ORIGINAL PAPER

The Shapiro–Procalcitonin algorithm (SPA) as a decision tool for blood culture sampling: validation in a prospective cohort study

Géraldine Wyss¹ · Silvia Berger¹ · Sebastian Haubitz^{1,2} · Hans Fankhauser³ · Ulrich Buergi⁴ · Beat Mueller^{1,5} · **Philipp Schuetz1 · Christoph A. Fux1,2 · Anna Conen[2](http://orcid.org/0000-0002-5216-4109)**

Received: 30 December 2019 / Accepted: 3 April 2020 / Published online: 14 April 2020 © Springer-Verlag GmbH Germany, part of Springer Nature 2020

Abstract

Purpose Blood cultures (BC) are the gold standard for bacteremia detection despite a relatively low diagnostic yield and high costs. A retrospective study reported high predictive values for BC positivity when combining the clinical Shapiro score with procalcitonin (PCT).

Methods Single-center, prospective cohort study between 01/2016 and 02/2017 to validate SPA algorithm, including a modifed Shapiro score≥3 points (S) PLUS admission PCT>0.25 µg/l (P), or presence of overruling safety criteria (A) in patients with systemic infammatory response syndrome. The diagnostic yield of SPA compared to non-standardized clinical judgment in predicting BC positivity was calculated and results presented as odds ratios (OR) with 95% confdence intervals. **Results** Of 1438 patients with BC sampling, 215 (15%) had positive BC which increased to 31% (173/555) in patients fulflling SP criteria (OR for BC positivity 9.07 [6.34–12.97]). When adding 194 patients with overruling safety criteria (i.e., SPA), OR increased to 11.12 (6.99–17.69), although BC positivity slightly decreased to 26%. With an area under the receiver operating curve of 0.742, SPA indicated better diagnostic performance than its individual components. Positive BC in 689 patients not fulflling SPA (sampling according to non-standardized clinical judgment) were rare (3%; OR for BC positivity 0.09 [0.06–0.14]). Eight out of 21 missed pathogens were still identifed by sampling the primary infection focus. **Conclusions** This study validates the high predictive value of SPA for bacteremia, increasing true BC positivity from 15 to 26%. Restricting BC sampling to SPA would have reduced BC sampling by 48%, while still detecting 194/215 organisms (90%), which makes SPA a valuable diagnostic stewardship tool.

Keywords Blood culture · Shapiro · Procalcitonin · Diagnostic stewardship

Géraldine Wyss and Silvia Berger have contributed equally to this work.

 \boxtimes Anna Conen anna.conen@ksa.ch

- ¹ Medical University Clinic, Kantonsspital Aarau, Aarau, Switzerland
- ² Department of Infectious Diseases and Hospital Hygiene, Kantonsspital Aarau, Aarau, Switzerland
- ³ Department of Clinical Microbiology, Institute of Laboratory Medicine, Kantonsspital Aarau, Aarau, Switzerland
- ⁴ Department of Emergency Medicine, Kantonsspital Aarau, Aarau, Switzerland
- ⁵ Department of Endocrinology, Diabetology and Metabolism, Kantonsspital Aarau, Aarau, Switzerland

Introduction

Infectious diseases with bacteremia are increasing worldwide, with mortality rates as high as 18–42%, particularly when diagnosis is delayed and treatment is inappropriate $[1–7]$ $[1–7]$ $[1–7]$ $[1–7]$. Blood cultures (BC) are the gold standard for the detection of bacteremia, allowing pathogen identifcation and antimicrobial susceptibility testing [[1,](#page-9-0) [4,](#page-9-2) [6](#page-9-3)]. The reported sensitivity of BC to detect bacteremia varies between 73 and 100%, depending on the number of BC drawn, the infectious source, patient's characteristics, and antibiotic pretreatment $[8-11]$ $[8-11]$ $[8-11]$. The diagnostic yield of BC in a clinical routine test is limited. Of all BC drawn in emergency departments, only 7–20% turned out to be positive [\[8,](#page-9-4) [12](#page-9-6)–[15\]](#page-9-7). Despite this limitation, the threshold to draw BC is low, thereby increasing health care costs with a limited beneft or even causing harm in case of contaminated (false positive) BC $[16]$. As there are no universal guidelines on when to draw BC, non-standardized clinical judgment by treating physicians is followed in the clinical routine practice [\[17,](#page-9-9) [18\]](#page-9-10). Several studies improved the prediction of bacteremia by applying clinical and laboratory scores, including biomarkers [[8,](#page-9-4) [19](#page-9-11)[–22](#page-9-12)]. A score published by Shapiro et al. combined three major and nine minor criteria (Table [1\)](#page-1-0), suggesting BC sampling if at least one major or two minor criteria were present [[19](#page-9-11)]. The sensitivity of 98%, with a resulting negative likelihood ratio (LR) of 0.08, allowed to safely abstain from BC in an important number of patients.

At our hospital, the performance of diferent bacteremia prediction scores was analyzed in 1083 consecutive patients with BC collection in the medical emergency department by Laukemann et al. [[23](#page-9-13)]. Thereby, BC had been collected according to non-standardized clinical judgment. The best predictor for the 104 (9.6%) bacteremia episodes was an adapted Shapiro score with≥3 points (when applying two points for each major and one point for each minor criterion), with an admission procalcitonin (PCT) level of > 0.25 µg/l in patients with a systemic infammatory response syndrome (SIRS) and suspected infection [\[23\]](#page-9-13). Applying this algorithm, the number of BC could have been reduced by 42% while missing 4 bacteremia only (4%).

The aim of the present study was to prospectively validate these criteria for predicting bacteremia and to analyze the potential for saving unnecessary BC.

Table 1 Modifed Shapiro score

Major criteria (two points each)				
1	Fever > 39.4 °C			
2	Suspected endocarditis			
\mathcal{E}	Indwelling endovascular catheter			
Minor criteria (one point each)				
1	Fever $38.3 - 39.3$ °C			
2	Shivering			
3	Vomiting			
4	Systolic blood pressure < 90 mmHg			
5	Leucocyte count (WBC) > 18 G/l			
6	Neutrophilic granulocytes $>80\%$			
	Non-segmented neutrophils $> 5\%$			
8	Thrombocyte count < 150 G/l			
9	Creatinine > 176 μ mol/l (> 2 mg/dl)			

Shapiro score positive if≥3 points

Methods

Study design

Between 4 January 2016 and 28 February 2017, we conducted a prospective cohort study in the Medical and Neurological Emergency Department of the Kantonsspital Aarau, a 600-bedded tertiary-care hospital in Switzerland. All adult patients who had at least one BC collected and were admitted to medical or neurological wards (including intensive-care unit) were included (Fig. [1](#page-2-0)). For the sample size calculation, the following parameters were used: power 90%, α =5%, and a signifcance level 5% (one-sided). 1500 patients over a period of 1 year were, therefore, planned to be included.

The study was performed in a quasi-experimental design that compared patients meeting predefined SPA (Shapiro score, procalcitonin, and overruling criteria) criteria for BC collection with patients not meeting these criteria, but with BC collected according to non-standardized clinical judgment by the treating physician, which represents the current clinical practice.

Defnitions

Systemic infammatory response syndrome (SIRS):≥2/4 criteria of body temperature > 38° C or < 36° C, heart rate > 90 beats per minute (bpm), respiratory rate > 20/min or PaCO₂ < 32 mmHg, and WBC count > 12 G/l or < 4 G/l.

Shapiro–Procalcitonin-Overruling criteria (SPA): "S" for modified Shapiro score \geq 3 points (Table [1](#page-1-0)), "P" for admission PCT level > 0.25 µg/l, and "A" for overruling safety criteria including severe sepsis and septic shock (i.e., SIRS with hypotension and end-organ damage or refractory hypotension), suspected endovascular infection or bacterial meningitis, immunosuppression, neutropenic fever (neutrophil count < 0.5 G/l), and fever in hematologic stem cell (HSCT) and solid organ (SOT) transplantation patients.

Shapiro–Procalcitonin (SP): "S" for modified Shapiro score≥3 points (Table [1](#page-1-0)) and "P" for admission PCT $level > 0.25$ μ g/l.

Quick sepsis-related organ failure assessment (qSOFA score): 1 point each for respiratory rate \geq 22 breaths/ min, Glasgow coma scale<15 points, and systolic blood pressure ≤ 100 mmHg.

Positive BC: Pathogen compatible with the clinical presentation.

False positive BC: Contaminated BC (were counted as negative).

Fig. 1 Study population according to the presence of SIRS with suspected infection, SP criteria as well as overruling safety criteria (A). Presented are numbers and rates of positive blood cultures. *SIRS* systemic infammatory response syndrome, *SP* modifed Shapiro score≥3 points AND Procalcitonin>0.25 µg/l, *SPA* modifed Shapiro score≥3 points AND Procalcitonin>0.25 µg/l, OR overruling safety criteria (A); *BC* blood culture

Indication for BC sampling and control group

BC sampling was suggested in all patients with suspected infection and SIRS fulflling SPA criteria, i.e., fulflling either a modifed Shapiro score≥3 points plus admission PCT level > 0.25 µg/l, or fulfilling overruling safety criteria. The latter were defned to identify the most vulnerable patients.

The study was approved by the local ethics committee (Ethikkommission Nordwest- und Zentralschweiz). Given that this was an observational quality-control study, the need for an individual informed consent was waived.

Data collection and laboratory data

Data on comorbidities and symptoms were collected using a case report form (CRF). In addition, laboratory and clinical parameters, data on prior hospitalizations, antibiotic pre-treatment within 30 days before admission, use of antipyretic drugs within 6 h before hospitalization, symptom

onset (> 7 days and \leq 1 day), and acute deterioration (< 24 h and>2 days) before admission were recorded. Every completed CRF was reviewed by an infectious disease (ID) specialist.

The PCT measurement was performed on an ADVIA Centaur (Siemens Healthineers, D) using license partner reagents (B·R·A·H·M·S PCT).

Blood culture sampling

One BC included aerobic and anaerobic bottles containing 8–10 ml blood each. Routinely, two BC were collected per patient. After skin disinfection with 77% ethanol, BC were drawn from one venous puncture [[24\]](#page-9-14). In suspected infective endocarditis, three BC were collected within 6 h in stable or within 20–30 min in unstable patients. For the bacterial culture, BACTEC FX (BD Becton Dickinson) and BacT/Alert (BioMérieux) were used according to the manufacturer's instructions. Pathogen identifcation and susceptibility testing were performed according to the European Committee on Antimicrobial Susceptibility Testing (EUCAST) [[25](#page-9-15)]. Every positive BC was categorized by an ID specialist during CRF review as positive or false positive, i.e., contaminated. Low-virulent organisms of the skin fora (i.e., coagulase-negative staphylococci (CNS), *Corynebacterium* spp., *Cutibacterium* spp., and *Bacillus* spp.), without evidence of an endovascular infection, were classifed as contaminants, particularly if cultured in one bottle only or after>24 h of culture [[26](#page-9-16)].

Statistical analysis

For the primary outcome, the diagnostic yield of SPA compared to non-standardized clinical judgment was calculated by logistic regression analysis and presented as odds ratios (OR) with 95% confdence intervals (95% CI) for positive BC. Secondary outcomes included missed bacteremia by SPA, individual performance of SIRS with suspected infection, modified Shapiro score \geq 3 points (S), PCT > 0.25 µg/l (P), overruling safety criteria (A), and the combination of them (SP and SPA, respectively) to predict blood culture positivity. For each single item of the algorithm, logistic regression analysis was performed providing OR and 95% CI for positive BC. In addition to sensitivities and specifcities, areas under the receiver-operating characteristic curves (AUROC) for every item of the algorithm were calculated, as well as negative (NPV) and positive (PPV) predictive values for SPA.

Continuous variables were presented as medians with interquartile ranges (IQR), and categorical variables as numbers and percentages. Continuous variables were compared with the Wilcoxon rank-sum test and frequencies by Chisquare test. All tests were two-tailed and a p -value < 0.05 was considered statistically signifcant. The statistical analysis was done using STATA 15.1 (Stata Corp, College Station, TX).

Results

Baseline characteristics

We included 1438 patients with BC sampling during the 14-month study period. The median age was 72 years (IQR 59–80), among whom 594 (41%) were females.

In Table [2,](#page-4-0) baseline characteristics of patients with and without true positive BC were compared. Patients with positive BC had more comorbidities; sepsis, severe sepsis, and septic shock were more common as was acute deterioration (<24 h) before admission. Bacteremia patients had higher modified Shapiro scores with median 4 vs. 3 points, higher qSOFA scores with median 1 vs. 0 points, and higher infammatory markers including median PCT with 2.89 vs. 0.32 µg/l, median C-reactive protein (CRP) with 120 vs. 80 mg/l and median neutrophil count with 9.6 vs. 8.2 G/l; platelet and lymphocyte counts were lower. The duration of symptoms before hospitalization was similar between the two groups, but patients with negative BC were pretreated with antibiotics more often. Urogenital, hepatobiliary, endovascular, and bone and joint infections were more common in patients with positive BC, while ear/nose/throat and pulmonary infections were more common in patients with negative BC.

Blood culture collection according to SPA versus non‑standardized clinical judgment

Of the 1438 patients with BC sampling, 989 (69%) presented with SIRS and suspected infection, of whom 555 met SP criteria (39% of all) and additional 194 (13% of all) overruling safety criteria (A) (Fig. [1](#page-2-0)). Thus, 749 (52%) fulflled SPA criteria and qualifed for BC. The remaining 689 patients (48%) were sampled according to non-standardized clinical judgment. Of the 215 patients (15%) with positive BC, 194 patients (13% of all) fulflled SPA (173 (12% of all) fulflled SP and 21 (1% of all) overruling safety criteria), and 21 patients (1% of all) were sampled according to nonstandardized clinical judgment.

The logistic regression analysis, to test the efficiency of SPA for positive BC compared to non-standardized clinical judgment, is shown in Fig. [2](#page-5-0). In patients fulflling SP criteria $(n=555)$, the proportion of positive BC increased to 31% (173/555) with an OR of 9.07 (95% CI 6.34–12.97) for positive BC. When SPA was fulfilled $(n = 749)$, the number of positive BC increased to 194 with a slightly decreased BC positivity rate (26%), but an increased OR

Table 2 Baseline characteristics of the 1438 patients with blood culture sampling, comparing patients with positive and negative blood cultures

BC blood culture, *IQR* interquartile range, *SOT* solid organ transplantation, *HSCT* hematopoietic stem cell transplantation, *SIRS* systemic infammatory response syndrome, *eGFR* estimated glomerular fltration rate, *NYHA* New York heart association, *GOLD* Global Initiative for Chronic Obstructive Lung Disease, *qSOFA* quick sepsis-related organ failure assessment score, *Sepsis* SIRS with suspected infection, *severe sepsis* with hypotension and end-organ damage, *septic shock* severe sepsis with refractory hypotension

qSOFA score (1 point each for respiratory rate≥22 breaths/min, Glasgow coma scale<15 points, and systolic blood pressure≤100 mmHg). Immunosuppression:>20 mg prednisone daily or equivalent, SOT or

Table 2 (continued) HSCT; diabetes: use of antidiabetic drugs or HbA1c≥6.5%

Fig. 2 Performance of SP criteria with and without overruling safety criteria (A) for positive blood cultures compared to non-standardized clinical judgment: Logistic regression analysis. Analyzed are 1438 patients with blood culture sampling. *SP* modifed Shapiro score≥3

of 11.12 (95% CI 6.99–17.69) for positive BC. Among the 194 patients not fulflling SP, but with at least one overruling safety criterion (A), 21 patients (11%) had positive BC. In SPA-negative patients with BC collection, according to non-standardized clinical judgment, only 3% (21/689) had positive BC with an OR of 0.09 (95% CI 0.06–0.14) for positive BC. Similarly, the yield was low for patients without SIRS (OR 0.15, 95% CI 0.09–0.25) or with SIRS but not SPA, either due to a modifed Shapiro score<3 points (OR 0.26, 95% CI 0.08–0.82) or a PCT≤0.25 µg/l (OR 0.13, 95% CI 0.03–0.54).

Performance of SPA and its individual components to predict positive blood cultures

Figure [3](#page-6-0) shows the BC positivity rates and the number of patients qualifying for BC sampling comparing SPA with its individual components, i.e., SIRS and suspected infection, overruling safety criteria (A), modified Shapiro score \geq 3 points (S), $PCT > 0.25 \mu g/l$ (P), and the combination of the latter (SP). Sensitivity, specifcity and AUROC for BC diagnostic are shown for each single item of the algorithm. Adding the overruling safety criteria to SP increased the sensitivity from 81% (SP) to 90% (SPA) at the price of reducing the specifcity from 69% (SP) to 52% (SPA). While S and P criteria alone showed sensitivities (each 91%) similar to SP

points AND Procalcitonin (PCT)>0.25 µg/l, *SPA* modifed Shapiro score \geq 3 points AND Procalcitonin (PCT) > 0.25 μ g/l, OR overruling safety criteria (A), *BC* blood culture, *SIRS* systemic infammatory response syndrome, *OR* odds ratio, *CI* confdence interval

or SPA, they required a signifcantly higher number of BC collections $[n=926$ for (S) and $n=900$ for (P) vs. $n=555$ (SP) and *n*=749 for (SPA), respectively]. By applying SPA, 48% (689/1438) of BC could have been, by applying SP even 61% (883/1438).

The AUROC illustrates the higher diagnostic yield of SP (0.746) and SPA (0.724) compared to S (0.657) and P (0.667). Logistic regression analysis also calculated more favorable OR for BC positivity for the combinations SPA and SP with 11.12 (95% CI 6.99–17.69) and 9.07 (95% CI 6.34–12.97), respectively, as compared to Shapiro score \geq 3 points and $PCT > 0.25$ µg/l each alone with 6.97 (95% CI 4.31–11.30) and 7.53 (95% CI 4.66–12.20), respectively. PPV and NPV of SPA were 25.9% (95% CI 22.8–29.2%) and 97% (95.4–98.1%), respectively.

Patients with positive blood cultures missed by SPA and contaminated blood cultures

The 21 patients (10%) with positive BC missed by SPA suffered from urogenital infections $(n=5)$, cholangitis $(n=4)$, infective endocarditis or infected aneurysma spurium $(n=4)$, and skin and soft-tissue infections $(n=3)$ as the most common infections (Table [3\)](#page-7-0). While 16 of these patients were missed, because SIRS was not fulflled, 3 did not meet the modifed Shapiro score and 2 had PCT below the threshold

Fig. 3 Performance of SP criteria with and without overruling safety criteria (A) for positive blood cultures in comparison to the performance of its individual components: sensitivity, specificity, ROC, and logistic regression analysis. Analyzed are 1438 patients with blood culture sampling. The height of the white columns indicates the number of patients requiring BC sampling. The black columns indicate the number of positive BC with the percentage of positive BC within

of 0.25 µg/l. Eight pathogens could be detected alternatively (4 by urine, 2 by synovial fuid, and 1 by vertebral tissue culture; 1 by pneumococcal antigen testing in urine). If only SP criteria were applied, 42/215 (20%) true positive BC would have been missed, thereby underlining the importance of the safety overruling criteria.

Of the 1438 BC, 52 (4%) were considered contaminated (false positive). The most common contaminants were CNS (65%), followed by viridans group *streptococci* (13%), *Cutibacterium* spp. (6%), and *Corynebacterium* spp. (4%).

Discussion

Without general guideline recommendations regarding indications, BC in clinical practice are drawn based on the physicians' subjective judgment [[27](#page-9-17)]. This is associated with a low diagnostic yield (positivity rates between 7 and 15% on average) and unnecessary costs [\[1](#page-9-0), [12](#page-9-6)–[14,](#page-9-18) [23,](#page-9-13) [28](#page-9-19)]. False positive BC may prompt unnecessary, expensive,

the column. *SP* modifed Shapiro score≥3 points AND Procalcitonin (PCT)>0.25 µg/l, *SPA* modifed Shapiro score≥3 points AND Procalcitonin (PCT) >0.25 µg/l, OR overruling safety criteria (A), *BC* blood culture, *SIRS* systemic infammatory response syndrome, *ROC* receiver-operating characteristic, *CI* confdence interval, *OR* odds ratio

and potentially harmful examinations [[27,](#page-9-17) [28\]](#page-9-19). Therefore, we aimed to prospectively validate a prediction tool for bacteremia in unselected patients admitted to internal medicine and neurology wards by introducing SPA, which is an algorithm combining a modifed Shapiro score (S) with PCT values (P) and overruling safety criteria (A) in patients admitted with SIRS and suspected infection. We explicitly decided to validate the algorithm against the current clinical practice, i.e., BC sampling according to non-standardized clinical judgment, as this allowed to demonstrate the net beneft.

A modifed Shapiro score≥3 points plus an admission $PCT > 0.25$ µg/l (i.e., SP) doubled the BC positivity rate in our cohort from 15 to 31%. Following that, SP criteria would have allowed to abstain from 61% of all BC while missing 20% of positive BC. The safety overruling criteria (A) add indications for supplementary BC for the most vulnerable patients. The 86% specificity of these criteria to predict bacteremia underlines their clinical beneft. The full algorithm increased the sensitivity from 81% (SP) to 90% (SPA), which

Table 3 Patients with positive blood cultures, but no formal indication for blood culture sampling (i.e., missed by SPA)

	Patient Diagnosis		Reason for SPA negativity Pathogen detected in blood culture	Alternative pathogen detection
1	Urinary tract infection	$SIRS < 2$ criteria	Escherichia coli	Urine culture
2	Urinary tract infection	$SIRS < 2$ criteria	Escherichia coli	Urine culture
3	Cholangitis	$SIRS < 2$ criteria	Escherichia coli	
4	Cholangitis	$SIRS < 2$ criteria	Escherichia coli	
5	Cholangitis	SIRS < 2 criteria	Escherichia coli	
6	Prosthetic valve endocarditis	$SIRS < 2$ criteria	Enterococcus faecalis	
7	Cellulitis	$SIRS < 2$ criteria	Pseudomonas aeruginosa	
8	Septic arthritis	$SIRS < 2$ criteria	Staphylococcus aureus	Synovial fluid culture
9	Spondylodiscitis	$SIRS < 2$ criteria	Staphylococcus lugdunensis	Culture of vertebral body aspirate
10	Pneumonia	$SIRS < 2$ criteria	Streptococcus pneumoniae	Pneumococcal antigen test in urine
11	Native valve endocarditis	$SIRS < 2$ criteria ^a	Enterococcus faecalis	Synovial fluid culture
12	Prosthetic valve endocarditis	$SIRS < 2$ criteria ^a	Staphylococcus aureus	
13	Infected inguinal aneurysma spurium	$SIRS < 2$ criteria ^a	Staphylococcus aureus	
14	Erysipelas	$SIRS < 2$ criteria ^a	Streptococcus agalactiae	
15	Neutropenic colitis	$SIRS < 2$ criteria ^a	Citrobacter braakii	
16	Translocation due to gastrointesti- nal angiodysplasia	$SIRS < 2$ criteria ^a	Polymicrobial (Pseudomonas aer- uginosa, Streptococcus milleri, Fusobacterium necrophorum, Bacteroides fragilis)	
17	Urinary tract infection	Shapiro < 3 points	Escherichia coli	
18	Urinary tract infection	Shapiro $<$ 3 points	Escherichia coli	Urine culture
19	Cholangitis	Shapiro $<$ 3 points ^a	Escherichia coli	
20	Urinary tract infection	$PCT \leq 0.25 \text{ µg/l}^a$	Escherichia coli	Urine culture
21	Erysipelas	$PCT \leq 0.25 \text{ µg/l}^a$	Group G β-hemolytic streptococci	

Shapiro modifed Shapiro score, *SIRS* systemic infammatory response syndrome, *PCT* Procalcitonin

a Symptom duration<24 h (in 9/21 patients)

would have allowed to detect 90% bacteremia despite omitting 48% of all BC. SPA, herein, outperformed its individual components with an OR of 11.1 in predicting BC positivity.

Selection of an algorithm

Several studies have identifed clinical and laboratory predictors of bacteremia and established prediction scores for defned infections, such as community-acquired pneumonia or cellulitis [[22,](#page-9-12) [29–](#page-9-20)[31\]](#page-9-21), and also for general internal and emergency medicine [\[8](#page-9-4), [19](#page-9-11), [32,](#page-9-22) [33\]](#page-9-23).

Jones et al. demonstrated that 95% of bacteremia fulfilled SIRS criteria $[20]$. With a negative LR of 0.09 (95%) CI 0.03–0.26), we made SIRS a prerequisite to consider BC in our study [\[27](#page-9-17)]. Lee and Metersky retrospectively developed scores to predict bacteremia including SIRS criteria, liver disease, and recent antibiotic treatment amongst others [\[21,](#page-9-25) [22\]](#page-9-12). If BC were abandoned in patients with a low risk of bacteremia, 38% and 79% of BC could be saved in these studies. However, opposite to SPA, both scores only applied to patients with pneumonia, and the most vulnerable patients were not represented by overruling safety criteria.

Modifed Shapiro score, procalcitonin, and SPA

Looking at individual biomarkers for bacteremia, PCT with an AUROC of 0.880–0.760 outperformed CRP with an AUROC of 0.600–0.650 [[23](#page-9-13), [34](#page-10-0)[–36\]](#page-10-1). By combining a PCT > 0.4 μ g/l and a Charlson comorbidity index ≥ 2, Tudela et al. identifed a subgroup of 26% among 412 study patients with a bacteremia rate of 3% as compared to 10% overall, which is low enough to abstain from BC sampling if the criteria were not fulflled [[32\]](#page-9-22). Takeshima et al. included 11 clinical and laboratory parameters and separated 759 (38%) of their 1982 study patients with a bacteremia rate of 2–3% as compared to 16% overall, again low enough to abstain from BC sampling [[33\]](#page-9-23). Unfortunately, no safety information regarding missed bacteremia was provided in that study. Shapiro et al. combined twelve clinical and laboratory parameters and evaluated them in 3730 emergency room patients with a bacteremia rate of 8% [\[19](#page-9-11)]. The

sensitivity of Shapiro's algorithm was 98%. The negative LR of 0.08 allowed a 27% BC reduction with only 2% of bacteremia missed. Still, important overruling safety criteria were missing.

Combination scores have shown AUROC values for bacteremia of 0.800–0.827 in derivation and 0.730–0.770 in validation cohorts, which are very similar to Laukemann's algorithm (AUROC 0.827; derivation) and the SP criteria (AUROC 0.746) [\[19](#page-9-11), [23,](#page-9-13) [32](#page-9-22)–[37\]](#page-10-2). With a 48% reduction in BC sampling, SPA showed a similar efficiency as the retrospective analysis by Laukemann et al., while the overall BC positivity rate was comparable (i.e., 10% and, in our study, 15%) [\[23\]](#page-9-13). The relatively low sensitivity of SP criteria for bacteremia (81%) could be increased to 90% by introducing the overruling safety criteria and, therefore, was in the range of the 96% in the Laukemann study [[23\]](#page-9-13).

The very low additional yield (3%) of BC collected solely, according to the physicians' clinical judgment from patients not fulflling SPA, underlines the robustness of the algorithm. Other studies also found unguided clinical judgment to be inefficient to guide BC sampling in febrile medical in-patients [[17,](#page-9-9) [18](#page-9-10)].

Missed bacteremia

By applying SPA, 21 patients (10%) with bacteremia would have been missed and 4% were missed in the Laukemann study [[23\]](#page-9-13). Two factors might have reduced the sensitivity of SPA: 29% of patients described a symptom duration ≤ 1 day and 53% an acute deterioration in the 24 h before admission. This may have been too short to reach SIRS or PCT cut-ofs. As a matter of fact, 18/21 bacteremia were missed, lacking inflammation criteria ($16 \times SIRS$, $2 \times PCT$), with nine (50%) reporting a symptom duration $\langle 24 \text{ h (Table 3)}.$ $\langle 24 \text{ h (Table 3)}.$ $\langle 24 \text{ h (Table 3)}.$ Consistently, both patients with normal PCT on admission reached the threshold on day 3 (data not shown). Therefore, BC may be considered in SIRS- or PCT-negative patients with a symptom duration $<$ 24 h. This might be particularly true for patients presenting with a safety overruling criterion as seen in 5 SIRS-negative patients in our analysis $(4 \times end$ ovascular infection, $1 \times$ neutropenic colitis). This fact was recently shown in patients with a central venous catheter infection with *S. aureus*, where the interval between clinical symptom onset and the diagnostic confrmation by BC collection was very short [\[38](#page-10-3)]. Second, prescribed betablockers and antipyretics may have mitigated the two SIRS criteria, tachycardia and fever.

In 8/21 patients (38%), the causative pathogen was identifed by alternative testing, particularly by urine culture or antigen testing, thereby causing a rise in the overall pathogen detection rate to 94%. From a more general point of view, bacteremia detection might be dispensable if treatment decisions are not altered, such as in febrile urinary tract infections or in community-acquired pneumonia which often are also of viral etiology [\[39](#page-10-4)–[42\]](#page-10-5).

Limitations and strengths

The main weakness is the lack of characterization of the SIRS-negative patients, for whom we only collected data if they underwent BC sampling. With our quasi-experimental study design, the control group of non-standardized clinical judgment is indeed heterogenous. The algorithm might appear complex and some physicians might be reluctant to calculate SPA, although almost all parameters are routinely collected in patients with suspected infections. Finally, the availability, turnaround time, and costs of PCT may vary in diferent settings and could impair applicability. However, future point-of-care testing (POCT) could facilitate testing and decrease costs.

The main strength was the large, unselected and, therefore, representative cohort of internal medicine and neurologic patients presenting in an emergency department. Consequently, generalizability to surgical and outpatients may not be given. Patient safety was a core component with the inclusion of overruling criteria for the most vulnerable patients. With the study duration of 14 months, the seasonality of infectious diseases is represented. SPA was tested against non-standardized clinical judgment, which includes individualized decisions (i.e., the "gut feeling" of the treating physicians) and, therefore, is able to show the net beneft to the current clinical practice.

Conclusions

As infectious diseases with bacteremia are associated with high morbidity and mortality, a reliable diagnostic is essential. A rational diagnostic can limit unnecessary costs and potentially harmful false positive results. We prioritized patient safety by including overruling criteria for the most vulnerable patients. SPA outcompetes SIRS, modifed Shapiro score, and PCT, both alone and in combination. Compared to non-standardized clinical judgment, which is the current clinical practice, SPA could reduce BC sampling by 48% while still detecting 194/215 (90%) of the relevant pathogens. This makes SPA a valuable diagnostic stewardship tool. Nevertheless, one should bear in mind that patients with an acute symptom onset or deterioration $(< 24$ h) should be considered for BC sampling despite not fulflling the algorithm, as SIRS and PCT could still be false negative. This should especially be considered in patients fulflling overruling safety criteria.

Acknowledgements We thank the nurses in the emergency department responsible for BC and biomarker sampling throughout the study period, the study nurses involved in data collection, and the laboratory and microbiology teams who performed all blood tests and microbiological analyses needed. In addition, we thank all the patients who participated in this study.

Funding This work was supported by the Research Fund of the Kantonsspital Aarau, Switzerland. The sponsor had no role in the study design, data collection or interpretation, and writing and submitting the report.

Compliance with ethical standards

Conflict of interest On behalf of all authors, the corresponding author states that there is no confict of interest.

References

- 1. Ratzinger F, Dedeyan M, Rammerstorfer M, Perkmann T, Burgmann H, Makristathis A, et al. A risk prediction model for screening bacteremic patients: a cross sectional study. PLoS ONE. 2014;9:e106765.
- 2. Laupland KB. Incidence of bloodstream infection: a review of population-based studies. Clin Microbiol Infect. 2013;19:492–500.
- 3. Bearman GM, Wenzel RP. Bacteremias: a leading cause of death. Arch Med Res. 2005;36:646–59.
- 4. Weinstein MP, Towns ML, Quartey SM, Mirrett S, Reimer LG, Parmigiani G, et al. The clinical signifcance of positive blood cultures in the 1990s: a prospective comprehensive evaluation of the microbiology, epidemiology, and outcome of bacteremia and fungemia in adults. Clin Infect Dis. 1990s;24:584–602.
- 5. Yoshida T, Tsushima K, Tsuchiya A, Nishikawa N, Shirahata K, Kaneko K, et al. Risk factors for hospital-acquired bacteremia. Intern Med. 2005;44:1157–62.
- 6. Reimer LG, Wilson ML, Weinstein MP. Update on detection of bacteremia and fungemia. Clin Microbiol Rev. 1997;10:444–65.
- 7. Wisplinghoff H, Bischoff T, Tallent SM, Seifert H, Wenzel RP, Edmond MB. Nosocomial bloodstream infections in US hospitals: analysis of 24,179 cases from a prospective nationwide surveillance study. Clin Infect Dis. 2004;39:309–17.
- 8. Tokuda Y, Miyasato H, Stein GH. A simple prediction algorithm for bacteraemia in patients with acute febrile illness. QJM. 2005;98:813–20.
- Weinstein MP. Current blood culture methods and systems: clinical concepts, technology, and interpretation of results. Clin Infect Dis. 1996;23:40–6.
- 10. Lee A, Mirrett S, Reller LB, Weinstein MP. Detection of bloodstream infections in adults: how many blood cultures are needed? J Clin Microbiol. 2007;45:3546–8.
- 11. Khatib R, Simeunovic G, Sharma M, Fakih MG, Johnson LB, Briski L, et al. Blood culture series beneft may be limited to selected clinical conditions: time to reassess. Clin Microbiol Infect. 2015;21:332–6.
- 12. Wang SJ, Kuperman GJ, Ohno-Machado L, Onderdonk A, Sandige H, Bates DW. Using electronic data to predict the probability of true bacteremia from positive blood cultures. Proc AMIA Symp. 2000:893–7.
- 13. Paul M, Andreassen S, Nielsen AD, Tacconelli E, Almanasreh N, Fraser A, et al. Prediction of bacteremia using TREAT, a computerized decision-support system. Clin Infect Dis. 2006;42:1274–82.
- 14. Previsdomini M, Gini M, Cerutti B, Dolina M, Perren A. Predictors of positive blood cultures in critically ill patients: a retrospective evaluation. Croat Med J. 2012;53:30–9.
- 15. Bates DW, Cook EF, Goldman L, Lee TH. Predicting bacteremia in hospitalized patients. A prospectively validated model. Ann Intern Med. 1990;113:495–500.
- 16. Leibovici L, Greenshtain S, Cohen O, Mor F, Wysenbeek AJ. Bacteremia in febrile patients. A clinical model for diagnosis. Arch Intern Med. 1991;151:1801–6.
- 17. Poses RMAM. When you hear hoofbeats: physicians overvalue recent experience when diagnosing bacteremia [abstract]. Clin Res. 1989;37:781A.
- 18. Makadon HJ, Bor D, Friedland G, Dasse P, Komaroff AL, Aronson MD. Febrile inpatients: house officers' use of blood cultures. J Gen Intern Med. 1987;2:293–7.
- 19. Shapiro NI, Wolfe RE, Wright SB, Moore R, Bates DW. Who needs a blood culture? A prospectively derived and validated prediction rule. J Emerg Med. 2008;35:255–64.
- 20. Jones GR, Lowes JA. The systemic infammatory response syndrome as a predictor of bacteraemia and outcome from sepsis. QJM. 1996;89:515–22.
- 21. Lee J, Hwang SS, Kim K, Jo YH, Lee JH, Kim J, et al. Bacteremia prediction model using a common clinical test in patients with community-acquired pneumonia. Am J Emerg Med. 2014;32:700–4.
- 22. Metersky ML, Ma A, Bratzler DW, Houck PM. Predicting bacteremia in patients with community-acquired pneumonia. Am J Respir Crit Care Med. 2004;169:342–7.
- 23. Laukemann S, Kasper N, Kulkarni P, Steiner D, Rast AC, Kutz A, et al. Can we reduce negative blood cultures with clinical scores and blood markers? Results from an observational cohort study. Medicine (Baltimore). 2015;94:e2264.
- 24. Weinstein MP, Reller LB, Murphy JR, Lichtenstein KA. The clinical signifcance of positive blood cultures: a comprehensive analysis of 500 episodes of bacteremia and fungemia in adults. I. Laboratory and epidemiologic observations. Rev Infect Dis. 1983;5:35–533.
- 25. The European Commitee on Antimicrobial Suspectibility Testing. Breakpoint tables for interpretation of MICs and zone diameters, Version 6.0, 2016. [https://www.eucast.org/clinical_breakpoints/.](http://www.eucast.org/clinical_breakpoints/)
- 26. Hall KK, Lyman JA. Updated review of blood culture contamination. Clin Microbiol Rev. 2006;19:788–802.
- 27. Coburn B, Morris AM, Tomlinson G, Detsky AS. Does this adult patient with suspected bacteremia require blood cultures? JAMA. 2012;308:502–11.
- 28. Bates DW, Goldman L, Lee TH. Contaminant blood cultures and resource utilization. The true consequences of false-positive results. JAMA. 1991;265:365–9.
- 29. Kim B, Choi J, Kim K, Jang S, Shin TG, Kim WY, et al. Bacteremia prediction model for community-acquired pneumonia: external validation in a multicenter retrospective cohort. Acad Emerg Med. 2017;24:1226–344.
- 30. Kim B, Kim K, Lee J, Kim J, Jo YH, Lee JH, et al. Impact of bacteremia prediction rule in CAP: before and after study. Am J Emerg Med. 2018;36:758–62.
- 31. Lee CY, Kunin CM, Chang C, Lee SS, Chen YS, Tsai HC. Development of a prediction model for bacteremia in hospitalized adults with cellulitis to aid in the efficient use of blood cultures: a retrospective cohort study. BMC Infect Dis. 2016;16:581–91.
- 32. Tudela P, Lacoma A, Prat C, Modol JM, Gimenez M, Barallat J, et al. Prediction of bacteremia in patients with suspicion of infection in emergency room. Med Clin (Barc). 2010;135:685–90.
- 33. Takeshima T, Yamamoto Y, Noguchi Y, Maki N, Gibo K, Tsugihashi Y, et al. Identifying patients with bacteremia in communityhospital emergency rooms: a retrospective cohort study. PLoS ONE. 2016;11:e0148078.
- 34. Leli C, Ferranti M, Marrano U, Al Dhahab ZS, Bozza S, Cenci E, et al. Diagnostic accuracy of presepsin (sCD14-ST) and procalcitonin for prediction of bacteraemia and bacterial DNAaemia in patients with suspected sepsis. J Med Microbiol. 2016;65:713–9.
- 35. Tromp M, Lansdorp B, Bleeker-Rovers CP, Gunnewiek JM, Kullberg BJ, Pickkers P. Serial and panel analyses of biomarkers do not improve the prediction of bacteremia compared to one procalcitonin measurement. J Infect. 2012;65:292–301.
- 36. Jeong S, Park Y, Cho Y, Kim HS. Diagnostic utilities of procalcitonin and C-reactive protein for the prediction of bacteremia determined by blood culture. Clin Chim Acta. 2012;413:1731–6.
- 37. Jessen MK, Mackenhauer J, Hvass AM, Ellermann-Eriksen S, Skibsted S, Kirkegaard H, et al. Prediction of bacteremia in the emergency department: an external validation of a clinical decision rule. Eur J Emerg Med. 2016;23:44–9.
- 38. Smit J, Rieg SR, Wendel AF, Kern WV, Seifert H, Schonheyder HC, et al. Onset of symptoms, diagnostic confrmation, and

occurrence of multiple infective foci in patients with *Staphylococcus aureus* bloodstream infection: a look into the order of events and potential clinical implications. Infection. 2018;46:651–8.

- 39. Chalasani NP, Valdecanas MA, Gopal AK, McGowan JE Jr, Jurado RL. Clinical utility of blood cultures in adult patients with community-acquired pneumonia without defned underlying risks. Chest. 1995;108:932–6.
- 40. McMurray BR, Wrenn KD, Wright SW. Usefulness of blood cultures in pyelonephritis. Am J Emerg Med. 1997;15:137–40.
- 41. Pasternak EL 3rd, Topinka MA. Blood cultures in pyelonephritis: Do results change therapy? Acad Emerg Med. 2000;7:1170.
- 42. Metlay JP, Waterer GW, Long AC, Anzueto A, Brozek J, Crothers K, et al. Diagnosis and treatment of adults with communityacquired pneumonia. An official clinical practice guideline of the American Thoracic Society and infectious diseases society of America. Am J Respir Crit Care Med. 2019;200:e45–e67.