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A phthalimide-based ESIPT fluorescent probe for sensitive detection of Cu²⁺ in complete aqueous solution

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Abstract

A novel fluorescence-enhanced probe 2-butyl-1,3-dioxoisoindolin-4-yl picolinate (BDIP) has been developed for the detection of Cu^{2+} based on the excited-state intramolecular proton transfer (ESIPT) process. BDIP utilized a phthalimide derivative as the fluorophore and selected picolinate ester as the recognition site for Cu^{2+} . The probe displayed high selectivity, strong antiinterference ability, and a significant fluorescence enhancement effect for Cu^{2+} in phosphate buffer saline (PBS, 10 mM, pH 7.4) with the detection limit of 31 nM. BDIP also possesses the advantages of simple synthesis steps, large Stokes shift, and good water solubility. Moreover, BDIP was used for Cu^{2+} detection in real water samples, with the result being satisfactory.

 $\textbf{Keywords} \hspace{0.1cm} 3-hydroxyphthalimide \cdot Fluorescent \hspace{0.1cm} probe \cdot Copper \hspace{0.1cm} ion \hspace{0.1cm} (II) \cdot Excited-state \hspace{0.1cm} intramolecular \hspace{0.1cm} proton \hspace{0.1cm} transfer$

Introduction

Copper is the third most abundant transition metal in the human body, widely distributed in human blood and tissues. Cu²⁺ plays vital roles in physiological processes such as energy production, signal transduction, cellular oxygen transport, and activation [1-3]. Abnormal levels of Cu²⁺ in the human body can cause several diseases such as Menkes disease [4], Wilson's disease [5], Alzheimer's disease [6], Parkinson's disease [7], coronary heart disease [8], amyotrophic lateral sclerosis [9], and myelopathy [10]. In addition, due to its wide range of applications in agriculture and industrial production, Cu²⁺ is causing serious environmental problems [11, 12]. The US Environmental Protection Agency (EPA) has set the threshold limit of Cu^{2+} in drinking water to $20 \,\mu M$ [13]. Therefore, the development of sensitive and selective detection methods for Cu²⁺ is vital for human health and environmental safety.

The current methods for detecting Cu^{2+} include atomic absorption spectroscopy [14], atomic emission spectroscopy [15], inductively coupled plasma mass spectrometry [16],

Shouxin Wang shouxinwang@mail.jnmc.edu.cn electrochemical [17], and so on. However, these methods are time-consuming, unsuitable for real-time analysis, and need tedious sample preparation and expensive instruments. Compared with the above methods, fluorescent probes have received more and more attention in detecting heavy metal ions in recent years because of their advantages of low cost, easy operation, real-time analysis, and non-destructive biological imaging [18–20]. So far, many Cu²⁺ fluorescent probes have been developed [21–23], but most probes still have some limitations, such as poor selectivity, need for organic solvents, cumbersome synthesis steps, and fluorescence quenching [24–29]. Therefore, it is still highly desirable to develop a fluorescent probe for Cu²⁺ with good water solubility, simple synthesis, and excellent performance.

Organic molecules with excited-state intramolecular proton transfer (ESIPT) property usually have a relatively high Stokes shift due to their unique luminescence mechanism, effectively avoiding self-absorption and inner filter effect [30]. In recent years, they have received extensive attention in the field of fluorescent probes. Recently, a new fluorophore, 3-hydroxyphthalimide, based on the ESIPT mechanism, has been developed and used for the detection of ions and small molecules and biological imaging research [31–33]. In addition to a relatively high Stokes shift, it also has the advantages such as high fluorescence quantum yield, high optical stability, and easy synthesis [34]. In this study, a fluorescent probe BDIP for Cu^{2+} was synthesized using *N*-butyl-3-hydroxyphthalimide (BHIO) as the fluorescent

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Scheme 1 Synthetic route of probe BDIP



matrix and 2-pyridinecarbonyl as the recognition group, based on the ESIPT mechanism (Scheme 1). We reason that the 2-pyridinecarbonyl group can prohibit the ESIPT process and quench the fluorescent emission of the fluorophore BHIO by esterification. When reacts with Cu^{2+} , the BHIO can be released from probe BDIP by Cu^{2+} -promoted hydrolysis reaction of the picolinate ester and the ESIPT process is recovered, which result in fluorescent enhancement simultaneously. Compared with the representative fluorescent probe that used picolinate ester as the recognition site for Cu^{2+} , BDIP showed a larger Stokes shift, wider linear range and lower detection limit (Table S1, Supporting Information).

Experimental

Instruments and reagents

All reagents and solvents were of analytical grade which purchased from commercial suppliers, used directly in the experiment without further purification. Deionized water was used throughout all experiments. Column chromatography was performed using silica gel 200-300 mesh from Qingdao Haiyang Chemical Co., Ltd. The solutions of Na⁺, K^+ , Ca^{2+} , Mg^{2+} , Zn^{2+} , Cd^{2+} , Hg^{2+} , Co^{2+} , Ni^{2+} , Fe^{3+} and Al^{3+} were prepared from their chloride salts; the solutions of Cu²⁺, Mn²⁺, Cr³⁺, Fe²⁺, Ag⁺ and Pb²⁺ were prepared from their nitrate salts; the solution of Cu⁺ was prepared from $[Cu(MeCN)_{4}][PF_{6}]$. ¹H NMR and ¹³C NMR spectra were measured on a Bruker AV-400 spectrometer with chemical shifts reported in ppm (in DMSO- d_6 , TMS as internal standard). High-resolution mass spectra (HRMS, ESI) were taken on a 7.0 T FTICR-MS (Varian). Absorption spectra were recorded with a Shimadzu UV-2550 spectrophotometer (Japan). Fluorescence spectra measurement was performed on a Hitachi F-4600 spectrofluorimeter (Japan). The pH was measured with a Model pHs-3C meter (Shanghai, China).

Synthesis of the fluorescent matrix BHIO

A mixture of 3-hydroxyphthalic anhydride (0.50 g, 3.05 mmol) and *n*-butylamine (0.33 g, 4.58 mmol) in acetic acid (8 mL) was stirred for 6 h at 120 °C. After cooling to room temperature, the reaction solution was poured into

100 mL ice water, and the precipitate was collected by filtration, washed with water, and dried in vacuo. BHIO was obtained as a white solid (yield: 72%). ¹H NMR (400 MHz, DMSO- d_6) δ : 10.98 (s, 1H), 7.58 (dd, 1H, J = 7.2 Hz, J = 8.4 Hz), 7.25 (d, 1H, J = 6.8 Hz), 7.18 (d, 1H, J = 8.0 Hz), 3.49 (t, 2H, J = 7.2 Hz), 1.49–1.56 (m, 2H), 1.21–1.30 (m, 2H), 0.87 (t, 3H, J = 7.6 Hz); ¹³C NMR (100 MHz, DMSO- d_6) δ : 167.70, 166.65, 155.03, 135.82, 133.50, 123.14, 114.59, 113.86, 36.65, 30.00, 19.45, 13.45; ESI-HRMS (m/z) calculated for C₁₂H₁₃NNaO₃[M + Na]⁺: 242.07876, found, 242.07878.

Synthesis of the fluorescent probe BDIP

A mixture of BHIO (0.10 g, 0.46 mmol), N, N-Diisopropylethylamine (DIPEA, 0.18 g, 1.38 mmol), pyridine-2-carbonyl chloride hydrochloride (0.12 g, 0.69 mmol), and a catalytic amount of 4-Dimethylaminopyridine (DMAP) was dissolved in anhydrous dichloromethane (20 mL), and stirred at room temperature for 10 h. Then the solvent was evaporated under reduced pressure and the obtained residue was purified by silica gel chromatography with petroleum ether/ ethyl acetate (2:1, V/V) as eluent to afford the desired product as white solid (yield: 85%). ¹H NMR (400 MHz, DMSO d_6) δ : 8.84—8.86 (m, 1H), 8.28—8.30 (m, 1H), 8.10–8.15 (m, 1H), 7.95 (t, 1H, J=7.5 Hz), 7.84 (d, 1H, J=7.4 Hz), 7.75—7.80 (m, 2H), 3.50 (t, 2H, J = 7.0 Hz), 1.48–1.55 (m, 2H), 1.20–1.29 (m, 2H), 0.85 (t, 3H, J = 7.4 Hz); ¹³C NMR $(100 \text{ MHz}, \text{DMSO-}d_6) \delta$: 167.02, 165.53, 162.54, 150.21, 146.02, 145.96, 137.87, 136.56, 133.27, 128.73, 128.31, 126.03, 122.34, 121.13, 37.20, 29.85, 19.46, 13.43; ESI-HRMS (m/z) calculated for $C_{18}H_{16}N_2NaO_4$ [M + Na]⁺: 347.10023, found, 347.10040.

Spectroscopy analysis

The probe BDIP was dissolved in DMSO to prepare a 1 mM stock solution. The metal cations (K⁺, Na⁺, Ag⁺, Ca²⁺, Mg²⁺, Cu²⁺, Zn²⁺, Al³⁺, Fe³⁺, Fe²⁺, Cd²⁺, Cr³⁺, Co²⁺, Hg²⁺, Ni²⁺, Pb²⁺, and Mn²⁺) were dissolved in deionized water to prepare 2 mM stock solutions, respectively. Cu⁺ stock solution (2 mM) was prepared in MeCN. 30 μ L of the probe BDIP stock solution was mixed with 30 μ L of the metal ions stock solutions. Then the resulting solution was diluted to

3 mL with phosphate buffer saline (PBS, 10 mM, pH 7.4) and placed in a cuvette. In the anti-interference experiment, 30 μ L of other metal cations stock solutions, 30 μ L of the probe BDIP stock solution, and 30 μ L Cu²⁺ stock solution were sequentially added into the PBS. The final volume of the test solution was set at 3 mL and placed in a cuvette. The fluorescence measurements were carried out after reacting for 15 min at room temperature. The following parameters were set for measurement: $\lambda_{ex} = 344$ nm, $\lambda_{em} = 515$ nm, slit width: 5 nm and 5 nm, and voltage: 700 V.

Application in real water samples

Two real water samples were taken to validate the analytical application of the probe BDIP. One water sample was taken from tap water in the laboratory, and the other was taken from the Jian Lake of Jining Medical University. Before the tests, each sample was filtered to remove the impurities using filter paper. Then the two water samples were used to prepare PBS (10 mM, pH 7.4) and a probe BDIP stock solution (3 mM). Cu²⁺ was determined by the standard addition method. BDIP stock solutions (30 μ L) and Cu²⁺ stock solution (0, 1, 3 or 5 μ L) were placed into a cuvette and diluted to 3 mL with the two water samples, and gave the test solutions. The final mixtures contained BDIP (10 μ M) and Cu²⁺ $(0, 1, 3 \text{ or } 5 \mu \text{M})$. The test solution was incubated for 15 min at room temperature, and the fluorescence was measured at 515 nm. For each Cu²⁺ concentration, water samples were tested three times. Comparing with the standard curve, the concentrations of Cu²⁺ can be observed.

Results and discussion

Synthesis of probe BDIP

The probe BDIP was prepared with 3-hydroxyphthalic anhydride as the starting material by a two-step reaction, as shown in Scheme 1. First, the condensation reaction of 3-hydroxyphthalic anhydride with *n*-butylamine was carried out at a high temperature, and the resulting fluorescent matrix BHIO was directly precipitated from the reaction solution. Then, with DIPEA as the acid-binding agent and DMAP as a catalyst, the acylation reaction of BHIO with pyridine-2-carbonyl chloride produced the probe BDIP. The structures of BHIO and BDIP were confirmed by ¹H NMR, ¹³C NMR, and HRMS (see Figs. S1–S6).

Effect of the reaction time of probe BDIP with Cu²⁺

To obtain the best response time for Cu^{2+} detection, we investigated the effect of the reaction time on the fluorescence intensity. As shown in Fig. 1, in PBS (10 mM, pH



Fig. 1 The plot of fluorescence intensity at 515 nm versus reaction time (0-40 min)

7.4), the fluorescence intensity of the probe BDIP solution (10 μ M) was very low. When 2 equivalents of Cu²⁺ were added to the probe BDIP solution, the fluorescence intensity at 515 nm gradually increased. With the progress of the reaction, the increased rate of the fluorescence intensity gradually decreased and the fluorescence intensity remained unchanged after 15 min. Therefore, in the subsequent experiments, a reaction time of 15 min was used.

Effect of Cu²⁺ concentration

Through the Cu²⁺ fluorescence titration experiment, we investigated the sensitivity and linear range of the probe BDIP. As shown in Fig. 2A, the fluorescence emission spectra of probe BDIP (10 μ M) in the PBS at different concentrations of Cu²⁺ from 0 to 50 μ M were recorded. Upon excitation at 344 nm, the free BDIP solution showed a very weak emission peak at 515 nm. With an increase in Cu²⁺ concentration, the fluorescence intensity at 515 nm increased significantly and reached a plateau with an eightfold enhancement at the concentration of 20 μ M (Fig. 2B). The Stokes shift of probe BDIP is as high as 171 nm, which can effectively prevent self-absorption and increase the signal-to-noise ratio of fluorescence imaging.

The inset in Fig. 2B indicated a good linear relationship between the fluorescence intensity of BDIP and the concentration of Cu^{2+} , and the regression equation was y = 532.9x + 909.7 ($R^2 = 0.9938$). According to the formula for detection limit (LOD): LOD = $3\sigma/K$ (wherein, σ is the standard deviation of the blank fluorescence spectrum and *K* is the slope of the straight line), the detection limit of the probe BDIP for Cu²⁺ was 31 nM, lower than the threshold limit (20 µM) set by the US EPA. This indicated that the probe BDIP had relatively good detection sensitivity for Cu²⁺.





Fig. 3 Effect of pH on fluorescence intensity of probe BDIP (10 μ M) in the absence and presence of Cu²⁺ (20 μ M), respectively

Effect of pH on the detection performance of the probe BDIP

Considering the significance of pH in practical applications, the pH effects on the fluorescent response of probe BDIP (10 μ M) in the absence and presence of Cu²⁺ were investigated (Fig. 3). In the absence of Cu²⁺, weak fluorescence signal of probe BDIP was observed within a wide pH range from 3.0 to 11.0, indicating that the probe BDIP is relatively stable under acidic and weakly alkaline conditions. On the other hand, after adding 2 equivalents of Cu²⁺, the fluorescence was gradually increased in the region of pH 3.0–7.4 and reached a maximum at pH 7.4. When pH was further increased, the fluorescence intensity gradually decreased. Therefore, considering the pH experiment results and the biological applications, the physiological conditions (pH 7.4) were selected to detect Cu²⁺.

Selectivity and anti-interference of probe BDIP

The selectivity of the fluorescent probe is an important parameter for performance evaluation. To investigate the selectivity of the probe BDIP, We examined the fluorescence response of BDIP to various interfering cations including K⁺, Na⁺, Ag⁺, Ca²⁺, Mg²⁺, Zn²⁺, Al³⁺, Fe³⁺, Fe²⁺, Cd²⁺, Cr³⁺, Co²⁺, Hg²⁺, Ni²⁺, Pb²⁺, Cu⁺ and Mn²⁺. As shown in Fig. 4A, the fluorescence intensity of the probe BDIP (10 μ M) did not change significantly after the addition of potentially interfering cations (20 μ M), whereas, after adding two equivalents of Cu²⁺, the fluorescence intensity increased significantly, indicating that the probe BDIP had specific selectivity for Cu²⁺ and could be used for the analysis and detection of Cu²⁺.

To further investigate the Cu²⁺ detection ability of the probe BDIP in the presence of other competing ions, we also conducted anti-interference experiments. 20 μ M different metal ions and 10 μ M probe BDIP were mixed in PBS first. And then 20 μ M Cu²⁺ was added. As shown in Fig. 4B, similar fluorescence responses of probe BDIP to Cu²⁺ were observed regardless of the presence or absence of various other interfering cations. These results indicated that the probe BDIP could be used to detect Cu²⁺ in a complex environment.



Recognition mechanism of probe BDIP for Cu²⁺

According to the previously reported work [35, 36], we proposed the possible response mechanism of BDIP for Cu²⁺ (Scheme 2). Due to the presence of the 2-pyridinecarbonyl group, the ESIPT process based on the keto-enol tautomer in BHIO is inhibited, resulting in weak fluorescence of probe BDIP. When Cu²⁺ coexists with BDIP, 2-pyridinecarbonyl group in BDIP acts as a "recognition group", which brings Cu^{2+} in close proximity to the ester bond. Then the Cu^{2+} coordinates with nitrogen and neighboring carbonyl oxygen from the 2-picolinic ester. The formation of this complex promotes the hydrolysis of 2-picolinic ester which acts as an "anchoring group" [37, 38]. Once hydrolysis occurs, the fluorescent matrix BHIO is released from BDIP and Cu²⁺ is regenerated which participates in the hydrolysis process of the remaining ester. Finally, this catalytic sensing cycle causes the fluorescent signal to gradually increase. This transformation was confirmed by MS, UV-vis and emission spectroscopic studies. UV-vis and fluorescence emission spectra of Cu²⁺-treated BDIP solution were compared with those of fluorescent matrix BHIO. As shown in Fig. S7 and Fig. S8, the spectra of these two compounds were nearly identical. Furthermore, MS spectrum of Cu²⁺-treated BDIP solution showed a major peak at m/z 218.08215 (see Fig. S9), which was consistent with the deprotonated molecular weight ([M-H]⁻ 218.08227) of BHIO.

Detection of Cu²⁺ in real water samples

To validate the practical application potential of the probe BDIP, we detected Cu²⁺ in tap water and lake water under the same above conditions. The probe BDIP (10 μ M) and a certain amount of Cu^{2+} were added to the water samples. No obvious fluorescence enhancement was observed when probe BDIP was added directly to the water samples. After Cu^{2+} was added to the water sample, significant fluorescence enhancement can be observed. The recovery tests were carried out and the relative standard deviation RSD were calculated. Table 1 indicated that the recoveries of Cu²⁺ were 95.0-104.0%, and the RSD were 1.35-3.80%. This showed that the probe BDIP could be used for the quantitative detection of Cu²⁺ in real water samples with high accuracy and precision.



Table 1	Determination of Cu ²⁴
in tap w	ater and lake water

Cu²⁺

Real sample	Added Cu ²⁺ /µM	Found/µM	Recovery, %	RSD $(n=3), \%$
Tap water	0	Not detected		
	1.00	0.95	95.0	3.80
	3.00	2.96	98.7	2.25
	5.00	5.03	100.6	3.67
	0	Not detected		
Lake water	1.00	1.04	104.0	1.52
	3.00	3.09	103.0	1.35
	5.00	5.15	103.0	2.32

Conclusions

In conclusion, we developed a new ESIPT-based fluorescent probe (BDIP) for Cu^{2+} based on the specific hydrolysis reaction of the picolinate moiety. The fluorescence intensity of the probe BDIP at 515 nm increased significantly when treated with Cu^{2+} , which indicated a consequence of the ESIPT turn-on. In PBS (10 mM, pH 7.4), probe BDIP displayed high selectivity and sensitivity for Cu^{2+} with excellent anti-interference ability. Moreover, the probe was successfully utilized for the quantitative detection of Cu^{2+} in real water samples, and the results indicated that BDIP possessed potential applicability for Cu^{2+} detection in environmental samples.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s44211-022-00084-9.

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