



Overlooked Adsorptive Route and Challenges in Arsenic Decontamination Using Iron Oxide Nanomaterials

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Abstract

Heavy metal such as arsenic in water has gained major global attention because of their carcinogenicity to a living being in the environment. By different routes, arsenic enters the water environment and decreases the water quality even at a very lower concentration (parts of billion (ppb)). Several approaches have been studied for arsenic abatement, but the adsorption process is efficient and economical to decontaminate water from arsenic. Iron oxide nanoparticles (IONPs) can act as low-cost adsorbents because of their magnetic properties, abundance, biocompatibility, and high selectivity towards arsenic. Further to increase the adsorption capacity of IONPs, a sustainable approach has been taken, and decorated with several materials such as activated carbon, biochar, chitosan, cellulose, and mining waste materials (which are environmentally friendly, low cost, number of functional groups, high reactivity, thermal stability, chemical stability and easily available everywhere). This review has outlined the overlooked adsorptive routes for arsenic removal with importance on arsenic sources, pollution, and removal strategies. A detailed influence parameter for arsenic removal by iron oxide and iron oxide-based composite and the interaction of arsenic species with the various functional groups of adsorbents were studied.

Keywords Iron oxide nanoparticles · Composite · Arsenic · Adsorption · Removal

1 Introduction

1.1 Water Pollution

Without water, the sustainability of life cannot be imagined. The level of both surface and groundwater is decreasing day by day and only 3% of usable water is present on the earth's surface apart from groundwater [1]. This warns that very less amount of freshwater is available on the earth's surface to fill up the necessity of living beings. A recent report from the World Health Organization (WHO) shows that about one billion people don't find usable drinking water and four

billion people will rigorously be affected by the shortage of drinking water by 2050 [2, 3]. Despite that, every second thousands of tons of toxic materials are coming to the water sources (pond, rivers, well, etc.) and creates problems for living beings [4, 5]. This water pollution or water quality degradation is mainly due to rapid population growth, urbanization, and industrial growth [6]. Every day tons of waste materials from different sources are coming to the natural water sources and pollute water severely as they contain pharmaceutical waste, dyes, toxic heavy metals, pesticides, organic solvents, and other pollutants [7–12].

Heavy metals are high-density elements and one of the prime toxic pollutants in water. These elements are coming to the water environments through different natural and anthropogenic activities which create problems in public health and ecology [13, 14]. However, the trace elements like iron (Fe), selenium (Se), chromium (Cr), cobalt (Co), magnesium (Mg), copper (Cu), nickel (Ni), zinc (Zn), manganese (Mn) and molybdenum (Mo) are the essential heavy metals (within their tolerance value) for the physiological and biochemical functioning of the living organisms [14, 15]. But heavy metals like lead (Pb) [16], cadmium (Cd) [17], mercury (Hg) [18], chromium (Cr) [19], and arsenic

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(As) [20] are very toxic and can damage the organ even at minute concentration. Also, these elements are categorized as ‘probable’ or ‘known’ carcinogens by different environmental agencies. Figure 1 shows the problems that occur due to these heavy metals. Arsenic has secured the first position in the toxic elements list and is very toxic as compared to others, so details of arsenic are studied in this review.

1.2 Arsenic

Arsenic is a toxic metalloid with a density of 5.75 g/cm^3 , an atomic weight of 74.92 amu, a boiling point of $613 \text{ }^\circ\text{C}$, and a melting point of $817 \text{ }^\circ\text{C}$ [21]. The main reasons for arsenic contamination in water are volcanic eruption, industrial chemical waste disposal, uses of arsenic in the coating, printing pigments, dyes, microbiological activities, rapid practices of arsenic fertilizers, and burning of fossil fuels [22, 23]. Around the world, several countries (India, Pakistan, Bangladesh, China, Taiwan, Vietnam, Japan, United States, Canada, New Zealand, Argentina, Poland, Mexico, and Hungary) have drinking water problems as the arsenic concentration is very much high (0.1–73 mg/L) in both ground and surface water [24, 25]. According to the World Health Organization (WHO) guidelines, the maximum contaminant limit (MCL) of arsenic in drinking water is 0.01 mg/L. Previously the arsenic MPL was 0.05 mg/L, but in January 2006, the United States environmental protection agency disclose new guidelines and reduce the standard of concentration from 0.05 to 0.01 mg/L [26]. Arsenic is present in both organic (monomethyl arsenic acid (MMA), dimethylarsinic acid (DMA), arsine derivatives, etc.) and

inorganic (arsenous and arsenic acid) forms in water [22]. Among these, organic arsenic is less toxic as it is soluble and less mobile as compared to inorganic arsenic. Again the inorganic arsenic is found in different oxidation states like +3, +5, 0, and -3 [27], but two major oxidation states of inorganic arsenic are As(III) (arsenite) and As(V) (arsenate) [28]. These arsenic species in the aquatic environment are extremely pH-dependent. Figure 2 shows the different speciation of As(III) and As(V) concerning pH and redox potential respectively [29]. Arsenite (As(III)) is the predominant species in anaerobic reducing environmental conditions and is present as hard arsenious acid (H_3AsO_3 , H_2AsO_3^- and HAsO_3^{2-}) while arsenate (As(V)) is the main species in oxidizing aerobic environmental conditions and occurs in soft arsenic acid (H_3AsO_4 , H_2AsO_4^- , HAsO_4^{2-} and AsO_4^{3-}) [27, 30]. As(III) is comparatively 20 times more toxic than As(V) as it reacts faster rate to the biological system [31]. As(III) is directly attached to the body protein through sulfhydryl groups and rapture the enzyme activities of the body, while As(V) can replace the phosphate groups in the mitochondria, which disturbs the oxidative phosphorylation cycles and affects the energy bond formation of adenosine triphosphate [22, 32, 33]. Hence, both oxidation states are toxic in different environmental conditions.

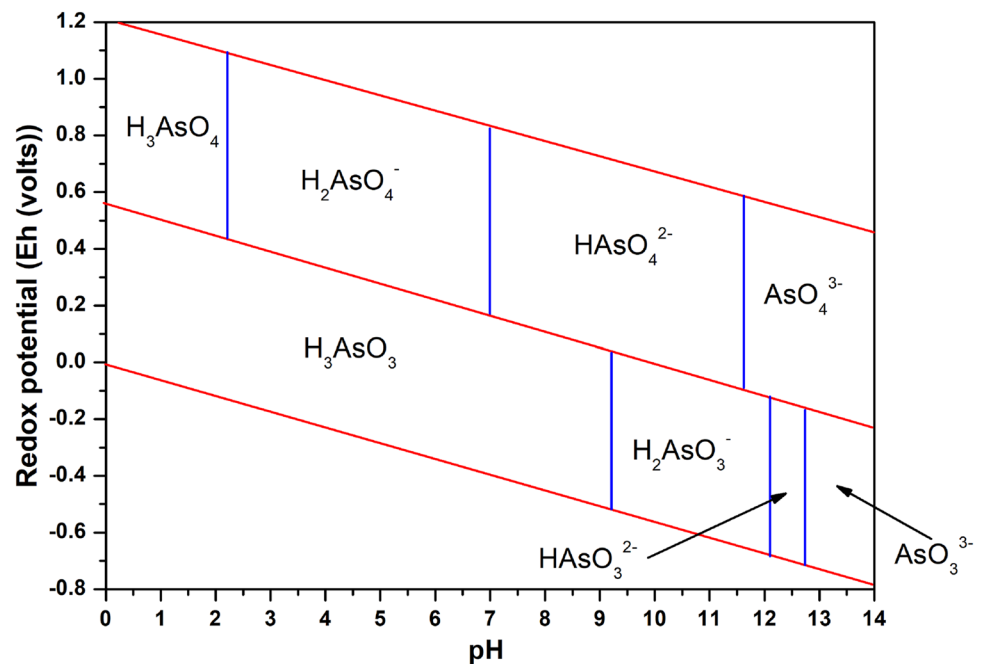
1.3 Sources of Arsenic

Both natural processes and man-made activities are liable for the existence of arsenic in a usable water environment. Naturally, arsenic is mainly present in 245 types of minerals including arsenopyrite (FeAsS), realgar (As_4S_4), orpiment (As_2S_3), and enargite (Cu_3AsS_4) which are the most common mineral found in nature [34]. The report indicates that erosion and leaching of these arsenic-containing minerals results in the release of arsenic into aquatic environments [35]. Another important thing is that about 0.5–2.4 $\mu\text{g/g}$ and 3–10 $\mu\text{g/g}$ of arsenic also present in rocks and sediments and the leaching of these materials brings arsenic to the water system [36]. Some other natural activities like volcanic erosion, geothermal water, soil media, seasonal flooding, and sedimentation are key processes for the presence of arsenic in water systems [37–43]. On the other side of view, the main sources of arsenic in water are the day-to-day activities which are done by human beings knowingly or unknowingly. Arsenic is rummage-sale used in the production of semiconductors, lasers, transistors, and solar cells, processing of ceramics, glass, paints, pigments, soaps and dyes, catalysts, metal adhesives, ammunition, and pyrotechnics [44, 45]. Hence, the waste coming from these industries brings arsenic to the water environment. Again arsenic compounds are also applied in the production of insecticides and pesticides, which transfer the arsenic to both vegetation and water system and ultimately pollute the environment. Lastly, wood



Fig. 1 Toxicity of heavy metals

Fig. 2 Arsenic speciation at different pH according to redox potential (Eh) [29]



preservation and burning of mining have brought arsenic to 60% of the total anthropogenic activities [36].

1.4 Arsenic Toxicity

Arsenic in usable water is the major source of contact with living beings and excess concentration of arsenic in drinking water is a major environmental concern, which can cause serious carcinogenic effects to mankind [46]. Among the top twenty hazardous elements, arsenic compounds occupy the number one position in this list as recognized by the Agency for toxic substances and disease registry (ATSDR), International Agency for Research on Cancer (IARC), and World Health Organization (WHO) [47, 48]. Arsenic exposure can occur through various routes, including ingestion, inhalation, and dermal contact. When it comes to skin exposure, arsenic can be absorbed through the skin and its ability to enter the bloodstream, could affect the vascular system, central nervous system, hematopoietic system, and cancer in the liver, skin, renal, bladder, lungs, and kidneys [22, 49]. Studies have indicated that chronic exposure to arsenic is associated with an increased risk of developing various types of cancers, such as lung, bladder, and liver cancer. Dermal exposure to arsenic can lead to skin-related health problems such as: (a) Skin Irritation: Arsenic can cause skin irritation, redness, and inflammation upon direct contact, (b) hyperpigmentation: Chronic exposure to arsenic can lead to hyperpigmentation, where patches of skin become darker in color. This is particularly evident in areas that are frequently exposed to arsenic-containing substances. (c) Keratosis: Arsenic exposure can result in the development of

skin lesions known as arsenical keratoses. These are rough, scaly patches that may appear on the palms, soles, and other parts of the body and (d) Skin Cancer. Excess concentration of arsenic also enhances the glutathione peroxidase and mitochondrial superoxide dismutase (MSOD) actions in the lungs and liver [30]. Other problems like anorexia, colitis, gastritis, vomiting, diarrhoea, hair loss, weight loss, muscle cramps, hallucination, and abdominal pain happen due to acute poisoning [22]. Similarly, arsenic shows its toxic effects towards the plant species which results in changes of physiological, morphological, and metabolic properties like decreases in chlorophyll content, short root length, membrane disruption, low photosynthetic and transpiration rate, lower relative water content of leaf, minimize the stomatal conductance, alter the sugar metabolism and production of toxic hydrogen peroxide (H_2O_2) and superoxide ions (O_2^-) in cell tissues of the plant [50, 51]. Due to arsenic poisoning, aquatic species like fish is highly affected and higher exposure of arsenic leads to alter the antioxidant enzyme activity, increase the oxidative stress, and modulation of antioxidant system of liver respectively [52].

1.5 Arsenic Removal Technologies

Several remediation technologies (membrane filtration, precipitation, ion exchange, and adsorption as shown in Fig. 3) have been applied to reduce the arsenic concentration in industrial and agricultural wastewater [53]. A detailed analysis of these arsenic removal technologies is presented in Table 1. The membrane filtration process for arsenic removal is an advanced technique and the output results are also very

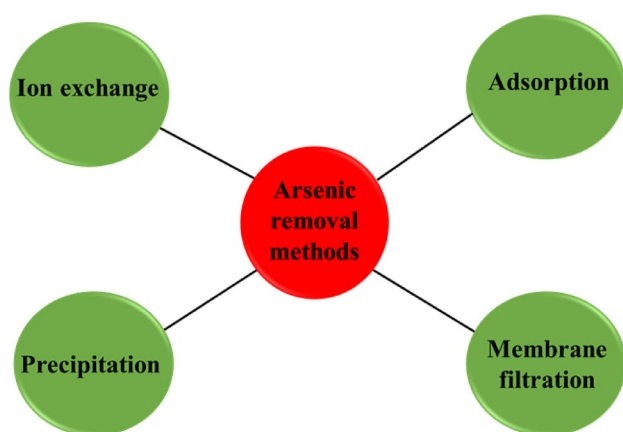


Fig. 3 Different arsenic removal methods

good [54, 55]. Different types of membranes are used concerning selective pollutants and are also used without any chemicals. But the main disadvantages of these techniques are high maintenance cost, high-pressure wastewater rejection, problems with regeneration and repeated use, and difficulty to remove As(III) [56]. Precipitation/flocculation is a different type of process for arsenic removal where aluminium, calcium, and iron salts are used to precipitate arsenic as aluminium arsenate, calcium arsenate, and ferric arsenate [53]. This process applies to the high concentration of arsenic (above 0.1 mg/L) and is difficult to remove low-concentration arsenic. Hence secondary treatments require for arsenic removal in every one cycle of experiment in the precipitation/flocculation process. Again difficult to remove As(III), hence oxidants are required to convert As(III) to As(V) [57]. One more physicochemical process is the ion-exchange process, arsenic is removed with some other ions in water [58]. Here, a column with different types of ion-exchange resin (cation-exchange resins, anion-exchange resin, and chelating resin) are inserted and arsenic water is passed through it, and an exchange of ions takes place. But this method highly depends on arsenic concentration,

pH, other anions and types of resin used [24]. Hence, most of these technologies are costly, has high water discharge with secondary sludge production, and can remove the high initial concentration of arsenic but fail for lower concentrations. Thus these processes are unable for real practical application in rural areas where it is not possible for high operational setup. In the adsorption process, the adhesion of arsenic takes place on the adsorbent surface. This process can be used for practical applications of arsenic removal as the process is effective in terms of low maintenance, highly efficient, and possible to recover arsenic from the adsorbent surface [24, 59]. Here, the adsorbent can be reused many times with successful regeneration [60]. In the adsorption process, the selectivity of adsorbent plays a crucial role and these days' nanomaterials have gained visible attraction in adsorption studies [61]. The high surface area of nanomaterial provides strength and active sites which will increase the removal rate. The nanoparticles commonly used as an adsorbent for arsenic removal are aluminium [62], cerium [63], magnesium [64], manganese [65], titanium [66], zinc [67], and zirconium [68]. Compared to other environmentally friendly iron oxide has more attraction towards arsenic with magnetic separation properties offering it a suitable adsorbent. But single iron oxide nanoparticles are agglomerated which decreases the efficiency for removal studies. Therefore, modifications are done with zeolite [69], mixed oxide [9], polymeric membrane [70], activated carbon [71], graphene oxide [72], etc. to increase the performance of iron oxides and a higher percentage of As(III) and As(V) are removed.

2 Iron Oxide-Based Adsorbents for Arsenic Removal

Iron oxide nanoparticles are frequently implemented in wastewater remediation. Iron oxide and iron oxide-based adsorbents are used for arsenic removal for their unique properties. Many iron oxide nanoparticle supporting

Table 1 Advantages and disadvantages of different arsenic removal techniques

Removal methods	Advantages	Disadvantages	References
Membrane filtration	Separated by size, no phase change, advanced technique, low energy required, good output	Membrane fouling effect, expensive, high flow rates, high maintenance cost	[54, 55]
Precipitation	Simple, good sludge setting, bacterial inactivation capacity, efficient for insoluble contaminants	Generate a large volume of sludge, no reusable chemical produced, pH-dependent, require additional treatment and low removal for arsenic	[53]
Ion exchange	Simple, easy control and maintenance, rapid, efficient technology	Selective, high cost, large volume requires, precipitation causes blocking of the reactor, pH-sensitive, and resin elimination	[24]
Adsorption	Use on a laboratory scale, low cost, high efficiency, used several times	Nano-sized adsorbents require host materials, regeneration could result in secondary pollution	[24, 59]

materials are used in several studies but some of them are very costly, toxic in nature, not biodegradable, technically not feasible, have complex synthesis processes, and are unstable at normal room temperature [39, 73]. In this review, iron oxide decorated on the surface of low-cost, easily available, and environmentally friendly materials is discussed thoroughly. The materials sources, synthesis, and iron oxide decoration have been discussed. The properties of synthesizing materials are explained with suitable characterization techniques. The optimum arsenic removal conditions obtained from every study have been analyzed. The adsorption capacity obtained from the different materials has been compared with each other. The adsorption mechanism of arsenic to the adsorbent surface has been explained respectively.

2.1 Iron Oxide Nanoparticles for Arsenic Removal

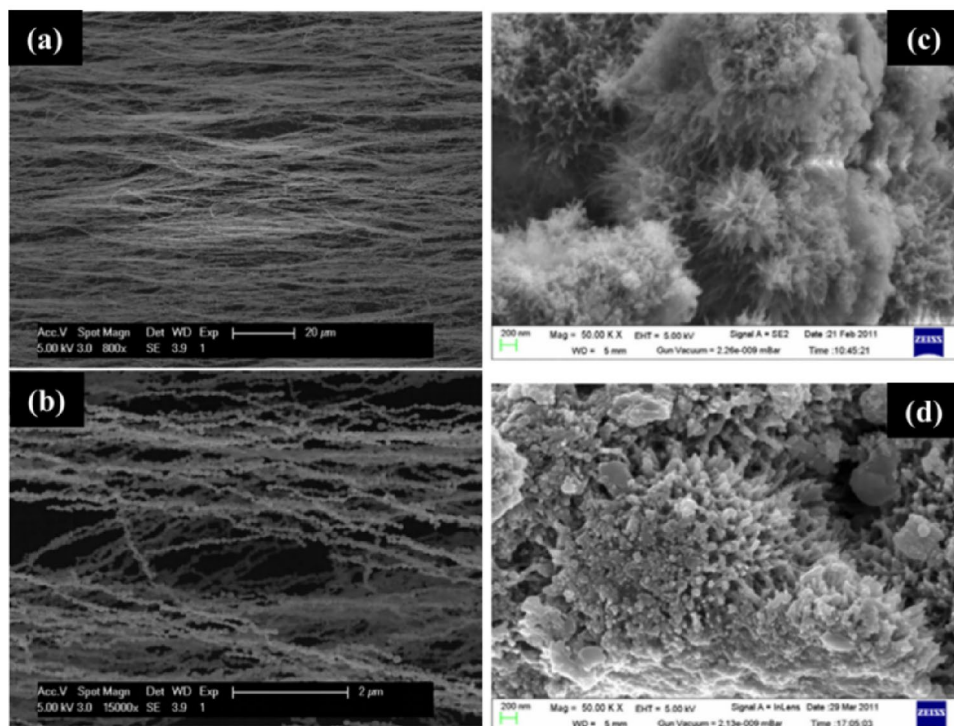
Iron oxide nanoparticles (Fe_3O_4 , Fe_2O_3 , FeOOH , nZVIO) have a greater ability towards arsenic. Therefore, studies have been done using these iron oxide nanoparticles for wastewater treatment due to arsenic contaminations. A detailed analysis of iron oxide nanoparticles synthesis process with surface area

and different conditions for arsenic removal are presented in Table 2. Raul et al. synthesized a nanoflower iron oxide hydroxide for As(III) removal [74]. Here, the Field emission scanning electron microscopy (FESEM) image confirmed the formation of nanoflower, and after As(III) adsorption changes were obtained respectively, as presented in Fig. 4. They obtained the optimum condition of adsorbent dose 1.0 g/L, contact time 300 min, pH 7.28, stirring speed 180 rpm, and temperature 25 °C for maximum As(III) removal with the uptake capacity was 475 $\mu\text{g/L}$ respectively. Desorption of arsenic from exhausted IOH nanoparticles was achieved through regeneration using dilute solutions of HCl (0.5 M) and NaOH (0.2 M and 0.55 M). A desorption efficiency of 75% was observed when NaOH was utilized, whereas HCl as an eluent recorded a desorption efficiency of 54%. Again, a combination of magnetite (55.8 wt%) and maghemite (44.2 wt%) nanoparticles was synthesized by the electrical wire explosion process for the removal of arsenic [75]. Here, 20 cm long 0.3 mm ϕ iron wires were exploded with a capacitance of 3.5 μF and voltage of 11.4 kV respectively, after which iron oxide nanoparticles were formed. It had a high magnetic saturation value of 92.4 emu/g and a surface area of 12 m^2/g respectively. XRD studies confirmed that it had a mixture of maghemite and

Table 2 Iron oxide nanoparticles synthesis process with surface area and different conditions for arsenic removal

Adsorbent	Synthesis process	Surface area (cm^2/g)	Arsenic	Optimum conditions	Isotherm	q_{max} (mg/g)	References
Iron oxide hydroxide	Precipitation	6.577	As(III)	pH 7.28; time 180 min; initial conc. 0–1000 $\mu\text{g/L}$	Redlich–Peterson	0.475	[74]
Iron oxide nanoparticles	Electrical wire explosion	12	As(III) As(V)	pH 6; time 24 h; initial conc. 1–7 mg/L pH 6; time 24 h; initial conc. 1–7 mg/L	Langmuir Langmuir	2.9 3.1	[75]
Magnetite particles	Commercial	–	As(III) As(V)	pH 2; time 3 h; initial conc. 2 mg/L pH 2; time 3 h; initial conc. 2 mg/L	Langmuir Langmuir	3.70 3.70	[77]
Fe_3O_4 nanoparticles	Microemulsion	84.68	As(III)	pH 7.7, time 120 min; initial conc. 32.32 mg/L	Langmuir	7.18	[80]
$\gamma\text{-Fe}_2\text{O}_3$	precipitation	100	As(III)	pH 5–7.5; time 1 h; initial conc. 100 mg/L	Freundlich	45	[81]
Mixed $\alpha\text{-Fe}_2\text{O}_3$ and $\gamma\text{-Fe}_2\text{O}_3$	dispersion-precipitation	121	As(III)	pH 7; time 200 min; initial conc. 0–60 mg/L	Langmuir	46.5	[82]
$\gamma\text{-Fe}_2\text{O}_3$ nano chains	Flame synthesis	151.12	As(V)	pH 7; time 45 min; initial conc. 0.25–	Langmuir	162	[83]
Fe_3O_4 nanoparticles	Precipitation	–	As(III)	300 mg/L	Langmuir	–	[84]
Nano zerovalent iron	Commercial	–	As(V) As(III)	pH 8; time 2 min; initial conc. 50–200 mg/L pH 8; time 2 min; initial conc. 50–200 mg/L pH 6.6–6.8; time 60 min; initial conc. 10 mg/L	– – –	– – –	[84] [85]

Fig. 4 SEM images of the ultra-long nano chains of iron oxides (a, b) [83] and FESEM of iron oxide hydroxide nanoparticles before (c) and after (d) adsorption of As(III) [74]



magnetite phases. The adsorption capacity was 2.9 mg/g for As(III) and 3.1 mg/g for As(V) respectively. Some researchers showed that laboratory-prepared iron oxide nanoparticles were much more efficient than commercial nanoparticles. Mayo et al. synthesized 12 nm size nanoparticles and were able to remove 98% of As(III) and As(V) respectively. The findings demonstrate a significant impact of nanoparticle size on the adsorption and desorption of arsenic, with no loss of Fe_3O_4 nanoparticles being observed. The results indicate irreversible desorption of both As(III) and As(V) from 20 nm Fe_3O_4 nanoparticles, with approximately 1% of the adsorbed As(III) and As(V) being desorbed at pH 6.1 [76]. Choudhury et al. also applied commercial Fe_3O_4 nanoparticles having a size of 20 nm for As(III) and As(V) removal. Furthermore, the study found that arsenic removal from contaminated water depends on various factors, including pH, contact time, initial arsenic concentration, phosphate concentration, and adsorbent concentration. The results suggest that the adsorption of arsenic involves the formation of weak arsenic-iron oxide complexes on the surface of magnetite particles. These findings indicate the potential application of magnetite particles in the design of permeable reactive barriers for groundwater remediation, particularly for in situ remediation of groundwater contaminated with redox-active metals [77]. Here, the spontaneous nature of the adsorption process was found from the thermodynamic study with ΔG value -35.5 kJ/mol. About 2 g/L adsorbent was enough to remove 95% of arsenic from water and the approximate adsorbent cost was 0.09\$ to remove arsenic from one-liter water. They concluded that at

pH 2, the maximum amount of arsenic was removed and the uptake capacity of Fe_3O_4 nanoparticles as obtained from the Langmuir isotherm model in synthetic and groundwater were 3.7 mg/g and 62.66 $\mu\text{g/g}$ for As(III) and As(V) respectively. They had done another study also where they used mixed maghemite and magnetite nanoparticles and all other conditions were same except pH 6.5 [78]. The adsorption capacity was 4.75 mg/g for As(III) and 4.85 mg/g for As(V) respectively. Luther and coworkers also found good results for As(III) and As(V) by using Fe_3O_4 and Fe_2O_3 nanoparticles [79]. They had done comparison studies between both iron oxide nanoparticles and found that the adsorption capacity of Fe_3O_4 was 5680 $\mu\text{g/g}$ and 4780 $\mu\text{g/g}$ for As(III) and As(V). Which was lower than the adsorption capacity of Fe_2O_3 nanoparticles of 20,000 $\mu\text{g/g}$ and 4904 $\mu\text{g/g}$ for As(III) and As(V) respectively. In further studies, Fe_3O_4 nanoparticles were synthesized by the microemulsion process using a low-cost phosphate-free surfactant Extran and used for As(III) removal [80]. Very small size 5 nm particles were formed with a magnetic saturation value of 47 emu/g, which was high enough for rapid magnetic separation in 20 s. Here, the Box-Behnken Design model was used for As(III) removal, and 90.5% of As(III) was removed in the obtained optimum condition of adsorbent dose 0.7 g/L, initial concentration of 33.32 mg/L, and pH 7.7 respectively. In another experiment, As(V) was removed by using $\gamma\text{-Fe}_2\text{O}_3$ nanoparticles [81]. The obtained particle size was very less nearly 10 nm, hence it had superparamagnetic with a magnetic saturation value of 39.6 $\text{A m}^2/\text{kg}$. They observed that when the synthesis condition was Fe: As 20:1, 100% As(V)

was removed and the material had an adsorption capacity of 45 mg/g respectively. The study also noticed that as the pH increases to an alkaline environment, the efficiency of arsenate adsorption from the solution gradually decreases due to the repulsive electrostatic effect. This effect could be potentially harnessed for regenerating the adsorbent. However, the study did not present any data regarding the desorption process. Cheng et al. also prepared mixed α and γ Fe_2O_3 nanoparticles in the dispersion-precipitation method for As(III) removal [82]. In the synthesis of iron oxide nanoparticles, they used acetone for precipitation, and the final materials were calcined for 2 h at 100–500 °C (named FeMag100–FeMag500) respectively. In the FTIR studies, the methyl group vibrations (1552 cm^{-1}) were seen below 200 °C but after that, it was diminished due to the decomposition of organic moiety at the higher temperature. It had a magnetic saturation value of 20 emu/g and shows a second-order chemisorption process. Another observation was with an increase in temperature (250 to 300 °C) the surface area was a decrease from 121 to 99 m^2/g and reached 16 m^2/g at 500 °C. Hence, the final adsorption capacity for As(III) was calculated by FeMag250, which was 46.5 mg/g from the Langmuir isotherm model, respectively. In the presence of phosphate and silicate, As(III) removal rate by these nanoparticles was decreased from 93.9% to 60.9% and 78.5%, respectively. Das et al. synthesized γ - Fe_2O_3 nano chains by a novel flame synthesis process and found very ultra-small size nanoparticles of 4 nm [83]. The SEM images at lower and higher magnifications are shown in Fig. 4. The TEM image showed very beautiful hexagonal structure nanoparticles with a surface area of 151 m^2/g . This material had a very high adsorption capacity of 162 mg/g and proceeds with the second-order kinetics model. After adsorption, it was rapidly separated from arsenic solutions because of its high magnetic saturation value of 77.1 emu/g. Recently, Martinez and team used Fe_3O_4 nanoparticles for As(III) and As(V) removal in a different way [84]. They directly used iron precursor ($\text{FeCl}_2 \cdot 4\text{H}_2\text{O}$ and $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$) to arsenic solution and found about 91% and 96% of As(III) and As(V) removed from the waste solution. In another research, zerovalent was used for As(III) removal where the research team mainly showed the role of UVA-Vis irradiation wavelength for the removal As(III) [85]. They found that under UVA-Vis irradiation the removal capacity was increased to 94% as compare to normal analysis and dark condition analysis respectively.

2.2 Activated Carbon Modified with Iron Oxide Nanoparticles

In recent days activated carbon prepared from different waste materials or commercially available activated carbon is used for arsenic removal. Its surface is negatively charged which decreases the arsenic removal efficiency and also difficulties with its separation. Hence, iron oxide modified with

activated carbon helps to solve these problems, which not only increases the removal capacity but also magnetically separated from the arsenic solution. Different iron-based activated carbon adsorbent with surface area, activated carbon source, and different conditions for arsenic removal are summarized in Table 3. Pirajan et al. used activated carbon prepared from bamboo waste and applied it for both arsenite and arsenate removal [86]. They synthesized bamboo activated carbon (BAC) from bamboo waste in the thermal process at a temperature of 950 °C, after that iron was modified on the BAC surface in the same process to get Fe-BAC. SEM studies of BAC indicated that it had a large porous structure which provides pores for Fe loading and ultimately increases the arsenic removal. The maximum adsorption took place at higher (pH nearly 11) for both arsenite and arsenate by BAC and Fe-BAC. The adsorption capacity for arsenite and arsenate was 0.20 and 0.28 mg/g by Fe-BAC as obtained by the Langmuir isotherm model. In another study, the waste biomass (pinewood sawdust; the carbon content was 43.25%) was collected from a wood factory in Beijing for activated carbon preparation [87]. In the thermal pyrolysis process, Fe_3O_4 nanoparticles were immersed on the activated carbon surface. The maximum adsorption for As(V) was seen in pH 8, contact time 6 h, and initial concentration of 40 mg/L. The adsorption capacity for As(V) was found to be 43.7 and 204.2 mg/g by the activated carbon and Fe_3O_4 -loaded activated carbon, respectively. Maiti et al. used the Tamarind hull for activated carbon preparation and used for As(V) removal from synthetic and real groundwater [88]. Iron oxide was impregnated on the activated carbon surface by the thermal pyrolysis process. The maximum adsorption capacity of this adsorbent for As(V) was 1.17 mg/g. The adsorbent was used for real groundwater (As concentration 264 $\mu\text{g}/\text{L}$) and in the optimum condition of adsorbent dose 3 g/L, contact time 150 min, and pH 3–5, higher than 98% of As(V) was removed. The desorption of arsenic from As(V)-loaded IOITHC was investigated using aqueous solutions covering a pH range from 3 to 12. Subsequent adsorption tests using the regenerated adsorbent displayed an adsorption efficiency of approximately 80% compared to the fresh IOITHC adsorbent, with an initial As(V) concentration of 2.0 mg L^{-1} . Another agricultural by-product ‘Apricot stone’ was used for activated carbon (IAC) preparation and iron (oxy-hydro) oxides (Fe(II) and Fe(III)) was deposited on the IAC surface to prepare IAC-Fe(II) and IAC-Fe(III) [71]. Here As(V) adsorption studies were done with a variety of process parameters like initial As(V) concentration, pH, and temperature. The experiments performed 0.05–0.3 g adsorbent dose with 0.5, 4.5, and 8.5 mg/L As(V) concentration, pH 3, 5, and 7, and temperature 298, 318, and 338 K. The maximum adsorption took place at pH 3, and about 99.5% of As(V) was removed by IAC-Fe(III) adsorbent. The adsorption process was endothermic, and the adsorption capacity

Table 3 Different iron-based activated carbon adsorbent with surface area, activated carbon source, and different conditions for arsenic removal

Adsorbent	Activated carbon source	Arsenic	Optimum conditions	Surface area (cm ³ /g)	Adsorption isotherm	q _{max} (mg/g)	References
Fe-BAC	bamboo waste	As(III) As(V)	pH 10.5; time 180 min; initial conc. 0.5 mg/L pH 10.5; time 180 min; initial conc. 0.5 mg/L	1357	Langmuir Langmuir	0.20 0.28	[86]
Iron oxide-impregnated carbon	Tamarind hull	As(V)	pH 3–5; time 150 min; initial conc. 2 mg/L	304.6	Langmuir	1.17	[88]
GAC-Fe	-	As(V)	pH 6; time 4–5 h; initial conc. 100 µg/L	876	Freundlich	1.430	[90]
IAC-Fe(III)	Apricot stone (agricultural byproduct)	As(V)	pH 3; time 5 h; initial conc. 4.5 mg/L	987	Freundlich and Dubinin–Radushkevich	3.009	[71]
(NZVI/AC)	Coal	As(III) As(V)	pH 6.5; time 72 h; initial conc. 0.20 mg/L pH 6.5; time 72 h; initial conc. 0.20 mg/L	64.9	Langmuir Langmuir	18.19 12.02	[91]
Iron oxide loaded on ACO	Activated charcoal	As(V)	pH 4; time 24 h; initial conc. 5 mg/L	26	Langmuir	27.78	[89]
Activated carbon modified with Fe ₃ O ₄ nanoparticles	Cigarette soot	As(III) As(V)	pH 7; time 90 min; initial conc. 0.5 mg/L pH 3 As(V); time 90 min; initial conc. 0.5 mg/L	575.604	Langmuir Langmuir	80.99 107.96	[29]
Fe ₃ O ₄ -loaded activated carbon	Pinewood sawdust	As(V)	pH 8; time 6 h; initial conc. 40 mg/L	349	Langmuir	204.2	[87]

of IAC, IAC-Fe (II), and IAC-Fe(III) was 0.034, 2.023, and 3.009 mg/g, respectively.

Similarly, Yurum et al. used activated charcoal (AC) to prepare oxidized activated carbon (OAC), and iron oxide nanoparticles were deposited on its surface by microwave hydrothermal technique [89]. In microwave hydrothermal synthesis, three different times i.e., 3, 6, and 9 min were chosen for iron oxide nanoparticles decoration on AC and OAC surfaces. From XRD data it was found that first β -FeOOH was formed in 3 min and within 6 min α -Fe₂O₃ was formed on the AC and OAC surface and in 9 min also same results were obtained. So in microwave heating, very little time is required for magnetic iron oxide nanoparticles deposition on AC and OAC surfaces. The adsorption process was proceeding with second-order kinetics and 99.97% of As(V) was removed in just 5 min. The maximum adsorption capacity was 27.78 mg/g. In a separate study, iron oxide (Fe) was supported on granular activated carbon (GAC) using the precipitation method for As(V) removal [82]. Column studies were conducted to investigate arsenic removal by filling the column with Fe-GAC. The column had a diameter of 2 cm and a height of 30 cm, and it was filled with 32 g of the adsorbent. Different concentrations of As(V) (100, 250, and 500 µg/L) were passed through the column at flow rates of

2.5 and 5 m/h. The maximum adsorption capacity observed was 470 µg/g at 250 µg/L and a flow rate of 2.5 m/h. Furthermore, adsorption onto GAC-Fe beds is considered a preferable method for reducing arsenic (As) concentration in water compared to co-precipitation. The adsorption process offers practical advantages, including ease of operation and management, shorter treatment times, and resulting in cleaner water production with a safer As disposal potential. GAC-Fe filters exhibit higher efficiency in adsorbing As, as well as iron (Fe) and organic pollutants, in comparison to commonly used sand filters [90].

Sahu et al. used cigarette shoots, the waste product of burned cigarettes for activated carbon preparation [29]. Fe₃O₄ nanoparticles were decorated on the cigarette shoot activated carbon (CSAC) by thermal pyrolysis process and applied to reduce As(III) and As(V) from synthetic water. The results were remarkable, and the adsorbent was used for up to five cycles without any difficulties. The maximum adsorption capacities were 80.99 and 107.96 mg/g for As(III) and As(V) at pH 3 and 7 respectively. In one more work, Zhu et al. synthesized nano zero-valent iron oxide supported on an activated carbon surface (NZVI/AC) in the chemical reduction process [91]. About 8.2% of Fe loaded on needle shape carbon surface. They mainly focused on

the effects of both cations and anions on As(III) and As(V) removal and found that phosphate and silicate anions and ferrous cations sharply decreased the removal rate. The maximum adsorption capacity for As(III) and As(V) were 18.2 and 12.0 mg/g at pH 6.5. Moreover the arsenic-loaded adsorbent, NZVI/AC, was regenerated by shaking it with 0.1 M NaOH at room temperature. Within 12 h, approximately 100% of the adsorbed arsenic was successfully desorbed by the alkaline solution. In contrast, desorption using phosphate at pH 3.5 and 6.5 only achieved stripping efficiencies of 36% and 46%, respectively.

2.3 Biochar Modified with Iron Oxide Nanoparticles

Biochar is another important material that can support iron oxide for arsenic removal. Here, the adsorbent is prepared by the direct impletion of iron oxide on the biochar surface or iron precursor loaded on the biomass surface in the pyrolysis process. The iron oxide was deposited on the inner, outer, and surface pores of biochar which would rapidly adsorb arsenic from an aqueous environment. Different iron-based biochar adsorbent with surface area, biochar source, and different conditions for arsenic removal are presented in Table 4. Hence, Zhang et al. used hyacinth biomass for biochar (MW2501) preparation and used precipitation process for iron oxide decoration on its surface, later on, used for As(V) removal [92]. This magnetic biochar had an adsorption capacity of 7.41 mg/g, as obtained from the Langmuir–Freundlich model. The maximum adsorption took place at pH 5.3 and was followed by a pseudo-second-order kinetics model. Whereas the sorbent MW2501 exhibited excellent initial As(V) removal efficiency, achieving

around 100% removal at an initial concentration of 5 mg/L and a solid/solution ratio of 1:200. Upon regeneration and subsequent sorption cycles, the As(V) removal remained at approximately 80%. However, in the third and fourth cycles, the efficiency declined to 50.8–65.5%. Despite this reduction, the sorbent could still be easily separated from the solution using a magnet after four cycles and provide a cost-effective option for treating most As-containing water in nature, particularly those with As concentrations below 5 mg/L. Sometimes fibrous biochar had been synthesized to increase the As(III) and As(V) adsorption capacity [93]. Here, fibrous biochar was prepared in a furnace at 800 °C for 2 h from the cotton fiber source. Then iron oxide was decorated with fibrous biochar surface in the hydrothermal process at 120 °C for 6 h and denoted as Fe-NN/BFs. As the name indicates, the SEM image (Fig. 5) confirmed the formation of a nanoneedle array of iron oxide on the fibrous biochar surface. The maximum adsorption took place at 15 and 120 min for As(V) and As(III), followed by a second-order kinetics model. The maximum adsorption capacity was 70.22 and 93.94 mg/g for As(III) and As(V) obtained from the Langmuir model. The materials were also applied for a real arsenic-contaminated water system where the initial arsenic concentration was 275 µg/L. The As(III) concentration was reduced to 5.31 µg/L in 15 min with an adsorbent dose of 2 g/L, whereas As(V) concentration was decreased to 1.22 µg/L in 30 min using 1.5 g/L of Fe-NN/BFs.

Again, bamboo waste was used for bamboo charcoal preparation and iron oxide was modified in the precipitation process [94]. This modified adsorbent showed a good surface area of 277.875 m²/g which would help for a higher percentage of As(III) and As(V) removal. The adsorption

Table 4 Different iron-based biochar adsorbent with surface area, biochar source, and different conditions for arsenic removal

Adsorbent	Biochar source	Arsenic	Optimum conditions	Surface area (m ² /g)	Adsorption isotherm	q _{max} (mg/g)	References
Rice husk iron oxide composite biochar	Rice husk	As(III)	pH 7.5; time 12 h; initial conc. 0.05–0.2 mg/L	300	Langmuir	0.096	[96]
Wheat husk iron oxide composite biochar	Wheat husk	As(III)	pH 7.5; time 12 h; initial conc. 0.05–0.2 mg/L	339	Langmuir	0.116	[96]
Iron-loaded biochar	Waste walnut shell	As(V)	pH 7; time 10 h; initial conc. 0.1–5 mg/L	418	Langmuir	1.19	[97]
Iron-Modified Bamboo Charcoal	Bamboo	As(III) As(V)	pH 4–5; time 30 h; initial conc. 2.343 mg/L pH 9–4; time 35.5 h; initial conc. 2.343 mg/L	277	Freundlich Freundlich	7.237 19.771	[94]
Bark-based magnetic iron oxide particles	<i>Tamarindus Indica</i>	As(III)	pH 7; time 11 h; initial conc. 92–1981 µg/L	40.87	Langmuir	19.61	[98]
Magnetic gelatin-modified biochar	Chestnut shell	As(V)	pH 4; time 24 h; initial conc. 0–50 mg/L	-	Langmuir	45.8	[95]
AMP@Fe ₂ O ₃ nanocomposite	<i>Aegle marmelos</i>	As(V)	pH 3, time 250 min, initial conc. 0.1–50 mg/L	135.29	Langmuir	69.65	[99]

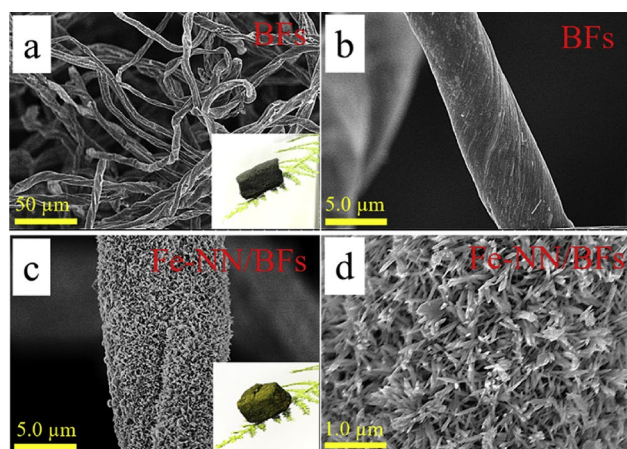


Fig. 5 SEM images of **a, b** BFs and **c, d** Fe-NN/BFs [93]

process for arsenic (As(III) and As(V)) was followed by a pseudo-second-order kinetics model with higher R^2 values than the pseudo-second-order and Elovich model kinetics model. Here, the intraparticle diffusion model showed multiple natures of the relationship and indicated a complex mechanism for As(III) and As(V). Zhou et al. prepared gelatin-modified magnetic biochar for As(V) removal [95]. Here, the chestnut shell was used for biochar formation, and gelatin a natural polymer rich in amino acids modified on the biochar surface to enhance adsorption capacity. Again magnetic iron oxide was loaded for magnetic separation of adsorbent after As(V) removal from the solution. It has a magnetic saturation value of 42 emu/g, which was high enough for magnetic separation. The kinetics studies showed that the adsorption process was chemisorption in nature and interaction took place between As(V) anions and positively charged functional groups of adsorbent through ion exchange or chelating process. The adsorption capacity was 45.8 mg/g for As(V) by gelatin-modified magnetic biochar and As–O interactions and electrostatic interactions was the main adsorption mechanism. Further two novel types of biochar were prepared from rice husks and white husks, then iron oxide was decorated on these biochar surfaces by the pyrolysis process at 600 °C for 1 h under nitrogen temperature [96]. Batch adsorption studies were performed for As(III) removal using RHIOB and WHIOB. Adsorption isotherm studies were performed at 10, 25, and 40 °C, pH 6.5–7.5 with a variation of As(III) concentration from 50 to 5000 $\mu\text{g/L}$. The output of this experimental data fitted with Langmuir, Freundlich, Temkin, Sips, Radke and Prausnitz, Redlich–Peterson, and Toth isotherms models. The different models were fitted to experimental data obtained from different temperature but adsorption capacities obtained from both cases in the Langmuir model were highly close to the experimental adsorption capacities. The As(III) adsorption capacity for RHIOB and WHIOB were 0.096 and 0.116 mg/g,

respectively. In another work, the waste walnut shell was used for iron-loaded biochar (ILB) preparation in the microwave pyrolysis process where the microwave frequency was 2.45 GHz and tubular furnace power was 800 W during ILB preparation [97]. The adsorption capacity of ILB was 1.19 mg/g respectively. In another study, the bark of *Tamarindus Indica* was used to prepare biochar, and Fe_3O_4 nanoparticles were deposited on its surface by thermal pyrolysis process [98]. The materials had a surface area of 40.87 m^2/g and a magnetic saturation value of 38.62 emu/g respectively. Langmuir isotherms model was best fitted as compared to Freundlich, and Dubinin–Radushkevich isotherms models. The adsorption capacity for As(III) and As(V) calculated from the individual study was 19.61 mg/g for As(III) and 13.58 mg/g for As(V) respectively. They also showed that when both As(III) and As(V) were present in the solution, the adsorption capacity for As(III) and As(V) was 7.04 and 9.43 mg/g respectively. In another research, *Aegle marmelos* leaves (Indian bael) were used for magnetic bio-adsorbent preparation, and Fe_2O_3 nanoparticles were incorporated on its surface by pyrolysis process at 300 °C [99]. The material showed good results for As(V) removal in optimum condition of pH 3, adsorbent dose 0.1 g/L, initial concentration 0.5 mg/L and contact time 250 min. The adsorbent had an uptake capacity of 69.65 mg/g and can be used for five cycles without any problems.

2.4 Magnetic Chitosan Adsorbents

Chitosan (CS) is produced from chitin (exoskeleton of animals such as crabs, prawns, crayfish, woodlice, shrimps, lobsters, krill, and barnacles) by the deacetylation process. Hence, it is easily obtained from nature and biodegradable. It has a higher number of amino and hydroxyl groups which can act as possible adsorption sites [100]. Thus, the specific surface properties of iron oxide can be increased by chelating with CS will be more feasible for arsenic removal [101, 102]. Table 5 shows the different magnetic chitosan adsorbent with surface area and different conditions for arsenic removal. Liu et al. used chitosan and prepared magnetic chitosan nanoparticles for arsenic removal [103]. Here, the particles were 10 nm in size and about 95% of both As(III) and As(V) were removed within the time period of 15 min. The adsorption capacities observed were 60.2 and 65.5 mg/g for As(III) and As(V) respectively. This adsorbent could be used for real applications as 95% of the adsorption capacity was stable even after 10 consecutive cycles. Abdollahi et al. also used chitosan-coated Fe_3O_4 nanoparticles for As(III) removal and adsorption capacity was less about 10.5 mg/g at pH 3 by this adsorbent [104]. Chauhan and coworkers synthesized Chitosan/PVA/zerovalent iron nanofibrous (CPZ) materials in the electrospinning process to remove arsenic from water [105]. The CS fiber was formed, and a

Table 5 Different magnetic chitosan adsorbent with surface area and different conditions for arsenic removal

Adsorbent	Arsenic	Optimum Conditions	Surface area (m ² /g)	Adsorption Isotherm	q _{max} (mg/g)	References
Iron chitosan granules	As(III)	pH 7; time 4 h; initial conc. 1–10 mg/L	96.8	Langmuir	2.32	[109]
	As(V)	pH 7; time 4 h; initial conc. 1–10 mg/L		Langmuir	2.24	
chitosan-coated magnetic nanoparticles	As(III)	pH 9; time 90 min; initial conc. 1–100 mg/L	–	Langmuir	10.5	
Iron chitosan flakes	As(III)	pH 7; time 4 h; initial conc. 1–10 mg/L	1.44	Langmuir	16.15	[109]
	As(V)	pH 7; time 4 h; initial conc. 1–10 mg/L		Langmuir	22.47	
Magnetic chitosan biochar (MBC)	As(V)	pH 5; time 3 h; initial conc. 0.2–50 mg/L	–	Langmuir	17.876	[107]
iron-chitosan-coated sand (ICCS)	As(III)	pH 7; time 2 h; initial conc. 100–1000 mg/L	0.3875	Freundlich	26	[108]
	As(V)	pH 7; time 2 h; initial conc. 100–1000 µg/L		Freundlich	56	
Magnetic chitosan nanoparticle	As(III)	pH 6.8; time 15 min; initial conc. 50 mg/L	108.32	Langmuir	60.2	[103]
	As(V)	pH 6.8; time 15 min; initial conc. 50 mg/L		Langmuir	65.5	
Chitosan zerovalent iron nanoparticle	As(III)	pH 7; time 180 min; initial conc. 1–10 mg/L	69	Langmuir	94	[110]
	As(V)	pH 7; time 180 min; initial conc. 1–10 mg/L		Langmuir	119	
CS functionalized iron nanocomposite	As(III)	pH 7; time 10 h; initial conc. 2–60 mg/L	111.8	Langmuir	108.6	[106]
Chitosan/PVA/zerovalent iron nanofibrous	As(III)	pH 7; time 4 h; initial conc. 0.1–10 mg/L	–	Langmuir	142.9	[105]
	As(V)	pH 7; time 4 h; initial conc. 0.1–10 mg/L		Freundlich	200	

zerovalent iron oxide nanoparticle was decorated on its surface with a size of less than 100 nm. These fibrous materials showed good results for arsenic removal with high uptake capacities for As(III) (142.9 mg/g) and As(V) (200 mg/g). This CPZ adsorbent material was regenerated with 0.01 M NaOH solutions and regenerated materials removed 20% less in the fifth cycle than in fresh cycles which suggested that there was a strong interaction between adsorbent and adsorbate. Chitosan (CS) functionalized iron nanosheet was synthesized by doping method and aimed for As(III) removal [106]. Here they doped 0–0.5% CS on an iron oxide surface and CS doping was chemically confirmed by FTIR studies. Where the bends were shifted from 3340 cm⁻¹ bends for –NH₂ decreased and 1637 cm⁻¹ for –OH was moved to 1019 cm⁻¹. This confirmed that CS (0.5%) was doped on an iron oxide surface with a surface area of 118 m²/g. Again, the As(III) removal capacity was increased with an increase in % of CS. The maximum removal was achieved at 0.5% with an uptake capacity of 108.6 mg/g.

To increase the surface area or in other words to increase the adsorption capacity of As(V), the biochar surface was

modified with magnetic chitosan composite [107]. In this study, rice straw biomass was used to prepare biochar, and magnetic chitosan was modified on its surface. The authors compared two materials: chitosan-modified biochar (CB) and magnetic chitosan biochar composite (MBC) for their adsorption capabilities. MBC showed maximum adsorption compared to CB. The adsorption capacity of MBC for As(V) was 14.92 mg/g, 17.16 mg/g, and 17.876 mg/g at temperatures of 25 °C, 35 °C, and 45 °C, respectively, indicating an endothermic nature of the adsorption process with a ΔH value of 232.630 kJ/mol.

Even after five cycles of reuse, the regenerated MCB still retained about 8.078 mg/g uptake capacity for As(V), indicating good reusability for As(V) removal. However, the decrease in adsorption capacity could be attributed to the reduction in surface area and pore volume, along with relatively weak functional groups. Another study had been done where iron-chitosan-coated sand was used for As(III) and As(V) removal [108]. Here, a comparison study had been done between chitosan-coated sand (CCS) and iron-chitosan-coated sand (ICCS). The surface area of CCS and

ICCS were 0.3978 and 0.3875 m²/g respectively. For both As(III) and As(V), maximum removal took place at near-neutral pH. They applied this material for real water application in the Shuklaganj area of Kanpur district, UP, India where the arsenic concentration was higher than 500 µg/L. Column experiments study has been performed with ICCS and arsenic concentration reduced to less than 5 µg/L.

Gupta et al. used iron chitosan flakes (ICF) and iron chitosan granules (ICB) to remove arsenic and found an adsorption capacity of 16.15 and 2.32 mg/g for As(III) and 22.47 and 2.24 mg/g for As(V) at pH 7 by ICF and ICB adsorbent respectively [109]. Here, they performed column studies, where 112-bed volumes of As(V) and 147-bed volumes of As(III) pointed wastewater were passed through the column (filled with iron chitosan flakes) and it was found that the 500 µg/L arsenic reduces to less than 10 µg/L. Again Gupta et al. used chitosan zerovalent iron nanoparticle (CIN) for arsenic removal [110]. The adsorption capacity of CIN was 94 mg/g for As(III) and 119 mg/g for As(V), respectively. This adsorbent had higher efficiency than the previous one.

2.5 Magnetic Cellulose Nanocomposite

Currently, biopolymers like cellulose act as a promising adsorbent for arsenic removal [111, 112]. The agricultural waste product or natural fiber which is biodegradable and economical has lots of components like cellulose, lignin, protein, hemicellulose, pectin, and fatty acid. This contains several functional groups and are more suitable for surface

modifications which can also be used as a source of cellulose [113–115]. Hence, a combination of these cellulose and cellulose obtained from natural materials can be combined with iron oxide and act as an efficient biocomposite with high adsorption capacity and easily recovered using an external magnet [116, 117]. Table 6 represents the different magnetic cellulose nanocomposite adsorbent with surface area, cellulose source, and different conditions for arsenic removal. Yu and their coworker synthesized cellulose-iron oxide nanoparticles to remove arsenic from aqueous solutions [118]. They used cotton as a cellulose source and Fe₂O₃ nanoparticles were dispersed on the cellulose matrix by a step precipitation process. To confirm material formation, there were characterized by XRD, TGA, XPS, FTIR, VSM, SEM, and TEM instrumental techniques. The size of Fe₂O₃ on the cellulose matrix was 61 nm and had a magnetic saturation value of 13.2 emu/g respectively. The effects of anions on arsenic removal on this adsorbent follow the order of sulfate < nitrate < phosphate. The adsorption efficiency was 23.16 and 32.11 mg/g for As(III) and As(V) respectively. In another study, Zhou et al. also synthesized cellulose-nZVI composites using medical cotton and nZVI was decorated in a reduction process [119]. The cellulose formation was confirmed with XRD peak at 22.7° (002) and 34.5° (040) respectively and nZVI formation was confirmed with 2θ of 44.7° (110) and 65.02° (200) planes of Fe⁰ with JCPDS card no. 06–0696. The cellulose-nZVI composites had a saturation magnetization value of 52.7 emu/g and could be easily separated after adsorption. Maximum As(III) was removed

Table 6 Different magnetic cellulose nanocomposite adsorbent with surface area, cellulose source, and different conditions for arsenic removal

Adsorbent	Cellulose source	Arsenic	Optimum Conditions	Surface area (m ² /g)	Adsorption Isotherm	q _{max} (mg/g)	References
iron-coated honeycomb	Honeycomb	As(V)	pH 7.5; time 14 h; initial conc. 100–500 µg/L	–	Langmuir	0.961	[125]
Magnetic wheat straw	wheat straw	As(III) As(V)	pH 7; time 12 h; initial conc. 1–28 mg/L pH 3; time 12 h; initial conc. 1–28 mg/L	–	Langmuir Langmuir	3.898 8.062	[121]
Fe-JF	Jute fiber	As(III)	pH 7; time 200 min; initial conc. 1–150 mg/L	–	Langmuir	12.66	[122]
Cellulose@iron oxide nanoparticles	Cotton	As(III) As(V)	pH 7; time 4 h; initial conc. 50 mg/L pH 2; time 4 h; initial conc. 100 mg/L	113	Langmuir Langmuir	23.16 32.12	[118]
JF@Fe ₂ O ₃ nanocomposite	Jute fiber	As(V)	pH 3; time 90 min; initial conc. 0.1–50 mg/L	95.43	Langmuir	48.06	[123]
Cellulose@nZVI composites	Medical cotton	As(III)	pH 8; time 40 min; initial conc. 10 mg/L	–	Langmuir	92.25	[119]
Tea waste to modified Fe ₃ O ₄ nanoparticles	Tea waste	As(III) As(V)	pH 7; time 12 h; initial conc. 2 mg/L pH 7; time 12 h; initial conc. 0.2–0.8 mg/L	–	Langmuir Langmuir	188.69 153.8	[124]

at pH 8 and the adsorption capacity was 92.25 mg/g as calculated from the Langmuir model. The composite was regenerated with 0.1 M HCl solution, where the removal efficiency was 99.31%, and after use for four cycles the removal rate was 92.17% respectively. Similarly, Hokkanen et al. synthesized magnetic iron nanoparticles modified microfibrillated cellulose (FeNP/MFC) and remove As(V) from aqueous solutions [120]. 100% of As(V) was removed at pH 2 with a contact time of 75 min. The maximum adsorption capacity was 2.460 mmol/g, respectively.

Tian and their coworker synthesized magnetic wheat straw (MWS) by incorporating different Fe₃O₄ nanoparticles on the wheat straw surface [121]. Here, XRD studies of WS indicated that cellulose was the main component present on the WS, so Fe₃O₄ nanoparticles were easily attached through its functional groups present on the cellulose. MSW had a magnetic saturation value of 11.87 emu/g, which was also high enough for magnetic separations. The maximum adsorption capacity was 3.898 mg/g for As(III) and 8.062 mg/g for As(V), respectively. Hao et al. synthesized jute fiber modified with iron oxyhydroxide (FeOOH) in normal precipitation for As(III) removal [122]. First, jute fibers were esterified with succinic anhydride to graft with carboxyl groups to increase the iron(III) amount on the JF surface and the maximum Fe on the JF surface was 208.2 mg/g. They performed a comparative study between JF and Fe-JF removal efficiency for As(III). Here, iron leaching was about 0.178 mg/g to the water which meets the requirements of iron in the water. The As(III) adsorption capacity was 12.66 mg/g. Again Sahu et al. also used jute fiber and decorate Fe₂O₃ nanoparticles (JF@Fe₂O₃ nanocomposite) for As(V) removal [123]. They found that iron oxide nanoparticles were dispersed on the jute fiber surface and increased the surface area from 5 to 95 m²/g respectively. The adsorption capacity of this composite material was 48.06 mg/g at pH 3. The kinetic was followed by the pseudo-second-order model and maximum adsorption took place in 90 min. The JF@Fe₂O₃ nanocomposite was also able to remove As(V) in the presence of other competitive ions. Similarly, Lunge et al. used tea waste to modify Fe₃O₄ nanoparticles on its surface [124]. The Fe₃O₄ nanoparticles were 5–25 nm in size which would increase the surface area. The adsorption capacity was 188.69 mg/g for As(III) and 153.8 mg/g for As(V), respectively. To synthesize 1 g of MION-Tea (magnetic iron oxide nanoparticles) adsorbent, only 136 rupees are required, and the adsorbent can be used for 5 cycles without any decrease in removal rate. To remove 100 L of As(III) contaminated water by using this MION-Tea adsorbent, the cost was only 14 rupees. Hence, this can be potentially applied to arsenic water treatment. Some researchers used novel waste like honeycomb briquette cinder and Fe₂O₃ nanoparticles decorated on its surface and used for As(V) removal [125]. They found about

961.5 µg/g of As(V) was removed at pH 7.5 and time 14 h. Langmuir's model was fitted to experimental data with an R² value of 0.999 and R_L value of 0.118, respectively. The other negatively charged species had the following order: PO₄³⁻ > HCO₃⁻ > F⁻ > Cl⁻ on As(V) removal.

2.6 Iron Oxide Nanoparticles Doped Mineral Oxides Composite

In recent days minerals, and adsorbents (sand, rock, and clay materials) have been applied for wastewater treatment for their low cost, easy availability, and environmentally friendly nature [126, 127]. But due to its negative surface charge, the efficiency of arsenic is very low [128, 129]. Hence, magnetic composite materials are prepared with iron oxide nanoparticles and used to remove arsenic from wastewater [130]. Table 7 gives details information of different iron oxide doped minerals nanocomposite, surface area, mineral source, and different conditions for arsenic removal. Devi and their coworker also used iron oxide-coated sand and limestone for As(III) removal from contaminated water [130]. The As(III) removal percentage was very high and 96% to 98.6% of As(III) was removed from 100 and 400 µg/L initial concentration arsenic solution. The effect of pH studies was performed in pH 4.8, 7.1, 8.5, and pH 10.4, and a near-neutral pH maximum amount of As(III) was removed. The pseudo-second-order was more accurate to the experimental data which was performed in the arsenic concentration of 200 and 500 µg/L. Column studies were completed where column height and diameter were 120 cm and 7.0 cm respectively, then iron oxide-coated sand filled and 200 µg/L arsenic were passed through it which reduced to 10 µg/L respectively. Maji and his coworker synthesized iron-oxide-coated natural rock (IOCNR) to remove arsenic from the contaminated groundwater of Lanyang Plain, North Eastern Taiwan [131]. About 75% of arsenic was removed from real water in the optimum conditions of an adsorbent dose of 15 g/L, initial concentration of 40 µg/L, pH 7, agitation speed of 180 rpm, and contact time 6 h. The arsenic concentration was reduced to less than 10 µg/L, which is below maximum arsenic accessible limit. Here, the pH of water was 7.5 as As(III) was the major species than As(V). First order kinetic model and Langmuir isotherm models were fitted to the experimental data and adsorption was spontaneous in nature as per thermodynamic data. The maximum arsenic adsorption capacity was 0.36 mg/g. They also used this iron-oxide-coated natural rock as an adsorbent for As(III) removal in column experiments and found satisfactory results [132]. Srivastava et al. synthesized modified iron-coated sand (DMICS) for As(III) elimination from contaminated water [133]. The sand was collected from river tons in Allahabad (India). Maximum As(III) was removed in the pH range of 6.5–7.5. The isotherm study

Table 7 Different iron oxide doped minerals nanocomposite, surface area, mineral source, and different conditions for arsenic removal

Adsorbent	Source	Arsenic	Optimum Conditions	Surface area (m ² /g)	Adsorption Isotherm	q _{max} (mg/g)	References
Iron oxide-coated sand and limestone	Sand and limestone	As(III)	pH 7.2; time 2 h; initial conc. 100–400 µg/L	–	Langmuir	0.075	[130]
Iron-oxide-coated natural rock	Rock	As(III)	pH 7.5; time 6 h; initial conc. 40 µg/L	15.312	Langmuir	0.36	[131]
Iron-coated sand (DMICS)	Sand	As(III)	pH 6.5–7.5; time 8 h; initial conc. 0.5–2.0 mg/L	–	Langmuir	0.29	[133]
Iron-modified light-expanded clay	Clay	As(V)	pH 6–7; time 24 h; initial conc. 1 mg/L	–	Langmuir	3.31	[134]
Iron oxide-modified clay-activated carbon	Clay	As(V)	pH 3; time 24 h; initial conc. 5–75 mg/L	433	Langmuir	5.0	[135]
Clay-supported zerovalent iron nanoparticles	Clay	As(III)	pH 2.75–9; time 30 min; initial conc. 100 mg/L	200.66	–	–	[136]

was followed by the order Langmuir > Freundlich > Temkin with the experimental data. The obtained adsorption capacity was 0.29 mg/g respectively. Same time, iron-modified light-expanded clay was used to remove As(V), and found that the material had an adsorption capacity of 3.31 mg/g at pH 6–7 [134]. Power et al. used iron oxide-modified clay-activated carbon for As(V) removal [135]. Here, Na-alginate and coconut shells were used as a source of clay and activated carbon. The prepared adsorbent had a high surface area of 433 m²/g, which was very high compared to single iron oxide nanoparticles. The FTIR studies suggested that a higher number of hydroxyl and carboxyl groups were present on the adsorbent surface which was responsible for As(V) adsorption. About 85% of arsenic was removed at pH 3 and decreased to 42% at pH 9 at the same time with an increase in As(V) concentration from 5 to 75 mg/L the adsorption capacity increased from 2.06 to 4.9 mg/g respectively. The adsorption experiments were done at low-level concentration and As(V) concentration was decreased from 50 to 7.45 µg/L, respectively. Some research had been done where the clay surface was decorated with zerovalent iron oxide nanoparticles for As(III) removal [136]. In the synthesis process, a green way of approach had been conducted and tea liquor was used as a reducing agent and successfully prepared nZVI on the clay surface. This adsorbent had the particle size and high surface area of 59.08 nm and 200.66 m²/g and was able to remove 99% of As(III) over a large pH range.

3 Adsorption Mechanism

Several studies verified that the adsorption of arsenic on the iron-based adsorbent surface have been carried out by inner-sphere complexation i.e. monodentate, bidentate, and

tridentate ligand exchange, electrostatic attraction, and ion exchange mechanism [137–139]. Furthermore, instrumentals such as Fourier-transform infrared spectroscopy (FTIR), and X-ray photoelectron spectroscopy (XPS) are used to elucidate the attachment of arsenic species on the adsorbent surface [114, 140].

The adsorption of As(III) on the iron oxide nanoparticles' surface was due to the inner sphere mechanism [82]. In FTIR spectra after adsorption, the appeared bend at 780 cm⁻¹ was for As(III)-O stretching vibrations. This indicated that surface hydroxyl was replaced by arsenic and the Fe–O–As bond was formed. Sahu et al. had also shown that As(V) is attached to CSAC/Fe₃O₄ composite surface through both ligand exchange (monodentate and bidentate) and electrostatic attraction as shown in Fig. 6 [29]. Here, they used FTIR studies to clear the adsorption mechanism. The adsorbent had Fe–OH vibration at 1080 cm⁻¹, but after adsorption, this bend was completely removed, and two new bends were seen at 792 and 807 cm⁻¹ for Fe–O–As(III) and Fe–O–As(V) vibrations respectively. This suggested that –OH groups were replaced by arsenic species by the inner sphere ligand exchange mechanism. Andjelkovic et al. also proposed similar types of adsorption mechanisms for As(III) and As(V) on the surface of iron-oxide nanowires on the basis of FTIR studies [48]. The iron-oxide nanowires had a spectrum of 952 cm⁻¹ for Fe–OH bending vibration. After the adsorption of As(V) and As(III), this Fe–OH bend completely disappeared and the new bend was seen at 812 cm⁻¹ for As(V)-O vibrations but in the case of As(III), the 912 cm⁻¹ bends were decreased. This indicated As(V) and As(III) were attached to where –OH groups on the adsorbent surface played a major role. They also said that the As(V) was attached by electrostatic attraction of Fe–OH₂⁺ groups of adsorbent and H₂AsO₄⁻ species. Similar studies were obtained by another researcher, where the adsorption

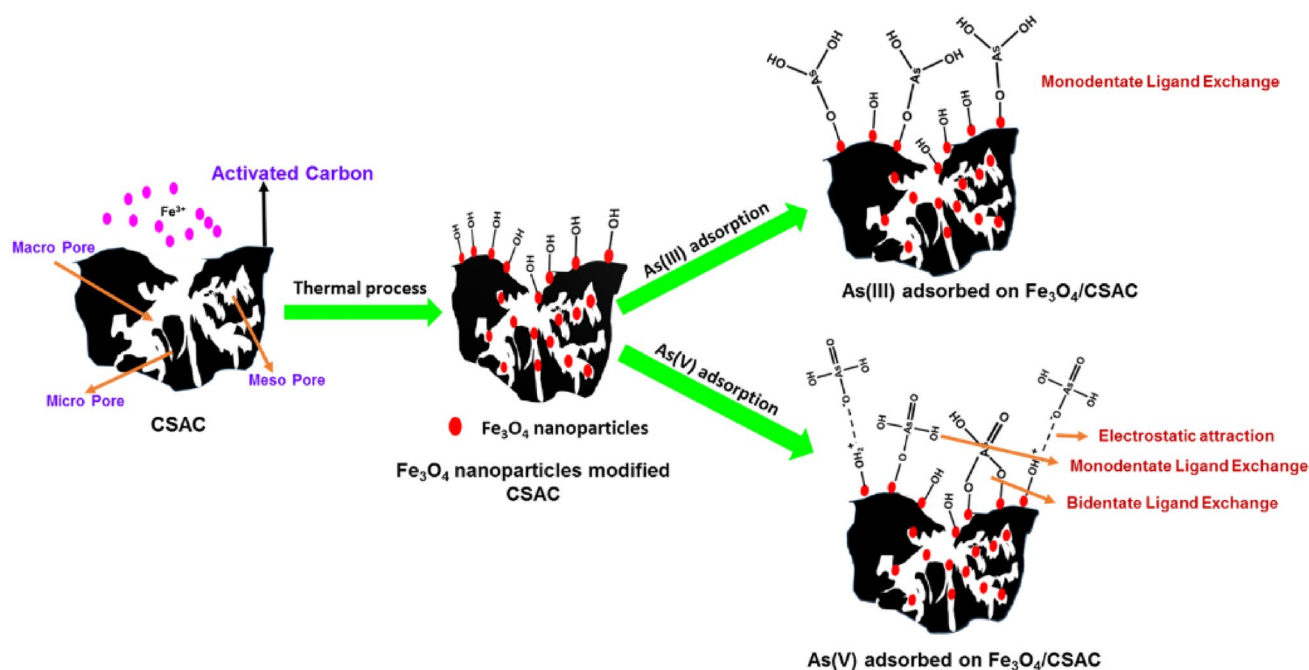


Fig. 6 Adsorption of As(III) by ligand exchange and As(V) by ligand exchange and electrostatic attraction process [29]

mechanism for As(V) to the iron oxide-loaded activated carbons was mainly due to electrostatic attraction going on between positively charged surface (Fe-OH^+ and Fe-OH_2^+) and negatively charged As(V) species (H_2AsO_4^- and HAsO_4^{2-}) [71, 86–91]. They showed that below pH_{PZC} the surface had a high positive charge and maximum adsorption took place in this region and above pH_{PZC} , the adsorbent had negatively charged so repulsion took place. So As(V) is attached through electrostatic attraction on the adsorbent surface.

Zhang et al. explained the As(V) adsorption mechanism on the basics of FTIR and XPS analysis. The FTIR of MW250 (magnetic biochar prepared at 250 °C) after As(V) adsorption, the bend at 3417, 3472, 3554, and 1319 cm^{-1} for surface hydroxyl groups were decreased to a negligible level, which suggested that surface hydroxyl groups attached to Fe were replaced by $\text{H}_2\text{AsO}_4^-/\text{HAsO}_4^{2-}$ groups by ligand exchange mechanism [141]. Also, the bend at 1602 cm^{-1} for C=O bend in $-\text{COO}$ groups was decreased which suggested that a hydrogen bonding was held between $-\text{COO}$ groups and hydroxyl groups of $\text{H}_2\text{AsO}_4^-/\text{HAsO}_4^{2-}$ [142]. XPS spectra after As(V) adsorption showed the presence of three high-resolution spectra of $\text{As}2\text{p}_{3/2}$. The peak at 1326.02 eV was for As(III) due to oxidative transformation and two other peaks at 1327.01 and 1327.67 eV were for As(V). This also suggested that As(V) was adsorbed on the MW250 adsorbent surface. The bark-based magnetic iron oxide particles for As(III) and As(V) also showed ligand exchange of hydroxyl groups [98].

The adsorption mechanism of As(III) and As(V) by Fe-NN/BFs was also examined on the basics of XPS studies. After adsorption, the binding energies of As(III) and As(V) were 44.7–5.2 eV and 45.9–46.3 eV respectively. This mainly showed that arsenic was attached to the Fe-NN/BFs surface but the mechanism was explained by O 1 s XPS patterns of Fe-NN/BFs. Before adsorption, the peak at 529.8, 531.4, and 532.5 eV represented the O lattice of MO, O in hydroxyl groups (MOH), and O in adsorbed H₂O, respectively [143]. But after adsorption of As(III) and As(V), the ratio of $\text{O}_{\text{M-OH}}$ to $\text{O}_{\text{M-O}}$ was changed from 0.96 to 0.87 and 0.88 respectively, which confirmed that Fe-OH groups were responsible for As(III) and As(V) adsorption by forming Fe–O–As bond through inner complex ligand exchange mechanism. The As(V) adsorption mechanism was explained on the surface of the AMP@Fe₂O₃ nanocomposite. Here, they found a point of zero charges of adsorbent before and after adsorption were 6.5 and 5.8 respectively. This shift in charge after adsorption was due to electrostatic attraction. Same time the FTIR bend of AMP@Fe₂O₃ nanocomposite for Fe–OH (1083 cm^{-1}) was removed and a new bend was seen at 807 cm^{-1} which also suggested that $-\text{OH}$ groups were replaced by ligand exchange mechanism [144]. Zhou et al. also suggested oxygen-containing functional groups were responsible for As(V) on magnetic gelatin-modified biochar surfaces based on FTIR and XPS analysis. Singh et al. proposed the As(III) mechanism by monodentate corner-sharing 2C or 1 V complex formation and conversion to a bidentate edge-sharing 2C or 1 V complex [96].

In other studies, researchers found that As(III) was oxidized to As(V) by nano-zerovalent iron oxide nanoparticles, and then As(V) was attached to the adsorbent surface through electrostatic attraction [145–147]. Chitosan fibre-supported zero-valent iron nanoparticles were found the same types of results for As(III) and As(V) adsorption [148]. After the adsorption of As(III), the oxidation of Fe(0) was marked as the binding energy at 707 eV was shifting to higher energy bands which will be more favourable for As(V) adsorption. Again, the binding energy at 533.1 eV was for the more distinguished features for O in the As(V). This suggested that As(III) was largely oxidized to As(V) by nZVIO on the adsorbent surface (Fig. 7). The As(V) took place through electrostatic adsorption between protonated hydroxyl groups on the chitosan surface and $\text{H}_2\text{AsO}_4^-/\text{HAsO}_4^{2-}$ of As(V) species, respectively. Gupta et al. also found similar types of observation of As(III) adsorption through oxidation/adsorption on chitosan zerovalent iron nanoparticle surface [110].

Arsenic also adsorbs on the adsorbent surface by an ion exchange mechanism [149]. Majhi et al. found an ion-exchange mechanism for the adsorption of As(III) on the iron-oxide-coated natural rock surface [131, 132]. They found cation exchange was going on between Fe oxyhydroxides ($=\text{FeOOH}$) and As(III)-contained synthetic water, respectively. They found that first As(III) was converted to As(V) by the adsorbent and then As(V) was attached to iron-oxide-coated surfaces through $-\text{OH}$ by weak hydrogen bonding and lastly attached on the adsorbent surface by elimination water.

4 Scope for Further Research

Based on the present review following future perspectives and challenges can be noted:

1. More agricultural by-products that are widely spread and go to waste without any use should be taken for activated carbon or biochar preparation, and then suitable iron

oxide nanoparticles will be decorated to further increase the adsorption capacity for arsenic and other heavy metals.

2. Some biowaste and mining waste are combined with other materials, so without separating those creating a new bio-adsorbent will be helpful for arsenic removal studies.
3. The applicability of the adsorbent should not be for a single metalloid in the water. It should be applied for simultaneous decontamination of the groups of toxic pollutants such as Pb(II), Cd(II), Cr(VI), F^- , PO_4^{3-} , and toxic dyes.
4. Simulation studies should be done in pollutant removal studies which will increase the adsorption capacity, make easy the system, and be applied in real exercise.
5. Many studies are in lab-scale batch experiments, so it is very necessary to extend them to a specific contaminated area and calculate their potential at the industrial level.

5 Conclusion

The present review describes the use of iron oxide nanoparticles and a composite of iron oxide for arsenic removal in synthetic/natural water. The uses of chitosan, cellulose, activated carbon, biochar, and mining waste are biodegradable, low cost, environmentally friendly, easily available, and green in nature which acts as supporting material for iron oxide nanoparticles. The details of these composite properties and arsenic removal have been presented stepwise. The surface area, particle size, morphology, magnetic properties, and functional groups of the iron oxide-based adsorbent have been analyzed overly. On the other hand, the adsorption capacity, removal rate, and optimum conditions for arsenic removal are given in this study. The adsorption mechanism of arsenic has been point wise analyzed on the adsorbent surface. Overlay, the inner surface complex mechanism including monodentate and bidentate ligand exchange mechanism, and electrostatic attraction are responsible for the arsenic adsorption. The iron oxide with activated carbon,

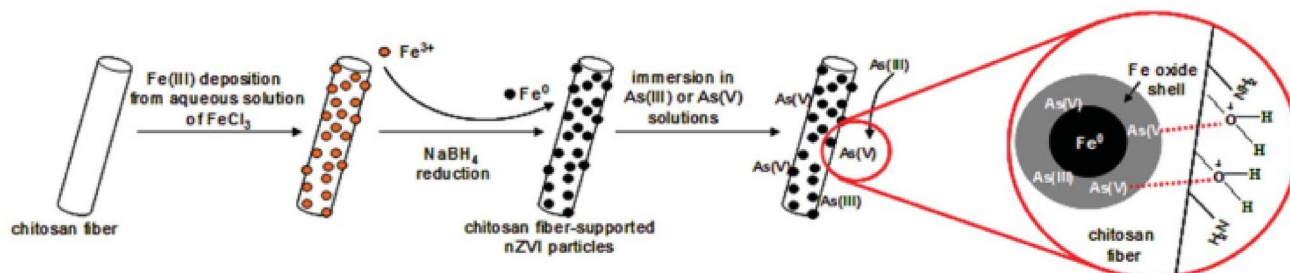


Fig. 7 Experimental steps for the preparation of chitosan fiber-supported nZVI particles and proposed mechanism of As(III)/As(V) sorption [148]

biochar, cellulose, and chitosan shows very good results for arsenic removal. Hence, they can play a crucial role in a bright future for arsenic removal in real wastewater and are highly helpful for rural area people across the world.

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Data availability Data is available on request.

Declarations

Conflict of Interest The authors declare no conflict of interest.

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