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# Glioma-associated macrophages: unraveling their dual role in the microenvironment and therapeutic implications



Jiachen Wang<sup>1</sup>, Shenglan Li<sup>1</sup>, Yanjie Lan<sup>1</sup>, Xinrui Liu<sup>1</sup> and Wenbin Li<sup>1\*</sup>

# **Abstract**

Glioblastoma (GBM) is a malignant brain glioma characterized by a high number of tumor-associated macrophages (TAMs) within its tissues. These TAMs have a close relationship with tumor grade and prognosis. Targeting TAMs has been identifed as a promising therapeutic strategy. However, TAM cells play both tumor-killing and tumorpromoting roles, making them a double-edged sword in the immune environment. The diferent subtypes of macrophages and their efects on the tumor microenvironment remain poorly understood. This study comprehensively elucidates the immunobiology of glioma-associated macrophages (GAMs), including their origin, classifcation, molecular mechanisms underlying glioma promotion and inhibition, polarization strategies, targeted therapy for GAMs and the current challenges and perspectives in immune modulation. Further research on macrophage function and mechanism may provide a new immunological basis for treating GBM patients and enhancing the efficacy of glioma immunotherapy.

**Keywords** Glioma, Glioma-Associated macrophages, Tumor-Associated macrophages, Tumor Microenvironment, Immunotherapy

# **1 Introduction**

Macrophages are multifunctional immune cells that exhibit remarkable plasticity and adaptability to diverse tissue environments. The specific niche in which macrophages reside plays a critical role in their homeostatic diferentiation. Glioblastoma (GBM) is a highly aggressive brain tumor with a signifcant population of gliomaassociated macrophages (GAMs) within its complex tumor microenvironment (TME). The TME encompasses a diverse array of cellular and molecular constituents that dynamically interact, exerting profound efects on tumor progression and therapeutic response.

\*Correspondence: Wenbin Li

liwenbin@ccmu.edu.cn

GAMs possess a dual function, combating and promoting tumor growth. Usually, they contribute to tumor progression by promoting angiogenesis, facilitating tumor invasion and metastasis, and shaping a microenvironment favorable for tumor growth. Moreover, tumors hijack GAMs through multiple molecular mechanisms, including the regulation of macrophage polarization signals, augmented immunosuppression, epigenetic modifcations, and interactions with other components of the immune system.

Considering the essential role of GAMs in tumor development, targeted therapies aimed at GAMs have emerged as promising strategies for glioma treatment. These approaches involve reducing the number of tumorassociated macrophages (TAMs), inducing a phenotypic transformation toward an antitumor M1 polarization state, enhancing their immune regulatory functions, and utilizing various immune modulatory therapies.



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<sup>&</sup>lt;sup>1</sup> Department of Neuro-oncology, Cancer Center, Beijing Tiantan Hospital, Capital Medical University, Beijing 100071, China

This article comprehensively elucidates the intricate immunobiology of GAMs, focusing on their origins, functional classifcations and the dynamic interplay between GAMs and the TME. Furthermore, we delve into the potential of targeted therapies of GAMs, discussing strategies to decrease GAMs. By unraveling the complexities of GAMs, this study aims to pave the way for novel immunological approaches in the treatment of GBM, ultimately augmenting the efficacy of glioma immunotherapy.

# **1.1 Macrophages grow in diferent tissue environments** *1.1.1 Macrophage niche*

The macrophage niche refers to the situation in which macrophages regulate their phenotype and function in diferent tissue environments to maintain the normal structure and function of the tissue. The macrophage niche is mainly determined by the interaction between macrophages and the surrounding cells and molecules (Mass et al. [2023\)](#page-15-0). Guilliams et al. summarized the key features of the macrophage niche, highlighting four crucial aspects. First, the niche provides a physical environment and survival support for macrophages. Second, it provides essential nutritional factors for macrophages, such as CSF-1, IL-32, and CSF-2. Third, it marks the tissue specifcity of macrophages by key transcription factors. Finally, macrophages participate in the formation of environmental homeostasis (Guilliams et al. [2020](#page-14-0)). Macrophages in distinct organs or tissues exhibit unique developmental trajectories, transcriptional programs and life cycles to adapt to their respective niches and fulfll specific functional requirements. These macrophages are called tissue-specific macrophages. They play important roles in various biological processes, such as angiogenesis, adipogenesis, metabolism, and neural function, contributing to organ development, homeostasis, and the immune response (Mass et al. [2023\)](#page-15-0).

#### **1.2 Homeostatic diferentiation and steady state**

Homeostatic diferentiation refers to the process of monocytes adapting to the tissue environment and exerting benefcial functions under normal conditions (Park et al. [2022](#page-15-1)). During this process, embryonic macrophage precursors or mature monocytes express distinct tissue-specifc markers, making them resident tissue macrophages (RTMs) in various tissues, achieving a steady state (Park et al. [2022](#page-15-1)). In the central nervous system (CNS), RTMs encompass microglia and macrophages located around the dura mater, pia mater, cerebral vessels, and choroid plexus (Kierdorf et al. [2019](#page-14-1)). Under normal conditions, these RTMs play an essential role in regulating CNS homeostasis (Mundt et al. [2022\)](#page-15-2), including maintaining blood–brain barrier (BBB)

integrity (Ronaldson and Davis [2020\)](#page-15-3), promoting blood flow and cerebrospinal fluid (CSF) drainage and facilitating nutrient transport (Bordon  $2023$ ). They also actively engage in waste and impurity clearance, as well as the phagocytosis of senescent apoptotic cells (Chen et al. [2023\)](#page-13-1). Microglia also contribute to the development of the nervous system, such as the establishment of neural circuits (Fujita and Yamashita [2021\)](#page-14-2), the maintenance of synaptic plasticity and the support of oligodendrocyte activity (Zhou et al. [2019](#page-16-0)). In the face of inflammation or pathological damage, these macrophages not only constitute the frst line of defense against pathogens by activating innate immune components but can also regulate adaptive immune responses, presenting necessary antigens to the periphery and suppressing unnecessary immune responses (Muzio et al. [2021\)](#page-15-4). The dysregulation of these responses can precipitate numerous CNS diseases (Kierdorf and Prinz [2017](#page-14-3)).

Nonhomeostatic diferentiation stands in contrast to homeostatic diferentiation when disease-related signals are much stronger than steady-state signals, leading to a disruption in monocyte-macrophage diferentiation status. This disruption is usually found in the inflammatory state, but in the case of CNS tumors, the opposite efect is observed (Park et al. [2022](#page-15-1); Kierdorf and Prinz [2017](#page-14-3)). Macrophage niches play a regulatory role between these two states by reprogramming transcription in response to diferent stimulation conditions (Wculek et al. [2022\)](#page-16-1).

When glioma occurs, this homeostasis of macrophages in the CNS is disrupted. Under the action of disease-related signals released by the tumor, the macrophage niche is rewritten as a component of the TME and actively promotes tumor progression (Yin et al. [2017](#page-16-2)). Under the hijacking of the tumor, RTMs undergo nonsteady-state diferentiation toward immune suppression, becoming part of TAMs. Another part of TAMs originates from peripheral bone marrow-derived macrophages (BMDMs) (Hambardzumyan et al. [2016](#page-14-4)). Consequently, macrophage homeostasis is regulated by the tumor and surrounding environment, creating dynamic heterogeneity.

#### **1.3 GBM**

GBM is a malignant brain glioma that is highly invasive, recurrent, and drug-resistant. It is one of the most common and lethal tumors of the central nervous system (Wang et al. [2017\)](#page-15-5). Glioma stem cells (GSCs) are a subgroup of cells with stem cell characteristics that can continuously self-renew and promote GBM diferentiation into diferent subtypes (Xuan et al. [2021](#page-16-3)). Single-cell RNA sequencing (RNA-seq) and lineage tracing techniques have elucidated the impact of the TME on the transcriptional characteristics of GBM subtypes, consequently influencing the efficacy of tumor treatments (Neftel et al. [2019](#page-15-6)).

#### **1.4 The TME**

The TME is a complex system composed of tumor cells, nontumor cells and molecules, including blood vessels, immune cells, stromal cells, extracellular matrix and other signaling molecules. The components of the TME engage in dynamic interactions with each other, including angiogenesis promotion (Palma et al. [2017;](#page-13-2) Jiang et al. [2020\)](#page-14-5), the regulation of immune cell polarization, function and migration (Pijuan et al. [2019;](#page-15-7) Li et al. [2017](#page-14-6)), modulation of stromal cell phenotype and secretome (Wei et al.  $2020$ ; Cai et al.  $2021a$ , [b](#page-13-4)), and important efects on tumor growth, invasion, metastasis and treatment response. It is one of the main factors leading to tumor progression and resistance. The cellular populations of the glioma TME include macrophages/microglia, lymphocytes, astrocytes, oligodendrocytes, and neurons (Nunno et al. [2023\)](#page-13-5). Among these, TAMs emerge as the dominant cell population. TAMs have a symbiotic relationship with tumors and possess strong immunosuppressive properties that promote tumor progression. They enhance the stemness, proliferation, survival, and migration abilities of GSCs while inhibiting adaptive immune responses (Xuan et al. [2021](#page-16-3)).

Therefore, understanding the changes in macrophage homeostasis is important for studying the glioma-associated TME and identifying potential targets for glioma treatment. This article aims to explore the immunobiological characteristics of GAMs, including their origin, classifcation, function and regulation mechanisms, while delving into the symbiotic relationship between TAMs and gliomas. Finally, this review will concentrate on several therapeutic approaches that target TAMs, holding substantial promise in the quest for efective glioma treatments.

#### **2 Diferent origins of GAMs**

GAMs in the CNS originate from two primary sources: microglia located in the brain parenchyma and macrophages located around the dura mater, pia mater, brain blood vessels, and choroid plexus (Andersen et al. [2021](#page-13-6)). In the context of neuroglioma, these two types of cells occupy  $30 \sim 50\%$  of the TME (Andersen et al. [2021](#page-13-6)). However, they difer in their developmental origins, phenotypic markers, gene expression profles, and functional characteristics.

Microglia are derived from yolk sac progenitors during embryonic development and self-renew throughout life. They express unique phenotypic markers, including TMIGD3, APOC2, SCIN, P2RY12, TREM2, Sall1, Tmem119 and CX3CR1, and exhibit a distinctive gene

expression profle that difers from that of other tissue macrophages. In contrast, macrophages in the CNS are derived from bone marrow progenitor cells that migrate across the BBB and diferentiate into macrophages in response to local signals. They express different phenotypic markers, such as CD45, CD11b, and F4/80, and their gene expression profle is more closely related to that of other tissue macrophages (Pombo Antunes et al. [2021](#page-15-8); Khan et al. [2023](#page-14-7)).

The traditional method for distinguishing monocyte-derived macrophages and microglia is to use flow cytometry to identify the level of CD45 expression, but microglia in the glioma TME can also exhibit high levels of CD45 (Bian et al. [2020\)](#page-13-7). Currently, TMEM119, Siglec-H, Olfml3 and Sall1 are widely used to distinguish the two (Brandenburg et al. [2020](#page-13-8); Andersen et al. [2022](#page-13-9)). The spatial and temporal distribution of the two in glioma is diferent. Microglia-derived GAMs are mainly distributed at the tumor margin and infltration zone, accounting for 82–97% of the cells in the TME of newly diagnosed gliomas, while monocyte-derived GAMs are mainly distributed in the tumor core area and are more prominent in recurrent tumors and necrotic hypoxic areas (Pombo Antunes et al. [2021](#page-15-8)). Most of the highly invasive macrophages come from the patient's peripheral blood monocyte-macrophage system (Wang et al. [2022](#page-16-5)), and the expression of SEPP1 is closely related to tumor recurrence (Pombo Antunes et al. [2021](#page-15-8)).

# **3 Functional classifcation of GAMs**

According to the efect of GAMs on tumor growth, they can be roughly divided into two subtypes: antitumor GAMs and protumor GAMs. Antitumor GAMs are similar to M1-type macrophages and express CD68, CD40, CD86 and MHC-II molecules to promote anti-infection and infammatory immune activation.

They secrete proinflammatory factors such as TNF- $\alpha$ , IL-1β, IL-6, IL-12, IL-23, and NO, which activate the T-cell immune response and induce tumor apoptosis and necrosis (Zhang et al. [2020](#page-16-6); Li et al. [2022a](#page-14-8), [b,](#page-14-9) [c](#page-14-10), [d](#page-14-11), [e](#page-15-9), [f](#page-15-10), [g](#page-15-11)). CD169 is considered a marker of proinfammatory macrophages, and M1-like macrophages can produce CXCL9/10 and other T-cell and NK cell chemokines, enhancing the killing efect on tumors (Kim et al. [2022\)](#page-14-12) (Kim et al. [2022\)](#page-14-12). Pro-tumor GAMs are similar to M2-type macrophages, highly expressing CD163, CD204, CD206 and other molecules, secreting anti-infammatory cytokines such as IL-4, IL-10, IL-13 and TGF-β, and inhibiting the infammatory response and adaptive immune intensity. M2-like macrophages lack the key expression of T-cell costimulation activation, such as CD86, CD80, and CD40 expression, thus promoting tumor metastasis and immune escape (Zhang et al. [2020;](#page-16-6) Li et al. [2022a](#page-14-8), [b,](#page-14-9) [c](#page-14-10), [d,](#page-14-11) [e,](#page-15-9) [f,](#page-15-10) [g\)](#page-15-11). The number of M2-type macrophages

is positively correlated with the pathological grading of WHO tumors (Ding et al. [2014\)](#page-13-10).

For the sake of research convenience, GAMs are generally classifed into two categories according to their function, but in fact, the GAM population is a dynamic continuum, with phenotypic and functional heterogeneity and plasticity among its members, and is regulated by various factors in the TME. Several methods have been developed to classify GAMs, and Antunes et al. divided monocyte-derived GAMs (M0) into six types based on single-cell sequencing of human patients and mouse models (Pombo Antunes et al. [2021\)](#page-15-8).

The statement describes the six different types of monocyte-derived GAMs (M0) identifed by Antunes et al. based on single-cell sequencing. These include transitional Mo-GAMs, which are in the process of differentiating and express monocyte-related genes but have low expression of mature macrophage markers. GAMs are related to phagocytosis and lipid metabolism and express genes such as *GPNMB*, *LGALS3*, and *FABP5*. GAMs are related to hypoxia and glycolysis and express genes such as *BNIP3*, *ADAM8*, *MIF*, and *SLC2A1*. GAMs are similar to tissue-resident microglia, expressing genes such as CX3CR1, BIN1, and SCIN, with low expression of SEPP1. GAMs are potentially related to infammatory activity and highly express genes such as *SEPP1*, *SLC40A1*, *FOLR2*, *MRC1*, and *RNASE1*. GAMs with interferon-induced gene expression (Pombo Antunes et al. [2021](#page-15-8)). Yin et al. also distinguished six diferent transcriptionally and functionally distinct monocyte/microglia (Mo/Mg) subtypes, as shown in Table [1](#page-3-0) (Yin et al. [2022](#page-16-7)).

Therefore, different GAM subtypes have significant differences, which are refected in many aspects, such as immune regulation, antigen presentation, and phagocytosis function.

The distribution of GAMs in different molecular subtypes of glioma is different. The proportion of peripheralderived monocytes in IDH wild-type GBM is signifcantly higher than that in IDH-mutated grade IV glioma, and their function is mainly infammation-related (Mo-GAMinf.), and their tumors show more powerful stemness and richer angiogenesis, while IDH-mutated tumors have a higher proportion of GAMs involved in antigen presentation (Mo/Mg-GAM-APP) (Yin et al. [2022](#page-16-7)). Different GAM subgroups in the tumor core are related to the overall survival rate of IDH wild-type patients, and studies have found that when the phenotype is CD68−, CD163<sup>−</sup> and CD206+, it is more conducive to the survival of patients (Zeiner et al. [2019](#page-16-8)). In IDH-mutated anaplastic astrocytoma, the M2 marker CD168 is negatively correlated with the patient survival rate (Prosniak et al. [2013](#page-15-12)). In low-grade glioma, GAMs are mainly derived from microglia, and their function is mainly antigen presentation and immune regulation, while high-grade glioma GAMs are mainly derived from peripheral blood monocyte subtypes and are related to immune suppression and inflammation (Andersen et al. [2022\)](#page-13-9). These differences may be related to factors such as chemokines produced by diferent tumor grades, vascular permeability and necrosis area sizes.

Therefore, further understanding of the TME composition and GAM subtypes of diferent subtypes of glioma will help to identify appropriate GAM targets and determine accurate immunotherapy.

# **4 Can GAMs fght against tumors?**

M1-like GAMs are a type of immune cell with antitumor activity that are generally considered to have some antitumor ability in the early stages of tumor development. M1 macrophages can directly phagocytose or kill tumor cells

<span id="page-3-0"></span>



through antibody-dependent cytotoxicity. The glycolysis pathway is the basis for M1 macrophages to fght against tumors, and the production of ROS and phagocytosis are highly dependent on glucose supply, while afecting this pathway is the main means for tumors to "hijack" GAMs (Mantovani et al. [2022](#page-15-13)). M1 macrophages drive adaptive immunity through antigen presentation, promoting T-cell proliferation and IFN-γ secretion (Cendrowicz et al. [2021](#page-13-11)). Macrophages stimulated by IFN-γ can secrete IL-12, which is a potent antitumor proinfammatory cytokine, and restore the costimulatory activity between GAMs and T cells (Cendrowicz et al. [2021](#page-13-11)). M1 macrophages can also cause vascular damage and tumor necrosis by producing other proinfammatory factors, such as NO, TNF-α, and IL-12. These factors can activate NK cells, T cells, and B cells, forming efective adaptive immunity (Mantovani et al. [2022](#page-15-13); Peng et al. [2020](#page-15-14)). M1 macrophages can also release various extracellular vesicles (M1EVs), carry antitumor molecules, and regulate the TME (Wang et al. [2022\)](#page-16-5).

As tumor invasion progresses and anti-infammatory cytokines increase, the latter enhances the suppression of the TME, and GAMs gradually lose all of the above functions, eventually becoming "accomplices" of tumor cells. This process reflects a continuous change in transcription between M1 and M2 macrophages rather than an absolute change (Lanza et al.  $2021$ ). Therefore, the general conclusion is that GAMs can coexist with tumors (Cassetta and Pollard [2023\)](#page-13-12). It is currently believed that the immune function of the TME is infuenced by PTEN status. The expression level of PTEN is positively correlated with M1 polarization, while reducing the expression of CCL2 and VEGFA inhibits M2 polarization. PTEN cooperates with NHERF-1 to afect macrophage polarization by regulating its expression and membrane localization (Li et al. [2015;](#page-14-14) Zhou et al. [2022a,](#page-16-9) [b\)](#page-16-10). Sahin et al. reported the role of PTEN in inhibiting macrophage secretion of arginase I (Arg-I), which is considered to be a key enzyme for M2 activation. PTEN regulates Arg-I expression and secretion by inhibiting the Akt signaling pathway, which activates Arg I gene transcription through the transcription factor CCAAT/enhancer binding protein β (C/EBPβ) (Sahin et al. [2014\)](#page-15-15).

# **5 The promoting efect of GAMs on tumor growth**

GAMs mainly promote tumor growth in the following aspects, as shown in Fig. [1.](#page-5-0)

#### **5.1 Promoting angiogenesis**

GAMs can secrete various cytokines, including VEGF, FGF1/2, PDGF, and TGF-β, which promote angiogenesis. These cytokines stimulate endothelial cell proliferation, migration, and lumen formation, ultimately providing nutrients and oxygen to tumors. This phenomenon is referred to as "infammation-driven angiogenesis. "(Cui et al. [2018](#page-13-13)).

Research by Cui et al. demonstrated that GBM angiogenesis depends on the interaction between TGF-β1 secreted by tumor cells and integrin (αvβ3) on the surface of endothelial cells. Dual blockade of both can efectively inhibit GAM-related angiogenesis and extracellular matrix changes (Cui et al. [2018](#page-13-13)). Wei et al. found that TNF-α expression leads to vascular endothelial activation in a GBM mouse model, reducing the efficacy of antiangiogenic therapy. Therefore, combining a TNF- $\alpha$ antibody with targeting VEGF can efectively improve treatment efficacy (Wei et al. [2021\)](#page-16-11).

#### **5.2 Promoting tumor invasion and metastasis**

GAMs also secrete various proteases, such as MMP2, MMP9, and uPA, that degrade the extracellular matrix and basement membrane. This destroys the normal communication between cells and provides conditions for tumor cell infltration and migration (Lanza et al. [2021](#page-14-13); Quintero-Fabián et al. [2019](#page-15-16)). In the glioma TME, GM-CSF is upregulated, promoting GAM secretion of large amounts of CCL5, which upregulates MMP2 expression and activity through the calcium-dependent protein kinase CaMKII pathway, achieving directional migration of glioma (Yu-Ju Wu et al. [2020\)](#page-16-12). M2-like GAMs can directly induce tumor cells into the mesenchymal type  $(CD44^+$ , YKL-40<sup>+</sup>, MET<sup>+</sup>, NOTCH<sup>+</sup>) by mediating the binding of oncostatin M (OSM) to its receptor OSMR or the IF-6 receptor LIFR. This endows tumors with stronger metabolic activity and drug resistance (Hara et al. [2021](#page-14-15)). After surgical removal of the tumor, GAMsecreted trophic factor (PTN) can even "revive" GSCs (Knudsen et al. [2022\)](#page-14-16). Hypoxia-induced M2 macrophages activate PI3K/Akt/Nrf2 by releasing VEGF, which promotes GBM cell proliferation, epithelial-mesenchymal transition (EMT), GSC characteristics and resistance to TMZ, thereby signifcantly enhancing tumor invasion ability (Zhang et al. [2022](#page-16-13)).

# **5.3 Promoting the formation of a tumor‑favorable TME**

GAMs secrete various immunosuppressive factors, such as IL-10, TGF-β, PGE2, and IDO, which inhibit the activation and function of T cells, NK cells, and DCs. These factors induce apoptosis of  $CD4^+$  and CD8<sup>+</sup> T cells and increase the number and activity of Treg cells, enabling tumor cells to escape immune surveillance (Tan et al. [2021a,](#page-15-17) [b](#page-15-18)). M2-like GAMs can also induce T-cell exhaustion by expressing PD-1 ligands (PD-L1 and PD-L2), CTLA-4 ligands (CD80 and CD86) and VISTA (a potent negative regulator of



<span id="page-5-0"></span>**Fig. 1** Promoting efects of homeostatic imbalance of GAMs on gliomas: (1) Promotes GBM cell metabolism, proliferation, and stemness, (2) Promotes extracellular matrix (ECM) degradation (3) Promotes GBM cell migration and invasion (4) Promotes angiogenesis around GBM (5) Promotes an immunosuppressive TME

T-cell function) (Lin et al. [2021\)](#page-15-19). GAMs recruit more immunosuppressive GAMs by releasing IL-11, which activates the STAT3-MYC signaling pathway in GSCs, a process that also depends on the activation of PI3K-γ (Li et al. [2021\)](#page-14-17).

## **5.4 Promoting tumor cell growth and metabolism**

GAMs can interact with tumor cells through direct contact or indirect secretion of factors, afecting tumor cell proliferation, diferentiation, metabolism, apoptosis and stem cell characteristics. TGFβ-1 is a protein secreted by GAMs that promotes the growth and stemness of GSCs via the integrin avβ5-Src-Stat3 signaling pathway (Xiao et al. [2022\)](#page-16-14). Macrophage-released osteopontin (OPN)/SPP1 activation of CD44 is critical for tumor stemness, and SPP1 interaction with CD44 can activate downstream pathways of MES-like transformation in glioma cells, increasing their invasive-ness and drug resistance (He et al. [2021\)](#page-14-18). This process can also regulate the TME to increase the number of M2-like GAMs while inhibiting  $CD4^+$  and  $CD8^+$  T-cell activity (Pietras et al. [2014](#page-15-20)) (Zhao et al. [2020\)](#page-16-15). M2-like macrophages promote glycolysis in tumor cells by releasing IL-6 and IL-1β, phosphorylating PGK1 and GPD2, etc., increasing tumor cell lactate production and ATP levels and enhancing tumor energy metabolism (Zhang et al. [2018;](#page-16-16) Lu et al. [2020](#page-15-21)). Overexpression of Chitinase-3 like-protein-1 (CHI3L1) in TME often indicates poor prognosis, which is widely expressed by macrophages and others, binds to its receptors IL-13Ra2, TMEM219, Gal-3, CRTH2 and CD44, activates NF-κB and other pathways, and promotes glioma proliferation (Zhao et al. [2020](#page-16-15)).

# **6 Tumor "hijacking" of GAMs**

Tumor "hijacking" of GAMs refers to the tumor altering the phenotype and function of GAMs through various mechanisms, making them a phenotype that is conducive to tumor cell survival. These mechanisms are shown in Fig. [2](#page-6-0).

# **6.1 Regulating macrophage polarization signals**

Tumor cells can secrete various polarization factors, such as CSF-1, IL-4, IL-10, IL-13, and IL-34, which can induce macrophage polarization to M2 (Tong et al.



<span id="page-6-0"></span>**Fig. 2** Glioma GAM imbalance mechanism: (1) Immunosuppressive molecules, such as TGF-β and IL-10, promote GAM transformation to the M2 type, reduce the immune activity of macrophages, and inhibit the killing efect of immune cells on tumor cells (2) Cytokines, such as CSF-1 and VEGF, recruit and activate GAMs to tumor tissues and promote the transformation of GAMs to the M2 type, promoting the growth of blood vessels. (3) miRNA: In glioma, the expression of certain miRNAs is water fat and related to the polarity of GAMs. For example, miR-142-3p can promote the conversion of GAMs to the M1 type, while miR-511 can promote the conversion of GAMs to the M2 type

[2021](#page-15-22)). Diferent polarization mechanisms are shown in Fig. [3](#page-7-0). Glioma cells can release various miRNAs through exosomes, including miR-1246, miR-155-3p, miR-10b-5p, miR-32 and miR-3591-3p. These miRNAs can be taken up by GAMs and afect the expression of genes such as IRF1, PTEN, CBLB, and MAPK1, inhibiting the corresponding signaling pathways IRF1/IFN-β, PTEN/ PI3K/AKT, JAK2/STAT3, and MAPK, thereby promoting GAMs to polarize to the M2 type (Qian et al. [2020;](#page-15-23) Xu et al. [2021;](#page-16-17) Li et al. [2022a,](#page-14-8) [b,](#page-14-9) [c](#page-14-10), [d](#page-14-11), [e,](#page-15-9) [f](#page-15-10), [g;](#page-15-11) Bao and Li [2019](#page-13-14); Shi et al. [2020](#page-15-24); Pan et al. [2022\)](#page-15-25).

High-grade gliomas often overexpress EWSR1, mediating the production of circular RNA circNEIL3 and afecting IGF2BP3 stability, thereby afecting the polarization of M2-like GAMs and releasing IL-10, GF-β, and other immunosuppressive factors (Li et al. [2022a,](#page-14-8) [b,](#page-14-9) [c,](#page-14-10) [d](#page-14-11), [e](#page-15-9), [f,](#page-15-10) [g](#page-15-11)). In GBM, the β2-microglobulin (B2M) subunit of MHC-II is activated by TGFβ-1 and promotes M2 transformation through SMAD and PI3K/AKT (Li et al. [2022a,](#page-14-8) [b,](#page-14-9) [c](#page-14-10), [d](#page-14-11), [e](#page-15-9), [f,](#page-15-10) [g;](#page-15-11) Ma et al. [2021\)](#page-15-26). GSCs have the ability to regulate CD90 expression and increase macrophage immune suppression through the USF1/CD90 axis while enhancing their own stemness (Zhou et al. [2022a,](#page-16-9) [b](#page-16-10)). ARS2/MGAL signaling in GSCs can also promote GAM self-renewal and M2 polarization (Yin et al. [2020](#page-16-18)). TERM2 expression is increased in glioma and is closely related to the enrichment of GAMs, especially M2-like macrophages. It can also inhibit the macrophage response to TLR ligands (Yu et al. [2022](#page-16-19)).

# **6.2 Enhanced immunosuppression of the TME**

Glioma cells release the secretory peptides SLIT2 and OPN, which bind to ROBO receptors and integrin ITGαvβ5 on the surface of macrophages, respectively, activating the PI3K-γ signaling pathway, mediating macrophage chemotaxis to the TME, and expressing cytokines to promote tumor growth and angiogenesis (Geraldo et al. [2021](#page-14-19); Wei et al. [2019](#page-16-20)). ATRX-deficient IDH-mutant gliomas exhibit more pronounced astrocytic characteristics while reducing the binding of CCCTCbinding factor (CTCF) to DNA. This disrupts the expression of chemokines such as GFAP, CXCL12, and CXCL14, mediating more GAM infltration and remodeling the immune microenvironment (Babikir et al. [2021](#page-13-15)). METoverexpressing tumors recruit more immunosuppressive GAMs by activating MET-STAT-PD-L1 (Wang et al. [2021](#page-16-21)). The nuclear function of tumor-secreted IL-33 can induce macrophages to produce a series of chemokines that recruit and activate circulating monocytes, such as CCL-2, CCL-5 and CXCL10, creating a glioma 'habitable environment' (Boeck et al. [2020](#page-13-16)). Tumor cells can consume large amounts of oxygen and glucose, resulting in a state of hypoxia and hypoglycemia in the tumor microenvironment. This state can activate HIF-1 $\alpha$  and



<span id="page-7-0"></span>**Fig. 3** The polarization of GAMs: As a duplicitous in the immune environment, GAM cells play a two-way role: tumor killing efect vs. pro-tumor efect (M1: infammation, phagocytosis, attack; M2: immunosuppression, fbrosis formation, tissue repair, angiogenesis). GAMs are continuous: the state of continuous transition between M1 and M2 morphologies, and the proportion of each morphology depends on the type and concentration of diferent secretions in the receiving tumor environment

AMPK signaling pathways in macrophages, promoting M2 polarization (Xiao et al. [2022](#page-16-14)). RNA regulatory factor (HuR) is a key factor in regulating tumor immune suppression. Knockout of HuR animal models showed tumor shrinkage, relatively increased M1-like GAMs, reduced PD-L1 expression, increased infiltration of  $CD4+T$  cells (including Th1 and CTL), and reduced tumor-associated polymorphonuclear myeloid-derived suppressor cells (Wang et al. [2019\)](#page-15-27). Matrix remodeling-associated protein 8 (MXRA8) is a novel prognostic indicator that may be involved in ferroptosis. MXRA8 expression in gliomas is signifcantly higher than that in normal brain tissue, and high expression of MXRA8 is associated with poor prognosis (Xu et al. [2022a](#page-16-22), [b\)](#page-16-23). MXRA8 is related to various immune suppressive factors, such as TGF-β1, IL-10, PD-L1 and CTLA4. Studies have found that knocking down this gene can reduce M2 infltration and rescue glioma progression (Xu et al. [2022a,](#page-16-22) [b](#page-16-23)).

# **6.3 Regulating the immune microenvironment by epigenetic modifcation**

Epigenetic modifcation is the process of regulating gene expression without changing the DNA sequence. Common epigenetic dysregulation in tumors includes mutations and abnormal expression of epigenetic modifcation enzymes and changes in the levels of related cofactors, which alter the structure and dynamics of chromatin, leading to changes in gene expression and ultimately promoting tumor initiation and progression

(Gangoso et al. [2021\)](#page-14-20). Epigenetic modifcation of gliomas greatly increases the opportunity to regulate the immune microenvironment. Mesenchymal-like GBM cells promote immune escape under host defense by epigenetic modifcations, including loss of CpG island methylation associated with IRF1, CCL2, and IRF8 (Gangoso et al.  $2021$ ). This immune escape mainly involves increased bone marrow-derived GAMs and T-cell exhaustion (Goswami et al. [2020;](#page-14-21) Mohme and Neidert [2020\)](#page-15-28). Under hypoxic conditions, glioma cells express ALKBH5. This enzyme can remove N6-methyladenosine  $(m<sup>6</sup>A)$  modification on the long noncoding RNA (lnc-RNA) NEAT1, which can increase CXCL8/ IL8 expression by inhibiting SFPQ and increase chemotaxis to GAMs (Dong et al. [2021](#page-13-17); Wei et al. [2022](#page-16-24)).

# **6.4 Regulating the interaction of macrophages with other immune components**

There are various immunosuppressive factors in the tumor microenvironment. M2 macrophages inhibit the activation of CD8+T cells and cDC1s by expressing molecules such as PD-L1, CD47 and ARG1 and secreting factors such as IL-10 and TGF-β while reducing the efficacy of targeting PD-1 (Zhu et al.  $2020$ ; Lee et al. [2021](#page-14-22)). Hu et al. found that gliomas escape the phagocytosis of GAMs by expressing high levels of LRIG2, which increases the expression of CD47 on M2 macrophages (Hu et al. [2022](#page-14-23)).



<span id="page-8-0"></span>**Fig. 4** Several targeted GAMs therapies. **A**: The CSF1/CSF1R signaling axis is overexpressed in high-grade gliomas and is associated with poor prognosis. Monoclonal antibodies or small molecule inhibitors targeting CSF-1/CSF1R can block CSF1R signaling. **B**: Immunovirotherapy: Oncolytic viruses may dissolve GBM, releasing tumor antigens and danger signals, or induce polarization of GAMs into M1 by upregulating PD-L1, TLR, or NF-kB, either due to therapeutic factors carried by OV such as GM-CSF, IFN-γ, or IL-12. **C**, **D**: Two classic immune checkpoint therapies: PD-1 antibodies, on one hand, restore the function of T cells, inhibit tumor growth and metastasis. On the other hand, even in the case of CD8+T cell exhaustion, PD-1 antibodies can still induce GAMs to polarize towards the M1 phenotype, producing anti-tumor efects. CD47 sends a 'do not eat me' signal to macrophages, preventing them from engulfng tumors. The combination of CD47 and CD24 antibodies efectively activates the bone marrow immune response, kills cancer cells, and simultaneously interrupts the malignant cycle caused by GSCs revival

Canine uric acid produced by GBM cells activates AHR in GAMs, promoting CCR2 expression. At the same time, AHR promotes the expression of ectonucleotidase CD39 in GAMs, which cooperates with CD73 to produce adenosine, leading to CD8<sup>+</sup> T-cell dysfunction (Takenaka et al. [2019](#page-15-29)). At the same time, AHR promotes the expression of ectonucleotidase CD39 in GAMs, which cooperates with CD73 to produce adenosine, leading to CD8<sup>+</sup>T-cell dysfunction (Takenaka et al. [2019\)](#page-15-29). Recurrent gliomas encode temozolomide-associated lnc-RNA (lnc-TALC), which activates the p38-MAPK signaling pathway to promote GAM release of complement C5/C5a after binding to ENO1, thereby promoting tumor cell DNA repair. Therefore, combined immunotherapy targeting C5a can

overcome lnc-TALC-mediated TMZ resistance (Boeck et al. [2020](#page-13-16)).

# **7 Targeted GAM therapy**

Targeted GAM therapy refers to the use of drugs or biological agents to intervene in the number, phenotype and function of GAMs, thereby inhibiting tumor growth and metastasis. The mechanism of targeted GAM therapy is visualized in Fig. [4.](#page-8-0)

# **7.1 Reduce the number of GAMs**

By inhibiting the survival or recruitment of macrophages, the number of GAMs in the TME can be reduced, thereby alleviating the protumor efect of the TME.

#### *7.1.1 Targeting CSF‑1 therapy*

Macrophages depend on CSF-1 for diferentiation and survival, and its paracrine signaling pathway plays a key role in the symbiotic relationship between glioma cells and GAMs (Pyonteck et al.  $2013$ ). Therefore, inhibiting the CSF-1/ CSF-1R signaling pathway is a representative strategy for targeting GAM depletion, which blocks the migration and diferentiation of peripheral blood monocytes into tumor tissues and significantly enhances the efficacy of other therapeutic measures (Pombo Antunes et al. [2021](#page-15-8); Akkari et al. [2020](#page-13-18)). This therapy has also achieved some results in medullo[b](#page-15-18)lastoma models (Tan et al.  $2021a$ , b). The efficacy of CSF-1 inhibitors depends on whether glioma cells are driven by *PDGFR* or *RAS* genes (Rao et al. [2022](#page-15-31)). The efficacy of CSF-1 inhibitors depends on whether glioma cells are driven by PDGFR or RAS genes. PDGFR-driven gliomas induce the proliferation and activation of microglia, which depend on the CSF-1 signaling pathway for differentiation and survival. RAS-driven gliomas have GAMs rich in proinfammatory and proangiogenic signals, which can maintain survival through other pathways (such as GM-CSF and IFN-γ), even when CSF-1R is inhibited (Rao et al. [2022\)](#page-15-31). Therefore, combined antiangiogenic therapy can significantly improve the efficacy of CSF-1 inhibitors against RAS tumors (Rao et al. [2022](#page-15-31)). However, Antunes et al. used a mouse model of glioma and showed that oral administration of the CSF1R inhibitor PLX3397 did not attenuate monocyte infltration into GBM but rather increased the percentage of monocytes in tumors. They believe that the mechanism of targeting CSFR is to block monocyte diferentiation into GAMs (Pombo Antunes et al. [2021\)](#page-15-8).

Several studies have reported methods to observe the efficacy of targeting CSF-1 by imaging methods. Foray et al. developed a method to noninvasively observe the efficacy of a CSF-1R inhibitor using  $18$  F-DPA-714 by PET/MRI, which showed that macrophage infltration was signifcantly reduced and tumor volume was signifcantly reduced after treatment (Foray et al. [2022\)](#page-14-24). Li et al.'s T2-weighted MRI imaging methods can also detect this process (Li et al. [2022a,](#page-14-8) [b](#page-14-9), [c,](#page-14-10) [d](#page-14-11), [e](#page-15-9), [f,](#page-15-10) [g](#page-15-11)).

# *7.1.2 Other therapies to reduce GAMs*

CCR2 is a key molecule for the chemotaxis and activation of macrophages (Boissonnas and Combadière [2022\)](#page-13-19). Silva et al. reported an inhibitor that targets and inhibits glutamyl cyclotransferase-like protein (QPCTL), which can inhibit the N-terminal degradation of CCR2 and CCL7, preventing macrophages from being released from the bone marrow and migrating to tumor tissues and limiting the recruitment of GAMs (Barreira da Silva et al. [2022](#page-13-20)). The targeting CCR2 inhibitor CCX872 also showed similar efects in glioma and other tumor models (Tu et al. [2020](#page-15-32); Flores-Toro et al. [2020\)](#page-14-25). PI3K is a key enzyme in tumor signal transduction, and its subtype PI3Kγ plays an important role in GAM polarization and migration. Inhibition or knockout of PI3Kγ can reduce the number of GAMs in the TME and enhance the chemotherapeutic efect of TMZ (Li et al. [2021\)](#page-14-17).

# **7.2 Phenotype of transformed GAMs**

In theory, activating M1 polarization signals or inhibiting M2 polarization signals can achieve GAM repolarization, and there are currently multiple ways to deliver regulatory factors to the TME to exert efects.

## *7.2.1 Promotion of M1 polarization*

On the one hand, activating TLR, IFN-γ, IL-12, STAT1 and other signaling pathways can promote GAM polarization to the M1 type (Birocchi et al. [2022](#page-13-21)). Xue et al. found that chlorogenic acid (CHA) afects the STAT1 and STAT6 signaling pathways, promoting macrophages to transform into the M1 phenotype and inhibiting tumor growth in animal models (Xue et al. [2017](#page-16-26)). Ginsenoside Rg3 is a plant sterol with antitumor activity. Zhu et al. developed liposomes coloaded with Rg3 and paclitaxel (Rg3-PTX-LPs), which can increase the ratio of M1/M2 macrophages, reduce the expression of the M2 markers CD206 and Arg-1, and increase the expression of the M1 markers iNOS and TNF-α, possibly through the activation of STAT1 (Zhu et al. [2021](#page-16-27)). Li et al. found that high mobility group protein B1 (HMGB1) is a key regulator between glioma cells and M1 polarization. HMGB1 induces macrophages to polarize into M1 through the RAGE-NFκB-NLRP3 infammasome pathway and enhances tumor cell sensitivity to TMZ (Li et al. [2022a](#page-14-8), [b,](#page-14-9) [c,](#page-14-10) [d,](#page-14-11) [e,](#page-15-9) [f,](#page-15-10) [g](#page-15-11)). Magnolol and disulfram/copper (DSF/ Cu) coloaded peptide-modifed liposomes can target the TME, induce apoptosis through ROS and autophagy inhibition, efectively inhibit angiogenesis, repolarize GAMs into the M1 phenotype, and enhance T-cell and NK cell antitumor immune responses (Zheng et al. [2020](#page-16-28); Li et al. [2022a](#page-14-8), [b,](#page-14-9) [c,](#page-14-10) [d,](#page-14-11) [e](#page-15-9), [f,](#page-15-10) [g](#page-15-11)). Birocchi made engineered hematopoietic stem cells (HSCs), which can improve GAM immune suppression by releasing IFN-α and IL-12 in the TME, improving the survival rate of glioma model mice, and this study is currently undergoing a phase I clinical trial in Italy (Birocchi et al. [2022\)](#page-13-21). In vitro transcription mRNA (IVT mRNA) is a novel potential targeted drug. Zhang et al. developed an IVT mRNA nanocarrier for GAM repolarization. After entering the TME, the carrier polymer is hydrolyzed, releasing mRNA that is transcribed by ribosomes into IRF5 and IKKβ proteins, which then activate M1 polarization-related signaling pathways, expressing IL-12, IFN-γ and TNF-α and other

infammatory cytokines, while inhibiting M2 polarization-related immune suppression and protumor factors such as IL-6, Serpinb2 and CCL11 (Zhang et al. [2019](#page-16-29)). These approaches show promise in achieving GAM repolarization and improving the efficacy of immunotherapy in gliomas and other cancers. However, further research is needed to optimize delivery methods and dosing strategies to maximize therapeutic benefts while minimizing potential side efects.

#### *7.2.2 Inhibition of M2 polarization*

On the other hand, inhibiting signaling pathways such as IL-4, IL-10, IL-13, STAT3 and STAT6 can suppress GAM polarization to M2 (Yang et al. [2022a,](#page-16-30) [b\)](#page-16-31). Carnosic acid (CA) is a natural compound derived from rosemary that can inhibit the Wnt/β-catenin-WISP1 signaling pathway. WISP1 is a protein secreted by GSCs that can promote GSC survival and proliferation and afect immune cells in the tumor microenvironment (Tao et al. [2020\)](#page-15-33). CA reduces WISP1 expression, decreases the M2 phenotype, and prolongs survival time in mouse models (Tao et al. [2020](#page-15-33)). Yang et al. recently developed the nanodrug SPP-ARV-825, which works by inhibiting IFR4 promoter transcription, afecting STAT3/6 and AKT phosphorylation, and thereby suppressing M2 polarization (Yang et al. [2022a](#page-16-30), [b](#page-16-31)). MK-8931 was originally a BACE1 inhibitor for treating Alzheimer's disease and has entered phase III clinical trials. It was recently shown to alter the GAM phenotype by inhibiting the IL6/STAT3 pathway and synergistically enhance radiotherapy efficacy in animal models (Zhai et al. [2021](#page-16-32)). Palbociclib is a CDK6 inhibitor that can reduce M2 polarization markers. This substance can also inhibit glioma growth by various methods, such as reducing lncRNA SNHG15 expression and increasing miR-627-5P (Li et al. [2019](#page-14-26)).

## *7.2.3 miRNA therapy*

miRNA is a small noncoding RNA that regulates mRNA degradation or translation by binding to mRNA with incomplete base pairing (Guo et al. [2019\)](#page-14-27). miRNA therapy provides a highly selective method to achieve precise modulation of the immune microenvironment. miR-124 is the most abundant miRNA in the CNS and is involved in neural development and functional regulation (Xu et al. [2022a](#page-16-22), [b\)](#page-16-23). Several studies have found that miR-124 vectors have therapeutic potential by inhibiting M2 polarization through targeting the STAT3 and C/ EBP $\alpha$  pathways and alleviating radiation-induced cognitive dysfunction (RICD) to some extent after radiotherapy (Yin et al. [2020;](#page-16-18) Leavitt et al. [2020](#page-14-28); Romano et al. [2020](#page-15-34)). miR-22 is a signifcantly downregulated miRNA in glioma cells and GAMs that can inhibit the NF-κB signaling pathway by targeting HDAC6, thereby enhancing macrophage phagocytosis, antigen presentation and T-cell activation abilities and inhibiting glioma growth and progression (Tu et al. [2022\)](#page-15-35). Currently, some studies have explored the possibility of miR-22 involvement in therapy, including using nanoparticles or macrophages as carriers to deliver miR-22 to tumor sites or combining with chemotherapy or radiotherapy to enhance therapeutic efects (Müller Bark et al. [2020;](#page-15-36) Kirstein et al. [2020\)](#page-14-29).

#### **7.3 Improving the immune regulatory function of GAMs**

The function of GAMs depends on various expressed receptors and secretions, and regulating them can regulate their immune function.

#### *7.3.1 Antiangiogenic therapy*

Inhibiting angiogenic factors secreted by GAMs, such as VEGF, bFGF, and PDGF, or blocking their binding to corresponding receptors can cut off the nutrient supply of tumors (Gilbert et al. [2017](#page-14-30)). Bevacizumab is a monoclonal antibody against VEGF-A that can inhibit angiogenesis and tumor growth (Ahir et al. [2020\)](#page-13-22) and signifcantly improved PFS-6 in phase III clinical trials (Gilbert et al.  $2014$ ). The VEGF receptor tyrosine kinase inhibitor cediranib also showed significant efficacy in a GBM phase II clinical trial (Batchelor et al. [2010\)](#page-13-23). Inhibiting proteases such as MMPs can reduce tumor invasion and metastasis (Peng et al. [2022\)](#page-15-37), and potential MMP inhibitors with therapeutic value include DX-2400, AB0041, AB0046 and GS-5745, which are currently in clinical trials (Fields [2019](#page-14-32); Levin et al. [2017](#page-14-33)). Glioma cells can stimulate GAMs to produce TNF-α by secreting IL8 and CCL2, which then mediates endothelial activation (Wei et al. [2021](#page-16-11)). This is the main cause of antiangiogenic therapy failure, and inhibiting  $TNF-α$  effectively blocks endothelial activation and increases the survival rate in model mice (Wei et al. [2021](#page-16-11)).

#### *7.3.2 Immune checkpoint regulation*

Immune checkpoints are a class of molecules that can regulate the intensity and duration of immune responses, and tumor cells often use them to evade immune surveillance and clearance. CD47 and CD24 are two immune checkpoint molecules that send "don't eat me" signals to macrophages, preventing macrophage phagocytosis of tumors (Barkal et al. [2019\)](#page-13-24). The combined use of CD47 and CD24 dual antibodies efectively activates bone marrow immunity, signifcantly kills cancer cells, and interrupts the vicious cycle caused by GSC revival (Wu et al. [2021\)](#page-16-33). Etomoxir combined with a CD47 monoclonal antibody inhibits CD47 expression and function and restores the macrophage phagocytosis ability of tumors (Jiang et al. [2022](#page-14-34)).

B7-H3 is also an immune checkpoint molecule that can inhibit macrophage phagocytosis of tumor cells by interacting with Siglec-10 on the surface of macrophages and is associated with poor prognosis of tumors (Huang et al. [2020](#page-14-35)). B7-H3 promotes glioma progression through the JAK2/STAT3/Slug signaling pathway, and the STAT3 inhibitor napabucasin can signifcantly inhibit B7-H3-mediated in vivo and in vitro tumor growth (Tang et al. [2019\)](#page-15-38). CXCL14 is a potential immunotherapy adjuvant factor that can increase MHCI expression on the surface of gliomas, chemoattract CTL cells in vivo and in vitro, and inhibit glioma growth in model mice but is downregulated in IDH-mutated gliomas (Kumar et al. [2022](#page-14-36)). High B7-H3 expression also inhibits CXCL14 signaling; therefore, targeting both B7-H3 and CXCL14 can achieve optimal therapeutic efects (Kumar et al. [2022](#page-14-36)).

PD-1 antibody is the most representative immune checkpoint inhibitor, and blocking the PD-1 and PD-L1 interaction can restore T-cell function and inhibit tumor growth and metastasis (Cai et al.  $2019$ ). However, the efficacy of PD-1 antibodies does not depend solely on T cells, and recent studies have found that innate immunity also plays an important role in this process (Cai et al. [2019](#page-13-25); Fierro et al.  $2022$ ). In the case of  $CD8<sup>+</sup>$  T-cell depletion, an anti-PD-1 antibody can still induce GAM polarization to the M1 phenotype by producing antitumor efects (Liu et al. [2023\)](#page-15-39). M1-like GAMs express high levels of PD-L1, providing more targets for PD-1 antibodies (Ruan et al. [2021](#page-15-40)). VEGF not only promotes angiogenesis but also induces tumor cells and GAMs to overexpress PD-L1 by activating the VEGFR1 signaling pathway (Liu et al. [2023](#page-15-39)). However, this process can be inhibited by soluble VEGFR1 (sVEGFR1). When the PD-L1 antibody blocks PD-L1, it can promote GAMs to secrete more sVEGFR1, suggesting the potential of immune checkpoint inhibitors combined with antiangiogenic therapy (Liu et al. [2023](#page-15-39); Ruan et al. [2021](#page-15-40)). Dendritic cell vaccines can induce tumor-specifc T-cell immunity but lead to upregulation of PD-L1 on macrophages (Antonios et al. [2017\)](#page-13-26). When anti-PD-1 monoclonal antibodies and CSF-1R inhibitors are used in combination with dendritic cell vaccines, they can signifcantly prolong mouse survival, increase T-cell number and activity, and produce long-term memory immunity (Antonios et al. [2017](#page-13-26)). NAMPT inhibitors can reduce NAD levels in GBM cells and change the immune status in the tumor microenvironment, leading to upregulation of PD-L1, reduction in M2-type macrophages, and inhibition of tumor cell growth (Li et al. [2020\)](#page-14-38). IL-6 is an immunosuppressive cytokine that induces PD-L1 expression through the STAT3 pathway (Lamano et al. [2019](#page-14-39)). In a mouse GBM model, IL-6 transduction inhibition reduced PD-L1 expression on bone marrow-derived macrophages, reduced tumor growth, and prolonged mouse survival. This effect is  $CD8<sup>+</sup>$  T-cell-dependent and has a synergistic effect with anti-PD-1 monoclonal antibody treatment (Lamano et al. [2019](#page-14-39)).

# *7.3.3 Oncolytic viruses*

Oncolytic viruses (OVs) can directly lyse tumor cells or induce antitumor immune responses (immunotherapy). OV can afect the phenotype and function of GAMs in various ways. On the one hand, OV can induce GAM polarization to the M1 type, thereby producing antitumor immune responses (Sanchez Gil and Rabkin [2022](#page-15-41)). This effect may be due to GAM death caused by OV infection, releasing tumor antigens and danger signals, or due to OVs carrying 'weapons', such as GM-CSF, IFN-γ or IL-12 (Sanchez Gil and Rabkin [2022\)](#page-15-41). On the other hand, OV can also produce antiviral responses by upregulating PD-L1, TLR or NF-kB, limiting virus replication and spread (Blitz et al. [2022](#page-13-27); Christie and Chiocca [2022](#page-13-28)). Kiyokawa et al. developed an oncolytic adenovirus (ICO-VIR17) expressing hyaluronidase (PH20), which promotes M1 polarization through NF-kB while degrading hyaluronic acid (HA) in the TME, showing broader efficacy (Kiyokawa et al. [2021](#page-14-40)). Researchers combined the use of oncolytic virus expressing IL-12 (G47Δ-mIL12), PD-1 antibody and CTLA-4 antibody. On the one hand, oncolytic viruses disrupt tumor cells to release antigens and promote antigen presentation. On the other hand, M1 cells activated by IL-12 further enhance CD8<sup>+</sup> T-cell killing activity while reducing Treg number and function. The treated animal models showed signs of tumor eradication (Saha et al. [2017\)](#page-15-42).

#### *7.3.4 Targeting CD40 therapy*

CD40 is a costimulatory molecule expressed on GAMs that can bind to the ligand CD40L, activate GAM immune function, and induce tumor cell apoptosis and the immune response. Currently, two clinical trials are evaluating the efficacy of humanized IgG1CD40 monoclonal antibody treatment for GBMs (NCT02304393, NCT04164901). Studies have found that IL-6 can induce GAMs to express CD40 through two signaling pathways, STAT3 and HIF-1α, thereby counteracting the immunosuppressive effect of IL-6 on GAMs. Therefore, targeting both IL-6 and CD40 is a potential therapeutic strategy (Yang et al. [2021\)](#page-16-34).

## *7.3.5 Other GAM‑related immune regulatory therapies*

Ferroptosis is the main mode of cell death for glioma cells, but it also enhances M2 activity, leading to immune suppression and tumor growth (Cai et al. [2021a](#page-13-3), [b](#page-13-4)). Therefore, Liu et al. demonstrated that patients with weak ferroptosis activity had better efficacy with PD-L1 antibody in animal models and the IMvigor210 clinical immunotherapy cohort (Liu et al. [2022](#page-15-43)). Recently, it was found that NH1 inhibitors restored GAM mitochondrial oxidative phosphorylation (OXPHOS) function, increased GAM glucose uptake and mitochondrial activity, and reduced aerobic glycolysis, thereby promoting cellular immunity and enhancing TMZ and PD-1 anti-body efficacy (Hasan et al. [2021](#page-14-41)). Yang et al. prepared zinc cyclic di-AMP nanoparticles (ZnCDA), which targeted STING binding of macrophages and activated the TBK1/IRF3 and IKK/NF-κB signaling pathways, thereby inducing the production of IFN-I and other infammatory factors, stimulating macrophages to express MHC-II molecules and costimulatory molecules, enhancing antigen presentation ability, and promoting T-cell activation and proliferation (Yang et al. [2022a,](#page-16-30) [b](#page-16-31)).

# **8 Discussion and conclusions**

This review discusses the immunobiological characteristics of GAMs, including their origin, classifcation, function and regulation mechanisms, and discusses potential therapeutic strategies. GBM is a malignant brain glioma with high invasiveness, recurrence and drug resistance and is one of the most common and fatal tumors in the CNS (Wang et al.  $2017$ ). The impact of GBM on TME homeostasis is multifaceted and intricate. By promoting angiogenesis, immune evasion, invasion, metastasis, and drug resistance, GBM exerts an infuence on various components of the TME, thus creating a conducive environment for tumor growth, progression, and responses to treatment. GAMs are the major cell population in the TME and have an extensive symbiotic relationship with the tumor. They can promote the stemness of GSCs, enhance cell proliferation, survival, and migration, and suppress T-cell-mediated immune responses, thereby promoting tumor progression (Xuan et al. [2021\)](#page-16-3).

The classification of GAMs remains a subject of intense debate within the academic community. Currently, there is widespread recognition that GAMs exhibit a heterogeneous and plastic range of phenotypes (Pombo Antunes et al. [2021](#page-15-8); Yin et al. [2022\)](#page-16-7). However, consensus on their identifcation and classifcation has not yet been achieved. Although researchers have discovered various biomarkers to delineate subsets and functions of GAMs, these classifcation methods are often overly complex and have not provided efective guidance for foundational experiments. Consequently, current experimental approaches often rely on a binary categorization of GAMs into two subtypes: tumor-suppressive GAMs and tumor-promoting GAMs. In the context of GBM, GAMs primarily exhibit immunosuppressive characteristics. The tumor-promoting effects of GAMs in GBM are evident in several crucial aspects. First, they actively promote angiogenesis, thereby facilitating the formation of new blood vessels to sustain tumor growth (Cui et al. [2018](#page-13-13); Wei et al. [2021\)](#page-16-11). Second, GAMs play a role in facilitating tumor invasion and metastasis, enabling the spread of cancer cells to distant sites (Xiao et al. [2022](#page-16-14); He et al. [2021](#page-14-18); Pietras et al. [2014](#page-15-20); Zhang et al. [2018;](#page-16-16) Lu et al. [2020](#page-15-21); Zhao et al. [2020\)](#page-16-15). Additionally, GAMs create a TME that is conducive to tumor growth and progression (Tan et al. [2021a](#page-15-17), [b](#page-15-18); Lin et al. [2021;](#page-15-19) Li et al. [2021](#page-14-17)). Finally, GAMs enhance tumor cell proliferation and metabolism, further contributing to the aggressive nature of GBM (Xiao et al. [2022](#page-16-14); He et al. [2021;](#page-14-18) Pietras et al. [2014](#page-15-20); Zhang et al. [2018](#page-16-16); Lu et al. [2020](#page-15-21); Zhao et al. [2020\)](#page-16-15). To ensure their survival, tumors employ various mechanisms to modify the phenotype and function of GAMs, efectively transforming them into phenotypes that support tumor cell survival. Therapeutic strategies targeting GAMs encompass a range of approaches, such as inhibiting their infltration or polarization and reshaping their phenotype or function (Birocchi et al. [2022](#page-13-21); Xue et al. [2017;](#page-16-26) Zhu et al. [2021](#page-16-27); Li et al. [2022a,](#page-14-8) [b,](#page-14-9) [c](#page-14-10), [d](#page-14-11), [e,](#page-15-9) [f](#page-15-10), [g;](#page-15-11) Zheng et al. [2020](#page-16-28); Zhang et al. [2019](#page-16-29)). These strategies can be employed individually or in combination with other treatment modalities to improve the prognosis of GBM patients. By understanding the complex role of GAMs and their contribution to tumor progression, researchers can develop more efective therapeutic interventions that specifcally target these cells, ultimately leading to improved outcomes for GBM patients.

Currently, there are several limitations to the targeted treatment strategies for glioma by focusing on GAMs. First, GAMs exhibit extensive heterogeneity and plasticity, necessitating precise timing and a clear understanding of the individual TME status for efective targeted therapy. Second, gliomas themselves demonstrate signifcant heterogeneity, requiring further investigation into the interactions between diferent types of GAMs and each subtype of glioma, as well as exploring their dynamic changes over time. GAMs likely play distinct roles in glioblastoma subtypes, grades, and mutational states, necessitating personalized approaches to optimize targeted GAM therapy. Third, although mice and human gliomas have some conservation of immune components, it is still difficult to use animal models to study the relationship between the glioma microenvironment and macrophages. For example, there is an isolation barrier between transplanted glioma tissue and normal brain tissue in animal models, but human primary brain tumors exhibit infiltrative growth. Therefore, using primary mouse glioma models can more accurately simulate the immune microenvironment of human gliomas. Fourth, most of the therapeutic strategies targeting GAMs are still in preclinical or early clinical stages, and more

clinical trials are needed to evaluate their safety, efficacy and tolerability. We hope that more efective methods can be found in the future to regulate the immune microenvironment of gliomas to enhance the efficacy of immunotherapy.

#### **Authors' contributions**

Jiachen Wang drafted the initial manuscript and created the original fgures. Shenglan Li critically reviewed the intellectual content of the manuscript and made substantial revisions. Yanjie Lan and Xinrui Liu critically reviewed the intellectual content of the manuscript. Wenbin Li designed the study and was responsible for it. He also critically reviewed the intellectual content of the manuscript and obtained funding for the project. All authors read and approved the fnal manuscript.

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#### **Declarations**

#### **Ethics approval and consent to participate**

This manuscript does not report on or involve any animals, humans, human data, human tissue or plants, so no ethical approval and consent to publish were required.

#### **Consent for publication**

This manuscript contains original images, which have been obtained with the consent of the original author. The author has given permission to use the images for the purpose of this manuscript and has agreed to publish them. The author has also confrmed that the images do not infringe any third-party rights or privacy.

#### **Competing interests**

Author Wenbin Li is a member of the Editorial Board for Current Medicine. The paper was handled by the other Editor and has undergone rigorous peer review process. Author wenbin Li was not involved in the journal's review of, or decisions related to, this manuscript.

The other authors of this manuscript declare that they have no competing interests related to the content of this article.

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