

Immunological harmony: the dynamic influence of cellular and humoral immunity on pregnancy success

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Abstract

This study is a crucial step in understanding the dynamics of the maternal immune response directed at paternal human leukocyte antigen (HLA) molecules. HLA molecules are proteins on cell surfaces that play a critical role in immune system regulation. Our findings focus on the pivotal role of maternal antibodies targeting fetal HLA molecules in inhibiting antigen-induced activation of uterine immune cells, which is essential for successful pregnancies. Antibodies are proteins produced by the immune system that recognize and neutralize foreign substances. The primary focus is to unravel maternal anti-fetal rejection by drawing parallels to transplant rejection and emphasizing the role of allorecognition—the process by which an individual's immune system recognizes and responds to antigens from another individual of the same species—in both cellular (involving immune cells) and humoral (involving antibodies) refusal. Although exploring anti-HLA antibodies in preventing fetal loss in patients with recurrent spontaneous abortion is captivating, there are still significant knowledge gaps that need to be addressed. Further studies are imperative to reveal the precise mechanism by which these antibodies generate and prevent maternal immune responses, critical determinants of pregnancy outcomes. It is vital to investigate the specificity of these antibodies and whether they exclusively target specific HLA molecules on trophoblasts (cells forming the outer layer of a blastocyst, providing nutrients to the embryo). This review paper not only offers insights into the development of these protective antibodies in pregnancy but also lays the foundation for future research on therapeutic implications, particularly in cases of recurrent spontaneous abortion.

Keywords Pregnancy · Abortion · Therapeutic targets

1 Introduction

Pregnancy introduces a unique challenge to the maternal immune system as it encounters the fetus, typically considered “semi-foreign” due to the presence of paternal antigens alongside maternal antigens [1, 2]. The success of pregnancy hinges on the sophisticated balance between maternal immune tolerance to the semi-allogeneic fetus and the defense mechanisms guarding against fetal loss. Rather than dismissing paternal antigens, a complex interplay of regulatory mechanisms unfolds, shielding the fetal allograft from rejection [3]. This narrative investigates the development and potential failure of these mechanisms, particularly focusing on the formation of antibodies to paternal human leukocyte antigen (HLA). These antibodies play a crucial role in maintaining normal pregnancy by preventing maternal immune cells from attacking the fetus. However, in patients with recurrent spontaneous abortion

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(RSA), the formation of anti-HLA antibodies can become dysregulated, leading to an adverse immune response that triggers fetal loss [4–6].

RSA is defined by the loss of three or more consecutive pregnancies before 20–28 weeks of gestation, affecting approximately 2.5% of women attempting to conceive [6–9]. The risk increases with maternal age and history of RSA elevates the risk for subsequent occurrences [10]. Chromosomal abnormalities contribute to 50–85% of RSA cases, underscoring their significant role [11, 12]. Other contributing factors include autoantibodies such as antiphospholipid antibodies, anti-thyroid antibodies, antinuclear antibodies, anti-transglutaminase antibodies, and anti-endothelial antibodies, as well as smoking, caffeine intake, contraceptive drug use hormonal problems, abnormal glucose metabolism, stress, and depression [13–22].

The elusive etiology of RSA remains unknown in over half of cases, but the deficiency of antibodies to paternal HLA molecules has been observed, suggesting their role in maintaining healthy pregnancies [5, 23–29]. Paternal cell immunization, akin to methods dampening organ allograft rejection, emerges as a hopeful therapeutic avenue. This approach aims to instill tolerance to paternal antigens, orchestrating the suppression of allogeneic immune reactions and fostering immune tolerance to the growing fetus. Clinical trials and preclinical studies demonstrated promising results, not only improving pregnancy outcomes but also reducing abortion rates in RSA patients [5, 25].

In the landscape of pregnancy success and recurrent pregnancy loss, anti-HLA antibodies stand as an ideal, offering a promising therapeutic strategy against allogeneic immune responses contributing to abortion. This paper serves as a current and insightful update on the impactful role of the critical HLA molecules, steering the course toward controlling adverse immune reactions that precipitate fetal loss in resilient women grappling with RSA.

1.1 Embarking on the odyssey of fetal development: a chronicle of marvels in human pregnancy

The immunology governing fetal survival during pregnancy remains a profound paradox, and deciphering this mystery requires understanding the intricate interplay between embryonic developmental events and fundamental immunological mechanisms.

Pregnancy unfolds as a series of events, encompassing fertilization, implantation, embryonic and fetal growth, culminating in birth after approximately 266 days or later (the gestation period), which is divided into the first, second and third trimesters [30]. The initial gestational period, particularly the first trimester is particularly critical, marked by a underlying cascade of events that shape the outcome of the pregnancy [31].

1.2 Conception chronicles: showing the details of fertilization in human reproduction

Fertilization is a complex process wherein the genetic material from spermatozoa and ovum converges into a single nucleus. Despite the introduction of approximately 300 to 500 million sperm cells into the vagina, less than 1% manage to reach the secondary oocyte. Normally occurring in the uterine (Fallopian) tube about 12 to 24 hours after ovulation, fertilization is a precise event. Sperm undergo maturation in the epididymis, becoming capable of fertilizing an oocyte after spending around 10 hours in the female reproductive tract [32, 33].

During fertilization, only one spermatozoon penetrates and enters a secondary oocyte (Fig. 1A) a process termed syngamy [34]. This event induces depolarization, triggering the release of calcium ions inside the cell [33]. These calcium ions stimulate the release of granules by the oocyte, promoting changes that block the entry of other sperm and prevent polyspermy [35].

Once a spermatozoon has entered a secondary oocyte, equatorial division (meiosis II) is completed. This division results in a larger ovum (mature egg) and a smaller second polar body that fragments and disintegrates. The tail is shed, and the nucleus in the head develops into a structure known as the male pronucleus. Simultaneously, the nucleus of the ovum develops into a female pronucleus. Upon the formation of pronuclei, they fuse to create a segmentation nucleus containing 23 chromosomes (n) from the male pronucleus and 23 chromosomes (n) from the female pronucleus. This fusion restores the diploid number (2n), and the fertilized ovum, now a zygote (Fig. 1B), comprises a segmented nucleus, cytoplasm, and zona pellucida [33, 36].

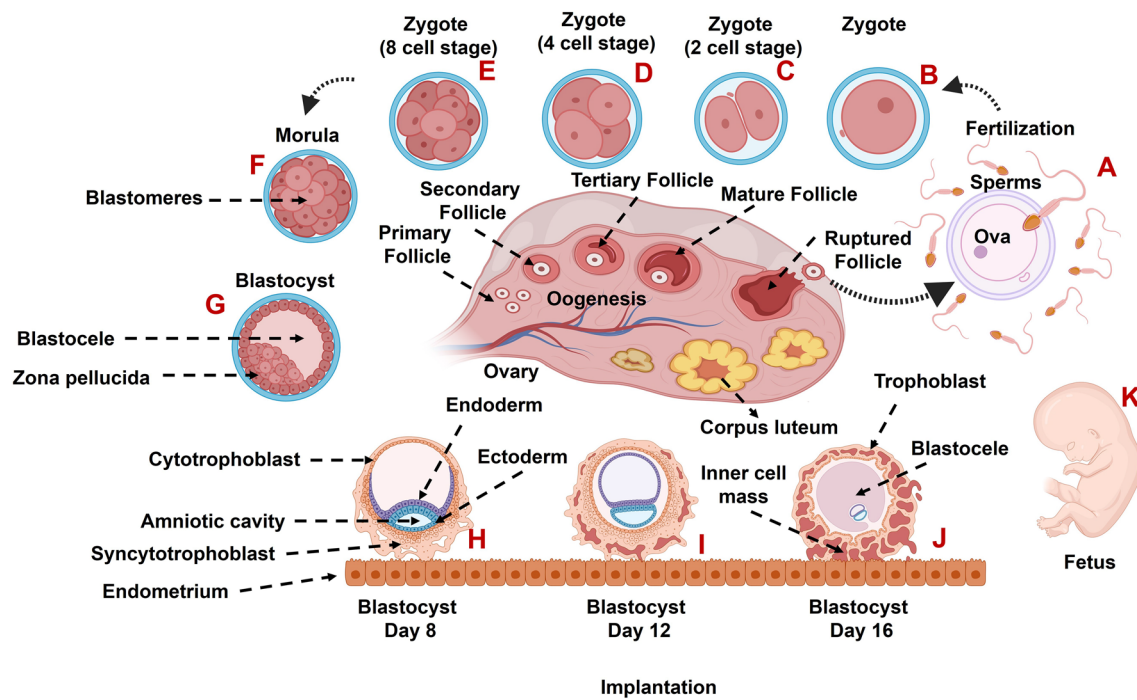


Fig. 1 Sequential events from fertilization to implantation in human pregnancy. **A** Fertilization: Illustrates the convergence of genetic material from spermatozoa and ovum into a single nucleus, resulting in the formation of a zygote. The process includes syngamy, equatorial division (meiosis II), and the formation of male and female pronuclei. **B–G** Development of Blastocyst: Demonstrates the progression from zygote to blastocyst through rapid mitotic cell divisions. The stages include the formation of blastomeres, the development of the morula, and the transformation into a blastocyst with an outer trophoblast, inner cell mass, and blastocoel. **H–K** Implantation: Depicts the process of blastocyst attachment to the uterine wall. The disintegration of the zona pellucida, development of syncytiotrophoblast and cytotrophoblast, and enzymatic penetration into the endometrium are highlighted. The blastocyst becomes embedded in the endometrium, with the trophoblast contributing to placental development

1.3 Blossoming potential: the enigmatic journey of blastocyst development in early embryogenesis

On the fifth day after fertilization, the zygote undergoes rapid mitotic cell divisions, leading to an increase in the number of cells. However, the size of the embryo does not enlarge, as it remains confined within the zona pellucida. The first cleavage is accomplished within approximately 36 hours, with subsequent divisions taking slightly less time. By the second day post-fertilization, the second cleavage is completed, resulting in 2, 4, 8, and 16 cells (Fig. 1C–F). These progressively smaller cells, formed through cleavage, are referred to as blastomeres [33].

Successive cleavages give rise to a solid mass of cells, still enveloped by the zona pellucida, known as the Morula (Fig. 1F). A few days after fertilization, the morula reaches a size like that of the original zygote. By the end of the fourth day, the number of cells in the morula increases, and it progresses along the uterine (Fallopian) tubes toward the uterine cavity [37]. Around four and a half to five days post-fertilization, the dense cell clusters transform into a hollow ball of cells, entering the uterine cavity and now referred to as a blastocyst (Fig. 1G). The blastocyst comprises an outer covering of cells called the trophoblast, an inner cell mass (embryoblast), and an internal fluid-filled cavity known as the blastocoel [33, 38].

1.4 Nesting perfection: decoding the enigmatic ballet of implantation in human reproduction

Implantation is a critical step in the initial stages of pregnancy, marking the attachment of the fertilized egg to the lining of the uterus. This process typically occurs around six to ten days after fertilization, just as the blastocyst, a ball of cells resulting from the transformation of the morula into the blastocyst, consisting of two main cell types, i.e., the inner cell mass and the outer layer known as the trophoblast. The blastocyst, now with the differentiated trophoblast, undergoes implantation into the endometrial lining of the uterus [39–41]. As the blastocyst burrows into the endometrial tissue,

trophoblast undergoes differentiation into two layers: the cytotrophoblast and the syncytiotrophoblast [39–41] indicated in the Fig. 1H, I, and J.

The cytotrophoblast is a layer of individual, undifferentiated cells closest to the inner cell mass and serve as the stem cells of the placenta and differentiate into the other two types of trophoblasts, namely syncytiotrophoblasts and extravillous trophoblast [39, 40, 42, 43]. The syncytiotrophoblast expands and facilitates nutrient and gas exchange between maternal and fetal circulations, while extravillous trophoblasts act as a barrier between maternal blood vessels and the embryo, enabling exchange of gases, nutrients, and waste products [44–47]. As pregnancy advances, extravillous trophoblasts differentiate into endovascular and interstitial trophoblasts, contributing to placental structure formation and growth of the fetus [42, 44–49]. In essence, implantation is a remarkable example of the complex and finely orchestrated series of events that occur during the initial stages of pregnancy. It lays the groundwork for the subsequent development of the placenta and the maturation of the embryo into a fully formed fetus (Fig. 1K)

1.5 Unlocking genetic identity: human leukocyte antigen system operation in pregnancy

The successful formation of the placenta and embryo during pregnancy presents a unique immunological challenge due to the fetus being a semi-allograft, inheriting half of its genes from the father, which differ from those of the mother [50–52]. This genetic disparity, particularly evident in the human leukocyte antigen (HLA) genes, poses a significant obstacle to maternal immune tolerance, as the maternal immune system typically recognizes and eliminates foreign tissues [53].

Despite defying traditional transplantation principles, the embryo remarkably survives within the potentially hostile maternal immune environment [54, 55]. The HLA gene, which is the human equivalent of the major histocompatibility complex (MHC), was first identified in mice as a genetic locus associated with organ transplant acceptance or rejection. In humans, Jean Dausset and Jan van Rood described this genetic system in 1954, naming it the HLA [56].

Comprising more than 200 genes located on short arm of chromosome 6, the HLA system is recognized as the most polymorphic genetic system in humans [57–59]. It consists of three groups known as Class I, II, and III [60–67]. Risk alleles associated with pregnancy failure predominantly involve HLA I and HLA II [63, 68–76].

1.6 HLA-class I: orchestrating immune vigilance in pregnancy

The HLA I gene is categorized into classical HLA Ia and non-classical HLA Ib. HLA Ia includes HLA A, B, and C, while HLA Ib comprises HLA E, F, and G [77–80]. HLA Ia encodes HLA A, B, and C molecules, consisting of a glycosylated heavy chain with $\alpha 1$, $\alpha 2$, and $\alpha 3$ domains, and a light chain composed of $\beta 2$ macroglobulin protein [81]. HLA Ib encodes HLA E, F, and G molecules. The HLA I molecule's α chain has a transmembrane domain facilitating association with the cell membrane, determining antigenic specificities. The molecular weights of the α chain and $\beta 2$ macroglobulin protein are approximately 45 kDa and 12 kDa, respectively. They are expressed on the membrane surface in a non-covalently bound state [82]. HLA I-encoded proteins are expressed on nearly all nucleated cells and platelets, presenting peptides from endogenous proteins of virus-infected or tumor cells. These peptides bind to the peptide-binding cleft of HLA I proteins, presenting them on CD8+ T cells [83–85]. Classical HLA Ia molecules (A and B) are exclusively found on fetal tissues, while HLA C is present on both trophoblast and placenta (Table 1). Certain HLA I alleles, such as HLAC1 C1 and HLA C2C2, have shown an increased proportion in patients with RSA compared to control women [75]. Non-classical HLA-Ib molecules (E, F, and G) are detected on both trophoblast and placenta (Table 1). In contrast to HLA-Ia, HLA-Ib molecules demonstrate reduced polymorphisms and diminished cell surface expression [86, 87].

The extravillous trophoblast, integral to the success of pregnancy, is characterized by the lack of expression of HLA-II molecules, while prominently expressing HLA-Ia, (e.g., HLA-C) and HLA-Ib molecules, such as HLA-E, HLA-F, and HLA-G. (Table 1). Notably, the upregulation of HLA Class I molecules in the placenta is observed in villitis, an inflammatory condition associated with miscarriage and stillbirth [88, 89]. Studies have also underscored the essential interaction between HLA-Ib molecules and NK cells in promoting pregnancy success [63, 76, 77, 87, 90–96]. This raises the intriguing possibility that the differential expression of HLA-Ib compared to HLA-Ia could play a distinct role in the mechanisms underlying pregnancy loss. Exploring this nuanced interplay between HLA-Ib expression and immune regulation in the placental environment may uncover new insights into the etiology of pregnancy complications and inform innovative therapeutic strategies.

Table 1 Presence of HLA molecules on embryo and fetus

	Cytotrophoblast	Syncytio-trophoblast,	Extravillous trophoblast		Placenta	Fetal tissue	References
			Endovascular trophoblast	Interstitial trophoblast			
HLA-A		–	–	–		S+	[63, 97]
HLA-B		–	–	–		S+	[63, 97]
HLA-C	IC+	IC+	S+	S+	S+	S+	[63, 76, 97–101]
HLA E	SI-ICs+	SI-ICs+	SI-ICs+	SI-ICs+	SI-ICs+	S+	[63, 97, 102, 103]
HLEF	SI-ICs+	SI-ICs+	SI-ICs+				[76, 104–106]
HLAG	SI-ICs+	SI-ICs+	SI-ICs+			S–	[63, 76, 77, 97, 104, 107]
HLADP						S+	[63]
HLADQ						S+	[63]
HLADR			–			S+	[63, 97]

S, Surface expression, IC, Intracellular expression, S-ICs, Surface and intracellular expression, + positive expression, –, no expression

1.7 HLA-class II: simplifying immune complexity in pregnancy

HLA-Class II genes play a vital role in regulating immune responses during pregnancy. These genes, including HLA DRA1, DQA1, DPA1, DQB1, and DPB1, encode a variety of HLA DR, DP, and DQ proteins [84, 108, 109]. Among these, HLA DRA1, DQA1, and DPA1 encode the α chain, while HLA DRB1, DRB3, DRB4, DRB5, DQB1, and DPB1 encode the β chain. HLA DRA1 forms heterodimers with HLA DRB1, DRB3, DRB4, or DRB5, whereas HLA DQA1 and DPA1 are associated respectively with HLA DQB1 and DPB1 [110–112].

HLA DR is classified into five distinct groups: DR1, DR51, DR52, DR53, and DR8, based on antigen groups. While the DR1 and DR8 groups exclusively consist of DRB1, the DR51, DR52, and DR53 groups include DRB1 along with additional expressions of DRB5, DRB3, and DRB4, attributed to DRB1 gene duplication [113–116]. The primary function of HLA-Class II proteins lies in processing and presenting peptides derived from exogenous antigens to CD4⁺ T cells [108, 117–119]. Initial analyses of HLA-Class II began with the discovery of HLA-D via the mixed lymphocyte culture test, followed by the identification of HLA-DR and HLA-DQ through subsequent test [120–122]. Studies have shown that patients with RSA exhibit an increased proportion of specific HLA alleles, such as HLA DQA105/B102, HLA DQA10505, HLA DQ2, HLA DQ8, HLA-DRB103, and HLA-DRB107, compared to control subjects [68–74].

Unlike HLA Class I molecules, HLA Class II molecules have not been observed on the trophoblast and placenta. However, their presence has been documented in fetal tissues (Table 1) indicating a potential connection between HLA-Class I and HLA-Class II risk alleles and pregnancy failure. Nevertheless, the mechanisms driving HLA sensitization during pregnancy are primarily associated with fetal cell trafficking, feto-maternal hemorrhage, organ transplantation, and blood transfusion [123–129]. Further research is imperative to unravel the complexities of HLA sensitization and its impact on maternal–fetal immune tolerance.

1.8 HLA function in defeating the cellular ballet in graft rejection

HLA-induced cellular immune responses play a pivotal role in graft rejection, particularly in transplant scenarios where both donor-derived and recipient-derived immune cells are implicated. This includes CD4⁺ T cells, CD8⁺ T cells, myeloid-derived suppressor cells, neutrophils, and natural killer cells, collectively contributing to allograft rejection [130–137].

There are mainly three pathways characterizing the interaction between donor and recipient cells. In the direct pathway, donor-derived antigen-presenting cells (APCs) directly present donor antigens via HLA to recipient-derived CD4⁺ T/ CD8⁺ T cells [138]. The indirect pathway involves recipient APCs processing donor antigen and presenting specific molecules through their HLA to recipient derived CD4⁺ T/ CD8⁺ T cells [139]. The semi-direct pathway, intriguingly, utilizes non-processed donor antigens through recipient APCs HLAII and recipient-derived CD4⁺ T cells [140]. These pathways lead to the activation of alloantigen-specific CD4⁺ T/ CD8⁺ T cells, recognizing alloantigen through HLA II and CD4 TCR or HLA I and CD8 TCR [137–146].

HLA I-mediated antigen presentation involves the processing of peptides derived from endogenous proteins, which happens through peptide-binding cleft of HLA I proteins and their presentation on CD8⁺ T cells. These peptides are typically 8 to 11 amino acids in length. In contrast, HLA II-mediated antigen presentation involves the processing of non-self-peptides derived from exogenous proteins, which happens through peptide-binding cleft of HLA II proteins and their presentation on CD4⁺ T cells. The peptides binding to the peptide-binding cleft of HLA II-encoded proteins are longer, approximately 15 to 30 amino acids [83–85, 147–151].

Notably, primary villous trophoblast cells do not express HLA I or HLA II molecules, suggesting that CD4⁺ and CD8⁺ T cells may not engage with the placental barrier, providing a highly effective mechanism for protecting the placenta from harm [97, 152]. Extra villous trophoblasts express HLA C, E, F, and G, but not HLA A, B, and DR molecules (Table-1). Furthermore, interaction between decidual HLA E and maternal CD8⁺ T cells has been observed [153]. Antigen presentation by HLA I-CD8⁺ T cells pathway causes the activation of CD8⁺ T cells and resulting massive release of cytotoxic granules containing perforin or granzymes, which lead to the lysis of virus-infected cells, cancer cells, and non-self-cells while preventing the growth of non-autologous cells [83–85, 147–151]. It is proposed that extra villous trophoblasts expressing HLA I molecules (e.g., C, G, E, and F) may utilize endogenous antigens, activating maternal CD8⁺ T cell effector functions, potentially leading to adverse pregnancy outcomes.

Conversely, the polymorphic nature of HLA C categorizes it into two allotypes, HLA C1 and HLA C2. Corresponding receptors expressed on decidual natural killer cells (dNKs), termed killer cell immunoglobulin receptors (KIRs), comprise inhibitory receptors (KIR2DL2 or KIR2DL3 specific for HLA C1 and KIR2DL1 specific for HLA C2) and activating receptors (KIR2DS1 specific for HLA C2) [100, 154]. The interplay of KIR inhibitory receptors with HLA-C is crucial for dNK cells to recognize and tolerate fetal antigens, while the absence of appropriate activation mediated by KIR activating receptors may result in increased interferon-gamma (IFN γ) production. This dysregulation in dNK cell function is pivotal in inducing adverse pregnancy outcomes such as RSA and preeclampsia [76, 155–157]. Additionally, besides HLA C, trophoblasts also express HLA E, F and G (Table 1) These findings suggest that the interaction of KIR inhibitory receptors with certain HLA I molecules (e.g., C, E, F, and G) could skew immune responses towards a tolerogenic rather than an immunogenic response.

The activation process of CD4⁺ T cells rely on three pivotal signals. First, the antigen-specific signal is conveyed through the T cell antigen receptor (TCR), which interacts with peptide-HLA class II complexes present on the surface of antigen-presenting cells. Second, the co-stimulatory signal, which is antigen nonspecific, emerges from the interaction between co-stimulatory molecules (e.g., CD80 and CD86) expressed on APCs. Third, the establishment of the third signal occurs through the interaction of stimulatory molecules (e.g., CD28/CD40L) on CD4⁺ T cells with TCR/CD40 present on antigen-presenting cells [158, 159]. This cellular activation leads to the differentiation of CD4⁺ T cells into distinct types of effectors CD4⁺ T cells T helper 1 (Th1), Th2, Th17, T regulatory (Treg), and T follicular helper (Tfh) cells, each associated with signature cytokines [160–163]. These T helper cell subsets, along with their respective cytokines such as interferon-gamma (IFN γ ; Th1), interleukin 4 (IL4; Th2), IL17 (Th17), transforming growth factor-beta (TGF β ; Treg), and IL6 (Tfh), contribute to tissue inflammation in various visceral and brain diseases [164–170].

Th1 cells produce IFN γ , IL2, and tumor necrosis factor-alpha (TNF α) to combat intracellular pathogens and evoke cell-mediated immunity. On the other hand, Th2 cells produce IL4, IL5, and IL13 to eliminate extracellular organisms and trigger robust allergic responses. Notably, Th17 cell differentiation, unlike Th1 and Th2, does not require IL17 but is critically dependent on TGF β and IL6 [148, 160, 171–176]. Treg cells, on the other hand, produce IL10 and TGF β , promoting immune tolerance and inhibiting IFN γ synthesis. They also play a role in blocking the differentiation of naïve T cells into effector T cells, contributing to immune homeostasis. Additionally, T helper cell subsets have the capacity to produce IL-10, a cytokine with broad immunoregulatory properties [177, 178]. These findings suggest that systemic differentiation and cytokine production by CD4⁺ T cells contribute to the complex orchestration of immune responses and immune regulation in various physiological and pathological contexts.

Multiparous women, including blood donors, showed heightened mismatching with paternal/fetal HLA I and II alleles (e.g., HLA A, B, C, E, F, G, DQ, and DR), leading to increased pregnancy success [25, 179–187]. Studies indicate that patients with RSA exhibit a higher proportion of specific HLA alleles (e.g., HLA DQA105/B102, HLA-DQA10505, HLA DQ2, HLA DQ8, HLA-DRB103, and HLA-DRB1*07) compared to control women [68–74]. In addition altered infiltration of decidual APCs, (e.g., dendritic cells, and macrophages), NK cells, B cells, and T cells as well as their effector Th1, Th2, Th17, and Treg cytokines have been linked to both pregnancy success and adverse pregnancy outcomes, encompassing RSA, preterm birth and pre-eclampsia [22, 188–211].

Th1 cells are associated with pro-inflammatory responses, producing cytokines like IFN γ [212–215]. In the context of pregnancy, an overactive Th1 response can pose a threat by promoting inflammation and potentially leading to fetal rejection [216–221]. On the other hand, Th2 cells are recognized for their anti-inflammatory nature, producing cytokines

such as IL4, IL5, and IL13 [222, 223]. A balanced TH2 response is crucial for fostering an environment conducive to implantation and maintaining pregnancy [216, 218–220, 224]. Th17 cells, known for generating pro-inflammatory cytokines like IL-17, contribute to tissue inflammation and the defense against pathogens [214, 215, 225, 226]. However, an exaggerated Th17 response may contribute to adverse pregnancy outcomes by promoting inflammation and tissue damage [192, 195, 227]. T reg cells suppress immune responses and help prevent the immune system from attacking fetal tissues [228–231]. Insufficient T reg cell activity can result in immune-mediated complications during pregnancy [195, 227].

Overall, the delicate interplay of these T-helper cell subsets is vital for a successful pregnancy. Imbalances, such as an excessive Th1 or Th17 response or inadequate T reg cell function, can contribute to conditions like RSA or preeclampsia. Understanding the critical network of these T-cell subsets and their effector functions provides insights into the immunological complexities of pregnancy.

Many of the immune regulatory and inflammatory molecules found in the circulation of women with normal pregnancy and RSA play critical roles in protecting against fetal loss triggered by various infectious agents. This protection is particularly important as trophoblast cells, expressing exclusively HLA I molecules, can face direct lysis through interaction with maternal CD8⁺ T cells. Future research in this area should explore the specific interactions between different HLA I molecules (such as HLAC, E, F, and G) and CD8⁺ T cells. Understanding these interactions will illuminate how they contribute to immune responses at the maternal-fetal interface, emphasizing the precise balance necessary for ensuring a healthy and successful pregnancy.

1.9 HLA operates humoral immune responses

The role of HLA-induced humoral immune responses is critical in the context of organ transplantation. When there are mismatches in HLA alleles between the donor and recipient, it can initiate the production of donor-specific anti-HLA antibodies, ultimately leading to organ rejection [232, 233]. In the indirect pathway, B cells play a crucial role by presenting processed alloantigens to CD4⁺ T cells in a manner restricted by self-HLA. This interaction serves as the catalyst for initiating humoral allo-immunity. Consequently, this activation process triggers responses in both CD4⁺ T cells and B cells, ultimately leading to the formation of long-lived plasma cells [234–237]. The presence of donor-specific anti-HLA antibodies, particularly in large quantities, poses significant concerns due to their profound impact on transplantation outcomes. These antibodies can manifest categorically as acute or chronic reactions, which exhibit clear pathological distinctions in kidney and liver transplantation settings [238–240].

However, in heart, lung, pancreas, and small bowel transplantation, universally articulated differential definitions are lacking, despite the presence of diagnostic criteria for antibody-mediated reactions. [241, 242]. Generally, class I donor-specific antibodies are pivotal in crossmatch positive transplantation, while de novo class II donor-specific antibodies significantly influence long-term graft loss post-transplantation [243, 244].

In theory, HLA II antigens are generally limited in abundance across most graft tissues, except for endothelial cells, smooth muscle cells, and certain APCs, whereas class I antigens exhibit widespread expression. As a result, HLA I donor-specific antibodies offer numerous targets, while those for HLA II donor-specific antibodies are comparatively fewer, with antibody-mediated reactions primarily influenced by the presence of HLA I donor-specific antibodies.

Nevertheless, the expression of HLA II antigens in pre-transplant tissues varies based on circulating cells, and the initial presence of HLA II antigens may impact transplantation outcomes [245]. Conversely, in cases of persistent tissue injury from rejection, infection, or drug toxicity, there may be an up-regulation of HLA II antigens, rendering them susceptible targets for HLAII donor-specific antibodies [246–249]. Antibody-mediated reactions mediated by class II donor-specific antigens seem to present a persistent and substantial burden on graft function in kidney, liver, heart, and lung transplants, compared to rejection due to class I donor-specific antibodies [243, 250–252].

Multiparous women, encompassing both blood donors and individuals in the first, second, and third trimesters, as well as the postpartum period, exhibited elevated mismatching when compared to paternal/fetal HLA class I alleles (e.g., HLA A, B, C, E, F, and G) and HLA class II alleles (e.g., HLA DQ and DR). This enhanced mismatching was associated with an increased development of corresponding IgG antibodies targeting HLA class I and II molecules and the pregnancy success [25, 179–187].

Notably, patients with RSA, experiencing consecutive spontaneous abortions of unknown origin, demonstrated a significantly increased frequency of shared HLA alleles (A, B, C and DQ/DRI) with their spouses compared to control women [69, 70, 253–262]. The development of IgG antibodies to HLA I molecules (e.g., A and B) has been noted in the first, second, and third trimesters, whereas IgG antibodies to HLA II molecules (e.g., DQ and DR) show an increase only in

the third trimester [182–187, 262, 263]. Conversely, patients with RSA, compared to normal pregnant women, exhibit a marked reduction in IgG antibodies against several specified HLA I and II molecules [25, 253, 254, 261–267].

Studies have demonstrated the positive impact of immunotherapy using paternal lymphocytes on pregnancy success in patients with RSA [24, 268–275]. These therapies have been shown to stimulate the production of antibodies against paternal HLA I and HLA II molecules, thereby contributing significantly to successful pregnancies in RSA patients [4–6, 27, 253, 261, 276–280]. Overall, the efficacy of immunotherapy with paternal lymphocytes is well-supported by extensive research, which emphasizes the beneficial immune responses that these treatments elicit. Specifically, the increased production of anti-paternal HLA antibodies is a key factor in promoting immune tolerance and enhancing pregnancy outcomes in women with RSA. However, it is important to note that while most studies report positive effects, a few have failed to detect significant benefits [281–283]. These studies, however, did not assess the development of anti-HLA antibodies, which is a critical component of the therapeutic mechanism.

This disparity underscores the need for comprehensive evaluation criteria in future research. By incorporating the measurement of all classes of anti-HLA antibody development, we can gain a more accurate understanding of the effectiveness of immunotherapy with paternal lymphocytes. Continued investigation in this area holds the promise of refining therapeutic approaches and improving pregnancy success rates for patients with RSA.

The human fetus produces IgM and IgA antibodies [284–286]. IgM, the initial antibody class generated by B lymphocytes, displays reduced specificity and antigen affinity [287]. Upon encountering antigens, B lymphocytes switch from IgM to IgG, an immunoglobulin with diverse effector functions in human health and disease [285, 288, 289]. Interestingly, the fetus does not produce its own IgG antibodies [285, 288, 289]. Studies have demonstrated the development of CD8 T follicular cells, which acquire CD4 T follicular helper (Tfh)-like functionality termed as CD8 T follicular like cells (CD8TFLCs). These cells produce Tfh effector cytokines and co-receptors, promoting B cell antibody class switching [290–293]. In vitro addition of purified IgG (from normal pregnant women or paternal lymphocyte-immunized patients with RSA) in a lymphocyte proliferation assay resulted in reduced cell proliferation [294]. This suggests the involvement of maternal CD8TFLCs and the fetal HLA I/a/b axis in inducing maternal B cell activation and trans differentiation to plasma B cells, resulting in the development of IgG antibodies to fetal HLA I molecules. These antibodies serve to protect against cytotoxic CD8 T cell-mediated fetal damage during pregnancy.

However, the precise mechanism underlying the development of IgG antibodies to specific paternal HLA I molecules in women with normal pregnancy and lymphocyte-immunized patients with RSA remains poorly defined. Further investigation is crucial to elucidate the nature of the fetal antigen and its specificity for distinct HLA I molecules expressed on trophoblast cells. It is imperative to determine whether IgG antibodies developed against these specific HLA I molecules belong to IgG1, IgG2, IgG3, or IgG4 subclasses. Additionally, identifying which isotype optimally inhibits the interaction between fetal-specific HLA I molecules and maternal CD8⁺ T cells, as well as the resulting effector functions that determine pregnancy success or failure, is essential.

Further studies aimed at elucidating the specific mechanisms underlying antibody production against distinct paternal HLA I molecules, as well as identifying and developing specific HLA I molecule targets, could significantly enhance the clinical relevance of this research. This focused approach is essential for bridging current knowledge gaps and maximizing the impact of these findings.

This deeper understanding not only enriches our comprehension of the immunological factors that influence pregnancy outcomes but also lays the groundwork for potential therapeutic interventions designed to modulate immune responses at the maternal-fetal interface. These advancements are crucial for maintaining the delicate balance required for a healthy and successful pregnancy.

2 Discussion

Pregnancy immunology suggests the fetus as a semi-allograft, an entity tolerated during pregnancy [1, 3, 295–304]. In transplantation, significant differences in donor and recipient HLA antigens may trigger T cell recognition of the donor's HLA molecules as foreign, initiating an immune response leading to the production of anti-HLA IgG antibodies. HLA II molecules, presenting peptides to CD4⁺ T cells, can similarly induce T cell activation and antibody production, leading to organ rejection [123, 235, 238, 305–307].

In the context of pregnancy, increased matching between maternal and paternal HLA can prevent the development of IgG antibodies to paternal HLA molecules. This situation enhances the interaction between maternal T cells and paternal HLA molecules, resulting in increased processing and presentation of endogenous/exogenous antigens and the

generation of pro-inflammatory cytokines, leading to fetal loss in patients with RSA [25, 253, 254, 262–267]. Conversely, when maternal and paternal HLA significantly differ, maternal T cells recognize the paternal HLA molecules as foreign, prompting massive production of IgG antibodies to these HLA molecules. These antibodies block the paternal HLA molecules, inhibiting maternal CD8⁺T cell-induced cellular activation and secretion of cytolytic mediators, perforin, and granule serine proteases (granzymes), and protecting against fetal rejection in normal pregnancy [308, 309]. Additionally, immunization with paternal lymphocytes in RSA patients has shown increased IgG antibody development to paternal HLA molecules, restoring T cell effector function, and protecting against fetal loss [27–29, 261, 270, 276–281, 310–317]. While such immunization has demonstrated promising results in enhancing pregnancy success rates among women with RSA [4–6, 24, 27, 253, 261, 268–280], a conscientious review of the studies is warranted.

Based on the exclusive presence of HLA I molecules (e.g., HLA C, E, F, and G) on trophoblasts (Table 1) and the discovery of CD8 T follicular cytotoxic cells (CD8TFLCs), which can promote T cell-dependent differentiation of plasma cells and antibody class-switching [290–293], we propose an additional layer to our understanding. This evidence suggests that a decrease in maternal-paternal HLA molecule matching specifically triggers the activation of the HLA I-CD8TFLCs axis. This activation leads to B cell plasmacytosis and the massive production of IgG antibodies against paternal HLA I molecules in early pregnancy. These antibodies play a crucial role in inhibiting the expression of HLA I molecules on trophoblast cells, thereby preventing maternal CD8⁺ cytotoxic T cells from targeting and inducing cell death in trophoblast cells through the secretion of cytolytic mediators such as perforin and granule serine proteases (granzymes). This proposed mechanism points out a potential pathway contributing to the success of pregnancy (Fig. 2A–G).

Conversely, increased maternal-paternal HLA I matching suppresses the development of maternal IgG antibodies against paternal HLA I a and b molecules, i.e., HLA- C, E, F, and G in early pregnancy. This scenario increases antigen processing through indicated HLA I molecules on trophoblast cells and their presentation to maternal CD8⁺ T cells. Such antigen processing activates CD8⁺ T cells, differentiating them into effector CD8⁺ T cells (Cytotoxic T cells). These cells directly target trophoblast cells, inducing cell death through the secretion of cytolytic mediators, perforin, and granule serine proteases (granzymes). This cascade contributes to fetal loss in recurrent spontaneous patients (Fig. 3A–E).

This study proposes an added layer to our understanding of paternal lymphocyte immunization-induced processing and presentation of fetal antigens by HLA I molecules on trophoblast cells to maternal CD8⁺ T cells. This leads to

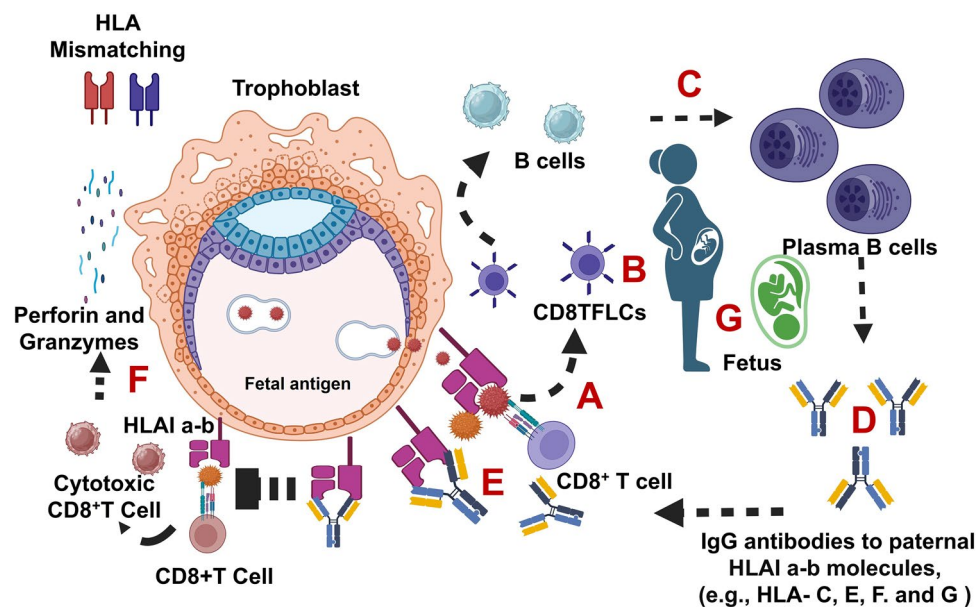


Fig. 2 The crucial role of decreased matching between maternal and paternal HLA I on trophoblast cells in initiating a successful pregnancy. **A** Reduced matching prompts the process and presentation of fetal antigens to maternal CD8⁺ T cells. **B** This antigen processing leads to the differentiation of CD8⁺ T cells into CD8T follicular-like cells (CD8TFLCs). **C** These cells interact with maternal B cells, resulting in the formation of plasma B cells. **D** Plasma B cells generate IgG antibodies to paternal HLA I a and b molecules, i.e., HLA- C, E, F and G (**E**). These antibodies effectively block paternal HLA I a and b molecules, preventing the interaction between maternal CD8⁺ T cells and trophoblast cells. (**F, G**) This blockade inhibits the HLA I-CD8⁺ T cell interaction mediated formation of cytotoxic CD8⁺ T cells and the production of fetal-damaging cytolytic mediators, (e.g., perforin and granzymes), ultimately protecting trophoblast cells from apoptosis and contributing to the success of pregnancy, ensuring the delivery of a healthy fetus

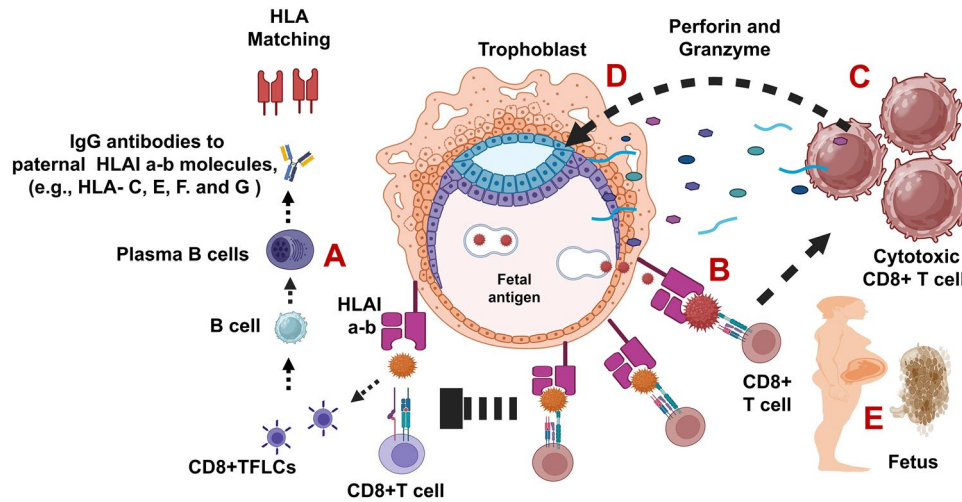


Fig. 3 The impact of increased matching between maternal and paternal HLA I on trophoblast cells, leading to fetal loss in women with recurrent spontaneous abortion. **A** Increased matching hinders maternal CD8⁺ T cell-mediated processing of fetal HLA I molecules, preventing CD8⁺ T cell differentiation into CD8⁺ T follicular-like cells (CD8⁺TFLCs). This, in turn, impacts maternal B cell activation, inhibiting the formation of plasma B cells and the production of IgG antibodies specific to paternal HLA I molecules. **B** The deficiency or absence of these specific maternal IgG antibodies to binding to HLA I molecules on trophoblast cells intensifies antigen processing and presentation to maternal CD8⁺ T cells. **C** Antigen processing activates CD8⁺ T cells, differentiating them into cytotoxic CD8⁺ T cells. **D** These cells directly target trophoblast cells, inducing cell death through the secretion of cytolytic mediators, perforin, and granule serine proteases (granzymes). **E** This cascade contributes to fetal loss in recurrent spontaneous patients

the development of CD8⁺TFLCs, fostering B cell activation, differentiation of plasma cells, and IgG antibody production targeting paternal HLA I molecules. These antibodies prevent the interaction between maternal naïve CD8⁺ T cells and paternal HLA I molecules on trophoblast cells, inhibiting CD8⁺ T cell activation, cytolytic mediator production, and promoting trophoblast cell survival, contributing to the success of pregnancy (Fig. 4A–G).

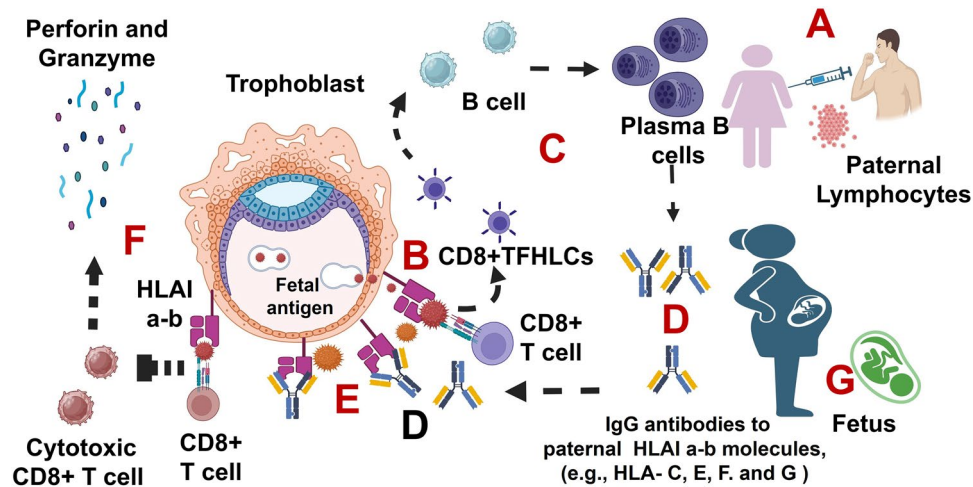


Fig. 4 The potential therapeutic approach of enhancing anti-paternal HLA I IgG antibodies through paternal lymphocyte immunization for promoting successful pregnancy. **A** Paternal lymphocytes, isolated from the peripheral blood mononuclear cells of the male partner, are administered to the female partner. **B** The injected cells facilitate the process and presentation of fetal antigens by HLA I molecules on trophoblast cells to maternal CD8⁺ T cells. **C** Antigen processing leads to the differentiation of T cells into CD8⁺ T follicular-like cells (CD8⁺TFHLCs), fostering interaction with maternal B cells and the formation of plasma B cells. **D** Plasma B cells generate IgG antibodies to paternal HLA I a and b molecules, i.e., HLA-C, E, F, and G. **E** These antibodies target indicated paternal HLA I molecules on the trophoblast cell surface. **F** This blockade prevents the interaction between maternal naïve CD8⁺ T cells and paternal HLA I a and b molecules on trophoblast cells, inhibiting CD8⁺ T cell activation and the production of cytolytic mediators, such as perforin and granzymes. **G** Consequently, this protective mechanism promotes the survival of trophoblast cells, contributing to the success of pregnancy and the delivery of a healthy fetus

The maternal immune response during pregnancy is influenced by several additional factors, including immune metabolic adaptations, hormonal changes, and the impact of the maternal microbiome [318–323]. A groundbreaking study has investigated into the complexities of immune tolerance during pregnancy, revealing critical elements such as Treg cells, trophoblast HLA interactions affecting T, NK, and NKT cell activity, tryptophan metabolism, T cell apoptosis, galectins, glycodelin co-stimulatory molecules, and mixed lymphocyte blocking factors (MLR-Bf) [4–6, 22, 25, 26, 63, 76, 77, 87, 91–96, 98, 104, 294, 300, 324–335]. These findings underscore the complex web of interactions shaping immune dynamics in pregnancy, highlighting avenues for further exploration and potential therapeutic interventions.

3 Conclusion

While our current review paper primarily investigates the dynamics of HLA matching between maternal and fetal molecules and its role in the development of IgG antibodies to fetal HLA molecules, the implications extend significantly to various pregnancy-related complications. These include conditions such as preeclampsia, gestational diabetes, villitis, and preterm labor, all of which are linked to inflammation and pregnancy failure [88, 336–338]. However, investigating the role of specific IgG antibodies to HLA molecules in sustaining successful pregnancies and protecting against fetal loss in RSA patients is a complex and intriguing research area.

However, exploring the role of specific IgG antibodies to HLA molecules in sustaining successful pregnancies and protecting against fetal loss in RSA patients remains a complex and intriguing research area.

Further studies focusing on the specific mechanisms, antibody targets, and broader therapeutic implications are essential to uncover how the axis involving HLA-CD8TFLCs mediates the development of IgG antibodies to specific paternal HLA molecules. This research could potentially shield the fetus from maternal immune responses in women with normal pregnancy and in response to lymphocyte immunization in RSA patients. Addressing these aspects through future studies will contribute to a more comprehensive understanding of the immune processes involved in maternal-fetal interactions. This deeper understanding holds the potential to advance diagnostic and therapeutic strategies aimed at maintaining successful pregnancies and addressing RSA, thereby advancing knowledge in this crucial field.

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Author contributions MKP has authored the manuscript, from conceptualization to meticulous preparation, thorough literature review, and creation of figures, ensuring clarity and coherence. MKP has been responsible for editing, refining, and guaranteeing the manuscript adheres to high academic standards before finalization.

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Data availability No datasets were generated or analysed during the current study.

Declarations

Competing interests The authors declare no competing interest related to the present work.

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References

1. Mor G, Cardenas I. The immune system in pregnancy: a unique complexity. *Am J Reprod Immunol*. 2010;63:425–33.
2. Mor G, Aldo P, Alvero AB. The unique immunological and microbial aspects of pregnancy. *Nat Rev Immunol*. 2017;17:469–82.
3. Trowsdale J, Betz AG. Mother's little helpers: mechanisms of maternal-fetal tolerance. *Nat Immunol*. 2006;7:241–6.
4. Pandey MK, Thakur S, Agrawal S. Lymphocyte immunotherapy and its probable mechanism in the maintenance of pregnancy in women with recurrent spontaneous abortion. *Arch Gynecol Obstet*. 2004;269:161–72.
5. Pandey MK, Agrawal S. Induction of MLR-Bf and protection of fetal loss: a current double blind randomized trial of paternal lymphocyte immunization for women with recurrent spontaneous abortion. *Int Immunopharmacol*. 2004;4:289–98.
6. Pandey MK, Rani R, Agrawal S. An update in recurrent spontaneous abortion. *Arch Gynecol Obstet*. 2005;272:95–108.
7. Practice Committee of the American Society for Reproductive Medicine. Evaluation and treatment of recurrent pregnancy loss: a committee opinion. *Fertil Steril*. 2012;98:1103–11.
8. Bender AR, et al. ESHRE guideline: recurrent pregnancy loss. *Hum Reprod Open*. 2018;2018:hoy004.
9. Dimitriadis E, Menkhorst E, Saito S, Kutteh WH, Brosens JJ. Recurrent pregnancy loss. *Nat Rev Dis Primers*. 2020;6:98.
10. du Fossé NA, van der Hoorn MP, van Lith JMM, le Cessie S, Lashley E. Advanced paternal age is associated with an increased risk of spontaneous miscarriage: a systematic review and meta-analysis. *Hum Reprod Update*. 2020;26:650–69.
11. Vaiman D. Genetic regulation of recurrent spontaneous abortion in humans. *Biomed J*. 2014. <https://doi.org/10.4103/2319-4170.133777>.
12. Zhu D, et al. Chromosomal abnormalities in recurrent pregnancy loss and its association with clinical characteristics. *J Assist Reprod Genet*. 2023;40:1713–20.
13. Hosseini S, Shokri F, Pour SA, Khoshnoodi J, Jeddi-Tehrani M, Zarnani AH. Diminished frequency of menstrual and peripheral blood NKT-like cells in patients with unexplained recurrent spontaneous abortion and infertile women. *Reprod Sci*. 2019;26:97–108.
14. Nigro G, Mazzocco M, Mattia E, Di Renzo GC, Carta G, Anceschi MM. Role of the infections in recurrent spontaneous abortion. *J Matern Fetal Neonatal Med*. 2011;24:983–9.
15. Nielsen A, et al. Maternal smoking predicts the risk of spontaneous abortion. *Acta Obstet Gynecol Scand*. 2006;85:1057–65.
16. Rouse CE, et al. Spontaneous abortion and ectopic pregnancy: case definition and guidelines for data collection, analysis, and presentation of maternal immunization safety data. *Vaccine*. 2017;35:6563–74.
17. Dominguez-Rojas V, de Juanes-Pardo JR, Astasio-Arbiza P, Ortega-Molina P, Gordillo-Florencio E. Spontaneous abortion in a hospital population: are tobacco and coffee intake risk factors? *Eur J Epidemiol*. 1994;10:665–8.
18. Ng KYB, et al. Systematic review and meta-analysis of female lifestyle factors and risk of recurrent pregnancy loss. *Sci Rep*. 2021;11:7081.
19. D'Ippolito S, et al. The pathogenic role of autoantibodies in recurrent pregnancy loss. *Am J Reprod Immunol*. 2020;83: e13200.
20. Bahar AM, et al. Antibodies to phospholipids and nuclear antigens in non-pregnant women with unexplained spontaneous recurrent abortions. *J Reprod Immunol*. 1993;24:213–22.
21. Andalib A, Rezaie A, Oreizy F, Shafiei K, Baluchi S. A study on stress, depression and NK cytotoxic potential in women with recurrent spontaneous abortion. *Iran J Allergy Asthma Immunol*. 2006;5:9–16.
22. Marsili L, Magnusen AF, Trivedi VS, Slavotinek AM, Pandey MK. Embracing the science of motherhood: pregnancy's transformative effects on the central nervous system and the radiance of maternal hormones and immune responses. *Discov Med*. 2023;35:673–96.
23. Günther V, et al. Live birth rates after active immunization with partner lymphocytes. *Biomedicines*. 2021;9:1350.
24. Meng L, et al. The effects of LIT and MLR-Bf on immune biomarkers and pregnancy outcomes in women with previous early recurrent miscarriage: a retrospective study. *Front Immunol*. 2021;12: 642120.
25. Agrawal S, Pandey MK, Mandal S, Mishra L, Agarwal S. Humoral immune response to an allogenic foetus in normal fertile women and recurrent aborters. *BMC Pregnancy Childbirth*. 2002;2:6.
26. Agrawal S, Pandey MK, Pandey A. Prevalence of MLR blocking antibodies before and after immunotherapy. *J Hematother Stem Cell Res*. 2000;9:257–62.
27. Aslanian-kalkhoran L, et al. The effect of lymphocyte immunotherapy (LIT) in modulating immune responses in patients with recurrent pregnancy loss (RPL). *Int Immunopharmacol*. 2023;121: 110326.
28. Chen J, et al. Effect of immunotherapy on patients with unexplained recurrent spontaneous abortion. *Ann Palliat Med*. 2020;9:2545–50.
29. Hajipour H, et al. Lymphocytes immunotherapy for preserving pregnancy: mechanisms and challenges. *Am J Reprod Immunol*. 2018;80: e12853.
30. Bainbridge DRJ. The evolution of pregnancy. *Early Human Dev*. 2014;90:741–5.
31. Bouariu A, Panaitescu AM, Nicolaidis KH. First trimester prediction of adverse pregnancy outcomes-identifying pregnancies at risk from as early as 11–13 weeks. *Medicina (Kaunas)*. 2022;58:332.
32. Clift D, Schuh M. Restarting life: fertilization and the transition from meiosis to mitosis. *Nat Rev Mol Cell Biol*. 2013;14:549–62.
33. Familiari G, Heyn R, Relucenti M, Nottola SA, Sathananthan AH. Ultrastructural dynamics of human reproduction, from ovulation to fertilization and early embryo development. *Int Rev Cytol*. 2006;249:53–141.
34. Amargant F, et al. The human sperm basal body is a complex centrosome important for embryo preimplantation development. *Mol Hum Reprod*. 2021;27:gaab062.
35. Castillo J, Jodar M, Oliva R. The contribution of human sperm proteins to the development and epigenome of the preimplantation embryo. *Hum Reprod Update*. 2018;24:535–55.
36. Familiari G, Heyn R, Relucenti M, Sathananthan H. Structural changes of the zona pellucida during fertilization and embryo development. *FBL*. 2008;13:6730–51.
37. Gerri C, et al. Initiation of a conserved trophoblast program in human, cow and mouse embryos. *Nature*. 2020;587:443–7.
38. Coticchio G, Lagalla C, Sturmey R, Pennetta F, Borini A. The enigmatic morula: mechanisms of development, cell fate determination, self-correction and implications for ART. *Hum Reprod Update*. 2019;25:422–38.
39. Roberts RM, et al. Syncytins expressed in human placental trophoblast. *Placenta*. 2021;113:8–14.

40. Hemberger M, Hanna CW, Dean W. Mechanisms of early placental development in mouse and humans. *Nat Rev Genet.* 2020;21:27–43.
41. Knöfler M, Haider S, Saleh L, Pollheimer J, Gamage TK, James J. Human placenta and trophoblast development: key molecular mechanisms and model systems. *Cell Mol Life Sci.* 2019;76:3479–96.
42. Chang C-W, Wakeland AK, Parast MM. Trophoblast lineage specification, differentiation and their regulation by oxygen tension. *J Endocrinol.* 2018;236:R43–56.
43. Gamage TK, Chamley LW, James JL. Stem cell insights into human trophoblast lineage differentiation. *Hum Reprod Update.* 2017;23:77–103.
44. Turco MY, et al. Trophoblast organoids as a model for maternal–fetal interactions during human placentation. *Nature.* 2018;564:263–7.
45. Horii M, Touma O, Bui T, Parast MM. Modeling human trophoblast, the placental epithelium at the maternal fetal interface. *Reproduction.* 2020;160:R1–11.
46. Staud F, Karahoda R. Trophoblast: the central unit of fetal growth, protection and programming. *Int J Biochem Cell Biol.* 2018;105:35–40.
47. Carrasco-Wong I, et al. Syncytiotrophoblast stress in early onset preeclampsia: the issues perpetuating the syndrome. *Placenta.* 2021;113:57–66.
48. Pijnenborg R, Vercruyse L, Hanssens M. The uterine spiral arteries in human pregnancy: facts and controversies. *Placenta.* 2006;27:939–58.
49. Ji L, Brkić J, Liu M, Fu G, Peng C, Wang Y-L. Placental trophoblast cell differentiation: physiological regulation and pathological relevance to preeclampsia. *Mol Aspects Med.* 2013;34:981–1023.
50. Petroff MG, Nguyen SL, Ahn SH. Fetal-placental antigens and the maternal immune system: Reproductive immunology comes of age. *Immunol Rev.* 2022;308:25–39.
51. Medawar PB. Some immunological and endocrinological problems raised by the evolution of viviparity in vertebrates. *Symp Soc Exp Biol.* 1953;7:320–38.
52. Petroff MG. Review: fetal antigens—identity, origins, and influences on the maternal immune system. *Placenta.* 2011;32(Suppl 2):S176–81.
53. Koch CA, Platt JL. T cell recognition and immunity in the fetus and mother. *Cell Immunol.* 2007;248:12–7.
54. Medawar P. Some immunological and endocrinological problems raised by the evolution of viviparity. *SSEB N VI Evolution.* Cambridge: Cambridge University Press; 1953.
55. Billington WD. The immunological problem of pregnancy: 50 years with the hope of progress. A tribute to Peter Medawar. *J Reprod Immunol.* 2003;60:1–11.
56. Dausset J, Souillou JP. An interview with Jean Dausset. *Am J Transpl.* 2004;4:4–7.
57. Choo SY. The HLA system: genetics, immunology, clinical testing, and clinical implications. *Yonsei Med J.* 2007;48:11–23.
58. Moreso F, et al. Treatment of chronic antibody mediated rejection with intravenous immunoglobulins and rituximab: a multicenter, prospective, randomized, double-blind clinical trial. *Am J Transpl.* 2018;18:927–35.
59. Degos L. Jean Dausset a scientific pioneer: intuition and creativity for the patients (1916–2009). *Haematologica.* 2009;94:1331.
60. MHC Sequencing Consortium. Complete sequence and gene map of a human major histocompatibility complex. *Nature.* 1999;401:921–3.
61. Sabbatino F, et al. Role of human leukocyte antigen system as a predictive biomarker for checkpoint-based immunotherapy in cancer patients. *Int J Mol Sci.* 2020;21:7295.
62. Hoek M, Demmers LC, Wu W, Heck AJ. Allotype-specific glycosylation and cellular localization of human leukocyte antigen class I proteins. *J Proteome Res.* 2021;20:4518–28.
63. Tersigni C, et al. Role of human leukocyte antigens at the feto-maternal interface in normal and pathological pregnancy: an update. *Int J Mol Sci.* 2020;21:4756.
64. Albert ED, et al. Nomenclature for factors of the HLA system 1984. In: Albert ED, Baur MP, Mayr WR, editors., et al., *Histocompatibility testing: report on the ninth international histocompatibility workshop and conference held in munich, West Germany, , May 6–11, 1984 and in Vienna, Austria, May 13–15.* Berlin: Springer; 1984. p. 4–8.
65. Slatkin M. Linkage disequilibrium—understanding the evolutionary past and mapping the medical future. *Nat Rev Genet.* 2008;9:477–85.
66. Hviid TVF, Christiansen OB. Linkage disequilibrium between human leukocyte antigen (HLA) class II and HLA-G—possible implications for human reproduction and autoimmune disease. *Hum Immunol.* 2005;66:688–99.
67. Evseeva I, Nicodemus KK, Bonilla C, Tonks S, Bodmer WF. Linkage disequilibrium and age of HLA region SNPs in relation to classic HLA gene alleles within Europe. *Eur J Hum Genet.* 2010;18:924–32.
68. D'Ippolito S, et al. Human leukocyte antigen (HLA) DQ2/DQ8 prevalence in recurrent pregnancy loss women. *Autoimmun Rev.* 2016;15:638–43.
69. Kruse C, Steffensen R, Varming K, Christiansen OB. A study of HLA-DR and -DQ alleles in 588 patients and 562 controls confirms that HLA-DRB1*03 is associated with recurrent miscarriage. *Hum Reprod.* 2004;19:1215–21.
70. Thomsen CK, et al. HLA-DRB1 polymorphism in recurrent pregnancy loss: new evidence for an association to HLA-DRB1*07. *J Reprod Immunol.* 2021;145: 103308.
71. Triggianese P, et al. Human leukocyte antigen (HLA) typing study identifies maternal DQ2 susceptibility alleles among infertile women: potential associations with autoimmunity and micronutrients. *Nutrients.* 2021;13:3270.
72. Varla-Leftherioti M, et al. HLA-DQA1*0505 sharing and killer immunoglobulin-like receptors in sub fertile couples: report from the 15th international histocompatibility workshop. *Tissue Antigens.* 2010;75:668–72.
73. Aruna M, et al. Novel alleles of HLA-DQ and -DR loci show association with recurrent miscarriages among South Indian women. *Hum Reprod.* 2011;26:765–74.
74. Oikonomou G, et al. Human leukocyte antigen alleles compatibility and immunophenotypic profile associations in infertile couples. *Cureus.* 2023;15: e36584.
75. Nowak I, et al. HLA-C C1C2 heterozygosity may protect women bearing the killer immunoglobulin-like receptor AA genotype from spontaneous abortion. *J Reprod Immunol.* 2011;88:32–7.
76. Hackmon R, Pinnaduwege L, Zhang J, Lye SJ, Geraghty DE, Dunk CE. Definitive class I human leukocyte antigen expression in gestational placentation: HLA-F, HLA-E, HLA-C, and HLA-G in extravillous trophoblast invasion on placentation, pregnancy, and parturition. *Am J Reprod Immunol.* 2017;77: e12643.

77. Dahl M, Hviid TVF. Human leucocyte antigen class Ib molecules in pregnancy success and early pregnancy loss. *Hum Reprod Update*. 2012;18:92–109.
78. Geraghty DE, Koller BH, Orr HT. A human major histocompatibility complex class I gene that encodes a protein with a shortened cytoplasmic segment. *Proc Natl Acad Sci USA*. 1987;84:9145–9.
79. Koller BH, Geraghty DE, Shimizu YO, DeMars RO, Orr HT. A novel HLA class I gene expressed in resting T lymphocytes. *J Immunol* (Baltimore, Md: 1950). 1988;141:897–904.
80. Geraghty DE, Wei X, Orr HT, Koller BH. Human leukocyte antigen F (HLA-F). An expressed HLA gene composed of a class I coding sequence linked to a novel transcribed repetitive element. *The J Exp Med*. 1990;171:1–18.
81. Natarajan K, Li H, Mariuzza R, Margulies D. MHC class I molecules, structure and function. *Rev Immunogenet*. 1999;1:32–46.
82. Ploegh HL, Orr HT, Strominger JL. Major histocompatibility antigens: the human (HLA-A,-B,-C) and murine (H-2K, H-2D) class I molecules. *Cell*. 1981;24:287–99.
83. Lundegaard C, Lamberth K, Harndahl M, Buus S, Lund O, Nielsen M. NetMHC-3.0: accurate web accessible predictions of human, mouse and monkey MHC class I affinities for peptides of length 8–11. *Nucleic Acids Res*. 2008;36:W509–12.
84. Schott G, Garcia-Blanco MA. MHC class III RNA binding proteins and immunity. *RNA Biol*. 2021;18:640–6.
85. Gabriel C, et al. HLA typing by next-generation sequencing—getting closer to reality. *Tissue Antigens*. 2014;83:65–75.
86. Rodgers JR, Cook RG. MHC class Ib molecules bridge innate and acquired immunity. *Nat Rev Immunol*. 2005;5:459–71.
87. Nilsson LL, Hviid TVF. HLA Class Ib-receptor interactions during embryo implantation and early pregnancy. *Hum Reprod Update*. 2022;28:435–54.
88. Enninga EAL, et al. Upregulation of HLA-class I and II in placentas diagnosed with villitis of unknown etiology. *Reprod Sci*. 2020;27:1129–38.
89. Cornish EF, McDonnell T, Williams DJ. Chronic inflammatory placental disorders associated with recurrent adverse pregnancy outcome. *Front Immunol*. 2022;13: 825075.
90. Yang Y, et al. Advances in the study of HLA class Ib in maternal-fetal immune tolerance. *Front Immunol*. 2022;13: 976289.
91. Maejima M, Fujii T, Kozuma S, Okai T, Shibata Y, Taketani Y. Presence of HLA-G-expressing cells modulates the ability of peripheral blood mononuclear cells to release cytokines. *Am J Reprod Immunol*. 1997;38:79–82.
92. Ishitani A, Sageshima N, Hatake K. The involvement of HLA-E and -F in pregnancy. *J Reprod Immunol*. 2006;69:101–13.
93. Tripathi P, Naik S, Agrawal S. HLA-E and immunobiology of pregnancy. *Tissue Antigens*. 2006;67:207–13.
94. Tripathi P, Naik S, Agrawal S. Role of HLA-G, HLA-E and KIR2DL4 in Pregnancy. *Int J Hum Genet*. 2007;7:219–33.
95. Langkilde CH, et al. Variation in the HLA-F gene locus with functional impact is associated with pregnancy success and time-to-pregnancy after fertility treatment. *Hum Reprod*. 2020;35:705–17.
96. Agrawal S, Pandey MK. The potential role of HLA-G polymorphism in maternal tolerance to the developing fetus. *J Hematother Stem Cell Res*. 2003;12:749–56.
97. Apps R, Murphy SP, Fernando R, Gardner L, Ahad T, Moffett A. Human leucocyte antigen (HLA) expression of primary trophoblast cells and placental cell lines, determined using single antigen beads to characterize allotype specificities of anti-HLA antibodies. *Immunology*. 2009;127:26–39.
98. King A, et al. Surface expression of HLA-C antigen by human extravillous trophoblast. *Placenta*. 2000;21:376–87.
99. Xiong S, et al. Maternal uterine NK cell-activating receptor KIR2DS1 enhances placentation. *J Clin Invest*. 2013;123:4264–72.
100. Parham P, Moffett A. Variable NK cell receptors and their MHC class I ligands in immunity, reproduction and human evolution. *Nat Rev Immunol*. 2013;13:133–44.
101. Lv H, Zhou Q, Li L, Wang S. HLA-C promotes proliferation and cell cycle progression in trophoblast cells. *J Matern Fetal Neonatal Med*. 2021;34:512–8.
102. Wei XH, Orr HT. Differential expression of HLA-E, HLA-F, and HLA-G transcripts in human tissue. *Hum Immunol*. 1990;29:131–42.
103. Bhalla A, Stone PR, Liddell HS, Zanderigo A, Chamley LW. Comparison of the expression of human leukocyte antigen (HLA)-G and HLA-E in women with normal pregnancy and those with recurrent miscarriage. *Reproduction*. 2006;131:583–9.
104. Ishitani A, et al. Protein expression and peptide binding suggest unique and interacting functional roles for HLA-E, F, and G in maternal-placental immune recognition. *J Immunol*. 2003;171:1376–84.
105. Nagamatsu T, Fujii T, Matsumoto J, Yamashita T, Kozuma S, Taketani Y. Human leukocyte antigen F protein is expressed in the extra-villous trophoblasts but not on the cell surface of them. *Am J Reprod Immunol*. 2006;56:172–7.
106. Shobu T, et al. The surface expression of HLA-F on decidual trophoblasts increases from mid to term gestation. *J Reprod Immunol*. 2006;72:18–32.
107. Ellis SA, Sargent IL, Redman CW, McMichael AJ. Evidence for a novel HLA antigen found on human extravillous trophoblast and a choriocarcinoma cell line. *Immunology*. 1986;59:595–601.
108. Dilthey AT. State-of-the-art genome inference in the human MHC. *Int J Biochem Cell Biol*. 2021;131: 105882.
109. Yao Y, et al. HLA Class II Genes HLA-DRB1, HLA-DPB1, and HLA-DQB1 are associated with the antibody response to inactivated Japanese encephalitis vaccine. *Front Immunol*. 2019;10:428.
110. Shiina T, Hosomichi K, Inoko H, Kulski JK. The HLA genomic loci map: expression, interaction, diversity and disease. *J Hum Genet*. 2009;54:15–39.
111. Megiorni F, Pizzuti A. HLA-DQA1 and HLA-DQB1 in Celiac disease predisposition: practical implications of the HLA molecular typing. *J Biomed Sci*. 2012;19:1–5.
112. Furukawa H, Oka S, Shimada K, Hashimoto A, Tohma S. Human leukocyte antigen polymorphisms and personalized medicine for rheumatoid arthritis. *J Hum Genet*. 2015;60:691–6.
113. Svensson AC, Anderson G. Presence of retroelements reveal the evolutionary history of the human DR haplotypes. *Hereditas*. 1997;127:113–24.
114. Berdoz J, et al. Constitutive and induced expression of the individual HLA-DR beta and alpha chain loci in different cell types. *J Immunol* (Baltimore, Md: 1950). 1987;139:1336–41.

115. Kotsch K, Blasczyk R. The noncoding regions of HLA-DRB uncover interlineage recombinations as a mechanism of HLA diversification. *J Immunol.* 2000;165:5664–70.
116. Baek IC, Choi EJ, Shin DH, Kim HJ, Choi H, Kim TG. Allele and haplotype frequencies of human leukocyte antigen-A, -B, -C, -DRB1, -DRB3/4/5, -DQA1, -DQB1, -DPA1, and -DPB1 by next generation sequencing-based typing in Koreans in South Korea. *PLoS ONE.* 2021;16:e0253619.
117. Xu X, et al. Genetic variants in human leukocyte antigen-DP influence both hepatitis C virus persistence and hepatitis C virus F protein generation in the Chinese Han population. *Int J Mol Sci.* 2014;15:9826–43.
118. Unanue ER, Turk V, Neeffjes J. Variations in MHC class II antigen processing and presentation in health and disease. *Annu Rev Immunol.* 2016;34:265–97.
119. Sadegh-Nasseri S, Kim A. Exogenous antigens bind MHC class II first, and are processed by cathepsins later. *Mol Immunol.* 2015;68:81–4.
120. Bach FH, Amos DB. Hu-1: Major histocompatibility locus in man. *Science.* 1967;156:1506–8.
121. Solheim B, Fuks A, Smith L, Strominger J, Thorsby E. Possible detection of HLA-DR alloantigenic specificities in man with unabsorbed rabbit antisera. *Scand J Immunol.* 1978;8:15–20.
122. Bradley B, Termijtelen A, Franks D, Van Rood J. Interpretation of data obtained from primed lymphocyte tests (PLTs). *Transpl Proc.* 1977;9:421–4.
123. Ayala García MA, González Yebra B, López Flores AL, Guaní Guerra E. The major histocompatibility complex in transplantation. *J Transpl.* 2012;2012:842141.
124. Mahdi BM. A glow of HLA typing in organ transplantation. *Clin Transl Med.* 2013;2:1–5.
125. Cohen F, Zuelzer WW, Gustafson DC, Evans MM. Mechanisms of isoimmunization. I. The transplacental passage of fetal erythrocytes in homospecific pregnancies. *Blood.* 1964;23:621–46.
126. Sebring E, Polesky H. Fetomaternal hemorrhage: incidence, risk factors, time of occurrence, and clinical effects. *Transfusion.* 1990;30:344–57.
127. Khosrotehrani K, Johnson KL, Cha DH, Salomon RN, Bianchi DW. Transfer of fetal cells with multilineage potential to maternal tissue. *JAMA.* 2004;292:75–80.
128. Sanfilippo F, Vaughn WK, Bollinger RR, Spees EK. Comparative effects of pregnancy, transfusion, and prior graft rejection on sensitization and renal transplant results. *Transplantation.* 1982;34:360–6.
129. Rebibou J-M, et al. Flow cytometric evaluation of pregnancy-induced anti-HLA immunization and blood transfusion-induced reactivation. *Transplantation.* 2002;74:537–40.
130. Dierselhuis M, Goulmy E. The relevance of minor histocompatibility antigens in solid organ transplantation. *Curr Opin Organ Transplant.* 2009;14:419–25.
131. Toldo S, Quader M, Salloum FN, Mezzaroma E, Abbate A. Targeting the innate immune response to improve cardiac graft recovery after heart transplantation: implications for the donation after cardiac death. *Int J Mol Sci.* 2016;17:958.
132. Scozzi D, Ibrahim M, Menna C, Krupnick AS, Kreisel D, Gelman AE. The role of neutrophils in transplanted organs. *Am J Transplant.* 2017;17:328–35.
133. Harmon C, Sanchez-Fueyo A, O'farrelly C, Houlihan D. Natural killer cells and liver transplantation: orchestrators of rejection or tolerance? *Am J Transpl.* 2016;16:751–7.
134. Nakamura T, Ushigome H. Myeloid-derived suppressor cells as a regulator of immunity in organ transplantation. *Int J Mol Sci.* 2018;19:2357.
135. Nakamura T, Nakao T, Yoshimura N, Ashihara E. Rapamycin prolongs cardiac allograft survival in a mouse model by inducing myeloid-derived suppressor cells. *Am J Transplant.* 2015;15:2364–77.
136. Grazia TJ, Pietra BA, Johnson ZA, Kelly BP, Plenter RJ, Gill RG. A two-step model of acute CD4 T-cell mediated cardiac allograft rejection. *J Immunol.* 2004;172:7451–8.
137. Taylor AL, Negus SL, Negus M, Bolton EM, Bradley JA, Pettigrew GJ. Pathways of helper CD4 T cell allorecognition in generating alloantibody and CD8 T cell alloimmunity. *Transplantation.* 2007;83:931–7.
138. Kreisel D, et al. The role of passenger leukocyte genotype in rejection and acceptance of rat liver allografts1. *Transplantation.* 2002;73:1501–7.
139. Golding H, Singer A. Role of accessory cell processing and presentation of shed H-2 alloantigens in allospecific cytotoxic T lymphocyte responses. *J Immunol (Baltimore, Md: 1950).* 1984;133:597–605.
140. Herrera OB, et al. A novel pathway of alloantigen presentation by dendritic cells. *J Immunol.* 2004;173:4828–37.
141. Kreisel D, et al. Non-hematopoietic allograft cells directly activate CD8+ T cells and trigger acute rejection: an alternative mechanism of allorecognition. *Nat Med.* 2002;8:233–9.
142. Harper SJ, et al. CD8 T-cell recognition of acquired alloantigen promotes acute allograft rejection. *Proc Natl Acad Sci USA.* 2015;112:12788–93.
143. Sánchez-Fueyo A, Domenig CM, Mariat C, Alexopoulos S, Zheng XX, Strom TB. Influence of direct and indirect allorecognition pathways on CD4+ CD25+ regulatory T-cell function in transplantation. *Transpl Int.* 2007;20:534–41.
144. Schenk S, et al. Alloreactive T cell responses and acute rejection of single class II MHC-disparate heart allografts are under strict regulation by CD4+ CD25+ T cells. *J Immunol.* 2005;174:3741–8.
145. Abrahimi P, et al. Blocking MHC class II on human endothelium mitigates acute rejection. *JCI insight.* 2016;1: e85293.
146. Cui J, et al. Ex vivo pretreatment of human vessels with siRNA nanoparticles provides protein silencing in endothelial cells. *Nat Commun.* 2017;8:191.
147. Srinivasan M, Domanico SZ, Kaumaya PT, Pierce SK. Peptides of 23 residues or greater are required to stimulate a high affinity class II-restricted T cell response. *Eur J Immunol.* 1993;23:1011–6.
148. O'Garra A. Cytokines induce the development of functionally heterogeneous T helper cell subsets. *Immunity.* 1998;8:275–83.
149. Elliott T, Cerundolo V, Elvin J, Townsend A. Peptide-induced conformational change of the class I heavy chain. *Nature.* 1991;351:402–6.
150. Peters PJ, et al. Cytotoxic T lymphocyte granules are secretory lysosomes, containing both perforin and granzymes. *J Exp Med.* 1991;173:1099–109.

151. Fehniger TA, et al. CD56bright natural killer cells are present in human lymph nodes and are activated by T cell–derived IL-2: a potential new link between adaptive and innate immunity. *Blood*. 2003;101:3052–7.
152. Moffett A, Colucci F. Uterine NK cells: active regulators at the maternal-fetal interface. *J Clin Investig*. 2014;124:1872–9.
153. Gumá M, et al. The CD94/NKG2C killer lectin-like receptor constitutes an alternative activation pathway for a subset of CD8+ T cells. *Eur J Immunol*. 2005;35:2071–80.
154. Gong H, et al. The regulation of ovary and conceptus on the uterine natural killer cells during early pregnancy. *Reprod Biol Endocrinol*. 2017;15:1–10.
155. Long W, et al. Association of maternal KIR and fetal HLA-C genes with the risk of preeclampsia in the Chinese Han population. *Placenta*. 2015;36:433–7.
156. Xiong S, et al. Maternal uterine NK cell–activating receptor KIR2DS1 enhances placentation. *J Clin Investig*. 2013;123:4264–72.
157. Kennedy PR, et al. Activating KIR2DS4 is expressed by uterine NK cells and contributes to successful pregnancy. *J Immunol*. 2016;197:4292–300.
158. Shah K, Al-Haidari A, Sun J, Kazi JU. T cell receptor (TCR) signaling in health and disease. *Signal Transduct Target Ther*. 2021;6:412.
159. Couture A, et al. HLA-Class II artificial antigen presenting cells in CD4(+) T cell-based immunotherapy. *Front Immunol*. 2019;10:1081.
160. Mosmann TR, Coffman RL. TH1 and TH2 cells: different patterns of lymphokine secretion lead to different functional properties. *Annu Rev Immunol*. 1989;7:145–73.
161. Fort MM, et al. IL-25 induces IL-4, IL-5, and IL-13 and Th2-associated pathologies in vivo. *Immunity*. 2001;15:985–95.
162. Ivanov II, et al. The orphan nuclear receptor ROR γ directs the differentiation program of proinflammatory IL-17+ T helper cells. *Cell*. 2006;126:1121–33.
163. Kolls JK, Linden A. Interleukin-17 family members and inflammation. *Immunity*. 2004;21:467–76.
164. Elsaesser H, Sauer K, Brooks DG. IL-21 is required to control chronic viral infection. *Science*. 2009;324:1569–72.
165. Harker JA, Lewis GM, Mack L, Zuniga EI. Late interleukin-6 escalates T follicular helper cell responses and controls a chronic viral infection. *Science*. 2011;334:825–9.
166. O’Shea JJ, Paul WE. Mechanisms underlying lineage commitment and plasticity of helper CD4+ T cells. *Science*. 2010;327:1098–102.
167. Machhi J, et al. Harnessing regulatory T cell neuroprotective activities for treatment of neurodegenerative disorders. *Mol Neurodegener*. 2020;15:32–32.
168. Sommer A, Winner B, Prots I. The Trojan horse - neuroinflammatory impact of T cells in neurodegenerative diseases. *Mol Neurodegener*. 2017;12:78.
169. Solleiro-Villavicencio H, Rivas-Arancibia S. Effect of chronic oxidative stress on neuroinflammatory response mediated by CD4+T cells in neurodegenerative diseases. *Front Cell Neurosci*. 2018;12:114.
170. Jankovic D, Kugler DG, Sher A. IL-10 production by CD4+ effector T cells: a mechanism for self-regulation. *Mucosal Immunol*. 2010;3:239–46.
171. Sher A, Coffman RL. Regulation of immunity to parasites by T cells and T cell-derived cytokines. *Annu Rev Immunol*. 1992;10:385–409.
172. Romagnani S. Lymphokine production by human T cells in disease states. *Annu Rev Immunol*. 1994;12:227–57.
173. Bottomly K. A functional dichotomy in CD4+ T lymphocytes. *Immunol Today*. 1988;9:268–74.
174. Murphy CA, et al. Divergent pro- and antiinflammatory roles for IL-23 and IL-12 in joint autoimmune inflammation. *J Exp Med*. 2003;198:1951–7.
175. Langrish CL, et al. IL-23 drives a pathogenic T cell population that induces autoimmune inflammation. *J Exp Med*. 2005;201:233–40.
176. Bettelli E, et al. Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory T cells. *Nature*. 2006;441:235–8.
177. Sojka DK, Fowell DJ. Regulatory T cells inhibit acute IFN- γ synthesis without blocking T-helper cell type 1 (Th1) differentiation via a compartmentalized requirement for IL-10. *Proc Natl Acad Sci U S A*. 2011;108:18336–41.
178. Amarnath S, et al. The PDL1-PD1 axis converts human TH1 cells into regulatory T cells. *Sci Transl Med*. 2011;3: 111ra120.
179. Densmore TL, Tim Goodnough L, Ali S, Dynis M, Chaplin H. Prevalence of HLA sensitization in female apheresis donors. *Transfusion*. 1999;39:103–6.
180. MacLennan S, et al. Prevalence of HLA and HNA antibodies in donors: correlation with pregnancy and transfusion history. *Vox Sang*. 2004;87:S2–16.
181. Powers A, Stowell CP, Dzik WH, Saidman SL, Lee H, Makar RS. Testing only donors with a prior history of pregnancy or transfusion is a logical and cost-effective transfusion-related acute lung injury prevention strategy. *Transfusion*. 2008;48:2549–58.
182. Slimane M, et al. Maternal HLA-G*01:01:01:04 protects from anti-HLA-class II immunization in pregnant women. *Hum Immunol*. 2019;80:120–5.
183. Masson E, et al. Incidence and risk factors of anti-HLA immunization after pregnancy. *Hum Immunol*. 2013;74:946–51.
184. Rizzo R, et al. Soluble human leukocyte antigen-G isoforms in maternal plasma in early and late pregnancy. *Am J Reprod Immunol*. 2009;62:320–38.
185. Steinborn A, Varkonyi T, Scharf A, Bahlmann F, Klee A, Sohn C. Early detection of decreased soluble HLA-G levels in the maternal circulation predicts the occurrence of preeclampsia and intrauterine growth retardation during further course of pregnancy. *Am J Reprod Immunol*. 2007;57:277–86.
186. Persson G, et al. Maternal HLA Ib polymorphisms in pregnancy allo-immunization. *Front Immunol*. 2021;12: 657217.
187. Morin-Papunen L, Tiilikainen A, Hartikainen-Sorri AL. Maternal HLA immunization during pregnancy: presence of anti HLA antibodies in half of multigravidous women. *Med Biol*. 1984;62:323–5.
188. Wang W, Sung N, Gilman-Sachs A, Kwak-Kim J. T Helper (Th) cell profiles in pregnancy and recurrent pregnancy losses: Th1/Th2/Th9/Th17/Th22/Tfh cells. *Front Immunol*. 2020;11:2025.
189. Yang X, Tian Y, Zheng L, Luu T, Kwak-Kim J. The update immune-regulatory role of pro- and anti-inflammatory cytokines in recurrent pregnancy losses. *Int J Mol Sci*. 2022;24:132.
190. Heaton J, Shippey S, Macri C, Macedonia C. Intestinal helminthes infestation in pregnancy: a case report and literature review. *Mil Med*. 2002;167:954–5.

191. Marbán-Castro E, Goncá A, Fumadó V, Romero-Acevedo L, Bardají A. Zika virus infection in pregnant women and their children: a review. *Eur J Obstet Gynecol Reprod Biol.* 2021;265:162–8.
192. Lu D, Peng Q, Chen D, Chen X, Jiang M. Expression imbalance of IL-17/IL-35 in peripheral blood and placental tissue of pregnant women in preeclampsia. *Taiwan J Obstet Gynecol.* 2020;59:409–14.
193. Pourakbari R, et al. Preeclampsia-derived exosomes imbalance the activity of Th17 and treg in PBMCs from healthy pregnant women. *Reprod Sci.* 2022. <https://doi.org/10.1007/s43032-022-01059-x>.
194. Simionescu AA, Danciu BM, Stanescu AMA. State-of-the-art review of pregnancy-related psoriasis. *Medicina (Kaunas).* 2021;57:804.
195. Roomandeh N, et al. Comparing serum levels of Th17 and treg cytokines in women with unexplained recurrent spontaneous abortion and fertile women. *Iran J Immunol.* 2018;15:59–67.
196. Aggarwal R, Jain AK, Mittal P, Kohli M, Jawanjal P, Rath G. Association of pro- and anti-inflammatory cytokines in preeclampsia. *J Clin Lab Anal.* 2019;33: e22834.
197. Szarka A, Rigó J Jr, Lázár L, Beko G, Molvarec A. Circulating cytokines, chemokines and adhesion molecules in normal pregnancy and preeclampsia determined by multiplex suspension array. *BMC Immunol.* 2010;11:59.
198. Rabiú OR, Dada-Adegbola H, Falade CO, Arinola OG, Odaibo AB, Ademowo OG. Serum cytokine profile of pregnant women with malaria, intestinal helminths and HIV infections in Ibadan. *Nigeria Parasitol Res.* 2022;121:1983–92.
199. True H, Blanton M, Sureshchandra S, Messaoudi I. Monocytes and macrophages in pregnancy: the good, the bad, and the ugly. *Immunol Rev.* 2022;308:77–92.
200. Mofenson LM. Risk of HIV acquisition during pregnancy and postpartum: a call for action. *J Infect Dis.* 2018;218:1–4.
201. Chetty T, Vandormael A, Thorne C, Coutsooudis A. Incident HIV during pregnancy and early postpartum period: a population-based cohort study in a rural area in KwaZulu-Natal, South Africa. *BMC Pregnancy Childbirth.* 2017;17:248.
202. la Cour Freiesleben N, et al. SARS-CoV-2 in first trimester pregnancy: a cohort study. *Hum Reprod.* 2021;36:40–7.
203. Chan LM, Lin HH, Hsiao SM. Successful treatment of maternal listeria monocytogenes bacteremia in the first trimester of pregnancy: a case report and literature review. *Taiwan J Obstet Gynecol.* 2018;57:462–3.
204. Clark RL. Safety of treating malaria with artemisinin-based combination therapy in the first trimester of pregnancy. *Reprod Toxicol.* 2022;111:204–10.
205. Vazquez-Alejo E, et al. SARS-CoV2 infection during pregnancy causes persistent immune abnormalities in women without affecting the newborns. *Front Immunol.* 2022;13: 947549.
206. Yang F, Zheng Q, Jin L. Dynamic function and composition changes of immune cells during normal and pathological pregnancy at the maternal-fetal interface. *Front Immunol.* 2019;10:2317.
207. Arenas-Hernandez M, et al. Effector and activated T cells induce preterm labor and birth that is prevented by treatment with progesterone. *J Immunol.* 2019;202:2585–608.
208. Leng Y, et al. Are B cells altered in the decidua of women with preterm or term labor? *Am J Reprod Immunol.* 2019;81: e13102.
209. Solano ME. Decidual immune cells: guardians of human pregnancies. *Best Pract Res Clin Obstet Gynaecol.* 2019;60:3–16.
210. Bulmer JN, Williams PJ, Lash GE. Immune cells in the placental bed. *Int J Dev Biol.* 2009;54:281–94.
211. Faas MM, De Vos P. Innate immune cells in the placental bed in healthy pregnancy and preeclampsia. *Placenta.* 2018;69:125–33.
212. Sun L, Su Y, Jiao A, Wang X, Zhang B. T cells in health and disease. *Signal Transduct Target Ther.* 2023;8:235.
213. Lei X, et al. CD4+ T cells produce IFN- γ to license cDC1s for induction of cytotoxic T-cell activity in human tumors. *Cell Mol Immunol.* 2024;21:374–92.
214. Pandey MK, Rani R, Zhang W, Setchell K, Grabowski GA. Immunological cell type characterization and Th1-Th17 cytokine production in a mouse model of Gaucher disease. *Mol Genet Metab.* 2012;106:310–22.
215. Pandey MK, et al. Complement drives glucosylceramide accumulation and tissue inflammation in Gaucher disease. *Nature.* 2017;543:108–12.
216. Tezabwala B, Johnson P, Rees R. Inhibition of pregnancy viability in mice following IL-2 administration. *Immunology.* 1989;67:115.
217. Mattsson R, Holmdahl R, Scheynius A, Bernadotte F, Mattsson A, Van der Meide P. Placental MHC class I antigen expression is induced in mice following in vivo treatment with recombinant interferon-gamma. *J Reprod Immunol.* 1991;19:115–29.
218. Druckmann R, Druckmann M-A. Progesterone and the immunology of pregnancy. *J Steroid Biochem Mol Biol.* 2005;97:389–96.
219. Lin H, Mosmann TR, Guilbert L, Tuntipopipat S, Wegmann TG. Synthesis of T helper 2-type cytokines at the maternal-fetal interface. *J Immunol.* 1993;151:4562–73.
220. Marzi M, et al. Characterization of type 1 and type 2 cytokine production profile in physiologic and pathologic human pregnancy. *Clin Exp Immunol.* 1996;106:127–33.
221. Tranchot-Diallo J, et al. Modulations of cytokine expression in pregnant women. *Am J Reprod Immunol.* 1997;37:215–26.
222. Gieseck RL, Wilson MS, Wynn TA. Type 2 immunity in tissue repair and fibrosis. *Nat Rev Immunol.* 2018;18:62–76.
223. Zhu J. T helper 2 (Th2) cell differentiation, type 2 innate lymphoid cell (ILC2) development and regulation of interleukin-4 (IL-4) and IL-13 production. *Cytokine.* 2015;75:14–24.
224. Chaouat G, et al. IL-10 prevents naturally occurring fetal loss in the CBA x DBA/2 mating combination, and local defect in IL-10 production in this abortion-prone combination is corrected by in vivo injection of IFN- τ . *J Immunol (Baltimore, Md: 1950).* 1995;154:4261–8.
225. Mills KHG. IL-17 and IL-17-producing cells in protection versus pathology. *Nat Rev Immunol.* 2023;23:38–54.
226. Huangfu L, Li R, Huang Y, Wang S. The IL-17 family in diseases: from bench to bedside. *Signal Transduct Target Ther.* 2023;8:402.
227. Pourakbari R, et al. Preeclampsia-derived exosomes imbalance the activity of Th17 and Treg in PBMCs from healthy pregnant women. *Reprod Sci.* 2023;30:1186–97.
228. Guerin LR, Prins JR, Robertson SA. Regulatory T-cells and immune tolerance in pregnancy: a new target for infertility treatment? *Hum Reprod Update.* 2009;15:517–35.
229. Wan YY. Regulatory T cells: immune suppression and beyond. *Cell Mol Immunol.* 2010;7:204–10.
230. Sakaguchi S, Yamaguchi T, Nomura T, Ono M. Regulatory T cells and immune tolerance. *Cell.* 2008;133:775–87.
231. Burt TD. Fetal regulatory T cells and peripheral immune tolerance in utero: implications for development and disease. *Am J Reprod Immunol.* 2013;69:346–58.

232. Copley HC, Elango M, Kosmoliaptsis V. Assessment of human leukocyte antigen immunogenicity: current methods, challenges and opportunities. *Curr Opin Organ Transplant*. 2018;23:477–85.
233. Kim JJ, et al. Molecular HLA mismatching for prediction of primary humoral alloimmunity and graft function deterioration in paediatric kidney transplantation. *Front Immunol*. 2023;14:1092335.
234. Karahan GE, Claas FHJ, Heidt S. Pre-existing alloreactive T and B cells and their possible relevance for pre-transplant risk estimation in kidney transplant recipients. *Front Med (Lausanne)*. 2020;7:340.
235. Conlon TM, et al. Germinal center alloantibody responses are mediated exclusively by indirect-pathway CD4 T follicular helper cells. *J Immunol*. 2012;188:2643–52.
236. Cyster JG, Allen CDC. B cell responses: cell interaction dynamics and decisions. *Cell*. 2019;177:524–40.
237. Ise W, et al. T follicular helper cell-germinal center B cell interaction strength regulates entry into plasma cell or recycling germinal center cell fate. *Immunity*. 2018;48:702–715.e4.
238. Demetris AJ, et al. 2016 comprehensive update of the Banff working group on liver allograft pathology: introduction of antibody-mediated rejection. *Am J Transplant*. 2016;16:2816–35.
239. McCaughan J, Xu Q, Tinkam K. Detecting donor-specific antibodies: the importance of sorting the wheat from the chaff. *Hepatobiliary Surg Nutr*. 2019;8:37–52.
240. Yue W, Liu J, Li X, Wang L, Li J. Memory B cells and long-lived plasma cells in AMR. *Ren Fail*. 2022;44:1604–14.
241. Berry GJ, et al. The 2013 International Society for Heart and Lung Transplantation Working Formulation for the standardization of nomenclature in the pathologic diagnosis of antibody-mediated rejection in heart transplantation. *J Heart Lung Transpl*. 2013;32:1147–62.
242. Levine DJ, et al. Antibody-mediated rejection of the lung: a consensus report of the International Society for Heart and Lung Transplantation. *J Heart Lung Transplant*. 2016;35:397–406.
243. Clerkin KJ, et al. Donor-specific anti-HLA antibodies with antibody-mediated rejection and long-term outcomes following heart transplantation. *J Heart Lung Transplant*. 2017;36:540–5.
244. Beyzaei Z, Geramizadeh B, Bagheri Z, Karimzadeh S, Shojazadeh AD. Novo donor specific antibody and long-term outcome after liver transplantation: a systematic review and meta-analysis. *Front Immunol*. 2020;11: 613128.
245. Mine KL, et al. Heightened expression of HLA-DQB1 and HLA-DQB2 in pre-implantation biopsies predicts poor late kidney graft function. *Hum Immunol*. 2018;79:594–601.
246. Stevanovic S, et al. HLA class II upregulation during viral infection leads to HLA-DP-directed graft-versus-host disease after CD4+ donor lymphocyte infusion. *Blood*. 2013;122:1963–73.
247. Rose ML, Coles MI, Griffin RJ, Pomerance A, Yacoub MH. Expression of class I and class II major histocompatibility antigens in normal and transplanted human heart. *Transplantation*. 1986;41:776–80.
248. Fuggle SV, McWhinnie DL, Chapman JR, Taylor HM, Morris PJ. Sequential analysis of HLA-class II antigen expression in human renal allografts. Induction of tubular class II antigens and correlation with clinical parameters. *Transplantation*. 1986;42:144–50.
249. Steinhoff G, Wonigeit K, Pichlmayr R. Analysis of sequential changes in major histocompatibility complex expression in human liver grafts after transplantation. *Transplantation*. 1988;45:394–401.
250. Sablik KA, et al. Chronic-active antibody-mediated rejection with or without donor-specific antibodies has similar histomorphology and clinical outcome—a retrospective study. *Transpl Int*. 2018;31:900–8.
251. Wozniak LJ, et al. Donor-specific HLA antibodies are associated with late allograft dysfunction after pediatric liver transplantation. *Transplantation*. 2015;99:1416–22.
252. Roux A, et al. Characteristics of donor-specific antibodies associated with antibody-mediated rejection in lung transplantation. *Front Med (Lausanne)*. 2017;4:155.
253. Beer AE, Quebbeman JF, Ayers JW, Haines RF. Major histocompatibility complex antigens, maternal and paternal immune responses, and chronic habitual abortions in humans. *Am J Obstet Gynecol*. 1981;141:987–99.
254. Johnson PM, Barnes RM, Risk JM, Molloy CM, Woodrow JC. Immunogenetic studies of recurrent spontaneous abortions in humans. *Exp Clin Immunogenet*. 1985;2:77–83.
255. Thomas ML, Harger JH, Wagener DK, Rabin BS, Gill TJ 3rd. HLA sharing and spontaneous abortion in humans. *Am J Obstet Gynecol*. 1985;151:1053–8.
256. Koyama M, et al. Probabilistic assessment of the HLA sharing of recurrent spontaneous abortion couples in the Japanese population. *Tissue Antigens*. 1991;37:211–7.
257. Schacter B, Weitkamp LR, Johnson WE. Parental HLA compatibility, fetal wastage and neural tube defects: evidence for a T/t-like locus in humans. *Am J Hum Genet*. 1984;36:1082–91.
258. Reznikoff-Etievant MF, et al. HLA antigen-sharing in couples with repeated spontaneous abortions and the birthweight of babies in successful pregnancies. *Am J Reprod Immunol*. 1991;25:25–7.
259. Meuleman T, Lashley LE, Dekkers OM, van Lith JM, Claas FH, Bloemenkamp KW. HLA associations and HLA sharing in recurrent miscarriage: a systematic review and meta-analysis. *Hum Immunol*. 2015;76:362–73.
260. Gharesi-Fard B, Askarinejad-Behbahani R, Behdin S. The effect of HLA-DRB1 sharing between the couples with recurrent pregnancy loss on the pregnancy outcome after leukocyte therapy. *Iran J Immunol*. 2014;11:13–20.
261. Agrawal S, et al. Outcome of pregnancy in women with recurrent spontaneous abortion following immunotherapy with allogeneic lymphocytes. *Hum Reprod*. 1995;10:2280–4.
262. Kishore R, Agarwal S, Halder A, Das V, Shukla BR, Agarwal SS. HLA sharing, anti-paternal cytotoxic antibodies and MLR blocking factors in women with recurrent spontaneous abortion. *J Obstet Gynaecol Res*. 1996;22:177–83.
263. Cerci Gurbuz B, Soyoz M, Ozkale Okyay D, Kilicaslan Ayna T, Pirim I. Comparison of anti-HLA antibody production according to gestational periods in pregnant women. *Transplant Proc*. 2017;49:464–6.
264. Van Rood JJ, Eernisse JG, Van Leeuwen A. Leucocyte Antibodies in Sera from Pregnant Women. *Nature*. 1958;181:1735–6.
265. Umapathy S, Shankarkumar A, Ramrakhiyani V, Ghosh K. Role of anti-human lymphocyte culture cytotoxic antibodies in recurrent spontaneous pregnancy loss women. *J Hum Reprod Sci*. 2011;4:17–9.

266. Orgad S, Loewenthal R, Gazit E, Sadetzki S, Novikov I, Carp H. The prognostic value of anti-paternal antibodies and leukocyte immunizations on the proportion of live births in couples with consecutive recurrent miscarriages. *Hum Reprod.* 1999;14:2974–9.
267. Meuleman T, Van Beelen E, Kaaja R, van Lith J, Claas F, Bloemenkamp K. HLA-C antibodies in women with recurrent miscarriage suggests that antibody mediated rejection is one of the mechanisms leading to recurrent miscarriage. *J Reprod Immunol.* 2016;116:28–34.
268. Liang X, Qiu T, Qiu L, Wang X, Zhao A, Lin Q. Female third party lymphocytes are effective for immunotherapy of patients with unexplained primary recurrent spontaneous abortion: a retrospective analysis of outcomes. *Eur J Contracept Reprod Health Care.* 2015;20:428–37.
269. Adachi H, Takakuwa K, Mitsui T, Ishii K, Tamura M, Tanaka K. Results of immunotherapy for patients with unexplained secondary recurrent abortions. *Clin Immunol.* 2003;106:175–80.
270. Khonina NA, Broitman EV, Shevela EY, Pasman NM, Chernykh ER. Mixed lymphocyte reaction blocking factors (MLR-Bf) as potential biomarker for indication and efficacy of paternal lymphocyte immunization in recurrent spontaneous abortion. *Arch Gynecol Obstet.* 2013;288:933–7.
271. Nonaka T, et al. Results of immunotherapy for patients with unexplained primary recurrent abortions—prospective non-randomized cohort study. *Am J Reprod Immunol.* 2007;58:530–6.
272. Mowbray J, Liddell H, Underwood J, Gibbings C, Reginald P, Beard R. Controlled trial of treatment of recurrent spontaneous abortion by immunisation with paternal cells. *The Lancet.* 1985;325:941–3.
273. Gatenby PA, et al. Treatment of recurrent spontaneous abortion by immunization with paternal lymphocytes: results of a controlled trial. *Am J Reprod Immunol.* 1993;29:88–94.
274. Liang P, et al. Comprehensive analysis of peripheral blood lymphocytes in 76 women with recurrent miscarriage before and after lymphocyte immunotherapy. *Am J Reprod Immunol.* 2012;68:164–74.
275. Wu L, et al. Alteration of Th17 and Treg cells in patients with unexplained recurrent spontaneous abortion before and after lymphocyte immunization therapy. *Reprod Biol Endocrinol.* 2014;12:74.
276. Wegener S, et al. Immunotherapy with paternal lymphocytes for recurrent miscarriages and unsuccessful in vitro fertilization treatment. *Transfus Med Hemother.* 2006;33:501–7.
277. Cavalcante MB, Sarno M, Barini R. Lymphocyte immunotherapy in recurrent miscarriage and recurrent implantation failure. *Am J Reprod Immunol.* 2021;85: e13408.
278. Li J, Gu Y, Zhang S, Ju B, Wang J. Effect of prepregnancy lymphocyte active immunotherapy on unexplained recurrent miscarriage, pregnancy success rate, and maternal-infant outcome. *Biomed Res Int.* 2021;2021:7878752.
279. Liu M, et al. Low-dose lymphocyte immunotherapy rebalances the peripheral blood Th1/Th2/Treg paradigm in patients with unexplained recurrent miscarriage. *Reprod Biol Endocrinol.* 2017;15:95.
280. Francisco PD, Tan-Lim CSC, Agcaoili-De Jesus MSL. Efficacy of lymphocyte immunotherapy in the treatment of recurrent pregnancy loss from alloimmunity: a systematic review and meta-analysis. *Am J Reprod Immunol.* 2022;88: e13605.
281. Liu Z, Xu H, Kang X, Wang T, He L, Zhao A. Allogenic lymphocyte immunotherapy for unexplained recurrent spontaneous abortion: a meta-analysis. *Am J Reprod Immunol.* 2016;76:443–53.
282. Ober C, et al. Mononuclear-cell immunisation in prevention of recurrent miscarriages: a randomised trial. *Lancet.* 1999;354:365–9.
283. Wong LF, Porter TF, Scott JR. Immunotherapy for recurrent miscarriage. *Cochrane Database Systematic Rev.* 2014. <https://doi.org/10.1002/14651858.CD000112.pub3>.
284. Malek A. Ex vivo human placenta models: transport of immunoglobulin G and its subclasses. *Vaccine.* 2003;21:3362–4.
285. Merbl Y, Zucker-Toledano M, Quintana FJ, Cohen IR. Newborn humans manifest autoantibodies to defined self molecules detected by antigen microarray informatics. *J Clin Invest.* 2007;117:712–8.
286. Simon AK, Hollander GA, McMichael A. Evolution of the immune system in humans from infancy to old age. *Proc Roy Soc B Biol Sci.* 2015;282:20143085.
287. Kumpel BM, Manoussaka M. Placental immunology and maternal alloimmune responses. *Vox Sang.* 2012;102:2–12.
288. Fu C, et al. Placental antibody transfer efficiency and maternal levels: specific for measles, coxsackievirus A16, enterovirus 71, poliomyelitis I-III and HIV-1 antibodies. *Sci Rep.* 2016;6:38874.
289. Palmeira P, Quinello C, Silveira-Lessa AL, Zago CA, Carneiro-Sampaio M. IgG placental transfer in healthy and pathological pregnancies. *J Immunol Res.* 2012;2012:1.
290. Valentine KM, et al. CD8 follicular T cells promote b cell antibody class switch in autoimmune disease. *J Immunol.* 2018;201:31–40.
291. Leong YA, et al. CXCR5+ follicular cytotoxic T cells control viral infection in B cell follicles. *Nat Immunol.* 2016;17:1187–96.
292. He R, et al. Follicular CXCR5-expressing CD8+ T cells curtail chronic viral infection. *Nature.* 2016;537:412–6.
293. Xing J, et al. CXCR5+ CD8+ T cells infiltrate the colorectal tumors and nearby lymph nodes, and are associated with enhanced IgG response in B cells. *Exp Cell Res.* 2017;356:57–63.
294. Pandey MK, Saxena V, Agrawal S. Characterization of mixed lymphocyte reaction blocking antibodies (MLR-Bf) in human pregnancy. *BMC Pregnancy Childbirth.* 2003;3:2.
295. Guleria I, Sayegh MH. Maternal acceptance of the fetus: true human tolerance. *J Immunol.* 2007;178:3345–51.
296. Mor G, Abrahams VM. Potential role of macrophages as immunoregulators of pregnancy. *Reprod Biol Endocrinol.* 2003;1:1–8.
297. von Rango U. Fetal tolerance in human pregnancy—a crucial balance between acceptance and limitation of trophoblast invasion. *Immunol Lett.* 2008;115:21–32.
298. Aluvihare VR, Betz AG. The role of regulatory T cells in alloantigen tolerance. *Immunol Rev.* 2006;212:330–43.
299. Kallikourdis M, Betz AG. Periodic accumulation of regulatory T cells in the uterus: preparation for the implantation of a semi-allogeneic fetus? *PLoS ONE.* 2007;2: e382.
300. Terness P, Kallikourdis M, Betz AG, Rabinovich GA, Saito S, Clark DA. Tolerance signaling molecules and pregnancy: IDO, galectins, and the renaissance of regulatory T cells. *Am J Reprod Immunol.* 2007;58:238–54.
301. Makrigiannakis A, Karamouti M, Drakakis P, Loutradis D, Antsaklis A. Fetomaternal immunotolerance. *Am J Reprod Immunol.* 2008;60:482–96.
302. Betz AG. Have you seen your mother, baby.... *Science.* 2010;330:1635–6.

303. Abu-Raya B, Michalski C, Sadarangani M, Lavoie PM. Maternal immunological adaptation during normal pregnancy. *Front Immunol.* 2020;11: 575197.
304. Liu X, et al. A dynamic peripheral immune landscape during human pregnancy. *Fundam Res.* 2022. <https://doi.org/10.1016/j.fmre.2022.06.011>.
305. Nakamura T, Shirouzu T, Nakata K, Yoshimura N, Ushigome H. The Role of Major Histocompatibility Complex in Organ Transplantation- Donor Specific Anti-Major Histocompatibility Complex Antibodies Analysis Goes to the Next Stage. *Int J Mol Sci.* 2019;20:4544.
306. Ballet C, et al. Indirect CD4+ TH1 response, antidonor antibodies and diffuse C4d graft deposits in long-term recipients conditioned by donor antigens priming. *Am J Transplant.* 2009;9:697–708.
307. Loupy A, et al. The Banff 2015 kidney meeting report: current challenges in rejection classification and prospects for adopting molecular pathology. Hoboken: Wiley; 2017.
308. Kim MJ, et al. Villitis of unknown etiology is associated with a distinct pattern of chemokine up-regulation in the feto-maternal and placental compartments: implications for conjoint maternal allograft rejection and maternal anti-fetal graft-versus-host disease. *J Immunol.* 2009;182:3919–27.
309. Kim CJ, et al. The frequency, clinical significance, and pathological features of chronic chorioamnionitis: a lesion associated with spontaneous preterm birth. *Mod Pathol.* 2010;23:1000–11.
310. Carp HJ, et al. Immunization by paternal leukocytes for prevention of primary habitual abortion: results of a matched controlled trial. *Gynecol Obstet Invest.* 1990;29:16–21.
311. Beer A. Pregnancy outcome in couples with recurrent abortions following immunological evaluation and therapy. In: Sharp F, Beard RW, editors. *Early pregnancy loss: mechanisms and treatment.* London: Springer; 1988. p. 337–49.
312. Recurrent Miscarriage Immunotherapy Trialists Group, Coulam CB, Clark DA, Collins J, Scott JR, Schlesselman JS, Aoki K, Carp HJ, Cauchi MN, Lim D, Christiansen OB. Worldwide collaborative observational study and meta-analysis on allogeneic leukocyte immunotherapy for recurrent spontaneous abortion 1. *Am J Reprod Immunol.* 1994;32:55–72.
313. Yu H, Deng X, Chao L, Chen C, Han Y. Study on positive rate of blocking antibody in women with recurrent spontaneous abortion administered by route and frequency of paternal lymphocyte immunotherapy. *Zhonghua Fu Chan Ke Za Zhi.* 2013;48:903–6.
314. Recurrent Miscarriage Immunotherapy Trialists Group, et al. Worldwide collaborative observational study and meta-analysis on allogeneic leukocyte immunotherapy for recurrent spontaneous abortion 1. *Am J Reprod Immunol.* 1994;32:55–72.
315. Sarkesh A, et al. Allogeneic lymphocytes immunotherapy in female infertility: lessons learned and the road ahead. *Life Sci.* 2022;299: 120503.
316. Liu S, Gu X, Weng R. Clinical effect of lymphocyte immunotherapy on patients with unexplained recurrent spontaneous abortion. *Immun Inflamm Dis.* 2021;9:1272–8.
317. Fainboim L, Belén S, González V, Fernández P. Evaluation of paternal lymphocyte immunotherapy and potential biomarker mixed lymphocyte reaction-blocking factor in an Argentinian cohort of women with unexplained recurrent spontaneous abortion and unexplained infertility. *Am J Reprod Immunol.* 2021;86: e13422.
318. Sharma S, Rodrigues PRS, Zaher S, Davies LC, Ghazal P. Immune-metabolic adaptations in pregnancy: a potential stepping-stone to sepsis. *EBioMedicine.* 2022;86:104337.
319. Lisova KM, Kalinovska IV, Pryimak SH, Tokar PY, Varlas VN. Changes in the level of fetoplacental complex hormones in pregnant women with miscarriage. *J Med Life.* 2021;14:487–91.
320. Jukic AM, Weinberg CR, Wilcox AJ, Baird DD. Effects of early pregnancy loss on hormone levels in the subsequent menstrual cycle. *Gynecol Endocrinol.* 2010;26:897–901.
321. Sinha T, Brushett S, Prins J, Zhernakova A. The maternal gut microbiome during pregnancy and its role in maternal and infant health. *Curr Opin Microbiol.* 2023;74: 102309.
322. Di Simone N, et al. Recent insights on the maternal microbiota: impact on pregnancy outcomes. *Front Immunol.* 2020;11: 528202.
323. *EBioMedicine.* The maternal microbiome: another bridge linking mothers and infants. *EBioMedicine.* 2021;71:103602.
324. Ernerudh J, Berg G, Mjösberg J. Regulatory T helper cells in pregnancy and their roles in systemic versus local immune tolerance. *Am J Reprod Immunol.* 2011;66(Suppl 1):31–43.
325. Williams Z. Inducing tolerance to pregnancy. *N Engl J Med.* 2012;367:1159–61.
326. Ernerudh J, Berg G, Mjösberg J. Regulatory T helper cells in pregnancy and their roles in systemic versus local immune tolerance. *Am J Reprod Immunol.* 2011;66:31–43.
327. Munn DH, et al. Prevention of allogeneic fetal rejection by tryptophan catabolism. *Science.* 1998;281:1191–3.
328. Zhu BT. Development of selective immune tolerance towards the allogeneic fetus during pregnancy: role of tryptophan catabolites. *Int J Mol Med.* 2010;25:831–5.
329. Hunt JS, Vassmer D, Ferguson TA, Miller L. Fas ligand is positioned in mouse uterus and placenta to prevent trafficking of activated leukocytes between the mother and the conceptus. *J Immunol (Baltimore, Md: 1950).* 1997;158:4122–8.
330. Bulla R, Bossi F, Fischetti F, De Seta F, Tedesco F. The complement system at the fetomaternal interface. *Immunol Pregnancy.* 2005;89:149–57.
331. Blois SM, et al. A pivotal role for galectin-1 in fetomaternal tolerance. *Nat Med.* 2007;13:1450–7.
332. Mor G, Romero R, Aldo PB, Abrahams VM. Is the trophoblast an immune regulator?: The role of toll-like receptors during pregnancy. *Crit Rev TM Immunol.* 2005;25:375.
333. Blois SM, et al. Dendritic cells: key to fetal tolerance? *Biol Reprod.* 2007;77:590–8.
334. Hempstock J, Cindrova-Davies T, Jauniaux E, Burton GJ. Endometrial glands as a source of nutrients, growth factors and cytokines during the first trimester of human pregnancy: a morphological and immunohistochemical study. *Reprod Biol Endocrinol.* 2004;2:1–14.
335. Huang N, Chi H, Qiao J. Role of regulatory T cells in regulating fetal-maternal immune tolerance in healthy pregnancies and reproductive diseases. *Front Immunol.* 2020;11:1023.

336. Preda A, et al. Gestational diabetes and preterm birth: what do we know? Our experience and mini-review of the literature. *J Clin Med.* 2023;12:4572.
337. Yang Y, Wu N. Gestational diabetes mellitus and preeclampsia: correlation and influencing factors. *Front Cardiovasc Med.* 2022;9: 831297.
338. Sandsæter HL, Horn J, Rich-Edwards JW, Haugdahl HS. Preeclampsia, gestational diabetes and later risk of cardiovascular disease: women's experiences and motivation for lifestyle changes explored in focus group interviews. *BMC Pregnancy Childbirth.* 2019;19:448.

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