



An accurate, high-speed, portable bifunctional electrical detector for COVID-19

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ABSTRACT Coronavirus disease 2019 (COVID-19), caused by SARS-CoV-2, has rapidly spread and caused a severe global pandemic. Because no specific drugs are available for COVID-19 and few vaccines are available for SARS-CoV-2, accurate and rapid diagnosis of COVID-19 has been the most crucial measure to control this pandemic. Here, we developed a portable bifunctional electrical detector based on graphene field-effect transistors for SARS-CoV-2 through either nucleic acid hybridization or antigen-antibody protein interaction, with ultra-low limits of detection of ~ 0.1 and ~ 1 fg mL⁻¹ in phosphate buffer saline, respectively. We validated our method by assessment of RNA extracts from the oropharyngeal swabs of ten COVID-19 patients and eight healthy subjects, and the IgM/IgG antibodies from serum specimens of six COVID-19 patients and three healthy subjects. Here we show that the diagnostic results are in excellent agreement with the findings of polymerase chain reaction-based optical methods; they also exhibit rapid detection speed (~ 10 min for nucleic acid detection and ~ 5 min for immunoassay). Therefore, our assay provides an efficient, accurate tool for high-throughput point-of-care testing.

Keywords: COVID-19, biosensor, nucleic acid detection, immunoassay, point-of-care testing

INTRODUCTION

In the past 20 years, humans have suffered several serious epidemics from emerging viruses, such as SARS, swine flu, Ebola, MERS and (most recently) SARS-CoV-2 [1–3]. During each epidemic, an accurate, rapid, and accessible molecular diagnostic test is highly essential for the control and prevention of viral diseases. In particular, a cluster of cases of pneumonia resulting from a new coronavirus, SARS-CoV-2, was initially described in late 2019 (or earlier); this disease was later named as COVID-19 [4]. During the epidemic, the Chinese government and people implemented approaches to aid in the diagnosis, isolation, and treatment of affected patients; they also strictly restricted the flow of people, which constituted the so-called “people’s war”. Therefore, the outbreak and spread of COVID-19 in China was largely contained after approximately 1.5 months. However, in most other countries, the disease has spread rapidly since late February 2020; on 12 March 2020, the World Health Organization recognized the COVID-19 as a global pandemic [5]. As of 30 August 2020, over 24 million confirmed cases and 838,924 deaths had been reported worldwide [6]. Thus far, effective specific drugs and vaccines specific for COVID-19 are unavailable; accordingly, rapid and accurate early detection of the COVID-19 causative virus (i.e.,

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SARS-CoV-2) is expected to aid in controlling the ongoing pandemic and support resumption of normal life and economic conditions.

SARS-CoV-2 mainly consists of a single-stranded RNA genome (approximately 30,000 nucleotides) and four structural proteins that include the spike surface glycoprotein (S), envelope protein (E), membrane protein (M), and nucleocapsid protein (N). There are generally two strategies for identification of the virus: (1) detection of viral RNA and (2) detection of host antibodies. Currently, the presence of specific viral RNA sequences in the patient samples is considered definitive proof of COVID-19, while immunodetection is regarded as a useful auxiliary technique. Reverse transcriptase quantitative polymerase chain reaction (RT-qPCR) is the primary method for detection of nucleic acid-based genetic sequences from any organism, including SARS-CoV-2 [7–11]. Because of the labor-intensive sample preparation, which must be conducted in a biological laboratory by professionals using specialized instruments, the typical turn-around time of the RT-qPCR method is longer than 24 h. The complex sample preparation steps prior to testing might also reduce clinical sensitivity (the percentage of actual positive individuals identified as positive individuals), resulting in false negative results [12,13]. In addition, point-of-care detection is not accessible by the RT-qPCR method. Hence, the development of highly sensitive and specific nucleic acid testing methods, in combination with the characters of fewer sample preparation steps and the use of a portable detection instrument, is an extremely attractive prospect for accurate and rapid diagnosis of COVID-19.

Biosensors, as an alternative and reliable solution for the analysis of biomolecules, has been drawing considerable research interest [14–17]. Among different kinds of optical and electrical biosensing technologies currently available, field-effect transistor (FET)-based biosensors are particularly attractive because of their remarkable features, including no fluorescent labeling requirement, highly sensitive detection, mass-production capability and low cost. The types of the FET biosensors mainly include silicon nanowire FETs, graphene/carbon nanotube FETs, organic FETs, compound-semiconductor FETs and ion sensitive FETs [18–20]. Graphene materials possess excellent properties of large surface area, high electronic conductivity and high carrier mobility, making the graphene FET an ideal platform for biomolecule detection [14,21].

In the present study, we developed an unprecedented accurate, rapid, and portable electrical detector based on

the use of graphene FETs (G-FETs) for detection of RNA from COVID-19 patients. As shown in Fig. 1, the detection system mainly consists of two parts: a plug-and-play packaged biosensor chip and a home-developed electrical measurement machine. Each packaged chip contains ten G-FETs; specific ss-DNA probes are immobilized onto the graphene surface *via* π - π stacking of a typical linker (1-pyrenebutyric acid *N*-hydroxysuccinimide ester, PBASE) [22–24]. The unique feature of this method is that the extent of hybridization between the ss-DNA probe and viral RNA can be directly converted to the current change of graphene channels without repetition of the PCR process. Our G-FET biosensors exhibited excellent performance for the detection of the RNA-dependent RNA polymerase (RdRp) gene target of SARS-CoV-2 with an ultra-low limit of detection (LOD) of $\sim 0.1 \text{ fg mL}^{-1}$. Furthermore, we validated our method using clinical samples collected from ten patients with COVID-19 infection and eight healthy individuals, and the testing results were in full agreement with those of PCR-based optical methods. The entire process, precluding the extraction of detection targets from oropharyngeal swabs, requires approximately 10 min. Because it does not involve time-consuming PCR step nor expensive instruments, our detection system has the potential to enable massive point-of-care testing of COVID-19, outside of specialized diagnostic laboratories, with the advantage of high sensitivity and low cost.

Notably, false negative results are inevitable in the course of nucleic acid testing; thus, the use of immunodetection as an auxiliary technique is important in the diagnosis of COVID-19 patients, especially those with suspected diseases [25–28]. By replacing the ss-DNA probe with a SARS-CoV-2 antigen protein, our detection system can also detect SARS-CoV-2 IgM and IgG antibodies with an ultra-low LOD of $\sim 1 \text{ fg mL}^{-1}$. Immunoassays of serum specimens of six COVID-19 patients and three healthy subjects matched excellently with those of PCR-based optical methods.

EXPERIMENTAL SECTION

Fabrication of graphene-based biosensor chips

High-quality chemical vapor deposition (CVD)-grown monolayer graphene on 4-inch silicon dioxide (300 nm)/Si wafer substrate was purchased from Jiangsu XFNANO Materials Tech Co., Ltd. The source/drain metal electrode arrays (8 nm Cr/600 nm Au) were patterned by photolithography and thermal evaporation, followed by electron beam evaporation of 40-nm-thick SiO₂ to passivate

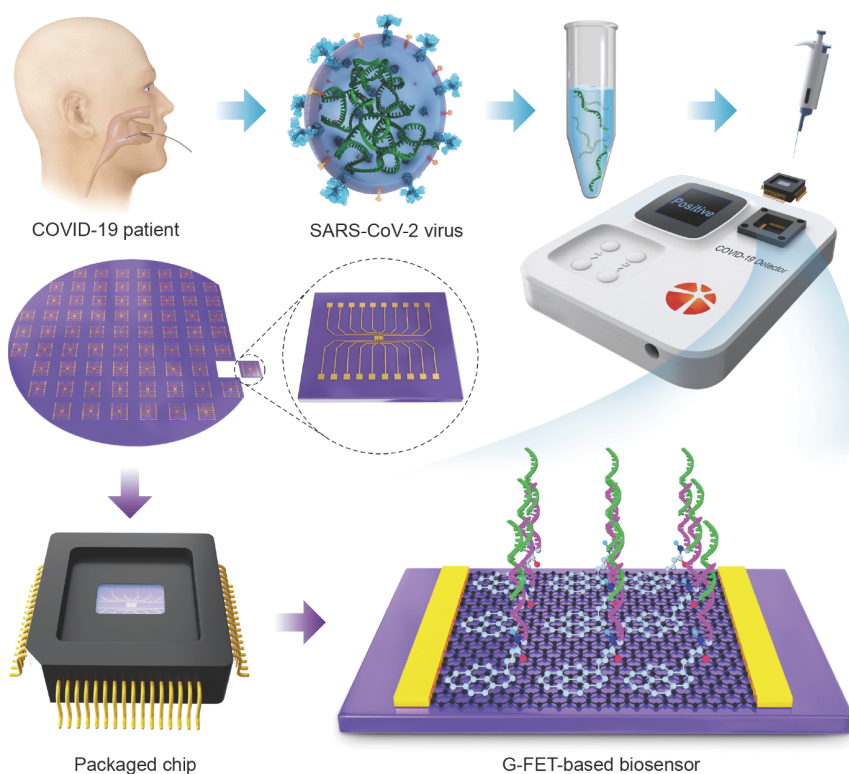


Figure 1 Schematic diagram of the operation procedure of our G-FET-based biosensing system for COVID-19. Top left: Extraction of viral RNA. Bottom left: from Si wafer to plug-and-play graphene packaged chips. Top right: home-developed portable electrical detector. Bottom right: illustration of the ss-DNA probe immobilization onto graphene using a typical PBASE linker, followed by hybridization with an RNA target.

the electrode surface (Fig. S1a, b).

Then, the 4-inch G-FET wafer was packaged into biosensor chips through Plastic Quad Flat Package (PQFP) in the following steps: (1) cut into approximately 425 small chips ($4\text{ mm} \times 4\text{ mm}$); (2) bond the chip with QFP64 ceramic package (Fig. S1c); (3) cover the contact point with glue to protect the electrical circuit.

Modification of the graphene surface

The freshly as-fabricated G-FET biosensor chips were immersed in 1 mmol L^{-1} PBASE (Sigma-Aldrich) in CH_3CN at room temperature for 6 h, then rinsed several times with CH_3CN and dried using mild stream of N_2 flow [22,23]. Subsequently, the PBASE-modified chips were exposed to either $10\text{ }\mu\text{g mL}^{-1}$ ss-DNA probe (Beijing RuiBiotech Co., Ltd.) or $100\text{ }\mu\text{g mL}^{-1}$ antigen protein in PBS solution at room temperature overnight; they were then being dried by a mild stream of N_2 gas, yielding ss-DNA probe-immobilized chips and antigen protein-immobilized chips, respectively. The antigen protein employed in this study was a recombinant protein

comprising the S and N protein of SARS-CoV-2, developed by Qingdao Hightop Biotech Co., Ltd. (For S protein, the receptor-binding domain (RBD) was chosen. The coincidence rates of N protein to those of SARS or MERS were $\sim 78.12\%$ and $\sim 39.67\%$, respectively. Although the coincidence rate between SARS-CoV-2 and SARS is high, SARS has almost disappeared. There are abundant antigenic determinants in N protein with high specificity, which can avoid missing inspection. Therefore, the full length of N protein was chosen and recombined with the RBD domain of S protein.).

Surface characterization and electrical measurements

The morphology of the G-FET devices was determined by optical microscopy (Nikon Eclipse LV100 POL Microscope). Raman spectroscopy was performed on a confocal Raman spectrograph (JY Horiba HR800) in the back-scattering geometry; the laser (532 nm) was focused on the sample by means of a $100\times$ objective lens (numerical aperture (NA) = 0.9). Analysis of the chemical binding component was determined by X-ray photoelectron

spectroscopy (XPS, Kratos Analytical Ltd., AXIS Supra), with the incident beam produced by an Al X-ray source (150 W) and a pass energy of 160 eV. Surface roughness analysis was conducted by atomic force microscopy (AFM, Bruker, Dimension Icon with Nanoscope V controller). Electrical characterizations, including the transfer curves and current-voltage (I_D - V_D) curves, were carefully carried out at room temperature using a Keysight B1500A semiconductor device analyzer (direct-current measurements) and a Karl Suss (PM5) manual probe station. IgG antibody protein was purchased from Beijing Sino Biological Inc. (isoelectrical point of ~ 6.47 and negatively charged at pH 7).

Extraction of viral RNA and detection of clinical samples

RNA samples from both COVID-19 patients and healthy subjects were extracted from oropharyngeal swabs using QiAamp Viral RNA Mini Kits (Qiagen), which required approximately about 15 min for each sample. The extracted RNA solution (25 μL) was directly added to the packaged chip surface, and heated at 85°C for 9 min. Afterwards, the chip was gently washed with deionized (DI) water to remove the unbonded RNA and dried before measurements with our home-developed electrical detector (Fig. S2). Similarly, for immunoassays, each serum specimen (25 μL) was added to the packaged chip surface at room temperature for 4 min; the surface was rinsed with DI water and dried with a stream of N_2 gas before electrical measurements were taken.

RESULTS AND DISCUSSION

Surface characterization

Fig. 2a displays an optical image of the ten G-FETs in one packaged biosensor chip. The dimensions of the sensing area are 40 $\mu\text{m} \times 8 \mu\text{m}$ (*Length* \times *Width*) for each G-FET. To avoid undesired adsorption of nucleic acids on the surface of gold electrodes and reduce interference during electrical measurements, the electrodes were protected by evaporation of an insulating layer of 40 nm SiO_2 . Details of the fabrication process of a G-FET array and the packaging procedure of the chip are provided in the EXPERIMENTAL SECTION and Supplementary information. As displayed in Fig. 2b–d, Raman spectrum analysis and high-resolution XPS were performed to demonstrate that PBASE were efficiently modified onto the graphene surface. In the Raman spectrum of pristine graphene, two typical major peaks (G and 2D peaks) were observed, which corresponded to the lattice vibration mode and second-order Raman scattering, respectively.

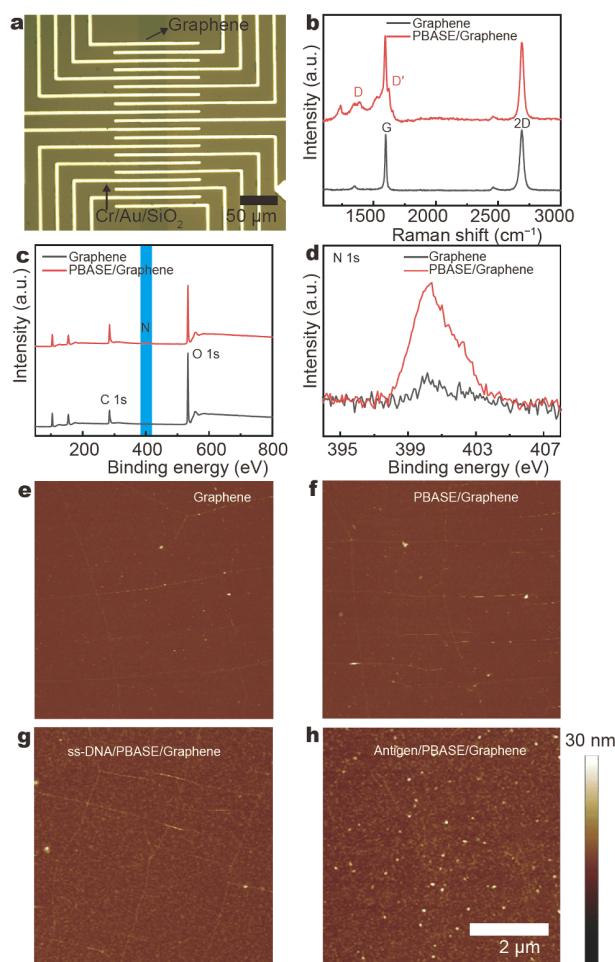


Figure 2 Surface analyses of pristine, PBASE-modified, ss-DNA-immobilized, and antigen-immobilized graphene using Raman spectroscopy, XPS, and AFM. (a) Optical image of a G-FET array in a packaged biosensor chip. (b) Raman spectra of pristine and PBASE-modified graphene. (c, d) High-resolution XPS spectra and enlarged N 1s region for pristine and PBASE-modified graphene. (e–h) AFM images of pristine graphene (e), PBASE-modified graphene (f), ss-DNA probe-immobilized graphene (g) and antigen protein-immobilized graphene (h). Scale bar = 2 μm .

The single Lorentz type 2D peak and the high integrated peak area ratio value of ~ 2.52 between 2D and G peaks indicated high-quality monolayer graphene [29]. While in the Raman spectrum of PBASE-modified graphene, D and D' peaks apparently appeared, stemming from the binding between the pyrene group and graphene. Fig. 2d shows the comparison of the N 1s region before and after the PBASE functionalization of graphene, enlarged from the XPS spectrum (Fig. 2c). For pristine graphene, the N 1s peak is nearly absent; the obviously increased intensity of the N 1s peak after modification indicates successful immobilization of PBASE on the graphene

surface.

We further exploited AFM to visualize the quality of surface functionalization. Fig. 2e–h are AFM images of pristine graphene, PBASE-modified graphene, ss-DNA probe-immobilized graphene, and antigen protein-immobilized graphene, respectively. The corresponding height profiles and surface roughness (RMS) of graphene are depicted in Fig. S3. After PBASE assembly, the RMS of graphene slightly increased from ~0.21 to ~0.40 nm, further reaching ~0.77 and ~1.25 nm with following immobilization of the ss-DNA probe and antigen protein, respectively. The AFM results are in consistence with the larger antigen protein size, compared with the ss-DNA probe. These findings confirmed the successful modification of graphene.

Electrical characterization

Transfer curves of G-FETs were also measured to confirm efficient immobilization of the ss-DNA probe on the graphene surface and to monitor the specific interaction between the ss-DNA probe and its complementary gene sequence (i.e., RdRp target). Ionic liquid as the dielectric layer has been proven to constitute an effective strategy for modulation of charge transport in semiconductor devices [30,31]. Here, we employed ionic liquid containing *N,N*-diethyl-*N*-(2-methoxyethyl)-*N*-methylammonium bis(trifluoromethylsulfonyl)imide (DEMETFSI), to prepare an ionic liquid-gated FET (structure shown in Fig. S4a). For pristine graphene, the transfer curve (Fig. S4b, black line) exhibits the typical ambipolar character of graphene with the Dirac point localized at a gate voltage of ~0.38 V (V_{Dirac}). The positive value of V_{Dirac} indicates that the graphene is *p*-doped, resulting from chemical doping by the residues of the processing chemicals; this phenomenon is common in CVD-grown graphene processed *via* wet transfer [32]. After immobilization of the ss-DNA probe and hybridization with the complementary RdRp target, the value of V_{Dirac} shifted by ~0.38 and ~0.31 V, sequentially. These results are in consistence with findings reported in previous literatures [33,34]. The transfer curve measurements of the pristine, antigen protein-modified, and antibody-bound graphene show a similar tendency (Fig. S4c).

Sensitivity and reusability characterizations

To evaluate the performance of the as-fabricated G-FET biosensors, we measured the $I_{\text{D}}-V_{\text{D}}$ curves of ss-DNA probe-modified devices in response to various concentrations of fully complementary RdRp target (fragment of the RdRp gene sequence in SARS-CoV-2) in

phosphate buffer saline (PBS), with V_{D} ranging from –50 to 50 mV (Fig. 3a, inset is the partially enlarged image). The sequences of the ss-DNA probe, RdRp target, and mismatched DNA are shown in Table S1. An excellent linear relationship between I_{D} and V_{D} was observed over the entire V_{D} scanning range. Therefore, the values of the I_{D} change ratio, in comparison with the origin of ss-DNA probe-modified biosensors ($\Delta I_{\text{D}}/I_0$) at $V_{\text{D}} = 50$ mV, was extracted and plotted as a function of the concentration of RdRp target, as displayed in Fig. 3b. As the concentration of RdRp target increased, the slope of the curve gradually decreased, such that it reached a close equilibrium between the hybridization and electrostatic repulsion of ss-DNA probe and RdRp target at high concentrations. For control experiments, the G-FET biosensor was exposed to different concentrations of a mismatched DNA chain of identical length of or to PBS solution, a slight reduction in the value of $\Delta I_{\text{D}}/I_0$ was observed (Fig. S5), revealing that the ss-DNA probe-modified biosensor was highly specific to its complementary RdRp target. Remarkably, our G-FET biosensor exhibited very high sensitivity with an LOD as low as ~0.1 fg mL⁻¹, which constituted approximately 1800 copies per mL; this was comparable to the LOD of the RT-qPCR method. Notably, our electrical method does not require the amplification steps inherent to RT-qPCR; it can also directly measure the gene target, which reduces the delay for results and lowers the possibility of interference. To examine the reusability of the G-FETs biosensor, the ss-DNA probe-RdRp target hybridization and dehybridization processes were conducted sequentially three times, as shown in Fig. 3e. The values of $\Delta I_{\text{D}}/I_0$ were ~7.2%, ~5.4%, and ~4.4% for the first, second, and third cycles, respectively, indicating that the G-FET biosensor could be reused multiple times. The reusability of the devices will enable cost reduction for each test, which is of considerable importance to the future commercialization of our product.

In addition to nucleic acid detection, we investigated the performance of antigen protein-modified G-FET biosensors in dynamic response to the SARS-CoV-2-specific IgG antibody protein. Similarly, the I_{D} value of the graphene channel increased gradually with increasing IgG antibody protein concentration; a good linear relationship was preserved with V_{D} ranging from –50 to 50 mV, thereby providing an excellent LOD of ~1 fg mL⁻¹ (Fig. 3c, d), which ranks the highest among all immunodetection techniques and at least three orders of magnitude higher than the immune colloidal gold technique [25,28]. Collectively, these results indicated that our

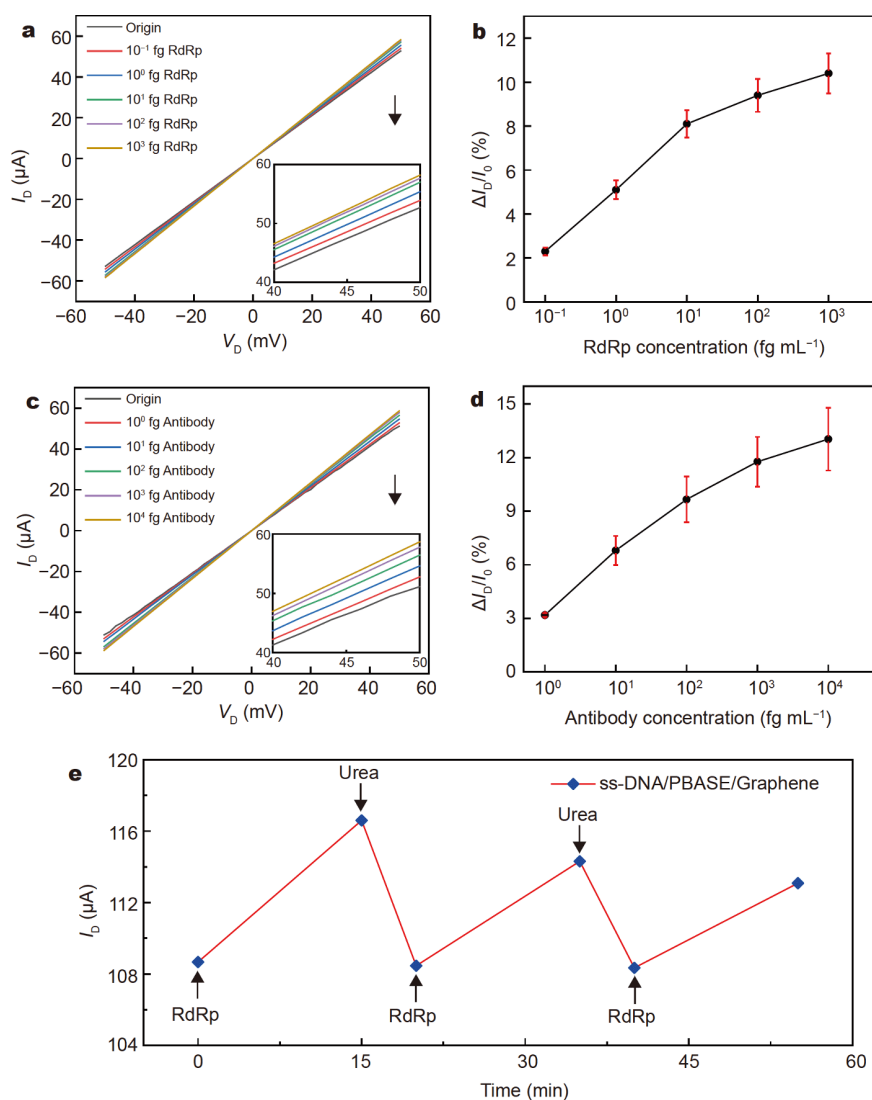


Figure 3 Sensitivity and reusability. (a, c) I_D - V_D curves of G-FET biosensors responding to different concentrations of RdRp target (a) and antibody protein (c). Insets are partially enlarged curves. (b, d) Values of $\Delta I_D/I_0$ at $V_D = 50$ mV extracted and plotted as a function of the concentration of RdRp target (b) and antibody protein (d). (e) Hybridization and dehybridization cycles of the same device.

G-FET biosensors have the potential to rapidly and sensitively detect both the gene RNA sequence of SARS-CoV-2 and an IgG antibody protein specific for the virus, thus laying the foundation for the following clinical diagnostic assay.

SARS-COV-2 detection from clinical samples

Because our G-FET biosensors displayed excellent performance for detection of both SARS-CoV-2 RNA and SARS-CoV-2-specific IgG antibody protein in PBS, we tested their performance for detection of viral RNA in clinical samples, which were provided by Beijing Ditan Hospital Capital Medical University. To evaluate the

performance of ss-DNA probe (RdRp gene) in detection of viral RNA in clinical samples, we firstly developed a DNA-RNA in-site hybridization (RNA-ISH) method to detect SARS-CoV-2 virus in tissue slides (Fig. S6). RNA-ISH is a non-amplification genetic test and a specific test due to long probe design (100 bps in our RNA-ISH system). Fluorescent image (Fig. S6b) of the bronchial brush specimen of COVID-19 patients clearly indicated effectiveness of the RdRp probe in detection of viral RNA. Afterwards, with ss-DNA probe 01, concentration-dependent measurements of viral RNA of clinical samples were performed, indicating an LOD of 1000 copies per mL (Fig. S7). For clinical detection, both COVID-19 patients

and healthy subjects underwent sampling with oropharyngeal swabs, followed by the extraction of viral RNA. Then, the viral RNA was diluted by 100 folds before testing, without further sample processing. All tests of clinical samples were performed using the packaged biosensor chips in our home-developed portable COVID-19 detector (Fig. S8). To achieve maximum accuracy, we designed and integrated ten G-FETs in each packaged biosensor chip. The resistance of ss-DNA probe-modified graphene was recorded as R_n ($n = 1-10$); the resistance was then recorded as R'_n after the addition of clinical samples from either COVID-19 patient or healthy subject. Subsequently, the value of the relative change in resistance $(R'_n - R_n)/R_n$ of each graphene channel was calculated and averaged (defined as $\Delta R/R_0$) as the basis to determine whether a sample yielded positive or negative results. Table 1 summarizes the test results of ten COVID-19 patients and eight healthy subjects. The cutoff value of $\Delta R/R$ was set at -2 ; thus results with $\Delta R/R \leq -2$ were considered positive and those with $\Delta R/R > -2$ were considered negative. All ten COVID-19 patients were correctly identified as COVID-19 infected individuals while eight healthy individuals were correctly identified as healthy, indicating that our G-FET biosensors possess both high sensitivity and specificity in clinical diagnosis of COVID-19 patients. The detection process requires only 10 min after the extraction of viral RNA from oropharyngeal swabs (Fig. S9). In comparison with the RT-

qPCR approach, this method avoids the time-consuming step of viral RNA amplification, in combination with the home-developed portable detector. Future development of our portable detector will be focused on point-of-care testing outside of specialized diagnostic laboratories, which is of considerable importance for large-scale nucleic acid detection of COVID-19 and the ability of large populations to return to work and school.

As an auxiliary measure to complement nucleic acid detection of COVID-19, we assessed the efficiency of immunodetection by validating the performance of our method in the immunoassays of clinical samples. The serum specimens of six COVID-19 patients and three healthy subjects (provided by Beijing Ditan Hospital Capital Medical University) were diluted 100-fold before assessment using our G-FET biosensors. Analysis of each sample required approximately 5 min. As summarized in Table 2, the testing results were in excellent agreement, indicating our biosensors provided accurate, rapid immunological diagnosis of COVID-19.

CONCLUSIONS

In summary, we developed an unprecedented reliable G-FET-based detection system for convenient diagnosis of COVID-19, which consists of two parts: a plug-and-play packaged biosensor chip and a portable electrical measurement machine. This detecting system exhibits obvious advantages of high sensitivity, rapid speed (~ 10 min

Table 1 Nucleic acid analysis of COVID-19 patients and healthy subjects^a

| | Patient 1 | Patient 2 | Patient 3 | Patient 4 | Patient 5 | Patient 6 | Patient 7 | Patient 8 | Patient 9 |
|--------------------|------------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|
| $\Delta R/R_0$ (%) | -9 | -5.8 | -2.8 | -3.6 | -5.9 | -8.9 | -6.2 | -3.1 | -6.1 |
| G-FET results | + | + | + | + | + | + | + | + | + |
| Agreement | Yes | Yes | Yes | Yes | Yes | Yes | Yes | Yes | Yes |
| | Patient 10 | Health 1 | Health 2 | Health 3 | Health 4 | Health 5 | Health 6 | Health 7 | Health 8 |
| $\Delta R/R_0$ (%) | -5.4 | 1.6 | -0.3 | 2 | 3.2 | 4.1 | 5.3 | 2.9 | 2.2 |
| G-FET results | + | - | - | - | - | - | - | - | - |
| Agreement | Yes | Yes | Yes | Yes | Yes | Yes | Yes | Yes | Yes |

a) "Cutoff value" was set at -2 ; "+" represents positive, and "-" represents negative; "Yes" indicates that the G-FET result is in consistency with the clinical standard samples.

Table 2 Antibody analysis of COVID-19 patients and healthy subjects^a

| Number | Patient 1 | Patient 2 | Patient 3 | Patient 4 | Patient 5 | Patient 6 | Health 1 | Health 2 | Health 3 |
|--------------------|-----------|-----------|-----------|-----------|-----------|-----------|----------|----------|----------|
| $\Delta R/R_0$ (%) | -2.6 | -4.5 | -4.5 | -2.8 | -1.3 | -2.6 | 0.7 | 0.2 | 1.1 |
| G-FETs results | + | + | + | + | + | + | - | - | - |
| Agreement | Yes | Yes | Yes | Yes | Yes | Yes | Yes | Yes | Yes |

a) "Cutoff value" was set at -1 ; "+" represents positive, and "-" represents negative; "Yes" indicates that the G-FET result is in consistency with the clinical standard samples.

for RNA analysis and ~5 min for immunoassay), and bifunction (both RNA analysis and immunoassay). These advantages enable high-throughput point-of-care testing, which may facilitate management of the current severe public health crisis. We are firmly of the opinion that this detection system offers a universal methodology that is ready for immediate application and rapid detection of various biomolecules and viruses, such as nucleic acids, proteins, biomarkers, SARS, swine flu, Ebola, and MERS.

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Conflict of interest The authors declare that they have no conflict of interest.

Supplementary Information Experimental details and supporting data are available in the online version of this paper.



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一种针对新型冠状病毒肺炎的准确、快速、便携式双功能电检测仪

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摘要 新型冠状病毒肺炎(COVID-19)正在多个国家快速传播, 已经导致了严重的全球大流行. 由于目前没有针对此类病人的特效药和针对此病毒的疫苗, 准确、快速地进行新冠病人检测成为了控制大流行最有效的措施. 本文中我们开发了一种基于石墨烯场效应晶体管的便携式双功能电检测仪, 其通过核酸互补杂交或者抗原-抗体特异性结合作用, 能分别进行病毒核酸序列检测和抗体检测, 检测限分别低至0.1和1 fg mL⁻¹. 我们通过临床样品检测进一步评估了此方法: 从10个新冠病人和8个正常人咽拭子中提取RNA直接用于核酸检测; 从6个新冠病人和3个正常人血清中提取抗体用于抗体检测. 临床样品检测结果和基于聚合酶链反应的光学方法结果完全吻合, 同时此方法拥有超快的检测速度(核酸检测需10 min, 抗体检测需5 min). 因此, 我们的实验提供了一种有效、准确、高通量的新冠现场即时检测工具.