#### **LEADING ARTICLE**



# **Alzheimer's Disease in Down Syndrome: Progress in the Design and Conduct of Drug Prevention Trials**

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Published online: 6 June 2020 © Springer Nature Switzerland AG 2020

#### **Abstract**

Individuals with Down syndrome (DS) are at high risk for developing Alzheimer's disease (AD) pathology and this has provided signifcant insights into our understanding of the genetic basis of AD. The present review summarizes recent clinical, neuropathologic, imaging, and fuid biomarker studies of AD in DS (DSAD), highlighting the striking similarities, as well as some notable diferences, between DSAD and the more common late-onset form of AD (LOAD) in the general population, as well as the much rarer, autosomal-dominant form of AD (ADAD). There has been signifcant progress in our understanding of the natural history of AD biomarkers in DS and their relationship to clinically meaningful changes. Additional work is needed to clearly defne the continuum of AD that has been described in the general population, such as the preclinical, prodromal, and dementia stages of AD. Multiple therapeutic approaches, including those targeting not only β-amyloid but also tau and the amyloid precursor protein itself, require consideration. Recent developments in the feld are presented within the context of such eforts to conduct clinical trials to treat and potentially prevent AD in DS.

#### **Key Points**

There is a need for novel therapeutics for Alzheimer's disease in Down syndrome (DSAD).

There is a lack of randomized controlled trials for DSAD therapeutics.

New biomarker strategies may enhance diagnostics and clinical trial designs, and new therapeutic targets may assist in the development of disease-modifying therapies for DSAD.

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# **1 Introduction**

In 1866, John Langdon Down described patients under his care at the Earlswood Asylum who exhibited intellectual disability along with abnormal physical growth and a distinct appearance [[1\]](#page-7-0). Nearly a century later, in 1959, trisomy for chromosome 21 was identifed as the molecular basis for Down syndrome (DS) [[2](#page-7-1)]. In 1987, mapping of the gene that encodes the amyloid precursor protein to chromosome 21 [[3\]](#page-7-2), along with further discovery of mutations on chromosome 21 that cause autosomal-dominant forms of AD (ADAD) [\[4,](#page-7-3) [5](#page-7-4)], suggested that overexpression of amyloid precursor protein is causative for AD pathology in both AD in DS (DSAD) and ADAD.

Over the past 4 decades, tremendous strides have been made in addressing treatable medical comorbidities in DS, which has led to a doubling in life-expectancy [\[6](#page-7-5)]. A child with DS born today can expect to live into his or her 60s and often beyond. However, with this increased lifespan, the prevalence of AD dementia in this population has surged. A number of studies have shown that the risk of AD dementia in DS is approximately 23% at 50 years of age, 45% at 55 years of age, and 88% or more at 65 years of age [\[7](#page-7-6)[–10](#page-7-7)]. Dementia, cardiovascular disease, and pneumonia make up the most common causes of death in people with DS over the age of 36 years  $[11]$  $[11]$ .

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As mentioned, DS is caused by trisomy of chromosome 21. There are in fact three cytogenetic forms of DS: (1) free trisomy 21, which consists of a supplementary chromosome 21 in all cells [\[12](#page-7-9)]; (2) mosaic trisomy 21, where there are two cell lineages, one with the normal number of chromo-somes and another with an extra chromosome 21 [[13\]](#page-7-10); and (3) robertsonian translocation trisomy 21, which occurs in only 2–4% of cases [[14\]](#page-7-11). All of these result in an increased copy of the *APP* gene and overexpression of the APP protein. Some individuals in the general population develop an early-onset form of AD as a result of duplication of a small region of chromosome 21 that includes the *APP* gene (dup-*APP*). These individuals develop an early-onset form AD and the pathogenic mechanisms in these individuals are thought to be analogous to those in individuals with DS [[15,](#page-7-12) [16](#page-7-13)], since an additional copy of *APP* is present in both dup-*APP* and DS forms of AD. This is in contrast to the autosomal-dominant forms of AD (ADAD) in which other genes (i.e. *PSEN1*, which encodes presenilin 1, or *PSEN2*, which encodes presenilin 2) are mutated and where the processing of APP is altered independently of gene copy number. Both of these mutations also lead to overproduction of the neurotoxic APP cleavage product Aβ (β-amyloid) [[17](#page-7-14)]. As expected, individuals with dup-*APP* share many traits with people with DS, including early age of dementia onset (mean age 52 years for dup-*APP*), ubiquitous AD neuropathology [\[18,](#page-7-15) [19](#page-7-16)], and an increased prevalence of cerebral amyloid angiopathy  $(CAA)$  [[20](#page-7-17)]. Although almost all people with DS have AD neuropathology by age 40 years, the variability in the prevalence of dementia is more marked in DS than in dup-APP. In addition, although Alzheimer's disease pathology is virtually inevitable in DS, the age of onset of clinical dementia varies tremendously. Indeed, not all individuals with DS will develop clinical dementia in their lifetime. Furthermore, the manifestation of CAA is less prevalent in DS than in dup-APP [\[21](#page-7-18)], but is more prevalent than in sporadic AD (22).

Case reports of individuals with DS who have partial trisomy 21 have provided deep insights into the critical role that APP overexpression plays in AD pathogenesis [\[23](#page-7-19), [24](#page-7-20)]. These rare and unusual cases point to the crucial importance of APP overexpression as the key driver of AD in DS. Moreover, the discovery of a mutation in the *APP* gene that results in diminished production of Aβ and is protective against the development of AD pathology and dementia, further supports the notion that AD can be solely driven by APP dysmetabolism, resulting in increased production of Aβ [[25\]](#page-7-21). Subsequent to accumulation of Aβ, the second key neuropathological hallmark of AD, neurofbrillary tangles, begin to develop. The abnormal hyperphosphorylation of tau protein leading to neurofbrillary tangles is seen later in the course of AD, both in the general population as well as in ADAD and DSAD, and correlates with the constellation

of symptoms based on its regional location in the brain [[26](#page-7-22)]. Tau pathology also appears to correlate temporally more closely with symptom onset in addition to spatially with symptom type, whereas  $\mathbf{A}\beta$  accumulation into fibrillar plaques occurs decades earlier [\[27](#page-7-23), [28](#page-7-24)].

To sum up, the discovery of the genetic causes for DSAD and ADAD catalyzed research into the relationship of chromosome 21, the gene for amyloid precursor protein, and the pivotal role of  $\overrightarrow{AB}$  in AD pathogenesis. In the past decade, the clinical, imaging, pathologic, and biochemical relationships of AD in DSAD and ADAD have been individually described by groups from around the world. Nonetheless, although DSAD and ADAD have fundamentally diferent initial pathways compared with the late-onset form of AD (LOAD), they share a remarkably similar pathophysiology, resulting in overproduction and subsequent accumulation of  $A\beta$ , which sets off a cascade of pathogenic events leading to neurodegeneration and the syndrome of dementia [\[29–](#page-7-25)[32\]](#page-8-0).

## **2 Clinical Presentation of Alzheimer's Disease (AD) in Down Syndrome (DS)**

In general, the clinical presentation of DSAD is similar to both ADAD and LOAD. DSAD cases present with an insidious onset of episodic memory difficulties followed by inevitable progression of cognitive defcits resulting in loss of function in activities of daily living, which is the cornerstone of dementia. Variability in baseline intellectual function in DS makes the diagnosis of dementia challenging. The most obvious diference between DSAD and ADAD with LOAD is the younger age at onset in individuals with DSAD and ADAD. Symptoms typically appear between the ages of 30 and 50 years, average age 52 years, and approximately 80% have dementia by the age of 65 years [\[33\]](#page-8-1). Behavioral symptoms seem to be more prevalent in DSAD compared with all other forms of AD [[34](#page-8-2), [35](#page-8-3)] and seem to present more frequently in those with DS who experience an earlier onset of dementia [[36–](#page-8-4)[39](#page-8-5)].

Focal neurological signs and symptoms appear to be more common in ADAD than in DSAD and LOAD. CAA, myoclonus, and seizures are a key feature of the *APP* duplication pedigrees [[19\]](#page-7-16) and are observed less frequently in DSAD. When myoclonus is observed in DSAD, it is more closely correlated with the onset of dementia and also appears to be associated with a more rapid cognitive decline in demented individuals with DS [\[40](#page-8-6)]. Up to 84% of demented individuals with DS develop seizures [\[41](#page-8-7)], with myoclonic epilepsy appearing to be the most common form in DS [\[42\]](#page-8-8).

### **3 Neuropathology**

For several decades, it has been known that fibrillar  $A\beta$ plaque accumulation is nearly universal in people with DS over age 40 years, as demonstrated by autopsy studies [[43\]](#page-8-9). Autopsy studies have demonstrated that  $\mathbf{A}\beta$  plaques begin to develop in very young people with DS [[44,](#page-8-10) [45\]](#page-8-11) and are found in all adults with DS by 40 years of age decades earlier than that observed in LOAD within the general population. Aβ initially appears in difuse deposits that over time progress to form compact neuritic plaques with increasing age. After 40 years of age, the accumulation of brain amyloid is not linear but rather exponential and spreads throughout the cortex  $[46]$ . A $\beta$  also accumulates within the cerebral vasculature in DS, as has been observed to occur in sporadic LOAD and, to a greater extent, in ADAD, resulting in CAA [\[21\]](#page-7-18). On a cellular level, increases in the endocytic recycling of Aβ have been observed, with consequent endosomal enlargement that has been reported in DS as early as 28 weeks of gestation [[47\]](#page-8-13).

Amyloid pathology begins in the late teens, with deposits of difuse plaques initially within the temporal lobe, then spreading to the neocortical regions and hippocampus, reaching the subcortical regions by the late 40s. After 50 years of age, every region of the brain, including the cerebellum, is littered with amyloid plaques [\[48,](#page-8-14) [49\]](#page-8-15). CAA begins at approximately 40 years of age, approximately 25 years after initial deposition of  $\mathbf{A}\beta$  as plaques [[49\]](#page-8-15).

Tau is a microtubule-related protein that becomes hyperphosphorylated in AD, leading to a conformational change in its structure, and in turn leading to the establishment of neurofbrillary tangles, which disrupts the neuronal cytoskeleton [[50](#page-8-16), [51\]](#page-8-17). The earliest sites of tau pathology include the entorhinal and trans-entorhinal cortex, spreading to the hippocampus, then the temporal cortex, and eventually to other regions of the cerebral cortex, fnally reaching the visual association cortex and primary visual cortex [[51](#page-8-17)]. Tau pathology begins at approximately 35 years of age within the hippocampus and spreads to the neocortical regions after 45 years of age [[51\]](#page-8-17).

Interestingly, critical infammatory genes on chromosome 21 are triplicated in DS and are believed to infuence the infammatory state of the DS brain [[52](#page-8-18)]. There are two macrophage types in the CNS immune system: M1 macrophages, which are proinfammatory and responsible for infammatory signaling, and M2 macrophages, which are anti-infammatory and participate in the resolution of the infammatory process. Indeed, M2 macrophages produce anti-infammatory cytokines, thereby contributing to tissue healing [[53\]](#page-8-19). As many genes located on chromosome 21 are primarily associated with M1, i.e. proinfammatory

macrophages, it has been recently shown that the DS brain has a proinflammatory status [[54](#page-8-20)]. It remains unclear to what extent this proinflammatory state affects the manifestation of AD neuropathology in DS.

## **4 Neuroimaging**

#### **4.1 Amyloid Positron Emission Tomography (PET)**

Amyloid PET imaging has been performed, using various tracers, in adults with DS over the past decade [[55–](#page-8-21)[58](#page-8-22)]. Amyloid PET positivity increases dramatically over age 40 years, refecting the binding of these tracers to the fbril-lar form of Aβ in plaques [[59,](#page-8-23) [60\]](#page-8-24). The progression of amyloid accumulation in the brain appears to be unique in individuals with DS compared with LOAD. Recent studies have demonstrated that the frst area of accumulation is the striatum  $[61, 62]$  $[61, 62]$  $[61, 62]$ , which is similar to the striatal binding observed in ADAD [\[63](#page-8-27)–[65\]](#page-8-28). Rates of accumulation of amyloid appear similar to rates in the sporadic population [[62\]](#page-8-26). The decades-long period between the emergence of amyloid plaques in the brain until the development of symptoms defnes the 'preclinical stage of AD' in the sporadic population. The next stage, when symptoms are restricted to memory loss, is referred to as the 'prodromal stage of AD', and the fnal stage, where memory and other cognitive domains are afected and contribute to functional decline, is referred to as the 'dementia stage of AD' [\[66](#page-8-29)]. It is believed that these same stages exist as a continuum DS. Although detection of elevated brain amyloid using PET has helped defne the preclinical stage of AD in the general population, and thereby refned sample selection for AD clinical trials [\[66](#page-8-29)], its applicability to clinical practice (both in the general and DS population) will likely be limited. In the general population, amyloid PET imaging will likely be most useful for determining if AD is the underlying cause in symptomatic individuals. In people with DS, who are genetically predetermined to develop AD, the utility of amyloid PET imaging in the clinic may have a very limited role since almost everyone will exhibit elevated brain amyloid.

#### **4.2 Tau PET**

As mentioned earlier, tau pathology in the form of neurofbrillary tangles is a key hallmark of AD pathology and a more proximal marker of subsequent AD-related cognitive decline than amyloid pathology [[67\]](#page-8-30). Indeed, the regional distribution of tau pathology in the brain mirrors clinical symptoms in the general population [\[68](#page-8-31)]. A similar relationship has been seen regarding neurofbrillary tangles and cognitive decline in DS [\[69](#page-9-0)]. The PET tracer 18F-AV-1451 has been shown to bind to neurofibrillary tangles [[70](#page-9-1), [71](#page-9-2)]. Tau pathology in adults with DS has recently been studied using the PET tracer 18F‐AV‐1451, as well as its relationship to regional amyloid deposition, regional cerebral glucose metabolism, regional brain atrophy, and cognitive and functional status  $[69]$  $[69]$ . The relationship between neurofibrillary tangles and amyloid load is consistent with that observed in LOAD. The presence of tau extending beyond the medial temporal lobes appears to correlate with subsequent cognitive decline [[70\]](#page-9-1). It has been noted that higher cognitive scores correlate with lower tau pathology in DS, just as has been observed in the general population [\[69](#page-9-0)]. Tau PET may therefore represent an appealing outcome measure for clinical trials as it is more closely linked in time with AD-related cognitive decline compared with amyloid PET,

#### **4.3 Fluorodeoxyglucose (FDG) PET**

Fluorodeoxyglucose (FDG)-PET is useful in measuring regional neuronal activity based on local glucose metabolism. Various patterns in regional FDG signal (both hypometabolic and hypermetabolic) have been shown to be associated with cognitive function in people with DS [\[71](#page-9-2)]. Recently, it has been shown that individuals with DS demonstrate the same regional hypometabolism observed in the posterior cingulate/precuneus as individuals with AD in the general population [[72](#page-9-3), [73\]](#page-9-4). There also appears to be an inverse relationship between amyloid accumulation and regional glucose metabolism [\[57](#page-8-32)], as well as an inverse relationship between tau pathology and FDG-PET signal [\[69](#page-9-0)]. More work is needed in studying longitudinal changes in FDG-PET in DSAD to better understand its potential role as an outcome measure for clinical trials.

#### **4.4 Magnetic Resonance Imaging**

Just as in the general population, neuropathological studies indicate that older adults with DS exhibit medial temporal lobe atrophy as part of the development of AD [[74–](#page-9-5)[76](#page-9-6)]. Magnetic resonance imaging (MRI) studies demonstrate hippocampal atrophy in DS, as has been reported in LOAD. Furthermore, hippocampal atrophy in DS correlates with changes in memory measures, but is a late fnding of neurodegeneration, as in the general population [\[77](#page-9-7)]. Regional grey matter changes can be detectable before signs of clinical dementia [[78](#page-9-8)]. Individuals with DSAD eventually show atrophy of the amygdala, caudate, posterior cingulate, parietal, temporal, and frontal regions, as well as enlargement of cerebrospinal fuid (CSF) spaces [\[78\]](#page-9-8). Although MRI exquisitely depicts brain atrophy, the fact that it refects neurodegeneration, which defnes the fnal stage of AD, severely limits its use as an AD biomarker; however, MRI will continue a critical role in safety assessments for adverse events in clinical trials.

## **5 Fluid Biomarkers**

Blood-based assays have clear advantages as biomarkers because they are easily accessible and non-invasive. The role of plasma levels of Aβ as a biomarker for DSAD has been studied extensively. People with DS have consistently shown higher plasma Aβ concentrations compared with individuals without DS [[79,](#page-9-9) [80](#page-9-10)]. There have only been a limited number of CSF studies in individuals with DS due to the challenges in collecting CSF from individuals with DS  $[81, 82]$  $[81, 82]$  $[81, 82]$  $[81, 82]$ . These studies show elevated levels of Aβ42. However, as these individuals age, CSF Aβ42 levels decline (as expected with their deposition into plaques) and CSF tau levels increase [[82](#page-9-12)].

Recent work on CSF and plasma levels of neuroflament light (NfL), a component of the axonal cytoskeleton and marker of neuronal damage and degeneration, has shown strong correlation with cognitive status in adults with DS [[83,](#page-9-13) [84\]](#page-9-14). Specifcally, plasma NfL levels appear to increase with age, and can distinguish between normal aging in DS and DSAD [\[84\]](#page-9-14). Indeed, there appears to be excellent diagnostic performance of plasma NfL for the asymptomatic versus dementia group, with an AUC of 0.95 (95% confdence interval [CI] 0.92–0.98) and a sensitivity and specificity of 90% and 92%, respectively [[82\]](#page-9-12). Plasma NfL levels have also been shown to correlate with standard biomarkers of AD pathology, as well as with markers of neurodegeneration (regional cerebral glucose metabolism as assessed with FDG-PET and hippocampal atrophy), and even cognitive and functional decline [[83](#page-9-13)].

Beyond standard AD biomarkers such as Aβ species, total-tau and phospho-tau species, and Nf, metabolomic analyses may represent another biomarker with which we can further characterize AD in the DS population. Specifcally, recent work has shown metabolic perturbations characterize individuals with DS who were cognitively stable and have mild cognitive impairment or DSAD [[85](#page-9-15)]. Whether these metabolomic changes will be more applicable to screening purposes versus outcome measures for clinical trials will be based on longitudinal data on rates of change in metabolomic profles in conjunction with clinically meaningful cognitive and functional changes.

## **6 The Alzheimer's Biomarker Consortium for Down Syndrome (ABC‑DS)**

As the largest AD biomarker study in DS to date, Alzheimer's Biomarker Consortium for Down Syndrome (ABC-DS) is setting the stage for powering secondary prevention trials for DSAD. Similar to the two largest ongoing longitudinal studies of AD biomarkers (the Alzheimer Disease Neuroimaging Initiative [ADNI] and the Dominantly Inherited Alzheimer Network [DIAN], all data are being made available in an open-access format using a model similar to ADNI. This 5-year longitudinal study launched in 2015 is examining the progression of ADrelated biomarkers (Aβ-, tau- and FDG-PET, structural and functional MRI, CSF  $\text{A} \beta$  and tau, plasma  $\text{A} \beta$  and proteomics, genetics, neuropathology), as well as cognitive and functional measures, in over 500 adults with DS [[86](#page-9-16)]. The consortium stages participants based on clinical and neuropsychological assessments as being located on the continuum of AD in DS. Participants were classifed as (1) *cognitively stable (CS)* if they were without cognitive or functional decline, beyond what would be expected with adult aging per se; (2) having *mild cognitive impairment (MCI*-*DS)* if they demonstrated some cognitive and/or functional decline over and above what would be expected with aging per se, but not severe enough to indicate the presence of dementia; and as having dementia (*DS*-*AD)* if there was evidence of substantial progressive declines in cognitive functioning and daily living skills. An *'unable to determine'* category was utilized to indicate that declines were observed but could be caused by signifcant life circumstance (e.g. staff changes, family death) or conditions unrelated to AD (e.g. severe sensory loss, poorly resolved hip fracture, and psychiatric diagnosis, in particular depression) [[86](#page-9-16)]. These data will allow for a deeper, richer understanding of the natural history of AD biomarkers in DS (Fig. [1\)](#page-4-0).

# **7 Clinical Trials**

Clinical trials require identifying the appropriate sample population and outcome measures that will best assess the safety and efficacy of any given intervention. The US Code of Federal Regulations outlines specifc requirements to enhance protections for 'vulnerable populations', including

individuals with DS, which also raises issues that need to be addressed as part of the informed consent process. In developing disease-modifying drugs for DSAD, there is a need to understand the ideal sample (e.g. age range), the interventions (e.g. pharmacologic mechanism of action, pharmacokinetic/pharmacodynamic) and outcome measures (e.g. clinically meaningful and sensitivity to change) for a successful study design. In addition, the level of intellectual disability, as well as language abilities, will need to be carefully considered when selecting cognitive and clinical outcome measures. Specifcally, foor efects may limit the feasibility of many cognitive instruments used to assess longitudinal change in memory or the impact of a particular drug on this change. Few empirically evaluated, psychometrically sound outcome measures appropriate for use in clinical trials with individuals with DS have been identifed. Moreover, for clinical trials that aim to prevent the dementia stage of AD in DS by intervening during its earliest stages (i.e. preclinical stage), assessment tools will need to be created and validated to assess subtle changes in the asymptomatic or minimally symptomatic stage of DSAD. The National Institutes of Health (NIH)-sponsored meeting on Outcomes Measures for Clinical Trials in Down Syndrome resulted in guidelines for the selection of outcome measures for clinical trials for DSAD, although most involve the dementia stage of AD in DS [[87](#page-9-17)].

The DS population will undoubtedly require a unique set of considerations in terms of sensitive and valid assessments of early AD-related cognitive decline. Baseline assessments will be important, as will the need for carefully planned and consistent testing sessions. Some direct neuropsychological measures will likely only be sensitive and valid in verbal and/or higher functioning adults with DS. Such measures will allow researchers to track individuals with DS prospectively (perhaps as part of a trial-ready cohort) and are most likely to be relevant in clinical trials for therapies aimed at delaying the onset of and/or preventing AD-related cognitive decline. Indeed, treatments are likely to be most efective

<span id="page-4-0"></span>**Fig. 1** The natural history of AD biomarkers in DS. The continuum of AD in DS is thought to be similar to LOAD but beginning at a younger age, much as in ADAD [\[101\]](#page-9-18). *AD* Alzheimer's disease, *DS* Down syndrome, *LOAD* late-onset Alzheimer's disease, *ADAD* autosomal-dominant form of Alzheimer's disease



if introduced prior to irreversible losses of critical neural pathways in the later stages of the disease [\[88,](#page-9-19) [89\]](#page-9-20).

### **8 The Horizon21 DS Consortium**

In addition to the ABC-DS effort, the Horizon21 DS consortium in Europe is leading a parallel project to assess the longitudinal change in aging adults with DS. This consortium consists of various existing DS cohorts from the UK (the London Down Syndrome Consortium [LonDownS], the Cambridge Dementia in Down's Syndrome [DiDS] cohort), The Netherlands (the Rotterdam Down Syndrome Study), Germany (AD21 study group, Munich), France (TriAL21 for Lejeune Institute, Paris), and Spain (the Down Alzheimer Barcelona Neuroimaging Initiative [DABNI]). The consortium is providing deep insights into cognitive and clinical outcome measures of AD in DS in over 1000 participants, setting the stage for clinical trials [[90\]](#page-9-21).

## **9 Current Treatment Trials**

The conduct of clinical trials in the DS population raises many methodological challenges. Given the wide variations in baseline intellectual capabilities and cognitive functioning, these diferences must be taken into account for accurate assessment in testing situations. The main defning feature of dementia in the general population is a decline from the baseline level of function and the performance of daily skills. Consensus guidelines for the evaluation and management of dementia in DS have been proposed that focus on the dementia stage and do not include prodromal AD or preclinical criteria [\[91](#page-9-22)]. These criteria are currently lacking in the DS feld. The consensus guidelines are meant to serve as recommendations for clinicians evaluating individuals with DS who are experiencing cognitive decline. The guidelines provide a step-by-step and comprehensive approach to clinical evaluation. The key hallmark defning dementia is functional decline compared with previously attained levels, and, beyond that which would be explained by age-related changes. However, the earliest signs of ADrelated cognitive decline, distinct of a normal age-related change in adults with DS, may be subtle and will often require an astute observer. In addition, concomitant medications, recent medical illnesses (including laboratory testing such as thyroid-stimulating hormone and vitamin B12), or recent life events that can impact psychosocial functioning must also be considered when interpreting results. Although some validated measures exist [\[92](#page-9-23), [93](#page-9-24)], there is currently no single, standard clinical instrument for the assessment of mild cognitive impairment or dementia in adults with DS. In this regard, the consensus guidelines are meant to

aid clinicians attempting to diagnosis dementia in people with DS, and have limited utility for defning earlier stages of AD in DS such as preclinical and prodromal AD. The development of research guidelines for the defnition of the preclinical, prodromal, and dementia stages of AD in DS are critical for designing clinical trials to treat or prevent AD in DS. Nonetheless, a few small multicenter clinical trials have been successfully conducted in the DS population and have provided important insights into trial design and implementation in this group.

#### **9.1 Scyllo‑Inositol**

ELND005 (scyllo-inositol) is hypothesized to reduce amyloid toxicity to improve cognitive function in patients with DS. ELND005 (scyllo-inositol; cyclohexane-1,2,3,4,5,6 hexol) has also been studied as a potential disease-modifying treatment of LOAD [[94\]](#page-9-25). In preclinical studies, ELND005 showed amyloid anti-aggregation efects in vitro, reduced oligomer-induced neuronal toxicity, and improved learning in animal models of AD [[94](#page-9-25)[–96\]](#page-9-26). ELND005 showed amyloid-lowering efects in CSF and brain, in patients with AD in the general population [[97\]](#page-9-27). In addition, ELND005 showed beneficial trends on cognition in mild AD [\[97](#page-9-27)]. A phase II, randomized, double-blind, placebo-controlled study of oral ELND005 in 26 adults with DS without dementia demonstrated the safety and tolerability of the anti-amyloid compound in DS, as well as the feasibility of conducting a pharmacokinetic/pharmacodynamic study in adults with DS [[98\]](#page-9-28).

#### **9.2 ACI‑24**

ACI-24 is a liposomal vaccine that elicits an antibody response against aggregated Aβ peptides without concomitant proinfammatory T cell activation [\[99](#page-9-29)]. ACI-24 is based on the truncated Aβ1-15 sequence, which is devoid of T-cell epitopes located closer to the peptide's C-terminus. The peptide sequence is anchored into the surface of liposomes such that that the peptides adopt an aggregated β-sheet structure as a conformational epitope. Previous active vaccines (e.g. AN-1792) elicited a T-cell response, which led to an increased risk of meningoencephalitis [[100](#page-9-30)]. A phase I/II trial of ACI-24 is ongoing in Europe and aims to address safety, immunogenicity, and efficacy in mild to moderate Alzheimer's dementia in the general population. A phase Ib, placebo-controlled, multicenter study with ACI-24 for the treatment of AD in individuals with DS was launched in 2016 (NCT02738450). The study enrolled 16 adults with DS aged 25–45 years and treated them with ACI-24 for 1 year with an additional year of follow-up. Primary endpoints included measures of safety, tolerability and immunogenicity, while secondary endpoints of this clinical trial included the effects on biomarkers of AD pathology as well as cognitive and clinical function and neuroinfammatory markers. The study was completed in 2020, with topline results expected in the near future.

## **10 Future Directions**

Clearly, genetic support for the role of *APP* in AD in the pathogenesis of AD in DS is undeniably compelling. As described above, the presence of an extra copy of *APP*, whether as trisomy 21 or dup-*APP*, results in overproduction of Aβ and subsequent development of amyloid plaques and neurofbrillary tangles, as well as the manifestation of dementia. Furthermore, individuals with DS who have trisomy 21 but are disomic for *APP* develop no amyloid plaques or neurofbrillary tangles and have no symptoms of dementia. Finally, a mutation in the *APP* gene that prevents its cleavage to produce Aβ protects against the development of amyloid plaques and neurofbrillary tangles as well as dementia. Therefore, Aβ remains a prime target for therapeutics, including both active and passive immunization, just as it is being actively studied in the ADAD and LOAD populations.

Beyond anti-Aβ drugs, compounds that modulate *APP* expression in DS may have great potential for efectiveness. In addition, the role of tau appears to be more closely related to subsequent cognitive decline and drugs that prevent hyperphosphorylation of tau and/or drugs that remove hyperphosphorylated species of tau may represent another therapeutic avenue. Perhaps as importantly, understanding protective genetics and epigenetics in DS may help provide novel therapeutic avenues to consider for the prevention of dementia in DS.

## **11 The Alzheimer's Clinical Trial Consortium for Down Syndrome**

The tight link between genetic determinants of AD and the overproduction of Aβ provides compelling support for the amyloid cascade hypothesis and has been the focal point in the development of disease-modifying drugs for AD [[89\]](#page-9-20). It is hypothesized that disease-modifying treatments for AD and DS should begin prior to the onset of cognitive symptoms to prevent extensive neurodegeneration, and thus necessitates a clear understanding of biomarker changes throughout the course of the disease. The ABC-DS project is setting the stage for conducting secondary prevention trials for DSAD. The NIH-funded Alzheimer's Clinical Trial Consortium for Down Syndrome (ACTC-DS) recently launched the Trial Ready-Cohort for Down Syndrome (TRC-DS) [Fig. [2](#page-6-0)]. TRC-DS will enroll 120 non-demented participants with DS into a longitudinal run-in study with MRI, amyloid PET, cognitive testing, and biofuid biomarker analysis in advance of upcoming randomized, placebo-controlled clinical trials for DSAD. Data from the ABC-DS project are providing key insights into study design, including sample size selection and duration of the treatment period.

## **12 Conclusions**

Over the past decade, great progress has been made in understanding DSAD utilizing brain MRI, amyloid and tau PET, and biofuid biomarkers. Indeed, several research groups from around the world have shown that there exist remarkable similarities (and some diferences) between LOAD, ADAD, and DSAD. We are now poised to ask whether therapies that are currently being tested for LOAD and ADAD can be evaluated in people with DS. Undoubtedly, many

<span id="page-6-0"></span>**Fig. 2** The Trial-Ready Cohort—Down Syndrome. A clinical trial platform has been developed to utilize a collaborative approach to design and conduct clinical trials for AD in DS across expert international sites using widely accepted tools and standards. *AD* Alzheimer's disease, *DS* Down syndrome, *NIH* National Institutes of Health, *ABC-DS* Alzheimer's Biomarker Consortium for Down Syndrome, *RCT* randomized controlled trial



questions about DSAD still remain unanswered. We will need to identify which neuropsychological tests (or composite of tests) will provide a reasonable measure of clinical meaningfulness. We will need to understand how removal of amyloid will afect AD-related cognitive decline and to what extent. We will also need to consider how early in the course of DSAD we should intervene and for how long treatment should last to provide measurable beneft to patients. Data from the ABC-DS and Horizon21 natural history studies are setting the stage for conducting secondary prevention trials to prevent AD in DS while ACTC-DS will utilize a collaborative approach to design and conduct such trials across expert international sites using widely accepted tools and standards to bring the latest therapies to the DS population.

**Acknowledgements** Michael S. Rafi has received funding support from NIH R61AG066543.

#### **Compliance with Ethical Standards**

**Funding** No funding sources were used to assist with the preparation of this manuscript.

**Conflict of interest** Michael S. Rafi has served as consultant to AC Immune.

## **References**

- <span id="page-7-0"></span>1. Down JLH. Observations on an ethnic classifcation of idiots. Lond Hosp Clin Lect Rep. 1866;3:259–62.
- <span id="page-7-1"></span>2. Lejeune J, Gautier M, Turpin R. A study of somatic chromosomes in nine infants with mongolism. CR Acad Sci (Paris). 1959;240:1026–7.
- <span id="page-7-2"></span>3. Tanzi RE, Gusella JF, Watkins PC, et al. Amyloid beta protein gene: cDNA, mRNA distribution, and genetic linkage near the Alzheimer locus. Science. 1987;235:880–4.
- <span id="page-7-3"></span>4. Murrell J, Farlow M, Ghetti B, et al. A mutation in the amyloid precursor protein associated with hereditary Alzheimer's disease. Science. 1991;254:97–9.
- <span id="page-7-4"></span>5. Goate A, Chartier-Hardin M-C, Mullan M. Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. Nature. 1991;349:704–76.
- <span id="page-7-5"></span>6. Yang Q, Rasmussen SA, Friedman JM. Mortality associated with Down's syndrome in the USA from 1983 to 1997: a populationbased study. Lancet. 2002;359(9311):1019–25.
- <span id="page-7-6"></span>7. Rubenstein E, Hartley S, Bishop L. Epidemiology of Dementia and Alzheimer Disease in Individuals with Down Syndrome. JAMA Neurol. 2020;77(2):262–4.
- 8. McCarron M, McCallion P, Reilly E, et al. A prospective 20-year longitudinal follow-up of dementia in persons with Down syndrome. J Intellect Disabil Res. 2017;61:843–52.
- 9. Hithersay R, Startin CM, Hamburg S, et al. Association of dementia with mortality among adults with down syndrome older than 35 years. JAMA Neurol. 2019;76(2):152–60.
- <span id="page-7-7"></span>10. Bayen E, Possin KL, Chen Y, et al. Prevalence of aging, dementia, and multimorbidity in older adults with down syndrome. JAMA Neurol. 2018;75(11):1399–406.
- <span id="page-7-8"></span>11. Landes SD, Stevens JD, Turk MA. Obscuring efect of coding developmental disability as the underlying cause of death on

mortality trends for adults with developmental disability: a cross-sectional study using US Mortality Data from 2012 to 2016. BMJ Open. 2019;9(2):e026614.

- <span id="page-7-9"></span>12. Antonarakis SE. Human chromosome 21: genome mapping and exploration circa 1993. Trends Genet. 1993;9:142–8.
- <span id="page-7-10"></span>13. Thuline HC, Pueschel SM. Cytogenetics in Down syndrome. In: Pueschel SM, Rynders JE, editors. Down syndrome. Advances in biomedicine and the behavioral sciences. Cambridge: Ware Press; 1982. p. 133.
- <span id="page-7-11"></span>14. Asim A, Kumar A, Muthuswamy S, et al. Down syndrome: an insight of the disease. J Biomed Sci. 2015;22(1):41.
- <span id="page-7-12"></span>15. Sleegers K, Brouwers N, Gijselinck I, et al. APP duplication is sufficient to cause early onset Alzheimer's dementia with cerebral amyloid angiopathy. Brain. 2006;129:2977–83.
- <span id="page-7-13"></span>16. McNaughton D, Knight W, Guerreiro R, et al. Duplication of amyloid precursor protein (APP), but not prion protein (PRNP) gene is a signifcant cause of early onset dementia in a large UK series. Neurobiol Aging. 2012;33:426.e13–21.
- <span id="page-7-14"></span>17. Rumble B, Retallack R, Hilbich C, et al. Amyloid A4 and its precursor in Down's syndrome and Alzheimer's disease. N Engl J Med. 1989;320:1446–62.
- <span id="page-7-15"></span>18. Rovelet-Lecrux A, Hannequin D, Raux G, et al. APP locus duplication causes autosomal dominant early-onset Alzheimer disease with cerebral amyloid angiopathy. Nat Genet. 2006;38:24–6.
- <span id="page-7-16"></span>19. Cabrejo L, Guyant-Maréchal L, Laquerrière A, et al. Phenotype associated with APP duplication in fve families. Brain. 2006;129:2966–76.
- <span id="page-7-17"></span>20. Wallon D, Rousseau S, Rovelet-Lecrux A, et al. The French series of autosomal dominant early onset Alzheimer's disease cases: mutation spectrum and cerebrospinal fuid biomarkers. J Alzheimers Dis. 2012;30:847–56.
- <span id="page-7-18"></span>21. Carmona-Iragui M, Balasa M, Benejam B, et al. Cerebral amyloid angiopathy in Down syndrome and sporadic and autosomal-dominant Alzheimer's disease. Alzheimers Dement. 2017;13:1251–60.
- 22. Head E, Phelan MJ, Doran E, et al. Cerebrovascular pathology in Down syndrome and Alzheimer disease. Acta Neuropathol Commun. 2017;5(1):93.
- <span id="page-7-19"></span>23. Prasher VP, Farrer MJ, Kessling AM, et al. Molecular mapping of Alzheimer-type dementia in Down's syndrome. Ann Neurol. 1998;43:380–3.
- <span id="page-7-20"></span>24. Doran E, Keator D, Head E, et al. Down syndrome, partial trisomy 21, and absence of Alzheimer's disease: the role of APP. J Alzheimers Dis. 2017;56(2):459–70.
- <span id="page-7-21"></span>25. Jonsson T, Atwal JK, Steinberg S, et al. A mutation in APP protects against Alzheimer's disease and age-related cognitive decline. Nature. 2012;488(7409):96–9.
- <span id="page-7-22"></span>26. Brier MR, Gordon B, Friedrichsen K, et al. Tau and Aβ imaging, CSF measures, and cognition in Alzheimer's disease. Sci Transl Med. 2016;8(338):338ra66.
- <span id="page-7-23"></span>27. Ossenkoppele R, Schonhaut DR, Schöll M, et al. Tau PET patterns mirror clinical and neuroanatomical variability in Alzheimer's disease. Brain. 2016;139(Pt 5):1551–67.
- <span id="page-7-24"></span>28. Margallo-Lana ML, Moore PB, Kay DW, et al. Fifteen-year follow-up of 92 hospitalized adults with Down's syndrome: incidence of cognitive decline, its relationship to age and neuropathology. J Intellect Disabil Res. 2007;51(Pt. 6):463–77.
- <span id="page-7-25"></span>29. Beyreuther K, Masters CL. Amyloid precursor protein (APP) and beta A4 amyloid in the etiology of Alzheimer's disease: precursor-product relationships in the derangement of neuronal function. Brain Pathol. 1991;1:241–51.
- 30. Selkoe DJ. The molecular pathology of Alzheimer's disease. Neuron. 1991;6:487–98.
- 31. Hardy JA, Higgins GA. Alzheimer's disease: the amyloid cascade hypothesis. Science. 1992;256:184–5.
- <span id="page-8-0"></span>32. Jack CR Jr, Knopman DS, Jagust WJ, et al. Hypothetical model of dynamic biomarkers of the Alzheimer's pathological cascade. Lancet Neurol. 2010;9(1):119–28.
- <span id="page-8-1"></span>33. Wiseman FK, Al-Janabi T, Hardy J, et al. A genetic cause of Alzheimer disease: mechanistic insights from Down syndrome. Nat Rev Neurosci. 2015;16(9):564–74.
- <span id="page-8-2"></span>34. Ball SL, Holland AJ, Hon J, et al. Personality and behaviour changes mark the early stages of Alzheimer's disease in adults with Down's syndrome: fndings from a prospective populationbased study. Int J Geriatr Psychiatry. 2006;21:661–73.
- <span id="page-8-3"></span>35. Ball SL, Holland AJ, Treppner P, et al. Executive dysfunction and its association with personality and behaviour changes in the development of Alzheimer's disease in adults with Down syndrome and mild to moderate learning disabilities. Br J Cli. Psychol. 2008;47:1–29.
- <span id="page-8-4"></span>36. Dekker AD, Sacco S, Carf A, et al. The Behavioral and Psychological Symptoms of Dementia in Down Syndrome (BPSD-DS) Scale: comprehensive Assessment of Psychopathology in Down Syndrome. J Alzheimers Dis. 2018;63(2):797–819.
- 37. Lautarescu BA, Holland AJ, Zaman SH. The early presentation of dementia in people with Down syndrome: a systematic review of longitudinal studies. Neuropsychol Rev. 2017;27(1):31–45.
- 38. Dekker AD, Strydom A, Coppus AM, et al. Behavioural and psychological symptoms of dementia in Down syndrome: early indicators of clinical Alzheimer's disease? Cortex. 2015;73:36–61.
- <span id="page-8-5"></span>39. Holland AJ, Hon J, Huppert FA, et al. Incidence and course of dementia in people with Down's syndrome: fndings from a population-based study. J Intellect Disabil Res. 2000;44:138–46.
- <span id="page-8-6"></span>40. Lott IT, Doran E, Nguyen VQ, et al. Down syndrome and dementia: seizures and cognitive decline. J Alzheimers Dis. 2012;29(1):177–85.
- <span id="page-8-7"></span>41. Menéndez M. Down syndrome, Alzheimer's disease and seizures. Brain Dev. 2005;27(4):246–52.
- <span id="page-8-8"></span>42. De Simone R, Puig XS, Gélisse P, et al. Senile myoclonic epilepsy: delineation of a common condition associated with Alzheimer's disease in Down syndrome. Seizure. 2010;19(7):383-9.
- <span id="page-8-9"></span>43. Mann DM, Yates PO, Marcyniuk B. Alzheimer's presenile dementia, senile dementia of Alzheimer type and Down's syndrome in middle age form an age related continuum of pathological changes. Neuropathol Appl Neurobiol. 1984;10(3):185–207.
- <span id="page-8-10"></span>44. Lemere CA, Blusztajn JK, Yamaguchi H, et al. Sequence of deposition of heterogeneous amyloid beta-peptides and APOE in Down syndrome: implications for initial events in amyloid plaque formation. Neurobiol Dis. 1996;3:16–32.
- <span id="page-8-11"></span>45. Leverenz JB, Raskind MA. Early amyloid deposition in the medial temporal lobe of young Down syndrome patients: a regional quantitative analysis. Exp Neurol. 1998;150:296–304.
- <span id="page-8-12"></span>46. Abrahamson EE, Head E, Lott IT, et al. Neuropathological correlates of amyloid PET imaging in Down syndrome. Dev Neurobiol. 2019;79(7):750–66.
- <span id="page-8-13"></span>47. Cataldo AM, Peterhoff CM, Troncoso JC, et al. Endocytic pathway abnormalities precede amyloid beta deposition in sporadic Alzheimer's disease and Down syndrome: diferential efects of APOE genotype and presenilin mutations. Am J Pathol. 2000;157:277–86.
- <span id="page-8-14"></span>48. Thal DR, Rüb U, Orantes M, et al. Phases of A beta-deposition in the human brain and its relevance for the development of AD. Neurology. 2002;58(12):1791–800.
- <span id="page-8-15"></span>49. Davidson YS, Robinson A, Prasher VP, et al. The age of onset and evolution of Braak tangle stage and Thal amyloid pathology of Alzheimer's disease in individuals with Down syndrome. Acta Neuropathol Commun. 2018;6(1):56.
- <span id="page-8-16"></span>50. Alonso AD, Grundke-Iqbal I, Iqbal K. Alzheimer's disease hyperphosphorylated tau sequesters normal tau into tangles of flaments and disassembles microtubules. Nat Med. 1996;2:783–7.
- <span id="page-8-17"></span>51. Alonso AC, Zaidi T, Novak M, et al. Hyperphosphorylation induces self-assembly of tau into tangles of paired helical filaments/straight filaments. Proc Natl Acad Sci USA. 2001;98(12):6923–69238.
- <span id="page-8-18"></span>52. Wilcock DM, Griffin WS. Down's syndrome, neuroinflammation, and Alzheimer neuropathogenesis. J Neuroinfammation. 2013;10:84.
- <span id="page-8-19"></span>53. Wilcock DM. Neuroinfammation in the aging down syndrome brain; lessons from Alzheimer's disease. Curr Gerontol Geriatr Res. 2012;2012:170276.
- <span id="page-8-20"></span>54. Wilcock DM, Hurban J, Helman AM, et al. Down syndrome individuals with Alzheimer's disease have a distinct neuroinfammatory phenotype compared to sporadic Alzheimer's disease. Neurobiol Aging. 2015;36(9):2468–74.
- <span id="page-8-21"></span>55. Landt J, D'Abrera JC, Holland AJ, et al. Using positron emission tomography and Carbon 11-labeled Pittsburgh Compound B to image Brain Fibrillar β-amyloid in adults with down syndrome: safety, acceptability, and feasibility. Arch Neurol. 2011;68(7):890–6.
- 56. Handen BL, Cohen AD, Channamalappa U, et al. Imaging brain amyloid in nondemented young adults with Down syndrome using Pittsburgh compound B. Alzheimers Dement. 2012;8(6):496–501.
- <span id="page-8-32"></span>57. Rafi MS, Wishnek H, Brewer JB, et al. The down syndrome biomarker initiative (DSBI) pilot: proof of concept for deep phenotyping of Alzheimer's disease biomarkers in down syndrome. Front Behav Neurosci. 2015;9:239.
- <span id="page-8-22"></span>58. Jennings D, Seibyl J, Sabbagh M, et al. Age dependence of brain β-amyloid deposition in Down syndrome: an [18F] forbetaben PET study. Neurology. 2015;84(5):500–7.
- <span id="page-8-23"></span>59. Annus T, Wilson LR, Hong YT, et al. The pattern of amyloid accumulation in the brains of adults with Down syndrome. Alzheimers Dement. 2016;12(5):538–45.
- <span id="page-8-24"></span>60. Cole JH, Annus T, Wilson LR, et al. Brain-predicted age in Down syndrome is associated with beta amyloid deposition and cognitive decline. Neurobiol Aging. 2017;56:41–9.
- <span id="page-8-25"></span>61. Lao PJ, Betthauser TJ, Hillmer AT, et al. The efects of normal aging on amyloid-β deposition in nondemented adults with Down syndrome as imaged by carbon 11-labeled Pittsburgh compound B. Alzheimers Dement. 2016;12(4):380–90.
- <span id="page-8-26"></span>62. Lao PJ, Handen BL, Betthauser TJ, et al. Longitudinal changes in amyloid positron emission tomography and volumetric magnetic resonance imaging in the nondemented Down syndrome population. Alzheimers Dement (Amst). 2017;9:1–9.
- <span id="page-8-27"></span>63. Klunk WE, Price JC, Mathis CA, et al. Amyloid deposition begins in the striatum of presenilin-1 mutation carriers from two unrelated pedigrees. Version 2. J Neurosci. 2007;27(23):6174–84.
- 64. Villemagne VL, Ataka S, Mizuno T, et al. High striatal amyloid beta-peptide deposition across diferent autosomal Alzheimer disease mutation types. Arch Neurol. 2009;66(12):1537–44.
- <span id="page-8-28"></span>65. Cohen AD, McDade E, Christian B, et al. Early striatal amyloid deposition distinguishes Down syndrome and autosomal dominant Alzheimer's disease from late-onset amyloid deposition. Alzheimers Dement. 2018;14(6):743–50.
- <span id="page-8-29"></span>66. Sperling RA, Aisen PS, Beckett LA, et al. Toward defning the preclinical stages of Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement. 2011;7(3):280–92.
- <span id="page-8-30"></span>67. Schöll M, Lockhart SN, Schonhaut DR, et al. PET imaging of tau deposition in the aging human brain. Neuron. 2016;89(5):971–82.
- <span id="page-8-31"></span>68. Lowe VJ, Curran G, Fang P, et al. An autoradiographic evaluation of AV-1451 Tau PET in dementia. Acta Neuropathol Commun. 2016;4:58.
- <span id="page-9-0"></span>69. Rafi MS, Lukic AS, Andrews RD, et al. PET imaging of tau pathology and relationship to amyloid, longitudinal mri, and cognitive change in down syndrome: results from the down syndrome biomarker initiative (DSBI). J Alzheimers Dis. 2017;60(2):439–50.
- <span id="page-9-1"></span>70. Forman MS, Zhukareva V, Bergeron C, et al. Signature tau neuropathology in gray and white matter of corticobasal degeneration. Am J Pathol. 2002;160:2045–53.
- <span id="page-9-2"></span>71. Haier RJ, Head K, Head E, et al. Neuroimaging of individuals with Down's syndrome at-risk for dementia: evidence for possible compensatory events. Neuroimage. 2008;39(3):1324–32.
- <span id="page-9-3"></span>72. Matthews DC, Lukic AS, Andrews RD, et al. Dissociation of Down syndrome and Alzheimer's disease efects with imaging. Alzheimers Dement (N Y). 2016;2(2):69–81.
- <span id="page-9-4"></span>73. Meltzer CC, Zubieta JK, Brandt J, et al. Regional hypometabolism in Alzheimer's disease as measured by positron emission tomography after correction for effects of partial volume averaging. Neurology. 1996;47(2):454–61.
- <span id="page-9-5"></span>74. Hof PR, Bouras C, Perl DP, et al. Age-related distribution of neuropathologic changes in the cerebral cortex of patients with Down's syndrome. Quantitative regional analysis and comparison with Alzheimer's disease. Arch Neurol. 1995;52(4):379–91.
- 75. Teipel SJ, Alexander GE, Schapiro MB, et al. Age-related cortical grey matter reductions in non-demented Down's syndrome adults determined by MRI with voxel-based morphometry. Brain. 2004;127(Pt 4):811–24.
- <span id="page-9-6"></span>76. Mullins D, Daly E, Simmons A, et al. Dementia in Down's syndrome: an MRI comparison with Alzheimer's disease in the general population. J Neurodev Disord. 2013;5(1):19.
- <span id="page-9-7"></span>77. Krasuski JS, Alexander GE, Horwitz B, et al. Relation of medial temporal lobe volumes to age and memory function in nondemented adults with Down's syndrome: implications for the prodromal phase of Alzheimer's disease. Am J Psychiatry. 2002;159(1):74–81.
- <span id="page-9-8"></span>78. Beacher F, Daly E, Simmons A, et al. Alzheimer's disease and Down's syndrome: an in vivo MRI study. Psychol Med. 2009;39(4):675–84.
- <span id="page-9-9"></span>79. Schupf N, Patel B, Pang D, et al. Elevated plasma beta-amyloid peptide Abeta(42) levels, incident dementia, and mortality in Down syndrome. Arch Neurol. 2007;64(7):1007–13.
- <span id="page-9-10"></span>80. Schupf N, Zigman WB, Tang MX, et al. Change in plasma Aß peptides and onset of dementia in adults with Down syndrome. Neurology. 2010;75(18):1639–44.
- <span id="page-9-11"></span>81. Dekker AD, Fortea J, Blesa R, et al. Cerebrospinal fuid biomarkers for Alzheimer's disease in Down syndrome. Alzheimers Dement (Amst). 2017;8:1–10.
- <span id="page-9-12"></span>82. Fortea J, Carmona-Iragui M, Benejam B, et al. Plasma and CSF biomarkers for the diagnosis of Alzheimer's disease in adults with Down syndrome: a cross-sectional study. Lancet Neurol. 2018;17(10):860–9.
- <span id="page-9-13"></span>83. Strydom A, Heslegrave A, Startin CM, et al. Neuroflament light as a blood biomarker for neurodegeneration in Down syndrome. Alzheimers Res Ther. 2018;10(1):39.
- <span id="page-9-14"></span>84. Rafi MS, Donohue MC, Matthews DC, et al. plasma neurofilament light and Alzheimer's disease biomarkers in down

syndrome: results from the down syndrome biomarker initiative (DSBI). J Alzheimers Dis. 2019;70(1):131–8.

- <span id="page-9-15"></span>85. Mapstone M, Gross TJ, Macciardi F, et al. Metabolic correlates of prevalent mild cognitive impairment and Alzheimer's disease in adults with Down syndrome. Alzheimers Dement (Amst). 2020;12(1):e12028.
- <span id="page-9-16"></span>86. Handen BL. The search for biomarkers of Alzheimer's disease in down syndrome. Am J Intellect Dev Disabil. 2020;125(2):97–9.
- <span id="page-9-17"></span>87. Esbensen AJ, Hooper SR, Fidler D, Outcome Measures Working Group, et al. Outcome Measures for Clinical Trials in Down Syndrome. Am J Intellect Dev Disabil. 2017;122(3):247–81.
- <span id="page-9-19"></span>88. Sperling RA, Jack CR Jr, Aisen PS. Testing the right target and right drug at the right stage. Sci Transl Med. 2011;3:111–33.
- <span id="page-9-20"></span>89. Rafi MS, Aisen PS. Alzheimer's disease clinical trials: moving toward successful prevention. CNS Drugs. 2019;33(2):99–106.
- <span id="page-9-21"></span>90. Strydom A, Coppus A, Blesa R, et al. Alzheimer's disease in Down syndrome: an overlooked population for prevention trials. Alzheimers Dement (N Y). 2018;4:703–13.
- <span id="page-9-22"></span>91. Moran JA, Rafi MS, Keller SM, et al. The National Task Group on Intellectual Disabilities and Dementia Practices consensus recommendations for the evaluation and management of dementia in adults with intellectual disabilities. Mayo Clin Proc. 2013;88(8):831–40.
- <span id="page-9-23"></span>92. Walsh DM, Doran E, Silverman W, et al. Rapid assessment of cognitive function in down syndrome across intellectual level and dementia status. J Intellect Disabil Res. 2015;59(11):1071–9.
- <span id="page-9-24"></span>93. Sheehan R, Sinai A, Bass N, et al. Dementia diagnostic criteria in Down syndrome. Int J Geriatr Psychiatry. 2015;30(8):857–63.
- <span id="page-9-25"></span>94. Fenili D, Brown M, Rappaport R, et al. Properties of scylloinositol as a therapeutic treatment of AD-like pathology. J Mol Med (Berl). 2007;85:603–11.
- 95. McLaurin J, Golomb R, Jurewicz A, et al. Inositol stereoisomers stabilize an oligomeric aggregate of Alzheimer amyloid beta peptide and inhibit abeta-induced toxicity. J Biol Chem. 2000;275:18495–502.
- <span id="page-9-26"></span>96. Townsend M, Cleary JP, Mehta T, et al. Orally available compound prevents defcits in memory caused by the Alzheimer amyloid-beta oligomers. Ann Neurol. 2006;60:668–76.
- <span id="page-9-27"></span>97. Salloway S, Sperling R, Keren R, et al. ELND005-AD201 Investigators A phase 2 randomized trial of ELND005, scylloinositol, in mild to moderate Alzheimer disease. Neurology. 2011;77:1253–62.
- <span id="page-9-28"></span>98. Rafi MS, Skotko BG, McDonough ME, et al. A randomized, double-blind, placebo-controlled, phase II study of oral ELND005 (scyllo-Inositol) in young adults with down syndrome without dementia. J Alzheimers Dis. 2017;58(2):401–11.
- <span id="page-9-29"></span>99. Muhs A, Hickman DT, Pihlgren M, et al. Liposomal vaccines with conformation-specific amyloid peptide antigens define immune response and efficacy in APP transgenic mice. Proc Natl Acad Sci USA. 2007;104(23):9810–5.
- <span id="page-9-30"></span>100. Orgogozo JM, Gilman S, Dartigues JF, et al. Subacute meningoencephalitis in a subset of patients with AD after Abeta42 immunization. Neurology. 2003;61(1):46–54.
- <span id="page-9-18"></span>101. Lott IT, Head E. Dementia in down syndrome: unique insights for Alzheimer disease research. Nat Rev Neurol. 2019;15(3):135–47.