -induced IKK phosphorylation and subsequent NF- κ B activation^[80]. The role of RXR α is further implicated by numerous studies showing that rexinoids are critical regulators of various inflammatory pathways in different cell types, including T cells^[81], macrophage^[82], dendritic cells^[83], and microglia and astrocytes^[84]. Targretin downregulates COX-2 expression in breast cancer cells^[85], inhibits angiogenesis and metastasis in solid tumors^[86], reduces the expression of TNF α and IL-1 β protein in *Apc(Min/+)* mice^[65], and suppresses inflammation in patients with plaque-type psoriasis^[87]. Rexinoids are also protective against colon inflammation that is induced by 2,4,6-trinitrobenzene sulfonic acid^[88]. RXR α antagonists are capable of altering the maturation process from human monocytes to dendritic cells in response to TNF α or lipopolysaccharide (LPS)^[83], and block T Helper 2 cell differentiation and IL-5

production in T cells^[89]. Thus, the diverse anti-inflammatory effects of RXR α and its ligands in various cell types underscore their function in the prevention and treatment of inflammatory and metabolic disorders, such as cancer, atherosclerosis, insulin resistance, autoimmunity, and neurodegeneration.

The mechanisms by which RXR α and its ligands modulate inflammation and immunity remain an important unanswered question that is currently being actively investigated. Both genomic and nongenomic actions of RXR α could account for its modulation of inflammation in macrophages and other cell types. For genomic action, the most potent anti-inflammatory effects of RXR α appear to result from protein-protein interactions between RXR α and pro-inflammatory transcription factors, particularly NF- κ B and AP-1, through the trans-repression mechanism, which has been reviewed elsewhere^[90]. The nongenomic mechanisms of RXR α may involve inhibition of the activation/phosphorylation of JNK and subsequent phosphorylation of c-Jun^[91]. Interestingly, the subcellular localization of RXR α is altered in response to inflammation^[17, 45]. LPS alters the subcellular location of RXR α in animals^[17], while RXR α undergoes rapid nuclear export in response to IL-1 β in hepatoma cells^[18]. The effect is rapid, occurring within 30 min of exposure to IL-1 β , and is likely due to RXR α phosphorylation by JNK and through a CRM-1-mediated nuclear export process^[18]. IL-1 β , IL-6, and TNF α also alter the intracellular distribution of RXR in Schwann cells, which occurs when cells are exposed to cytokine for as little time as 5 minutes^[45].

Our recent discovery that TNF can induce cytoplasmic localization of tRXR α underscores the significance of tRXR α cytoplasmic action in the regulation of inflammation (Figure 3). As discussed above, TNF α is a cytokine that induces not only the extrinsic apoptotic and PI3K/AKT pathways but also the NF- κ B and AP-1 inflammatory pathways. In this regard, TNF α receptor-1 (TNF-R1) recruits TNF receptor-associated factor 2 (TRAF-2) and receptor-interacting protein (RIP) kinase, which results in the initiation of pathways that culminate in the activation of transcription factors NF- κ B, c-Jun, c-Fos, and ATF-2 via the activation of various kinases including I κ B kinase (IKK) and MAPKs. These pathways control the inducible expression of genes important for inflammation. It remains to be seen whether tRXR α or other forms of RXR α are directly involved in the activation and regulation of the inflammasome. Intriguingly, TNF α induction of tRXR α -dependent responses is inhibited by Sulindac, which is currently used for treating inflammation, implying that the drug may exert its anti-inflammatory effects by targeting tRXR α pathways.

Nongenomic action of RXR and the PI3K/AKT survival pathway

The role of PI3K/AKT activation in oncogenesis and drug resistance has been validated by multiple studies, demonstrating that aberrations in this pathway are potential causes of cell transformation, metabolic disorders, and neurodegenerative diseases, as well as drug resistance^[92]. The pathway has therefore been targeted extensively for the development of thera-

peutics against cancer and related diseases, and for overcoming drug resistance. However, current targeting strategies that rely on direct inhibition of PI3K/AKT activities have caused profound adverse events and have thus far been confined to preclinical and clinical evaluation due to toxicity and lack of selectivity. Thus, identification of key molecules involved in the aberrant activation of PI3K/AKT pathway will offer new strategies for drug development.

We recently reported that tRXR α , but not the full-length RXR α , could act to mediate TNF α activation of PI3K/AKT in a number of cancer cell lines^[21] (Figure 3). The tRXR α protein is detected in a variety of cancer cell lines and in primary tumors, but not in tissues surrounding the tumor or in distant normal tissues from the same cancer patients^[21], suggesting its oncogenic potential. Our finding that tRXR α , but not RXR α , acts nongenomically to interact with p85 α indicates that tRXR α acquires a new function that is different from that of the full-length RXR α protein. Such activation or conversion of a protein's phenotype by limited proteolytic cleavage is not without precedent. Limited proteolytic processing of RXR α occurs in many types of cancer cells^[15, 39-43], suggesting that it may represent an important mechanism that regulates the biological activity of RXR α . Regulated proteolysis is a key step in a number of different signaling pathways that respond to developmental cues or external stimuli. Caspase-mediated cleavage of the BH3-only protein Bid into a truncated protein (tBid) and subsequent translocation of tBid to mitochondria is implicated in death receptor signaling^[93], whereas proteolytic processing of Notch and nuclear translocation of the truncated product is a crucial step in transduction of Notch signaling^[94]. Cleavage of the androgen receptor by calpain produces a truncated receptor protein that may play a role in the development of androgen-independent prostate cancer^[95]. An intriguing question regarding tRXR α -mediated activation of PI3K/AKT relates to the proteases responsible for RXR α cleavage. Our recent study^[96] identified calpain II as one of the proteases that can cleave RXR α protein *in vitro* and *in vivo*. Activation of calpain II by ionomycin enhances the production of tRXR α in cancer cells, which is regulated in a glycogen synthase kinase 3 beta (GSK-3 β)-dependent manner^[96]. However, proteases other than calpain II are likely involved in the cleavage of RXR α and remain to be identified.

Many more important questions remain regarding the nongenomic regulation of the PI3K/AKT pathway by RXR α and its ligands. Unlike the full-length RXR α that resides in the nucleus, tRXR α is cytoplasmic and interacts with the p85 α subunit of PI3K to activate the PI3K/AKT survival pathway and to induce anchorage-independent cell growth *in vitro* and cancer cell growth in animals (Figure 3). It is unclear whether the cytoplasmic localization of tRXR α results from its nuclear export or cytoplasmic retention due to its interaction with cytoplasmic proteins such as p85 α . In either case, the N-terminal region that is deleted from RXR α is expected to play a critical role in regulating RXR activities. As the N-terminal region of RXR α is subject to regulation by phosphorylation, it remains to be determined whether phosphorylation or other

modifications of RXR α are involved in the regulation of RXR α cytoplasmic localization and its interaction with p85 α . How tRXR α interacts with p85 α is also currently unknown. Several nuclear receptors including RAR, PPAR, and T3R have been shown to interact with p85 α , implying the existence of a more general mechanism for their interaction. Nevertheless, our results reveal a nongenomic regulation of the PI3K/AKT signaling pathway by tRXR α , which provides not only an explanation for abnormal activation of the pathway in cancer cells but also new strategies to inhibit the activation of PI3K/AKT in cancer cells by targeting tRXR α . Such tRXR α -based PI3K/AKT inhibitors are likely more specific and tumor selective than conventional PI3K/AKT inhibitors.

TNF α controls diverse cellular events such as cell survival and death that determine the destiny of cancer cells^[75]. Although TNF α is capable of inducing the apoptosis of cancer cells through death receptor-dependent mechanisms, such an effect is often antagonized by TNF α 's own survival function through its activation of NF- κ B and PI3K/AKT pathways^[75]. Since TNF α is produced by malignant or host cells in the tumor microenvironment but not in normal cells, there has been tremendous interest in developing strategies to shift TNF α signaling from survival to death. Sulindac and its K-80003 and K-8008 analogs can bind to tRXR α to inhibit the TNF α -induced interaction of tRXR α with p85 α and the activation of PI3K/AKT, resulting in the activation of the TNF α -dependent apoptotic pathway^[21]. Thus, binding of tRXR α by Sulindac and analogs could convert TNF α signaling from survival to death. It is anticipated that many RXR α modulators exert their therapeutic effect by targeting this pathway.

Novel surface binding sites of RXR α as alternate sites for targeting

Canonical ligands bind to the LBP to directly mediate transcriptional activity, and so identifying and optimizing molecules that bind to RXR α 's canonical LBP have so far been the focus of drug discovery efforts targeting RXR α . However, there are key limitations of treatment with rexinoids, including unwanted side effects such as an increase in plasma triglyceride levels, suppression of the thyroid hormone axis, and induction of hepatomegaly. Therefore, targeting alternate sites on RXR α for regulating its activities could become a new strategy for RXR α -based drug discovery. Compounds that bind to alternate sites have been successfully demonstrated for other nuclear receptors^[97-99], including estrogen receptor, androgen receptor, VDR and T3R. Among the reported alternate sites on nuclear receptors, the coregulator-binding site is the most studied. Recently, by employing a docking-based virtual screening approach, we identified some small molecules that bind to the coregulator-binding surface of RXR α , a region where the binding sites of the corepressor and the coactivator overlap (Figure 4A). One of the identified binders, **23**, can regulate the biological functions of tRXR α , including inhibition of TNF α -induced interaction of tRXR α with p85 α , inhibition of AKT activation *in vitro* and in animals, and induction of apoptosis^[56]. Compound **23** doesn't bind to the LBP and rep-

resents the first example of an RXR α modulator that acts via the coregulator-binding site rather than the classical ligand-binding pocket. Thus, targeting alternate binding sites on the surface of RXR α for therapeutic intervention may become a new paradigm for nuclear receptor-based drug discovery.

In addition to the coregulator-binding groove, another alternate binding site was identified on the surface of RXR α . Our recently determined crystal structure of the RXR α LBD in complex with the Sulindac analogs K-8008 or K-8012^[57] demonstrates the existence of a different binding site. The complex structure exists as a noncrystallographic homo-tetramer similar to the previously reported apo-homotetramer^[11, 13], in which the bottoms of 2 homodimers interface to form a tetramer (Figure 4B). In a tetramer, 2 K-8008 molecules bind to one homotetramer at a hydrophobic region that is near the entry and the edge of the cognate LBP^[57]. The K-8008 binding region is close to the dimer-dimer interface and does not overlap with the binding region of 9-*cis*-RA. With respect to the monomeric and the dimeric RXR α LBD, the K-8008 binding region is located on the surface of the RXR α molecules. RXR α has been shown to form homo-tetramers in solution, which is transcriptionally silent, but to rapidly dissociate into active homodimers upon binding of agonists or antagonists^[11-13]. Therefore, it is intriguing that K-8008, an RXR α antagonist, binds to a novel region and that the binding does not result in dissociation of the tetramer, similar to binding by danthron^[100]. The structural basis of K-8008 binding suggests that RXR α tetramerization represents a key mechanism for the regulation of the nongenomic actions of RXR.

Conclusions and perspectives

It is evident that both genomic and nongenomic mechanisms contribute to the pleiotropic effects of RXR α and its ligands. Recent advances have revealed the roles of the nongenomic actions of RXR α and its ligands in the control of apoptosis, survival, and inflammation, which likely account for their therapeutic effects in cancer and metabolic and neurodegenerative disorders, although their physiological and pathophysiological relevance remains to be fully established. The mechanisms that regulate the nongenomic actions of RXR α need to be further elucidated. Despite the recognition that RXR α is an innovative drug target, development of RXR α -based drugs has been hampered by the side effects associated with targeting its cognate LBP. The findings that RXR α is cleaved in tumor cells and that Sulindac-derived small molecules and others act at the alternate binding sites of the surface of RXR α will provide new rational drug design and screening approaches by targeting functionally important surface-binding sites. Such an approach will likely target tumor- or disease-selective RXR α (*ie*, tRXR α or RXR α with abnormal modifications) rather than unmodified RXR α and may also circumvent the side effects associated with binding to the cognate RXR α LBP. However, many unanswered questions regarding the production, function, and the underlying mechanisms of tRXR α need to be answered. The binding of Sulindac analogs to the tetrameric form of the RXR α -LBD is interesting. However, little is known

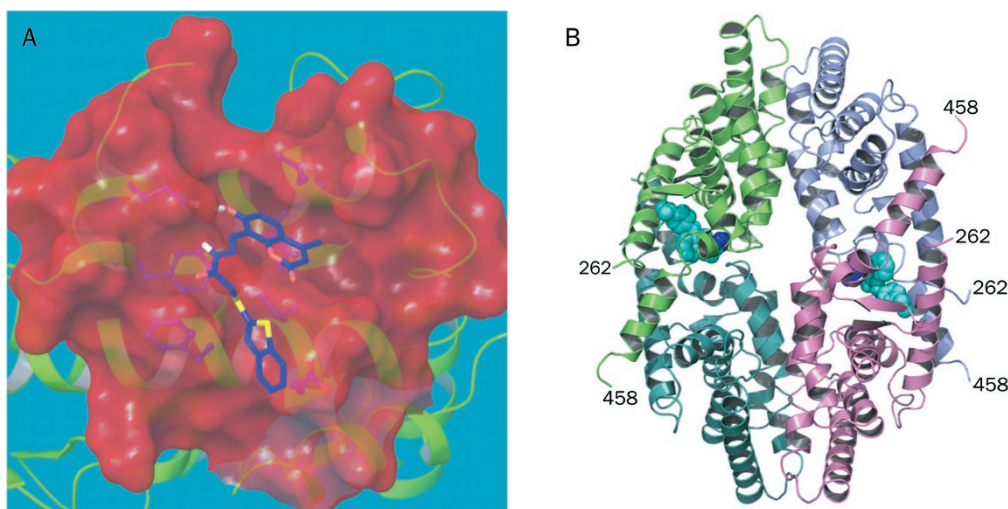


Figure 4. Alternate sites on the surface of RXR α . (A) The recently identified compound **23** bound to the coregulator-binding groove of RXR α , a docking model. (B) The newly identified site for K-8008 binding, PDB code 4N8R.

about the biological function of the RXR α tetramer with respect to the regulation of the nongenomic function of RXR α . The characterization of the surface binding sites in RXR α and the development of selective inhibitors targeting the surface-binding sites may support a departure from the traditional paradigm of targeting the LBP.

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Abbreviations

RXR α , retinoid X receptor- α ; tRXR α , truncated retinoid X receptor- α ; PI3K, phosphatidylinositol 3-kinase; NSAID, nonsteroidal anti-inflammatory drug; LBD, ligand-binding domain; LBP, ligand-binding pocket; RAR, retinoic acid receptor; T3R, thyroid hormone receptor; VDR, vitamin D receptor; PPAR, peroxisome proliferator-activated receptor; LXR, liver X receptor; FXR, farnesoid X receptor; RA, retinoic acid; DHA, docosahexaenoic acid; TZD, thiazolidinedione; A β , β -amyloid; TNF, tumor necrosis factor; IGF1BP, insulin-like growth factor binding protein; MAPK, *mitogen-activated protein kinase*; NGF, nerve growth factor; TRADD, TNF receptor-associated death domain; FADD, Fas-associated death domain; TNF-R1, TNF α receptor-1; JNK, c-Jun N-terminal kinase; LPS, lipopolysaccharide; GSK-3 β , glycogen synthase kinase 3 beta.

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