

Reversal Strategies for NOACs: State of Development, Possible Clinical Applications and Future Perspectives

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Abstract The non-vitamin K antagonist oral anticoagulants (NOACs) are used for thromboembolic prophylaxis of patients with atrial fibrillation and in the treatment as well as secondary prophylaxis of patients with venous thromboembolism. Even though NOACs have a better safety profile than vitamin K antagonists (VKAs), there will still be bleeding complications on NOAC treatment. In some cases, stopping the NOAC and non-drug-related management such as manual compression and interventional endoscopy will be sufficient to stop the bleeding. In more serious bleeding events and before acute surgery, coagulation factor concentrates or NOAC-specific antidotes could be used. Coagulation factor concentrates can be used in patients with haemophilia and to reverse the effect of VKAs but, in NOAC-treated patients, results are inconsistent and these agents could potentially have pro-thrombotic effects. Specific antidotes for NOACs are expected to be on the market soon. Phase III clinical trials with a humanized antibody fragment directed against dabigatran (idarucizumab) and recombinant, modified factor Xa (andexanet alfa) are ongoing. A molecule (aripazine) with broad activity against various anticoagulants including NOACs is currently undergoing phase II trials. For use of these specific antidotes, it is desirable that measurements for coagulation activity with a short response delay are

widely available for the different NOACs and further research in this field is needed. Furthermore, guidelines for antidote use, including general measures for the treatment of NOAC-related bleeding, should be available.

Key Points

Specific antidotes for non-vitamin K antagonist oral anticoagulants (NOACs) are expected to be available on the market soon.

Clinical trials are ongoing with idarucizumab (against dabigatran), andexanet alfa (against all factor Xa inhibitors) and aripazine (against all NOACs, fondaparinux and heparin).

Indications for use of NOAC antidotes are serious bleeding complications and before acute surgery in NOAC-treated patients.

1 Introduction

The non-vitamin K antagonist oral anticoagulants (NOACs) are used as alternatives to the vitamin K antagonists (VKAs) in thromboembolic prophylaxis of patients with atrial fibrillation (AF) and in the treatment as well as secondary prophylaxis of patients with venous thromboembolism (VTE) [1]. Treatment with NOACs has reached the guidelines in Europe for these indications and also for AF in the US.

On the market for AF and VTE are apixaban (Eliquis[®]), rivaroxaban (Xarelto[®]), dabigatran etexilate (Pradaxa[®]) and edoxaban (Lixiana[®]). Of importance for the use of

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NOACs are effective treatment options in case of serious bleeding complications on treatment, and to stop the treatment effect fast in case of a need for an operation or another procedure with a high risk of bleeding. At the moment, there is no registered antidote for these drugs, but several antidotes are under development. NOACs have a relatively short half-life in patients with normal kidney function, and therefore stopping therapy is part of the strategy in these critical situations.

The present review gives the status for the development of antidotes for NOACs, discusses the potential clinical applications and provides perspectives for the use of NOACs once the antidotes are available.

2 Pharmacological Properties of Non-Vitamin K Antagonist Oral Anticoagulants (NOACs)

Dabigatran etexilate is a prodrug that is rapidly converted by liver and plasma esterases to dabigatran, which is a competitive and reversible inhibitor of free and clot-bound thrombin. The half-life is 12–17 h and 80 % is excreted unchanged by the kidneys.

Rivaroxaban, apixaban and edoxaban are selective, competitive and direct inhibitors of both free factor Xa (FXa) and FXa associated with the prothrombinase complex, decreasing the activation of prothrombin to thrombin. The half-life of rivaroxaban is 7–11 h and 33 % is excreted

unchanged by the kidneys. The half-life of apixaban is 12 h and 27 % of the drug is renally excreted unchanged. For edoxaban, the half-life is 9–11 h and 50 % is renally excreted unchanged.

3 Measurement of Plasma Concentrations

The golden standard for measurement of plasma concentrations of NOACs is liquid chromatography tandem mass-spectrometry (LC-MS/MS) [2]. Currently, the restricted availability of the test is one of the drawbacks, especially in acute situations [3].

Expected therapeutic and toxic plasma concentrations for each NOAC are shown in Table 1. Peak levels are measured 1–4 h after intake. Trough levels are measured 12 h after intake with twice daily dosing and 24 h after intake with once daily dosing, just before the next dose.

Increasing plasma concentrations of edoxaban and dabigatran showed increasing bleeding risk in clinical trials; however, these were limited by the fact that measurements of plasma concentration were not related to the time of the bleeding [4, 5]. There is evidence that NOAC plasma concentrations might not fully reflect bleeding risk, as major bleeding in some subgroups of patients (dabigatran: after implantation of a mechanical heart valve; edoxaban: in those with clinical risk factors for high bleeding risk such as reduced kidney function and body

Table 1 Plasma concentrations of NOACs

| | Dabigatran ^a | Rivaroxaban ^b | Apixaban ^c | Edoxaban ^d |
|----------------|---|--|---|---|
| Trough (ng/mL) | 110 mg BID: 65 (28–155) 150 mg BID: 91 (40–215) [5] | 10 mg QD: 9 (1–38) [53] 20 mg QD: 26 (6–87) [54] 20 mg QD: 22 (4–96) [53] | 2.5 mg BID: 32–97 5 mg BID: 63–103 [55, 56] 10 mg BID: 120 (41–335) [56] | 30 mg QD: 25 60 mg QD: 48 [4] |
| Peak (ng/mL) | Time after last dose: 1–3 h 110 mg BID: 126 (52–275) 150 mg BID: 175 (74–383) [5] | Time after last dose: 2–4 h 10 mg QD: 125 (91–195) [53] 20 mg QD: 270 (189–419) [54] 20 mg QD: 222 (160–360) [53] | Time after last dose: 1–3 h 2.5 mg BID: 67–123 5 mg BID: 132–171 10 mg BID: 251 (111–572) [56] | Time after last dose: 1–2 h 30 mg QD: 85 60 mg QD: 175 [57] |
| Toxic | Trough 210 ng/mL (major bleeding rate doubled) [5] | No clinical data available | No clinical data available | Trough ca. 110 ng/mL (major bleeding rate doubled) [4] |

BID twice daily, *NOACs* non-vitamin K antagonist oral anticoagulants, *QD* once daily

^a Mean plasma concentrations are shown with 10th to 90th percentiles in parentheses

^b Median predicted concentrations are shown with 5th to 95th percentiles in parentheses

^c Ranges for the median of the different studies available are shown. For the 10 mg BID dose, the median is shown with 5th and 95th percentiles in parentheses

^d Mean trough and median peak plasma concentrations are shown

weight under 60 kg) was not associated with increased plasma concentrations [4, 6]. A limitation that must be kept in mind is that the same plasma concentration can induce a variable degree of anticoagulation in different patients [7].

4 Coagulation Assays

Determining the anticoagulant activity of the drug using a coagulation assay is a more practical and possibly more accurate tool to assess a bleeding patient and the biological effect of the reversal agent. The ideal test is quick, cheap and widely available. The international normalized ratio (INR), prothrombin time (PT) and activated partial prothrombin time (APTT) are examples of such tests. Unfortunately, they are not suitable for quantitative evaluation of the anticoagulant activity of NOACs. Results of these assays vary according to the reagent used for each NOAC and the test is either insensitive or relatively insensitive already at plasma concentrations considered to be therapeutic [8]. The PT-derived INR was developed specifically for the monitoring of VKAs and cannot be used [9, 10].

Specific coagulation assays for the NOACs have been developed. These assays are currently not widely available but their use is becoming more prevalent. The Hemoclot[®] diluted Thrombin Time (dTT) has been advocated as the preferable assay for quantification of dabigatran [11]. The reagent for Hemoclot[®] dTT, however, requires an hour stabilization time before use according to the manufacturer's current recommendations [12], making this assay less suitable for use in acute clinical situations [13]. Other dTTs calibrated for dabigatran concentration are available, but clinical experience with these assays is currently lacking [14].

The Ecarin Clotting Time (ECT) and Ecarin Chromogenic Assay (ECA) have also shown strong correlations and a linear relationship with dabigatran plasma concentrations [15, 16]. The shorter incubation times of the reagents make them more useful for monitoring in acute situations. Limitations of ECT are its dependence on plasma fibrinogen and prothrombin levels [17]. These limitations are overcome by the ECA [18].

For FXa inhibitors, chromogenic anti-FXa assays with specific calibrators and controls are available and demonstrated strong correlations and linear relationships with plasma concentrations of the drug [7].

It has been suggested that Dilute Russell's Viper Venom Time (DRVV-T) can be used for monitoring of all NOACs [19], but this requires additional validation in clinical studies.

Because of large inter-individual variation in coagulation factor levels and therefore varying potency and activeness of clotting, global coagulation assays such as

Thrombin Generation Assay (TGA) and thromboelastography (TEG[®]/ROTEM[®]) are better at taking the complexity of the coagulation cascade into account [20] and may be more sensitive for testing the reversal of NOAC effects [21, 22]. Drawbacks of TGA include the relatively long duration of the test and inadequate standardization for broad clinical use [23]. Use of thromboelastography is promising in studies with spiked plasma samples but requires further investigation in clinical studies [21].

Using the information currently available, for quantitative monitoring in acute situations we recommend ECA for dabigatran and anti-Xa assays for the FXa inhibitors. Further research is needed to determine the relation between results of coagulation assays and clinical outcomes.

A summary of the available coagulation assays for monitoring NOACs is shown in Table 2.

5 Clinical Application of NOAC Antidotes

Major bleeding rates in patients treated with NOACs in clinical trials are lower in comparison with VKAs (5.3 vs 6.2 %) and a substantial reduction in the rate of intracranial haemorrhage was observed (0.7 vs 1.5 %) [1]. Data from NOAC registries show even lower bleeding rates [24, 25]. Nevertheless, bleeding events will still be an important issue in patients treated with NOACs.

It can be expected that NOAC antidotes will be used in relation to serious bleeding complications on treatment with these anticoagulants as a supplement to more general measures [26]. In addition, the antidotes may be used in case of urgent surgery and procedures with a high risk of bleeding related to ongoing anticoagulant treatment with NOACs.

It is, however, important to note that the location and size of the organ lesion leading to a bleeding complication while on a NOAC and the general condition of the patient may be the most important factors for prognosis and should be taken care of simultaneously with antidote administration.

It is important in relation to the ongoing clinical studies with antidotes to focus on the risk of thromboembolic complications when anticoagulant therapy is rapidly interrupted with antidotes in patients with a high thrombogenicity related to the condition leading to anticoagulant therapy and the clinical status related to the serious bleeding or a need for urgent surgery.

The experience so far from the limited number of dabigatran etexilate-treated patients receiving idarucizumab is that no biochemical indications of a prothrombotic state were observed [27], but a potential prognostic impact of the antidote in the treated cohort cannot be evaluated. The patients had a high mortality and a control group could not be included.

Table 2 Coagulation assays in patients treated with NOACs

| | Dabigatran etexilate | Rivaroxaban | Apixaban | Edoxaban |
|--------------------------------------|---|--|--|--|
| INR | Unsuitable | Unsuitable | Unsuitable | Unsuitable |
| PT | Unsuitable | Not suitable for quantitative measurement | Not suitable for quantitative measurement | Not suitable for quantitative measurement |
| APTT | Not suitable for quantitative assessment | Unsuitable | Unsuitable | Unsuitable |
| dTT | Linear relationship with plasma concentration. Hemoclot [®] has a 1 h-reagent stabilization time before use, less suitable in acute situations | Unsuitable | Unsuitable | Unsuitable |
| ECT | Linear relationship with plasma concentration Can be used | Unsuitable | Unsuitable | Unsuitable |
| ECA | Linear relationship with plasma concentration Recommended | Unsuitable | Unsuitable | Unsuitable |
| Anti-Xa chromogenic assay | Unsuitable | Linear relationship with plasma concentration Recommended | Linear relationship with plasma concentration Recommended | Linear relationship with plasma concentration Recommended |
| DRVV-T | Further studies required | Further studies required | Further studies required | Further studies required |
| TGA | Further studies required Long test time, not suitable in acute situations | Further studies required Long test time, not suitable in acute situations | Further studies required Long test time, not suitable in acute situations | Further studies required Long test time, not suitable in acute situations |
| ROTEM [®] /TEG [®] | Further studies required | Further studies required | Further studies required | Further studies required |

APTT activated partial prothrombin time, *DRVV-T* Dilute Russell's Viper Venom Time, *dTT* diluted Thrombin Time, *ECA* Ecarin Chromogenic Assay, *ECT* Ecarin Clotting Time, *INR* international normalized ratio, *NOACs* non-vitamin K antagonist oral anticoagulants, *PT* prothrombin time, *TGA* Thrombin Generation Assay

Issues of concern regarding antidotes for NOACs might be that availability of these agents could result in a more careless attitude to timing of invasive procedures and that the agents may be subject to overuse. Patients with a prescription for a NOAC may not have circulating anticoagulant in the blood [27] in relation to serious bleeding or the need for urgent surgery and, to avoid overutilization of antidotes and a delay in the use of other more relevant interventions [26], measurement of NOAC exposure in the specific situation must be available. It is desirable that measurement of coagulation activity using different methods dependent on the type of NOAC treatment and with a short response delay is available and can be established in hospitals responsible for handling bleeding and urgent surgery of patients on treatment with oral anticoagulants including NOACs [8, 28].

For patients with normal kidney function, accumulation of NOAC in the body is not expected to occur and a low plasma concentration can be expected 12 h after the last

dose. It is very important to have exact information on type of NOAC prescribed to the patient, responsible physician, indication for anticoagulant therapy and other medications with an effect on haemostasis. All patients treated with a NOAC should carry a card with this information [26].

There is no data to date evaluating a possible formation of antibodies to the antidotes leading to serious allergic reactions and/or neutralization of treatment effect on repeated antidote administration.

6 Coagulation Factor Concentrates

Prothrombin complex concentrate (PCC) can be used to reverse the effect of VKAs. In this situation, the coagulation factors in PCC substitute for the reduced concentration of coagulation factors during therapy with a VKA. It has been suggested that PCC can also be used to reverse the effect of rivaroxaban, as the administration of PCC 50 IU/

kg to healthy volunteers treated with a supratherapeutic dose of rivaroxaban (20 mg twice daily) rapidly normalized the prolonged PT [29]. In a similar Dutch study, the same effect was found on the PT, but a prolongation of the APTT was seen [30]. PCC contains a small amount of heparin to avoid coagulation factor activation during preparation and storage. The heparin content, especially when PCC is given at higher doses, could cause this APTT prolongation. Considering the prolongation of the APTT, it is questionable whether serious rivaroxaban-induced bleeding can be stopped using PCC.

The fact that PCC augments thrombin generation, however, suggests that it may have an effect [30].

When PCC is used in experiments to reverse the effect of NOACs, it is not a substitution therapy, and may also potentially lead to a prothrombotic state, as it induces a supra-physiological rise in the plasma concentration of coagulation factors that are not inhibited by the NOAC in use.

This situation is also different from the PCC treatment of patients with haemophilia that have developed an inhibitor to factor VIII or those missing another coagulation factor. For serious bleeding in these patients, PCC can be used to augment coagulation activity. For VKA-treated patients, a more potent effect can be obtained by activated PCC (Feiba[®]) or recombinant factor VIIa (FVIIa; NovoSeven[®]). For NOACs, consistent clinical data is lacking. Animal studies show conflicting results and in several studies with NOAC-induced bleeding, FVIIa and PCC improved results of coagulation assays but failed to improve haemostasis [31–33].

PCC, activated PCC and FVIIa should only be used in serious and potentially life-threatening bleeding complications during treatment with NOACs, as these patients, in contrast to patients with haemophilia, have a high risk of thromboembolic complications. Surgical and local-acting treatments should be considered and used if relevant and possible before the use of these products.

6.1 NOAC-Specific Antidotes

6.1.1 Idarucizumab

Idarucizumab is a monoclonal antibody fragment, which binds specifically to dabigatran with an affinity that is 350 times higher than is seen for thrombin [34]. Therefore, idarucizumab binds both free and thrombin-bound dabigatran and neutralizes its anti-thrombin activity [34]. The antidote has been tested in healthy young volunteers, in volunteers aged 65–80 years and in volunteers aged 45–80 years with mild or moderate renal impairment [35–37]. Idarucizumab reversed the anticoagulant effect of dabigatran immediately and completely with normalization of the Hemoclot[®] dTT without a procoagulant effect.

In a study including 30 pigs treated with dabigatran etexilate, idarucizumab significantly decreased total blood loss after blunt liver trauma and reduced mortality in comparison with a control group not treated with the antidote [38]. Whether these results are also applicable to humans has yet to be confirmed.

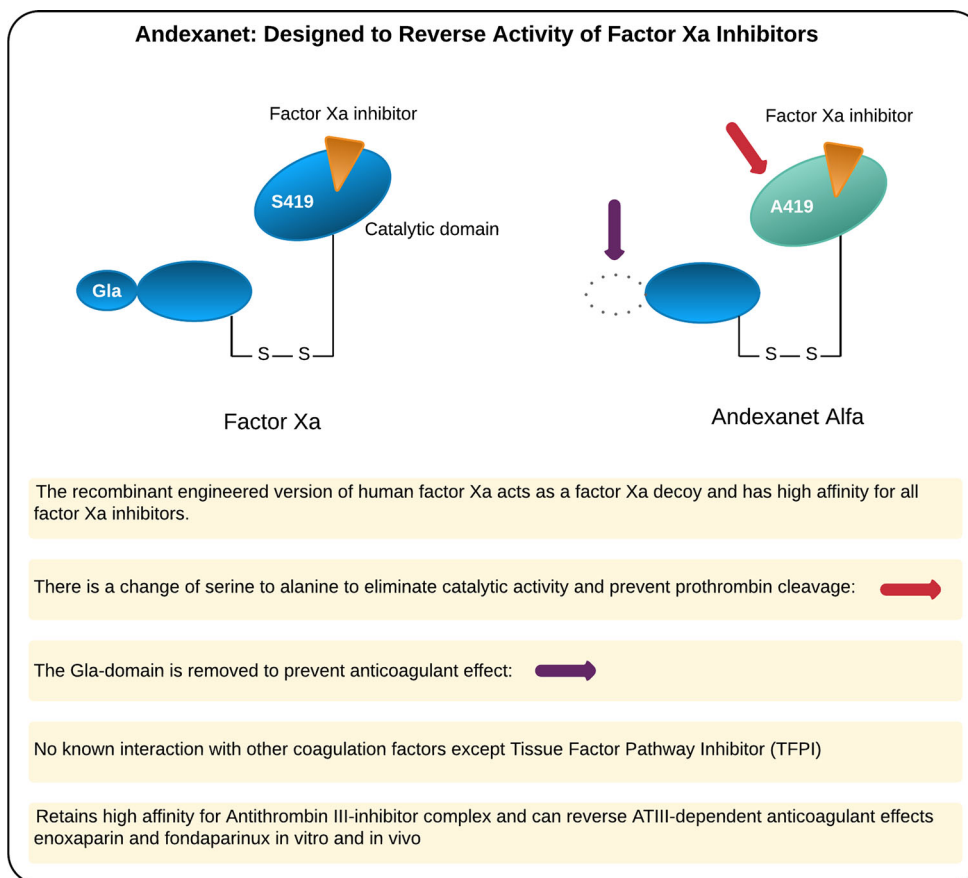
RE-VERSE AD (Reversal Effects of Idarucizumab on Active Dabigatran) is an ongoing, open, prospective cohort study in 300 patients, testing the efficacy and safety of idarucizumab 5 g intravenously for reversal of the anticoagulant effect of dabigatran in patients with severe bleeding and in patients requiring urgent surgery or intervention. An interim analysis including 90 patients has been published [27]. The primary endpoint of reversal of anticoagulation after 4 h could not be determined in 22 % of patients with severe bleeding and in 28 % of patients undergoing acute surgery, as coagulation assays (Hemoclot[®] dTT alone or in combination with ECT) were normal at baseline. In patients with dTT above the upper limit of normal, reversal of dTT was shown in 88–98 % of the patients within minutes. In 79 % of these patients, concentrations of unbound dabigatran remained at a level near the lower limit of quantification (20 ng/mL) at 24 h. Increases in dabigatran blood levels at 12 h were seen in 15 % (bleeding) and 21 % (surgery) of patients, respectively, probably due to redistribution of extravascular dabigatran into the intravascular compartment. In patients with severe bleeding, the median time to cessation of bleeding was 11.4 h, but this was difficult to assess in a large proportion of the patients, such as those with intracranial or retroperitoneal bleeding. Overall, there were 18 deaths (20 % of patients), ten of which were due to vascular causes (five due to cardiac arrest/circulatory collapse/ischaemic stroke and five due to fatal bleeding events). There was one thrombotic event within 72 h after idarucizumab administration in a patient in whom anticoagulants had not been reinitiated [27]. A controlled study design could have given a clearer view on clinical outcome, but seems unethical.

6.1.2 Andexanet Alfa

Andexanet alfa (PRT 064445) is a truncated human recombinant FXa, is catalytically inactive and lacks the membrane-binding domain of native FXa, but has the ability, like native FXa, to bind direct FXa inhibitors as well as low molecular weight heparin- and fondaparinux-activated antithrombin III with high affinity ([39]; Fig. 1).

Andexanet alfa acts as a decoy receptor and reverses the anticoagulant effects of all FXa inhibitors by competing with native FXa with a dose-dependent reversal of the anticoagulant effects and correction of clot formation *ex vivo* [39]. Reduced blood loss and restored haemostasis

Fig. 1 Working mechanism of andexanet alfa, a reversal agent for factor Xa-inhibitors [39]



was demonstrated in animal bleeding models after rivaroxaban, fondaparinux and enoxaparin pretreatment [40–42].

No effect on normal FXa function in haemostasis and no anticoagulant activity were seen. In placebo-controlled trials, the reversal effect of andexanet alfa was tested in healthy volunteers receiving direct (apixaban, rivaroxaban and edoxaban) or indirect (low molecular weight heparin and fondaparinux) acting FXa inhibitors [43–45]. When an intravenous bolus of 240 mg with subsequent infusion of 4–8 mg/min was used, a dose-dependent reversal of the anticoagulant effect of apixaban was observed with a >90 % decrease in anti-FXa activity within 2 min [44]. The anti-FXa activity returned to placebo level 2 h after stopping infusion. Thrombin generation and clotting time were also reversed dose-dependently by andexanet alfa. No thrombotic events or adverse effects were registered.

In the ANNEXA (Andexanet Alfa a Novel Antidote to the Anticoagulant Effects of FXa Inhibitors) trials (NCT02207725 and NCT02220725), older healthy volunteers pretreated with apixaban [46] or rivaroxaban (ongoing; [47]), respectively, received andexanet alfa bolus 400 mg intravenously followed by an infusion of 4 mg/minute for 120 min [47]. A 94 % rapid reversal of the

anticoagulant effect of apixaban was seen without serious adverse events or thrombotic complications in 33 volunteers aged 50–70 years [46].

A phase III, open label and uncontrolled study (NCT02329327) with andexanet alfa treatment is ongoing in patients with major bleeding episodes on treatment with an FXa inhibitor.

6.2 Synthetic Antidote

6.2.1 Aripazine

Aripazine (ciraparantag; PER977) is a small synthetic, peptide-like molecule that is designed with a distance between the positive guanidyl groups in the end of aripazine that matches the distance between the negative regions in heparin ([48]; Fig. 2). Aripazine not only binds to and reverses the anticoagulant effect of low molecular weight and unfractionated heparins, but also fondaparinux and NOACs. Aripazine does not bind coagulation factors and has no anti- or procoagulant effect.

In an animal bleeding model (rat tail transection), aripazine reduced blood loss on treatment with high doses of NOACs [48–50] and global coagulation tests were restored

Molecular structure of PER977

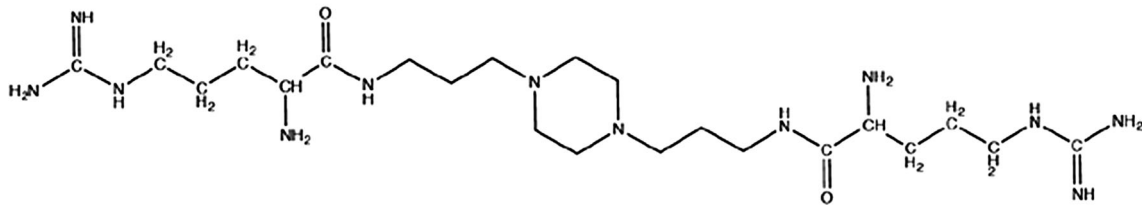


Fig. 2 Molecular structure of arripazine, a potential reversal agent for all NOACs (non-vitamin K antagonist oral anticoagulants)

to baseline levels in 20 min [48]. Aripazine safety, tolerability and pharmacokinetics were evaluated in healthy human volunteers in a phase I trial (NCT01826266) with single intravenous doses (5–300 mg) before and after a single 60 mg dose of edoxaban [51]. The anticoagulant activity of edoxaban was reversed by >100 mg of arripazine within 10 min and clot formation was restored without rebound signals for 24 h [52]. No procoagulant effect was observed and arripazine was well tolerated and safe.

With a similar design in normal human volunteers, re-anticoagulation with edoxaban and a second reversal with arripazine are currently being studied (NCT02207257).

Phase III clinical trials are needed to assess the clinical potential of this antidote in case of severe bleeding complications on treatment with NOACs.

7 Conclusion

Several types of NOAC antidotes are undergoing clinical evaluation and may, within a relatively short time period, be available for use in NOAC-treated patients with serious bleeding complications or who need urgent surgery. Availability of antidotes may lead to more frequent prescription of a NOAC in patients with an indication for anticoagulant therapy, but at the same time this will also lead to more hospital admissions with NOAC-related bleeding complications. At hospitals handling these NOAC-related bleeding complications and urgent surgery in anticoagulated patients, not only must antidotes be available, but also established coagulation assays with a rapid response rate should be on hand to test NOAC exposure, as well as guidelines for antidote use and for general measures for optimal treatment of these high-risk patients.

Ongoing studies will also show if a sudden interruption of anticoagulant therapy with antidotes increases the risk of thromboembolic complications and will demonstrate whether repeated antidote administration is safe.

Compliance with Ethical Standards

Funding No sources of funding were used to assist in the preparation of this study.

Conflicts of interest Steen Husted has participated on the advisory board for Bayer, Bristol-Myers Squibb, Pfizer and Boehringer Ingelheim and has received speaker fees from Bayer, Bristol-Myers Squibb and Boehringer-Ingelheim. Freek Verheugt has received consulting fees from Bayer, Bristol-Myers Squibb/Pfizer, Boehringer Ingelheim and Daiichi-Sankyo. Willemijn Comuth has received the Young Thrombosis Researchers Group laboratory exchange grant from the European Society of Cardiology Working Group on Thrombosis and a hospital donation from Boehringer Ingelheim. She has received speaker fees from Bayer and Boehringer Ingelheim, and non-financial support from Bayer, Bristol-Myers Squibb/Pfizer, Boehringer Ingelheim, Stago, Siemens and ANIARA. Steen Husted, Freek Verheugt and Willemijn Comuth have not been involved in any activities related to NOAC reversal agents, which are the topic of this article.

References

- Ruff CT, Giugliano RP, Braunwald E, Hoffman EB, Deenadayalu N, Ezekowitz MD, et al. Comparison of the efficacy and safety of new oral anticoagulants with warfarin in patients with atrial fibrillation: a meta-analysis of randomised trials. *Lancet*. 2014;383(9921):955–62.
- Blaich C, Muller C, Michels G, Wiesen MH. Multi-analyte analysis of non-vitamin K antagonist oral anticoagulants in human plasma using tandem mass spectrometry. *Clin Chem Lab Med*. 2015. <http://www.degruyter.com/view/j/cclm.ahead-of-print/cclm-2014-1108/cclm-2014-1108.xml>.
- Douxflis J, Mani H, Minet V, Devalet B, Chatelain B, Dogne JM, et al. Non-VKA oral anticoagulants: accurate measurement of plasma drug concentrations. *Biomed Res Int*. 2015;2015:345138.
- Ruff CT, Giugliano RP, Braunwald E, Morrow DA, Murphy SA, Kuder JF, et al. Association between edoxaban dose, concentration, anti-Factor Xa activity, and outcomes: an analysis of data from the randomised, double-blind ENGAGE AF-TIMI 48 trial. *Lancet*. 2015;385(9984):2288–95.
- Reilly PA, Lehr T, Haertter S, Connolly SJ, Yusuf S, Eikelboom JW, et al. The effect of dabigatran plasma concentrations and patient characteristics on the frequency of ischemic stroke and major bleeding in atrial fibrillation patients: the RE-LY Trial (Randomized Evaluation of Long-Term Anticoagulation Therapy). *J Am Coll Cardiol*. 2014;63(4):321–8.
- Eikelboom JW, Connolly SJ, Brueckmann M, Granger CB, Kappetein AP, Mack MJ, et al. Dabigatran versus warfarin in patients with mechanical heart valves. *N Engl J Med*. 2013;369(13):1206–14.
- Samama MM, Meddahi S, Samama CM. Pharmacology and laboratory testing of the oral Xa inhibitors. *Clin Lab Med*. 2014;34(3):503–17.
- Cuker A, Siegal DM, Crowther MA, Garcia DA. Laboratory measurement of the anticoagulant activity of the non-vitamin K oral anticoagulants. *J Am Coll Cardiol*. 2014;64(11):1128–39.

9. Samama MM, Guinet C. Laboratory assessment of new anticoagulants. *Clin Chem Lab Med*. 2011;49(5):761–72.
10. Garcia D, Barrett YC, Ramacciotti E, Weitz JI. Laboratory assessment of the anticoagulant effects of the next generation of oral anticoagulants. *J Thromb Haemost*. 2013;11(2):245–52.
11. Cohen D. Dabigatran: how the drug company withheld important analyses. *BMJ*. 2014;23(349):g4670.
12. Hyphen Biomed. Application sheet for dabigatran with hemoclot thrombin inhibitors v3. Available at: <http://www.aniara.com/pdf/ADA-ACK002KLCS2000i-2100i.pdf>. Accessed July 2015.
13. Faaborg L, Comuth WJ, Bloch-Münster AM, Henriksen LØ, Husted S. Handling of the hemoclot thrombin inhibitor assay reagents on Sysmex CS-2100i when monitoring dabigatran in acute clinical situations. Poster, XXV International Society on Thrombosis and Haemostasis Congress, Toronto, Canada 2015, Abstract (June). http://www.postersessiononline.eu/pr/aula_poster.asp?congreso=740245997. Accessed 19 Oct 2015.
14. Schmitz EM, Boonen K, van den Heuvel DJ, van Dongen JL, Schellings MW, Emmen JM, et al. Determination of dabigatran, rivaroxaban and apixaban by ultra-performance liquid chromatography–tandem mass spectrometry (UPLC–MS/MS) and coagulation assays for therapy monitoring of novel direct oral anticoagulants. *J Thromb Haemost*. 2014;12(10):1636–46.
15. Antovic JP, Skeppholm M, Eintrei J, Boija EE, Soderblom L, Norberg EM, et al. Evaluation of coagulation assays versus LC–MS/MS for determinations of dabigatran concentrations in plasma. *Eur J Clin Pharmacol*. 2013;69(11):1875–81.
16. Hawes EM, Deal AM, Funk-Adcock D, Gosselin R, Jeanneret C, Cook AM, et al. Performance of coagulation tests in patients on therapeutic doses of dabigatran: a cross-sectional pharmacodynamic study based on peak and trough plasma levels. *J Thromb Haemost*. 2013;11(8):1493–502.
17. Bates SM, Weitz JI. Coagulation assays. *Circulation*. 2005;112(4):e53–60.
18. Lange U, Nowak G, Bucha E. Ecarin chromogenic assay—a new method for quantitative determination of direct thrombin inhibitors like hirudin. *Pathophysiol Haemost Thromb*. 2003–2004;33(4):184–191.
19. Douxfils J, Chatelain B, Hjemdahl P, Devalet B, Sennesael AL, Wallemacq P, et al. Does the Russell Viper Venom time test provide a rapid estimation of the intensity of oral anticoagulation? A cohort study. *Thromb Res*. 2015;135(5):852–60.
20. Klufft C, Burggraaf J. Introduction to haemostasis from a pharmacodynamic perspective. *Br J Clin Pharmacol*. 2011;72(4):538–46.
21. Dinkelaar J, Patiwaal S, Harenberg J, Leyte A, Brinkman HJ. Global coagulation tests: their applicability for measuring direct factor Xa- and thrombin inhibition and reversal of anticoagulation by prothrombin complex concentrate. *Clin Chem Lab Med*. 2014;52(11):1615–23.
22. Gatt A, van Veen JJ, Woolley AM, Kitchen S, Cooper P, Makris M. Thrombin generation assays are superior to traditional tests in assessing anticoagulation reversal in vitro. *Thromb Haemost*. 2008;100(2):350–5.
23. Lance MD. A general review of major global coagulation assays: thrombelastography, thrombin generation test and clot waveform analysis. *Thromb J* 2015;13:1. doi:10.1186/1477-9560-13-1.
24. Beyer-Westendorf J, Forster K, Pannach S, Ebertz F, Gelbricht V, Thieme C, et al. Rates, management, and outcome of rivaroxaban bleeding in daily care: results from the Dresden NOAC registry. *Blood*. 2014;124(6):955–62.
25. Beyer-Westendorf J, Ebertz F, Forster K, Gelbricht V, Michalski F, Kohler C, et al. Effectiveness and safety of dabigatran therapy in daily-care patients with atrial fibrillation. Results from the Dresden NOAC Registry. *Thromb Haemost*. 2015;113(6):1247–57.
26. Heidbuchel H, Verhamme P, Alings M, Antz M, Hacke W, Oldgren J, et al. European Heart Rhythm Association Practical Guide on the use of new oral anticoagulants in patients with non-valvular atrial fibrillation. *Europace*. 2013;15(5):625–51.
27. Pollack CV Jr, Reilly PA, Eikelboom J, Glund S, Verhamme P, Bernstein RA, et al. Idarucizumab for Dabigatran Reversal. *N Engl J Med*. 2015;373(6):511–20.
28. Verheugt FW, Granger CB. Oral anticoagulants for stroke prevention in atrial fibrillation: current status, special situations, and unmet needs. *Lancet*. 2015;386(9990):303–10.
29. Eerenberg ES, Kamphuisen PW, Sijpkens MK, Meijers JC, Buller HR, Levi M. Reversal of rivaroxaban and dabigatran by prothrombin complex concentrate: a randomized, placebo-controlled, crossover study in healthy subjects. *Circulation*. 2011;124(14):1573–9.
30. Levi M, Moore KT, Castillejos CF, Kubitzka D, Berkowitz SD, Goldhaber SZ, et al. Comparison of three-factor and four-factor prothrombin complex concentrates regarding reversal of the anticoagulant effects of rivaroxaban in healthy volunteers. *J Thromb Haemost*. 2014;12(9):1428–36.
31. Martin AC, Le Bonniec B, Fischer AM, Marchand-Leroux C, Gaussem P, Samama CM, et al. Evaluation of recombinant activated factor VII, prothrombin complex concentrate, and fibrinogen concentrate to reverse apixaban in a rabbit model of bleeding and thrombosis. *Int J Cardiol*. 2013;168(4):4228–33.
32. Zhou W, Schwarting S, Illanes S, Liesz A, Middelhoff M, Zorn M, et al. Hemostatic therapy in experimental intracerebral hemorrhage associated with the direct thrombin inhibitor dabigatran. *Stroke*. 2011;42(12):3594–9.
33. Godier A, Miclot A, Le Bonniec B, Durand M, Fischer AM, Emmerich J, et al. Evaluation of prothrombin complex concentrate and recombinant activated factor VII to reverse rivaroxaban in a rabbit model. *Anesthesiology*. 2012;116(1):94–102.
34. Schiele F, van Ryn J, Canada K, Newsome C, Sepulveda E, Park J, et al. A specific antidote for dabigatran: functional and structural characterization. *Blood*. 2013;121(18):3554–62.
35. Glund S, Stangier J, Schmohl M. Idarucizumab, a specific antidote for dabigatran: immediate, complete and sustained reversal of dabigatran induced anticoagulation in elderly and renally impaired subjects. *Blood*. 2014;124(344). <http://www.bloodjournal.org/content/124/21/344>. Accessed 19 Oct 2015.
36. Glund S, Moschetti V, Norris S, Stangier J, Schmohl M, van Ryn J, et al. A randomised study in healthy volunteers to investigate the safety, tolerability and pharmacokinetics of idarucizumab, a specific antidote to dabigatran. *Thromb Haemost*. 2015;113(5):943–51.
37. Glund S, Stangier J, Schmohl M, Gansser D, Norris S, van Ryn J, et al. Safety, tolerability, and efficacy of idarucizumab for the reversal of the anticoagulant effect of dabigatran in healthy male volunteers: a randomised, placebo-controlled, double-blind phase 1 trial. *Lancet*. 2015;386(9994):680–90.
38. Grottko O, Honickel M, van Ryn J, Ten Cate H, Rossaint R, Spronk HM. Idarucizumab, a specific dabigatran reversal agent, reduces blood loss in a porcine model of trauma with dabigatran anticoagulation. *J Am Coll Cardiol*. 2015;66(13):1518–9.
39. Lu G, DeGuzman FR, Hollenbach SJ, Karbarz MJ, Abe K, Lee G, et al. A specific antidote for reversal of anticoagulation by direct and indirect inhibitors of coagulation factor Xa. *Nat Med*. 2013;19(4):446–51.
40. Hollenbach S, Tan S, Deguzman F, Malinowski J, Hutchaleelaha A, Inagaki M, et al. PRT064445 reverses rivaroxaban induced anticoagulation in a rabbit liver laceration “treatment” model. *Eur Heart J*. 2013;241. http://eurheartj.oxfordjournals.org/content/34/suppl_1/P241. Accessed 19 Oct 2015.
41. Lu G, Deguzman F, Hollenbach S, Luan P, Abe K, Siu G, et al. Reversal of low molecular weight heparin and fondaparinux by a

- recombinant antidote (r-Antidote, PRT064445). Abstract 12420. *Circulation*. 2010;122. http://circ.ahajournals.org/cgi/content/meeting_abstract/122/21_MeetingAbstracts/A12420. Accessed 19 Oct 2015.
42. Hollenbach S, Lu G, Deguzman F, Curnutte J, Conley P, Sinha U. Bolus administration of PRT064445, a recombinant factor Xa inhibitor antidote, reverses blood loss and PD markers in a rat model following enoxaparin-induced anticoagulation. Abstract P1857. *Eur Heart J*. 2012;33 (abstract supplement):309. http://www.postersessiononline.eu/pr/aula_poster.asp?congreso=740245997&direccion_posters=Seleccion&pagina_posters=5&ordenacion=n_poster&cod_congreso_integracion=&pst_clave=&buscar=&swAcceso=&swAccesoAdmin=&cod_congreso_aula_rel=&busqueda_rapida=&tipo=autor&texto=&texto2=&grupo2_aula=&grupo=&texto_ident=. Accessed 19 Oct 2015.
43. Crowther M, Levy G, Lu G. A phase 2 randomized, double-blind, placebo-controlled trial demonstrating reversal of edoxaban-induced anticoagulation in healthy subjects by andexanet Alfa (PRT064445), a universal antidote for factor Xa (FXa) inhibitors. *Blood*. 2014;124. <http://www.bloodjournal.org/content/124/21/4269?sso-checked=true>. Accessed 19 Oct 2015 (abstract).
44. Crowther M, Lu G, Conley P. Sustained reversal of apixaban anticoagulation with andexanet alfa using a bolus plus infusion regimen in a phase 2 placebo controlled trial. *Eur Heart J*. 2014;35(137). <http://spo.escardio.org/SessionDetails.aspx?eevid=69&sessId=14101&subSessId=3562#.VgqhNPntmko>. Accessed 19 Oct 2015 (abstract).
45. Crowther M, Mathur A, Kitt M. A phase 2 randomized, double-blind, placebo-controlled trial demonstrating reversal of rivaroxaban-induced anticoagulation in healthy subjects by andexanet alfa (PRT064445), an antidote for FXa inhibitors. *Blood*. 2013;122(A 3636). <http://www.bloodjournal.org/content/122/21/3636>. Accessed 19 Oct 2015 (abstract).
46. Crowther M, Levy G, Lu G. ANNEXA (TM)-R: A phase 3 randomized, double-blind, placebo-controlled trial, demonstrating reversal of apixaban-induced anticoagulation in older subjects by andexanet alfa (PRT064445), a universal antidote for factor Xa (FXa) inhibitors. *Circulation*. 2014;130:2116-7 Accessed 19 Oct 2015 (abstract).
47. Gold A, Crowther M, Levy G. ANNEXA (TM)-R: A phase 3 randomized, double-blind, placebo-controlled trial, demonstrating reversal of rivaroxaban-induced anticoagulation in older subjects by andexanet alfa (PRT064445), a universal antidote for factor Xa (FXa) inhibitors. *J Am Coll Cardiol*. 2015. <http://content.onlinejacc.org/article.aspx?articleid=2196748> (abstract).
48. Laulicht B, Bakhru S, Lee C. Small molecule antidote for anticoagulants. *Circulation*. 2012;126(A11395). http://circ.ahajournals.org/cgi/content/meeting_abstract/126/21_MeetingAbstracts/A11395 (abstract).
49. Laulicht B, Bakhru S, Jiang X. Antidote for new oral anticoagulants: mechanism of action and binding specificity of PER977. *J Thromb Haemost JTH*. 2013;11:75. <http://perosphere.com/content/media/documents/AntidoteforNewOralAnticoagulants-MechanismofActionandBindingSpecificityofPER977.pdf> (abstract).
50. Bakhru S, Laulicht B, Jiang X. PER977: a synthetic small molecule which reverses over-dosage and bleeding by new oral anticoagulants. *Circulation*. 2013;128. http://circ.ahajournals.org/cgi/content/meeting_abstract/128/22_MeetingAbstracts/A18809 (abstract).
51. Ansell J, Laulicht B, Bakhru S. Aripazine reverses unfractionated and low molecular weight heparins, fondaparinux and new Xa and IIa oral anticoagulants: report of Phase I/II clinical trial with edoxaban. *Eur Heart J*. 2014;35:854-5.
52. Ansell JE, Bakhru SH, Laulicht BE, Steiner SS, Grosso M, Brown K, et al. Use of PER977 to reverse the anticoagulant effect of edoxaban. *N Engl J Med*. 2014;371(22):2141-2.
53. Mueck W, Borris LC, Dahl OE, Haas S, Huisman MV, Kakkar AK, et al. Population pharmacokinetics and pharmacodynamics of once- and twice-daily rivaroxaban for the prevention of venous thromboembolism in patients undergoing total hip replacement. *Thromb Haemost*. 2008;100(3):453-61.
54. Mueck W, Lensing AW, Agnelli G, Decousus H, Prandoni P, Misselwitz F. Rivaroxaban: population pharmacokinetic analyses in patients treated for acute deep-vein thrombosis and exposure simulations in patients with atrial fibrillation treated for stroke prevention. *Clin Pharmacokinet*. 2011;50(10):675-86.
55. Skeppholm M, Al-Aieshy F, Berndtsson M, Al-Khalili F, Ronquist-Nii Y, Soderblom L, et al. Clinical evaluation of laboratory methods to monitor apixaban treatment in patients with atrial fibrillation. *Thromb Res*. 2015;136(1):148-53.
56. Bristol-Myers Squibb. Summary of product characteristics. Available at: http://www.ema.europa.eu/docs/en_GB/document_library/EPAR_-_Product_Information/human/002148/WC500107728.pdf. Accessed 9 Jul 2015.
57. Weitz JI, Connolly SJ, Patel I, Salazar D, Rohatagi S, Mendell J, et al. Randomised, parallel-group, multicentre, multinational phase 2 study comparing edoxaban, an oral factor Xa inhibitor, with warfarin for stroke prevention in patients with atrial fibrillation. *Thromb Haemost*. 2010;104(3):633-41.