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Itaconimides derivatives induce relaxation in mesenteric artery and negative inotropism by inhibition of CA²⁺ influx

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Abstract

Background The aim of this study was to evaluate the cardiovascular effects of *N*-phenyl-itaconimide (Imide-1), *N*-4-methyl-phenyl-itaconimide (Imide-2), *N*-4-methoxy-phenyl-itaconimide (Imide-3) and *N*-4-chloro-phenyl-itaconimide (Imide-4), and investigate the mechanisms of action involved in the observed responses.

Methods The relaxant effect was investigated in rat superior mesenteric arteries by using isometric tension measurements. Additionally, in isolated atria were evaluated the heart rate and force of cardiac contraction and in vivo experiments was evaluated blood pressure and heart rate.

Results Cumulative administration of itaconimides $(3 \times 10^{-8} \text{ to } 3 \times 10^{-4} \text{ M})$ in pre-contracted mesenteric artery rings with phenylephrine, 1 µM, induced endothelium-independent vasorelaxation. The itaconimides showed similar maximum efficacies. Additionally, Imide-3 induced vasorelaxation in rings exposed to a depolarizing-tyrode solution containing 60 mM KCl or 20 mM KCl similar to the control, suggesting the non-participation of K⁺ channels. Imide-3 attenuated Ca²⁺ influx in a concentration-dependent manner. As well, imide-3 reduced CaCl₂-induced contraction in nominally calcium-free medium, in the presence of cyclopiazonic acid (20 µM), phenylephrine (1 µM) and nifedipine (1 µM), indicating a reduction of Ca²⁺ influx by receptor-operated channels (ROC) and store-operated channels (SOC). The presence of SKF 96365 (10⁻⁵ M), SOC blocker, did not significantly alter the vasorelaxant effect induced by imide-3 lowered blood pressure and induced bradycardia. **Conclusions** These results suggest that itaconimides have concentration-dependent vascular effects and the vasorelaxation seems to be endothelium-independent. The vasodilatory effect induced by imide-3 may be due to a possible influence on the Ca_V and ROC. In addition, imide-3 is able to reduce force of cardiac contraction, blood pressure and promote bradycardia.

Keywords *N*-4-methoxy-phenyl-itaconimide \cdot Bradycardia \cdot Hypertension \cdot Ion channels as drug targets \cdot Itaconimide \cdot Vasorelaxation

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Introduction

Hypertension is an important public health problem and has been associated with the development of cardiovascular and renal diseases. Moreover, hypertension is a major preventable risk factor for early death and disability. In this way, controlling blood pressure is significantly related with reduction of cardiovascular morbidity and mortality [1]. Hypertension causes a substantial impact to cardiovascular disease worldwide. In 2011 to 2014, 103.3 million adults in the United States met the definition for hypertension according to the 2017 American College of Cardiology/American Heart Association (ACC/AHA) guideline, which represents approximately 45.6% of the adult population [2]. Hypertension is a treatable condition, as management includes healthy lifestyle modifications and anti-hypertensive therapy [3]. Although a wide range of treatment options are available for blood pressure control, many hypertensive individuals remain with uncontrolled hypertension [4].

Additionally, resistant hypertension is defined as persistent increase in blood pressure that remains above goal despite use of three or more antihypertensive agents of different classes, one of them being a diuretic, at the best tolerated doses [5, 6]. Several studies demonstrate that the prevalence of resistant hypertension can be predicted at ranges between 10 and 15% of treated hypertensive patients [7]. Therefore, novel drugs are needed that display multiple therapeutic effects due to the fact that the majority of current antihypertensive drugs require combination therapy based on their limited efficacy and side effects. Thus, the study of natural products, which are an important source of compounds with antihypertensive activity, can be used as a prototype to design more potent and safer drugs for the treatment of hypertension [8].

Phyllanthimide is an alkaloid isolated from *Phyllanthus sellowianus* (Euphorbiaceae) [9], which presents antispasmodic activity [10, 11]. This alkaloid was used as a prototype for the development of analog compounds belonging to the chemical class of cyclic imides [12, 13], which contain the group –CO–N(R)–CO–, are electrically neutral and hydrophobic molecules [14]. In addition, studies have demonstrated that cyclic imides are biologically active molecules in the cardiovascular system, promoting vasorelaxant [15, 16] and hypotensive effects [15].

In this way, cyclic imides are compounds with potential biological activities on the cardiovascular system, however to date, these molecules have been poorly studied in this system. It's important to evaluate the potential of these substances on cardiovascular system, thus a variety of cyclic imides were synthesized, in order to investigate their effects on the cardiovascular system. Specifically, *N*-phenyl-itaconimide (Imide-1), *N*-4-methyl-phenyl-itaconimide (Imide-2), *N*-4-methoxy-phenyl-itaconimide (Imide-3) and *N*-4-chlorophenyl-itaconimide (Imide-4), were synthesized to test their properties on the cardiovascular system, which has yet to be investigated (Fig. 1). Therefore, in this study, we aimed to investigate, for the first time, the cardiovascular activities of these itaconimides in the vascular and cardiac tissues, with propose of describing the mechanisms of action involved in the observed responses.

Material and methods

Animals

Male Wistar rats weighing 200–300 g were used in all experiments. The animals were supplied by the animal facility of the Neuroscience Laboratory, at the Institute of Health Sciences at the Federal University of Bahia and kept under controlled temperature $(21 \pm 1 \,^{\circ}C)$ a light–dark cycle of 12 h (6 a.m. to 6 p.m.) and free access to food and water. The study was conducted in accordance with the guide of care and use of laboratory rats, adopted by the National Council for Animal Experiments Control (CONCEA—BRAZIL) and National Institutes of Health guide for the care and use of Laboratory animals (NIH Publications No. 8023, revised 1978). It was approved by the Ethics Committee on Animal Use from the Institute of Health Sciences, Federal University of Bahia (CEUA/UFBA n°120/2017).

Fig. 1 Chemical structure. Chemical structure of *N*-phenyl-itaconimide (Imide-1), *N*-4-methyl-phenyl-itaconimide (Imide-2), *N*-4-methoxy-phenylitaconimide (Imide-3) and *N*-4-chloro-phenyl-itaconimide (Imide-4)



Drugs

The drugs used in this study were: (-)-phenylephrine hydrochloride, acetylcholine chloride, cremophor, nifedipine, sodium nitroprusside, SKF 96365, cyclopiazonic acid, dimethyl sulfoxide (DMSO), ketamine, xylazine, heparine, all acquired from Sigma-Aldrich (Sigma Chemical Co., Saint Louis, MO, USA). The cyclic imides were obtained from the itaconic anhydride with different aromatic amines [17]. For the preparation of the itaconimides solutions, drugs were solubilized in cremophor and diluted to the desired concentrations with distilled water (in vitro assay) or NaCl 0.9% (in vivo assay) just before use. The final concentration of cremophor in the organ bath never exceeded 0.003% and had no effect when tested in control preparations. Nifedipine, SKF 96365 and cyclopiazonic acid was dissolved in DMSO 100%. The other compounds were freely dissolved in distilled water. Tyrode's physiological solution was used throughout all arterial experiments with the following compositions (mM): NaCl, 158.3; KCl, 4.0; CaCl₂, 2.0; MgCl₂, 1.05; NaH₂PO₄, 0.42; NaHCO₃, 10.0, and glucose, 5.6. All reagents were purchased from Sigma (Sigma-Aldrich, St. Louis, USA).

Vascular studies using superior mesenteric artery

Wistar rats were euthanized in a CO₂ chamber. The superior mesenteric arteries were quickly removed as described by Silva and colleagues [18], and cleaned from connective tissue and fat, and segmentally cut into rings (2 mm), which were suspended in organ baths containing Tyrode's solution, bubbled with 95% O₂/5% CO₂, and maintained at 37 ° C and pH 7.4. Tissues were stabilized with an optimal resting tension of 7.5 mN for 60 min. Isometric tension of the superior mesenteric arterial ring was recorded by a force transducer (Insight, Ribeirão Preto, SP, Brazil) coupled to an amplifier-recorder (Insight, Ribeirão Preto, SP, Brazil) and to a computer equipped with a data acquisition software. An intact endothelium was assessed by the ability of Ach $(1 \mu M)$ to induce 90% or more of relaxation of rings pre-contracted with Phe $(1 \mu M)$. In the endothelium denuded rings, the relaxation to Ach was less than 10%. When necessary, the endothelium was mechanically removed by gentle rubbing the vessel with a thin wire.

Effect of itaconimides on the vasculature

After the stabilization period, two successive contractions of similar magnitude were produced by Phe, 1 μ M, (alpha₁-adrenergic agonist) in endothelium-intact and -denuded rings. In the tonic phase of the second contraction, different concentrations of itaconimides (Imide-1, 2, 3 and 4; 3×10^{-8} to 3×10^{-4} M) were cumulatively added to the organ bath and the effects were compared. Additionally, these itaconimides were also added when an endothelium-intact vessel was at basal tone in order to examine the effect of itaconimides on spontaneous muscle tone.

Evaluation of the K⁺-channels activity in the vasorelaxant response induced by Imide-3

Different concentrations of Imide-3 $(3 \times 10^{-8} \text{ to } 3 \times 10^{-4} \text{ M})$ were added cumulatively to the organ bath to evaluate if Imide-3 can attenuate the relaxation of vessels pre-contracted with 60 mM KCl or phenyle-phrine in Tyrode's solution with 20 mM KCl in endothe-lium-denuded rings.

Investigation of the effects induced by Imide-3 in Ca²⁺ influx

To investigate the effects of imide-3 on Ca^{2+} influx in endothelium denuded rings, cumulative concentrations of $CaCl_2$ ($10^{-6}-10^{-2}$ M) were added in Tyrode's solution (60 mM KCl) nominally without Ca^{2+} (15 min), depolarization medium, in the absence (control) or presence of different itaconimide concentrations (10^{-6} , 10^{-5} , 3×10^{-5} and 10^{-4} M).

In another set of experiments, the effects induced by imide-3 on Ca²⁺ influx through other channels instead of voltage-sensitive Ca^{2+} channels (Ca_{y}), such as, receptoroperated channels (ROC) and store-operated channels (SOC), was also investigated. First, rings were maintained in a Tyrode's solution nominally without Ca²⁺ and incubated with cyclopiazonic acid (20 µM; 20 min), inhibitor of the sarco/endoplasmic reticulum Ca²⁺-ATPase (SERCA), Phe (1 μ M; 5 min) and nifedipine (1 μ M; a Ca_v blocker; 20 min), which was then followed by a single application of $CaCl_2$ (10⁻² M) to induce contraction. In the same vessel, this experimental protocol was repeated by pre-incubation with cyclopiazonic acid, Phe and nifedipine following incubation of imide-3 at different concentrations $(10^{-5}, 3 \times 10^{-5} \text{ and } 10^{-4} \text{ M})$ for 10 min, which was followed by a single application of CaCl₂. The role of imide-3 on Ca²⁺ influx, most likely through SOC and ROC was assessed by comparing CaCl₂ mediated contraction in the absence or presence of different concentrations of imide-3 with nifedipine/Phe/cyclopiazonic acid. Another experiment was performed to investigate the role of SOC, where endothelium-denuded rings were pre-incubated with SKF 96365 $(10^{-5} \text{ M}, \text{ a SOC blocker})$, for a period of 30 min, before adding Phe (1 µM), and then concentration-response to imide-3 $(3 \times 10^{-8} \text{ to } 3 \times 10^{-4} \text{ M})$ was initiated.

Atrial tissue preparation

Rats were euthanized in a CO₂ chamber, hearts were quickly removed and the isolated left and right atria were maintained in an organ bath with krebs-bicarbonate (117.0 mM NaCl, 5.36 mM KCl, 2.5 mM CaCl₂, 1.03 mM KH₂PO₄, 0.57 mM MgSO₄, 25.0 mM NaHCO₃, and 11.1 mM glucose) at a temperature of 37 °C and gassed with 95% O₂ and 5% CO₂. Isometric tension recordings of suspended atria were performed. The initial tension of each atrium was adjusted to 5.0 mN and the tissues were maintained during a stabilization period for 45 min before beginning the experiments. The isometric tension was recorded by a force transducer (FORT-10; WPI, Sarasota, FL, USA) connected to an amplifier-recorder (Miobath-4, WPI, Sarasota, USA). In each left atrium, used for cardiac inotropic analyses, was measured the excitation threshold, the minimum voltage of electrical stimulation required to elicit sustained synchronous contractions of the whole atrium. During the experiment, the left atrium was continuously electrically stimulated by square voltage pulses with a frequency of 3 Hz for 3 ms to 1.5 times the excitation threshold voltage of each heart tissue, in order to ensure that all cardiomyocytes were excited, as previously described [19], with an AVS stimulator (AVS, Brazil). Right atrium were measured for rhythmic and spontaneous organic contractions and imide-3 was added in 3 different concentrations $(3 \times 10^{-7}, 3 \times 10^{-6} \text{ and } 3 \times 10^{-5} \text{ M})$.

Influence of imide-3 on blood pressure (BP) and heart rate (HR)

On the day prior to animal experimentation where rats were anesthetized with ketamine/xylazine (75 mg/kg and 20 mg/kg ip, respectively), a polyethylene catheter (PE-10 connected to PE-50) filled with heparinized saline solution (1000 U/mL) was inserted into the abdominal aorta through the left femoral artery to measure the pulsatile arterial pressure (PAP). A second catheter was inserted into inferior vena cava through the left femoral vein for intravenous drug administration. Twenty-four hours following these surgical procedures, experiments were performed in nonanesthetized rats, under conscious and free-moving conditions. After 30 min of cardiovascular parameters stabilization, sodium nitroprusside (10 µg/kg, iv) was administered to verify the effectiveness of the venous catheter implantation and subsequently different doses of Imide-3 (0.1, 1 and 10 mg/kg randomly, iv) were administered. The PAP were continuously monitored through the arterial catheter connected to a blood pressure transducer (AD Instruments) whose signal was amplified and digitally recorded by an analog-to-digital interface (Power/Lab 8 35, application for data acquisition, LabChart, New South Wales, Austrália) and recorded (1 kHz) on a microcomputer for later analysis.

The mean arterial pressure (MAP), systolic arterial pressure (SAP), diastolic arterial pressure (DAP) and heart rate (HR) were obtained from the PAP.

Data analysis

The values were expressed as means \pm standard error of the mean (S.E.M.), and *n* represents a number of rings obtained from different rats. All calculations were performed using GraphPad Prism© software, version 5.0 (GraphPad Software Inc., La Jolla, CA, USA). The maximal effect (E_{max}) corresponds to maximum relaxation of the itaconimides, and pD₂ to negative logarithm of the concentration of a substance that induces 50% of maximal effect ($-\log EC_{50}$). The pD₂ was calculated using non-linear regression. For statistical analyses, Student's *t* test and one-way ANOVA was used to compare 2 and 3 or more groups, respectively, followed by Bonferroni's multiple comparisons post-test. Values of p < 0.05 were considered statistically significant.

Results

Itaconimides induce endothelium-independent-relaxation in mesenteric artery

As shown in Fig. 2, imides-1, 2, 3 and 4 $(3 \times 10^{-8} \text{ to}$ 3×10^{-4} M) induced concentration-dependent relaxation in superior mesenteric artery segments with intact endothelium, pre-contracted with phe (1 µM). Imide-1 $(E_{\text{max}} = 92.9 \pm 14.1\%; \text{ pD}_2 = 5.08 \pm 0.15, n = 7), \text{ imide-2}$ $(E_{\text{max}} = 93.7 \pm 7.9\%; \text{ pD}_2 = 4.63 \pm 0.07, n = 7)$, imide-3 $(E_{\text{max}} = 108.7 \pm 3.8\%; \text{pD}_2 = 4.57 \pm 0.04, n = 7)$ and imide-4 $(E_{\text{max}} = 103.7 \pm 4.2\%; \text{pD}_2 = 4.67 \pm 0.04, n = 6)$. The efficacy and pharmacological potency for endothelium-denuded rings imide-1 ($E_{\text{max}} = 103.8 \pm 10.4\%$; pD₂ = 4.72 ± 0.11, n = 7), imide-2 ($E_{\text{max}} = 102.4 \pm 4.4\%$; pD₂ = 4.68 ± 0.07, n = 6), imide-3 ($E_{\text{max}} = 107.5 \pm 6.3\%$; pD₂ = 4.64 ± 0.09, n = 6) and imide-4 ($E_{\text{max}} = 100.2 \pm 2.6\%$; pD₂ = 4.69 ± 0.05, n=7) were statistically similar to endothelium intact. Thus, there were no statistical differences among the maximum efficacy of itaconimides. In addition, imide-1 showed higher pharmacological potency than imide-2 (**p < 0.05), Imide-3 (**p < 0.01) and imide-4 (*p < 0.05). Furthermore, there was no significant change in basal tone following administration of itaconimides (data not shown). Imide-1 was the most potent drug tested, however, the substance was quite unstable. Since the other itaconimides presented similar potency, we selected the imide-3 to characterize the possible mechanisms involved in pharmacological actions.



Fig. 2 Effects of itaconimides in superior mesenteric artery. Representative original recordings of the effects of itaconimides imide-1 (**a**), imide-2 (**b**), imide-3 (**c**) and imide-4 (**d**) on the isolated mesenteric artery rings, pre-contracted with phenylephrine. The arrows represent the time-course of the itaconimides administration $(3 \times 10^{-8} \text{ to } 3 \times 10^{-4} \text{ M})$. Concentration–response curves (**e**–**h**) showing the relaxant effect of itaconimides on the isolated mesenteric artery rings with endothelium-intact (E+, \blacksquare) or endothelium-denuded (E–, \blacklozenge)

Imide-3 inhibited contraction induced by depolarization with High K⁺ concentration

In endothelium-denuded rings, imide-3 $(3 \times 10^{-8} \text{ to } 3 \times 10^{-4} \text{ M})$ inhibited the sustained tonic contraction induced

and bar graph showing the E_{max} and pD_2 values in different condition. Bar graph comparing the E_{max} (i) and pD_2 (j) of itaconimides on the isolated mesenteric artery rings with endothelium-intact. Results are expressed as mean ± S.E.M. e Imide-1, E+ (n=7) and E- (n=7); f Imide-2, E+(n=7) and E- (n=6); g Imide-3, E+(n=7) and E-(n=6) and h Imide-4, E+(n=6) and E- (n=7). Statistical analysis was performed using unpaired Student's t tests

by 60 mM KCl in a concentration-dependent manner. The response of imide-3 was not significantly changed when compared to rings contracted with phe $(1 \mu M)$ (Fig. 3).



Fig. 2 (continued)

Effect of Imide-3 on arteries treated with 20 mM KCI

To evaluate the involvement of K^+ channels in the vasorelaxant response induced by Imide-3, mesenteric rings lacking endothelium, pre-contracted with phenylephrine were pre-treated with 20 mM KCl. Imide-3 did not alter the concentration–response curve when compared to the control. This finding indicates that K^+ channel activation does not play a role in the relaxant effect of Imide-3 in mesenteric arteries (Fig. 3).



Fig. 3 Effect of Imide-3 on the contractions induced by phenylephrine, KCl 60 mM or KCl 20 mM with phenylephrine. Relaxation responses induced by Imide-3 (3×10^{-8} to 3×10^{-4} M) in denuded rat mesenteric arteries rings pre-contracted with phenylephrine (1 μ M) (\bullet , n=6), KCl 20 mM+phenylephrine (1 μ M) (\blacksquare , n=6) and KCl 60 mM (\blacktriangle , n=6). Results are expressed as mean ± S.E.M. Statistical analysis was performed using one-way ANOVA followed by the Bonferroni post-test

Effect of Imide-3 on the CaCl₂-induced concentration-response curves

As shown in Fig. 4, the concentration–response curve to CaCl₂ in a depolarizing medium, while in the presence of Imide-3 (10^{-6} or 10^{-5} or 3×10^{-5} or 10^{-4} M) was shifted to the right when compared with the control. The maximal contraction of CaCl₂ was significantly (***p < 0.001) attenuated by 10^{-5} M, 3×10^{-5} M and 10^{-4} M concentrations of imide-3 ($69.5 \pm 10.6\%$, $36.3 \pm 9.0\%$, $-3.2 \pm 3.9\%$, respectively). These data indicate that the mechanism of action of imide-3 involves the reduction of Ca²⁺ influx.

Imide-3 reduced Ca²⁺ influx through ROC and SOC

The influence of Imide-3 on Ca²⁺ influx through ROC and SOC was evaluated in experiments performed with Tyrode's solution nominally without Ca²⁺ in the presence of Cyclopiazonic acid/Phe/Nifedipine. Figure 5 demonstrates that the contraction evoked by CaCl₂ in Tyrode's solution nominally without Ca²⁺, following intracellular Ca²⁺-store depletion and blocking Ca_v, no modification of the E_{max} values was observed following pretreatment using different concentrations (10⁻⁵ or 3×10⁻⁵ M) of Imide-3 (25.3±2.2%, 22.5±4.6%, respectively) compared to control (29.0±2.6%). However, the concentration of 10⁻⁴ M significantly reduced the contraction (7.0±1.9%).



Fig. 4 Effect of Imide-3 on Ca²⁺ influx. Concentration–response curve of CaCl₂ on rat mesenteric artery segments without endothelium, in the absence (\bigoplus , n=28) or in the presence of Imide-3 (\blacksquare , 10^{-6} M, n=8), (\blacktriangle , 10^{-5} M, n=7), (\blacktriangledown , 3×10^{-5} M, n=7) and (\blacklozenge , 10^{-4} M, n=6). *p < 0.05 versus control. **p < 0.01 versus control. **p < 0.01 versus control. Results are expressed as mean \pm S.E.M. Statistical analysis was performed using one-way ANOVA followed by the Bonferroni post-test



Fig. 5 Effect of Imide-3 on the contraction induced by CaCl₂ in the presence of phenylephrine, nifedipine and cyclopiazonic acid. Influence of Imide-3 (10^{-5} , 3×10^{-5} and 10^{-4} M) on CaCl₂ (10^{-2} M) mediated contraction in Tyrode's solution nominally without Ca²⁺, in the presence of phenylephrine (Phe) (1 µM), nifedipine (1 µM) and Cyclopiazonic acid (CPA) (20 µM). Imide-3 (10^{-5} M, n=7), (3×10^{-5} M, n=6) and (10^{-4} M, n=7). ***p < 0.001 Phe+CPA+nifedipine versus Imide-3 (10^{-4} M). Results are expressed as mean ± S.E.M. Statistical analysis was performed using one-way ANOVA followed by the Bonferroni post-test



Fig. 6 Vasorelaxant effect from Imide-3 in the presence of SOCinhibition. Relaxation responses induced by Imide-3 $(3 \times 10^{-8} \text{ to} 3 \times 10^{-4} \text{ M})$ in denuded rat mesenteric arteries rings, pre-contracted with phenylephrine, in the presence or absence of SKF 96365 (10^{-5} M) . Results are expressed as mean ± S.E.M. (*n*=6). Statistical analysis was performed using unpaired Student's *t* tests

Effect of SOC inhibition on the relaxation induced by imide-3

Figure 6 shows that SOC inhibition by SKF 96365 in endothelium-denuded rings did not attenuate relaxation induced by imide-3. This result suggests that SOC do not participate in the mechanism of action of imide-3.

Evaluation of the direct effect of Imide-3 on the isolated atria

The influence of imide-3 on chronotropism was analyzed based on isolated right atrium spontaneous beating frequency, quantified and defined as atrial rate. The addition of three cumulative concentrations of Imide-3 $(3 \times 10^{-7} \text{ M}, 3 \times 10^{-6} \text{ M})$ and 3×10^{-5} M) did not result in a significant negative chronotropic effect $(99.3\% \pm 1.63, 98.0\% \pm 1.33, 97.4\% \pm 1.22,$ respectively) compared to control (100 ± 0) (Fig. 7a). Moreover, with the purpose of analyzing the effect of Imide-3 on the strength of cardiac contraction (inotropic effect), these concentrations of Imide-3 were added to the isolated left atrium preparation. The concentrations of 3×10^{-7} M and 3×10^{-6} M were did not significantly reduce force of cardiac contraction $(86.1\% \pm 14.4, 61.5\% \pm 15.6, respectively)$. However, the last tested concentration, 3×10^{-5} M, demonstrated a significant negative inotropic effect $(47.0\% \pm 15.1)$ compared to control (100 ± 0) (Fig. 7b, c).

Imide-3 induces hypotensive effects in normotensive rats

Hemodynamics changes induced by Imide-3 (0.1; 1 and 10 mg/kg, *iv*, administered randomly) were investigated in normotensive non-anesthetized rats. Imide-3 administration did not alter blood pressure and heart rate at the following doses: 0.1 and 1 mg/kg (-2.06 ± 0.44 ; -6.48 ± 1.89 mmHg and 0.24 ± 0.86 ; 4.18 ± 2.17 bpm; respectively, n = 5). However, the 10 mg/kg dose induced hypotension associated with a decrease in heart rate (-57.3 ± 6.83 mmHg and -78.1 ± 1.76 bpm, n = 5) (Fig. 8).

Discussion

This study demonstrates that the itaconimides, imide-1, imide-2, imide-3 and imide-4, induce a pronounced endothelium-independent vasorelaxation in isolated rat superior mesenteric artery rings, mainly thought the inhibition of extracellular Ca^{2+} influx. We also demonstrated that imide-3 induced hypotensive and bradycardic effects in non-anaesthetized rats and negative inotropic action in isolated atria. Additionally, the reduction in peripheral vascular resistance and negative inotropic action could be, at least in part, responsible for the hypotensive and bradycardic effects promoted by imide-3. To the best of our knowledge, this is the first work that shows these cardiovascular effects of itaconimides.

The endothelium plays a pivotal role in regulating vascular tone by synthesizing and releasing an array of endothelium-derived relaxing factors, including nitric oxide (NO), prostacyclin (PGI₂), and endothelium-dependent hyperpolarization factors (EDHF), as well as endothelium-derived contracting factors [20, 21]. To evaluate the participation of the endothelium modulating itaconimides actions, we performed experiments in the absence and presence of a functional endothelium. Our experiments demonstrated that itaconimides relaxed contractions induced by phenylephrine, in a concentration-dependent manner, with similar pharmacological potency in endothelium-denuded and in endothelium-intact artery rings. This indicates that the vasorelaxant effects of itaconimides are independent of vasoactive mediator release from endothelium and that these itaconimides may act directly on the smooth muscle cell. However, another cyclic imide, 4-Nitro-N-phenylmaleimide, has been demonstrated to produce an endothelium-dependent and -independent vasorelaxation in superior mesenteric artery rings [16].

Moreover, the four itaconimides showed similar maximal effect. However, Imide-1 demonstrated a higher pharmacological potency than the other itaconimides tested, thus the most active itaconimide was unsubstituted compound.



Fig. 7 Effect of Imide-3 on isolated right and left rat atria. Bar graph showing the chronotropic effect of Imide-3 $(3 \times 10^{-7}, 3 \times 10^{-6} \text{ and } 3 \times 10^{-5} \text{ M})$ on isolated right rat atria (**a**). Representative tracings showing the effect of Imide-3 on left atrial contractile force (**b**). Bar

graph showing the negative inotropic effect of Imide-3 on isolated left rat atria (c). Results are expressed as mean \pm S.E.M. (n=5). *p <0.05 versus control. Statistical analysis was performed using one-way ANOVA followed by the Bonferroni post-test

These results suggest that there may be some unfavorable steric effects due to the substitution of the Imide-2 ($-CH_3$), Imide-3 ($-OCH_3$) and Imide-4 (-Cl) in the fourth position of the benzene ring because the introduction of these substituents (chloro, methyl and methoxy-substituent) decreased the pharmacological potency. As well, all itaconimides induced vasorelaxant activity, and therefore the complete structure is important for the interaction with the biological target, emphasizing the importance of the imidic ring for the biological effect. Under our experimental conditions, all

vasodilator responses to itaconimides 1, 2 and 3 were reversible until the concentration of 3×10^{-5} M, therefore these vasorelaxant actions are not related to putative toxic effects (data not shown).

Furthermore, itaconimides were unable to alter the resting tone of arterial smooth muscle. Thus, these results may indicate that the itaconimides do not present vasoconstriction or vasodilation activity on basal tone, however itaconimides may possibly induce vasorelaxation only in pre-contracted mesenteric artery rings. Moreover, the most



Fig.8 Effects of intravenous administration of Imide-3 on the mean arterial pressure and heart rate in normotensive rats. Representative tracing showed the hypotensive response and bradycardia, induced by addition of Imide-3 (10 mg/kg) in conscious normotensive rats (**a**). Bar graphs showing changes in mean arterial pressure and pressure rate (**a**).

potent itaconimide was imide-1, but the substance was quite unstable. Since the other itaconimides presented similar pharmacological potency, the imide-3 was chosen to characterize the possible mechanisms involved in pharmacological actions. In the present study, imide-3 inhibited KCl (60 mM)-induced contractions, suggesting that itaconimide might interfere with electromechanical coupling, and the concentration-response curve in this condition was similar to the response obtained in the presence of Phe. It is well known that KCl solution may induce contraction of the vascular smooth muscle mainly due to depolarization and the influx of extracellular Ca^{2+} though Ca_v [22, 23] and release of calcium from the intracellular stores [24, 25]. While phenylephrine selectively activates α_1 -adrenergic receptors, resulting in the production of inositol trisphosphate (IP_3) and diacylglycerol (DAG). IP₃ increases Ca²⁺ by activating sarco/endoplasmic reticulum Ca2+ channels and Ca2+ release. DAG can activate protein kinase C and provoke

sure (%) (b). Changes on heart rate (%) (c). Results are expressed as mean \pm S.E.M. (n=5). ***p<0.001 versus vehicle (control). Statistical analysis was performed using one-way ANOVA followed by the Bonferroni post-test

 Ca^{2+} entry through transient receptor protein cation channels [26]. Our current data indicate that imide-3 acts in a possible common pathway the both contractile agents, receptor independent mechanism and promote a mechanism that could be related to inhibiting calcium influx activity.

Another important mechanism that promotes vasorelaxation by a direct action on vascular smooth muscle cells can involve potassium channels. The opening of K⁺ channels causes diffusion of this cation out of the cells, resulting in membrane hyperpolarization and consequently decreases in the open probability state of Ca_v, ultimately leading to vasorelaxation [27–29]. Furthermore, elevated extracellular potassium ion concentration reduces the electrochemical gradient for K⁺ efflux and attenuates vasorelaxation mediated by K⁺ channels activation [30]. In our study, the increase in extracellular K⁺ concentrations (from 4 to 20 mM) did not significantly attenuate the vasorelaxation induced by imide-3, which was also observed when rings



Fig. 9 Schematic representation of the possible signaling pathway of the vasorelaxant response induced by Imide-3. Imide-3 acts blocking Ca^{2+} influx by Ca_v and ROC, reducing intracellular calcium concentration and leading to vasodilation in smooth muscle cells. Additional studies will be needed to assess whether the inhibition of SOC do

not participate of the mechanism of action of this itaconimide. IP_3R , inositol-1,4,5-trisphosphate receptor; RyR, Ryanodine receptor; Ca^{2+} , calcium ion; ROC, receptor-operated channels; SOC, store-operated channels; Ca_v, voltage-sensitive Ca²⁺ channels

were exposed to 60 mM KCl, suggesting that K⁺ channel activation does not appear to be involved. Moreover, we evaluated whether vasorelaxation induced by imide-3 is associated with the inhibition of Ca^{2+} influx. There are different Ca²⁺ entry channels, such as Ca_v, ROC and SOC in smooth muscle cells. Ca_v are the major way by which Ca²⁺ enters vascular smooth muscle cells and their function are regulated by membrane potential, so that depolarization opens these channels, leading to vasoconstriction [31]. Imide-3 markedly reduced the Ca²⁺-induced vasoconstriction in mesenteric rings lacking endothelium that were preincubated with a high K⁺ solution (KCl 60 mM) nominally without Ca²⁺, suggesting that Ca²⁺-influx through Ca_v channels was probably inhibited. This result corroborates previous observations describing cyclic imides, phyllanthimide and 4-Nitro-N-phenylmaleimide, that decrease the influx of Ca^{2+} in smooth muscle cells [10, 16, 32]. However, patch clamp technique is needed to confirm a blocking effect of imide-3 on Ca_v in smooth muscle cells.

Next, another experimental protocol was performed to assess whether the effect of imide-3 on inhibiting Ca^{2+} -influx could involve an additional calcium influx inhibition pathways, such as SOC and ROC. To evaluate this, arterial rings were studied in a nominally Ca^{2+} -free Tyrode solution in the presence of cyclopiazonic acid (20 µM), used to deplete intracellular Ca^{2+} stores and thus activate SOC [31, 33]. Ca^{2+} depletion in the sarco/endoplasmic reticulum is followed by a stimulated Ca^{2+} entry via SOC, replenishing intracellular calcium stores. Thus, SOC are inhibited when Ca^{2+} stores are filled [31, 34]. Furthermore, Phe evokes contraction by activation of ROC and Ca_{V} [35]. Furthermore, ROC are activated by ligand-mediated activation of G protein coupled receptors (GPCRs) and receptor tyrosine kinase (RTK), which after ligand binding activate phospholipase C, resulting to IP₃ and DAG production [36]. ROC play an important role in arterial tone regulation in response to vasoactive agonists, such as angiotensin II and α -adrenoceptor agonists [37]. Moreover, depolarization may occur after activation of ROC, leading an additional Ca^{2+} influx through Ca_{v} [38]. In our experiment, to exclude the participation of Ca_v , nifedipine (10⁻³ M) was used, which blocks voltage-gated L-type Ca^{2+} channels [39, 40], but ROC are unaffected by this blocker [41]. Under these conditions, the highest tested concentration of imide-3 significantly attenuated the CaCl₂-induced contraction. These results indicate that the vasorelaxant response of imide-3 may also involve, at least in part, the inhibition of Ca^{2+} influx through ROC and SOC. In addition, experiments were performed to assess the involvement of SOC in relaxation induced by imide-3, using the nonspecific channel blocker, SKF 96365. The results indicate that the itaconimide-mediated relaxation was similar in the presence and absence of SKF, indicating a lack of inhibition of Ca²⁺ entry through SKF-sensitive channels by imide-3. Therefore, the vasodilatory effect induced by imide-3 may be due to a possible influence on the $Ca_{\rm V}$ and ROC.

Since the itaconimide induced a marked vasodilation by Ca^{2+} -influx inhibition, we hypothesized that imide-3 could induce negative inotropic effect. To investigate this, experiments were carried out to evaluate the direct effect of imide-3 on the isolated atria preparations. Imide-3 induced a negative inotropic effect without a significant change in cardiac rhythmicity, but the mechanisms by which imide-3 acts in heart remain still unclear.

In vivo experiments were performed to investigate whether Imide-3 alters cardiovascular parameters in nonanesthetized normotensive rats. In these animals, we observed that intravenous administration of imide-3 induced a hypotensive response associated with bradycardia. The hypotensive effect may be due to reduced peripheral vascular resistance, and the bradycardia can be a direct or indirect action on cardiac activity.

Taken together, these results suggest that the itaconimides evaluated in this study is able to induce vasorelaxation, independent of endothelium-derived relaxing factors. In addition, imide 3 induces vasodilatation, most likely due to the inhibition of calcium influx through Ca_v and ROC (Fig. 9) and it also induces negative inotropic effect. These effects probably are responsible by the decreasing of blood pressure and bradycardia observed in the in vivo experiments. In this way, the results observed were promising, showing the possibility of new therapeutic option for hypertensive patients, in the future. However, further studies will be necessary to better clarify the mechanisms of action of the imide-3 on the cardiovascular system.

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Compliance with ethical standards

Conflict of interest The authors declare that there are no conflicts of interest. The founding sponsors had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, and in the decision to publish the results.

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