

Remarkable Pharmacokinetics of Monoclonal Antibodies: A Quest for an Explanation

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Abstract Monoclonal antibodies (MAbs) usually display slow and limited distribution with combined linear and non-linear elimination mechanisms. While studying individual pharmacokinetic profiles, it was noticed that MAb plasma concentration can vary abruptly over time, with one or more increases after the time to maximum plasma concentration when theoretically the concentration should only decline. This article summarizes the frequency of these additional peaks and assesses whether normal intra-subject and assay variability can explain the observations. For this analysis, we used a benchmark consisting of three registered (adalimumab, bevacizumab, and trastuzumab) and three unregistered immunoglobulin G1 MAbs. At a selected ‘normal’ intra-subject variability of 12%, at least 70% of the study participants (approximately 90% for certain MAbs) still had at least one additional peak, which decreased when the ‘normal’ variability was increased. There was no difference in occurrence between the high- and low-concentration ranges. Only a high sample density was associated with an increased likelihood of detecting additional peaks. Based on the analytical variability for the applied ligand-binding assays (5–10%, up to 15% at the lower limit of quantitation), the number of observed increases was extremely improbable ($p < 0.01$) for most MAbs, especially for the large excursions. Therefore, the

fluctuations are likely genuine. We discuss the possible explanations and the relevance for clinical practice.

Key Points

The plasma concentration–time course of monoclonal antibodies can show considerable fluctuations in individuals that cannot be explained by physiological or assay variability.

More research is required to elucidate which in vivo mechanism(s) are responsible for the observed fluctuations and to determine its relevance for clinical practice.

1 Introduction

Monoclonal antibodies (MAbs) are widely used to treat diseases in almost all fields of medicine. They display highly similar pharmacokinetics with a relatively small volume of distribution and a long half-life. Many of the mechanisms responsible for these properties have been extensively studied and are excellently reviewed elsewhere [1–5].

At the Centre for Human Drug Disease, multiple clinical trials with MAbs are performed annually. When studying their pharmacokinetics, it was noticed that plasma concentrations of MAbs in individuals can follow a remarkable, or even bizarre, time course, characterized by (large) excursions (Fig. 1), which seems to be in disagreement with the current understanding of drug distribution and/or elimination.

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Initially, these findings were disregarded as normal intra-subject and assay variability, also because the mean (group) pharmacokinetic profile usually follows a predictable time course of slow distribution combined with both linear and non-linear elimination. However, after observing fluctuating individual plasma concentrations for an increasing number of MAb, it was considered worthwhile to analyze individual profiles systematically, with the objective to determine if the fluctuations are genuine. A benchmark was used to assess if the result could indeed be explained by normal intra-subject and assay variability, or that other factors may be involved.

2 Methods

Pharmacokinetic data were taken from clinical trials with MAbs in healthy volunteers. Three registered MAbs (adalimumab [Humira[®], AbbVie Ltd, Maidenhead, United Kingdom], bevacizumab [Avastin[®], Roche Pharma AG, Grenzach-Wyhlen, Germany], trastuzumab [Herceptin[®], Roche Pharma AG, Grenzach-Wyhlen, Germany]) were analyzed, as well as three unregistered products (denoted A, B, and C). All drugs were immunoglobulin G1 antibodies and were administered as a single intravenous dose, except for adalimumab, which was administered subcutaneously. The trial participants were healthy subjects, mainly male individuals aged 18–50 years, but some trials included up to 50% female individuals.

All trial procedures were performed in accordance with the different trial protocols. Samples were collected and handled following standard operating procedures. Within each clinical trial, MAb plasma concentration was determined in a single external laboratory and in a single analytical run per participant using validated methods. Product C was quantitated in batches of samples across multiple subjects, resulting in more than one analytical run per participant.

To determine if intra-subject variability for MAb plasma concentration would be related to, for example, changes in circulating plasma volume, the time course of albumin plasma concentration and erythrocyte counts were studied. As albumin and erythrocytes are produced at a relatively constant rate and under normal circumstances do not leave the intravascular compartment, these analytes were considered suitable benchmarks.

Samples for albumin concentration and erythrocyte count were always collected concurrently with samples for MAb concentration, albeit at a lesser frequency. On the administration day, albumin and erythrocytes were quantitated one to five times, depending on the trial protocol. Thereafter, the ratio of the number of these samples to the number of pharmacokinetic samples ranged between 0.5 and 1. The mean of the individual coefficients of variation (CVs) of

albumin concentration per clinical trial ($70 < n < 200$) varied between 3.6 and 4.4% with standard deviations of 1.2–1.2%-point. For the erythrocyte count, the mean ranged between 2.7 and 3.7% and the standard deviations between 0.75 and 1.2%-point, irrespective if only samples collected on the same day (hours apart) or during the full length of the trial (days to weeks apart) were included in the calculation.

Hence, it appears that a conservative reference CV (CV_{ref}) of 4% for normal intra-subject variability (CV_i) is justified. It was subsequently investigated whether the observed fluctuations in MAb plasma concentration exceeded 1, 2, or 3 CV_{ref} , indicating an increasing unlikelihood a change in the profile can be explained by 'normal (physiological) variability'. Additionally, we considered a less conservative CV_{ref} of 8%, covering approximately two CVs of the observed variability for albumin and erythrocytes, and a very extreme variability of 50%. By applying this strategy to pharmacokinetic data for different MAbs, the number of relative maxima in the concentration–time profile (other than the absolute maximum plasma concentration) that cannot be ascribed to the chosen intra-subject variability was counted.

Excursions beyond normal intra-subject variability were identified based on a deviation in the exponential growth constant (λ) outside the margins determined by 1–3 CV_{ref} (4, 8, and 12%) of the conservative variability estimate or of the less conservative CV_{ref} (8, 16, and 24%). The margins for λ per observation were derived from the formula $A \pm CV_i = B \cdot e^{\lambda t}$, where A is the observed plasma concentration, CV_i is the chosen intra-subject variability, B is the plasma concentration of the previous sample, and t is the difference in sample collection time between A and B . Next, the minimum number of unique λ s was determined to describe the observations ($A \pm CV_i$) per individual. A deviation in λ beyond the chosen intra-subject variability was defined as the requirement of two or more unique λ s to describe the rising leg of the plasma concentration curve before a relative (local) maximum was reached (see Supplementary Figure for an example).

This approach assumes linear elimination kinetics, which is known not to be the case for MAbs. However, the plasma concentration–time profile of MAbs usually approximates linearity at the observed (high) MAb plasma concentrations in healthy volunteers where the non-linear elimination mechanism is saturated. Additionally, the non-linearity in elimination manifests as different negative λ s, whereas the focus of this analysis was on deviations from the surrounding data points in the λ in the rising parts of the plasma concentration–time profile. Therefore, this approach was considered fit for purpose.

A separation was made between peaks occurring within the first 24 h from intravenous administration, or 14 days from subcutaneous administration, and thereafter.

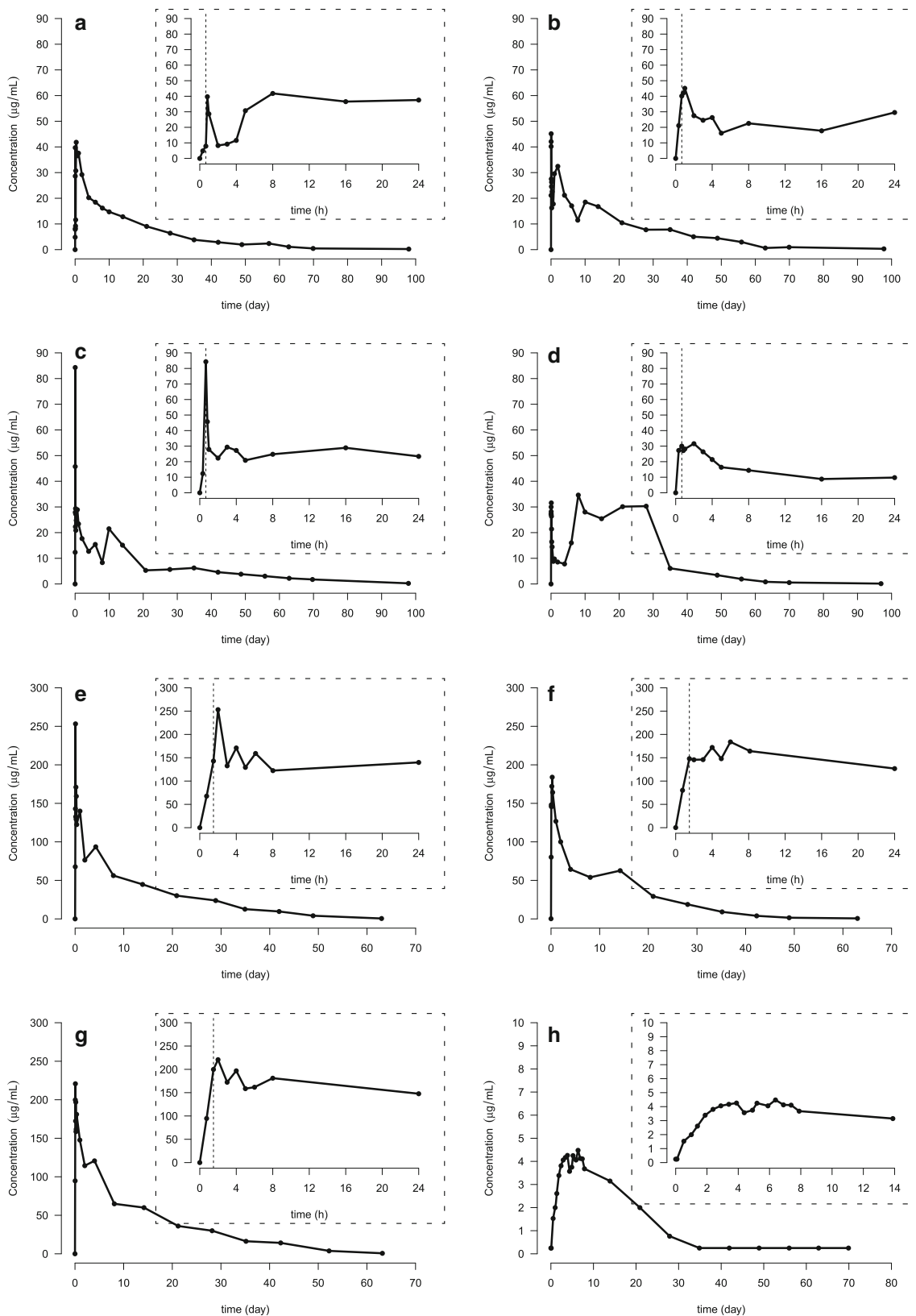


Fig. 1 Individual pharmacokinetic profiles. Representative patterns in individual plasma drug concentrations over time for registered monoclonal antibodies: 2 mg/kg intravenous bevacizumab (a–d), 6 mg/kg intravenous trastuzumab (e–g), and 40 mg subcutaneous

adalimumab (h–j). The insets depict an enlarged section of the first part of the profile. The dashed lines mark the end of intravenous administration

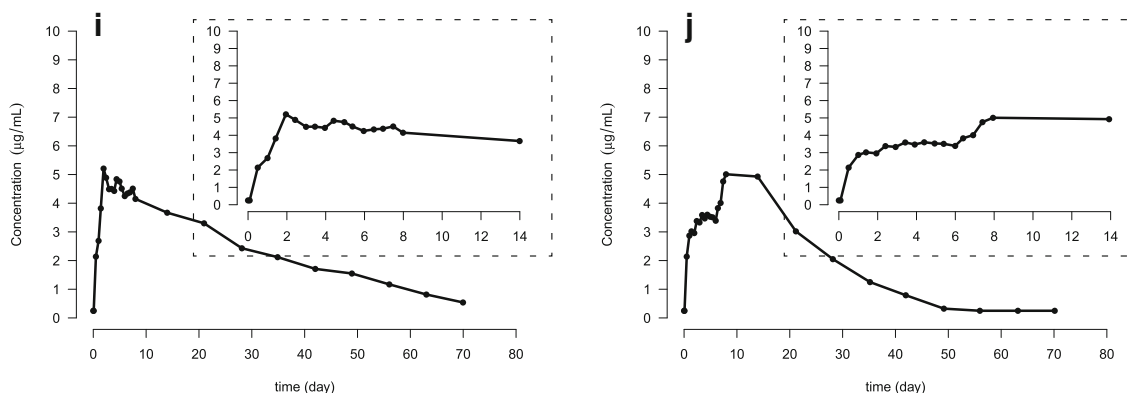


Fig. 1 continued

Additionally, the results of the intravenously administered compounds were stratified based on dose.

To study the potential impact of assay variability, the total number of increases between relative extremes $\geq k$ in the individual plasma concentration–time profiles was compared with the expected number based on the CV of the used bioanalytical assays (CV_{assay}). Here, k is a factor for which values were chosen as 1.12, 1.25, 1.5, 2, 3, 4, and 5, corresponding to increases between 12 and 400%. A one-tailed binomial test was performed to determine the probability (p) of finding at least the observed number of increases $\geq k$. Samples collected before or during infusion were excluded. This analysis could only be performed for intravenously administered MABs.

The expected number of increases $\geq k$ between two consecutive extremes was calculated using the method by Reed and co-workers [6]. This approach assumes that the plasma concentration between two samples remains constant, which results in an underestimation of the observed number of increases $\geq k$, as the plasma concentration theoretically declines after the completion of intravenous administration. Because the p values of increases $\geq k$ at any CV_{assay} derived with the binomial tests are thereby overestimated, thus favoring the probability of increases being attributed to assay variability, this methodological shortcoming was accepted.

According to the regulatory guidelines for ligand-binding assays (the type usually applied when measuring MABs in plasma), the CV_{assay} should not exceed 20%, except at the lower level of quantitation, where it should not exceed 25% [7, 8]. The actual CV_{assay} for the bio-analyses applied in the clinical studies ranged between 5 and 10%, with higher levels (up to 15%) found at the lower level of quantitation. Therefore, p values were obtained at different CV_{assays} from 5 to 25%.

Data analysis was performed with R (Version 2.15.2; R Foundation for Statistical Computing, Vienna, Austria [R Development Core Team 2012]).

3 Results

Pharmacokinetic observations were available for 130 subjects receiving adalimumab (mean 26.1 observations per subject), 90 subjects receiving bevacizumab (mean 26.3), and 46 subjects receiving trastuzumab (mean 19.8). For products A, B, and C, data were available from respectively 18, 33, and 7 subjects, with a mean number of observations of 18.9, 15.1, and 17.4, respectively. The samples collected during the first 7 days after administration summed 16 for adalimumab, 15 for bevacizumab, 12 for trastuzumab, 12 for A, 10 for B, and 10 for C.

Tables 1 and 2 present the number of subjects with additional maxima beyond increasing CV_i . This shows that with a conservative CV_i of 4% virtually all subjects had an additional peak in their profile. Even if the variability considered normal is increased to 3 CV_{ref} (12%), at least 70% of the study participants (approximately 90% for certain MABs) still had at least one additional peak, with the exception of products B and C. It should be noted, however, that for both products B and C a sparse sampling scheme was used compared with the other MABs, with fewer than five samples collected during the first 24 h. This may have limited the chance to identify short-lasting concentration changes.

At a variability of 24%, 60–70% of subjects who received bevacizumab, trastuzumab, or product A showed an unexplained additional relative maximum, a percentage that decreased further and became more dispersed among the MABs at a variability of 50%. For adalimumab, product B and C, the corresponding numbers were again lower, although the overall pattern observed with increasing intra-subject variability was similar for all investigated MABs. Even when considering an intra-subject variability of 24 or 50% as normal, which is well beyond the variability (CV_{ref}) observed for albumin and erythrocytes, additional peaks remain.

The probability of finding a number of increases with a certain magnitude in plasma concentration rose with increasing assay variability (Table 3). Conversely, the

Table 1 Additional peak concentrations after intravenous administration

MAB/period (h)	Variability, <i>n</i> (%)					
	4%	8%	12%	16%	24%	50%
Dose ≤3 mg/kg						
Bevacizumab (<i>n</i> = 90)						
Total	88 (97.8)	87 (96.7)	83 (92.2)	72 (80.0)	65 (72.2)	41 (45.6)
<24	82 (91.1)	79 (87.8)	68 (75.6)	54 (60.0)	46 (51.1)	25 (27.8)
≥24	73 (81.1)	70 (77.8)	66 (73.3)	61 (67.8)	56 (62.2)	30 (33.3)
A (<i>n</i> = 18)						
Total	18 (100)	17 (94.4)	16 (88.9)	16 (88.9)	12 (66.7)	1 (5.6)
<24	18 (100)	17 (94.4)	16 (88.9)	16 (88.9)	12 (66.7)	1 (5.6)
≥24	4 (22.2)	4 (22.2)	4 (22.2)	4 (22.2)	4 (22.2)	0
B (<i>n</i> = 33)						
Total	12 (36.4)	12 (36.4)	7 (21.2)	5 (15.2)	3 (9.1)	1 (3.0)
<24	8 (24.2)	8 (24.2)	4 (12.1)	2 (6.1)	1 (3.0)	0
≥24	4 (12.1)	4 (12.1)	3 (9.1)	3 (9.1)	2 (6.1)	1 (3.0)
Dose >3 mg/kg						
Trastuzumab (<i>n</i> = 46)						
Total	46 (100)	44 (95.7)	42 (91.3)	37 (80.4)	28 (60.9)	8 (17.4)
<24	45 (97.8)	42 (91.3)	39 (84.8)	33 (71.7)	22 (47.8)	3 (6.5)
≥24	21 (45.7)	21 (45.7)	20 (43.5)	20 (43.5)	16 (34.8)	5 (10.9)
C (<i>n</i> = 7)						
Total	3 (42.9)	3 (42.9)	2 (28.6)	1 (14.3)	0	0
<24	2 (28.6)	2 (28.6)	2 (28.6)	1 (14.3)	0	0
≥24	1 (14.3)	1 (14.3)	0	0	0	0

Number (%) of subjects with at least one peak concentration other than the maximum plasma concentration beyond 'normal' intra-subject variability, for which values between 4 and 50% were chosen. Values are displayed per MAB. A separation is made between peaks occurring within the first 24 h from administration and thereafter. A, B, and C are unregistered IgG1 antibodies

MAB monoclonal antibody, IgG1 immunoglobulin G1

Table 2 Additional peak concentrations after subcutaneous administration

Period (days)	Variability, <i>n</i> (%)					
	4%	8%	12%	16%	24%	50%
Total	128 (98.5)	118 (90.8)	90 (69.2)	72 (55.4)	54 (41.5)	19 (14.6)
<14	127 (97.7)	116 (89.2)	87 (66.9)	70 (53.8)	48 (36.9)	15 (11.5)
≥14	13 (10.0)	13 (10.0)	12 (9.2)	11 (8.5)	11 (8.5)	8 (6.2)

Number (%) of subjects with at least one peak concentration other than the maximum plasma concentration beyond 'normal' intra-subject variability after a single subcutaneous dose of 40 mg adalimumab (*n* = 130). Values for normal intra-subject variability were chosen between 4 and 50%. A separation is made between peaks occurring within the first 14 days from administration and thereafter

probability was lower for larger excursions. Within the actual CV range for the used ligand-binding assays (5–10%), the number of observed increases was extremely improbable based on assay variability, except for product C. Even at higher CV_{assays}, which are only accepted at the lower level of quantitation (up to 15% for the used assays), assay variability must be considered unlikely in causing the observed increases, especially for those with a large amplitude.

A relationship between standard demographic parameters (age, [lean] body weight, body mass index) and the number or magnitude of additional peaks could not be detected, although it should be noted that, as a result of the trial protocols, the populations were highly homogeneous with regard to these parameters. Additionally, across the different trials, demographic variability was limited.

Table 3 Probability of increases based on assay variability

MAb/factor (<i>k</i>)	Observed	CV_{assay}				
		5%	10%	15%	20%	25%
Dose ≤ 3 mg/kg						
Bevacizumab						
1.12	228	$<10^{-93}$	$<10^{-5}$	0.86	1.00	1.00
1.25	153	$<10^{-99}$	$<10^{-37}$	$<10^{-3}$	0.97	1.00
1.5	78	$<10^{-99}$	$<10^{-99}$	$<10^{-20}$	0.01	0.99
2	23	$<10^{-99}$	$<10^{-99}$	$<10^{-31}$	$<10^{-7}$	0.20
3	8	$<10^{-99}$	$<10^{-97}$	$<10^{-37}$	$<10^{-16}$	$<10^{-6}$
4	3	$<10^{-99}$	$<10^{-59}$	$<10^{-23}$	$<10^{-11}$	$<10^{-5}$
5	2	$<10^{-99}$	$<10^{-53}$	$<10^{-22}$	$<10^{-11}$	$<10^{-5}$
<i>A</i>						
1.12	34	$<10^{-18}$	$<10^{-3}$	0.10	0.39	0.62
1.25	19	$<10^{-39}$	$<10^{-5}$	0.08	0.64	0.92
1.5	10	$<10^{-70}$	$<10^{-14}$	$<10^{-3}$	0.16	0.73
2	1	$<10^{-20}$	$<10^{-4}$	0.05	0.47	0.89
3	0					
4	0					
5	0					
<i>B</i>						
1.12	3	0.76	1.00	1.00	1.00	1.00
1.25	1	0.06	0.99	1.00	1.00	1.00
1.5	1	$<10^{-6}$	0.14	0.86	1.00	1.00
2	1	$<10^{-20}$	$<10^{-4}$	0.04	0.38	0.82
3	1	$<10^{-52}$	$<10^{-12}$	$<10^{-5}$	$<10^{-2}$	0.06
4	0					
5	0					
Dose > 3 mg/kg						
Trastuzumab						
1.12	75	$<10^{-29}$	0.02	0.90	1.00	1.00
1.25	26	$<10^{-43}$	0.01	1.00	1.00	1.00
1.5	5	$<10^{-31}$	$<10^{-3}$	0.89	1.00	1.00
2	1	$<10^{-19}$	$<10^{-3}$	0.14	0.85	1.00
3	1	$<10^{-51}$	$<10^{-12}$	$<10^{-4}$	0.01	0.21
4	1	$<10^{-82}$	$<10^{-19}$	$<10^{-8}$	$<10^{-3}$	$<10^{-2}$
5	0					
<i>C</i>						
1.12	3	0.07	0.76	0.94	0.97	0.99
1.25	0					
1.5	0					
2	0					
3	0					
4	0					
5	0					

Probability (p) of finding the observed number of increases $\geq k$ in plasma concentration based on the CV_{assay} . Only increases after completion of intravenous administration are considered

CV_{assay} assay's coefficient of variation, *MAB* monoclonal antibody

4 Discussion

In this article, it is reported that MABs may show unexpected and remarkable pharmacokinetic behavior with increases in plasma concentrations at the time the compound is cleared. These increases, which are occasionally substantial and long lasting, cannot be explained by physiological or assay variability. There was no difference in occurrence between the high- and low-concentration ranges. Only a high sample density seemed to be associated with an increased likelihood of detecting additional peaks.

When observing fluctuations, especially increases, in the concentration of a drug over time that theoretically should decline steadily, there are a few explanations to consider. First, pre-analytical errors such as not disconnecting and removal of the infusion material upon stopping intravenous administration, sample switching, applying incorrect dilutions, or calculation errors, should be ruled out. Subsequently, the assay performance should be considered critically, including, among others, assay precision, within and between-run variability, limit of quantitation, and effects of freeze-thaw cycles.

For multiple reasons, assay variability or interference was considered unlikely to explain our observations. First, a vast number of additional peaks were counted (Table 3). Additionally, the finding that comparable fluctuations were observed for all investigated MABs, in each assay, both in the low- and high-concentration ranges, and at any moment in time after administration (Fig. 1) argues against an assay-related explanation. Furthermore, the data points before or after the peak often confirmed the relatively high concentration or suggested a steady increase towards the maximum, respectively, a decrease following the maximum. These observations are generally not compatible with the randomness one expects to arise from assay variability.

Another explanation to consider is physiological variability, as for instance, changes in volume status over time may alter the concentration of the MAB in plasma, while the absolute quantity in the body remains unchanged. Fluid shifts were recently postulated by Van Iersel et al. [9] as the underlying mechanism for the postural changes in MAB concentrations that they had observed. Similar day-to-day variability was seen in our study with adalimumab. In that clinical trial, pharmacokinetic samples were collected 12 h apart during the first week (Fig. 1h–j). The evening concentrations (0.5, 1.5, 2.5, ... days after administration) were higher than the morning concentrations (1, 2, 3, ... days after administration), with a mean difference of 13.3% (standard deviation 10.5%-point) per participant. It should be noted, however, that for the main part of the adalimumab trial the participants were ambulatory and traveled

both in the morning and evening to the clinical unit, making postural changes unlikely.

Additionally, the magnitude of many of the remaining fluctuations in plasma concentration for the investigated MABs (Fig. 1) exceeded by far the reported increases by Van Iersel et al. [9], and what would be physiologically achievable as a result of contraction of the plasma volume. Furthermore, concurrent changes of equal magnitude in intravascularly distributed endogenous substances with a relatively constant production, such as albumin and erythrocytes, were not seen, which is not in keeping with the fluid shift hypothesis. In conclusion, we argue that the majority of the observed fluctuations in the profiles cannot be explained by physiological or assay variability and should be considered genuine.

Now that we have demonstrated that the observed fluctuations in MAB pharmacokinetic profiles are likely to be genuine, a few considerations are warranted. First, the occurrence of additional peaks immediately following administration (<24 h for intravenous and <14 days for subcutaneous) was usually higher than in the period thereafter, regardless of the chosen value for normal intra-subject variability. An explanation for this phenomenon may be that the sampling frequency is usually decreased over time, thereby reducing the chance to identify relative extremes. Additionally, some MABs had relatively short profiles, and thus a limited number of data points after 24 h, as was the case for products A and B.

Next, the question arises as to which physiological mechanism may be responsible for the phenomenon of fluctuating plasma concentrations. One explanation comprises the capture and subsequent release of MABs by tissues or components, which would presumably be large quantities of MABs, given the observed magnitude of the excursions, with increases of 50% or more (Table 3). Moreover, a MAB is presumably released quite rapidly, as the changes over time in certain cases approaches the infusion rate of intravenous administration (Fig. 1). Earlier, we demonstrated the endothelium to be a potential candidate for dynamically binding biopharmaceuticals [10]. Nonetheless, there may be other locations where MABs can be stored temporally. For example, can MABs simply pool in the venous compartment or in less perfused organs? Does an extravascular reservoir exist? Which physiological or pathophysiological mechanism underlies the release ('auto-injection') of the MAB into the circulation?

Considering daily life, the redistribution of blood flow to various organs during alimentation (gastrointestinal system), resting, and physical exercise (muscles) may either mask or expose sites for adsorption, absorption, and elimination, or, in contrast, flush out pooled or adsorbed MABs in these organs. Possibly, changes in the local milieu

(e.g., pH), competition for adsorption sites by other substances, and modifications to structural components involved in the binding or transport of MAbs can mediate the release into the circulation. However, without dedicated research on the distribution of MAbs over the body, these options remain speculative at best.

MAbs are designed to specifically bind a particular target and the resulting complex is internalized and subsequently degraded by either immune cells or the target cell [3, 5]. Therefore, this elimination process cannot contribute to increases in the plasma concentration of a MAb. However, it is conceivable that an abrupt decrease in the target-mediated elimination route, for example, because of down-regulation of the target following exposure to an abundance of circulating MAbs, can acutely elevate plasma concentrations, provided that there is continuous absorption of the MAb into the plasma compartment, as with subcutaneous administration. Other prerequisites for this possible explanation are a relative high absorption rate and a significant contribution of the target-mediated pathway to the total elimination of the MAb, which does not seem to be the case, based on published values regarding absorption and elimination rates [2, 11]. By analogy, although variations in the absorption rate over time after subcutaneous administration can theoretically change plasma concentrations of MAbs, the absorption of MAbs from a subcutaneous depot into the circulation is generally understood to be slow [4], which is not in line with the observed rapid and large excursions.

The neonatal Fc receptor (FcRn, or Brambell receptor) requires special consideration. Binding of a MAb to this receptor does not result in lysosomal degradation, but returns the MAb-FcRn complex to the cell membrane [1–3, 12]. Such recycling of MAbs to the vascular compartment may contribute to fluctuations in plasma concentrations, as MAbs can be temporarily sequestered from the circulation [13]. However, research suggests the transportation of immunoglobulins by FcRn is quite rapid [14]. Another function of FcRn is transcytosis of immunoglobulins, including MAbs. According to current understanding, distribution of MAbs to tissues is slow and limited [2–5], which suggests this process cannot explain our observations. Furthermore, albumin is also a substrate of FcRn [12], and comparable fluctuations in its concentration have not been documented. Nonetheless, involvement of the FcRn cannot be ruled out, although it would be interesting to know which factors in that case can trigger abrupt changes in FcRn-mediated transcellular transport rate of MAbs.

An important question to be answered is what the clinical relevance of fluctuations in plasma concentrations over time could be. Assuming that plasma concentration is a key determinant to achieve therapeutic concentrations at the

site of action, measuring and understanding variations in plasma concentrations over time is probably pivotal. Therefore, we hope to initiate a broad discussion within the field on possible explanations for the observed phenomena, as well as how to increase more fundamental knowledge of the pharmacokinetics of MAbs.

5 Conclusion

The plasma concentration of MAbs can vary abruptly and to a great extent, which cannot be explained by normal physiological or assay variability. Future studies are required to elucidate this phenomenon and to determine its relevance for clinical practice.

Compliance with Ethical Standards

Funding No funding was received for this study.

Conflict of interest Authors JR, MM, and JB declare that they have no conflicts of interest.

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References

1. Wang W, Wang EQ, Balthasar JP. Monoclonal antibody pharmacokinetics and pharmacodynamics. *Clin Pharmacol Ther.* 2008;84:548–58.
2. Dirks NL, Meibohm B. Population pharmacokinetics of therapeutic monoclonal antibodies. *Clin Pharmacokinet.* 2010;49:633–59.
3. Keizer RJ, Huitema ADR, Schellens JHM, Beijnen JH. Clinical pharmacokinetics of therapeutic monoclonal antibodies. *Clin Pharmacokinet.* 2010;49:493–507.
4. Dostalek M, Gardner I, Gurbaxani BM, et al. Pharmacokinetics, pharmacodynamics and physiologically-based pharmacokinetic modelling of monoclonal antibodies. *Clin Pharmacokinet.* 2013;52:83–124.
5. Shi S. Biologics: an update and challenge of their pharmacokinetics. *Curr Drug Metab.* 2014;15:271–90.
6. Reed GF, Lynn F, Meade BD. Use of coefficient of variation in assessing variability of quantitative assays. *Clin Diagn Lab Immunol.* 2002;9:1235–9.
7. Committee for Medicinal Products for Human Use. Guideline on bioanalytical method validation. EMEA Comm Med Prod Hum Use. 2012;44:1–23.
8. US Food and Drug Administration. Guidance for industry: bioanalytical method validation (draft). Washington, DC: US Department of Health and Human Services; 2013.
9. Van Iersel MP, Velinova M, Lutgerink R. A change in posture significantly affects plasma concentrations of immunoglobulin G, such as monoclonal antibodies. *Clin Pharmacol Drug Dev.* 2016;5:6.

10. Reijers JAA, Dane MJC, Moerland M, et al. Biotherapeutics: how much do we actually administer and where does it go? *Proc Br Pharmacol Soc.* 2014;12:105.
11. Ternant D, Bejan-Angoulvant T, Passot C, et al. Clinical pharmacokinetics and pharmacodynamics of monoclonal antibodies approved to treat rheumatoid arthritis. *Clin Pharmacokinet.* 2015;54:1107–23.
12. Martins JP, Kennedy PJ, Santos HA, et al. A comprehensive review of the neonatal Fc receptor and its application in drug delivery. *Pharmacol Ther.* 2016;161:22–39.
13. Daugherty AL, Mersny RJ. Formulation and delivery issues for monoclonal antibody therapeutics. *Adv Drug Deliv Rev.* 2006;58:686–706.
14. Tzaban S, Massol RH, Yen E, et al. The recycling and transcytotic pathways for IgG transport by FcRn are distinct and display an inherent polarity. *J Cell Biol.* 2009;185:673–84.