Anaphylaxis (M Sánchez-Borges, Section Editor)



Penicillin and Cephalosporin-Induced Anaphylaxis: an Update

I. Doña, MD, PhD^{1,*} G. Bogas, MD, PhD¹ N. Pérez-Sánchez, MD¹ T. D. Fernández, PhD² E. Moreno, MD, PhD^{3,4,5} M. J. Torres, MD, PhD^{1,6}

Address

^{*,1}Allergy Unit, Pabellón 6, Primera Planta, Malaga Regional University Hospital (Pavillion C, Hospital Civil), Plaza del Hospital Civil, 29009, Malaga, Spain Email: inmadd@hotmail.com

²Research Laboratory, IBIMA-Regional University Hospital of Malaga-UMA, Málaga, Spain

³Allergy Service, University Hospital of Salamanca, Salamanca, Spain

⁴Biosanitary Institute of Salamanca, Salamanca, Spain

⁵Department of Biomedical and Diagnostics Sciences, Salamanca Medical School, Salamanca, Spain

⁶Andalusian Center for Nanomedicine and Biotechnology-BIONAND, Málaga, Spain

Published online: 23 May 2018 © Springer International Publishing AG, part of Springer Nature 2018

I. Doña, G. Bogas, E. Moreno and M. J. Torres contributed equally to this work. This article is part of the Topical Collection on *Anaphylaxis*

Keywords Anaphylaxis \cdot Beta-lactam \cdot Cephalosporin \cdot Desensitization \cdot Drug provocation test \cdot IgE \cdot Penicillin \cdot Serum-specific IgE \cdot Skin test

Abstract

Purpose of review Beta-lactams (BL) are the most widely used antibiotics and the firstchoice drugs for many bacterial infections. Their consumption has increased in the last decades being now three times higher than the second most highly consumed group. However, they have a high potential for inducing allergic reactions, being the compounds most frequently involved in drug reactions induced by specific immunological mechanisms. The purpose of this review is to summarize the sensitization patterns, the methods used for diagnosing, and the management of BL-induced anaphylaxis.

Recent findings BL can cause severe reactions such as anaphylaxis; in fact, they have been estimated to account for 75% of all fatal anaphylactic episodes in the USA. As a result,

physicians often recommend individuals reporting previous reactions to a given BL to avoid all others in the future. However, we consider this practice to be outdated and in need of revision. The decision of what drugs to recommend should be made based on our knowledge of cross-reactivity between BL groups, based on clinical data and chemical structure. A complete and accurate diagnostic workup must be performed to confirm such cross-reactivity or lack thereof. Our improved knowledge regarding cross-reactivity coincides with recent improvements of in vitro tests, which has decreased the need to perform potentially risky procedures such as drug provocation tests in some situations. This will allow physicians to re-evaluate previous cases and lead to an increase in their therapeutic arsenal to fight against infections.

Summary This review summarizes the complex diagnostic approach and management of BL-induced anaphylaxis focusing in recommendation of alternative BL according to the cross-reactivity between BL groups based on clinical data and chemical structure.

Introduction

Anaphylaxis is a potentially life-threatening, systemic hypersensitivity reaction that results from the sudden release of mediators derived from mast cells and basophils via degranulation [1, 2]. Symptoms of anaphylaxis can involve any organ; however, the most commonly affected ones are the cutaneous (affecting around 88% of cases), respiratory (76.1%), cardiovascular (41.9%), and gastrointestinal systems (12.8%) [3].

Drugs are the most common triggers of anaphylaxis triggers in adults [4–6], being beta-lactams (BL) and non-steroidal anti-inflammatory drugs the most common culprits, accounting for up to 28.1% of all cases of anaphylaxis[7–9] and for 42.6% of all cases of severe drug-induced anaphylaxis.[10••] Amoxicillin is the BL that induces anaphylaxis most frequently.[5, 10••] Recently, clavulanic acid, usually prescribed in combination with amoxicillin, has also been implicated.[11•, 12] Cases induced by cephalosporins, carbapenems, and monobactams are rare.[10••, 13, 14]

BL-induced anaphylaxis has been reported to involve IgE-dependent mechanisms.[15-17] IgE can be directed to the central ring and/or to the side chains (R1 and R2) of the BL molecule. Immunological side chain recognition is particularly relevant for amoxicillin and cephalosporin reactions. [18••] Diagnosis is not always straightforward, and regularly requires the use of skin tests (ST) or in vitro tests, which can be performed during the acute phase (assessing tryptase or histamine levels in serum or histamine metabolites in urine) and after the reaction (serum-specific IgE quantification or basophil activation test (BAT), to confirm the culprit agent. Drug provocation testing (DPT) is not recommended for severe anaphylaxis, due to the high risk of inducing another reaction; it is only recommended in some cases to assess tolerance to potentially cross-reactive drugs.[19] Desensitization should be considered if there is an absolute requirement for a specific BL in the presence of positive ST or DPT.[20••]

Epidemiology and sensitization patterns

Changes in BL antibiotic consumption patterns over time have gradually modified the allergic determinants to which patients can be sensitized.[21] In the1950s, it was estimated that 25 anaphylactic episodes occurred for every 100,000 patients treated with benzylpenicillin.[22] Gradually, benzylpenicillin has been replaced by semisynthetic penicillins with differences in chemical structure, such as amoxicillin, resulting in an increase in the appearance of selective reactions to these drugs.[23] In fact, amoxicillin is now considered the most frequent cause of anaphylaxis to BL.[5, 10••] Since clavulanic acid was introduced in the 1980s, anaphylactic reactions have also been reported to this drug.[11•, 12] In 2007, the rate of anaphylactic reactions to BL was estimated to lie between 0.001 and 0.002% for each treatment course, although these estimates have wide confidence intervals.[24, 25] BL account for up to 20% of all fatal anaphylactic episodes in Europe and up to 75% of all fatal anaphylactic episodes in the USA each year.[9] In the USA, this corresponds to 500–1000 deaths/year.[9] The risk of fatal anaphylaxis with penicillin has been estimated to be between 0.0015 and 0.002% of treated patients.[26] There were no cases of fatal anaphylaxis associated with oral amoxicillin use in a population exposed to 100 million treatment courses over 35-years period.[24]

Concerning cephalosporins, the incidence of anaphylaxis is not well established due to more limited data availability; however, it is generally thought to be lower than for penicillin with an estimated risk range around 0.0001% for each treatment course.[13, 27, 28] However, the incidence of severe anaphylactic reactions to cephalosporins is relatively high.[10••] Now-adays, cephalosporin-induced anaphylaxis is mainly due to the increased use of second-, third-, and fourth-generation cephalosporins,[29] being ceftriaxone, cephalexin, cefuroxime, and cefazolin the most frequent involved.[10••, 13, 28, 30] It is of note that there are more reported cases of anaphylaxis to cephalosporins in patients without known penicillin allergy compared to those with known penicillin allergy.[31, 32]

Penicillins and cephalosporin chemical structures and antigenic determinants

Investigation of allergic reactions requires knowledge of BL structural chemistry.

Chemical structure of penicillin and cephalosporin

Both penicillins and cephalosporins share a common BL ring that is attached to either a 5-membered thiazolidine ring or a 6-membered dihydrothiazine (cephem) ring, respectively. The former has 1 side chain (R1), and the latter has 2 (R1 and R2), with substitution at the R1 and R2 side chains resulting in different chemical structures with a broader spectrum of antibacterial activity and better pharmacokinetic properties.[18••, 33] The earliest generation of cephalosporins focused mainly on the R1 chemical group, whereas the later generations focused on modifications at both the R1 and R2 groups. The homology of aminopenicillin and cephalosporin side chains is shown in Table 1. Additional modifications to the basic core structures of both penicillins and cephalosporins have been made, leading to other types of BL-containing antibiotics such as carbapenems and monobactams.

Antigenic determinants

Penicillin undergoes spontaneous degradation because of a chemically unstable BL ring, forming reactive intermediate products which can bind to lysine residue aminogroups on soluble or cell-bound proteins.[34–36] This results in the

Table 1. Homology of side chains of the chemical structure of aminopenicillins and cephalosporins				
Identical R1 side chain (C7 position) • Amoxicillin, cefadroxil ¹ , cefatrizine ¹ , cefprozil ² • Ampicillin, cephalexin ¹ , cephradine ¹ , cephaloglycin ¹ , cefaclor, ² loracarbef ² • Cephalotin ¹ , cephaloridine ¹ , cefoxitin ² • Cefamandole ² , cefonicid ² • Ceftriaxone ³ , cefotaxime ³ , cefpodoxime ³ , cefditoren ³ , ceftizoxime ³ , cefmenoxime ³ , cefepime ⁴ Similar R1 side chain (C7 position) • Cefadroxil ¹ , cefaclor ² • Ceftazidime ³ , ceftriaxone ³ , cefotaxime ³ , cefixime ³ , cefpodoxime ³ , cefepime ⁴	Identical R2 side chain (C3 position) • Cephalexin ¹ , cefadroxil ¹ , cephradine ¹ • Cephapirin ¹ , cephalothin ¹ , Cephaloglycin ¹ , cefotaxime ³ • Cefuroxime ² , cefoxitin ² • Cefotetan ² , cefamandole ² , cefmetazole ² , cefpiramide ³ • Cefaclor, ² loracarbef ² • Ceftibuten, ³ ceftizoxime ³ Similar R2 side chain (C3 position) • Cefuroxime ² , cefotaxime ³			
 ¹First-generation cephalosporins ²Second-generation cephalosporins ³Third-generation cephalosporins ⁴Fourth-generation cephalosporins 				

formation of benzyl penicilloyl (BPO), known as the major antigenic determinant of penicillin.[35–38] The remaining part of the benzylpenicillin molecule degrades to a range of derivatives which can also act as haptens. These are minor determinants, accounting for allergic reactions in approximately 15–16% of patients.[20••] The minor determinants do not cross-react with each other and are known to provoke severe anaphylactic reactions.[33, 34] In addition to the BL ring, the side chains can also trigger allergic reactions.[35, 36]

The degradation process for cephalosporins leads to fragmentation of the BL ring as well as the thiazinic group, resulting in larger degradation products. This process is more rapid than the fragmentation of penicillin. The exact nature of these intermediate products has not been characterized, [39, 40] but the haptenization mechanism appears slower and possibly more complex than for penicillins.[41]

Because of these differences in degradation processes between penicillins and cephalosporins, the investigation of IgE-mediated reactions to BL must include major and minor penicillin determinants as well as to the whole molecule; conversely, the investigation of IgE-mediated reactions to cephalosporin should only include the native molecule.

Diagnosis

The diagnosis of BL-induced anaphylaxis is based on clinical history, physical examination (if signs or symptoms are present), ST, in vitro tests, and DPT.

Clinical history

Diagnosis of BL-induced anaphylaxis is based on recognition of characteristic signs and symptoms during the reaction (Table 2), which generally start within minutes after administration of the drug.[42] It essentially involves recording clinical history, including a detailed description of the symptoms and their

Acute onset of an illness with skin-mucosal involvement and at least one of the following		Respiratory compromise Decreased blood pressure, syncope, or collapse	
or Two or more of the following that occ exposure to a likely allergen for that or Decreased blood pressure after expose for that patient	cur rapidly after at patient ure to a known allergen	Skin-mucosal involvement Respiratory compromise Decreased blood pressure, syncope, or collapse Gastrointestinal symptoms	
	severity.[43] However, the clinica patient is evaluated a long tim established the diagnosis of ana identified the culprit agent, the p	l history can be imprecise in many cases as the e after the reaction. After the physician has phylaxis based on clinical symptoms and has atient should undergo an allergological study.	
Skin test	ST has been shown to be an im allergy to BL.[21, 44] Firstly, th performed. If this does not cause be carried out. In patients who phylactic reactions, IDT should s therapeutic drug concentration, centration is achieved (Table 3). personnel as systemic reactions previous history of anaphylaxis.[The current benzylpenicillin of benzylpenilloic acid in many Ef (PPL) in the USA and Canada. naturally metabolized penicillin initial ST.[16, 21, 45•] Currenth	portant method for confirming IgE-mediated he skin-prick test (SPT) technique is usually a reaction, an intradermal test (IDT) can then have previously suffered life-threatening ana- tart with a dilution of 1/1000 or 1/100 of the increasing 10-fold until a non-irritating con- [21, 45•] ST should be undertaken by trained may occur in up to 8% of patients with a 46] determinants consist of BPO octa-L-lysine and uropean countries, and of BPO poly-L-lysine A minor determinant mixture (MDM) of the h G products must also be included for the y, changes in patterns of consumption of BL	

Table 3. Non-irritating concentrations for ST to BL antibiotics

Reagent	Concentration
BP-OL	0.04 mg/ml
MDM	0.5 mg/ml
Benzylpenicillin	10,000 UI/ml
Amoxicillin	20 mg/ml
Clavulanic acid	5 mg/ml
Cephalosporins	2 mg/ml

Table 2. Clinical criteria for the diagnosis of anaphylaxis, taken from Simons FE et al. World Allergy Organ J 201542

has led to a fall in ST positivity rates for major and minor benzylepnicillin determinants[22, 47] and the need for other determinants, such as amoxicillin.[21, 44] In fact, amoxicillin has become the most important determinant of penicillin allergy and its inclusion is essential in the diagnosis of BL anaphylaxis nowadays.[48-51] Indeed, the inclusion of amoxicillin in ST could increase positivity up to 70%.[48] In the case of amoxicillin, the equivalent determinant for benzylpenicilloic acid (amoxicilloic acid) and benzylpenilloic acid (amoxilloic acid) are not of value for ST, and amoxicillin itself with the intact BL ring is the reagent used.[52] There are no clear benefits to adding benzylpenicillin to ST that already include PPL and MDM, in populations where amoxicillin and amoxicillin-clavulanic acid are the main culprit drugs.[53] However, it can be useful if PPL and MDM are not available, as it has been reported that up to 5% of BL allergic patients with negative ST to PPL and MDM gives positive results to benzylpenicillin in ST.[54] In addition, ST should include amoxicillin and clavulanic acid separately if anaphylaxis occurred after administration of amoxicillin-clavulanic acid, [21, 45•] as cases of selective hypersensitivity reactions to clavulanic acid have been reported in recent years.[11•, 12, 55] Including clavulanic acid in ST has been shown to increased sensitivity from 9 to 18.7% in SPT and from 63.6 to 81.2% in IDT.[11°, 12]Concerning cephalosporins, ST are done with the native molecule (intravenous preparations or crushed tablets solubilized in buffer) and can predict hypersensitivity only to the specific cephalosporin ST reagent or cephalosporins with similar side chains.[18••] Moreover, concentrations for ST with native molecule cephalosporins have to be standardized. [56, 57]

In general, the percentage of positive ST in patients with a clinical history of a BL allergic reaction varies between 7 and 76% according to different studies,[51, 58, 59] with the higher results given by patients with suggestive clinical histories of immediate reactions (urticaria and anaphylaxis), as well as when ST are made a short time after the reaction.[21, 59, 60] Prospective studies show that ST reactivity decreases over time in penicillin-allergic patients, with only 30–50% of patients with initial positive ST remaining positive after 5 years[60]; this percentage is even higher in the case of aminopenicillins.[60] Several studies suggest that between 1 and 27.9% of subjects may become positive again after BL administration (resensitization).[51, 58, 61, 62] For this reason, it is necessary to re-evaluate the patient after 1 month if they experienced anaphylaxis to BL but the allergological study gives negative results,[44, 51, 63] particularly if the reaction occurred more than a year ago.

However, despite using a large panel of BL, the sensitivity of ST is not optimal[44] and even in recent years, it has been decreasing, meaning that diagnosis must be achieved through DPT in a significant percentage of patients.[48, 64]

In vitro

These methods can be performed whilst the reaction is still ongoing (acute phase), during which we can analyze the release of different mast cell mediators that occurs after the symptoms onset, in order to confirm the diagnosis of anaphylaxis. In vitro methods can be performed once the reactions is over (diagnostic phase), in order to identify the culprit BL.

Acute phase

Several mediators have been studied as possible biomarkers of an anaphylactic reaction, although the release of tryptase and histamine during the acute phase are the most frequently used in the clinical practice. [65•] Both mediators can be determined in plasma by immunoassay, [66] although histamine can also be determined in urine.[67] These mediators are continuously released by resting mast cells, and therefore mast cell diseases can influence basal levels, [68] thus it is important to compare the values obtained at the time of the event with a recent baseline. [69, 70] The halflife of tryptase in serum is 90-120 min; therefore, the optimal timing for measuring its levels is between 30 and 120 min after the initiation of symptoms. Basal levels must be measured at least 24 h after resolution of the reaction.[19, 71-73] In the case of histamine, due to its short half-life of only 20 min, blood must be collected during the first hour after symptoms onset, which limits its use in clinical practice. [74, 75] Another possibility is the measurement of two histamine metabolites, Nmethylhistamine and N-methylimidazoleacetic acid, in urine. Both appear 30-60 min after the onset of a reaction and are detectable for 24 h.[67, 76, 77] There is a lack of studies with a sufficient number of cases to establish the sensitivity and specificity of this technique to diagnose penicillin and cephalosporin-induced anaphylaxis. In studies that also include other drugs, sensitivity of tryptase determination has been estimated to range between 37-94% and specificity between 92-94%, [65•, 78] depending on the cut-off point used.

Diagnostic phase

Immunoassays

Serum-specific IgE quantification can be performed using immunoassays, such as commercial assays or custom-made radioimmunoassays.[70] The sensitivity of these methods in patients with anaphylaxis to benzylpenicillin or amoxicillin is around 55%, with a specificity of 97%.[79]

Basophil activation test

It is a more functional cellular test that mimics the in vivo reaction. The technique is based on the measurement of basophil activation after drug stimulation using flow cytometry.[80] The use of BAT as a part of the allergological workup is increasing; in fact some authors have recommended its inclusion in diagnostic algorithms even before the performance of ST.[3] BAT can be of great value in decreasing the necessity to perform DPT, especially in patients suffering life-threatening reactions such as anaphylaxis.[81, 82] The sensitivity of BAT for penicillins has been estimated to be around 50%, with a specificity of 90%,[80, 83, 84] although when analyzing patients with more severe reactions sensitivity increases to 70%.[81] Interestingly, around 25% of amoxicillin allergic patients with a negative ST show a

positive BAT result.[81, 84] This value is even higher in clavulanic acid allergic patients, for whom nearly 50% of patients with a negative ST showed a positive BAT.[81] When combining the results of BAT and ST together, between 80 and 90% of patients suffering anaphylactic reactions after amoxicillin-clavulanic acid intake could be diagnosed without the need to perform DPT.[81]

Drug provocation test

DPT is considered to be the gold standard to establish or exclude the diagnosis of hypersensitivity to a certain substance.[85•] However, it is not recommended for severe anaphylaxis, due to the high risk of inducing another reaction. It is primarily indicated for patients where the drug being tested is thought unlikely to be the trigger, and for assessing tolerance to potentially cross-reactive drugs.[19] DPT is time and cost-consuming, and given the high possibility of inducing another allergic reaction, patients should undergo a risk-benefit analysis prior to the procedure. It should only be performed by trained personnel in a clinical setting where resuscitation facilities are available.[85•] Ideally, it should be performed 4-6 weeks after the episode due to the high rate of negativization of diagnostic tests over time. [60, 86•, 87] The drug is administered at increasing doses, with a minimum interval of 30 to 60 min between each administration, until the full therapeutic dose is reached.[33] Different protocols for DPT have been published, [21, 30, 55] with that of Messaad et al. being the most frequently used[88] (Table 4).

Management

This includes the treatment of the acute episode of anaphylaxis and its subsequent management, and a diagnostic workup that can include either recommendation of alternative BL or desensitization with the culprit BL.

Anaphylaxis treatment

An anaphylactic reaction is a life-threatening situation that needs urgent medical assessment, even in some cases intensive care.[89•, 90, 91] Immediate treatment is the same, regardless of the trigger (Airway, Breathing, Circulation, Disability, and Exposure approach). Intramuscular adrenaline (1 mg/ml) is recommended as the first-line treatment due to its agonist effects on α -1, β -1, and β -2 receptors.[92] Second-line treatment includes correct patient posture, fluids and oxygen support, and administration of short-acting β -2 agonists if bronchospasm symptoms are present. As a third-line treatment, H1 and H2 antihistamines and glucocorticosteroids should be given. It is important after a BL-induced anaphylactic reaction to provide the patient with oral and written information about BL avoidance in order to prevent another adverse event and to refer the patient for allergological study.

Recommendation of alternative BL: cross-reactivity between BL compounds

All too frequently, patients suffering anaphylaxis to a suspected BL are told to avoid all BL, without taking into account cross-reactivity between

Drug (route)	Messad et al. 2004[88] Doses every 30 min	Blanca et al. 2009[21] Doses every 45–60 min	Blanca-Lopez et al. 2015[55] Doses every 30 min	Romano et al. 2016[30] Doses every 60 min
Benzylpenicillin (intramuscular)		10 ³ , 10 ⁴ , 10 ⁵ , 5 × 105 IU/ml (CD 6 × 10 ⁵ IU/ml)		
Penicillin V (oral)		5, 50, 150, 200 (CD 400 mg)		
Amoxicillin (oral)	1, 5, 25, 100, 500, 1000 (CD 1000-2000 mg)	5, 50, 100, 150, 200 (CD 500 mg)	5, 50, 125, 250, 500 (CD 1000 mg)	
Amoxicillin-clavulanic acid (oral)			50/12.5; 125/31.25; 250/62.5; 500/125 (CD 925/231.25 mg)	
Ampicillin (oral)	1, 5, 25, 100, 500, 1000 (CD 1000-2000 mg)			
Cloxacillin (oral)	1, 5, 25, 100, 500, 1000 (CD 2000 mg)			
Cefaclor (oral)				5, 50, 445 (CD 500 mg)
Cefadroxil (oral)	1, 5, 25, 100, 500, 1000 (CD 2000 mg)			
Cefazolin (intravenous/intramuscular)	1, 5, 25, 100, 500, 2000 (CD 1500–3000 mg)			10, 100, 890 (CD 1000 mg)
Cefuroxime (oral)	1, 5, 20, 80, 400 (CD 500 mg)			5, 50, 445 (CD 500 mg)
Ceftazidime (intravenous)	1, 5, 25, 100, 500, 2000 CD 3000 mg)			
Cefixime (oral)	1, 5, 25, 100, 225 (CD 400 mg)			
Ceftriaxone (intravenous/intramuscular)	1, 5, 25, 100, 500, 1000 (CD 1000-2000 mg)			10, 100, 890 (CD 1000 mg)

Table 4. Doses recommended for DPT to BL. CD cumulative dose

different BL compounds and the existence of selective responders. Subjects with an IgE response to the BPO structure usually respond to several penicillin derivatives (including aminopenicillins such as amoxicillin) and first-generation cephalosporins.[93] This IgE-response differs in subjects with selective allergy to amoxicillin or cephalosporins; in these cases, the





antigenic determinants are predominantly side chain structures (R1 for the amoxicillin; R1 and R2 for cephalosporins).[33, 94] Furthermore, other parts of the molecule also account for cross-reactivity. Thus, the attributable risk of an allergic cross-reactivity between penicillins and cephalosporins, for all but a few cephalosporins with similar side chain structures to penicillin, is essentially nil.[18^{••}] With these preliminary concepts, the decision algorithm for the allergological workup is described in Figs. 1 and 2.



This procedure is recommended when alternative drugs are not available or not effective and the culprit drug is the only treatment option.[95] Desensitization is defined as the induction of a temporary state of tolerance to a compound that





caused a hypersensitivity reaction previously. This tolerance is achieved after several hours by the administration of increasing doses of the drug involved until the therapeutic dose is reached. [20^{••}, 97] Although desensitization protocols were first described for penicillin, [98, 99] there is a distinct lack of validated BL desensitization protocols in general, and the European Network on Drug Allergy has recommended that effort be put into establishing better protocols for desenzitation by replicating previous studies (consensus statement of the European Academy of Allergy and Clinical Immunology[95]). A good example of how to achieve this aim is provided by researchers at the Brigham and Women's Hospital in Boston. [100-102]

Conclusions

BL continue to be the most highly used antibiotics worldwide. They are also the most frequent triggers of hypersensitivity reactions to drugs. The rise in prescription of amoxicillin, clavulanic acid, and cephalosporins in recent decades means that the details of the original allergological assessments need to be updated. The major and minor determinants of benzylpenicillin are no longer the main allergenic molecules, side chains now play an important role, and the knowledge of cross-reactivity between BL is crucial for recommending alternatives. Additionally, the development of specific in vitro tests, particularly the BAT, is helping us to perform more accurate and safer diagnosis. Despite all these changes, desensitization remains essential in some cases, and more research is needed in this area to establish better procedures. We hope that this review will go some way towards improving BL hypersensitivity management, leading to a decrease in the use of risky procedures and unnecessary avoidance of important antibiotics.

Acknowledgments

We thank James Perkins for helping us revise the English version of the manuscript.

Funding Information

The present study has been supported in part by Institute of Health "Carlos III" of the Ministry of Economy and Competitiveness (grants cofunded by European Regional Development Fund (ERDF): ARADYAL RD16/0006/0001, PI15/01206) and Andalusian Regional Ministry Health (grant: PI-0241-2016). Doña holds a "Juan Rodes" research contract (JR15/00036) supported from the Institute of Health "Carlos III" of the Ministry of Economy and Competitiveness (grants cofunded from the European Social Fund (ESF)). Bogas G and Perez-Sanchez N hold a "Río Hortega" research contract (CM16/0067 and CM17/00141) supported from the Institute of Health "Carlos III" of the Ministry of Economy and Competitiveness (grants cofunded from the European Social Fund (ESF)). Fernández TD holds a "Ramon y Cajal" research contract (RYC-2013-01283) supported from the Ministry of Economy and Competitiveness (grant cofunded from the European Social Fund (ESF)).

Compliance with Ethical Standards

Conflict of Interest

I. Doña declares that she has no conflict of interest. G. Bogas declares that he has no conflict of interest. N. Perez-Sanchez declares that she has no conflict of interest. T. D. Fernandez declares that she has no conflict of interest. E. Moreno declares that she has no conflict of interest. M. J. Torres declares that she has no conflict of interest.

Human and Animal Rights and Informed Consent

This article does not contain any studies with human or animal subjects performed by any of the authors.

References and Recommended Reading

Papers of particular interest, published recently, have been highlighted as:

Of importance

- •• Of major importance.
- 1. Johansson SG, Bieber T, Dahl R, Friedmann PS, Lanier BQ, Lockey RF, et al. Revised nomenclature for allergy for global use: report of the Nomenclature Review Committee of the World Allergy Organization, October 2003. J Allergy Clin Immunol. 2004;113(5):832–6.
- Sampson HA, Munoz-Furlong A, Campbell RL, et al. Second symposium on the definition and management of anaphylaxis: summary report—Second National Institute of Allergy and Infectious Disease/Food Allergy and Anaphylaxis Network symposium. J Allergy Clin Immunol. 2006;117(2):391–7.
- Aun MV, Kalil J, Giavina-Bianchi P. Drug-induced anaphylaxis. Immunol Allergy Clin N Am. 2017;37(4):629–41.
- Liew WK, Williamson E, Tang ML. Anaphylaxis fatalities and admissions in Australia. J Allergy Clin Immunol. 2009;123(2):434–42.
- 5. Gonzalez-Perez A, Aponte Z, Vidaurre CF, Rodriguez LA. Anaphylaxis epidemiology in patients with and patients without asthma: a United Kingdom database review. J Allergy Clin Immunol 2010; 125(5): 1098–1104 e1.
- Wood RA, Camargo CA Jr, Lieberman P, Sampson HA, Schwartz LB, Zitt M, et al. Anaphylaxis in America: the prevalence and characteristics of anaphylaxis in the United States. J Allergy Clin Immunol. 2014;133(2):461–7.
- Gabrielli S, Clarke AE, Eisman H, et al. Disparities in rate, triggers, and management in pediatric and adult cases of suspected drug-induced anaphylaxis in Canada. Immunity, inflammation and disease 2017.
- Jares EJ, Baena-Cagnani CE, Sanchez-Borges M, et al. Drug-induced anaphylaxis in Latin American countries. J Allergy Clin Immunol Pract. 2015;3(5):780–8.
- 9. Neugut AI, Ghatak AT, Miller RL. Anaphylaxis in the United States: an investigation into its epidemiology. Arch Intern Med. 2001;161(1):15–21.
- 10.•• Renaudin JM, Beaudouin E, Ponvert C, Demoly P, Moneret-Vautrin DA. Severe drug-induced anaphylaxis:

analysis of 333 cases recorded by the Allergy Vigilance Network from 2002 to 2010. Allergy. 2013;68(7):929–37. In this paper, authors analyzed a large serie of cases with severe anaphylaxis providing data on clinical characteristics and drugs involved as well as the efficacy of a stepwise procedure for optimal diagnostic approach.

11.• Torres MJ, Ariza A, Mayorga C, et al. Clavulanic acid can be the component in amoxicillin-clavulanic acid responsible for immediate hypersensitivity reactions. J Allergy Clin Immunol. 2010;125(2):502–5 e2.

In this paper, authors show the relevance of new component (clavulanic acid) in skin tests for the study of immediate allergy reactions to current penicillins.

- Sanchez-Morillas L, Perez-Ezquerra PR, Reano-Martos M, Laguna-Martinez JJ, Sanz ML, Martinez LM. Selective allergic reactions to clavulanic acid: a report of 9 cases. J Allergy Clin Immunol. 2010;126(1):177–9.
- Macy E, Contreras R. Adverse reactions associated with oral and parenteral use of cephalosporins: a retrospective population-based analysis. J Allergy Clin Immunol. 2015;135(3):745–52 e5.
- 14. Iglesias Cadarso A, Saez Jimenez SA, Vidal Pan C, Rodriguez Mosquera M. Aztreonam-induced anaphylaxis. Lancet. 1990;336(8717):746–7.
- Antunez C, Fernandez T, Blanca-Lopez N, Torres MJ, Mayorga C, Canto G, et al. IgE antibodies to betalactams: relationship between the triggering hapten and the specificity of the immune response. Allergy. 2006;61(8):940–6.
- Joint Task Force on Practice P, American Academy of Allergy A, Immunology, et al. Drug allergy: an updated practice parameter. Ann Allergy, Asthma Immunol : Off Publ Am College Allergy, Asthma, Immunol. 2010;105(4):259–73.
- 17. Lagace-Wiens P, Rubinstein E. Adverse reactions to beta-lactam antimicrobials. Expert Opin Drug Saf. 2012;11(3):381–99.

18.•• Zagursky RJ, Pichichero ME. Cross-reactivity in betalactam allergy. The journal of allergy and clinical immunology In practice 2017.

In this review, authors focus on the structural involvement of the R1 and R2 chemical side chains of aminopenicillins and cephalosporins and the clinical relevance of immunologic cross-reactivity in patients with confirmed allergy to cephalosporins.

- 19. Kuruvilla M, Khan DA. Anaphylaxis to drugs. Immunol Allergy Clin N Am. 2015;35(2):303–19.
- 20.•• Mirakian R, Leech SC, Krishna MT, et al. Management of allergy to penicillins and other beta-lactams. Clin Exp Allergy: J Br Soc Allergy Clin Immunol. 2015;45(2):300–27.

This guideline adresses management of allergic reactions to betalactams.

- Blanca M, Romano A, Torres MJ, Férnandez J, Mayorga C, Rodriguez J, et al. Update on the evaluation of hypersensitivity reactions to betalactams. Allergy. 2009;64(2):183–93.
- Macy E. Penicillin and beta-lactam allergy: epidemiology and diagnosis. Curr Allergy Asthma Rep. 2014;14(11):476.
- 23. Silviu-Dan F, McPhillips S, Warrington RJ. The frequency of skin test reactions to side-chain penicillin determinants. J Allergy Clin Immunol. 1993;91(3):694–701.
- 24. Lee P, Shanson D. Results of a UK survey of fatal anaphylaxis after oral amoxicillin. J Antimicrob Chemother. 2007;60(5):1172–3.
- 25. Johannes CB, Ziyadeh N, Seeger JD, Tucker E, Reiter C, Faich G. Incidence of allergic reactions associated with antibacterial use in a large, managed care organisation. Drug Saf. 2007;30(8):705–13.
- 26. International Collaborative Study of Severe A. Risk of anaphylaxis in a hospital population in relation to the use of various drugs: an international study. Pharmacoepidemiol Drug Saf. 2003;12(3):195–202.
- 27. Kelkar PS, Li JT. Cephalosporin allergy. N Engl J Med. 2001;345(11):804–9.
- 28. Tejedor Alonso MA, Moro MM, Hernandez JE, et al. Incidence of anaphylaxis in hospitalized patients. Int Arch Allergy Immunol. 2011;156(2):212–20.
- Romano A, Torres MJ, Castells M, Sanz ML, Blanca M. Diagnosis and management of drug hypersensitivity reactions. J Allergy Clin Immunol. 2011;127(3 Suppl):S67–73.
- Romano A, Gaeta F, Valluzzi RL, et al. IgE-mediated hypersensitivity to cephalosporins: cross-reactivity and tolerability of alternative cephalosporins. J Allergy Clin Immunol 2015; 136(3): 685–91 e3.
- Pichichero ME. A review of evidence supporting the American Academy of Pediatrics recommendation for prescribing cephalosporin antibiotics for penicillinallergic patients. Pediatrics. 2005;115(4):1048–57.
- 32. Anne S, Reisman RE. Risk of administering cephalosporin antibiotics to patients with histories of penicillin allergy. Ann Allergy, Asthma Immunol: Off Publ

Am Coll Allergy, Asthma, Immunol. 1995;74(2):167–70.

33. Torres MJ, Blanca M. The complex clinical picture of beta-lactam hypersensitivity: penicillins, cephalosporins, monobactams, carbapenems, and clavams. Med Clin N Am. 2010;94(4):805–20.

xii

- Solensky R. Hypersensitivity reactions to beta-lactam antibiotics. Clin Rev allergy immunology. 2003;24(3):201–20.
- Baldo BA, Zhao Z, Pham NH. Antibiotic allergy: immunochemical and clinical considerations. Curr Allergy Asthma Rep. 2008;8(1):49–55.
- Baldo BA. Penicillins and cephalosporins as allergens—structural aspects of recognition and crossreactions. Clin Exp Allergy: J Br Soc Allergy Clin Immunol. 1999;29(6):744–9.
- 37. Macy E, Richter PK, Falkoff R, Zeiger R. Skin testing with penicilloate and penilloate prepared by an improved method: amoxicillin oral challenge in patients with negative skin test responses to penicillin reagents. J Allergy Clin Immunol. 1997;100(5):586–91.
- Bousquet PJ, Co-Minh HB, Arnoux B, Daures JP, Demoly P. Importance of mixture of minor determinants and benzylpenicilloyl poly-L-lysine skin testing in the diagnosis of beta-lactam allergy. J Allergy Clin Immunol. 2005;115(6):1314–6.
- Antunez C, Blanca-Lopez N, Torres MJ, Mayorga C, Perez-Inestrosa E, Montañez MI, et al. Immediate allergic reactions to cephalosporins: evaluation of crossreactivity with a panel of penicillins and cephalosporins. J Allergy Clin Immunol. 2006;117(2):404–10.
- Sanchez-Sancho F, Perez-Inestrosa E, Suau R, et al. Synthesis, characterization and immunochemical evaluation of cephalosporin antigenic determinants. J Mol Recognition: JMR. 2003;16(3):148–56.
- 41. Perez-Inestrosa E, Suau R, Montanez MI, et al. Cephalosporin chemical reactivity and its immunological implications. Curr Opin Allergy Clin Immunol. 2005;5(4):323–30.
- 42. Simons FE, Ebisawa M, Sanchez-Borges M, et al. Update of the evidence base: World Allergy Organization anaphylaxis guidelines. World Allergy Organ J. 2015;8(1):32.
- 43. Ring J, Behrendt H. Anaphylaxis and anaphylactoid reactions. Classification and pathophysiology. Clin Rev Allergy immunolo 1999; 17(4): 387–399.
- Torres MJ, Blanca M, Fernandez J, Romano A, Weck A, Aberer W, et al. Diagnosis of immediate allergic reactions to beta-lactam antibiotics. Allergy. 2003;58(10):961–72.
- 45.• Brockow K, Garvey LH, Aberer W, et al. Skin test concentrations for systemically administered drugs—an ENDA/EAACI Drug Allergy Interest Group position paper. *Allergy*. 2013;68(6):702–12.

In this paper, EAACI recommendations of skin test concentrations for systemically drugs are provided.

46. Co Minh HB, Bousquet PJ, Fontaine C, Kvedariene V, Demoly P. Systemic reactions during skin tests with beta-lactams: a risk factor analysis. J Allergy Clin Immunol. 2006;117(2):466–8.

- 47. Dona I, Blanca-Lopez N, Torres MJ, et al. Drug hypersensitivity reactions: response patterns, drug involved, and temporal variations in a large series of patients. J Investig Allergol Clin Immunol. 2012;22(5):363–71.
- Torres MJ, Romano A, Mayorga C, et al. Diagnostic evaluation of a large group of patients with immediate allergy to penicillins: the role of skin testing. Allergy. 2001;56(9):850–6.
- 49. Blanca M, Romano A, Torres MJ, Demoly P, DeWeck A. Continued need of appropriate betalactam-derived skin test reagents for the management of allergy to betalactams. Clinical and experimental allergy: journal of the British Society for Allergy and Clin Immunol. 2007;37(2):166–73.
- Lin E, Saxon A, Riedl M. Penicillin allergy: value of including amoxicillin as a determinant in penicillin skin testing. Int Arch Allergy Immunol. 2010;152(4):313–8.
- Moreno E, Laffond E, Munoz-Bellido F, et al. Performance in real life of the European Network on Drug Allergy algorithm in immediate reactions to betalactam antibiotics. Allergy. 2016;71(12):1787–90.
- Torres MJ, Ariza A, Fernandez J, et al. Role of minor determinants of amoxicillin in the diagnosis of immediate allergic reactions to amoxicillin. Allergy. 2010;65(5):590–6.
- Lacombe-Barrios J, Salas M, Gomez F, et al. The addition of benzylpenicillin does not increase the skin test sensitivity obtained with classic beta-lactam determinants. J Investig Allergol Clin Immunol. 2016;26(1):52–4.
- Romano A, Bousquet-Rouanet L, Viola M, Gaeta F, Demoly P, Bousquet PJ. Benzylpenicillin skin testing is still important in diagnosing immediate hypersensitivity reactions to penicillins. Allergy. 2009;64(2):249–53.
- 55. Blanca-Lopez N, Perez-Alzate D, Ruano F, Garcimartin M, de la Torre V, Mayorga C, et al. Selective immediate responders to amoxicillin and clavulanic acid tolerate penicillin derivative administration after confirming the diagnosis. Allergy. 2015;70(8):1013–9.
- Uyttebroek AP, Decuyper, II, Bridts CH, et al. Cefazolin hypersensitivity: toward optimized diagnosis. J Allergy Clin Immunol Pract 2016; 4(6): 1232–6.
- Kim MH, Lee JM. Diagnosis and management of immediate hypersensitivity reactions to cephalosporins. Allergy, Asthma Immunol Res. 2014;6(6):485–95.
- Goldberg A, Confino-Cohen R. Skin testing and oral penicillin challenge in patients with a history of remote penicillin allergy. Ann Allergy, Asthma & Immunol: Off Publ Am Coll Allergy, Asthma, Immunol. 2008;100(1):37–43.
- Moreno E, Davila I, Laffond E, Gracia T, Munoz F, Lorente F. Immediate allergic reactions to beta-lactams: diagnostic accuracy of skin tests. Ann Allergy, Asthma Immunol: Off Publ Am Colle Allergy, Asthma, Immunol. 2011;107(1):89–90.

- 60. Blanca M, Torres MJ, Garcia JJ, et al. Natural evolution of skin test sensitivity in patients allergic to beta-lactam antibiotics. J Allergy Clin Immunol. 1999;103(5 Pt 1):918–24.
- 61. Solensky R, Earl HS, Gruchalla RS. Lack of penicillin resensitization in patients with a history of penicillin allergy after receiving repeated penicillin courses. Arch Intern Med. 2002;162(7):822–6.
- 62. Garcia Nunez I, Barasona Villarejo MJ, Algaba Marmol MA, Moreno Aguilar C, Guerra PF. Diagnosis of patients with immediate hypersensitivity to beta-lactams using retest. J Investig Allergology Clin Immunol. 2012;22(1):41–7.
- 63. Demoly P, Adkinson NF, Brockow K, Castells M, Chiriac AM, Greenberger PA, et al. International consensus on drug allergy. Allergy. 2014;69(4):420–37.
- Torres MJ, Mayorga C, Leyva L, Guzman AE, Cornejo-Garcia JA, Juarez C, et al. Controlled administration of penicillin to patients with a positive history but negative skin and specific serum IgE tests. Clin Exp Allergy: J Br Soc Allergy Clin Immunol. 2002;32(2):270–6.
- 65.• Mayorga C, Celik G, Rouzaire P, et al. In vitro tests for drug hypersensitivity reactions: an ENDA/EAACI Drug Allergy Interest Group position paper. Allergy. 2016;71(8):1103–34.

In this position paper by ENDA/EAACI Drug Allergy Interest Group, data and recommendations regarding the available in vitro methods for drug hypersensitivity diagnosis are provided.

- Schwartz LB. Diagnostic value of tryptase in anaphylaxis and mastocytosis. Immunol Allergy Clin N Am. 2006;26(3):451–63.
- 67. Keyzer JJ, de Monchy JG, van Doormaal JJ, van Voorst Vader PC. Improved diagnosis of mastocytosis by measurement of urinary histamine metabolites. N Engl J Med. 1983;309(26):1603–5.
- Valent P, Akin C, Arock M, Brockow K, Butterfield JH, Carter MC, et al. Definitions, criteria and global classification of mast cell disorders with special reference to mast cell activation syndromes: a consensus proposal. Int Arch Allergy Immunol. 2012;157(3):215–25.
- 69. Garvey MDLH, Bech MDB, Mosbech MDDMSH, et al. Effect of general anesthesia and orthopedic surgery on serum tryptase. Anesthesiology. 2010;112(5):1184–9.
- Mayorga C, Celik G, Rouzaire P, Whitaker P, Bonadonna P, Rodrigues-Cernadas J, et al. In vitro tests for drug hypersensitivity reactions. An ENDA/EAACI Drug Allergy Interest Group position paper. Allergy. 2016;71(8):1103–34.
- 71. Vitte J. Human mast cell tryptase in biology and medicine. Mol Immunol. 2015;63(1):18–24.
- 72. Schwartz LB, Min HK, Ren S, Xia HZ, Hu J, Zhao W, et al. Tryptase precursors are preferentially and spontaneously released, whereas mature tryptase is retained by HMC-1 cells, mono-mac-6 cells, and human skin-derived mast cells. J Immunol. 2003;170(11):5667–73.
- 73. Schwartz LB, Yunginger JW, Miller J, Bokhari R, Dull D. Time course of appearance and disappearance of

human mast cell tryptase in the circulation after anaphylaxis. J Clin Invest. 1989;83(5):1551–5.

- 74. Lin RY, Schwartz LB, Curry A, Pesola GR, Knight RJ, Lee HS, et al. Histamine and tryptase levels in patients with acute allergic reactions: an emergency department– based study. J Allergy Clin Immunol. 2000;106(1, Part 1):65–71.
- 75. Williams KW, Sharma HP. Anaphylaxis and urticaria. Immunol Allergy Clin N Am. 2015;35(1):199–219.
- Stephan V, Zimmermann A, Kuhr J, Urbanek R. Determination of N-methylhistamine in urine as an indicator of histamine release in immediate allergic reactions. J Allergy Clin Immunol. 1990;86(6 Pt 1):862–8.
- 77. Greenberger PA, Ditto AM. Chapter 24: anaphylaxis. Allergy Asthma Proc. 2012;33(Suppl 1):S80–3.
- Berroa F, Lafuente A, Javaloyes G, et al. The usefulness of plasma histamine and different tryptase cut-off points in the diagnosis of peranaesthetic hypersensitivity reactions. Clin Exp Allergy: J Br Soc Allergy Clin Immunol. 2014;44(2):270–7.
- Blanca M, Mayorga C, Torres MJ, Reche M, Moya C, Rodriguez JL, et al. Clinical evaluation of Pharmacia CAP System RAST FEIA amoxicilloyl and benzylpenicilloyl in patients with penicillin allergy. Allergy. 2001;56(9):862–70.
- Torres MJ, Padial A, Mayorga C, Fernandez T, Sanchez-Sabate E, Cornejo-Garcia JA, et al. The diagnostic interpretation of basophil activation test in immediate allergic reactions to betalactams. Clin Exp Allergy : J Br Soc Allergy Clin Immunol. 2004;34(11):1768–75.
- 81. Salas M, Fernandez-Santamaria R, Mayorga C, et al. Use of the basophil activation test may reduce the need for drug provocation in amoxicillin-clavulanic allergy. J Allergy Clin Immunol Pract. 2017;
- 82. Mayorga C, Dona I, Perez-Inestrosa E, Fernandez TD, Torres MJ. The value of in vitro tests to diminish drug challenges. Int J Mol Sci. 2017;18(6):1222.
- De Weck AL, Sanz ML, Gamboa PM, et al. Diagnosis of immediate-type beta-lactam allergy in vitro by flowcytometric basophil activation test and sulfidoleukotriene production: a multicenter study. J Investig Allergol Clin Immunol: Off Organ Int Assoc Asthmol. 2009;19(2):91–109.
- Gamboa PM, Garcia-Aviles MC, Urrutia I, Antepara I, Esparza R, Sanz ML. Basophil activation and sulfidoleukotriene production in patients with immediate allergy to betalactam antibiotics and negative skin tests. J Investig Allergol Clin Immunol : Off Organ Int Assoc Asthmol. 2004;14(4):278–83.
- 85.• Aberer W, Bircher A, Romano A, et al. Drug provocation testing in the diagnosis of drug hypersensitivity reactions: general considerations. Allergy. 2003;58(9):854–63.

In this paper, EAACI reccommendations for drug provocation testing in the diagnosis of drug hypersensitivity are provided.

86.• Fernandez TD, Torres MJ, Blanca-Lopez N, et al. Negativization rates of IgE radioimmunoassay and basophil activation test in immediate reactions to penicillins. Allergy. 2009;64(2):242–8. In this review, author demostrate that levels of specific IgE antibodies tended to decrease over time in patients with immediate allergic reactions to amoxicillin; therefore, the diagnostic procedure must be done as soon as possible after the reaction.

- 87. Qiao HL, Yang J, Zhang YW. Relationships between specific serum IgE, cytokines and polymorphisms in the IL-4, IL-4Ralpha in patients with penicillins allergy. Allergy. 2005;60(8):1053–9.
- Messaad D, Sahla H, Benahmed S, Godard P, Bousquet J, Demoly P. Drug provocation tests in patients with a history suggesting an immediate drug hypersensitivity reaction. Ann Intern Med. 2004;140(12):1001–6.
- 89. Muraro A, Roberts G, Worm M, et al. Anaphylaxis: guidelines from the European Academy of Allergy and Clinical Immunology. Allergy. 2014;69(8):1026–45.

In this position paper by EAACI, guidelines about the management of anaphylaxis are provided.

- 90. Cardona V CN, Chivato T, De la Hoz B, Fernández Rivas M, Gangoiti Goikoetxea I, Guardia P, Herranz Sanz MA, Juliá Benito JC, Lobera Labairu T, Praena Crespo M, Prieto Romo, JI SSC, Sánchez JI, Uixera Marzal S, Vega A, Villarroel P. Guía de actuación en anafilaxia: GALAXIA 2016. Fundación SEAIC 2016.
- 91. Sheikh A, Sheikh Z, Roberts G, Muraro A, Dhami S, Sheikh A. National clinical practice guidelines for food allergy and anaphylaxis: an international assessment. Clin Trans Allergy. 2017;7:23.
- 92. Dhami S, Panesar SS, Roberts G, Muraro A, Worm M, Bilò MB, et al. Management of anaphylaxis: a systematic review. Allergy. 2014;69(2):168–75.
- 93. Pichichero ME, Casey JR. Safe use of selected cephalosporins in penicillin-allergic patients: a meta-analysis. Otolaryngol—Head Neck Surg: Off J Am Acad Otolaryngol-Head Neck Surg. 2007;136(3):340–7.
- Romano A, Gaeta F, Valluzzi RL, Caruso C, Rumi G, Bousquet PJ. The very limited usefulness of skin testing with penicilloyl-polylysine and the minor determinant mixture in evaluating nonimmediate reactions to penicillins. Allergy. 2010;65(9):1104–7.
- Cernadas JR, Brockow K, Romano A, Aberer W, Torres MJ, Bircher A, et al. General considerations on rapid desensitization for drug hypersensitivity—a consensus statement. Allergy. 2010;65(11):1357–66.
- Frumin J, Gallagher JC. Allergic cross-sensitivity between penicillin, carbapenem, and monobactam antibiotics: what are the chances? Ann Pharmacother. 2009;43(2):304–15.
- 97. Liu A, Fanning L, Chong H, et al. Desensitization regimens for drug allergy: state of the art in the 21st century. Clin Exp Allergy: J Br Soc Allergy Clin Immunol. 2011;41(12):1679–89.
- Sullivan TJ, Yecies LD, Shatz GS, Parker CW, Wedner HJ. Desensitization of patients allergic to penicillin using orally administered beta-lactam antibiotics. J Allergy Clin Immunol. 1982;69(3):275–82.
- 99. Wendel GD Jr, Stark BJ, Jamison RB, Molina RD, Sullivan TJ. Penicillin allergy and desensitization in

serious infections during pregnancy. N Engl J Med. 1985;312(19):1229–32.

- 100. Lee CW, Matulonis UA, Castells MC. Rapid inpatient/ outpatient desensitization for chemotherapy hypersensitivity: standard protocol effective in 57 patients for 255 courses. Gynecol Oncol. 2005;99(2):393–9.
- 101. Castells MC, Tennant NM, Sloane DE, Ida Hsu F, Barrett NA, Hong DI, et al. Hypersensitivity reactions to chemotherapy: outcomes and safety of rapid desensitization in 413 cases. J Allergy Clin Immunol. 2008;122(3):574–80.
- 102. Castells M. Desensitization for drug allergy. Curr Opin Allergy Clin Immunol. 2006;6(6):476–81.