### **ORIGINAL RESEARCH**



# **Magnetic dual‑responsive semi‑IPN nanogels based on chitosan/ PNVCL and study on BSA release behavior**

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### **Abstract**

Magnetic thermoresponsive nanogels present a promising new approach for targeted drug delivery. In the present study, bovine serum albumin (BSA) loaded thermo-responsive magnetic semi-IPN nanogels (MTRSI-NGs) were developed. At frst poly(*N*-vinyl caprolactam) (PNVCL) was synthesized by free radical polymerization and then MTRSI-NGs were prepared by crosslinking chitosan in presence of chitosan and  $Fe<sub>3</sub>O<sub>4</sub>$ . The formation of MTRSI-NGs has been confirmed by FTIR, and the average molecular weight of PNVCL was determined by GPC analysis. Rheological and turbidimetry analysis were used to determine lower critical solution temperature (LCST) of PNVCL and magnetic thermo-responsive nanogels (MTRSI-NGs) around 32 and 37 °C, respectively. FE-SEM analysis showed particle size at less than 20 nm in the dried state. Dynamic light scattering determined particle size at about 30 nm in a swelling state. The analysis of release behavior showed that the BSA release ratio at 40 °C was faster than 25 °C. The pH release behavior was evaluated at pH 5.5 and 7.4 and showed that the drug release rate at pH 5.5 was more rapid than pH 7.4. The results show MTRSI-NGs are applicable to protein targeted delivery by thermosensitive targeted drug delivery systems.

**Keywords** Magnetic thermoresponsive nanogels · Poly(*N*-vinyl caprolactam) · Chitosan nanogel · Albumin release behavior · Target drug delivery · Semi-IPN nanogels

# **Introduction**

Over the last quarter-century, several approaches have been utilized to improve drug efficacy. Different types of drugs such as small molecules, proteins, genes, nucleic acids, etc. are available. Diferent types of drugs make a great challenge between efficacy and bioavailability. Common drug usages have been encountered with short plasma circulation

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half-life, susceptibility to rapid degradation in plasma, low aqueous solubility, and high toxicity (Butler et al. [2014](#page-9-0)). Controlled drug delivery technology has offered many advantages in comparison to conventional drug usage. Smart polymers in drug delivery have excellent potential for improvement of pharmacokinetic properties and increase intracellular uptake of nanocarriers.

Environmental sensitive polymers are one of the most well-known biomaterials which have been applied in drug delivery systems. Macromolecules provide many ways of creating stimuli-sensitive behavior. Smart materials or stimuli-responsive materials can change their behavior by small environmental variations. Stimuli-sensitive polymers can respond to one or more stimuli such as chemical, temperature, light intensity, electric or magnetic pH, feld, biological application (Alibolandi et al. [2016;](#page-9-1) Aguilar et al. [2019](#page-9-2); Wang et al. [2014;](#page-10-0) Ward et al. [2011;](#page-10-1) Zhuang et al. [2013](#page-10-2)). Many proteins and drugs are not accessed to specific tissues in sufficient dosages. Drug carriers require precise physicochemical design for targeting such as nanoparticles developed for imaging purposes. Incorporation of magnetic nanoparticles into drugs or protein carriers provide



advantages, for instance, local protein release, reduction of other tissues drug exposure and minimize side efects, accelerate protein transfer across cell membranes, prevent rapid metabolism or excretion from the body. Magnetic nanocarriers are injected intravenously. The mechanistic action of MTRSI-NGs in the body occurs by three following successions: frst, magnetically conducting MTRSI-NGs to targeted tissue, second, the magnet leads to the increasing temperature of  $Fe<sub>3</sub>O<sub>4</sub>$  and then heat transfer to thermoresponsive nanogels to LCST until to protein release, third wipe out body from MTRSI-NGs (He et al. [2013;](#page-9-3) Smith et al. [2021](#page-9-4); Uhrich et al [1999](#page-9-5)).Cytotoxicity of PNVCL was investigated by Han et al. The results showed that PNVCL did not show toxicity with short-term cell exposure. PNVCL withstand increases in temperature, while others change from twophase to single-phase by increasing temperature (Dinari et al. [2021](#page-9-6); Fernández-Quiroz et al. [2015](#page-9-7); Siirilä et al. [2019](#page-9-8); Vihola et al. [2005](#page-9-9)).Temperature responsive polymers present an excellent hydrophobic-hydrophilic balance in their structure. The slight change in temperature near the cloud point makes the chains collapse. The presence of hydrophilic or hydrophobic polymers in thermo-responsive chains leads to change responding in an aqueous medium and eventually obtain a polymer with new hydrophilic and hydrophilic balances. Most polymers are soluble in water when it is heated, but some water-soluble polymers may collapse from the solution upon heating (Schmidt [2007](#page-9-10); Weber et al. [2012](#page-10-3)). These unique features of thermo-responsive polymers that exhibit biphasic when it's heated above the specifc temperature, recognized as a lower critical solution temperature (LCST). The phase transition temperature depends on the hydrogen bonding between water molecules and the functional groups of polymers. Phase or reversible conformational changes take place for thermo-responsive polymers over small variations of temperature. Diferent types of polymeric materials and systems are known which exhibit thermo-responsive behavior, including hydrogels with a temperature-depending swelling behavior, membranes of thermo-responsive permeability, and materials possessing a change of shape or color upon reaching a certain temperature (Rinaudo [2006](#page-9-11)). Poly(*N*-vinylcaprolactam) is one of the most popular thermo-responsive polymers that exhibit phase transition temperature around 32 °C. The phase transition temperature can be modifed by copolymerization with hydrophobic or hydrophilic polymers (Rao et al [2016](#page-9-12)). Polysaccharides are well-known hydrophilic biocompatible polymers which are used extensively in drug delivery systems (Yadav and karthikeyan [2019](#page-10-4)). Among various polysaccharides, chitosan is extensively studied as a nanocarrier of drugs and proteins. Biocompatibility, hydrophilic properties, pH-sensitive, electrostatic force are widely used for target delivery purposes. Phase transition temperature can tune by incorporation of chitosan to PNVCL and crosslinking of



chitosan in presence of PNVCL and  $Fe<sub>3</sub>O<sub>4</sub>$ , leads to form thermo-pH responsive magnetic carrier for targeted drug delivery system (Abedini et al. [2018\)](#page-9-13). Banihashemi et al. [\(2020](#page-9-14)) investigated the potential of nanocarriers of chitosang-PNVCL nanofbers coated with gold–gold sulfde nanoparticles, for controlled release of cisplatin. They found that the higher and faster rate of release was at T>LCST and pH 5.In this study, novel magnetic thermo/pH responsive semi-IPN nanogels were prepared. The phase transition temperature was controlled near human body temperature. Chitosan (CS) was used as an LCST modifer which increases the PNVCL phase transition temperature and it controls the human body. Bovine serum albumin (BSA) was loaded as a model of the protein drug. Two diferent temperatures were selected to determine drug release behavior at a lower and higher temperature of LCST point. The pH release behavior was reported under two diferent pH values at 7.4 and 5.5. The novelty of this study is the preparation of thermo-pH dual responsive magnetic nanogels containing hydrophilic protein which exhibits phase transition temperature in the human body, providing new routes for drug delivery systems. Furthermore, it is to investigate the effect of chitosan and magnetic nanoparticles on the phase transition temperature of PNVCL, and evaluate the potential of MTRSI-NGs for use in targeted drug delivery systems as a protein carrier.

# **Materials and methods**

### **Materials**

*N*-Vinylcaprolactam (NVCL 98%), chitosan medium molecular weight, thioglycolic acid (TG), isopropyl alcohol (IPA), azobisisobutyronitrile (AIBN), sodium tripolyphosphate, 2,2,6,6-tetramethylpiperidin-1-yl)oxyl or (2,2,6,6-tetramethylpiperidin-1-yl)oxidanyl(tempo), FeCl<sub>2</sub>·4H<sub>2</sub>O, FeCl<sub>3</sub>·6H<sub>2</sub>O, ammonia were purchased from Sigma Aldrich. NaCl, KCl, Na<sub>2</sub>HPO<sub>4</sub> 2H<sub>2</sub>O, KH<sub>2</sub>PO<sub>4</sub>, *N*-hexane supplied from Merck chemical company.

### **Synthesis of PNVCL**

PNVCL was synthesized by controlled free-radical polymerization in IPA solution. First, a mixture of NVCL, TG as a chain transfer agent, and the solvent was added to a sealed fask. The oxygen was removed using a vacuum. Then, AIBN was added to the reaction mixture. The reaction was carried out under a nitrogen atmosphere at 70 °C. After the desired time, the polymerization was stopped and the product was purifed by adding anti-solvent *N*-hexane and it was fltered through a Tefon flter under vacuum conditions.

#### **Preparation of magnetite nanoparticles**

Iron oxide nanoparticles (Fe<sub>3</sub>O<sub>4</sub>) were prepared through a controlled chemical co-precipitation method (Kozanoğlu et al.  $2011$ ). First, 1 mL of FeCl<sub>2</sub> 4H<sub>2</sub>O 2 M was added into 4 mL of FeCl<sub>3</sub> 6H<sub>2</sub>O, 1 M solution (1:2, molar ratio) and mixed for a half hour. Then, 30 mL of 1 M ammonia was incorporated into the mixture under a high-speed mechanical mixer equipped Tefon blade until the precipitate was formed. The precipitated iron oxide was washed three times with deionized water to remove the excess amount of ammonia.

# **Preparation of magnetic thermo‑responsive chitosan poly(N‑vinylcaprolactam) semi‑IPN nanoparticles (MTRSI‑NPs)**

Magnetic semi-IPN nanogels were prepared through the suspension cross-linking technique. This method was performed in the 150 mL three-necked fask equipped with a mechanical mixer equipped Tefon blade, condenser, and nitrogen. The poly(*N*-vinylcaprolactam) (PNVCL) was prepared as described previously. An amount of 1 g of PNVCL was dissolved in acetic acid (1% solution) and 0.12 g of chitosan and  $Fe<sub>3</sub>O<sub>4</sub>$ was added to a solution. The molar ratio of chitosan and  $Fe<sub>3</sub>O<sub>4</sub>$  nanoparticles was 1:1 (w/w). Finally, the resulting solution was mixed for 30 min under 1000 rpm and crosslinked by (sodium tripolyphosphate) TPP in a 1:6 (w/w) ratio. The solution was under high-speed mixing at about 1000 rpm until the nanogels formed. The obtained suspension was centrifuged at 4000 rpm for 15 min. To remove the non-reacted component, magnetic nanogels were washed with deionized water several times and centrifuged under previously described conditions. The purifed nanogels were dried in a vacuum oven at 30 °C.(see Fig. [1\)](#page-3-0).

### **Characterization techniques**

### **FTIR spectral studies**

Fourier transform infrared (FTIR) measurements were carried to characterize PNVCL polymerization and formation of magnetic chitosan-PNVCL semi-IPN according to the KBr technique using a Bruker-IFS-48 FT-IR spectrometer (Ettlingen, Germany) in the range of 400–5000  $cm^{-1}$ .

#### **Molecular weight analysis**

The apparent molecular weights and molecular weight distributions were measured by gel permeation chromatography (GPC) using Agilent-1100 series equipped with a refractive index detector. The solution containing the desired polymer is pumped into a column containing porous particles. GPC is a chromatographic method in which completely dissolved molecules of a polymer are separated by porous columns depending on the size of the molecules. The molecular weight of the test samples is relative and is obtained by comparing its exit time with the calibration curve. This test was performed by a permeable gel chromatography device to determine the average molecular mass of PNVCL polymer samples at 23 °C. First some of the sample was dissolved in tetrahydrofuran as eluent and then in an injection machine with a volume of 20 μL, which was performed at a flow rate of 1 mL/min.

#### **Rheological analysis**

Rheology measurements of 10% w/w thermo-responsive suspension/buffer phosphate solutions (pH 7.4) were undertaken using an Anton Paarrheometer with a 50 MM and angle 1 in cone geometry. Solutions were cooled to 5 °C Measurements of complex viscosity  $(g^*)$ , viscous modulus (G″), and elastic modulus (G′) were taken over a range of 5–65  $\degree$ C and a heating rate of 1  $\degree$ C/min.

#### **UV–visible spectroscopy**

The cloud point of the nanogels as a parameter of LCST determination was analyzed by Shimadzu UV–visible spectrophotometer equipped with a temperature controller. The average of three values was taken as the LCST of nanogels.

#### **Field‑emission scanning electron microscopy**

To investigate the particle size in dried state FE-SEMMIRA3TES-CAN-XMU was utilized. At frst, nanogels were dried by freezedrying technique at− 80 for 24 h, and then nanogels were set on brass equipped with doubled side adhesive and then sputter-coated with gold to enable electric conductivity (Polaron SC515 sputter coater, Fison instruments, Sussex, UK). The measurements were done with an accelerating voltage of 10 kV.

#### **Dynamic light scattering**

Dynamic light scattering is a physical technique for measuring particle size in emulsions and suspensions. It is non-destructive and fast to determine the particle size in the range of nm to the micrometer. The measurement was performed with SZ-100 Horbia Join Joyovin in 25 °C and 655 nm.



# **BSA loading efficacy**

Prior to the determination of protein release from the MTRSI-NGs reservoir, loading efficiency was evaluated. The protein encapsulation consists of mixing a certain amount of BSA and MTRSI-NGs (0.2 mg/mL) compound before forming MTRSI-NGs. The protein loading efficiency (LE) and loading capacity (LC) of MTRSI-NGs were evaluated by measuring free BSA unloaded after encapsulation. To measure residual free BSA, the MTRSI-NGs were centrifuged at 4000 rpm for 5 min, and the supernatant was measured using UV–spectrophotometry. The LE and LC were calculated as the following equation (Cheng et al. [2015\)](#page-9-16):

(0.2 mg/mL) were added into dialysis bags. Finally, dialysis bags were immersed in 250 mL and the temperatures were set at 25 and 40 °C under mechanical stirrer 50 rpm. The BSA pH release from MTRSI-NPs was measured against PBS (pH7.4 and 5.5) containing at 37 °C. The 3 mL of the medium was withdrawn at the determined time at every 24 h, and 3 mL fresh medium was supplemented. The released amount of BSA was determined using UV–Visible spectroscopy. The release of BSA was expressed as a percentage of the released BSA plotted as a function of time (Zuo et al. [2017](#page-10-5)).

$$
LE (\%) = \frac{\text{Total amount of BSA} - \text{Free BSA Total amount of BSA in supernatant}}{\text{Total amount of BSA}} \times 100
$$
 (1)

$$
LC (\%) = \frac{\text{Total amount of BSA} - \text{Free BSA}}{\text{Dried nanoparticle weight}} \times 100 \quad (2)
$$

# **BSA release study**

The study of drug release in a temperature-responsive state was performed using a phosphate buffer solution (pH 7.4) at  $25 \pm 1$  and  $40 \pm 1$  °C. BSA loaded MTRSI-NPs nanoparticles

# **Results and discussion**

# **FTIR spectral studies**

FTIR spectroscopy has been applied to confrm the polymerization of NVCL and other prepared systems. The presence of the amide I band depends on the degree of the hydrogen bonding and the physical state of the compound. FTIR spectra of poly(*N*-vinycaprolactam) and *N*-vinylcaprolactam



<span id="page-3-0"></span>**Fig. 1** Scheme of PNVCL synthesis method, and MTRSI-NGs preparation technique



<span id="page-4-0"></span>**Fig. 2** FTIR spectra of NVCL monomer and PNVCL





<span id="page-4-1"></span>**Fig. 3** FTIR spectra of crosslinked chitosan, PNVCL, chitosan PNVCL semi-IPN and MTRSI-NGs

monomer are shown in Fig. [2.](#page-4-0) The broadband at  $3441 \text{ cm}^{-1}$ is attributed to the OH stretching of carboxyl groups in the synthesized polymer. The vibration at 2930 and 1433 cm<sup>-1</sup> assigned to the aliphatic C–H and  $-CH<sub>2</sub>$  stretching. The dip of 1716 and 1637 cm<sup>-1</sup> are associated with C=O and amide I band vibrations. The vinyl peaks,  $(CH \text{ and } CH_2)$  in the spectrum of the monomer at 3105 and 981 cm<sup>-1</sup>are disappeared. To confrm the polymerization of NVCL, FTIR spectra of the monomer and the polymer are shown in Fig. [2.](#page-4-0) It can be concluded from FTIR spectra analysis that the polymer was successfully synthesized and the polymerization proceeds by carbon–carbon double bond opening without any change in the *N*-vinylcaprolactam ring (Kozanoğlu et al. [2011](#page-9-15)).

Figure [3](#page-4-1) demonstrated the FTIR spectra of crosslinked chitosan, PNVCL, chitosan-poly (*N*-vinylcaprolactam) semi IPN particles, and MTRSI-NGs. The bands in the FTIR spectrum of chitosan can be seen as follows: The amine peak at 1632 in crosslinked chitosan and 1637 in PNVCL were shifted to 1652 cm−1 in (PNVCL-CHI)-semi IPN due to the interaction of chitosan with PNVCL. These peaks were attributed to amide I band and carboxylic groups which overlapped with each other (Owens et al. [2007](#page-9-17)). The semi IPN sample two new peaks appeared. The depth of 1082 was assigned to C–O–C observed in PNVCL and PNVCL-chi semi-IPN samples. Another new peak at 1383 corresponding to  $CH<sub>2</sub>$  and OH bending vibrations were due to an interaction of  $CH<sub>2</sub>$ group presence in PNVCL and OH group in chitosan (Bouillot et al. [2000](#page-9-18); Sarkar et al. [2005](#page-9-19)). The stretching vibration bands at 3443 cm−1for chitosan chains and 3441 in PNVCL spectra are shifted to 3470 cm−1thatcorrespond to functional groups of the semi IPN (mainly N–H, O–H). Thus, the results suggested the presence of poly(*N*-vinylcaprolactam) between crosslinked chitosan chains (Patel et al. [2010\)](#page-9-20).

The shift of characteristic peak can explain by two mechanisms:

(1) TPP interaction with the protonated amide and/or amine from the residual chitosan in the semi IPN polymer (Gislén et al [2003](#page-9-21)),

(2) Interaction of chitosan via PNVCL chain can shift the original position of chitosan and PNVCL.

FTIR spectra of MTRSI-NGs exhibited peaks as can be seen in chitosan and semi-IPN nanogels around 1632 and 1589 cm−1 that were attributed to amine and N–H bending vibration of chitosan chains, respectively. The broad bands around 2930, 3440, 1455, and 1054 cm−1 were related to the presence of PNVCL chains in MTRSI-NGs. In MTRSI-NGs, a new sharp peak at 560 cm−1 appeared that's related to Fe–O groups (Karimzadeh et al. [2017](#page-9-22); Zhao et al. [2009\)](#page-10-6). The FTIR spectra demonstrated that chitosan/PNVCL chains were coated on  $Fe<sub>3</sub>O<sub>4</sub>$  nanoparticles. The negatively surface charged iron oxide has an affinity toward chitosan positively charged chains that could



coat the magnetic nanoparticles by the electrostatic selfassembly oppositely charged and the magnetic semi-IPN nanogels were formed successfully.

### **Gel permeation chromatography**

In order to investigate the molecular weight of poly(N-vinylcaprolactam) GPC was utilized. The reported GPC data of PNVCL were limited while it has been extensively studied. Molecular weight data from GPC studies illustrate that Mw and Mn were about 3.5 and 2.7 kDa, respectively. The polydispersity index (PDI) was 1.29 calculated from the ratio of Mw/Mn. Although the free radical polymerization has a high conversion rate, the PNVCL conversion rate is low due to NVCL strict hindrance. The phase transition temperature of thermo-responsive polymers is dependent on molecular weight and molecular weight distribution of PNVCL (Lee et al. [2013](#page-9-23)).

## **LCST analysis of magnetic nanogels**

The phase transition temperature was determined by a UV spectrophotometer-equipped heating system. To determine cloud point temperature, PNVCL and MTRSI-NGs were dispersed in certain concentrations in deionized water. The phase transition temperature for PNVCL was determined about 32 °C while MTRSI-NGs were about 37 °C. In lower temperatures, the solution or suspension is clear while increasing temperature causes clouds to increase in suspension and solution appearance (Namdeo et al. [2008](#page-9-24)). The increase in phase transition temperature was attributed to changes in the hydrophilic-hydrophobic balance of polymer chains. To confrm turbidimetry results, the rheological analysis in the temperature sweep mode was utilized. The rheological behavior studies are one of the ways to assess the temperature response behavior of these polymers, which is carried out in a wide range of temperatures. Rheological behavior can help explain the LCST processes



<span id="page-5-0"></span>**Fig. 4** Rheological analysis of MTRSI-NPs. The viscosity evolves over time during heating to a fnal temperature





<span id="page-5-1"></span>**Fig. 5** Rheological analysis of MTRSI-NPs. Variation storage and loss modulus against temperature

in nanoparticles with their performance in response to shear rates applied by changing temperatures. Figures [4](#page-5-0) and [5](#page-5-1) show the results of rheological test in a temperature state. Figure [4](#page-5-0) exhibited chitosan nanoparticles' rheological behavior that showed the decrease in complex viscosity, storage, and loss modulus by increasing temperature, while it demonstrated an increase in complex viscosity, storage modulus, and loss modulus for magnetic semi IPN nanogels. MTRSI-NGs curve demonstrates that when temperature rises to higher than LCST, the value of viscosity and modulus curve rises (Huang et al. [2017](#page-9-25); Smith et al. [2021](#page-9-4)). As shown in Figs. [4](#page-5-0) and [5,](#page-5-1) a signifcant increase in complex viscosity, storage modules, and the loss modulus are observed by increasing temperature. The modulus of elasticity shows the amount of energy storage and its recovery in each cycle of behavioral change. Therefore, the modulus of elasticity is low at temperatures below the LCST temperature and increases with increasing temperature up to the temperature of LCST. Rheometry results confrmed FTIR results for preparation thermo-responsive nanogels by exhibition increasing in modulus and complex viscosity for magnetic semi-IPN nanogels. On the one hand increasing in complex viscosity and modulus have occurred at 38 °C that is confrmed by turbidimetry results. Phase transition temperature has been evaluated in the human body temperature range that can provide a suitable carrier for target protein delivery. In most polymers, the elastic modulus and complex viscosity decrease by increasing temperature, in contrast to other polymers in temperature-responsive polymers that are attributed to chain conformation. This phenomenon can be explained by the disappearance of hydrogen interactions between thermo-responsive polymer nanoparticles and water molecules that caused conformation changes from a random coil to a globule (Cheng et al. [2013\)](#page-9-26). In the case of suspensions, the phase transition temperature is explained by their collapse. It means when the temperature is below the LCST, the particles are stable and are completely dispersed

due to strong hydrogen bonding between poly(*N*-vinylcaprolactam) chains and environmental aqueous. Increasing temperature in thermo-responsive suspension causes the collapse of the nanogels due to dissipated hydrogen bonding between thermo-responsive nanogels and environmental aqueous change in chain conformation leading to an increase in modulus. As seen in Fig. [5](#page-5-1) thermo-responsive nanogels exhibited higher loss modulus in comparison with storage modulus at lower LCST temperature, while storage modulus was higher than loss modulus at the upper phase transition temperature. The diferences in storage and loss modulus are due to the change in the conformation of thermo-responsive polymer chains at above LCST. In a monophasic state the PNVCL chains conformation is random coil that causes it to display a higher loss modulus, while in the biphasic state, the chains transform to a globule. Globule conformation can provide storage energy more than a random coil. By increasing temperature up to LCST, storage modulus and loss modulus signifcantly increased that illustrated phase transition temperature. Thermo-responsive nanogels behave like a hard-sphere which loss modulus is higher than storage modulus in lower temperature  $(G' \cap G)$ . By increasing temperature up to phase transition temperature, suspension behavior was changed, and storage modulus became almost equal to loss modulus.

#### **Particles size analysis**

Particle size analysis was evaluated in dried and swelled condition. FE-SEM analysis was applied to investigate chitosan and magnetic semi-IPN nanogels in the dehydrated state. Figure [6](#page-7-0) exhibits FE-SEM analysis of dehydrated pure chitosan and semi-IPN magnetic nanogels. By changing different parameters in preparation methods such as stirring speed, reaction temperature, sample pH can control nanogels size. FE-SEM images indicated that dehydrated chitosan nanogels size in the range of 30–40 nm was non-uniform. The size of MTRSI particles was about 20 nm. The reason for decreasing particle size in MTRSI particles is attributed to the presence of magnetic nanoparticles. The presence of magnetic nanoparticles caused to increase ionization degree that leads to reduced particle size. The ionization degree that depends on pH value is the most critical parameter that determines particle size during hybrid nanogels forming (Sahu et al. [2011\)](#page-9-27). MTRSI nanogels morphology is more spherical than chitosan. One reason for controlling the shape and structure of nano gels is the presence of chain caprolactam in the MTRSI nanogels, which prevents nanoparticle deformation and strengthens the nanoparticles in the dehydrated state. Preserving the spherical form of nanoparticles in pharmacological systems is the critical parameter (Sadighian et al. [2015\)](#page-9-28).

To investigate the size of nanogels in a swelling state, dynamic light scattering was utilized. DLS exhibited particles size around 30 nm and almost mono dispersed which was appropriate for local drug delivery. The present results can be obtained by the presence of acetic acid in preparation MTRSI-NGs that causes high dispersion of magnetic nanoparticles and decrease particles size while it was formed. The most important factor for decreasing particle size is attributed to the presence of PNVCL chains in chitosan nanoparticle cavities, which reduced free volume. Highspeed mixing is another substantial parameter caused by polymer and iron oxide dispersed well and its formation in the smaller size. The diference between particle size of FE-SEM and DLS measurement can be related to measurement conditions. The DLS evaluated the hydrodynamic particles size while FE-SEM was analyzed in a dried mood (Elzoghby et al. [2012](#page-9-29)).(see Fig. [7](#page-8-0)).

#### **Drug release behavior**

MTRSI-NGs loaded bovine serum albumin drug release behavior was evaluated under diferent temperatures at pH. The BSA was used as a type of protein for studying the controlled release of proteins from nanogels which were widely used (Tong et al. [1996](#page-9-30)). The various physical and chemical parameters afect drug release behavior such as temperature, pH, the molecular weight of polymer and drug, preparation condition of nanogels, drug concentration.

To investigate, the efect of temperature on drug release behavior two diferent temperatures were selected. The drug release test was carried out at  $25 \pm 1$  and  $40 \pm 1$  °C and in constant pH at 7.4. As shown in Fig. [8](#page-8-1) bovine serum albumin release ratio at 40 °C was faster in comparison with 25 °C which indicates the importance of phase transition temperature. The BSA release behavior at 40 °C suggests that more than 70% of the protein in MTRSI-NPs sample was released from the nanoparticles in 3 first days when the drug release rate at 25 °C was slower than 40 °C. The higher release ratio was at the frst three days of the test, which was attributed to BSA release from the vicinity of the surface of nanoparticles. By comparing drug release at 25 and  $40^{\circ}$ C, the LCST effect is understandable. The release behavior at a higher temperature of LCST is faster than at the lower temperature of LCST. The main reason for describing this behavior changes in hydrophilic properties at the higher temperature of LCST which caused a change in nature of the hydrophilic to hydrophobic (Ta and Pulat et al. [2012\)](#page-9-31).

Drug release kinetics evaluated in pH 7.4 and 5.5. The release behavior indicates that pH strongly influenced bovine serum albumin release through MTRSI-NGs. As shown in Fig. [9](#page-8-2), more than 50% of BSA was released after 3 days at pH 5.5, while this amount was about 30%at pH 7.4. This behavior is explained by shrinking MTRSI-NGs





<span id="page-7-0"></span>





<span id="page-8-0"></span>**Fig. 7** Dynamic light scattering analysis of MTRSI nanogels



<span id="page-8-1"></span>**Fig. 8** In vitro release profles of BSA from MTRSI-NGs in PBS solution (Temperature 25 and 40 °C)



<span id="page-8-2"></span>**Fig. 9** In vitro release profles of BSA from MTRSI-NGs in PBS solution (pH 5.5, 7.4)

in pH 7.4 that prevents albumin release through the nanogels. A fast release profile for pH 5.5 is attributed to the swelling behavior of nanogels due to the pH-responsive properties of MTRSI-NGs. The BSA release behaviors represented unstable behavior nanogels under different pH values and temperatures. According to MTRSI-NGs results, they only prevented the high ratio release of proteins in pH 7.4 and temperature at 25 °C. The obtained results illustrated that protein-carrying nanoparticles are suitable for use in protein delivery to internal organs and tissues due to the long release time of the protein (Ta and Pulat et al. [2012\)](#page-9-31).

## **Conclusion**

In the present study, magnetic thermo-responsive semi-IPN nanogels are optimized for the local protein delivery systems. At frst, PNVCL was synthesized through free radical polymerization and the MTRSI-NGs were prepared. FTIR spectra demonstrate that magnetic semi-IPN nanogels were formed successfully. Gel permeation chromatography exhibited a low molecular weight of PNVCL synthesis. Thermoresponsive properties of MTRSI-NGs were analyzed by turbidimetry and rheometry. Turbidimetry exhibits cloud point around 37 °C. The results of turbidimetry were confrmed by rheological analysis in temperature sweep mood. Phase transition results demonstrated that MTRSI-NGs are controlled near human body temperature. To study particle size in the dried and swelling mood, FE-SEM and dynamic light scattering were utilized. FE-SEM analysis displayed particle sizes less than 20 nm while pure chitosan was between 30 and 40 nm due to high ionization properties of magnetic nanoparticles present in MTRSI-NGs structure. Dynamic light scattering analysis showed nanogels size about 30 nm in swelling mood. The determination of protein release treatment of MTRSI-NGs showed a higher ratio of drug release at 40 in comparison with 25 °C that is attributed to the switching hydrophobicity properties of MTRSI-NGs after phase transition temperature. Furthermore, drug release behavior of MTRSI-NGs is given at diferent pH values. The release behavior at pH 5.5 showed a higher ratio in comparison with 7.4 that indicates the pH-sensitive properties of MTRSI-NGs. The present study suggests that chitosan leads to modifying the LCST behavior of PNVCL to the near-human body. These results indicate that MTRSI-NGs are appropriate for the target protein delivery system.

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#### **Declarations**

**Conflict of interest** The authors declare no confict of interest.

**Ethical approval** This article does not contain any studies with human participants or animals performed by any of the authors.



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