



Formaldehyde-free high-strength low-density wood biocomposites via corrugation and self-bonding of wooden cell

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Abstract

In alignment with global sustainable development strategies and the growing demand for green manufacturing practices in engineered wood production, an innovative method has been developed for incorporating hot pressing techniques, minimal energy consumption, and the complete elimination of adhesives. This approach achieved a 100% conversion of waste palm wood into sustainable natural biocomposites suitable for use in structures and furniture. Analysis shows that the biocomposites forms strong internal bonding through mechanical “nail like” nanomaterials, ester bonds, and ether bonds. Unlike conventional furniture materials, which rely on hazardous formaldehyde-based adhesives, this biocomposites boasts an internal bonding strength of 1.652 MPa—four times higher than typical materials. Additionally, it is lightweight, with a density of less than 1 g/cm³, offers excellent friction resistance, and is dense with only 0.67% internal porosity. The composite materials eliminate the use of toxic adhesives, addressing concerns regarding potential harmful emissions from formaldehyde-based VOCs and ensuring higher indoor air quality. This surpasses the performance of existing structures and furniture materials that rely on synthetic adhesives. The method achieves a 100% conversion of waste palm wood into biocomposites, offering a cost-effective and profitable alternative. This provides a novel solution for developing new structural and furniture materials.

Keywords Palm biomass · Biocomposites · Lightweight · Nano-scratch · Micro-CT

1 Introduction

Wood as one of the four major building materials in the world (including cement, masonry, and steel) [1, 2], is an extremely complex biocomposites [3]. Wood has inherent characteristics that differ from other materials: biological characteristics, porous properties, anisotropy and variability, durability, regeneration, and modifiability [4–8]. Although wood has made significant contributions to national economic and city construction, single wood products or wood materials (including various Engineered wood) can no longer meet the needs of human social development. Conventional biocomposites are limited by mechanical processing conditions, and the strength of wood cannot meet the requirements of many engineering structures [9].

Wood composites using formaldehyde-based adhesives have improved in this respect [10, 11], but the long-term release of formaldehyde in wood will cause disease and harm to human health, and increase the risk of indoor environmental pollution [12]. This is due to the fact that chemically synthesized adhesives, exemplified by urea–formaldehyde and phenol–formaldehyde, frequently harbor volatile organic compounds (VOCs), which, upon release, emit formaldehyde into the ambient surroundings [13, 14]. Formaldehyde and VOCs are detrimental, posing the potential to induce adverse health effects, such as irritation, headaches, and nausea. Prolonged exposure to these substances may result in severe consequences for humans, encompassing the development of liver disease and cancer [15, 16]. In order to solve the fundamental problem of formaldehyde release and weather resistance in wooden cells, since Cai Lun improved folk papermaking methods in 105 AD, the dream of wood scientists for 2000 years has been to achieve high-strength self-binding of wooden cells. However, the existing adhesive free bonding technology has problems and bottlenecks such as low

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bonding strength, cumbersome processes, low efficiency, and unclear self-bonding mechanism, which restrict the development of low-cost and efficient green composite processing technology for wood fibers.

The hot pressing method is a commonly employed processing technology for manufacturing composite materials [17, 18]. Within this method, raw materials typically manifest in either flake or granule form, undergoing direct compression into the desired shape amidst conditions of elevated temperature and pressure [19]. Conventional methods typically entail the mixing of raw materials with adhesives, followed by pressing, extrusion, and molding (including injection molding and compression) [20]. Moreover, they necessitate the setup of additional auxiliary equipment, such as specialized heating, exhaust, drying, and cooling systems, which prolongs the duration required for completing the curing and bonding processes of materials [21]. The hot pressing method involves the direct application of high temperature and pressure to compress the raw materials, facilitating the bonding of material molecules to create a robust composite structure. Additionally, processing temperatures and pressures in conventional methods may fluctuate depending on the characteristics of the material and binder, frequently necessitating individual adjustments for each material [21]. However, in the hot pressing method, the processing temperature and pressure remain generally stable and controllable, as they are regulated through the parameter settings of the hot press and remain unaffected by the type of raw material used. In general, compared with conventional methods, the hot pressing method has the advantages of simple process principle, stable processing temperature and pressure, and fast molding speed.

At the same time, alongside the swift progression of technology, elevated demands have been articulated for the wood science community [22–24]. Palm wood is a common tree that originated in China and is currently distributed around the world. Palm trees have high economic value, and Indonesia and Malaysia are one of the few palms' oil producing and exporting countries in the world, with palm cultivation areas reaching 14.8 million hm^2 and 5.4 million hm^2 , respectively [25–27]. To ensure palm oil production, 25-year-old palm trees need to be harvested and renewed [28]. Most of the fallen palm trees are placed in plantations or burned, seriously polluting the environment and causing a large amount of resource waste. The sustainable use of palm trees is an important way to protect the environment and promote economic development. In Malaysia, workers spin and cut some harvested palm trees into veneer for the production of plywood [29, 30]. Due to the lower density of the inner layer of palm trees compared to the outer layer, and their soft tissue, improving the quality and

compressive strength of plywood has become a major technical challenge.

Therefore, a series of experiments were conducted to explore the conversion of palm wood, a raw material, into biocomposites. Subsequently, a low-energy one-step production process using hot pressing as a potential technique. In contrast to conventional furniture materials dependent on formaldehyde-based adhesives, this biocomposites eschews toxic adhesives, thereby circumventing the issue of deleterious formaldehyde emissions. Concurrently, it transcends the conventional theoretical framework of adhesive-free wood bonding, thus achieving an original theoretical innovation in formaldehyde-free wood composite materials and offering a novel approach to advancing the development of new structural and furniture materials.

2 Materials and methods

2.1 Preparation of biocomposites

The palm wood obtained from Terengganu University in Malaysia was dried and crushed into powder (particle size: 0.25–0.425 mm) to imitate the wood waste generated by the furniture industry. Before conversion, all waste underwent a 12-h drying phase at 55 °C, followed by storage at 5 °C. Palm sawdust was pressed using a hot press at 147 MPa [31] for 10 to 35 min to produce biocomposites (size: 50 mm long \times 50 mm wide \times 7 mm thick). Biocomposites with varied mechanical properties (Fig. 1 and Table S1) were acquired through the manipulation of temperature, pressure, and pressing duration, denoted as Y1, Y2, Y3, Y4, Y5, Y6, Y7, Y8, Y9, and Y10, respectively. Through using various physical and chemical analysis methods, three tests were conducted on each biocomposites sample to determine and describe the reactivity, chemical transformation, and structural evolution of wood fiber biocomposites, thereby revealing their self-adhesion mechanism.

2.2 Internal bonding strength (IBS) and compressive strength

According to the Chinese national standard *GB/T 17657–2013*, the IBS (internal bonding strength) of the samples was determined [31]. The IBS was measured by subjecting the 50 mm \times 50 mm \times 7 mm sample to tension until failure. The IBS was computed by dividing the maximum failure tension by the vertical surface area. The sample underwent testing, positioned on a chuck and affixed with epoxy resin, enabling the measurement and assessment of the bonding quality between fibers within the sample. Low IBS values may lead to sample stratification, while a higher

IBS indicates better resistance to temperature variations and a reduced risk of cracking [32, 33].

The compressive strength of the samples was measured using a cement flexural and compressive constant stress testing machine (Model HYE-300B). The load was applied uniformly at a rate of $1.1 \text{ kN/s} \pm 100 \text{ N/s}$ on the side surface of a half-prism until failure occurred. The compressive strength R_c (MPa) of the sample was calculated according to Eq. (1). Extended Data Table S2 shows the compressive strength test results of the samples.

$$R_c = \frac{F_c}{A} \quad (1)$$

where: F_c — the maximum load at failure (kN), A — the area of the compressed section (m^2).

2.3 FTIR and thermogravimetric analyses

Extraction of samples was performed utilizing KBr discs incorporating 1.00% finely ground specimens. The FT-IR spectra for each sample were acquired employing a Thermo Fisher Scientific iS10 spectrophotometer, with a spectral range of $500\text{--}4000 \text{ cm}^{-1}$.

The powder of the sample was also analyzed by thermogravimetric analyzer (TGA-Q500, TA Instruments) to investigate the material content (such as water, volatile matter), thermal stability, reaction kinetics and decomposition stage of the sample [31]. The nitrogen release rate is 60 mL/min . The TG temperature program starts at $30 \text{ }^\circ\text{C}$ and increases at a heating rate of $25 \text{ }^\circ\text{C/min}$ to $750 \text{ }^\circ\text{C}$.

2.4 Nanoindentation and nanoscratch analyses

The samples were tested using the TI-980 nanoindentation instrument (Bruker, United States). The sensor used a 10-mN range two-dimensional mechanical sensor, with a maximum loading force of 8 mN and a loading time of

5 s. The load was maintained for 2 s and unloaded for 5 s. The testing method for the sample is 3×4 arrays, testing 12 points per sample, with a spacing of $100 \text{ }\mu\text{m}$ [34].

Use a 3D scratch tester (Rtec, China) to perform scratch testing on the sample. In the 3D scratch test, a tip scratch head with a radius of $0.1 \text{ }\mu\text{m}$ was used, with a loading force of 10 N, a running speed of 0.08 mm/s , and a scratch length of 3 mm [35]. A $20 \times$ confocal lens was used to image the scratch morphology.

2.5 Additional analyses

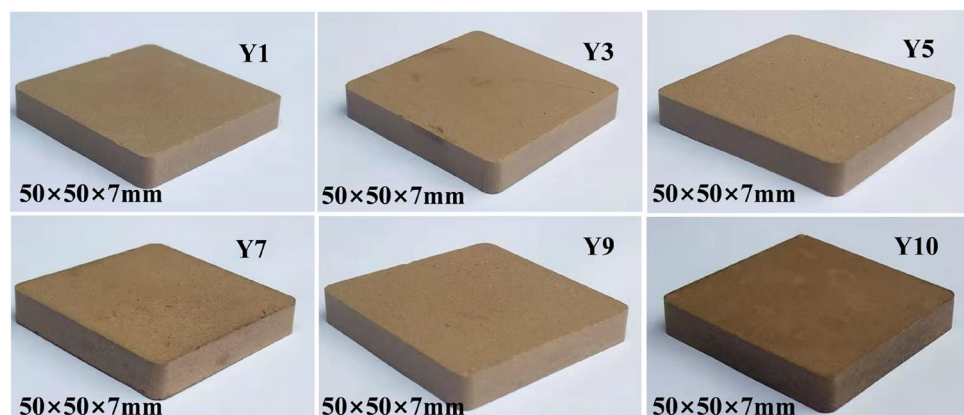
The anti-wear characteristics of the sample were tested using a friction tester (Rtec, China) [36, 37]. The testing instrument uses a grinding ball with a radius of 9.3 mm, a loading force of 5 N, a friction frequency of 1 HZ, a friction stroke of 10 mm, and a friction time of 10 min. A $20 \times$ confocal lens is used to image the morphology of the sample after friction.

Using XRadia Context 3D Micro CT (ZEISS, Germany) to scan and image the samples, observe the internal microstructure of the biocomposites. The sample was scanned and imaged using a voxel resolution of $22.0 \text{ }\mu\text{m}$, with a field of view range of $66.0 \times 44.0 \text{ mm}$, a scanning energy of 60 kV, a scanning power of 5 W, and a scanning time of 0.5–1 h. Use a $3.0\text{-}\mu\text{m}$ voxel resolution to perform high-resolution, high contrast scanning imaging of the local area of the sample. The field of view range is $9.0 \times 6.0 \text{ mm}$, with a scanning energy of 60 kV, a scanning power of 5 W, and a scanning time of 0.8 h.

The surface morphology of the biocomposites was analyzed using an S-3400N scanning electron microscope (SEM) (Hitachi Ltd., Japan).

Formaldehyde emission of biocomposites was measured in accordance with the Chinese national standard “Test methods of evaluating the properties of wood-based panels and surface decorated wood-based panels” (GB/T 17657–1999).

Fig. 1 Palm biocomposites (size: 50 mm long \times 50 mm wide \times 7 mm thick)



2.6 Statistical analyses

Graph Pad Prism9.0 software was used for the graphical presentations, while a paired Student's *t*-test was used to test for group references (significant level set to $p < 0.05$).

3 Results and discussion

3.1 Physical and chemical analyses

The sample biocomposites was tested using FT-IR and Fig. 2a shows several clear spectral peaks. The absorption peaks of each spectrogram reflect the types of functional groups contained in the substance and its chemical environment [38, 39]. In the 3390 cm^{-1} band, the O–H bond tensile vibration of aromatic and Aliphatic compound is mainly formed, and in the 1030 cm^{-1} band, the C–O bond tensile vibration of aldehydes, esters and carbohydrates is mainly formed. The peak absorption at 1610 cm^{-1} is caused by the stretching vibration of the carbon–carbon double bond (C = C) in alkanes. The peak absorption at 1740 cm^{-1} is attributed to the stretching vibration of the carbonyl group (C = O) present in aldehydes, ketones, acids, and esters. The ability to improve fiber self-adhesion is further evidenced by the strength of absorption peaks observed at the FT-IR transmission peak, encompassing various functional groups including O–H bonds, aldehydes, carbohydrates, ketones, alkanes, and ester C = O bonds [40]. Through Chemical bonds, including hydrogen bonds, ester bonds, and ether bonds, more robust chemical connections are established between compounds in biocomposites [31].

In a high-purity nitrogen atmosphere, palm wood undergoes a series of processes, including oxidation, dehydration, condensation, reduction, and decomposition, ultimately leading to weight loss [41]. The thermal weight loss curves for various biocomposites are depicted in Fig. 2b and Fig. S1. During the process of heating to $750\text{ }^{\circ}\text{C}$ at room temperature, the thermal weight loss curve of the sample biocomposites is divided into three stages. In the first stage of weight loss at $30\text{--}235\text{ }^{\circ}\text{C}$, the total weight loss rate of the sample biocomposites is around 2%, mainly due to the evaporation of some water vapor and low boiling point small molecules. When the temperature rises to $250\text{ }^{\circ}\text{C}$, the weight of the sample begins to sharply decrease, and the thermal weight loss rate reaches its maximum at $347\text{ }^{\circ}\text{C}$. This stage is mainly due to the cleavage of the heavy components (such as sugars, phenols, esters, and carbohydrates) in the biocomposites. At $500\text{ }^{\circ}\text{C}$, the weight loss rate of the sample reaches about 28%. With the ongoing increase in temperature, the weight of the sample biocomposites exhibited only a marginal decrease. When

the final temperature reached $750\text{ }^{\circ}\text{C}$, the weight of various sample biocomposites remained at 21–27%, among which the Y10 sample biocomposites with paraffin added had the most weight loss, and the final weight loss rate reached 79%.

Similarly, SEM analysis shows that the fiber structure is tightly bound in the hinge network (Fig. 2c). In addition, all broken and fragmented cell wall fibers will cause the intertwined fragments between the fibers to completely close (Fig. 2c and d). In summary, this creates a biocomposites with an intricate interface structure formed by interlacing networks of fibers. Micro CT analysis (Fig. 2d) confirmed that the Tight junction fiber network formed a dense interface structure in the biocomposites [42]. Figure 2c shows the fragmentation of cells, which generates various subminiature substances, enhances physical and chemical interconnections, thereby improving densification.

3.2 Strength and hardness

Internal Bond Strength (IBS) tests were conducted on each palm biocomposite, and the corresponding test results are illustrated in Fig. 3b. According to the results of Y1, Y2, Y3, and Y4, when the insulation time is 25 min, the IBS value of palm biocomposites obtained at $170\text{ }^{\circ}\text{C}$ is the highest, reaching 1.308 MPa (Fig. 3c). During the process of $150\text{--}170\text{ }^{\circ}\text{C}$, as the temperature continues to rise, the IBS value gradually increases. Comparing the results of Y3, Y5, Y6, Y7, Y8, and Y9 samples, it was found that under the same temperature ($170\text{ }^{\circ}\text{C}$) conditions, the IBS value of palm biocomposites obtained at a holding time of 25 min was the highest 1.308 MPa (Fig. 3d). The IBS value of palm Y10 with the addition of paraffin increased compared to the corresponding value of the control group, reach 1.652 MPa. The internal bond strength of palm biocomposites is also superior to that of fiber biocomposites reported by Shi et al. (1.52 MPa) [43]. The addition of paraffin made the wood fibers tightly bonded, enhanced their surface hardness and internal bonding strength, and made them more wear-resistant and wear-resistant. This process condition provides reference and reference for the production and processing of palm biocomposites.

Based on the aforementioned considerations, it is deduced that the selection of an appropriate hot-pressing temperature ($170\text{ }^{\circ}\text{C}$) and holding time (25 min) is pivotal in the fabrication of palm biocomposites. Utilizing hot-pressing technology proves effective in enhancing the interface compatibility between the matrix and filler, facilitating uniform filler dispersion, and elevating the overall performance of the composite material. In comparison with both Chinese National Standards and American National Standards for Medium Density Fiberboard (MDF) and Particleboard (PB) in China,

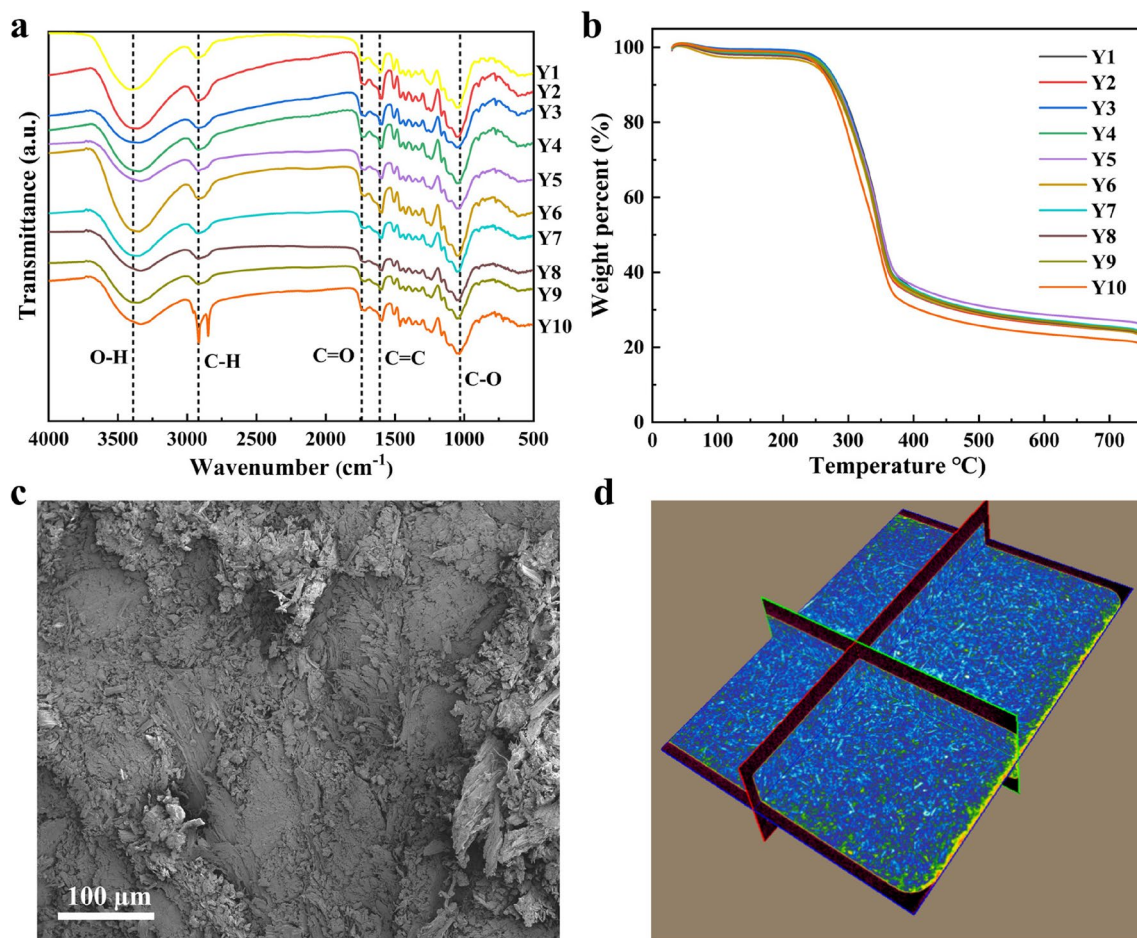


Fig. 2 **a** FTIR Spectrogram of palm biocomposites. **b** Thermal weight loss curve of palm biocomposites. **c** SEM micrographs of palm biocomposites (Y10). **d** The micro-CT (voxel resolution of 22 μm) of palm biocomposites (Y1)

this biological composite demonstrates significantly elevated adhesive strength, ranging between 0.648 and 1.652 MPa. This range is 2 to 4 times greater than the specified values for commercial MDF (U.S., ≥ 0.8 MPa; CN, ≥ 0.6 MPa) and PB (U.S., ≥ 0.6 MPa; CN, ≥ 0.4 MPa) outlined in the national standards [31]. The creation of shorter and finer wood particles led to improved packing efficiency, allowing for greater IB strength. The smaller size of the particles enabled them to fit together more snugly, much like the way puzzle pieces fit together to create a completed picture. This intimate contact between the particles translated into increased IB strength, as the pieces of wood were held together more securely, reminiscent of the way bricks in a wall are tightly mortared to provide support and stability.

In addition, compressive strength tests on palm biocomposites and found that the compressive strength values of Y1, Y2, Y3, and Y4 biocomposites were all greater than 90 MPa, with a compression rate of 11–15% (Table S2). The specific strength value (strength/density) of Y1 biocomposites is the highest, at 177.88 $\text{MPa}\cdot\text{cm}^3/\text{g}$ (Table S3), and

similar to aluminum alloy materials ($244 \text{ MPa}\cdot\text{cm}^3/\text{g}$) [44] commonly used in industry. The maximum compressive strength value of palm biocomposites (139.638 MPa) is 2 times the compressive strength specified for bridge bearings (70 MPa) as seen in Fig. 3a [45]. This groundbreaking development means that biocomposites have become an important breakthrough in replacing concrete and pine in structural applications such as architecture and furniture (Fig. 3g). This breakthrough technology not only breaks the limitations of traditional wood, but also provides strong support for sustainable development in the future.

Nanoindentation testing was conducted to evaluate the micro and nanomechanical properties of biocomposites. The exquisite nanoindentation technique carries the distinction of high resolution and depth sensing abilities, establishing it as a formidable toolbox for gauging the nanomechanical attributes of minute materials. This paves a path towards exploring the mechanical behavior of materials at the nanoscale, deciphering the complex interactions that occur between materials and their environment

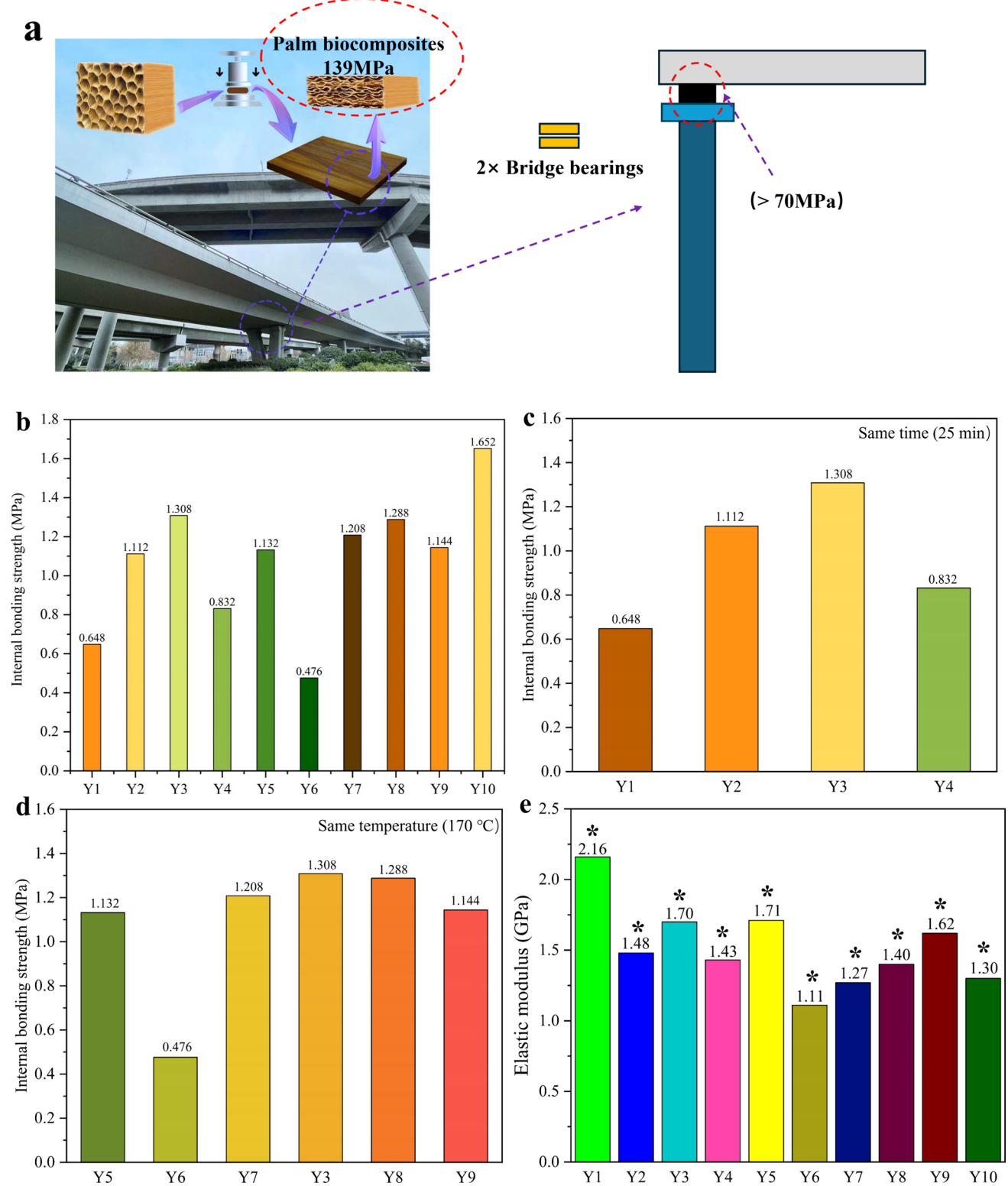


Fig. 3 **a** Compressive strength of bridge bearings and palm biocomposites. **b** Internal bonding strength of palm biocomposites. **c** Internal bonding strength of palm biocomposites obtained under the same insulation time. **d** Internal bonding strength of palm biocomposites obtained under the same temperature. **e** Elastic Modulus of palm biocomposites. **f** Hardness of palm biocomposites. **g** The compressive

strength of various common building materials. **h** Surface morphology images of palm biocomposites (Y10) after nanoindentation. **i** Surface morphology images of palm biocomposites (Y10) after nanoindentation, obtained using a profilometer. “*” Significance value < 0.05

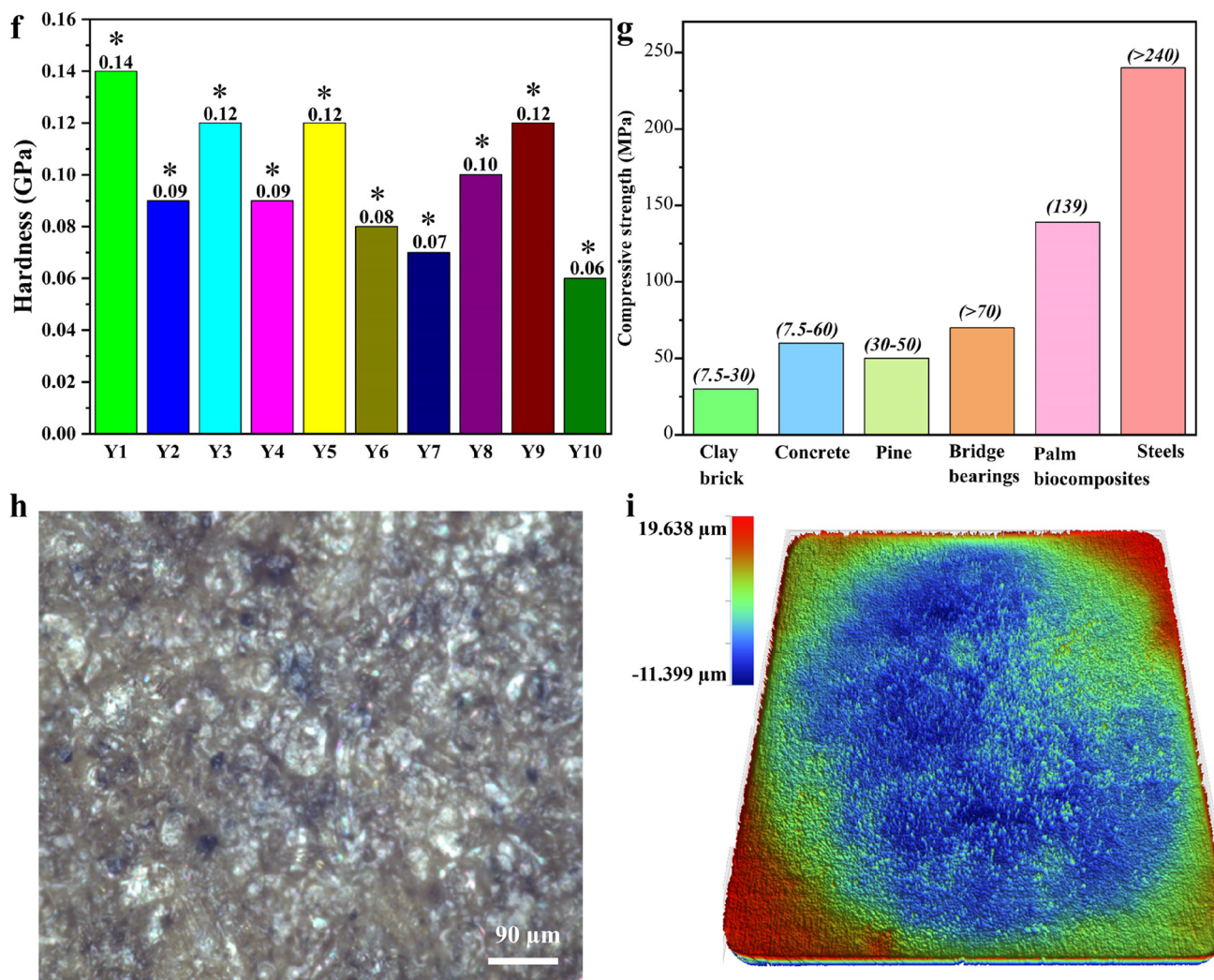


Fig. 3 (continued)

[46, 47]. The test results indicate that as the indentation depth increases, the elastic modulus (Fig. 3e) and hardness (Fig. 3f) of the material also change. From the figure, it can be seen that the Y1 sample has the highest modulus and hardness, reaching 2.16 GPa and 0.14 GPa, the scratch hardness is 7 times that of natural wood (0.02 GPa) [44]. The modulus and hardness of the Y3 sample with high internal adhesive strength are 1.70 GPa and 0.12 GPa, respectively. The Y10 sample with added paraffin has relatively low modulus and hardness, which are 1.30 GPa and 0.06 GPa, respectively. Figure 3h and 3i shows the surface morphology of palm biocomposites after nanoindentation. When heating and cooling palm wood biocomposites, a dense interface layer is formed, forming high adhesive strength, resulting in the interwoven cell walls leading to a high density of natural polymer biocomposites [48]. Therefore, this method enhances both bonding strength and interwoven cell walls, collectively contributing to the

method of a high-strength interface layer in natural fiber biocomposites.

3.3 Friction resistance and stability

To further investigate the hardness and internal structure of palm biocomposites, nano scratch tests were conducted on each biocomposite specimen. The study of adhesion between fiber and matrix is often achieved through micromechanical tests. Nano scratch test is an effective means to evaluate Strength of materials properties, and has a wide range of applications in the research of structural material surface [49, 50]. In the experiment, a scratch head with a radius of 0.1 μm was used to test the scratches on the biocomposites under a loading force of 10 N, and a 20× confocal lens was used to image the scratch morphology. Figure 4a and b shows the 3D interface of palm wood under nanoscratching, with each dark color representing different depths. The

maximum scratch depth of each palm biocomposites under the same conditions is shown in Fig. 4d, with Y1 and Y3 having the shallowest maximum scratch depths of 164.9 and 162.0 μm , respectively. Y5 has the maximum scratch depth of 284.9 μm . Under the same force, Y3 has the smallest deformation amplitude and Y5 has the largest change. The maximum scratch depth of Y10 with added paraffin is 183.7 μm .

Plastic resistance (PR) characterizes the ability of a scratched material to resist permanent deformation [51], calculated by dividing the normal force (F_n) by the residual depth (R_d): $PR = \frac{F_n}{R_d}$. Materials with higher PR values have better resistance to permanent scratching. Tests have shown that palm biocomposites Y1 and Y3 have higher PR values and better friction resistance.

In addition to nanoindentation and nanoscratch tests, friction and wear tests to study and compare the surface roughness of palm biocomposites. In the experiment, a grinding ball with a radius of 9.3 mm was used to rub the sample for 10 min under a loading force of 5 N, a frequency of 1 HZ, and a friction condition of 10 mm. The morphology of the friction sample was imaged (Fig. 4c). After 10 min of friction testing under a force of 5 N, the maximum friction

depth of Y1 sample is 279.8 μm , Y2 sample is 246.3 μm , Y3 sample is 331.6 μm , while Y7 sample is 48.4 μm , Y8 sample is 56.7 μm , Y9 sample is 24.0 μm , and Y10 sample is 16.6 μm (Fig. 4e). Among them, the Y10 sample has the smallest surface friction depth and the highest smoothness, which also reflects that after adding paraffin, the roughness of the formed biocomposites is reduced, making the surface smoother and more wear-resistant. Secondly, the maximum friction depth of Y7, Y8, and Y9 samples is also within 100 μm , and the wear resistance of the samples is better than other groups.

3.4 Lightweight, compact and environmentally friendly

The samples were scanned and imaged using recon resolutions of 7.5 μm and 22.0 μm , respectively, to obtain virtual slices of the Y1 biocomposites (Fig. 5a and b). From the displayed results, it can be seen the distribution of high-density substances, pores, and wood fibers in the biocomposites. Simultaneously, Dragonfly software was used to directly render the Y1 biocomposites sample to obtain its rendered image (Fig. 5c and d), with different colors representing

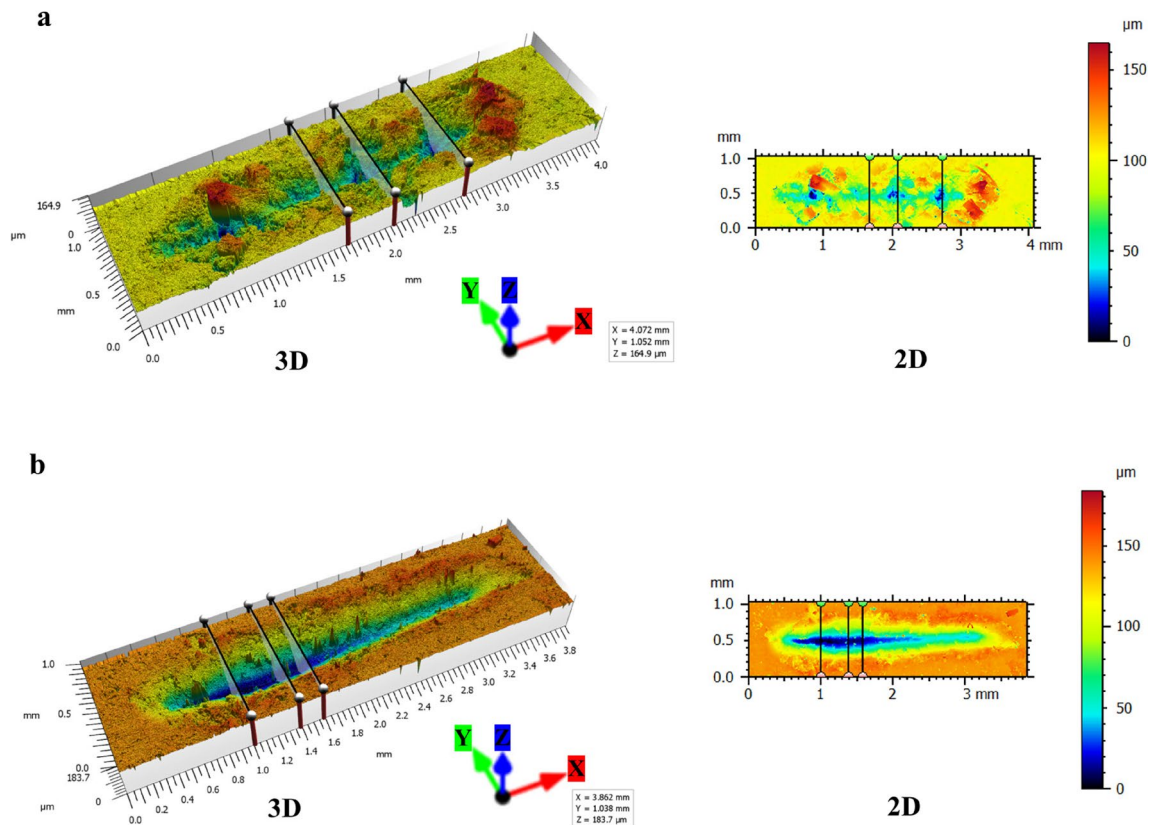


Fig. 4 **a** 3D imaging of palm biocomposites (Y1) by nano-scratch. **b** 3D imaging of palm biocomposites (Y10) by nano-scratch. **c** 3D imaging of palm biocomposites (Y1) by abrasion test. **d** Maximum

depth of palm biocomposites by nano-scratch. **e** Maximum depth of palm biocomposites by abrasion test

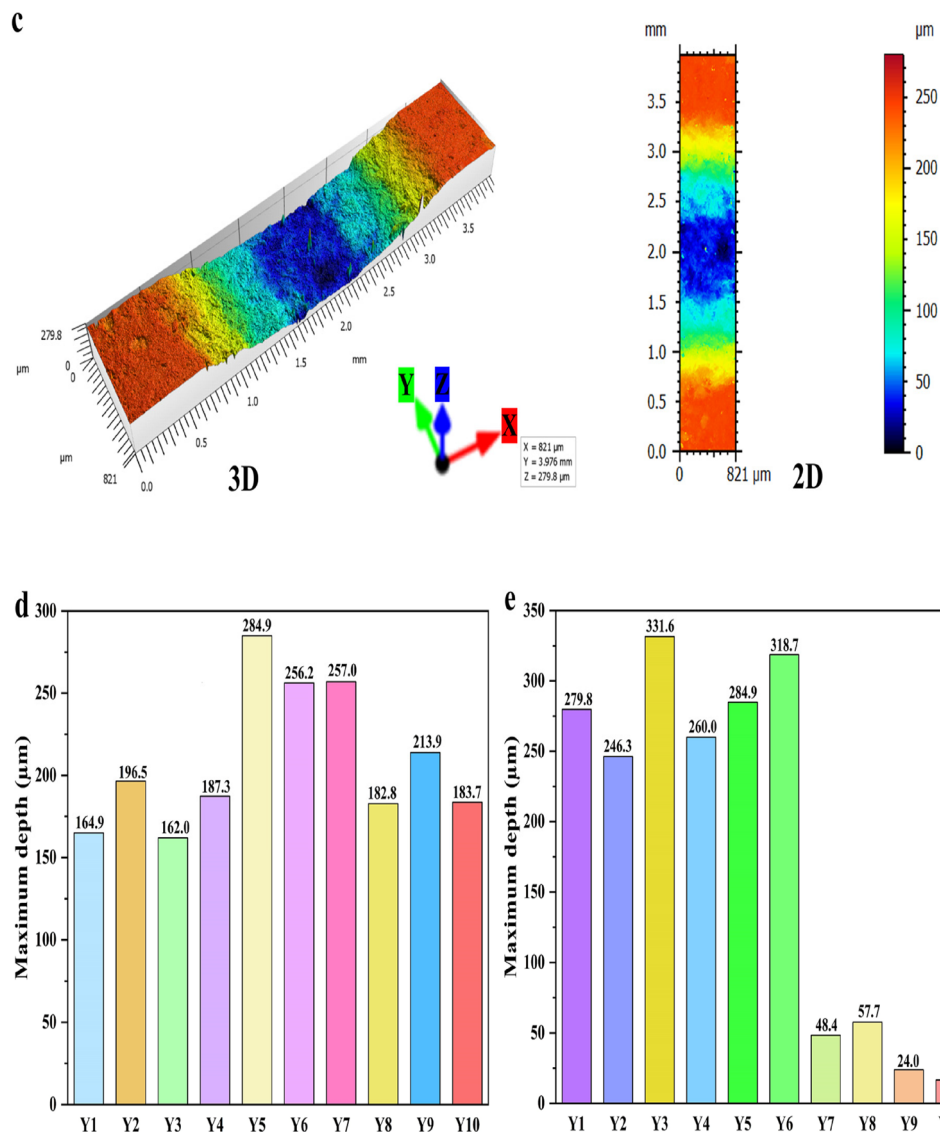


Fig. 4 (continued)

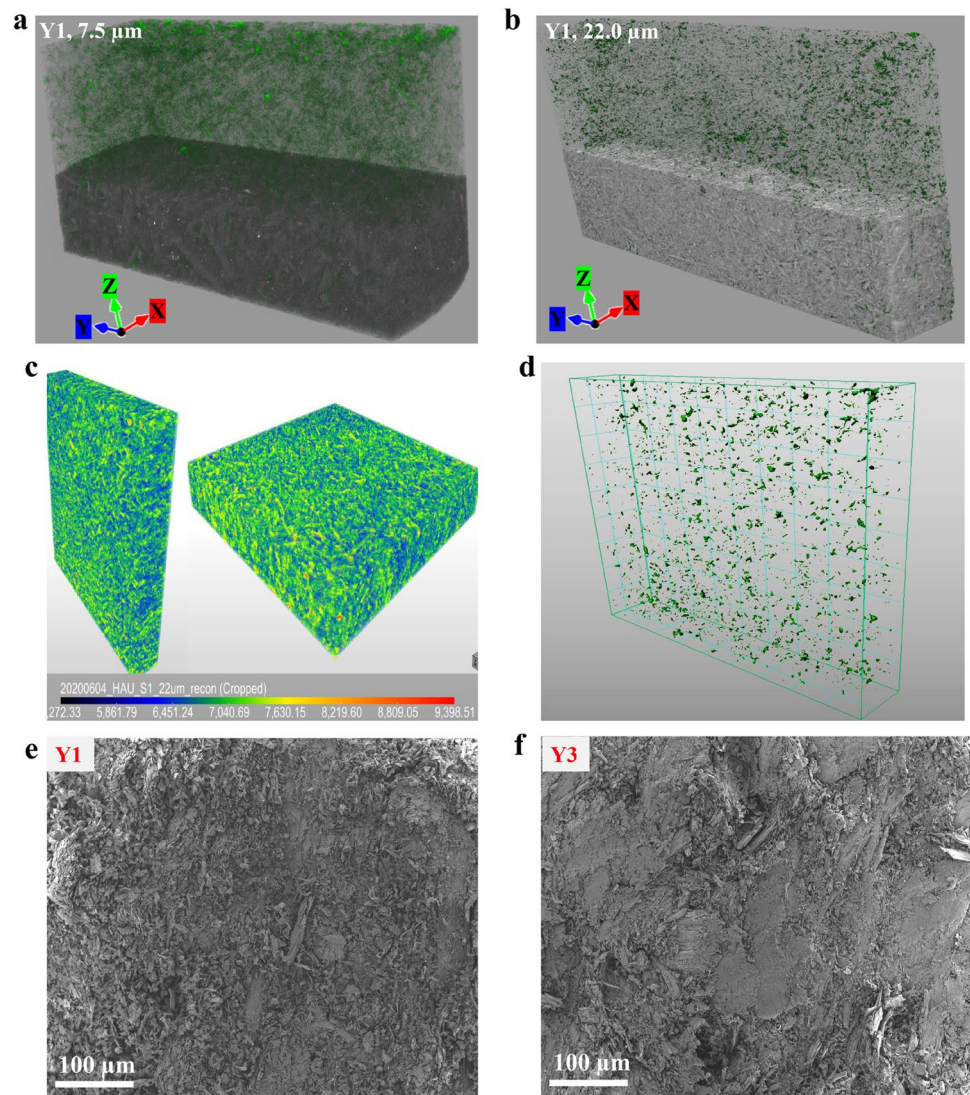
different grayscale values. The morphology of the pores inside the sample plate and their spatial distribution in three-dimensional space are visually displayed. By comparing the volume of pores with that of biocomposites, it was calculated that at a resolution of 22.0 μm recon, the pore volume of Y1 biocomposites in the selected area accounted for 0.67% of the total volume of the selected area. At the same time, under identical conditions, scanning was conducted on Y3 and Y10 biocomposites with high internal bonding strength. According to the results, the porosity of Y1 biocomposites is only 0.67%, while that of Y3 biocomposites is 0.75%. However, due to the addition of paraffin, the porosity of Y10 biocomposites is relatively large, reaching 1.42%. The microscopic CT analysis confirms that the Tight junction fiber network forms a dense interface structure in the

palm composite. Figure 5e and f shows the fragmentation of wood cells, which produce various subminiature bonding substances, enhancing the physical and chemical interconnections inside the palm biocomposites, thereby improving its densification.

The formaldehyde release from palm biocomposites was measured, and according to the measurement results (Table S4), the moisture content of the sample is 6.1%, and the formaldehyde release of the Y1-Y10 sample is between 0.9 and 1.3 mg/100 g. Among them, the formaldehyde release of the Y3 sample is the lowest at 0.9 mg/100 g, and the formaldehyde release of the Y10 sample is the highest at 1.3 mg/100 g.

According to the Chinese national standard “Indoor decorating and refurbishing materials Limit of formaldehyde

Fig. 5 **a** The recon resolution of palm biocomposites (Y1) scanned by Micro-CT is 7.5 μm . **b** The recon resolution of palm biocomposites (Y1) scanned by Micro-CT is 22.0 μm . **c** 3D rendering of palm biocomposites (Y1) in 22.0 μm recon. **d** Micro-CT photos of biocomposites (Y1) in 22.0 μm recon. **e** SEM micrographs of biocomposites (Y1). **f** SEM micrographs of biocomposites (Y3)



emission of wood based panels and finishing products” (*GB 18580–2001*), the formaldehyde emission of Class A solid wood composite flooring is less than or equal to 9 mg/100 g; the formaldehyde emission of Class B solid wood composite flooring is equivalent to 9–40 mg/100 g [52, 53], and palm biocomposites fully meet the standards for being used as solid wood composite flooring. According to the measurement results of Peng et al. [54] on formaldehyde emission from several kinds of wood, the formaldehyde emission from *Pinus massoniana* is 2.55 mg/100 g, that from *Fraxinus mandshurica* is 3.28 mg/100 g, that from *Cunninghamia lanceolata* is 2.01 mg/100 g, that from *Populus L.* is 1.17 mg/100 g, and that from *Eucalyptus salicifolia* is 1.43 mg/100 g. Compared with the reported results, the formaldehyde emission of palm biocomposites is lower than that of *Pinus massoniana*, *Fraxinus mandshurica*, *Cunninghamia lanceolata*, and *Liueucalyptus*, and close to that of natural *Populus*.

This innovative biocomposites not only represents a groundbreaking technological advancement but also presents sustainability advantages, characterized by low energy consumption and the absence of toxic adhesives. It’s a heraldic achievement in the green materials’ field that points towards a healthier, more environmentally sound future. The biomaterial’s eco-friendliness goes beyond its production process. It also boasts excellent mechanical properties—a key factor for various engineering applications—such as, biocomposites have shown significant improvements in internal bond strength, material density, and resistance to pressure, friction, and environmental influences. This statement highlights an innovative approach utilizing hot pressing, characterized by minimal energy consumption and the absence of adhesives, to transform palm wood biomass into natural and sustainable biocomposites. These biocomposites offer significant potential in the furniture and construction industries, aligning well with contemporary societal preferences

for environmentally friendly materials. This method not only underscores sustainability but also emphasizes the importance of reducing energy consumption and eliminating harmful chemicals, contributing to a greener future. Moreover, by repurposing palm wood biomass, it addresses both environmental concerns and the growing demand for eco-friendly alternatives in various sectors, thus reflecting a promising avenue for sustainable material innovation.

4 Conclusions

An innovative approach has been introduced for the conversion of palm biocomposites into natural and sustainable biocomposites, deemed suitable for application in construction and furniture manufacturing. This method effectively integrates hot pressing with minimal energy consumption, thereby completely eliminating the need for adhesives. The result analyses have shown that the exceptional internal strength of this biocomposites results from a combination of mechanical interlocking and the formation of hydrogen, ether, and ester bonds, which function synergistically like a meticulously assembled puzzle. Compared to conventional materials that rely on formaldehyde-based adhesives, this biocomposites demonstrates enhancements in internal bonding strength—increasing fourfold—while maintaining a low density, which enhances its resilience and environmental friendliness. Additionally, the absence of toxic adhesives not only mitigates concerns about harmful formaldehyde VOCs emissions but also significantly improves indoor air quality. This approach, which combines fragment riveting and cell folding, represents a breakthrough in adhesive-free bonding for wood, achieving original theoretical innovations in formaldehyde-free wood composite materials and offering a novel solution for the development of new structural and furniture materials.

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