



Mono vs. combo regimens with novel beta-lactam/beta-lactamase inhibitor combinations for the treatment of infections due to carbapenemase-producing *Enterobacteriales*: insights from the literature

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Received: 22 November 2020 / Accepted: 13 January 2021 / Published online: 3 February 2021
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Abstract

Ceftazidime–avibactam (CZA), meropenem–vaborbactam (MVB) and imipenem–relebactam (I–R) are combinations of old β -lactams with novel non- β -lactam β -lactamase inhibitors (BLBLIs) able to inhibit some carbapenemases, such as the KPC-type, thus are becoming the standard for difficult-to-treat carbapenemase-producing *Enterobacteriales* (CPE); a practical question is whether these novel BLBLIs should be used as monotherapy or as part of a combination regimen with other antibiotics, and if so, with which ones, to reduce the emergence of resistant strains and to optimize their efficacy. In this short review, we assessed clinical outcomes in patients with CPE-infections treated with the novel BLBLIs as mono- or combo-regimens, and laboratory studies on the synergistic effects with other antimicrobials. Available evidence on combination therapy is scarce and mainly limited to retrospective studies involving 630 patients treated with CZA: aminoglycosides were used in 39.6% of 336 patients treated with combo-regimens, followed by polymyxin B/colistin (24.4%), tigecycline (24.1%), carbapenems (13.4%) and fosfomycin (5.4%). Aminoglycosides could be useful in case of bloodstream and severe urinary infections. Pneumonia is a risk factor for CZA-resistance emergence: fosfomycin, due to favorable lung pharmacokinetics/pharmacodynamics, could represent an interesting partner; fosfomycin could be added also for osteomyelitis. Tigecycline could be preferred for intrabdominal and skin-soft tissue infections. Due to nephrotoxicity and lack of in vitro synergy, the association CZA/colistin seems not optimal. MVB and I–R were mostly used as monotherapies. Currently, there is no definitive evidence whether combinations are more effective than monotherapies; further studies are warranted, and to date only personal opinions can be provided.

Keywords Carbapenemases · *Enterobacteriaceae* · *Enterobacteriales* · Avibactam · Vaborbactam · Relebactam · Combination · Monotherapy

Introduction

Ceftazidime–avibactam (CZA, CAZ–AVI), meropenem–vaborbactam (MVB, MEM–VAB) and imipenem/cilastatin–relebactam (I–R, IPM–REL) represent recent or

upcoming combinations of old β -lactams with novel synthetic non- β -lactam β -lactamase inhibitors (BLBLIs) able to inhibit extended-spectrum β -lactamases (ESBLs), AmpCs and some carbapenemases, such as the *Klebsiella pneumoniae* carbapenemase (KPC). AVI displays activity also towards some class D β -lactamases (such as OXA-48); VAB also restored the activity of meropenem against class A and class C β -lactamase-producing *K. pneumoniae* strains with reduced permeability due to porin mutations [1–3]. Therefore, these novel BLBLIs represent an interesting option for managing severe infections due to carbapenem-resistant (CR) carbapenemase-producing *Enterobacteriales* (CPE), although they are not active on strains producing class B metallo- β -lactamases (MBLs), such as the New-Delhi

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metallo- β -lactamase (NDM). Zidebactam and nacubactam are ongoing enhancer derivatives of the diazabicyclooctane scaffold with promising in vitro activity against MBL-producing *Enterobacterales*; taniborbactam is a boronic-acid-containing β -lactamase inhibitor able to inhibit β -lactamases belonging to class A, B, C and D [3].

KPC-producing *Klebsiella pneumoniae* (KPC-Kp) is nowadays endemic in several countries, is mainly driven by nosocomial spread [4], and is associated with a mortality of about 40% [5], but even over 50% [6] for bloodstream infections (BSIs) when treated with the best available therapy (BAT) prior these new options. The optimal treatment of infections due to KPC-Kp is not known, nor whether combination therapy would be superior to monotherapy; lacking the results of well-conducted randomized clinical trials (RCTs), the therapeutic choice is mainly based on clinical experience and expert opinions. For CPE other than KPC-Kp, such as NDM-producers that recently resulted in sustained transmission in the North-West area of Tuscany, Italy [7], data on the most appropriate treatment is even more scarce.

BAT against KPC-producers classically consisted of various combinations of antibiotics displaying in vitro activity against the isolate (i.e., aminoglycosides, colistin, fosfomicin and tigecycline), often in associations with high-dose meropenem (especially for MIC \leq 8 mg/L) [8]; in the more severely ill patients the combination therapy has been shown to be associated with a lower mortality compared to monotherapy [5]. Other agents, such as cefiderocol, plazomicin and eravacycline, represent novel interesting not-BLBLI options for CPE treatment [9, 10].

Tumbarello et al. [6] showed that in patients with BSIs due to KPC-Kp, a salvage therapy with CZA was associated with a 30-day mortality significantly lower than that of a matched cohort treated with second-line regimens containing other antibiotics (36.5% vs 55.8%, $p=0.005$), suggesting a survival benefit relatively to the commonly used regimens.

Despite the unsatisfactory outcomes with the traditional BAT, and the obvious expectations placed on these novel carbapenemase-inhibitors, there is currently scarce evidence of clinical use of the novel BLBLIs for real-life management of the difficult-to-treat infections caused by CPE. Moreover, these drugs were often administered only as salvage therapy on a compassionate-use basis, because most phase 2 and 3 trials conducted to support their marketing authorization were conducted mainly in complicated intra-abdominal and urinary infections and compared with carbapenems so that the patients whose infections were caused by carbapenem-resistant strains were excluded from enrollment.

Many doubts are so nowadays unanswered by current literature. A practical question is whether these novel BLBLIs should be used, for the real-life treatment of CPE infections, as monotherapy or as part of a combination regimen with

other antibiotics. And if so, with which ones? A combination therapy would have, in fact, the advantage of exploiting the bactericidal synergy between different drugs, as well as limiting the risk of selection of resistant mutant strains with a single antimicrobial, but these remain only hypotheses needing proof.

The aim of this short review is to try to address this knowledge gap, extracting from the available literature data regarding the clinical outcomes, mainly the mortality and clinical success (as defined in the different studies), for patients with infections due to carbapenem-resistant *Enterobacterales* (CRE) treated with the novel BLBLIs as mono- or combo-regimens. In addition, we searched in vitro studies investigating the effects of the mono- and combo-regimens.

Methods

A comprehensive computerized search was performed using PubMed, through November 15, 2020, involving both Medical Subject Headings (MeSH) terminology and relevant keywords for search strings. The following terms were searched in combination: *Enterobacteriaceae*, *Enterobacterales*, carbapenemase, avibactam, relebactam, vaborbactam, combination, therapy, synergism/synergy, resistance, carbapenem-resistant. References of retrieved articles were manually searched to ensure the identification of studies not found in the initial literature search. The selection was limited to peer-reviewed publications written in English. After de-duplication, all authors independently screened titles and abstracts, and finally full texts, to identify all potentially relevant studies, resolving discrepancies through discussion and consultation between them. Moreover, a search was performed on <https://clinicaltrials.gov> for the studies on the new BLBLIs for CRE infections, in progress or waiting to begin or completed but not yet published. Authors of the included studies were contacted via e-mail if further study details were needed.

Results

We retrieved 11 observational retrospective [6, 11–20] and 2 prospective studies [21, 22] for a total of 630 patients treated with CZA (294 as monotherapy and 336 as combination therapy). Two of these studies specifically addressed the treatment of infections due to OXA-48 producing *Enterobacterales*, for a total of 81 patients treated with CZA as mono- or combo-therapy [18, 19]. We retrieved also a meta-analysis enrolling 11 retrospective studies comparing the clinical effects of CZA as monotherapy or combo against infections due to CRE and/or CR-*Pseudomonas aeruginosa*

[23]: eight [6, 13–19] of the 11 studies included in the meta-analysis have been included in the present review.

As for MVB, we retrieved one RCT (TANGO II) with a total of 32 patients with CRE infections all treated with monotherapy [24] and a retrospective multicenter study enrolling 26 patients most of which (84.6%) treated with monotherapy [20].

We retrieved one RCT (RESTORE-IMI 1) [25] on imipenem non-susceptible bacterial infections, with a total of 21 patients treated with I–R as monotherapy compared with 10 patients treated with imipenem plus colistin: only 5 patients were treated with I–R for infections due to CRE, being the remaining CR-*Pseudomonas aeruginosa* infections, which are not the topic of this review.

Regarding the studies currently in progress (consultable on <https://clinicaltrials.gov>), we retrieved a phase 3 prospective, randomized, multicenter, open-label, parallel group trial (NCT03580044) designed to evaluate the association of aztreonam-avibactam as monotherapy for MBL-producing Gram-negative bacteria, but to date is not yet recruiting patients.

Most available evidence on the novel BLBLIs as mono or combo regimens for the treatment of infections due to CR-CPE are thus limited to CZA, and relies on relatively small sample size cohorts and on observational retrospective studies.

Table 1 shows the main characteristics of the clinical studies assessing the effectiveness of the novel BLBLIs as mono- or combo-regimens for CRE infections.

Regimens containing CZA were usually classified as combination therapy if they included at least one other antimicrobial displaying activity against the isolate, but also carbapenems were administered in several cases: carbapenems were associated in 45/336 cases (13.4%), but in the study of van Duin et al. [21] they were used in 46% of cases. Aminoglycosides were overall used in 133/336 patients (39.6%), representing the main class usually associated with CZA, followed by polymyxin B or colistin (82/336, 24.4%), tigecycline (81/336, 24.1%), and fosfomycin (18/336, 5.4%).

A non-significant trend of improvement of clinical success was reported for CZA combination therapy only in 3 studies [11, 12, 15], while in 6 studies this outcome was (albeit not significantly) better in the monotherapy group [13, 16–20]

Shields et al. [12] reported a higher mortality for CZA as monotherapy when compared with combination regimen with gentamicin (12.5% vs 0%); also Tumbarello et al. [6] and Guimaraes et al. [22] reported higher mortality rate for patients treated with CZA as monotherapy, while other authors [13, 16, 18–20] found higher mortality in combo group.

Tumbarello et al. [6] showed that, among patients with KPC-Kp BSIs, CZA was significantly more effective than

alternative regimens in reducing 30-day mortality both as mono- or combination therapy. The mortality was 40.9% in the monotherapy group and 35.4% in the combination group: in the combo group, the mortality was 50% in association with amikacin, 38.5% with colistin, 37.5% with tigecycline, 36.9% with carbapenems, 32% with gentamicin, 28.6% with fosfomycin.

Guimaraes et al. [22], in case of severe infections caused by KPC-producing *Enterobacteriales* co-resistant to carbapenems and polymyxins and treated with CZA as salvage treatment, reported a trend (although not significant) for a lower 14-day mortality for combo- than mono-therapy (21.4% vs 40%).

In the meta-analysis of Onorato et al. [23], among 377 patients with infections due to CRE, the mortality rate was 38.7% (74/191) for combo and 31.2% (58/186) for monotherapy (RR = 1.20, 95% CI 0.89–1.61; $p=0.229$). Similarly, no difference was found analyzing the rate of microbiological cure reported in 6 studies (63.3% for combo- vs. 61.9% for mono-therapy; RR = 1.05, 95% CI 0.84–1.31, $p=0.653$). All studies except one reported data on the emergence of resistance to CZA, with a total of 8 patients (4.1%) in the mono and 6 patients (3.0%) in the combo group.

Combination therapy of MVB with other antibiotics has been evaluated by Ackley et al. [20], but data is limited to only 3 patients co-treated with polymyxins and 1 with tigecycline.

Table 2 shows some in vitro studies assessing the synergistic effects of the novel BLBLIs in combination with different antimicrobials on CRE. As clinical studies, most data refer to CZA.

Discussion

Currently, there is not enough high-level clinical evidence to make definitive conclusions whether the novel BLBLIs used as combo regimens are more effective than monotherapy for CPE infections; in some studies, the mortality rate among patients treated with combination therapy was even higher than in monotherapy group [13, 16, 18–20], but in retrospective studies an indication bias towards more severely ill patients could be the explanation for the lack of benefit of combo regimens observed.

Most of the studies we retrieved in the literature were, in fact, observational, a fact which does not allow assessment of efficacy, but only of effectiveness in real life. Definitive evidence from clinical trials of the adequate methodology is, therefore, crucial.

Despite the optimal combination regimen remains unknown, clinical and laboratory studies suggest some considerations; nevertheless, at the present time, only

Table 1 Clinical studies on the novel BLBLIs administered as mono- or combo-regimens for infections due to CRE

First author, year [ref.]	Study design (adjustment for confounding)	BLBLI	Source of CRE infection (number of patients)	CRE (number of isolates)	Carbapenemase type (% CRE isolates expressing)	Number of pts treated with mono vs combo
Shields, 2016 [11]	RSC (None)	CZA	Lung (12), BSI (10), IAI (4), SSTI (4), UTI (4), Other (3)	Kp (31), Ec (3), Ecl (2), Ka (1)	KPC (78%), OXA-1 (10.8%)	26 vs 11
Shields, 2017 [12]	RSC (MLR)	CZA	BSI (3 primary; 2 secondary to IAI, 3 to pneumonia, 5 to UTI)	Kp (13)	KPC (100%)	8 vs 5
Krapp, 2017 [13]	RSC (None)	CZA	UTI (5), Lung (1), BSI (1)	Kp (6)	KPC (100%), OXA-9 (100%)	2 vs 4
Caston, 2017 [14]	RMC (None)	CZA	BSI (2 primary; 1 secondary to CVC, 1 to IAI, 2 to pneumonia, 1 to SSTI, 1 to other)	Kp (6), Ko (1), Ec (1)	KPC (37.5%), OXA-48 (62.5%)	0 vs 8
Temkin, 2017 [15] ^a	RMC (None)	CZA	BSI (26), IAI (15), Lung (7), SSTI (4), UTI (3), Bone (3), Endocarditis (2), Other (3)	Kp (34), Ko (1), Ec (1)	KPC (63.9%), OXA-48 (36.1%)	13 vs 25
King, 2017 [16]	RMC (None)	CZA	BSI (23), UTI (17), Lung (16), Wound (8), IAI (4), Bone/ Joint (2)	Kp (50), Ec (5), Ecl (4), Ko (1), Sm (1), Ps (1)	NR	33 vs 27
Shields, 2018 [17]	RSC (MLR)	CZA	Lung (33), Primary BSI (20), UTI (8), IAI (7) SSTI (6), Other (3)	Kp (60), Ec (9), Ecl (5), Ka (1), Ko (1), Sm (1)	KPC (75%), OXA not-48 (23%)	53 vs 24
Sousa, 2018 [18]	RSC (MLR)	CZA	IAI (16), Lung (15), UTI (14), BSI (26), SSTI (3)	Kp (54), Ec (2), Ecl (1)	OXA-48 (100%)	46 vs 11
De la Calle, 2019 [19]	RMC (None)	CZA	IAI (7), UTI (6), Lung (5), SSTI (4), Other (2)	Kp (23), Ec (1)	OXA-48 (100%)	14 vs 10
van Duijn, 2018 [21]	PMC (IPTW)	CZA	BSI (15), Lung (9), UTI (6), SSTI (6), Other (2)	Kp (37), Ecl (1)	KPC (96.3%)	14 vs 24
Guimaraes, 2019 [22]	PMC (None)	CZA	BSI (12), UTI (8), IAI (4), Lung (3), SSTI (2)	Kp (28), Sm (2)	KPC (100%)	15 vs 14
Tumbarello, 2019 [6]	RMC (MLR)	CZA	BSI (104), Lung (13), IAI (12), UTI (6), Other (3)	Kp (138)	KPC (100%)	29 vs 109
Wunderink, 2018 [24]	MC, open-label RCT (Randomization)	MVB	BSI (14), UTI (12), Lung (4), IAI (2)	Kp (29), Ec (3), Ecl (1), Sm (1)	KPC (90.6%), NDM or OXA-48 (9.4%)	32 vs 0
Ackley, 2020 [20]	RMC (None)	CZA	BSI (44), UTI (13), Lung (37), SSTI (20), IAI (18), Other (5)	<i>Klebsiella</i> spp (76), <i>Enterobacter</i> spp. (20), Ec (9), <i>Citrobacter</i> spp (2)	KPC (71.9%) Data available only for 32 isolates	41 vs 64
Motsch, 2019 [25] ^c	MC double-blind RCT (Randomization)	MVB	BSI (9), UTI (1), Lung (12), SSTI (3), IAI (8), Other (1)	<i>Klebsiella</i> spp (15), <i>Enterobacter</i> spp. (8), Ec (3), <i>Citrobacter</i> spp (2), Sm (1)	KPC (76.9%) Data available only for 13 isolates	22 vs 4
NCT03580044	MC open-label RCT, not yet recruiting (Randomization)	I-R ATM-AVI NR	IAI (1), UTI (4) NR NR	Kp (3), Ecl (1), <i>Citrobacter freundii</i> (1) NR NR	KPC (80%) NR NR	5 vs 0 NR (all mono)

Table 1 (continued)

First author, year [ref.]	Additional antibiotic in combo group	Overall 30-day mortality (mono vs combo)	Overall clinical success (mono vs combo)
Shields, 2016 [11]	GEN (8), CST (2), TGC (1)	24% (NR)	59% (58% vs 64%, ns)
Shields, 2017 [12]	GEN (5)	8% (12.5% vs 0%)	85% (75% vs 100%)
Krapp, 2017 [13]	TGC (2), PMB (2)	50% (0% vs 75%)	83.3% (100% vs 75%) as initial clinical cure
Caston, 2017 [14]	AG (7), CB (3), FOF (2), TGC (2), CST (2)	25% (NA)	85.7% (NA)
Temkin, 2017 [15] ^a	TGC (11), AMK (9), FOF (4), Other	39.5% (NR)	73.7% (69.2% vs 76.0%, ns)
King, 2017 [16]	AG (11), PMB/CST (7), TGC (6), CB (3)	32% (30% vs 33%, ns)	65% (67% vs 63%, ns)
Shields, 2018 [17]	GEN (19), CST (2), AMK (1), FQ (1), TGC (1)	19% (NR)	55% (56% vs 50%, ns)
Sousa, 2018 [18]	CST (7), TGC (2), AMK (1), CB (1)	23% (22% vs 27%, ns)	77% (80% vs 64%, ns)
De la Calle, 2019 [19]	AMK (1), CST (1), TGC (3), two out of AMK/ CST/TGC (5)	8.3% (14.3% vs 30%, ns: 90-day mortality)	62.5% (64.3% vs. 60%, ns)
van Duijn, 2018 [21]	GEN (12), TGC (12), AMK (6), SXT (4), CB (11), FOF (1)	9% (NR)	NR
Guimaraes, 2019 [22]	PMB (9), TGC (7), MEM (5), AMK (3), GEN (1), FOF (1)	51.7% (40% vs 21.4%, ns: 14-day mortality)	82.7% (NR)
Tumbarello, 2019 [6]	GEN (34), CB (21), CST (22), TGC (16), FOF (10), Other (6)	34.1% (40.9% vs 35.4% ^b)	NR
Wunderink, 2018 [24]	NA	15.6% (NA)	65.6% (NA)
Ackley, 2020 [20]	CST (20), TGC (18), AG (15), PMB (10), FQ (8), STX (4), CB (1)	19.1% (22.0% vs 31.2%, ns: 90-day mortality)	61.9% (63.4% vs 60.9%, ns)
Motsch, 2019 [25] ^c	CST (2), PMB (1), TGC (1)	11.5% (NR)	69.2% (NR)
NCT03580044	NA	20% (NA)	40% (NA)
	NR	NR	NR

MC multicenter, PMC Prospective multicenter, PSC Prospective single centre, RMC Retrospective multicenter, RSC Retrospective single centre, RCT randomized controlled trial, MLR Multivariate logistic regression, IPTW inverse probability of treatment weighting, BSI bloodstream infection, IAI intra-abdominal infection, SSTI skin/soft tissue infection (wound included), UTI urinary tract infection, *Ec Escherichia coli*, *Ecl Enterobacter cloacae*, *Ka Klebsiella aerogenes*, *Ko Klebsiella oxytoca*, *Kp Klebsiella pneumoniae*, *Ps Providencia stuartii*, *Sm Serratia marcescens*, AG aminoglycosides, AMK amikacin, ATM aztreonam, AVI avibactam, CB carbapenems, CST colistin, CZA ceftazidime-avibactam, GEN gentamicin, FOF fosfomycin, FQ fluoroquinolones, I-R imipenem/relebactam, MEM meropenem, MVB meropenem-vaborbactam, PMB polymyxin B, SXT trimethoprim-sulfamethoxazole, TGC tigecycline, NR not reported, NA not applicable, ns not significant

^aIn this case series most data (aggregated) relate to 36 patients infected with CRE, but two cases are carbapenem-resistant *Pseudomonas aeruginosa*

^bData only for the BSI group

^cIn this study on imipenem-non-susceptible Gram-negative bacteria, CRE infections were only a minority, for a total of only 5 pts treated with I-R as monotherapy, the remaining data regarding 16 infections due to *Pseudomonas aeruginosa*

Table 2 In vitro studies on effects of the novel BLBLs in combination with different antimicrobials for CRE

First author, year [ref.]	Combination	Method	CRE species: number of strains	Effect of combination: % of total strains	Comments
Gaibani, 2017 [26]	CZA plus: GEN, CIP, IPM, MEM, ETP, TGC	Gradient diffusion test	KPC-Kp: 13	Indifference: 100% for GEN and CIP Synergy: 8% for TGC, 100% for IMP and MEM CZA restored meropenem and imipenem susceptibility in 50% and 80%, respectively, of KPC-3-producing strains, but never in case of KPC-2-producers	Combinations of CZA with IPM or MEM displayed high synergistic activity against KPC-Kp, including CZA-susceptible and resistant strains. For ETP, a synergistic effect was observed only against CZA-susceptible strains
Shields, 2018 [27]	CZA plus CST	Time-kill	KPC-Kp: 15; Other: 9	Synergy: 13% Antagonism: 46%	CST does not potentiate the in vitro killing of most CRE by CZA, nor suppress CZA resistance
Chew, 2018 [28]	ATM plus AVI	BMD	Dual-carbapenemase NDMs-KPC/OXA-48 producers. (<i>Escherichia coli</i> : 13, Kp: 44, <i>Citrobacter freundii</i> : 7, <i>Enterobacter cloacae</i> : 6)	AZT susceptibility restoration: 98.6%	Where MBLs and CPE predominate, ATM-AVI rather than CZA should be more therapeutically effective
Biagi, 2019 [29]	ATM plus AVI or VBR	Time-kill	<i>Escherichia coli</i> : 4 Kp: 4 All coproducing NDM and at least one serine β -lactamase	ATM/CZA was synergistic against all 7 aztreonam-resistant strains. ATM/MEM-VBR was synergistic against all strains (with the exception of an OXA-232-producing Kp)	ATM plus MVB has similar activity to ATM plus CZA against <i>Enterobacteriaceae</i> producing NDM and other non-OXA-48-like serine β -lactamases
Jayol, 2018 [30]	CZA plus ATM	MIC test strips-based synergy	CST-resistant carbapenemase-producing Kp -KPC-like: 11 -OXA-48: 32 -OXA-181: 5 -NDM-like: 8 -Dual-carbapenemase.(NDM-OXA): 7	Synergy: 100% for MBL-producers	The synergy of CZA-ATM combination against NDM producers could be explained by the neutralization of the ESBL activity by AVI restoring the susceptibility to ATM
Nath, 2018 [31]	CZA plus: AMK, MEM	Time-kill	KPC-Kp: 4 -2 resistant to PMB, AMK and MEM (with CZA MIC 8/4 mg/L) -2 PMB susceptible	-Synergy for CZA plus AMK or MEM -At 1/4 MIC, synergy for CZA plus MEM	Combination of CZA with another agent (such as MEM) should be considered, particularly for isolates with CZA MICs near the susceptibility breakpoint (8/4 mg/L)

Table 2 (continued)

First author, year [ref.]	Combination	Method	CRE species: number of strains	Effect of combination: % of total strains	Comments
Gottig, 2019 [32]	CZA plus IPM	Time-kill	KPC-Kp	Bactericidal activity Prevention of resistance development	Combination of CZA-IMP seems to be an effective strategy to prevent development of CZA resistance both in vitro and in vivo (better survival in <i>Galleria mellonella</i> infection model)
Ojdana, 2019 [33]	CZA plus: ETP, FOF, TGC	E-test MIC:MIC ratio synergy	Carbapenemase-producing Kp:19 -KPC-Kp:8 -NDM-Kp:10 -OXA-48-Kp:1	Synergy for CZA with ETP and FOF: 47% Synergy/Indifference for CZA-TGC: 5%/95% -Synergy for CZA-ETP: 100% -Synergy for CZA-FOF: 37.5% -Indifference for CZA-FOF: 12.5% -Synergy for CZA-FOF: 50% -Synergy for all combination but TGC	Combination of CZA with ETP or TGC reduced MIC to less than the SBP in all KPC- and OXA-48-Kp. However, CZA-FOF reduced MIC to less than the SBP among all tested carbapenemase-producers
Mikhail, 2019 [34]	CZA plus: AMK, ATM, CST, FOF, MEM	MIC combination and time-kill	Carbapenem-resistant Kp: 21	Synergy for CZA plus AMK or ATM or FOF	CZA in combination with these antibiotics has potential for therapeutic options in difficult to treat pathogens
Borjian, 2020 [35]	CZA plus: polymyxin B	Tandem in vitro time-kill/ in vivo <i>Galleria mellonella</i> survival model assay	KPC-3-producing Kp: 3	Combinations at 1/4x or 1/2xMIC were not bactericidal or synergistic against any of the 3 isolates Combining polymyxin B with CZA at 4xMIC did not improve survival compared to CZA alone	No improvement in in vitro bactericidal activity or in vivo efficacy when polymyxin B is combined with CZA against KPC-Kp

BMD broth microdilution method, MIC minimum inhibitory concentration, KPC-Kp KPC-producing *K.pneumoniae*, SBP susceptibility breakpoint, AMK amikacin, ATM aztreonam, AVI avibactam, CIP ciprofloxacin, CST colistin, CZA ceftazidime-avibactam, ETP ertapenem, FOF fosfomicin, IPM imipenem, MEM meropenem, MVB meropenem-vaborbactam, TGC tigecycline, VBR vaborbactam

considerations reflecting personal opinion can be made about this specific topic.

Drug combinations are widely used in clinical practice to prevent the emergence of resistance; however, unfavorable effects on resistant selection cannot be excluded: Liu et al. [36] showed that the rapid evolution of tolerance is a major survival factor for bacteria, and tolerance can promote the evolution of resistance under antimicrobial combinations that were instead expected to prevent resistance.

CZA was used as combo regimen in over 50% of cases with CPE infections, in real-life. Aminoglycosides, such as gentamicin or amikacin (depending by in vitro susceptibility results), are rapidly and durably bactericidal and effective in the treatment of carbapenem-resistant *K. pneumoniae* BSIs, and are especially useful when the sources of infection are amenable to reliable pharmacokinetics, such as BSIs due to urinary sources; intraabdominal sources (especially in presence of biliary obstruction or hepatic damage) and respiratory tract and bone are instead characterized by a very poor penetration [37]. Shields et al. [12] effectively used combination therapy with short-course gentamicin followed by de-escalation to CZA alone as a strategy for maximizing treatment effectiveness while limiting toxicity, mainly represented by acute kidney injury. In the study of Tumbarello et al. [6], 8.7% of patients, mostly with BSIs, experienced KPC-Kp infection relapses after CZA alone treatment was discontinued: in all cases the isolates remained susceptible to CZA, and clinical and/or microbiological cures were achieved after retreatment with CZA plus gentamicin. Nevertheless, using a gradient synergy test, the combination of CZA with gentamicin (the same resulted also for ciprofloxacin) displayed no synergy against any of the KPC-Kp isolates tested in an in vitro study [26]. However, synergistic activities observed in vitro may not exactly correlate with clinical efficacy, for the peculiar in vivo pharmacokinetic properties displayed by the antibiotics.

In effect, a combination regimen could result in a synergistic activity, very useful in the most serious infections and in case of sepsis and septic shock, and it might suppress CZA resistance selection. Pneumonia has been recognized as a risk factor for CZA resistance among patients with CRE infections [17], and Krapp et al. [13] previously showed that patients with pneumonia due to CR-Kp had poorer outcomes (failure or relapse), suggesting potential limitations in the treatment of CR-Kp pneumonia with CZA and highlighting the need for adequate studies about the activity and bioavailability of CZA in the lungs, and therefore the need to assess the optimal dosing (such as increased or loading doses) of CZA, especially in septic patients. In effect, clinical data on optimal dosing are generally lacking for the novel BLBLIs thus, variations in extracellular volume and renal dysfunction often observed in critically ill patients may impact the disposition of both

β -lactams and β -lactamase inhibitors [3]. Fosfomycin, which has excellent pharmacokinetic characteristics in the lung, could represent a very interesting option to guarantee a synergistic action and to intervene in cases where CZA resistance could more likely occur. The synergy of CZA in combination with fosfomycin against carbapenem-resistant *K. pneumoniae* has been shown in recent in vitro studies [33, 34].

Fosfomycin could be useful also for osteomyelitis, due to very high bone penetration of this drug. However, despite the manageability and the efficacy of this option, this association was rarely used in clinical practice, only in 5.4% of total patients we retrieved.

Polymyxin B and E (colistin, CST) have been used in combination with CZA in 24.4% of cases we retrieved, but despite the rationale that membrane permeabilization by colistin could facilitate increased access of CZA to its target sites, in an in vitro study [27] the combination of CZA and CST did not suppress CZA resistance and did not provide a benefit in potentiating the killing of most CRE isolates. Moreover, the study of van Duin et al. [21] showed that patients treated with CZA, compared to those treated with CST, were less likely to die and more likely to be discharged home at 30 days, and an analysis using the desirability of outcome ranking method showed that the IPTW (inverse probability of treatment weighting)-adjusted probability of a better outcome on CZA compared with colistin is 64%. No improvement of in vitro bactericidal activity and in vivo efficacy has been shown when polymyxin B is combined with CZA against KPC-Kp [35]. In addition, in consideration of the known nephrotoxicity of CST, if another option is available, the association of CZA with CST should be, in our opinion, avoided, reserving CST for cases in which it is the only active agent and the CZA monotherapy has the greatest risk of failure, as when the MIC of CZA is very close to the susceptibility breakpoint, or in case of pneumonia, as previously discussed.

In general, a combination therapy for CZA could be particularly useful when the MIC values for CZA are very close to the susceptibility clinical breakpoint (8 mg/L) [38, 39], as the in vitro study of Nath et al. [31] confirmed. However, it should also be noted that in the study of Shields et al. [17] a “monotherapy” did not represent a significant risk factor for CZA resistance among patients with microbiologic failure.

Tigecycline was often used in combination with CZA (24.1%), and could be useful when the infection source is the abdomen, especially the biliary tract, or skin and soft tissues, given its favorable pharmacokinetic in these districts, avoiding its use for pneumonia and BSIs. However, it is important to note that a recent in vitro study using the E-test MIC:MIC ratio synergy method [33] showed that the combination of CZA with tigecycline is synergistic only for 5% of carbapenemase-producing *K. pneumoniae* isolates

tested (this percentage reached 12.5% for KPC-producing strains), being indifferent in the remaining cases.

Carbapenems were sometimes used in combination with CZA (13.4% of cases). The rationale goes through the reduction of MIC of carbapenems provided by avibactam, allowing to recover the favorable pharmacokinetics/pharmacodynamics of these antibiotics. Notably, CZA reduced MIC of meropenem and particularly for imipenem below the resistance breakpoints against KPC-Kp strains tested in an in vitro model using the gradient synergy test, displaying a synergistic activity both in CZA susceptible and resistant isolates, while for ertapenem a synergistic effect was observed only against CZA-susceptible strains [26]. Recently, CZA showed to be very active and synergistic with meropenem against multiresistant *Serratia marcescens* isolates carrying bla_{KPC-2} [40], and a synergy for CZA-ertapenem was found in all KPC-Kp isolates tested by Ojdana et al. [33]. Furthermore, in certain isolates of *K. pneumoniae* the development of resistance-conferring bla_{KPC-3} mutations during CZA exposure is associated with restoration of carbapenem susceptibility [41], thus the carbapenem/CZA combination might mitigate the emergence of bla_{KPC} mutations or, alternatively, treat emerging resistant subpopulations. Notably, even 30% of 10 patients with microbiologic failure observed in the study of Shields et al. [11] developed CZA resistance (MIC > 8 mg/L) following a median of 15 days of treatment, but in 2 of these 3 patients the mutations so reduced the meropenem MIC to restore the full susceptibility and allowing an effective salvage treatment with the carbapenem therapy [41].

More recently, Gottig et al. [32], based on time-kill kinetics as well as an in vivo infection model, confirmed that the combination of CZA with IMP can represent an evaluable bactericidal strategy able to prevent the in vivo development of CZA resistance in KPC-producing *Enterobacterales*.

Unfortunately, the limit of combining CZA with a carbapenem is thwarting the carbapenem-sparing stewardship policies.

Awaiting the future marketing of aztreonam-avibactam association, the combination of the novel BLBLIs (CZA, MVB, I-R) with aztreonam (ATM) retains currently great importance in overcoming the resistance conferred by MBLs [42], taking advantage of the stability of aztreonam to hydrolytic activity of the MBLs and of the ability of the new inhibitors to block the activity of the β -lactamases frequently co-expressed by the MBL-producing *Enterobacterales*, both of Ambler class A (such as the ESBLs or the KPCs), class C (AmpCs) and class D [1, 28, 30, 42]. For the combinations involving a double β -lactam strategy (such as ATM plus ceftazidime or carbapenems), a possible benefit could arise by the simultaneous inhibition of multiple penicillin-binding proteins [42].

The combination with ATM seems particularly useful in geographical regions where MBL-producing *Enterobacterales* (e.g. NDM-producers) predominate, such as the Asian continent [43] and Tuscany, Italy [7]; if a molecular antibiogram directly detecting the resistance mechanisms at the molecular (mainly genic) level cannot be performed, and the only tool available to clinicians is represented by the conventional antibiogram based on phenotypic detection of bacterial growth, especially when the novel BLBLIs (such as CZA and MVB) are not tested or turn out in a resistance, it is very difficult to understand if the CRE-isolate is expressing class A (such as KPC) and/or class B (such as NDM) carbapenemases: in these cases, in areas endemic for MBL-producers, the use of a combination of ATM with a BLBLI could be the more effective and practical solution. ATM could be combined both with CZA and MVB and I-R; despite the novel BLBLIs could be realistically largely interchangeable for the treatment of infections due to CPE in real-life, the combination with AVI is preferable when OXA-48-producing *Enterobacterales* are present [29].

Only one study [20] directly compared clinical outcomes in patients who received CZA versus MVB as mono or combo regimens for CRE infections: KPC-Kp was the primary causative organism in both groups, but in the MVB group there were proportionally more patients with *ampC*-harboring *Enterobacterales* [44]. Patients in the CZA arm received combination therapy more often than patients in the MVB arm (61.0% versus 15.4%; $p < 0.01$), no difference of mortality was observed at 30-day (19.1% vs 11.5%, $p = 0.57$) and 90-day (28.6 vs 26.9%, $p = 0.48$) for CZA and MVB groups, respectively, and the rate of adverse events was similar [20]. Clinical success was observed in 61.9% of patients in the CZA group and 69.2% in the MVB group ($p = 0.49$). The median duration of combination therapy was 8.8 days with CZA and 3.1 days with MVB ($p = 0.08$). Emergence of MVB resistance was not observed within the study period, while among 15 patients who were treated with CZA and had recurrent infections three developed drug resistance within 90 days: interestingly, all three patients were found to have respiratory sources and received renal replacement therapy [20], confirming what has been previously discussed on pneumonia as a risk factor for CZA resistance and the need of adequate considerations about the pharmacokinetics/pharmacodynamics of CZA in particular high-risk populations [13, 17]. This study [20] supported the use of MVB as the preferred monotherapy agent for KPC-producing *Enterobacterales*.

In conclusion, the role of combination regimens with the novel BLBLIs for the treatment of infections due to CPE, in order to prevent bacterial resistance and optimize their overall efficacy, deserves further investigations. We hope

that well-conducted RCTs will be performed to clarify these important aspects so that these agents can be effectively used for the longest possible period of time and minimizing the selection of resistance.

Conflict of interest

C.T. has received fees for speaking at symposia organized on behalf of Pfizer, Novartis, Merck, Angelini, Zambon, Ther-mofischer, Biotest, Gilead, Hikma, Biomerieux and Astellas. S.M. and B.V.: none to declare.

Funding No funding has been received for this work. This study was carried out as part of our routine work.

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