

Brazilian Journal of Pharmacognosy revista brasileira de farmacognosia

www.elsevier.com/locate/bjp

Staphylococcus aureus ATCC 25923 and Bacillus subtilis ATCC 6633.



CrossMark

# Original Article

## Constituents from the bark resin of Schinus molle

Gonzalo Rodolfo Malca-García<sup>a,\*</sup>, Lothar Hennig<sup>a</sup>, Mayar Luis Ganoza-Yupanqui<sup>b,\*</sup>, Alejandro Piña-Iturbe<sup>c</sup>, Rainer W. Bussmann<sup>d</sup>

ABSTRACT

<sup>a</sup> Institut für Organische Chemie, Fakultät für Chemie und Mineralogie, Universität Leipzig, Leipzig, Germany

<sup>b</sup> Departamento de Farmacología, Facultad de Farmacia y Bioquímica, Universidad Nacional de Trujillo, Trujillo, Peru

<sup>c</sup> Escuela de Microbiología y Parasitología, Facultad de Ciencias Biológicas, Universidad Nacional de Trujillo, Trujillo, Peru

<sup>d</sup> William L. Brown Center, Missouri Botanical Garden, St. Louis, United States

#### ARTICLE INFO

Article history: Received 13 June 2016 Accepted 21 July 2016 Available online 15 September 2016

Dedicated to Prof. Dr. Lothar Beyer on the occasion of his 80th birthday.

Keywords: Schinus molle Resin Terpenes Cytotoxic activity MIC

Introduction

The family Anacardiaceae is of pantropical occurrence and includes a few representatives in temperate regions. The family comprises about 70 genera and 600 species. Many of them are used traditionally as healing, stomachic and antidiarrheal agents, due to the presence of tannins and oil resins (Duarte et al., 2006). In this family. Schinus molle L. (also known as California pepper and pink pepper) was introduced from South America to most tropical and subtropical areas of the world, as well as the Mediterranean (Rhouma et al., 2009). In Peruvian traditional medicine S. molle is used as antibacterial, topical antiseptic, digestive and purgative diuretic (Duke, 2002), for toothache, wound healing, rheumatism, and menstrual disorders, as well as stimulant and antidepressant (Machado et al., 2007), for respiratory and urinary infections (Perez and Anesini, 1994), and as analgesic and central depressant (Barrachina et al., 1997). Extracts of S. molle showed promising antitumoral effects and cytotoxic activity (IC<sub>50</sub> 50  $\pm$  7  $\mu$ g/ml) against a human hepatocellular carcinoma Cell Line, Hep G was also reported (Ruffa et al., 2002). Recent studies revealed that S. molle essential

\* Corresponding authors.

E-mails: gonzalo.malca@uni-leipzig.de

oil was cytotoxic in several cell lines, and it was more effective on breast carcinoma and leukemic cell lines (Díaz et al., 2008).

Extensive investigations of the fruit and leaf essential oils of *S. molle* identified a broad variety of constituents (Abdel-Sattar et al., 2010; Bendaoud et al., 2010), showing that the ingredients are similar, but with differences in their percentage depending on the region in which they are grown. Main components are monoterpene hydrocarbons and oxygenated monoterpene hydrocarbons. No reports were found concerning isolation and characterization of secondary metabolites from the bark resin. Therefore, we decided to concentrate our effort in *S. molle* resin.

#### Material and methods

#### NMR and MS infrastructure and methods

A total of five terpenes was isolated from the bark resin of Schinus molle L., Anacardiaceae, and their

structures were determined by spectroscopic techniques. Among these compounds the sesquiterpene

hydrocarbon terebinthene showed significant growth inhibitory activity against human colon carcinoma

HCT-116 cells. Furthermore, terebinthene and pinicolic acid (5) also showed antibacterial activity against

© 2016 Sociedade Brasileira de Farmacognosia. Published by Elsevier Editora Ltda. This is an open

access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

NMR spectra (<sup>1</sup>H, <sup>13</sup>C, APT, NOESY1D, *H*,*H*-COSY, edited HSQC, and HMBC) were recorded on a Varian Mercury 400 plus (400 MHz for <sup>1</sup>H, 100 MHz for <sup>13</sup>C) and a Varian Mercury 300 plus (300 MHz for <sup>1</sup>H, 75 MHz for <sup>13</sup>C) spectrometer, respectively, at 26 °C and with CDCl<sub>3</sub> as a solvent. The chemical shifts were reported relative to the residual solvent peak, used as an internal reference (<sup>1</sup>H 7.26 ppm, <sup>13</sup>C 77.16 ppm). Chemical shifts are given in  $\delta$  values, coupling constants *J* in Hz.

http://dx.doi.org/10.1016/j.bjp.2016.07.004

0102-695X/© 2016 Sociedade Brasileira de Farmacognosia. Published by Elsevier Editora Ltda. This is an open access article under the CC BY-NC-ND license (http:// creativecommons.org/licenses/by-nc-nd/4.0/).

<sup>(</sup>G.R. Malca-García), mganoza@unitru.edu.pe (M.L. Ganoza-Yupanqui).

#### Plant material

The resin bark of "molle" was collected in March 2009 from Trujillo-Peru and the species was identified as *Schinus molle* L., Anacardiaceae, by Eric F. Rodríguez Rodríguez at Herbarium Truxillense (HUT), National University of Trujillo, Peru. A voucher specimen under No. 58335 (HUT) documenting the collection was deposited at Herbarium Truxillense (HUT) in Peru.

#### Extraction and isolation

Spot tests were used for the qualitative determination of secondary metabolites present in the resin of *S. molle* (Dominguez, 1973; Harborne, 1984). We identified small amounts of steroids and triterpenoids (dark green) by the Liebermann-Burchard's test, and flavonoids (red) by the Shinoda's test. No alkaloids were detected using Dragendorff's test.

The resin of *S. molle* (10.14g; still containing pieces of wood and insoluble materials) was extracted with CH<sub>2</sub>Cl<sub>2</sub> and hexane (250 ml each). Both extracts were combined yielding 2.63 g after removal of the solvents. This residue was fractionated by column chromatography on silica gel eluting with hexane, called fraction 1 (0.92 g) and by using CH<sub>2</sub>Cl<sub>2</sub> system to give fraction 2 (1.50 g). Fraction 1 was subjected to column chromatography with hexane/CH<sub>2</sub>Cl<sub>2</sub> (100:0  $\rightarrow$  99.5:0.5  $\rightarrow$  99:1) system to give germacrene D (1) (264 mg; *R*<sub>f</sub>: 0.81) and terebinthene (**2**) (18.4 mg; *R*<sub>f</sub>: 0.91).

Fraction 2 was subjected to column chromatography with the eluent system CH<sub>2</sub>Cl<sub>2</sub>/EtOAc (1:0  $\rightarrow$  2:0.1  $\rightarrow$  3:0.1  $\rightarrow$  4:0.2  $\rightarrow$  5:0.3) to furnish isomasticadienonalic acid (**4**) (120 mg;  $R_f$  0.65) and to CH<sub>2</sub>Cl<sub>2</sub>/EtOAc (5:0.3  $\rightarrow$  3:1  $\rightarrow$  2:1.5  $\rightarrow$  1:2) resulting in the isolation of isomasticadienoic acid (**3**) (336 mg;  $R_f$ : 0.6) and pinicolic acid (**5**) (18.6 mg;  $R_f$ : 0.2).

#### Bioactivity assays

#### Cytotoxicity screen

The human HCT-116 colon carcinoma cell line (ACC-581) was obtained from the German Collection of Microorganisms and Cell Cultures (Deutsche Sammlung für Mikroorganismen und Zellkulturen, DSMZ) and was cultured under conditions recommended by the depositor. Cells were seeded at  $6 \times 10^3$  cells per well of 96-well plates in 180 µl complete medium (90% McCoy's 5A+10% FBS) and treated with sesquiterpene hydrocarbon terebinthene (2) in serial dilution after 2 h of equilibration. Terebinthene (2) was tested in duplicate as well as the internal solvent control. After 5 d incubation, 20 µl of 5 mg/ml MTT (3-(4,5-dimethylthiazol-2yl)-2,5-diphenyltetrazolium bromide) in PBS (phosphate-buffered saline; pH 7.4) was added per well and it was further incubated for 2h at 37 °C. The medium was then discarded and cells were washed with 100 µl PBS before adding 100 µl 2-propanol/10 N HCl (250:1) in order to dissolve formazan granules. The absorbance at 570 nm was measured using a microplate reader (Tecan Infinite M200Pro), and cell viability was expressed as percentage relative to the respective solvent control. IC<sub>50</sub> values were determined by sigmoidal curve fitting and values represent the average  $\pm$  SD of two independent measurements.

#### Bacteria strains assay

The strains used were *Staphylococcus aureus* ATCC 25923, *Bacillus subtilis* ATCC 6633, *Escherichia coli* ATCC 25922 and *Pseudomonas aeruginosa* (isolated at the Hospital Regional Docente de Trujillo, Peru). The bacterial cultures were grown in Mueller-Hinton agar media incubated at 37 °C for 24 h following literature (CLSI, 2012).

Agar-well diffusion methods: The agar-well with diameter of 5.5 mm were inoculated to a concentration equivalent to 0.5 McFarland. Compounds **2** and **5** were inoculated at  $40 \,\mu$ l in DMSO, at concentrations of 5 and  $10 \,mg/m$ l. After bacterial diffusion for  $10 \,min$ , those petri plates were incubated at  $37 \,^{\circ}$ C for 22 h.

#### Broth macrodilution MIC (minimal inhibition concentration)

Dry extract  $CH_2Cl_2$  (**3**–**5**) was diluted in DMSO, tween 80 and water in ratio 2:1:3 according to the following concentrations: 0.062, 0.125, 0.25, 0.5, 1.0, 2.0, 4.0, 8.0 and 16.0 mg/ml. Each MIC test was repeated twice and all tubes were incubated for 22 h at 37 °C. After incubation MIC results were determined as minimum concentration which showed no visible bacterial growth. Then 50  $\mu$ l of resazurin sodium salt (Sarker et al., 2007) were added to each tube to a final concentration of 0.02%, and the samples were incubated at 37 °C for 1 h. MIC was confirmed as minimum concentration when no color change of the resazurin sodium salt (from blue to pink) happened.

#### **Results and discussion**

The structure of all five already known compounds germacrene-D (**1**), terebinthene  $[(6R^*, 8R^*)$ -9-spiro(cyclopropa)-2,4,4,8-tetramethylbicyclo[4.3.0]non-1-ene(**2**)], isomasticadienoic acid (**3**), isomasticadienonalic acid (**4**), and pinicolic acid [3-oxo-5 $\alpha$ -lanosta-8,24-diene-21-oic acid (**5**)] was determined after careful purification of the compounds using spectroscopic methods and comparison of the data with literature results.



Germacrene *D* is a biogenetic precursor of many other sesquiterpene structures (Bülow and König, 2000), terebinthene was isolated from *S. terebinthifolius* (Richter et al., 2010), the triterpenoid acids **3** and **4** were found in the berries or the stem exudate of *S. molle* (Pozzo-Balbi et al., 1978; Abdel-Sattar et al., 2007) and **5** was identified as anti-inflammatory triterpenoid from *Ganoderma* genus (Ko et al., 2008). Surprisingly, there are no reports in literature for sesquiterpene hydrocarbon **2** as an ingredient of *S. molle*, but **2** was isolated for the first time from *S. terebinthifolius* (Richter et al., 2010). Compound **5** was not found to be active against any type of cancer, but induced platelet aggregation (Mosa et al., 2011).

Terebinthene (2):  $(6R^*, 8R^*)$ -9-spiro(cyclopropa)-2,4,4,8-tetramethylbicyclo[4.3.0]non-1-ene: <sup>1</sup>H NMR (400.1 MHz, CDCl<sub>3</sub>):  $\delta$ =0.49 (1H, m, H-10a), 0.53 (1H, m, H-11a), 0.60 (1H, m, H-10b), 0.69 (3H, s, CH<sub>3</sub>-15), 0.71 (1H, s, H-11b), 1.00 (1H, m, H-7a), 1.06 (3H, s, CH<sub>3</sub>-13), 1.08 (1H, H-5a), 1.10 (3H, s, CH<sub>3</sub>-14), 1.28 (3H, m, CH<sub>3</sub>-12), 1.63 (1H, m, J=18.5 Hz, H-5b), 1.83 (1H, m, H-7b), 2.00 (1H, m, H-8), 2.12 (1H, m, H-3a), 2.12 (1H, m, H-3b), 2.65 (1H, br, H-6); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$ =6.33 (C-10), 7.36 (C-11), 13.00 (C-12), 17.29 (C-15), 24.46 (C-9), 29.66 (C-13), 30.70 (C-14), 34.24 (C-8), 37.15 (C-4), 38.27 (C-7), 41.33 (C-6), 45.80 (C-3), 48.57 (C-5), 126.04 (C-2), 139.25 (C-1).

Table T	Ta	bl	e	1
---------	----	----	---	---

Diameters of inhibition zones at concentrations of 5 and 10 mg/ml.

Compounds	Concentration (mg/ml)	Diameter (mm)			
		S. aureus ATCC 25923	B. subtilis ATCC 6633	E. coli ATCC 25922	P. aeruginosa
2	10	9.3	9.3	0	0
	5	8.0	8.6	0	0
5	10	7.1	9.0	0	0
	5	6.8	8.8	0	0

Table 2

MIC from the bark resin of Schinus molle.

Resin	MIC		
	S. aureus ATCC 25923	B. subtilis ATCC 6633	
CH <sub>2</sub> Cl <sub>2</sub> extract	8.0 mg/ml	0.125 mg/ml	

### Cytotoxicity on HCT-116 cells

The *in vitro* cytotoxicity of **2** was evaluated in a tetrazolium salt-based assay (MTT: 3-(4,5-dimethylthiazol-2-yl)-2,5diphenyltetrazolium bromide) against human HCT-116 colon carcinoma cells. Compound **2** displayed a moderate inhibition of HCT-116 cells with IC<sub>50</sub> 14.23  $\pm$  1.3 µg/ml.

#### Antibacterial activity

Additionally, we determined the antibacterial activity of compounds **2** and **5** by screening against *Staphylococcus aureus* ATCC 25923, *Bacillus subtilis* ATCC 6633, *Escherichia coli* ATCC 25922 and *Pseudomonas aeruginosa* (Table 1). The other constituents (**1**, **3** and **4**) are well known for their antibacterial activity and pharmacological properties (Duarte et al., 2006; Rhouma et al., 2009; Chantraine et al., 1998). The MIC of CH<sub>2</sub>Cl<sub>2</sub> extract was reported in Table 2.

#### Conclusions

Here we report the isolation and characterization of five constituents form the bark resin of *S. molle*: germacrene *D* (1), terebinthene (2), isomasticadienoic acid (3), isomasticadienonalic acid (4), and pinicolic acid (5). The resin of *S. molle* contained 10% of germancrene *D*(1), which is very high compared with other natural sources (Kapoor et al., 2009). Terebinthene (2) was identified as a constituent of *S. molle* for the first time and it showed significant cytotoxic activity against a human colon carcinoma cell line.

### Authors' contributions

GRMG contributed running the laboratory work, and drafted the paper; LH did the NMR investigations; RWB contributed in collecting plant samples and revised the paper; MLGY and API carried out biological activity tests and wrote one part of the manuscript.

All the authors have read the final manuscript and approved the submission.

#### **Conflicts of interest**

The authors declare no conflicts interest.

#### Acknowledgements

We would like to thank Prof. R. Müller, J. Herrmann and Mrs. Viktoria Schmitt (HIPS, Department of Microbial Natural Products, Saarbrücken, Germany) for technical assistance with the cytotoxic assay.

#### Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.bjp.2016.07.004.

#### References

- Abdel-Sattar, E., Zaitoun, A.A., Farag, M.A., El Gayed, S.H., Harraz, F.M.H., 2010. Chemical composition, insecticidal and insect repellent activity of *Schinus molle* L. leaf and fruit essential oils against *Trogoderma granarium* and *Tribolium castaneum*. Nat. Prod. Res. 24, 226–235.
- Abdel-Sattar, E., Khaleel, A.A., Harraz, F.M., 2007. Triterpene acids from the stem exudate of Schinus molle L. Egypt. J. Biomed. Sci. 24, 68–74.
- Barrachina, M.D., Bello, R., Martinez Cuesta, M.A., Primoyufera, E., Espulgues, J., 1997. Analgesic and central depressor effects of the dichloromethanol extract from *Schinus molle L.* Phytother. Res. 11, 317–319.
- Bendaoud, H., Romdhane, M., Souchard, J.P., Cazaux, S., Bouajila, J., 2010. Chemical composition and anticancer and antioxidant activities of *Schinus molle* L. and *Schinus terebinthifolius* Raddi berries essential oils. J. Food Sci. 75, C466–C472.
- Bülow, N., König, W.A., 2000. The role of germacrene D as a precursor in sesquiterpene biosynthesis: investigations of acid catalyzed, photochemically and thermally induced rearrangements. Phytochemistry 55, 141–168.
- Chantraine, J.M., Laurent, D., Ballivian, C., Saavedra, G., Ibañez, R., Vilaseca, A., 1998. Insecticidal activity of essential oils on *Aedes aegypti* lavae. Phytother. Res. 12, 350–354.
- CLSI, 2012. Methods for Dilution Antimicrobial Susceptibility Tests for bacteria That Grow Aerobically. Clinical and Laboratory Standards Institute, Wayne, PA, Approved Standard-Ninth Edition M07-A9.
- Díaz, C., Quesada, S., Brenes, O., Aguilar, G., Cicció, J.F., 2008. Chemical composition of *Schinus molle* essential oil and its cytotoxic activity on tumour cell lines. Nat. Prod. Res. 22, 1521–1534.
- Dominguez, X.A., 1973. Métodos de investigación fitoquímica. 1st, Editorial Limusa, México.
- Duke, J.A., 2002. Handbook of Medicinal Herbs, 2nd ed. CRC Press, Boca Raton, FL.
- Duarte, M.C.T., Leme, D.C., Figueira, G.M., Sartoratto, A., Rehder, V.L.G., 2006. Effects of essential oils from medicinal plants used in Brazil against epec and etec *Escherechia coli*. Rev. Bras. Pl. Med. 8, 139–143.
- Harborne, B.J., 1984. Phytochemical Methods, a Guide to Modern Techniques of Plant Analysis, 2nd ed. Chapman and Hall, New York.
- Kapoor, I.P.S., Singh, B., Singh, G., De Heluani, C.S., De Lampasona, M.P., Catalan, C.A.N., 2009. Chemistry and in vitro antioxidant activity of volatile oil and oleoresins of black pepper (*Piper nigrum*). J. Agric. Food Chem. 57, 5358-5364.
- Ko, H.H., Hung, C.F., Wang, J.P., Lin, C.N., 2008. Antiinflammatory triterpenoids and steroids from *Ganoderma lucidum* and *G. tsugae*. Phytochemistry 69, 234–239.
- Machado, D.G., Kaster, M.P., Binfaré, R.W., Dias, M., Santos, A.R.S., Pizzolatti, M.G., Brighente, I.M.C., Rodrigues, A.L.S., 2007. Antidepressant-like effect of the extract from leaves of *Schinus molle* L. in mice: evidence for the involvement of the monoaminergic system. Prog. Neuro-Psychol. Biol. Psychiatr. 31, 421–428.
- Mosa, R.A., Oyedeji, A.O., Shode, F.O., Singh, M., Opoku, A.R., 2011. Triterpenes from the stem bark of *Protorhus longifolia* exhibit anti-platelet aggregation. Afr. J. Pharm. Pharmacol. 5, 2698–2714.
- Perez, C., Anesini, C., 1994. Inhibition of *Pseudomonas aeruginosa* by Argentinean medicinal plants. Fitoterapia 65, 69–172.
- Pozzo-Balbi, T., Nobile, L., Scapini, G., Cini, M., 1978. The triterpenoid acids of Schinus molle. Phytochemistry 17, 2107–2110.
- Richter, R., von Reuß, S.H., König, W.A., 2010. Spirocyclopropane-type sesquiterpene hydrocarbons from *Schinus terebinthifolius* Raddi. Phytochemistry 71, 1371–1374.
- Rhouma, A., Daoud, H.B., Ghanmi, S., Salah, H., Romdhane, M., Demak, M., 2009. Antimicrobial activities of leaf extracts of *pistacia* and *Schinus* species against some plant pathogenic fungi and bacteria. J. Plant Pathol. 91, 339–345.
- Ruffa, M.J., Ferraro, G., Wagner, M.L., Calcagno, M.L., Campos, R.H., Cavallaro, L., 2002. Cytotoxic effect of Argentine medicinal plant extracts on human hepatocellular carcinoma cell line. J. Ethnopharmacol. 79, 335–339.
- Sarker, S.D., Nahar, L., Kumarasamy, Y., 2007. Microtitre plate-based antibacterial assay incorporating resazurin as an indicator of cell growth, and its application in the *in vitro* antibacterial screening of phytochemicals. Methods 42, 321–324.