Review Paper

A review on environmental selenium issues

Getachew Dagnew Gebreeyessus1 [·](http://orcid.org/0000-0001-5171-0465) Feleke Zewge2

© Springer Nature Switzerland AG 2018

Abstract

Selenium is an essential element since it is necessary for metabolism and is also toxic to the living things if it is consumed above the desirable limit. For instance, up to 40 µg per d of dietary selenium is essential for human health; however, consumption of a little over 400 µg per d could result in adverse physiological problems. In the environment, selenium occurs in diferent species; of which selenate and selenite are its most toxic forms accounting for 95% of selenium toxicity. Though, selenium fortifcation is important for normal human physiology in selenium defcient environments, it needs to be minimized or removed from selenium rich environmental media. Thus, its quantifcation is signifcantly important to control per limit. Currently there is advancement in the detection and quantifcation of selenium in environmental including the use of a Hydride Generating Atomic Absorption Spectroscopy. Due to the excess occurrences of selenium in diferent locations the removal techniques from geogenic or anthropogenic sources include microbial metabolism, phytoremediation, sorption, membrane separation, coagulation, ion exchange and catalytic reduction. Most of all, the microbial removal is applied in drinking water supply and in selenium recovery from wastewater sludge. Though current selenium removal from sorption studies reported better efficiency, phytoremediation appears attractive due to its field scale application advantage. However, the fate of selenium following phytoremediation remains unaddressed.

Keywords Selenium · Toxicity · Occurrence · Environment

1 Introduction

Both geogenic and anthropogenic activities govern selenium in the environment. Environmental biogeochemical conditions control the bioavailability and the nature of selenium species [\[1](#page-15-0)[–5\]](#page-15-1). Geogenic pollution of groundwater is referred to as the natural occurrence of elevated concentration of certain elements in groundwater hav-ing negative health effects [[6](#page-15-2)]. In fact, selenium is listed among the minor constituents in geogenic water pollution $(0.1-100 \mu g/l)$. Moreover, the narrow tolerance limit of selenium brings serious concerns through defciency as well as toxicity in the biological and environmental systems. Toxicity generally results when certain ions are taken up with the soil–water and accumulate in some organs of plants and animals. Though geogenic selenium

contamination is a concern for toxicity from drinking water, plants are the main source of dietary selenium [[7\]](#page-15-3).

By elemental category, selenium is nonmetal and sometimes it is considered as trace element. Geographically, it is widely distributed throughout the world, but only in small quantities. It is more commonly found together with the sulfdes as the selenides in the ores of such metals as iron, lead, silver, and copper [\[8\]](#page-15-4). Selenium salts are toxic in large amounts, but trace amounts are necessary for cellular function in many organisms, including all animals [[9](#page-15-5), [10\]](#page-15-6).

Fortifed selenium is an ingredient in many multivitamins and other dietary supplements, including infant formula in many countries. In human, low dietary selenium intakes are associated with health disorders including oxidative stress-related conditions, reduced fertility and immune functions and an increased risk of cancers. For

 \boxtimes Getachew Dagnew Gebreeyessus, getachewdagn@yahoo.com | ¹Department of Urban Environmental Management, Kotebe Metropolitan University, P.O. Box 31248, Addis Ababa, Ethiopia. ²African Center of Excellence for Water Management, Addis Ababa University, P.O. Box 1176, Addis Ababa, Ethiopia.

SN Applied Sciences (2019) 1:55 | https://doi.org/10.1007/s42452-018-0032-9

Received: 27 July 2018 / Accepted: 23 October 2018 / Published online: 22 November 2018

that purpose the daily intake in the diets difer worldwide, for UK adult female and male 60 and 75 µgSe/d respectively are expected while for the World Health Organization (WHO), it is 10 μ q/l in water [\[11](#page-15-7), [12](#page-15-8)]. Contrasting the former, a study in Burundi of Africa that reported risk in defciency with average intake of 17 µgSe/d from diet [[13\]](#page-16-0). In fact, intake depends on age and sex $[14]$ $[14]$ $[14]$. However, selenium turns out to be toxic in low margin beyond limit causing hair and nail loss, cancer and even death as detailed later in the toxicity section of this article.

Until recently a lot of research have been done on selenium either as toxic or as nutrient. As of the middle of the 1990s approximately 100,000 research papers had been published on the topic of selenium since it was discovered by the Swedish chemist Jons Jakob Berzelius in 1817 [[5](#page-15-1)]. However, it is only recently that its geogenic contamination and bioremediation, selenium recovery as resource, selenium detection methods, and occurrence as well as distribution especially in the African continent have begun to be discovered.

Therefore, the objective of this mini review is to comprehensively present the nature, geochemistry, toxicity, biochemistry, detection and quantifcation methods reported on selenium in the past researches. Further it systematically reports the remediation techniques applied to selenium at various scales and shows fllable gaps for further research.

2 Selenium property and toxicity

2.1 Physicochemical properties of selenium

The physical, atomic properties and periodic table position of selenium is shown in fgure one. Selenium appears

SN Applied Sciences A SPRINGER NATURE journal as black and red allotropes (Fig. [1](#page-1-0)). It was found out long ago that it's electrical resistance is greater in the light than in the dark [\[15,](#page-16-2) [16](#page-16-3)]. Analogous to the behavior of other chalcogens, selenium forms hydrogen selenide, H_2 Se. It is a strongly odiferous, toxic, and colorless gas. It is more acidic than H₂S. In solution it ionizes into HSe[−] [[17\]](#page-16-4).

Chemically, selenium is reactive towards many reagents allowing their incorporation into organic compounds. Further, symmetric or non-symmetric diselenides are often used as starting materials for the preparation of more complex chalcogen-containing derivatives [[8\]](#page-15-4). Selenium also forms complexes with heavy metals including cadmium, sparing its biological effects as reported earlier [\[18\]](#page-16-5).

Although the element selenium itself is not toxic, many of its compounds are extremely toxic. For instance, H_2 Se is a colorless, foul-smelling gas that is poisonous. It also forms selenides with most metals (for example, aluminum selenide, cadmium selenide, and sodium selenide). In groundwater, selenium exists in the dissolved forms of Se (+VI), Se (+IV), and Se (−II). Selenium (+IV) is more toxic due to its higher bioavailability.

2.2 Selenium in plants: toxic dose, symptoms and distribution

Though debatable, it has been reported that selenium is not an essential element to plants but it can be toxic to them [[19](#page-16-6)]. Generally, the plant toxicity of inorganic selenium, namely selenate and selenite anions, is higher than the organic forms of selenium depending on three major factors. Absorption of the anions by the roots and the subsequent translocation into other parts of the plant is the frst factor. Secondly, the metabolism of the anions into organic forms in the cell and thirdly the interference of cellular biochemical reaction in plants due to selenium metabolite of which replacing the role of sulfur [\[20\]](#page-16-7) and protein functions are worth to mention.

The distribution of selenium in plants is all over it, from the root to its aerial parts. However, there is dissimilarity between selenate and selenite in distribution. Furthermore, the toxicity and tolerance character vary among plants [\[21\]](#page-16-8). The later mainly depends on their ability to divert selenium away from the accumulation of selenocysteine and selenomethionine that ranges from 2 mg/kg in non-accumulators, such as rice, and 330 mg/kg in white clover, to several thousands of mg/kg in the accumulator Astragalus bisulcatus. For instance, in non-accumulator's selenium toxicity occurs under about 10–100 mg Se/kg of dry weight (DW). Toxicity occurs mostly due to the accumulation of selenium in the plant tissues. The characteristic symptoms due to selenium toxicity includes stunting, chlorosis, and fading of leaves, interfering with chlorophyl combination and nitrate accumulation that is generally

assumed to occur from selenium levels higher than 1 mg/ kg [\[22–](#page-16-9)[25](#page-16-10)].

2.3 Selenium in animals: toxic dose and health symptoms

Earlier, selenium is identifed as micronutrient for bacteria, mammals and birds earlier; however, it is now recognized as a problem if it is consumed in excess. A case in point is the alkali disease in livestock that was observed in the American great plains since then concern on chronic and acute selenosis raised [[26](#page-16-11)]. Based on testes on laboratory animals, the minimum lethal dose of selenium as acute toxicity in rabbits, rats, and cats was between 1.5 and 3.0 mg/kg body weight as sodium selenite or selenate and irrespective of the mode of entry of the compound into their body. During acute toxicity, laboratory animals showed garlicky breath odor, vomiting, dyspnea, tetanic spasms, and death from respiratory failure accompanied by pathological changes. Chronically, a selenium dietary dose of 4–5 parts per million (ppm) proved to cause growth inhibition [[27\]](#page-16-12).

In farm animal's selenium acute toxicity occurs due to the ingestion of seleniferous accumulator plants or due to excess supplements with following signs of severe distress that include labored breathing, abnormal movement and posture, and prostration and diarrhea, and even death in few hours [[27](#page-16-12), [28\]](#page-16-13). The chronic selenosis also known as alkali disease occurs in animals consuming grains containing 5–40 mg Se/kg over a period of several weeks or months with signs and symptoms that include liver cirrhosis, lameness, hoof malformations, loss of hair, and emaciation [[29](#page-16-14)].

The determination of selenium level in animal tissues and organ has been investigated along with its metabolism in sheep. Findings revealed that the accumulation of selenium varies among the diferent organs, the liver being the highest accumulator. Further the share of selenium through enzymatic extracted in the form of amino acid compounds was 40% [\[30\]](#page-16-15). In a related earlier study in Burundi of Africa, kidney of goats and cows is found to be the accumulator organ for selenium ranging 994–1852 ng/g wet weight for goats and 1056–1801 ng/g wet weight for cows. This fnding has relevance to note the safety and adequacy of nutrients for people in developing countries; however South Africa is the only African state on the list of seleniferous areas which rather mentioned Africa in relation to selenium defciencies in crops and livestock as reported in a recent study [\[31](#page-16-16), [32\]](#page-16-17).

Selenium toxicity in animals or plants edible to human is a primary concern since bio-magnifcation of selenium starts when animals consume plant biomass that has impounded large amounts of selenium and get magnifed in food chains that can involve aquatic selenium pollution [[33](#page-16-18), [34](#page-16-19)]. Besides, due to irrigation run-off concentrations of selenium tend to be very high in aquatic organisms in many areas. Thus, the bioaccumulation potential of selenium can further cause toxicity to the natural ecosystem. When animals absorb or accumulate extremely high concentrations of selenium it can cause reproductive failure and birth defects [[35–](#page-16-20)[37](#page-16-21)].

2.4 Selenium in humans with epidemiological perspective

Water, food and air in decreasing order are identifed earlier as source of human toxicity from selenium [[38](#page-16-22)]. Moreover, drinking water sources remains to be a concern in geogenic selenium contamination. For instance, in many French regions, geogenic sedimentary sources of selenium were identifed as responsible for the selenium concentrations exceeding the European limit in many wells designated to produce drinking water [[36\]](#page-16-20). Selenium toxicity is prevalent in 1000 ha in Hoshiarpur and Nawanshahar of Nigerian districts where about 11 and 4% of groundwater samples were found unft for drinking and irrigation purposes, respectively [[35](#page-16-23)].

Selenium carries the narrowest range between its nutritional deficiency ($<$ 40 μ g/d) and toxicity ($>$ 400 μ g/d) with respect to the daily intake [[17\]](#page-16-4) (Table [1\)](#page-3-0). Selenate and selenite have an oxidant mode of action in living organisms and are very toxic. In humans, the toxic efects of longterm high level environmental selenium exposure are manifested in dermatological efects, nails, hair and liver problems [\[39](#page-16-24)].

Selenium exposure can result in either acute or chronic health problems. An acute exposure is explained by selenium neurotoxicity while the chronic exposure is explained by the toxic efect on endocrine function especially in the synthesis of thyroid hormones and if the dose exposed is relatively lower [\[39,](#page-16-24) [40\]](#page-16-25). For instance, a report from China indicated that clinical and biochemical signs occur at a daily intake above 0.8 mg. Conversely, inadequate concentration of selenium in Chinese diet is a cause for Keshan disease which is the degeneration of articular cartilage between joints, thyroid disease and diferent forms of cancer [[26,](#page-16-11) [38](#page-16-22)].

To get the nutrient beneft from selenium diet fortifcation became an alternative. Care should be taken as in product labeling; otherwise, toxicity outbreak is inevitable as exhibited in the identifed outbreak due to the selenium in a liquid dietary supplement in a certain town in the United States. Based on epidemiological observation, the median estimated dose of selenium consumed was 41 749 μg/d with frequently reported symptoms that include diarrhea, fatigue, hair loss, joint pain, nail discoloration or

brittleness, and nausea [\[41,](#page-16-26) [42\]](#page-16-27). Nevertheless, acute toxicity problem is rare due to diet diversity despite the rising concern over environmental selenium contamination in the United States.

Elsewhere in New Zealand and Venezuela, acute selenosis was observed due to the selenium rich food consumption [[43](#page-16-28)]. In a related fact the daily intake of Venezuelan children with clinical signs was estimated to be about 0.7 mg. Efects on synthesis of a liver protein were also seen in a small group of patients with rheumatoid arthritis given selenium at a rate of 0.25 mg/d in addition to selenium intake through food. No clinical or biochemical signs of selenium toxicity were reported in a group of 142 persons with a mean daily intake of 0.24 mg (maximum 0.72 mg) from food [\[44\]](#page-16-29).

3 Environmental fate of selenium

Selenium in the whole environment can be either from natural or anthropogenic sources forming cycles between them (Fig. [2\)](#page-4-0). The natural bases of selenium include the weathering of selenium-containing rocks and soils and volcanic eruptions [[45](#page-16-30)]. Anthropogenically, selenium pollution in the environment is hugely contributed by the coal mining and the use of selenium rich fossil fuels in industries that can account 40% of selenium in air as well as the aquatic systems that resulted in wastewater discharge limit of 5 μ g Se/l [[34,](#page-16-19) [37\]](#page-16-21). In that regard, selenium contamination in groundwater and surface water in numerous river basins worldwide has become a critical issue in recent decades [[46](#page-16-31), [47](#page-16-32)], especially due to the geogenic contamination. In order to foresee the mineralization, mobilization, and toxicity extent of selenium to plants, animals and human in natural systems understanding of its oxidation state is important. In geologic formations, selenium transformation and speciation depend on the presence of nitrate $(NO₃)$, organic

materials and clay content, pH and carbonates/bicarbonates for mobility and hence the risk associated [[48–](#page-16-33)[51](#page-16-34)].

Selenium concentrations in soils vary widely, from 5 to 1, 200, 000 µg/kg, being higher in soils of more recent volcanic origins. It is normally found in concentrations ranging 50–90 µg/kg of soil, but higher concentrations can be associated with some volcanic, sedimentary and carbonate rocks. Selenium occurs in environmental soils in several forms, according to its possible oxidation states: selenides $(Se^{2−})$, amorphous or polymeric elemental selenium (Se⁰), selenites (Se⁴⁺) and selenates (Se⁶⁺) (IPCS, 1987; UK EGVM, 2002).

Regarding selenium speciation, acidic and reducing conditions reduce inorganic selenites to elemental selenium, whereas alkaline and oxidizing conditions favor the formation of selenates in the environment. Since selenites and selenates are both soluble in water, selenium is leached from oxic alkaline soils that favor its oxidation. In contrast, the elemental and selenides forms are insoluble in water; consequently, selenium inclines to be retained in wet and anoxic soils. Thus, selenium in alkaline soils is accessible by plants; however, the availability of selenium in acidic soils inclines to be restricted by the adsorption of selenites and selenates to iron and aluminum oxide soils (NRC 1983).

Based on the species extraction using 0.1 mol/l NaOH, all the four selenium compounds are present as anionic species based on their pKa values at a pH of 13. Being in state of dissolution or precipitation and absorption or desorption, selenium is present in almost all soils, particularly in irrigated agricultural aquifer systems underlain by or adjacent to shale formations containing seleno-pyrite where $SeO₄$ can be released through autotrophic reduction of O_2 or NO_3 [\[52\]](#page-16-35) (Eqs. [1a](#page-3-1), [1b](#page-4-1)).

(1a) 2FeSexS2−^x ⁺ 7O2 ⁺ 2H2O [→] 2Fe²⁺ ⁺ ²xSeO2[−] 4 + (4 − 2X)SO²[−] ⁴ + 4H⁺

Fig. 2 Selenium cycle in the environment near the surface of the earth. (Redrawn from [\[1\]](#page-15-0) with permission from John Wiley and Sons. © 2016 WILEY–VCH Verlag GmbH & Co. KGaA, Weinheim)

$$
5FeSe_xS_{2-x} + 14NO_3^- + 4H^+ \rightarrow 5Fe^{2+} + 5xSeO_4^{2-} + (10-5X)SO_4^{2-} + 7N_2 + 2H_2O
$$
 (1b)

Selenate is mobilized in oxidizing condition and is interfered by reducing condition as well as adsorption. Irrigation and agriculture are associated with Se mobility. In addition, selenium is subject to cycling in the plant–soil system similar to other nutrients such as nitrogen (N), phosphorus (P), and sulfur (S).

Selenium mobility and risk of toxicity are not limited to physico-chemical phenomena. Microbially-mediated chemical reduction reactions reduce $SeO₄$ to $SeO₃$ [Eq. ([2a\)](#page-4-2)] and SeO $_3$ to either immobile Se 0 [Eq. [\(2b](#page-4-3))] or mobile selenomethionine (SeMet) suggesting the possible mitigation pathways for $SeO₄$. Almost all of the selenium removal techniques include reduction of SeO₄ to SeO₃, SeO, or SeMet that in most case it is inhibited by the presence of O_2 and NO_3 which act as electron acceptors. However, it is suggested that there is a concentration of $NO₃$ at which $NO₃$ reduction and SeO₄ reduction can even occur simultaneously [[53\]](#page-16-36).

$$
CH_2O + 2SeO_4^{2-} \to CO_2 + 2SeO_3^{2-} + H_2O
$$
 (2a)

$$
CH_2O + SeO_3^{2-} + 2H^+ \to CO_2 + Se^0 + 2H_2O
$$
 (2b)

The risk of selenium toxicity in the environment is afected by its mobility. In that regard experimental study by Bassil and co-workers revealed that even though the less fraction (10%) of selenium was mobilizable, 70% of it was Se (IV) which is the most soluble species [\[54](#page-16-37)]. Environmentally, the selenium pool exists much in the lithosphere followed by the fossil fuel deposits and oceans (Fig. [3](#page-5-0)), thus, its geochemistry and hence the associated risk is also afected by climate variability [\[55](#page-16-38)].

4 Geochemistry of selenium and bioavailability

Worldwide the occurrence of selenium is irregular. Selenium is widely distributed in the environment, but its natural geogenic concentration level is generally below 1 µg/1 in natural waters or below 1000 µg/kg in rocks and soils provided that anthropogenic influences are only moderate [[56](#page-17-2)]. Their distribution ranges from selenium defcit regions like the United Kingdom with a daily intake of<55 μg to selenium rich regions like Canada with a daily intake between 100 and 200 μg. Again, the intake amount is explained on the fact that selenium soil concentration is determined in many countries. Countries like Ireland, parts of the United State and India are in the toxic range while Yugoslav and Finland are deficient (Fig. [4](#page-5-1)).

Following its spatial variation, the daily selenium intake varies worldwide by country. For some regions of the world, selenium occurrence is scarce and raises issue of nutrient limitations rather than issue of toxicity. Accordingly, diferent countries attempted to trace its occurrence and dietary implication, whether its removal or fortifcation would be needed in their water supply scheme or not. Thus, selenium in Africa appears to be within limit, if not nutrient fortifcation is essential for most countries (Fig. [5\)](#page-6-0). **Fig. 3** Mass of selenium (in millions of kg) in various environmental pools worldwide. (Reprinted by permission from Springer-Verlag Berlin Heidelberg, Hydrogeol J [\[5\]](#page-15-1). © Springer-Verlag Berlin Heidelberg 2016)

Oceans

Interestingly, recent studies report the importance of selenium in reducing the risk of arsenicosis due to selenium mediation [\[57](#page-17-3), [58](#page-17-4)], arsenic hit regions may even need selenium supplement to tackle arsenicosis. Selenium is widely distributed in the environment with average occurrence of 0.09 ppm and its occurrence or distribution has been investigated in rocks, soils, minerals, groundwater and surface water [\[27\]](#page-16-12).

4.1 Selenium in rocks

High selenium concentrations are associated with some phosphatic rocks, organic-rich black shales, coals, and sulfide mineralization, whereas most other rock types contain very low concentrations. Selenium deficient environments are far more widespread than seleniumcontaining ones. However, it should be recalled that health outcomes are not only dependent on the total selenium content of rocks and soils but also on the amount of selenium taken up into plants and animals as bioavailable selenium from its cycle in the environment (Fig. [2\)](#page-4-0) [[32\]](#page-16-17).

4.2 Selenium occurrence and distribution in minerals

Selenium naturally occurs in a number of inorganic forms, namely selenide, selenate, and selenite, but these minerals are rare. Selenium rarely occurs in its elemental state or as pure ore compounds in the earth's crust. It is found in metal sulfde ores, where it partially replaces the sulfur. Likewise, the organic compounds of selenium and

Fig. 4 Outline of occurrence of selenium in diferent regions of the world as selenium-defcient, selenium-low and selenium-rich regions [[7](#page-15-3)]

tellurium are known since the late nineteenth century [\[8](#page-15-4)]. Selenium is found as a major constituent of 40 minerals and a minor component of 37 others, chiefy sulfdes [\[27](#page-16-12)].

Based on a reconnaissance survey conducted in South Africa by K Sami and AL Druzynski, selenium is not related to rock-forming silicates, and is most abundant in sulfde minerals, such as galena (commonly 0–15 ppm), arsenopyrite (42–57 ppm), pyrrhotite (5–63 ppm), marcasite (0–11 ppm), chalcopyrite (commonly between 10 and 50 ppm), pyrite (commonly between 0 and 50 ppm), and sphalerite (commonly between 10 and 50 ppm, can be as high as 900 ppm) [[60\]](#page-17-5).

Fig. 5 Dietary selenium availability and risk of inadequacy in Africa. (*Source*: [\[59\]](#page-17-8))

4.3 Selenium occurrence and distribution in soils

The occurrence of selenium in diferent soils is mainly a reflection of the underlying rock as well as practice of irrigation with selenium rich groundwater. The soluble selenite and selenate species in soil enter into plant metabolism; however, leaching and formation of insoluble complex by selenite accounted for losses into the environment like other insoluble selenium species. Most soils showed a selenium content ranging between 0.1 and 0.2 ppm while a maximum of 100 ppm is detected in the United States. However irregular concentrations have been measured in diferent regions of the world including China. In that regard sedimentary rocks are most reported contributor to soil selenium and to plants. In relation to that predicting selenium mobility in soils is proposed as researchable area in order to address either defciency or toxicity issues [[10,](#page-15-6) [27](#page-16-12), [51\]](#page-16-34). In fact, selenium is also present in air though it is detected to nano scale concentration especially in urban air [[43](#page-16-28)].

4.4 Selenium occurrence and distribution in groundwater and surface water

Based on reports, selenium content in natural water exists in the range of 0.1–400 µg/l and sometimes reaching up to 6000 µg/l. In such water bodies the predominant species are those biogeochemically relevant selenite and selenate from mobility and bioavailability points of views [[61](#page-17-6)].

Selenium poisoning may result from water systems whenever new agricultural runoff flow through a normally dry, undeveloped lands. This process leaches natural soluble selenium compounds (such as selenates) into the water, which may then be concentrated in emerging wetlands. Selenium pollution of waterways also occurs when selenium is leached from coal fue ash, mining and metal smelting, crude oil processing, and landfll.

Recently issues of selenium water contamination are associated with selenium biochemical risks. For instance, from the 210 ground water samples taken in Nigeria, selenium concentration of $7.33 \pm 6.22 - 46.3 \pm 22.4$ µg/L is reported following the needed assessment vis-à-vis risk of geogenic contamination which is almost greater than that of WHO's limit of 10 µg/l [\[35\]](#page-16-23) (Table [1\)](#page-3-0). The distribution of selenium was fairly uniform in the samples studied in diferent locations that suggested the infuence of geogenic factors. More than that, in some parts of the United States dark-gray shale sampled from 51 locations in Colorado, New Mexico, and Utah, many of the groundwater samples were found saline with selenium concentration of 1000 µg/l [[2\]](#page-15-9) which is 100 times beyond the limit recommended by the WHO.

Locally, selenium occurrence is rarely checked including the status in groundwater. There is a report noting that selenium content in Ethiopia is generally low, though a concentration of 15–7600 μ g/l [\[64](#page-17-7)] is reported for ground waters of the rift valley region of the country; and the median values being lower for spring and surface water

sources. Yet, this range of selenium concentration is over the limit value set by WHO. However, selenium health efect, either defciency or toxicity, is not known or unreported. Moreover, that same region is a concern for other geogenic water contamination in which fuoride is the most explored one [[65](#page-17-9)[–67\]](#page-17-10).

The selenium enrichment in groundwater may be caused by the contact with sedimentary formations. The selenium behavior as function of pH was similar in the ultrapure water that can determine its leaching behavior (Fig. [6\)](#page-7-0) [[36](#page-16-20), [68](#page-17-11)]. Thus, medium-range oxidation–reduction potential (ORP) and neutral pH are critical conditions for the toxic selenium to exist or to become dominant as species.

4.5 Reactivity and mobility conditions of selenium species

In the environment pH and reduction conditions play role in selenium species and mobility. Acidic and reducing conditions reduce inorganic selenites to elemental selenium, whereas alkaline and oxidizing conditions favor the formation of selenates as described earlier. Consequently, since selenites and selenates are soluble in water, selenium is leached from well-aerated alkaline soils that favors its oxidation. Conversely, elemental selenium and selenides are insoluble in water; therefore, selenium tends to be sorbed to wet and poorly aerated soils. Thus, reducing conditions of the environment favor those forms. As a result, selenium in alkaline soils is available for uptake by plants, whereas the availability of selenium in acidic soils tends to be limited by the adsorption of selenites and selenates to iron and aluminum oxide soils (Figs. [7,](#page-8-0) [8](#page-8-1)) [\[5,](#page-15-1) [69](#page-17-12)]. Further, temperature, moisture, the concentrations of water-soluble selenium, climate condition, organic matter content and microbial activity also determine selenium mobility.

As an anthropogenic activity, agriculture does not only increase the selenium content in soil but it can also increase selenium concentrations in surface water, as selenium is brought along with irrigation drainage water [[70](#page-17-13)]. In that regard, factors controlling selenium release into the groundwater system, soluble Na– and Mg–SO₄ salts represent the primary mobile reservoir of selenium in the Mancos Shale and associated alluvium, with only minor contributions from relatively insoluble gypsum. These salts are more prevalent in alluvium than Mancos Shale, and thus the alluvium represents a potential large source of readily mobilized selenium, which has been reported in other irrigated alluvial valleys particularly in the Western United States and China [[71\]](#page-17-14).

Generally, selenium species typically have greater reactivity towards oxidants and attract oxidants comparably greater than sulfur that is determined by their scavenging

SN Applied Sciences A SPRINGER NATURE journal

Fig. 6 Phase diagram of selenium speciation. (Reprinted with permission from Springer-Verlag Berlin Heidelberg, Hydrogeol J [[5](#page-15-1)]. © Springer-Verlag Berlin Heidelberg 2016)

efficient on oxidants including hypochlorous acid, especially in living systems. Such reactivity of selenium brings implications of pros and cons in living system [[72](#page-17-15), [73](#page-17-16)]. Besides, its acid forms in the activation of $π$ -bonds in diynyl esters giving to conjugate structures are recently reported [[74](#page-17-17), [75\]](#page-17-18).

5 The selenium biochemistry

In the environment whereby inorganic selenium is transformed into volatile forms including $(CH_3)_2$ Se by the microbial action of fungi and plants, organic selenium is found in biotic systems [[61\]](#page-17-6). Selenium metabolism is found in all domains of life, such as in bacteria, *archaea*, and eukarya and viruses. The chalcogen, selenium, plays an important role in animals, plants and microbes and their enzymes [[76](#page-17-19)] especially after the major finding that uracil guanine and adenosine (UGA) termination codon is used to direct the covalent insertion of selenocysteine. Selenocysteine is viewed as the 21st amino acid by many scientists in terms of ribosome-mediated protein synthesis-into certain selenium-dependent enzymes [\[77](#page-17-20), [78\]](#page-17-21). Selenium is incorporated into amino acids, such as cysteine, and then into selenium-containing proteins. Three types of redox enzymes that contain selenium in the form of **Fig. 7** Selenium species and transformation pathways between the species in **a** soilgroundwater-shale system; dissolution, organic incorporation, volatilization, precipitation, and **b** the succession of terminal e-accepting processes in a soil-groundwater system, showing the chemical reduction of $SeO₄$ to $SeO₃$ only after $O₂$ and NO₃ have been reduced [\[5\]](#page-15-1)

and role of organisms

selenocysteine have been isolated so far and they are characterized from bacterial sources, whereas only one, glutathione peroxidase, is known in mammals [\[79](#page-17-22)].

In nature the selenium cycle (Fig. 8) is partially maintained by selenium reducing bacteria (SeRB). The role of selenium cycling is played partly by the microorganisms that use selenate and selenite as terminal electron acceptors whose selenium biomineralization mechanisms gave ways to selenium environmental biotechno-logical applications [[80\]](#page-17-23).

In bacteria and *archaea*, selenium is readily metabolized and involved in a range of metabolic functions that include assimilation, methylation, detoxification, and anaerobic respiration. In that regard, anaerobic digesters treating organic waste and operating at high ammonia concentrations are operated stably with selenium supplement at 0.16 mg selenium per kg of fresh organic matter feed [[81](#page-17-24)[–83](#page-17-25)].

In a related fact, after ingestion, selenium is absorbed by the human body mainly in the duodenum and caecum via active transport through a sodium pump. The mechanisms of intestinal absorption of selenium differ depending on the chemical form of the element. When it is in excess, it is excreted through the urine and feces. Its

homeostasis is based on reserves of selenomethionine in the kidney and liver [[76,](#page-17-19) [84](#page-17-26)[–87\]](#page-17-27). Humans can get selenium from food items such as Brazil nuts, kidney, liver wheat (grown in selenium rich countries) and drinking water [[88\]](#page-17-28).

Regarding the biochemistry of selenium, whether it is essential for higher plants metabolism or not, is debatable though most argue it is not $[19, 21]$ $[19, 21]$ $[19, 21]$ $[19, 21]$. However, there is a possible incorporation of trace amount of the element into specifc selenoproteins for which the "S-assimilation" pathway is proposed where selenium is metabolized by the enzymes of sulphate assimilation both in accumulators and non-accumulators. Further, the accumulation selenium varies within the plant where shoot and root are most accumulators [[21\]](#page-16-8). Whether selenium benefts higher plants metabolism or not, a more concern as well as an opportunity is the issue of phytotoxicity and its implication to human. On one hand, hyper-accumulators do beneft through phytoremediation of selenium contaminated sites, on the other hand consumption of selenium pounded plants could result in human selenium toxicity while the less-accumulators would open the potential to human nutrient supplement [\[89\]](#page-17-29).

6 Selenium quantifcation in an environmental sample

Several instrumental and analytical methods including microwave digestion and fluorescence detection are applied to determine selenium in environmental samples in the recent past [\[61,](#page-17-6) [88](#page-17-28)]. Due to the narrow margin between the essential selenium and selenium toxicity, the detection sensor material, electrode surface modifers and chelators that overcome various challenges and achieve better detection limit, range, reproducibility, stability, selectivity, sensitivity, and so forth got advanced as reported by Devi et al., and Paikaray. Further the type of sample matters on the choice of a particular or group of detection methods as well as whether quantifcation or speciation is sought. In that regard a recipe depicting various environmental samples versus quantifcation and speciation is given in Fig. [9](#page-9-0) [\[1](#page-15-0), [17,](#page-16-4) [61](#page-17-6)].

Generally, selenium detection techniques can apply either the destructive method that includes the inductively coupled plasma mass spectroscopy (ICP-MS), a hydride generation atomic fluorescence spectrometer (HG-AFS) or the hydride generation atomic absorption spectroscopy (HGAAS). The non-destructive methods such as the Instrumental Neutron Activation Analysis (INAA) are also gaining attention. If the desire is routine detection but a simultaneous detection of multielement per sample, then the use of ICP-MS is advised [\[1,](#page-15-0) [90](#page-17-30)]. The INNA can, especially be applied to low detection (10−8–10−9 g) multielemental analysis in an air sample though better detection limits and selenium speciation in an environmental sample can be achieved using HGAAS $(0.1 \mu q)$ $(0.1 \mu q)$ $(0.1 \mu q)$ [1, [91–](#page-17-31)[94](#page-18-0)]. In spite of the advances in detection of selenium in an air sample, obtaining composite air sample is complicated and feld-deployable air sampling method has been challenging until a simultaneous preconcentrating analysis method is forwarded [[95](#page-18-1)].

However, conventionally, gas chromatography (GC), high-performance liquid chromatography (HPLC), ICP-MS, neutron activation analysis (NAA), atomic fuorescence spectrophotometry (AFS), AAS, electrothermal AAS, and graphite furnace AAS (GF-AAS) are widely applied. Moreover, for improved detection and speciation, hyphenated separation methods including Ion chromatography (IC) and capillary electrophoresis (CE) are used in conjugation with various atomic spectrophotometry detection systems such as Electrothermal AAS and plasmic optical emission spectroscopy (POES) [\[5](#page-15-1)].

Fig. 9 Selenium detection from an environmental sample for speciation and quantifcation. (Redrawn from [[1](#page-15-0)] with permission from John Wiley and Sons. © 2016 WILEY–VCH Verlag GmbH & Co. KGaA, Weinheim)

SN Applied Sciences A SPRINGER NATURE journal

For trace analysis of toxic elements, it is not only necessary to determine the total content of the element in question at lower and lower levels, but it is also necessary to elucidate the binding state in order to evaluate the ecotoxicity and to follow the pathways of these elements in the environment and their metabolism in biological systems. Speciation of toxic as well as the essential elements are therefore important through the detection of forms of selenium.

Therefore, as applied to unravel selenium sorption mechanisms that rely on an interaction of some source of X-ray excitation and a sample, diverse advanced detection techniques are emerging which include energy-dispersive X-ray spectroscopy (EDXRS), Fourier-transform infrared spectroscopy (FT-IR), powered X-ray difraction (PXRD), extended X-Ray absorption fne structure (EXAFS) and X-ray absorption spectroscopy (XAS) [\[96\]](#page-18-2).

7 Management of the excess selenium in water and soil media

Diverse technologies are being applied to remove selenium from water, soil, sediment and sludge [[97](#page-18-3)]. These technologies are applied at laboratory, pilot and full-scale levels [[46,](#page-16-31) [98](#page-18-4)]. By their working principle, these technologies encompass physico-chemical and biological methods. For instance, the full-scale application systems in selenium removal include fxed bed biological flters, constructed wetlands and reverse osmosis. Table [2](#page-10-0) below gives a glimpse of the classes of selenium remediation technologies with the corresponding examples that are partly detailed in Table [3](#page-11-0) including the advantage and disadvantage as well as process description.

7.1 Conventional selenium management techniques

Numerous physicochemical methods have been used to reduce the level of selenium from drinking water as well as to meet discharge limits by the process industries. For instance, the order of 1–5 µg/l is sought in the United States and other selenium rich nations. Conventionally

the following techniques are applied to meet standards in drinking water supply.

- 1. Biological reduction following pretreatment.
- 2. Ion exchange,
- 3. Activated alumina (AA),
- 4. Reverse osmosis (RO), and
- 5. Distillation.

Ion (anion) exchange can reduce selenium by up to 95%, in which the selenate ion is strongly preferred. Although Se (IV) is more difficult to oxidize compared to As (III) which is oxidized to As (V), this can readily be accomplished with the application of excess chlorine. The optimum amount of oxidation is obtained between pH 6.5 and 8.0 where Se (IV) can be converted to Se (VI) within fve minutes at a free chlorine concentration of 2 mg/l. At pH 9.0, only 15% of the Se (IV) has been found to be converted with 2 mg/l of free chlorine. Other techniques used for selenium removal include distillation (>98% reduction), reverse osmosis (RO) (>90% reduction) and activated alumina (85–95% reduction) [[87](#page-17-27), [99,](#page-18-5) [100](#page-18-6)] (Table [3](#page-11-0)).

Removal of the excess selenium from drinking water entered to a conventional scale especially in selenium rich countries through the application of the biological reduction technique. In such regard, fxed-flm fuidized bed reactor (FF-FBR), commercially called "ABMet", has got wider application. Recently, developments on the application of this biotechnology to mineralize selenium as extracellular and intracellular precipitation of selenium biominerals have been reported using selenate respiring bacterium [\[101\]](#page-18-7).

Comparatively, despite its pretreatment and management cost, pressure driven separations including RO are also applied to full scale. Some removal techniques are at pilot scale and few are still on a rigorous study especially the electrocoagulation method as explained in Table [3.](#page-11-0)

7.2 Recent developments on selenium management research

Since selenium contamination in groundwater and surface water in numerous river basins worldwide has become a

Table 2 Brief summary of classes of selenium removal technologies with examples

critical issue in recent decades [\[46](#page-16-31)], researches in selenium environmental management are gaining wider attention. Among the various endeavors attempted, phytoremediation, sorption and use of characteristically identifed microbes are worth to mention.

Basically, phytoremediation is a biological approach to pollution control or management activities that apply selenium selective plants species as portrayed on Fig. [10](#page-13-0) [[102](#page-18-9), [103\]](#page-18-10). The mechanism by which plants take up, transport and transform selenium is nearly similar as illustrated in Fig. [11.](#page-13-1) Accordingly, a selenium concentration of 1 g/ kg DW of plants is reported from phytoremediation of contaminated site using hyper accumulator plant species (Fig. [10\)](#page-13-0). In that regard, the hyper accumulators including giant reed to clean selenium-contaminated sites are also conveyed in recent publications [[7,](#page-15-3) [55,](#page-16-38) [76](#page-17-19)]. In a related study water hyacinth (*Eichhornia crassipes*) were tested for their pollutant metal sequestering potential [[104](#page-18-11)] that can also potentially be considered for selenium removal [[105](#page-18-12)]. However, the later management of the biomass will be a challenge for future research.

Organo-selenium, selenite and selenate in the soil are taken up through the plants' physiological system involving the root transport system and are either incorporated in selenium-protein, accumulated in their leaf or dissipated into the air after methylation (Fig. [11\)](#page-13-1) [[95](#page-18-1)].

Sorption of selenium is another attractive removal technology which is proved efficient in removing selenium from an environmental media and yet it is an older technique. For instance, sorption of selenium using minerals like hydrated lime and calcium-based particles proved to be efficient based on an XRD evaluation [[107,](#page-18-13) [108](#page-18-14)]. Adsorption of selenium is a phenomenon that co-occurs during a metabolic process [[10\]](#page-15-6).

Fig. 11 Selenium uptake by plants. (Adapted from [\[50\]](#page-16-39))

Hyper-accumulators >1000mg-Se/kg DW Secondary-accumulators 100-1000mg-Se/kg DW Non-accumulators <100mg-Se/kg DW Stanleya Brassica Grass

Fig. 10 Classifcation of plants depending upon selenium accumulation as hyperaccumulators, secondary accumulators and non-accumulators. (*Source*: [\[7\]](#page-15-3))

In a related current study, sorption of selenium is tested using rare earth hydroxide since the contents of selenium in industrial wastewater, agricultural runoff, and drinking water have to be constrained to a value of 50 μg/l, as the maximum concentration limit [[109](#page-18-15)]. This recent experimental research reported that selenium can be sequestered efficiently from its aqueous solution using a structurally well-defned cationic layered rare earth hydroxide, $Y_2(OH)_{5}Cl·1.5H_2O$ (Fig. [12](#page-14-0)) with maximum sorption capacities of selenite and selenate reported are 207 and 124 mg/g, respectively, both representing the new records among the inorganic sorbents. In the low concentration region, $Y_2(OH)_{5}Cl \cdot 1.5H_2O$ is able to almost completely remove selenium from aqueous solution even in the presence of competitive anions such as $NO₃⁻$, Cl⁻, CO₃²⁻, SO₄ ^{2–}, and HPO₄^{2–}. The resulting concentration 10 Å μg/l is achieved. The sorption kinetics is well ftted in the Langmuir model in two diferent pH levels, 7 and 8.5; however, the selenium sorption decreased at the higher pH (Fig. [13](#page-15-10)).

Observing the surface phenomena closely, the sorption mechanism was unraveled by the combination of advanced techniques like EDS, FT-IR, PXRD, and EXAFS. Selenate ions were exchanged with chloride ions in the interlayer space, forming outer-sphere complexes. In comparison, besides anion exchange mechanism, the selenite ions were directly bind to the Y^{3+} center in the positively charged layer of $[Y_2(OH)_5(H_2O)]$ ⁺ through strong bidentate binuclear inner-sphere complexation, consistent with the observation of the higher uptake of selenite over selenate [[109](#page-18-15)].

Waste sludge of domestic or industrial origin is becoming another selenium mine sites. Like other recovery operations, selenium is being obtained through sludge mining: *Pseudomonas stutzeri* NT- I to reduce the soluble selenium compounds to elemental selenium, Se(0) [[110](#page-18-16)] in another study. Michihiko Ike and colleagues at Osaka University, have developed a process to mineralize selenium through microbial recovery though the technique is obscured. This element is also present in industrial wastewater streams, for instance, from copper production [\[111](#page-18-17)]. However, much of the recovery efforts are on a laboratory scale which is mainly due to gaps in the method applied and the associated cost implications.

8 Conclusion

Selenium occurrence, geochemistry, nutrient and toxicity efect remained concerns worldwide. Advances in analytical methods and instrumentation, selenium detection and speciation helped in understanding selenium's environmental process in more detail. Selenium's natural distribution is globally uneven; for some it is scarce but for others, it needs removal from an environmental medium. In addition, the selenium intake by animals involves both food and water and hence the toxicity risk has to consider both factors. Furthermore, the selenium mobility associated risk is largely the selenium species specifc, environmental pH and redox potential dependent.

Regarding selenium management, reduction is the most feasible chemical technology to remove selenium. The biological methods are most established, especially in using phytoremediation and microbial reduction, the latter being largely applied in drinking water supply systems. Similarly, the application of surface phenomena to remove selenium is advancing with time. Thus, selenium sorption and isotherm are elucidated in recent works.

Yet, selenium remains to be a concern in geogenic and anthropogenic contamination as well as in nutrient supplement efect. The recovery of selenium from anthropogenic sources of release is also equally important. However, there is a huge geographical imbalance in selenium studies and publications, the least studies being in the African continent.

Research and development towards implementation and provision of an adequate institutional framework and marketing chains to increase the use of recovered selenium materials require feasibility studies. Furthermore, technologies for efficient recovery and reuse of recovered selenium with consideration of speciation, purity and

Fig. 12 Selenium adsorption in rare earth hydroxide. (Adapted with permission from [[109\]](#page-18-15). © 2017 American Chemical Society)

SN Applied Sciences A SPRINGER NATURE journal

 (b) ²⁵⁰ C(OH)_Cl-Se(VI) Y.(OH).Cl-Se(IV) q (mg Se/g sorbent) 200 150 100 50 pH 8.5 0 $\overline{0}$ 200 400 600 800 **Contact time (min)** (d) $Y_{2}(OH)_{2}Cl-Se(IV)-pH8.5$ 200 Langmuir model $Y_{2}(OH)_{2}Cl-Se(VI)$ -pH8.5 **Freundlich model** q (mg Se/g sorbent) 150 \triangleleft ¢ 100 50 pH 8.5 0 50 100 150 200 0 C_{eq} (mg/L)

Fig. 13 Sorption kinetics of Se (IV) and Se (VI) on $Y_2(OH)_{5}Cl·1.5H_2O$ at **a** pH 7 and **b** 8.5. Sorption isotherms of Se (IV) and Se (VI) on Y₂(OH)₅Cl-1.5H₂O at **c** pH 7 and **d** 8.5. (Phase ratio=2 g/l, contact

value are highly sought. In such regard, integrating selenium sequestered plants as source of selenium requires those advanced separation techniques. However, the extraction of the plant sequestered selenium is a future challenge that needs to be addressed.

Acknowledgements We are thankful to the African Center of Excellence for Water Management, Addis Ababa University, Ethiopia for the kind support it has made.

Compliance with ethical standards

Conflict of interest The authors declare that they have no confict of interest.

References

- 1. Paikaray S (2015) Origin, mobilization and distribution of selenium in a soil/water/air system: a global perspective with special reference to the indian scenario. CLEAN Soil Air Water 44(5):474–487
- 2. Morrison SJ, Goodknight CS, Tigar AD, Bush RP, Gil A (2012) Naturally occurring contamination in the Mancos Shale. Environ Sci Technol 46(3):1379–1387
- 3. Devitt DA, Wright LE, Shanahan SA, Hausrath E (2014) Fate of selenium in a small urban watershed. Environ Monit Assess 186(5):3181–3197

SN Applied Sciences

A SPRINGER NATURE journal

time=12 h, background=0.01 M NaCl. (Adapted with permission from [\[109](#page-18-15)]. © 2017 American Chemical Society)

- 4. Pettine M, McDonald TJ, Sohn M, Anquandah GAK, Zboril R, Sharma VK (2015) A critical review of selenium analysis in natural water samples. Trends Environ Anal Chem 5:1–7
- 5. Bailey RT (2017) Review: Selenium contamination, fate, and reactive transport in groundwater in relation to human health. Hydrogeol J 25(4):1191–1217
- 6. Grützmacher G, Kumar PS, Rustler M, Hannappel S, Sauer U (2013) Geogenic groundwater contamination—definition, occurrence and relevance for drinking water production. Zbl Geol Paläont Teil I 1:69–75
- 7. Gupta M, Gupta S (2017) An overview of selenium uptake, metabolism, and toxicity in plants. Front Plant Sci 7:1–14
- 8. Fernández-Lodeiro J, Pinatto-Botelho MF, Soares-Paulino AA, Gonçalves AC, Sousa BA, Princival C, Dos Santos AA (2014) Synthesis and biological properties of selenium- and telluriumcontaining dyes. Dyes Pigment 110:28–48
- 9. Gaskill C (2008) Veterinary toxicology—basic and clinical principles. Can Vet J 49(6):591
- 10. Lopes G, Ávila FW, Guilherme LRG (2017) Selenium behavior in the soil environment and its implication for human health. Ciência e Agrotec 41:605–615
- 11. Broadley MR, White PJ, Bryson RJ, Meacham MC, Bowen HC, Johnson SE, Hawkesford MJ, McGrath SP, Zhao F-J, Breward N, Harriman M, Tucker M (2007) Biofortifcation of UK food crops with selenium. Proc Nutr Soc 65(2):169–181
- 12. Levander OA, Burk RF (2006) Update of human dietary standards for selenium. In: Hatfeld DL, Berry MJ, Gladyshev VN (eds) Selenium: its molecular biology and role in human health. Springer, Boston, pp 399–410
- 13. Benemariya H, Robberecht H, Deelstra H (1993) Daily dietary intake of copper, zinc and selenium by diferent population groups in Burundi, Africa. Sci Total Environ 136(1):49–76
- 14. Kipp AP, Strohm D, Brigelius-Flohé R, Schomburg L, Bechthold A, Leschik-Bonnet E, Heseker H (2015) Revised reference values for selenium intake. J Trace Elem Med Biol 32:195–199
- 15. Gordon JEH (1875) Anomalous Behaviour of Selenium. Nature 12.187
- 16. Da Silva JF, Williams RJP (2001) The biological chemistry of the elements: the inorganic chemistry of life. Oxford University Press, New York
- 17. Devi P, Jain R, Thakur A, Kumar M, Labhsetwar NK, Nayak M, Kumar P (2017) A systematic review and meta-analysis of voltammetric and optical techniques for inorganic selenium determination in water. TrAC Trends Anal Chem 95:69–85
- 18. Gasiewicz TA, Smith JC (1978) Properties of the cadmium and selenium complex formed in rat plasma in vivo and in vitro. Chem Biol Interact 23(2):171–183
- 19. White PJ (2016) Selenium accumulation by plants. Ann Bot 117(2):217–235
- 20. Gupta M, Gupta S (2016) An overview of selenium uptake, metabolism, and toxicity in plants. Front Plant Sci 7:2074
- 21. Terry N, Zayed A, De Souza M, Tarun A (2000) Selenium in higher plants. Annu Rev Plant Biol 51(1):401–432
- 22. Brown TA, Shrift A (1982) Selenium: toxicity and tolerance in higher plants. Biol Rev 57(1):59–84
- 23. Garousi F (2017) Toxicity of selenium, application of selenium in fertilizers, selenium treatment of seeds, and selenium in edible parts of plants. Acta Univ Sapientiae Aliment 10(1):61–74
- 24. Sun G-X, Liu X, Williams PN, Zhu Y-G (2010) Distribution and translocation of selenium from soil to grain and its speciation in paddy rice (*Oryza sativa* L.). Environ Sci Technol 44(17):6706–6711
- 25. Kaur N, Sharma S, Kaur S, Nayyar H (2014) Selenium in agriculture: a nutrient or contaminant for crops? Arch Agron Soil Sci 60(12):1593–1624
- 26. Sharadamma K, Purushotham B, Radhakrishna P, Abhilekha P, Vagdevi H (2011) Role of selenium in pets health and nutrition: a review. Asian J Animal Sci 5(1):64–70
- 27. Council NR (1983) Selenium in nutrition: revised edition. National Academies Press, Washington, DC
- 28. Desta B, Maldonado G, Reid H, Puschner B, Maxwell J, Agasan A, Humphreys L, Holt T (2011) Acute selenium toxicosis in polo ponies. J Vet Diagn Invest 23(3):623–628
- 29. Olson OE (1986) Selenium toxicity in animals with emphasis on man. J Am Coll Toxicol 5(1):45–70
- 30. Kurek E, Ruszczyńska A, Wojciechowski M, Czauderna M, Bulska E (2009) Study on speciation of selenium in animal tissues using high performance liquid chromatography with on-line detection by inductively coupled plasma mass spectrometry. Chemia Analityczna 54:43–57
- 31. Benemariya H, Robberecht H, Deelstra H (1993) Zinc, copper, and selenium in milk and organs of cow and goat from Burundi, Africa. Sci Total Environ 128(1):83–98
- 32. Fordyce FM (2013) Selenium defciency and toxicity in the environment. In: Selinus O (ed) Essentials of medical geology: revised edition. Springer, Dordrecht, pp 375–416
- 33. Lemly AD (2004) Aquatic selenium pollution is a global environmental safety issue. Ecotoxicol Environ Saf 59(1):44–56
- 34. Lemly AD (1997) Environmental implications of excessive selenium: a review. Biomed Environ Sci 21:415–435
- 35. Etim EU (2017) Occurrence and distribution of arsenic, antimony and selenium in shallow groundwater systems of Ibadan Metropolis, Southwestern Nigerian. J Health Pollut 7(13):32–41
- 36. Bassil J, Naveau A, Bueno M, Razack M, Kazpard V (2018) Leaching behavior of selenium from the karst infillings of

the hydrogeological experimental site of Poitiers. Chem Geol 483:141–150

- 37. Tan LC, Nancharaiah YV, van Hullebusch ED, Lens PNL (2016) Selenium: environmental significance, pollution, and biological treatment technologies. Biotechnol Adv 34(5):886–907
- 38. Barceloux DG, Barceloux D (1999) Selenium. J Toxicol Clin Toxicol 37(2):145–172
- 39. Vinceti M, Wei ET, Malagoli C, Bergomi M, Vivoli G (2001) Adverse health effects of selenium in humans. Rev Environ Health 16(4):233–251
- 40. Vinceti M, Mandrioli J, Borella P, Michalke B, Tsatsakis A, Finkelstein Y (2014) Selenium neurotoxicity in humans: bridging laboratory and epidemiologic studies. Toxicol Lett 230(2):295–303
- 41. MacFarquhar JK, Broussard DL, Melstrom P, Hutchinson R, Wolkin A, Martin C, Burk RF, Dunn JR, Green AL, Hammond R, Schaffner W, Jones TF (2010) Acute selenium toxicity associated with a dietary supplement. Arch Intern Med 170(3):256–261
- 42. Aldosary BM, Sutter ME, Schwartz M, Morgan BW (2012) Case series of selenium toxicity from a nutritional supplement. Clin Toxicol 50(1):57–64
- 43. Fan AM, Kizer KW (1990) Selenium. Nutritional, toxicologic, and clinical aspects. West J Med 153(2):160–167
- 44. Kubota S, Tsuchiya Y (2009) Water quality and standards. Eolss Publishers, Oxford
- 45. Fan AM, Vinceti M (2015) Selenium and its compounds, in Hamilton & Hardy's industrial toxicology. Wiley, Hoboken
- 46. Bailey RT, Romero EC, Gates TK (2015) Assessing best management practices for remediation of selenium loading in groundwater to streams in an irrigated region. J Hydrol 521:341–359
- 47. Santos S, Ungureanu G, Boaventura R, Botelho C (2015) Selenium contaminated waters: an overview of analytical methods, treatment options and recent advances in sorption methods. Sci Total Environ 521–522:246–260
- 48. Bailey RT, Gates TK, Halvorson AD (2013) Simulating variablysaturated reactive transport of selenium and nitrogen in agricultural groundwater systems. J Contam Hydrol 149:27–45
- 49. Martens DA, Suarez DL (1997) Selenium speciation of soil/ sediment determined with sequential extractions and hydride generation atomic absorption spectrophotometry. Environ Sci Technol 31(1):133–139
- 50. Winkel LHE, Vriens B, Jones GD, Schneider LS, Pilon-Smits E, Bañuelos GS (2015) Selenium cycling across soil-plant-atmosphere interfaces: a critical review. Nutrients 7(6):4199–4239
- 51. Xing K, Zhou S, Wu X, Zhu Y, Kong J, Shao T, Tao X (2015) Concentrations and characteristics of selenium in soil samples from Dashan Region, a selenium-enriched area in China. Soil Sci Plant Nutr 61(6):889–897
- 52. Mast MA, Mills TJ, Paschke SS, Keith G, Linard JI (2014) Mobilization of selenium from the Mancos Shale and associated soils in the lower Uncompahgre River Basin, Colorado. Appl Geochem 48:16–27
- 53. Fellowes JW, Pattrick RAD, Boothman C, Al Lawati WMM, van Dongen BE, Charnock JM, Lloyd JR, Pearce CI (2013) Microbial selenium transformations in seleniferous soils. Eur J Soil Sci 64(5):629–638
- 54. Bassil J, Naveau A, Bueno M, Di Tullo P, Grasset L, Kazpard V, Razack M (2016) Determination of the distribution and speciation of selenium in an argillaceous sample using chemical extractions and post-extractions analyses: application to the hydrogeological experimental site of Poitiers. Environ Sci Pollut Res 23(10):9598–9613
- 55. El-Ramady H, Abdalla N, Alshaal T, El-Henawy A, Faizy S, Belal E, Shehata S, Amer M, Fa´ri MS, Sztrik A, Schnug E, Pilon-Smits Samia EAH, El-Marsafawy M, Domokos-szabolcsy E (2015) Giant reed for selenium phytoremediation under changing climate. Environ Chem Lett 13:359–380
- 56. Jakubowski N, Thomas C, Stuewer D, Dettlaff I, Schram J (1996) Speciation of inorganic selenium by inductively coupled plasma mass spectrometry with hydraulic high pressure nebulization. J Anal At Spectrom 11(11):1023–1029
- 57. Ponomarenko O, La Porte PF, Singh SP, Langan G, Fleming DEB, Spallholz JE, Alauddin M, Ahsan H, Ahmed S, Gailer J, George GN, Pickering IJ (2017) Selenium-mediated arsenic excretion in mammals: a synchrotron-based study of whole-body distribution and tissue-specifc chemistry. Met Integr Biometal Sci 9(11):1585–1595
- 58. Willett WC, MacMahon B (1984) Diet and cancer—an overview. N Engl J Med 310(11):697–703
- 59. Hurst R, Siyame EWP, Young SD, Chilimba ADC, Joy EJM, Black CR, Ander EL, Watts MJ, Chilima B, Gondwe J, Kang'ombe D, Stein AJ, Fairweather-Tait SJ, Gibson RS, Kalimbira AA, Broadley MR (2013) Soil-type infuences human selenium status and underlies widespread selenium defciency risks in Malawi. Sci Rep 3:1425
- 60. Sami K, Druzynski A (2013) South African Water Research Council [Online]. [http://www.wrc.org.za/Pages/DisplayIte](http://www.wrc.org.za/Pages/DisplayItem.aspx?ItemID=7713&FromURL=%2FPages%2FAllKH.aspx%3F) [m.aspx?ItemID=7713&FromURL=%2FPages%2FAllKH.aspx%3F](http://www.wrc.org.za/Pages/DisplayItem.aspx?ItemID=7713&FromURL=%2FPages%2FAllKH.aspx%3F). Accessed Sept 2018
- 61. Pyrzyńska K (2002) Determination of selenium species in environmental samples. Microchim Acta 140(1):55–62
- 62. Murray MP, Sharmin R (2015) Groundwater arsenic and education attainment in Bangladesh. J Health Popul Nutr 33:20
- 63. IS10500 B (2012) Indian standard drinking water–specifcation (second revision). Bureau of Indian Standards (BIS), New Delhi
- 64. Kebede S (2012) Groundwater in Ethiopia: features, numbers and opportunities. Springer, Berlin, pp 83–92
- 65. Dagnaw LA, Chandravanshi B, Zewge F, Ababa A (2018) Fluoride content of leafy vegetables, irrigation water, and farmland soil in the rift valley and in non-rift valley areas of Ethiopia. Fluoride 50:409–429
- 66. Wendimu G, Zewge F, Mulugeta E (2017) Aluminium-ironamended activated bamboo charcoal (AIAABC) for fluoride removal from aqueous solutions. J Water Process Eng 16:123–131
- 67. Zewge F (2017) Combined aluminium sulfate/hydroxide process for fluoride removal from drinking water. Bull Chem Soc Ethiop 30:391
- 68. Floor GH, Iglesías M, Román-Ross G, Corvini PFX, Lenz M (2011) Selenium speciation in acidic environmental samples: application to acid rain–soil interaction at Mount Etna volcano. Chemosphere 84(11):1664–1670
- 69. McNeal JM, Balistrieri LS (1989) Geochemistry and occurrence of selenium: an overview. In: Selenium in agriculture and the environment (seleniuminagric), pp 1–13
- 70. Violante A, Cozzolino V, Perelomov L, Caporale A, Pigna M (2010) Mobility and bioavailability of heavy metals and metalloids in soil environments. J Soil Sci Plant Nutr 10(3):268–292
- 71. Mills TJ, Mast MA, Thomas J, Keith G (2016) Controls on selenium distribution and mobilization in an irrigated shallow groundwater system underlain by Mancos Shale, Uncompahgre River Basin, Colorado, USA. Sci Total Environ 566–567:1621–1631
- 72. Tsui EY, Hartstein KH, Gamelin DR (2016) Selenium Redox reactivity on colloidal CdSe quantum dot surfaces. J Am Chem Soc 138(35):11105–11108
- 73. Carroll L, Pattison DI, Fu S, Schiesser CH, Davies MJ, Hawkins CL (2015) Reactivity of selenium-containing compounds with myeloperoxidase-derived chlorinating oxidants: second-order rate constants and implications for biological damage. Free Radic Biol Med 84:279–288
- 74. Wilkins Lewis C, Günther Benjamin AR, Walther M, Lawson James R, Wirth T, Melen Rebecca L (2016) Contrasting frustrated Lewis pair reactivity with selenium- and boron-based Lewis acids. Angewandte Chemie Int Ed 55(37):11292–11295
- 75. Freudendahl DM, Wirth T (2011) New selenium electrophiles and their reactivity. In: Woollins JD, Laitinen RS (eds) Selenium and tellurium chemistry. Springer, Berlin, pp 41–55
- 76. Stadtman TC (1974) Selenium biochemistry. Science 183(4128):915
- 77. Stadtman TC (1990) Selenium biochemistry. Annu Rev Biochem 59(1):111–127
- 78. Reddy CC, Massaro EJ (1983) Biochemistry of selenium: a brief overview. Fundam Appl Toxicol 3(5):431–436
- 79. Stolz JF, Basu P, Santini JM, Oremland RS (2006) Arsenic and selenium in microbial metabolism. Annu Rev Microbiol 60(1):107–130
- 80. Nancharaiah YV, Lens PNL (2015) Selenium biomineralization for biotechnological applications. Trends Biotechnol 33(6):323–330
- 81. Nancharaiah YV, Lens PNL (2015) Ecology and biotechnology of selenium-respiring bacteria. Microbiol Mol Biol Rev MMBR 79(1):61–80
- 82. Tang K, Baskaran V, Nemati M (2009) Bacteria of the sulphur cycle: an overview of microbiology, biokinetics and their role in petroleum and mining industries. Biochem Eng J 44(1):73–94
- 83. Bailey RT, Gates TK, Ahmadi M (2014) Simulating reactive transport of selenium coupled with nitrogen in a regional-scale irrigated groundwater system. J Hydrol 515:29–46
- 84. Jager T, Drexler H, Goen T (2016) Human metabolism and renal excretion of selenium compounds after oral ingestion of sodium selenite and selenized yeast dependent on the trimethylselenium ion (TMSe) status. Arch Toxicol 90(5):1069–1080
- 85. Bügel S, Larsen EH, Sloth JJ, Flytlie K, Overvad K, Steenberg LC, Moesgaard S (2008) Absorption, excretion, and retention of selenium from a high selenium yeast in men with a high intake of selenium. Food Nutr Res 52:1–8. [https://doi.org/10.3402/fnr.](https://doi.org/10.3402/fnr.v52i0.1642) [v52i0.1642](https://doi.org/10.3402/fnr.v52i0.1642)
- 86. Mehdi Y, Hornick J-L, Istasse L, Dufrasne I (2013) Selenium in the environment, metabolism and involvement in body functions. Molecules 18(3):3292–3311
- 87. Higashi RM, Cassel TA, Skorupa JP, Fan TW-M (2005) Remediation and bioremediation of selenium-contaminated waters. Water Encycl 2:355–360
- 88. Nahir TM, Sheffield M-C (2002) Analysis of selenium in Brazil Nuts by microwave digestion and fuorescence detection. J Chem Educ 79(11):1345
- 89. Natasha M, Shahid Niazi NK, Khalid S, Murtaza B, Bibi I, Rashid MI (2018) A critical review of selenium biogeochemical behavior in soil-plant system with an inference to human health. Environ Pollut 234:915–934
- 90. Ballihaut G, Pécheyran C, Mounicou S, Preud'homme H, Grimaud R, Lobinski R (2007) Multimode detection (LA-ICP-MS, MALDI-MS and nanoHPLC-ESI-MS2) in 1D and 2D gel electrophoresis for selenium-containing proteins. TrAC Trends Anal Chem 26(3):183–190
- 91. Uden PC (2002) Modern trends in the speciation of selenium by hyphenated techniques. Anal Bioanal Chem 373(6):422–431
- 92. Buchberger W (2001) Detection techniques in ion chromatography of inorganic ions. TrAC Trends Anal Chem 20(6–7):296–303
- 93. Vassileva E, Becker A, Broekaert J (2001) Determination of arsenic and selenium species in groundwater and soil extracts by ion chromatography coupled to inductively coupled plasma mass spectrometry. Anal Chim Acta 441(1):135–146

SN Applied Sciences

A SPRINGER NATURE journal

- 94. Mazej D, Falnoga I, Veber M, Stibilj V (2006) Determination of selenium species in plant leaves by HPLC-UV-HG-AFS. Talanta 68(3):558–568
- 95. Vriens B, Ammann AA, Hagendorfer H, Lenz M, Berg M, Winkel LH (2014) Quantifcation of methylated selenium, sulfur, and arsenic in the environment. PLoS ONE 9(7):e102906
- 96. Zhang C (2007) Fundamentals of environmental sampling and analysis. Wiley, New York, pp 172–241
- 97. van Hullebusch ED (2018) Remediation of selenium contaminated wastewater. In: Kallel A, Ksibi M, Ben Dhia H, Khélif N (eds) Recent advances in environmental science from the Euro-Mediterranean and surrounding regions. EMCEI 2017. Advances in science, technology & innovation (IEREK Interdisciplinary Series for Sustainable Development). Springer, Cham
- 98. Zhang Y, Frankenberger WT (2003) Factors afecting removal of selenate in agricultural drainage water utilizing rice straw. Sci Total Environ 305(1):207–216
- 99. Remediation and Bioremediation of Selenium‐Contaminated Waters, in Water Encyclopedia
- 100. Mahmoud MR, Soliman MA, Ali AH, Othman SH (2016) Kinetic studies on radio-selenium uptake by ion exchange resin. Sep Sci Technol 51(6):976–989
- 101. Butler CS, Debieux CM, Dridge EJ, Splatt P, Wright M (2012) Biomineralization of selenium by the selenate-respiring bacterium *Thauera selenatis*. Biochem Soc Trans 40(6):1239
- 102. Ansari AA, Gill SS, Gill R, Lanza GR, Newman L (2016) Phytoremediation. Springer, Berlin
- 103. Carvalho KM, Gallardo MT, McGettigan MJ, Martin DF (2000) Remediation of selenium contamination by plants and microbes: an annotated bibliography. Fla Sci 63:133–141
- 104. Newete SW, Erasmus BF, Weiersbye IM, Byrne MJ (2016) Sequestration of precious and pollutant metals in biomass of cultured water hyacinth (*Eichhornia crassipes*). Environ Sci Pollut Res Int 23(20):20805–20818
- 105. Banuelos G, Lin Z, Wu L, Terry N (2002) Phytoremediation of selenium-contaminated soils and waters: fundamentals and future prospects. Rev Environ Health 17(4):291–306
- 106. Hill CM (2010) Review of available technologies for the removal of selenium from water. Final Report, prepared for North American Metals Council (NAMC)
- 107. Ghosh-Dastidar A, Mahuli S, Agnihotri R, Fan LS (1996) Selenium capture using sorbent powders: mechanism of sorption by hydrated lime. Environ Sci Technol 30(2):447–452
- 108. Agnihotri R, Chauk S, Mahuli S, Fan LS (1998) Selenium removal using ca-based sorbents: reaction kinetics. Environ Sci Technol 32(12):1841–1846
- 109. Zhu L, Zhang L, Li J, Zhang D, Chen L, Sheng D, Yang S, Xiao C, Wang J, Chai Z (2017) Selenium sequestration in a cationic layered rare earth hydroxide: a combined batch experiments and EXAFS investigation. Environ Sci Technol 51(15):8606–8615
- 110. Kuroda M, Notaguchi E, Sato A, Yoshioka M, Hasegawa A, Kagami T, Narita T, Yamashita M, Sei K, Soda S, Ike M (2011) Characterization of *Pseudomonas stutzeri* NT-I capable of removing soluble selenium from the aqueous phase under aerobic conditions. J Biosci Bioeng 112(3):259–264
- 111. Kagami T, Narita T, Kuroda M, Notaguchi E, Yamashita M, Sei K, Soda S, Ike M (2013) Efective selenium volatilization under aerobic conditions and recovery from the aqueous phase by *Pseudomonas stutzeri* NT-I. Water Res 47(3):1361–1368