**ECOLOGY & BIOGEOGRAPHY - ORIGINAL ARTICLE**



# **The invader shrub** *Chrysanthemoides monilifera* **(boneseed) negatively impacts native plant communities in a Mediterranean zone in Central Chile**

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#### **Abstract**

Biological invasions have severe social, economic and ecological repercussions. Central Chile is a biodiversity hot spot but also a highly disturbed area containing many invasive plant species such as *Chrysanthemoides monilifera* (L.) Norl. This shrub is present close to the coast and is considered an aggressive invasive species with allelopathic efects in other countries. Despite being ubiquitous, its invasive potential and ecological impact has not yet been evaluated in Chile. Here, we aim to determine the efect of the invasive *C. monilifera* on local plant communities and test for possible allelopathic efects on the germination of native and other invasive species. To do this, we analyzed plant diversity in patches with and without the invader in two sites in Valparaíso, Chile. Additionally, we conducted an experiment testing the efect of diferent concentrations (0, 25, 50 and 100%) of the aqueous extract of the leaves on the germination of both a native and an invasive shrub. We found that *C. monilifera* negatively impacts local plant diversity, especially endemic species. Also, its leaf extract inhibited the germination of the native shrub, especially at higher concentrations (50 and 100%), but do not inhibit the germination of the invasive shrub. This will likely result in a rapid change in the plant community, with negative impacts on the native species and an increase in invasive and introduced species, further degrading this already altered ecosystem. Developing strategies for the control of *C. monilifera* are urgent to limit its spread and negative ecological impact in Chile.

**Keywords** Allelopathy · Biological invasions · Boneseed · Invasive species

# **1 Introduction**

Invasive species are a main component of global change (GC) that has become a major threat for biodiversity worldwide (Hobbs [2000](#page-6-0); Mack et al. [2000](#page-7-0); Bradley et al. [2011](#page-6-1)). Biological invasions not only have ecological impacts, but can have social and economic repercussions as well (Mack

 $\boxtimes$  Cristian Atala cristian.atala@pucv.cl et al. [2000](#page-7-0); Shackleton et al. [2018\)](#page-7-1). In Chile, invasive plant species are a serious threat to biodiversity (Arroyo et al. [2000](#page-6-2); Fuentes et al. [2008\)](#page-6-3), but there are no current specifc legislation to manage them. These species belong mainly to the Poaceae, Fabaceae or Asteraceae families, and many have been reported as invasive elsewhere (Fuentes et al. [2013\)](#page-6-4). Central Chile is where most introduced and naturalized species are found (Castro et al. [2005;](#page-6-5) Fuentes et al. [2020](#page-6-6)). This is mainly due to higher population density, agriculture, cattle and forestry, and other anthropic disturbances (Fuentes et al. [2008](#page-6-3); Figueroa et al. [2011](#page-6-7)). Moreover, Central Chile has been largely afected by deforestation and the original native vegetation only exists in very small and fragmented sites. Large areas have been replaced by exotic species like *Eucalyptus* spp., *Pinus* spp. and *Acacia* spp. (Becerra [2006\)](#page-6-8). All those species are highly pyrogenic, and added to the summer drought typical of this area, large fres of anthropogenic origin occur regularly, further afecting native fora (McWethy et al. [2018](#page-7-2)).

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The Valparaíso region in Central Chile is one of the most negatively afected zones in the country by the recent wildfres. These fres are, as mentioned above, not of natural origin (McWethy et al. [2018](#page-7-2)). It also has a long history of port activity and host many dangerous invasive species. Valparaíso has been invaded by *Genista monspessulana* (L.) L.A.S. Johnson, *Cytisus* sp., *Rosa* spp., *Rubus ulmifolius* Schott and *Ulex europaeus* L. (Fuentes et al. [2014\)](#page-6-9), among others. Ports are known to have a high risk of biological invasions due to the high traffic and trade with other parts of the world (Hulme [2021\)](#page-6-10). Additionally, fres promote the germination of introduced, often invasive, species and can negatively impact native and endemic plants (Pauchard et al. [2008;](#page-7-3) Gómez-González et al. [2009](#page-6-11); [2011;](#page-6-12) [2017\)](#page-6-13) which lack adaptations to survive intense fres. This can result in a positive feedback loop further increasing the likelihood of a fre in an area. For example, the shrub *Genista monspessulana*, an invasive Fabaceae present in Central Chile, regenerates vigorously after fres and this, in turn, increases the risk of a subsequent fre (Pauchard et al. [2008](#page-7-3)). Valparaíso is part of the Mediterranean biodiversity hot spot (Arroyo et al. [2008\)](#page-6-14) and includes many endemic plant species (Rodríguez et al. [2018\)](#page-7-4), some of them with severe conservation problems. In the disturbed areas close to Valparaíso and Viña del Mar, the vegetation is usually dominated by native shrubs and herbaceous species, including the critically endangered *Chloraea disoides* Lindl. (Atala et al. [2017](#page-6-15)) and the vulnerable endemic palm *Jubaea chilensis* (Rodríguez et al. [2018](#page-7-4)). Taking into account the frequent fres, the role of Valparaíso as a main Chilean port is essential to assess any further risk for the highly endemic biodiversity of Valparaíso, particularly the possible negative impact of new invasive plant species.

The shrub *Chrysanthemoides monilifera* (L.) Norl., commonly known as boneseed, is an Asteraceae native to South Africa. It is considered to be an aggressive invasive species in Australia (Adair [1992](#page-6-16)). It grows abundantly, forming dense canopies that negatively impact abundance and diversity of native species (Thomas et al. [2005](#page-7-5)). There is evidence that this plant thrives after fres (Bray [2006\)](#page-6-17) and its control could be very difficult due to its high seed output (personal observation). These traits put *C. monilifera* as a highly risky invader in Central Chile according to the criteria in Fuentes et al. ([2014\)](#page-6-9). To our knowledge, there are no records of how it was brought to Chile and established the frst populations, but it is likely that it may be related to port activity and/or tourism. This species is present in Valparaíso, especially in degraded areas. Its risk as an invader has not been evaluated nor the mechanisms related to its success. The native plant diversity of Valparaíso could be threatened by the presence of invasive

plant species such as *C. monilifera* and its impact should be evaluated in order to establish future control measures.

Invasive species usually thrive in invaded areas due to several mechanisms like the ability to show a wide range of phenotypes (Molina-Montenegro et al. [2013](#page-7-6)), high reproductive output, high competitive ability and the production of allelopathic compounds (Thorp and Lynch [2000;](#page-7-7) Rudman [2001](#page-7-8); Weiss et al. [2008](#page-8-0); Al-Harun et al. [2014\)](#page-6-18). These compounds negatively afect other plant species, reducing seed germination and/or seedling growth (i.e., Gómez-González et al. [2009;](#page-6-11) Al-Harun et al. [2014](#page-6-18)). Plants can introduce the allelochemicals into the environment through leaf lixiviation, through root exudates or through the incorporation of plant residues in the soil (Inderjit and Duke [2003\)](#page-6-19). *C. monilifera* has been shown to have an allelopathic effect on *Lactuca sativa* L. and *Isotoma axillaris* Lindl. (Al-Harun et al. [2014](#page-6-18)), possibly due to a high phenols concentration. Aqueous extract of *C. monilifera* reduced germination in these species, even at low concentrations (Al-Harun et al. [2014](#page-6-18)). This high allelopathic effect could partially explain the high invasiveness of this species and its success in Valparaíso, especially in disturbed areas.

The present study aims to assess the impacts of an invasive species on the local plant community. To achieve this, we characterized the plant community where *C. monilifera* is present and compared its diversity with patches without the invader. Additionally, we tested the existence of a possible allelopathic efect that this invasive plant could have on a coexisting native and exotic shrub species. This information could be essential for future management and control initiatives and for the conservation of the unique biodiversity of Valparaíso.

## **2 Materials and methods**

**Efect of** *C. monilifera* **on plant diversity** The study site was located in Valparaíso Region and was selected on the basis of the presence of *C. monilifera* subsp. *monilifera* and accessibility. This site was previously studied and is the natural habitat of some native orchids, including species classifed as critically endangered (Atala et al. [2017](#page-6-15)) by Ministry of Environment (Ministerio del Medio Ambiente [2011](#page-7-9)) from Chilean government. The site is anthropically disturbed and dominated by shrubs and with abundant herbaceous cover, especially in the spring. Here, patches with  $(Chr+)$  and without (Chr–) *C. monilifera* were selected and 1 × 1 m randomly put plots were used to study plant diversity and abundance following standard methods (see Atala et al. [2017](#page-6-15)). Sampling was conducted in late spring to ensure most plant species are in the reproductive stage in order to facilitate identifcation. We applied a simulation-based sampling protocol to assess the adequacy of our sampling effort for plots

with the presence of *C. monilifera* and without the species, as implemented in the SSP package (v. 1.0.1, Guerra-Castro et al. [2021\)](#page-6-20) in R (R Core Team [2022\)](#page-7-10). A rarefaction analysis was used to estimate sample completeness. Rarefaction analysis compares the accumulated number of species as a response to plot number (Chen et al. [2014](#page-6-21); Chao et al. [2020](#page-6-22)). When the curve becomes asymptotical, it is assumed that all (or almost all) species diversity in that site has been recorded, and that it determines the minimal number of plots required to appropriately estimate such diversity. In each plot, we registered vascular plant species present. A diversity index and rarefaction analyses were used to compare plant diversity between patches in both sites. We also compared the % of native and introduced plant species in diferent patches (with and without *C. monilifera*). Rarefaction were constructed in R using the *iNEXT* package (Hsieh et al. [2016](#page-6-23)). This package allows extrapolation of sample completeness, while estimating standard error. Rarefaction analyses were run with 1000 bootstrap pseudoreplicates with 95% confdence interval.

**Allelopathic efect of aqueous extract of** *C. monilifera* Leaves of *C. monilifera* were collected from the feld in "Palmar el Salto", Valparaíso (33°3′58.1″ S, 71°31′54.6″ W). We collected leaves of the invasive plant from at least 10 different individuals and were then pooled together. Samples were then taken to the laboratory, dried at room temperature for 48 h and then grounded with a manual mixer, adding 300 mL of distilled water per 300 g of leaves to facilitate the grounding. The obtained solution was strained using a metal strainer and then fltered using flter paper. The fnal fltered solution was stored in glass fasks in the dark at 4 °C for the subsequent experiment. From the stock solution, 3 dilutions with sterile distilled water were used in the experiment, 100, 50 and 25% of the obtained fltered solution. Additionally, we used a control of only distilled water (0% stock solution). All of these solutions, and the control, were stored as described above.

From natural populations, seeds of the native shrub, *Baccharis linearis* (Ruiz & Pav.) Pers., and the invader tree, *G. monspessulana*, were collected, aiming for at least 100 seeds per species from individuals near *C. monilifera*. We chose these species because both coexist in the same habitat where *C. monilifera* is found. Additionally, both species are shrubs and relatively similar in size to *C. monilifera*. This allowed us to assess potential allelopathic efects between exotic and native species with the same growth habit, and compare a model native and introduced species. Seeds were superfcially disinfected by immersion in a 70% ethanol solution for 30 s, followed by immersion in a 1% NaOCl solution with two drops of Tween 20 for 5 min under constant agitation (see Pereira et al. [2015](#page-7-11)). This was done to avoid external fungal contamination. Then, seeds were washed 5 times with sterile distilled water. Seeds, then, were put in distilled water for 12 h to initiate imbibition. Afterward, 20 seeds of *B. linearis* and 30 seeds of *G. monspessulana* were put in Petri dishes with tissue paper at the bottom and were assigned to one of the following treatments: (1) control treatment, where dishes were watered with 1 mL of distilled water every 2 days; (2) 100% where dishes were watered with 1 mL of the 100% extract solution; (3) 50%, where plates were watered with 1 mL of the 50% solution v/v; and 4) 25%, where dishes were watered with 1 mL of the 25% solution v/v. We used 25 Petri dishes for *B. linearis* per treatment and 27 for *G. monspessulana* per treatment (i.e., *n*=25 and 27, respectively). Petri dishes were put in the laboratory at room temperature and natural light. Seed germination was recorded every 2–3 days until 41 days. After that time, no further germination was detected.

Initial exploration of germination data under the diferent treatments was carried out with a time to event analysis, which has been used in other seed germination studies (Onofri et al. [2011;](#page-7-12) Ritz et al. [2013](#page-7-13)[2019\)](#page-7-14). The data were analyzed using the R environment (version 4.2.0, R Core Team [2022](#page-7-10)). For each Petri dish, the response was defned as the daily proportional cumulative germination curve (propCum), expressed as the number of germinated seeds. The analysis was conducted using the diferent treatments as predictors against germination. The "time event approach" is available in *drcSeedGerm* R package (Onofri et al. [2018\)](#page-7-15), and the function *makeDrm* was used to calculate the response. The distributional assumption for the germination times considered was the three-parameter log-logistic, a shifted log-logistic distribution (Ritz et al. [2015\)](#page-7-16) where ED50 was used as a parameter with lower limit at 0. We compared the curves per treatment using nonparametric Kruskal–Wallis one-way analysis of variance using *stats* R package, as our data were not normal distributed (Shapiro–Wilk normality test,  $p < 0.001$ ). This was followed by Dunn test post hoc analysis, available from *FSA* R library (Ogle et al. [2022](#page-7-17)). Differences were considered significant at  $p < 0.05$ .

### **3 Results**

*C. monilifera* **negatively impacts on a natural plant community** The invasive *C. monilifera* negatively impacted plant diversity in patches where it was present  $(Chr+)$  compared to patches without the species (Chr−) in the same site (Fig. [1](#page-3-0)). This was also evident when comparing Shannon–Weaver diversity index, 2.92 vs 3.41 in Chr+and Chr− patches, respectively (Table [1\)](#page-3-1). Additionally, Chr+patches had a higher proportion of introduced species and a lower proportion of native and endemic species, compared to Chr− patches (Table [1](#page-3-1)). The differences in the proportion of introduced and endemic species were statistically

<span id="page-3-0"></span>**Fig. 1** Interpolated and extrapolated rarefaction curves in sites with the presence of the invasive shrub, *C. monilifera* (Chr+) and without *C. monilifera* (Chr−). Shaded areas indicate the 95% standard error confdence intervals



<span id="page-3-1"></span>**Table 1** Shannon–Weaver diversity index, most common plant family and % of introduced, native and endemic species, in patches with the invasive shrub, *C. monilifera* (Chr+), and without *C. monilifera* (Chr−) in a site in Central Chile



\*z-score test,  $p < 0.05$ , \*\* *z*-score test,  $p < 0.01$ 

significant between patches ( $z$ -score test,  $p < 0.05$ , Table [1](#page-3-1)). No statistical diferences were found in the proportion of native species between Chr+and Chr− patches (*z*-score test,  $p > 0.05$ , Table [1\)](#page-3-1). In both patches, most species belonged to the Asteraceae family (Table S1), including *B. linearis*, followed by Poaceae.

Rarefaction curves showed that both conditions (Chr+and Chr−) reach the asymptote within a 95% confdence interval, but with an overlap in their extrapolated confdence, modeled up to 80 sampling sites in each condition. These results also shows that Chr+sites present a lower total richness, with an observed total of 32 species and an estimated of ~ 44 species, with a standard error of 9.5 species and a sample coverage of ~ 94%. On the other hand, Chr− sites showed a total of 42 species observed and an estimated of  $\sim$  50 species, with a standard error of  $\sim$  6 species and a sample coverage of ~93%. These results are supported by our simulated sampling (data not shown), indicating that with an average of 18 samples points per conditions we can reach a descriptive precision above 67% for either community.

**Allelopathic efect of** *C. monilifera* A log-logistic model was ftted to compare time to event analysis, which showed statistical diferences in treatment efect in all models for *B. linearis* and *G. monspessulana* (Table S2, Time to event analysis,  $p < 0.001$  in both cases). Our results showed that an increase in the solution concentration negatively afected the germination in *B. linearis* (Fig. [2](#page-4-0)a). Although a 25% solution was not signifcantly diferent from the control, the addition of a 50% or 100% solution of leaf extract signifcantly reduced germination (Dunn test,  $p < 0.001$ , Table [2](#page-4-1)). On the other hand, we observed no negative impact on *G. monspessulana* germination and even a slight positive efect (Fig. [2b](#page-4-0)). All treatments have higher germination compared with the control (Table [2,](#page-4-1) Dunn test,  $p < 0.001$ ), but there were no signifcant diferences between diferent concentrations of the extract (Dunn test,  $p > 0.05$ ). Finally, after close to 40 days, seeds did not further germinate in any treatment for both species.



<span id="page-4-0"></span>**Fig. 2** Time course of cumulative germination for all treatments: **a** native shrub *B. linearis*; and **b** invasive shrub *G. monspessulana*. \*Represent treatments with signifcant diferences compared to control (Dunn test,  $p < 0.001$ ), and ns indicates not significant (Dunn test,  $p > 0.05$ 

<span id="page-4-1"></span>**Table 2** Dunn test parameters for native shrub, *B. linearis* (a), and invasive shrub, *G. monspessulana* (b); *p* values are adjusted to Benjamini–Hochberg correction method

Comparison	z-value	$p$ -value
$(a)$ B. linearis		
$0 - 100$	16.848	< 0.001
$0 - 50$	11.150	< 0.001
$0 - 25$	0.952	0.341
$100 - 25$	$-15.896$	< 0.001
$100 - 50$	$-5.699$	< 0.001
$25 - 50$	10.198	< 0.001
(b) G. monspessulana		
$0 - 100$	$-5.506$	< 0.001
$0 - 50$	$-5.594$	< 0.001
$0 - 25$	$-4.823$	< 0.001
$100 - 25$	0.684	0.593
$100 - 50$	$-0.088$	0.930
$25 - 50$	$-0.772$	0.604

#### **4 Discussion**

Invasive species are a severe threat to global biodiversity (Bradley et al. [2011](#page-6-1); Linders et al. [2020](#page-7-18); Pyšek et al. [2020](#page-7-19)), but despite their ecological and socioeconomic impact they are seldom considered in predictive global models (Roura-Pascual et al. [2021](#page-7-20)). Here we showed that *C. monilifera* can negatively impact the plant community when present in a site in Central Chile. To our knowledge, this is the frst report on the impact of this invasive species in Chile. Particularly, patches with the invasive species have a lower richness and a lower proportion of endemic species and higher proportion of introduced species. There is evidence of the negative impact of invasive species on biodiversity (Pyšek et al. [2012](#page-7-21); Haider et al. [2018;](#page-6-24) Pauchard et al. [2018](#page-7-22)). However, some authors suggest that the impact of invasive plants on native plant communities is low and/or very slow, even that invasive species can add to the overall species pool of a region, increasing local diversity (Houlahan and Findlay [2004;](#page-6-25) Ellis et al. [2012](#page-6-26); Gilbert and Levine [2013;](#page-6-27) Thomas and Palmer [2015](#page-7-23); Corlett [2016\)](#page-6-28). This may likely depend on the spatial scale and on the particular site.

Our results indicate a negative impact at a local scale in Central Chile. Moreover, among the introduced species present in patches with *C. monilifera* are other invasive plants such as *G. monspessulana*, which may further degrade the community. In fact, patches with the invader looked very homogeneous compared to patches without *C. monilifera* (personal observation). Furthermore, the presence of invasive species reduced the plant richness, as seen in the comparison between patches with  $(Chr+)$  and without (Chr−) *C. monilifera*. Endemic species in particular were more severely affected compared to native species. Endemic species were represented by less than 10% of the total richness in Chr+patches, whereas endemic species richness was close to 25% in Chr− patches. This is consistent with other studies where invasive species reduce the richness in the communities where they live, specially impacting endemic and/or rare species (Thomson [2005;](#page-7-24) Hardman et al. [2012](#page-6-29)). In addition, the presence of native and endemic species in patches without *C. monilifera* may serve as a barrier for the introduction of this plant (see Reinhardt Adams and Galatowitsch [2008](#page-7-25); Vargas-Gaete et al. [2018](#page-8-1)). On the other hand, the study site show evidence of frequent disturbances (see Atala et al. [2017](#page-6-15)). It is likely that disturbances, such as fres and/or cattle, can alter patches without the invader, allowing seeds to germinate and slowly colonize new patches. We suggest that this species should be monitored to avoid its spread into nearby protected areas, where many unique endemic plant species are found, being a biodiversity hot spot and a biosphere reserve (Arroyo et al. [2008](#page-6-14); Moreira-Muñoz and Borsdorf [2014](#page-7-26)). This could have severe consequences for the conservation of these endemic species, which can have restricted distributions and small populations. Such is the case of *Chloraea disoides*, a critically endangered endemic species recorded close to the study site which has few existing populations (Atala et al. [2017\)](#page-6-15).

Leaf aqueous extract of *C. monilifera* inhibited the germination of a native shrub (*B. linearis*), especially at higher concentration of the extract. Low concentrations (25%) had no significant effect on seed germination on laboratory conditions. Previous studies have shown the allelopathic potential of this invasive species, reducing the germination of other plant species (Al-Harun et al. [2014,](#page-6-18) [2015a](#page-6-30)). This was attributed to a high phenolic content of the plant tissues (Al-Harun et al. [2014\)](#page-6-18). This effect seems to be dose-dependent and is stronger in roots and leaves, compared to stems (Al-Harun et al. [2015b](#page-6-31)). Other invasive species present in Central Chile, such as *Cytisus scoparius* (L.) Link and *Centaurea solstitialis* L., can also have allelopathic effects on native species (Gómez-González et al. [2009;](#page-6-11) Grove et al. [2012](#page-6-32)). These mechanisms could explain the success of *C. monilifera* in coastal Central Chile and should be taken into account for future control initiatives. However, direct application of aqueous extract may not exactly mimic natural conditions in the soil and further studies should be conducted in the feld and laboratory using *C. monilifera* litter to confrm its allelopathic efect on *Baccharis* spp. and other native Chilean plants. In the feld, below *C. monilifera* individuals, there are large amounts of shed leaves, forming a thick layer in the soil. This could also form a physical barrier, as well as a chemical barrier, for the germination of native species. Photoblastic seeds may not receive sufficient light to germinate, added to the litter weight and potential chemical inhibition of the seeds. Some Asteraceae species have been reported as having photosensitive seeds (Schutz et al. [2002](#page-7-27); Luo and Cardina [2012](#page-7-28)), varying in their response to light with seed traits such as seed weight and size (Schutz et al. [2002](#page-7-27)). In the feld, only *C. monilifera* seedlings are found under the canopy of the invader (personal observation, data not shown). Thus, further studies are required to unravel the full extent of the efect of *C. monilifera* litter on the soil and on other plant species.

Contrary to the native shrub, leaf extract of *C. monilifera* did not afect and even slightly promoted the germination of the invasive shrub *G. monspessulana*. This negative effect on native species and neutral (or even positive) effect on other invasive species could severely deteriorate the plant community, resulting in an invasion meltdown (see Simberloff and Von Holle [1999;](#page-7-29) Collins et al. [2020](#page-6-33)). Other authors, however, disagree with the concept and use the term "secondary invasion" (O'Loughlin and Green [2017\)](#page-7-30) for a facilitated invasion of an invader species due to the previous presence of another. The obtained germination % in *G. monspessulana* was relatively low, close to 10%. In this species, fre can increase germination

to 40–50%, whereas unburnt seeds reach only 10% (García et al. [2010](#page-6-34)), which may explain the obtained low germination. There is evidence that fre also can increase germination of *C. monilifera* (Bray [2006\)](#page-6-17). Anthropically caused wildfres have afected Central Chile, and Valparaíso in particular, especially during the dry summers (Peña and Valenzuela [2008;](#page-7-31) Atala et al. [2017\)](#page-6-15). These fres could have further negative impacts on the non-adapted native species (Gómez-González and Cavieres [2009](#page-6-35)) and potentiate the increase and dominance of invasive species such as *C. monilifera* and *G. monspessulana*.

Central Chile includes a Mediterranean biodiversity hot spot (Arroyo et al. [2008\)](#page-6-14) that has been largely altered by human activity. Here many unique endemic plant species can be found that are vulnerable to these anthropic disturbances and the possible negative efects of plant invasions. Based on our results, we suggest that urgent actions are needed to control the spread of *C. monilifera* in the country. This, especially considering that it can have a negative impact of endemic fora, already threatened by urban development, agriculture, wildfres, and other plant and animal invasive species (Castro et al. [2005;](#page-6-5) Fuentes et al. [2008;](#page-6-3) Pauchard et al. [2008;](#page-7-3) Figueroa et al. [2011;](#page-6-7) Gómez-González et al. [2011](#page-6-12)). Since its distribution in Chile is still relatively small, we suggest that manual control can be implemented as a frst step to manage this invader, especially close to protected natural areas such as the biosphere reserve La Campana–Peñuelas (Moreira-Muñoz and Borsdorf [2014\)](#page-7-26). In the mid-long term, other strategies such as biological control or the use of herbicides could also be applied to control its spread as used in other countries such as Australia (Brougham [2006](#page-6-36); Melland and Preston [2008](#page-7-32); Melland [2009\)](#page-7-33). Additionally, future studies could also address possible efects of this invader on soil microorganisms, its regeneration after the frequent summer fires of the areas and its possible effects on pollinators and other animals that could indirectly impact the native and endemic flora.

**Supplementary Information** The online version contains supplementary material available at<https://doi.org/10.1007/s40415-023-00905-9>.

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**Author contribution** CA was involved in conceptualization, resources, writing—original draft and writing—reviewing and editing. SAR was responsible for formal analysis, data curation, writing—reviewing and editing, and visualization. JO carried out investigation and data curation. OJ-C participated in data curation, investigation and visualization. RV contributed to formal analysis, writing—reviewing and editing, and visualization.

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