SYSTEMATICS, MORPHOLOGY AND PHYSIOLOGY

New Complex of Cryptic Species Discovered in Genus *Biblis* **(Papilionoidea: Nymphalidae: Biblidinae) in Mexico**

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Received: 14 November 2021 / Accepted: 5 May 2022 / Published online: 23 June 2022© Sociedade Entomológica do Brasil 2022

Abstract

Our research focuses on demonstrating the existence of cryptic species named under *Biblis aganisa* Boisduval. We used COI sequences to delimit *Biblis* species for Mexico using species delimitation analyses and examined phylogenetic relationships with sequences from Mexico, Costa Rica, Argentina, USA, and Guana Island using a Bayesian inference tree. We performed a discriminant analysis with quantitative traits using female and male wing and genitalia, and a tree of maximum parsimony based on 39 qualitative characters of wings, head, and male genitalia. The results were congruent in the three analyses. Three groups were formed based on DNA, ECO 01+ DHJ02, ECO 02+ DHJ01, and ECO 03. The characters that contributed over 50% separation were for wings: wing length, anal margin length, and distance from the band to the outer margin; for male genitalia, angle of the integument, uncus, and the length of the hypandrium, while for females, it was the angle of the anteapophysis and the length of the abdomen. For the analysis of qualitative characters, a tree of maximum parsimony was obtained where 20 characters were informative. We confrmed the existence of three cryptic *Biblis* species in Mexico, two not yet described, and one corresponding to *B. aganisa* (ECO 02), which is sympatric in Oaxaca and Sinaloa (ECO 03) and in the Yucatan Peninsula (ECO 01).

Keywords *Biblis aganisa* · COI · MDH and GAPDH nuclear genes · Yucatan Peninsula

Introduction

Identifying species based on the DNA barcode has proven to help distinguish and discover new species (Blaxter [2003](#page-9-0); Hallwachs et al. [2008](#page-10-0); Janzen et al. [2017](#page-10-1); Gaytán et al. [2020](#page-10-2); D'ercole et al. [2021\)](#page-10-3). The premise is that sequence diversity within a short, standardized segment of the mitochondrial genome (mtDNA) can provide a unique biological barcode, allowing species-level identifcations; it presents a rapid mutation rate, a signifcant variation in sequences between diferent

Edited by André VL Freitas

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 \boxtimes Carmen Pozo cpozo@ecosur.mx species, and a low variation within the same species (Hebert et al. [2003;](#page-10-4) Hajibabaei et al. [2006](#page-10-5)). A barcode system analyzes lineages whose species may present genetic divergence (generally greater than 2% in vertebrates and 3% in Lepidoptera: Hebert et al. [2003;](#page-10-4) Sperling [2003\)](#page-11-0). However, this is not a substitute for a complete taxonomic study, and the use of barcodes must be combined with morphological and ecological data for the results to be decisive for the fnal recognition of the species (Hebert et al. [2004b;](#page-10-6) Sperling [2003](#page-11-0)).

Diferent studies have used the barcode in the Insecta class and have uncovered many cryptic species (e.g., Hallwachs et al. [2008;](#page-10-0) Seraphim et al. [2014](#page-11-1); Nieukerken et al. [2012](#page-11-2); Kim et al. [2020;](#page-10-7) Moraes et al. [2021](#page-11-3)). Cryptic species are morphologically similar taxa determined under a single name (Bickford et al. [2007;](#page-9-1) Gill et al. [2016](#page-10-8)). They may difer in biological attributes, such as host specifcity, mating system, and susceptibility to diferent parasitoids, among others (Hebert et al. [2004a;](#page-10-9) Bickford et al. [2007](#page-9-1); Pfenninger and Schwenk [2007;](#page-11-4) Trontelj and Fier [2009\)](#page-11-5).

An excellent example of the usefulness of the COI, as a tool in molecular systematics, is in the genus *Hermeuptychya* Fabricius. The frst publication with the use of this

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gene (Seraphim et al. [2014\)](#page-11-1) revealed the unknown cryptic diversity within this genus and led to the discovery of new and corroborated species with morphological analysis of genitalia (Cong and Grishin [2014;](#page-10-10) Cong et al. [2021](#page-10-11); Nakahara et al. [2017\)](#page-11-6). This method has also been used to detect cryptic species of *Astraptes fulgerator* Walch (Hebert et al. [2004a](#page-10-9)) but not enough evidence has been presented to confrm this (Brower [2006](#page-9-2)). In another study, Janzen et al. [\(2017](#page-10-1)) confrmed the existence of cryptic species within the species previously known as *Udranomia kikkawai* Weeks (Hesperiidae), identifying two more species using complete genome sequencing and barcode (COI). These species presented subtle diference in adult color variation, size, ecological distribution, and natural history.

Jasso-Martínez et al. [\(2016](#page-10-12)) solved the taxonomic problem regarding the number of species within the Mexican species of the *Enantia jethys* (Boisduval) complex (Pieridae), using COI as a molecular character. They confrmed the three species reported in the literature and a fourth potential cryptic species not yet described. Later Jasso-Martínez et al. [\(2018](#page-10-13)) demonstrated hybridization events in three species of the genus *Enantia* that occur in partial sympatry using diferent genes. Burns et al. ([2008](#page-9-3)) analyzed a complex of cryptic species of the genus *Perichares* Scudder (Hesperiidae: Hesperiinae) from northwestern Costa Rica based on genetic information and their relationship with host plant. They found that information on the host plants of this genus constitutes an ecological characteristic which helps distinguish female oviposition preferences of each species.

The genus *Biblis* Fabricius, 1807 (Lepidoptera: Nymphalidae), is monotypic and presents fve described subspecies in the Americas and one putative subspecies from Peru (Lamas [2004\)](#page-11-7). The type localities of these subspecies are the following: *Biblis hyperia hyperia* Cramer from St. Thomas, "Indies Occidentales"; *Biblis hyperia laticlavia* Thieme from Rio Napo in Ecuador; *Biblis hyperia nectanabis* Fruhstorfer from Rio Grande do Sul, Brazil; *Biblis hyperia pacifca* Hall from Huigra, Ecuador; *Biblis hyperia aganisa* Boisduval from "Java" (although Godman and Salvin ([1879\)](#page-10-14) mention that the locality of this species is an error and that the type specimen originates from Mexico, without stating the specifc locality), and fnally, *Biblis hyperia* nov. ssp., from Peru (Lamas [2004](#page-11-7)).

Initially, *Biblis aganisa* Boisduval, was described as a species, then changed to a subspecies of *B. hyperia* with-out sufficient evidence to justify this change (Lamas [2004](#page-11-7)). The same process occurred with *B. laticlavia*, described as a species easily recognized by its broad submarginal band of the posterior wing on the dorsal and ventral view and the marked diference in the size of each of the spots that make up this band, very diferent to other species of this group. There are also considerable diferences in the male genitalia therefore could be considered a good species. Two more members, *B*. *h*. *pacifca*, and *B. h. nectanabis*, were described as subspecies and synonymized of *B. hyperia*, with very subtle diferences in the width and coloration of the submarginal band.

Prado et al. ([2011\)](#page-11-8) examined the diversity of larvae and adults of the Nymphalidae (Papilionoidea) from the Yucatan Peninsula using Cytochrome C Oxidase I (COI), including *B. hyperia*, having shown divergence percentages above 3% in the sequences in individuals from the same geographic area, suggesting the presence of new species. *B. hyperia* was split into two separate groups with an average divergence of 4.6% and were named with a provisional OTU: *Biblis hyperia* ECO 01 and *Biblis hyperia* ECO 02.

However, no detailed studies include molecular characters combined with morphological data to verify the status of subspecies of *B. hyperia*. Recently, Zhang et al. ([2021\)](#page-12-0) proposed *B. aganisa* as a valid species and will classifed as a distinct species from *B. hyperia*. However, morphological data are not included, and the only specimen used representing *B*. *aganisa* does not originate from Mexico (country where type specimen was collected). According to this study, the genus *Biblis* would be composed of two subgenera *Biblis* and *Vila* Kirby, 1871, with a separation percentage of 7% (46 bp) between both subgenera and 4.6% (30 bp) between species in the subgenera *Biblis*, which should be considered non-monotypic since it would be composed of the species of each of these subgenera. In the case of *Vila*, three valid species are known; two of these were considered in his study (*Vila azeca* E. Doubleday and *Vila euidiformis* Joicey & Talbot).

A similar case is found in the public database in Gen-Bank/Bold System where Janzen and Hajibabaei ([2009\)](#page-10-15) registered sequences of *Biblis* specimens from Costa Rica, which were assigned to three BINs (BOLD: AAC6005, ABY4876, and AAC0692). BINs are a group of sequence assigned to an operational taxonomic unit (OTU), that often correspond to species (Ratnasingham and Hebert [2013](#page-11-9)). Janzen and Hajibabaei assigned a name to each group of sequences: *Biblis* sp. *hyperia*DHJ01 (*n*=5), *Biblis* sp. *hyperia*DHJ02 ($n=4$), and *Biblis* sp. *hyperiaDHJ03* ($n=14$). Out of a total of four BIN registrations in BoldSystem ([http://](http://www.boldsystems.org/index.php/Public_BarcodeIndexNumber_Home) [www.boldsystems.org/index.php/Public_BarcodeIndexNum](http://www.boldsystems.org/index.php/Public_BarcodeIndexNumber_Home) [ber_Home\)](http://www.boldsystems.org/index.php/Public_BarcodeIndexNumber_Home), two *Biblis* BINs share Mexico and Costa Rica as occurrence points.

According to the 4.6% separation percentage between the two groups of the Yucatan Peninsula in Mexico, there are two possible cryptic species (Prado et al. [2011](#page-11-8)) and maybe three cryptic species in *Biblis* from Costa Rica (according to the OTUS of Janzen and Hajibabaei on GenBank, not yet thoroughly analyzed). The groups from Mexico were determined using only a few specimens (11 specimens: *Biblis hyperia*ECO01 (1) and *Biblis hyperia*ECO 02 (10)) and by a single gene fragment (COI).

The study by Zhang et al. [\(2021\)](#page-12-0) shows that the Mexican species difers from *B. hyperia*; therefore, continuing to consider the groups from Mexico as *B. hyperia,* as used in Prado et al. ([2011](#page-11-8)), would be incorrect. The Mexican species from Oaxaca, *B. aganisa*, which was previously described as a species and now its species status has re-emerged. From here on, we will refer to the *Biblis* of Mexico groups as *B. aganisa* stat. rev*.*

There is no study that includes an analysis of morphological characters and sufficient samples from different sites in Mexico to verify if there are cryptic species named as *B*. *aganisa*. Therefore, the evidence is insufficient to confirm whether two or more cryptic *Biblis* species occur in Mexico. Consequently, it is essential that more detailed morphological and genetic studies are carried out in order to confrm the potential new cryptic species of the genus *Biblis* in the Americas, and specifcally within the Yucatan Peninsula, Mexico.

The present study amplifes the molecular analyzes using the markers mtDNA (COI) and two nuclear genes (MDH and GAPDH). Furthermore, a morphological study verifes the existence of cryptic species in *B. aganisa*. Considering the following questions more specifcally: (1) Are there adult morphological characters that support the presence of two or more cryptic species of *Biblis*? (2) Is there congruence among the groups formed by molecular analysis and by the morphometric analysis of adults? (3) Does the information from points 1 and 2 allow us to identify how many cryptic *Biblis* species occur? And fnally (4) in which geographic areas are these species sympatric.

Methods

Biological Material

A total of 171 specimens of *B. aganisa* collected in the states of Campeche, Quintana Roo, Yucatan, Tabasco, Oaxaca, Queretaro, and Zacatecas and deposited in the Lepidoptera Collection of the Museo de Zoología of ECOSUR-Chetumal (ECOCH-L) and in the Lepidoptera Collection from the Museo de Zoología de la Facultad de Ciencias (MZFC) of the Universidad Nacional Autónoma de México (UNAM) were selected for morphological and genetic analyses. In addition, we downloaded COI gene sequences of the *Biblis* genus in GenBank (Benson et. al [2014](#page-9-4)): eleven sequences used by Prado et al. ([2011](#page-11-8)), 22 sequences from Costa Rica, nine sequences from Argentina, four *Biblis* (two from the subgenera *Biblis* and two from the *Vila*) sequences used by Zhang et al. [\(2021](#page-12-0)), and fnally one from *V. azeca* (GQ864818.1). Five sequences from Nymphalidae species of GenBank were taken as an outgroup (*Colobura annulata* Willmott, Constantino & J. Hall, *Archimestra teleboas* Ménétriés, *Mestra amymone* Ménétriés, *Mestra hersilia* Fabricius, and *Mestra dorcas* Fabricius) for molecular analyses, already used in other systematic studies (Chazot et al. [2020](#page-9-5); Wahlberg et al. [2005a](#page-11-10), [b,](#page-12-1) [2009](#page-12-2)) (complete data in Supplementary Information, excel sheet). To obtain the sequences of COI of *B. hyperia* (NVG-19094E05), *B. aganisa* (NVG-17117F03), *V. azeca* (NVG-19095B04), and *V. eueidiformis* (NVG-19095B05) of the project (PRJNA731937) (Zhang et al. [2021](#page-12-0)), we conducted the following treatment: cleaning the adapters with the Trim using BBDuk tool in Geneious Prime 2020.2.3 (Kearse et al. [2012](#page-10-16)); subsequently, once the readings were cleaned, we created an assembly or COI mapping of these four species with the reference sequence used by Zhang et al. ([2021](#page-12-0)); for later study, the consensus sequences were included in the alignment.

DNA Extraction, Sequencing, and Phylogenetic Analyses

The DNA extraction, amplifcation, and sequencing of the specimens of the *Biblis* from Mexico (*N*=160), Costa Rica $(N=22)$, and Argentina $(N=9)$, and the five outgroups, were made considering the protocols from Hajibabaei et al. [\(2005\)](#page-10-17) and Ivanova et al. ([2006\)](#page-10-18). Sequencing was carried out using COI primers with demonstrated efficacy in butterfly studies (Jasso-Martínez et al. [2016;](#page-10-12) Vodă et al. [2015](#page-11-11)), amplifying a ~ 657-bp fragment: LepR1 (5′-TAAACTTCTGGA TGTCCAAAAAATCA-3′) and LepF1 (5′-ATTCAACCA ATCATAAAGATATTGG-3′). Additionally, two nuclear genes were sequenced (for *N*=17 specimens of *Biblis* from Mexico) that have been efective for butterfy phylogenetic studies (Wahlberg and Wheat [2008;](#page-11-12) Wahlberg et al. [2016\)](#page-12-3) and cryptic species delimitation (Tóth et al. [2014\)](#page-11-13): (1) GAPDH (HybFrigga 5′-TAATACGACTCACTATAGGGA ARGCTGGRGCTGAATATG t-3′ and HybBurre 5′-ATT AACCCTCACTAAAGGGGWTTGAATGTACTTGAT RAG RTC-3′) and (2) MDH (HybMDHF 5′-TAATACGAC TCACTATAGGGGAYATNGCNCCNATGATGGGNGT-3′ and MDHmidR 5′-ATTAACCCTCACTAAAGGGAAYTG NGTRGATGARTGRTTNCC-3′).

A total of four alignments were realized consisting of one for each gene separately (COI, MDH, and GAPDH), and a concatenated alignment $(COI + MDH + GAPDH)$ with 17 Mexican specimens for which we have the three genes sequenced plus one specimen of *B. hyperia* from GenBank (voucher number NW106-3) from Campinas, Sao Paulo, Brazil. All alignments were made in the Geneious Prime 2020.2.3 program (Kearse et al. [2012](#page-10-16)) using the Muscle 3.8.425 option and a refned alignment (Robert [2004](#page-11-14); Edgar and Batzoglou [2006\)](#page-10-19). The Mega X program (Kumar et al. [2018](#page-11-15)) was used for selecting the evolutionary model, which selected the $GTR + G + I$ model with a lower BIC value (Huelsenbeck and Rannala [2004;](#page-10-20) Lecocq et al. [2013](#page-11-16)).

Before running the phylogenetic analysis, we applied two species delimitation methods to evaluate the informative potential of each gene and decide which ones to use for the phylogenetic tree: (1) ABGD (Automatic Barcode Gap Discovery) is an automatic procedure that classifes the sequences based on the width of the barcode, which is observed in the divergence between members within the same species and is always lower between diferent species (Puillandre et al. [2012](#page-11-17)). This analysis was performed with distance matrices according to the Kimura model (K80) and the default parameters; (2) Assemble Species by Automatic Partitioning (ASAP) is similar to ABGD and in both analyses the species hypothesis is indicated thanks to the partitions. In this analysis, distance matrices used the Kimura (K80) ts/tv 2.0 (Puillandre et al. [2021\)](#page-11-18). Both analyses were ran on the online servers (ASAP: [https://bioinfo.mnhn.fr/](https://bioinfo.mnhn.fr/abi/public/asap/) [abi/public/asap/](https://bioinfo.mnhn.fr/abi/public/asap/); ABGD: [https://bioinfo.mnhn.fr/abi/public/](https://bioinfo.mnhn.fr/abi/public/abgd/abgdweb.html) [abgd/abgdweb.html\)](https://bioinfo.mnhn.fr/abi/public/abgd/abgdweb.html).

A phylogenetic tree was constructed using the Bayesian inference (Huelsenbeck and Ronquist [2001](#page-10-21); Holder and Lewis [2003\)](#page-10-22). Trees under the Bayesian inference were constructed for the COI gene using MrBayes (Huelsenbeck and Ronquist [2001](#page-10-21)). Analyses considered a gamma distribution with fve categories and a percentage of invariant sites; MCMC chains ran for 5 million generations, 25% of the trees were discarded (burned), and a consensus tree was calculated. The posterior probabilities summarized in the MrBayes consensus tree were drawn on the clade maximum credibility tree as recommended by García-Sandoval [\(2014\)](#page-10-23) for better visualization. The mean intraspecifc and interspecifc sequence divergences were estimated using the Kimura two-parameter (K2P) distance model (Kimura [1980\)](#page-10-24) in MEGA X (Tamura et al. [2011\)](#page-11-19).

Finally, we constructed a Haplotype network with the alignment of 214 COI sequences based on statistical parsimony implemented in TCS version 1.23 (Clement et al. [2000](#page-10-25)). The haplotypes were classifed with the tcsBU program (Múrias Dos Santos et al. [2016\)](#page-11-20), and later the network was stylized in Photoshop 2020 to improve its presentation.

Morphological Analyses

Based on our results of the genetic analysis, we conducted morphological measurements on 148 specimens (85 males and 63 females) organized into three groups (ECO 01 (*N*=4), ECO 02 (*N*=137), and ECO 03 (*N*=7)). For each specimen of both sexes, quantitative and qualitative characters were taken from the dorsal and the ventral view of the wings and male and female genitalia. For the genitalia analysis, abdomens were removed and placed in labeled glass jars with a 10% sodium hydroxide (KOH) solution for 1 or 2 days (depending on how dry the abdomens were). Genitalia was removed using a stereoscopic microscope, the fat and tissue using forceps, and a fne-tip dissecting needle, and scales were removed with a fne-bristle brush. Clean genitalia (including hypandrium) were placed on concave slides with 70% ethanol or glycerol. Lateral, dorsal, and ventral view photographs were taken using a millimeter scale to obtain later measurements and comparisons (Zubek et al. [2015](#page-12-4)). Image J software was used for the measurements, calibrated with the millimeter scale each time a new photograph was used (Abramoff et al. [2004](#page-9-6); Collins [2007](#page-10-26)).

Quantitative Characters of Wings The following twelve dorsal (D) and ventral (V) measurements were taken with a digital vernier, following the nomenclature of the wings according to Miller [1970](#page-11-21). An alphanumeric key was assigned as described below: wing length (D1), distance from band to disc cell (D2), band distance to outer margin (D3), spot length in cell M3-CuA1 (D4), spot length in Rs-M1 (D5), spot length in cell $CuA2-1A+2A$ (D6), band to disc cell distance (V1), distance from band to outer margin (V2), spot length in cell M3-CuA1 (V3), length of the spot in cell Rs-M1 (V4), spot length in CuA2-1A + 2A (V5), and anal margin length (V6) (Supplementary Information: Fig. 1).

Quantitative Characters of Genitalia The following ten male genitalia measurements were annotated: aedeagus length (EL), valva length (VL), tegument length (TL), uncus length (UL), tegumen angle (TA), tegumen to saccus distance (TSD), uncus angle (UA), saccus length (SL), valva width (VA), and androchondrial patch length (APL) (Supplementary Information: Fig. 2a). Additionally, measurements on the hypandrium were made: length (HL), width (HW), and angle (HA) (Supplementary Information: Fig. 2b). For the female genitalia: abdomen length (AL), width of ostium bursae (A), shortest distance between anteapophysis (B), longest distance between anteapophysis (C), length of anteapophysis (D), height of ostium bursae (E) (Supplementary Information: Fig. 2c), length of copus bursae (F), length of ductus bursae (G), width of papilla analis (H), and angle of anteapophysis (I) (Supplementary Information: Fig. 2d).

Four discriminant analyses were made based on the female and male wings and genitalia looking for groups within *B. aganisa*. Additionally, we performed a similarity percentage analysis (SIMPER) considering Euclidian distances as a dissimilarity measure for female and male wings and genitalia separately, to evaluate the contribution of the diferent measures. These analyses were processed with PAST version 4.03 (Hammer et al. [2001](#page-10-27)).

Qualitative Characters of Wings and Genitalia A total of 23 characters were taken, fve from dorsal view and 18 from ventral view of female and male (Supplementary Information: Figs: 3–5). A total of ffteen characters from male genitalia and one of the head (labial palps) were encoded

561

(Supplementary Information: Figs. 5–9, and Tables 1 and 2). Using the matrix generated from the qualitative characters of wings of male and female, male genitalia, and head of male, a phylogenetic relationship based on maximum parsimony method was constructed using PAUP V4.0a (Swofford [2001](#page-11-22)).

Geographical Distribution

Using the coordinates of the 160 Mexican specimens belonging to the three groups (ECO 01 (4), ECO 02 (149), ECO 03 (7)), a distributional map was elaborate with the ArcGis program version 10.2.1

Results

Species Delimitation

Analyses ran for each nuclear gene did not allow the delimitation of species inside the *Biblis* genus while regarding COI gene, we obtained an adequate species delimitation. Consequently, all further analyses were run using exclusively COI sequences. The best partitions generated by ASAP analysis with COI (including outgroup) consider five, nine, 11, and 12 species delimitation which correspond to one, three, four, and five possible species delimitation within the *Biblis* genus (Fig. [1](#page-4-0)). Additionally, other ASAP partitions were less probable (two, three, four, 13, and 60 with outgroups), and therefore were not considered. Examination of the ASAP score (the lower the score, the best partition) revealed that the optimal partition for species delimitation in *Biblis* is three species (Fig. [1](#page-4-0); ASAP score 3). This implies that *B. hyperia*, *B. nectanabis*, and the group including ECO1 are only one species. We discarded this hypothesis because each of these species' group presented morphological differences at the level of the extern red band ([http://butterfliesofamerica.com/L/t/Biblis_hyperia_a.](http://butterfliesofamerica.com/L/t/Biblis_hyperia_a.htm) [htm\)](http://butterfliesofamerica.com/L/t/Biblis_hyperia_a.htm); thus, the next best partition is five internal species for the *Bilbis* genus (ASAP score = 5) which coincides with our phylogenetic analyses. With ABGD, four of the same groups were recovered with recursive partitioning $(p = 0.004642)$, while ECO $01 + NW106-3$ (Mexico-Brazil) separated from DHJ02 (Costa Rica), which also presents a difference at the level of the red band within the group, and that should be evaluated at a later stage at the genitalia level, as with the groups in Mexico (Fig. [1](#page-4-0)).

Phylogenetic Analyses

The Bayesian phylogeny constructed using COI gene dataset (Fig. [1](#page-4-0); 214 sequences: 22 from Costa Rica, nine from Argentina, 171 from Mexico, one from EU, and one from the British Virgin Islands: Guana Island) revealed two well-supported sister clades: (1) the first clade is confirmed by the majority of Costa Rica samples (DHJ03, DHJ02), samples from Argentina (*B. nectanabis*), and four samples from Mexico belonging to the ECO 01. The only sample of *B. hyperia* was combined with DHJ03. Remarkably, these four Mexican samples are pooled with some Costa Rica samples (DHJ02) and they are sister to Argentina samples; (2) a second clade consists of the remaining Mexican samples, with very good separation between the ECO 03 and ECO 02 samples. Some Costa Rica samples (DHJ01) and *B. aganisa* are pooled with ECO 02 samples from Mexico in this clade.

Fig. 1 Bayesian phylogenetic relationship based on COI gene for the species of *Biblis* Fabricius (Nymphalidae: Biblidinae). The numbers in the clades correspond to the posterior probability values supporting the clade. The colors of the branches indicate species, and the same color on the right side of the bars present the results of the two species delimitation methods. The numbers below the bars indicate the number of species or groups recovered

The results of percentages of identity among the species confirm the results of phylogeny (Table [1\)](#page-5-0). The samples from Mexico (ECO 01), Costa Rica, and Brazil present very high values (from 99.22 to 99.81%); in contrast, ECO 01 present lower values with Costa Rica (DHJ03; 97.48%), with Argentina (*B. nectanabis*) 97.67%, and with *B. hyperia* from Guana Island (95.54%). The clade including the two other groups from Mexico (ECO 02 and ECO 03) presents a percentage of identity of 96.12%. Our samples identified as ECO 02 (Mexico) present maximum values with samples from Costa Rica (DHJ01), the sample from Texas, and *B. aganisa*.

The genetic divergence values (Table [2\)](#page-5-1) were obtained in the groups of the first clade, comprised Costa Rica, Argentina, Guana Island, and Mexico. In the group composed by ECO 01, Costa Rica (DHJ02), and Brazil (NW106-3), the mean intraspecific distance was 0.42%. However, between this group and Costa Rica (DHJ03), the interspecific distance was 2.80%, and 2.44%. with Argentina (*B. nectanabis*). The second clade, composed primarily of samples from Mexico (ECO 02), some from Costa Rica (DHJ01), and a sample of *B. aganisa* from Hidalgo (Texas, USA), presents an intraspecific divergence of 0.22%. This clade presented 4.12% of divergence with ECO 03. The most considerable divergence between the groups from Mexico (ECO 01 and ECO 03) was 5.85%.

The haplotype network analysis of 214 specimens including outgroups and all specimens from *Biblis* resulted in the formation of five haplogroups (Fig. [2](#page-6-0)). In Fig. [2,](#page-6-0) the blue dotted circle is composed of specimens from Mexico (ECO 01), Costa Rica (DHJ02), and Brazil (NW106-3). Green haplotypes regroup specimens from Mexico (ECO 02), Costa Rica (DHJ01), and USA (*B. aganisa*). The specimens from Mexico (ECO 03) are shown in pink, while specimens from Argentina (*B. nectanabis*) are in yellow, and finally the haplogroup formed by individuals from Costa Rica (DHJ03) and the British Virgin Islands (*B. hyperia*) are in pistachio green.

Table 1 Means of identity with COI gene. Numbers in black are groups that present similarity above 98% in their bases and that are within the same clade

	Biblis. hyperia* DHJ03		Biblis nectanabis ECO 01 NW106-3 DHJ02 ECO 03 Biblis aganisa* ECO 02 DHJ01							
Biblis hyperia*										
DHJ03	99.03									
Biblis nectanabis	97.09	97.29								
ECO ₀₁	95.54	97.48	97.67							
NW106-3	96.90	97.29	97.48	99.81	$\overline{}$					
DHJ02	96.32	96.71	97.67	99.22	99.42					
ECO ₀₃	94.57	94.38	94.38	94.38	94.19	94.38	$\overline{}$			
Biblis aganisa*	95.35	95.54	96.12	95.54	95.35	95.54	96.12			
ECO ₀₂	95.35	95.54	96.12	95.54	95.35	95.54	96.12	100		
DHJ01	95.35	95.54	96.12	95.54	95.35	95.54	96.12	100	100	

* With only one sequence taken from Zhang et al. [2021.](#page-12-0) *Biblis hyperia* (Guana Island); *B*. *aganisa* (USA); *B*. *nectabis* (Argentina); DHJ01, DHJ02, DHJ03 (Costa Rica); NW106-3 (Brazil); ECO 01, ECO 02, and ECO 03 (Mexico)

Table 2 Distance matrix of the Kimura model (COI) shown under the diagonal, all above 2%. The percent in parentheses correspond to mean values of intraspecifc sequence divergence

Group	Biblis hyperia*/ DHJ03 (0.52%)	Biblis aganisa*/ECO 02/DHJ01 (0.22%)	ECO 01/NW106-3/ DHJ02 (0.42%)	Biblis nectanabis (0.57%)	ECO ₀₃ (0.06%)
Biblis hyperia*/DHJ03					
Biblis aganisa*/ECO 02/DHJ01	4.67%				
ECO 01/NW106-3/DHJ02	2.80%	4.55%			
Biblis nectanabis	2.67%	3.88%	2.44%	-	
ECO ₀₃	5.90%	4.12%	5.85%	5.78%	

* With only one sequence taken from Zhang et al. [2021.](#page-12-0) *Biblis hyperia* (Guana Island); *B*. *aganisa* (USA); *B*. *nectabis* (Argentina); DHJ01, DHJ02, DHJ03 (Costa Rica); NW106-3 (Brazil); ECO 01, ECO 02, and ECO 03 (Mexico)

Fig. 2 Haplotype network based on COI barcodes of *Biblis* and outgroup that provide evidence of the 5 haplogroups of *Biblis* that correspond to the phylogenetic tree. The size of the colored circle is proportional to the number of samples represented. Small uncolored circles represent unsampled haplotypes or mutational step diference

Morphological Analyses

Quantitative Characters

The discriminant analysis based on male wings' characters separated the samples into three groups (Fig. [3A](#page-6-1); ECO 01 in blue, ECO 02 in green, and ECO 03 in fuchsia). ECO 01 is completely separated from the two remaining groups, while a slight overlap is noted between ECO 02 and ECO 03. The SIMPER analysis based on male wing measurements (Fig. 10A, Supplementary Information) indicates that three variables could explain 50% of the separation between groups: wing length (D1), anal margin length (V6), and band to outer margin (V2).

The discriminant analysis, considering the characters of females' wings (Fig. [3B\)](#page-6-1), also resulted in the separation of three groups. In this case, ECO 03 (fuchsia) is separate from the two other groups, while a slight overlap is observed

Fig. 3 Discriminant analysis with morphometric measurements of the wings and genitalia of the male and female. **A** Male wings, **B** female wings, **C** genitalia in male, and **D** genitalia in female. Filled blue

squares: *Biblis aganisa* ECO 01, flled green triangles: *Biblis aganisa,* ECO 02, and pink flled diamonds: *Biblis aganisa* ECO 03

between ECO 02 (green) and ECO 01 (blue). The SIMPER analysis (Fig. 10B Supplementary Information) revealed that the same characters of wing females explain 50% of the separation between groups: wing length (D1), anal margin length (V6), and band to outer margin (V2).

The discriminant analysis based on male genitalia characters (Fig. $3C$) results in the formation of three groups (ECO 01, ECO 02, and ECO 03) without any overlap. The SIMPER analysis (Fig. 10C, Supplementary Information) indicates that only two characters explain 90% of the separation of the three groups: tegumen angle (TA) and uncus angle (UA).

The discriminant analysis based on female genitalia characters (Fig. [3D\)](#page-6-1) also results in the separation of three groups (ECO 01, ECO 02, and ECO 03) without any overlap. The SIMPER analysis (Fig. 3D, Supplementary Information) indicates that only two characters explain 90% of the separation of the three groups: angle of anteapophysis (I) and length of abdomen (AL).

Qualitative Characters

Maximum parsimony analysis of the qualitative characters of the wings, head (labial palps), and the male genitalia produced a tree with three terminal branches: ECO 01, ECO 02, and ECO 03, and two terminal branches in a diferent clade: *B. hyperia* and *B*. *h*. *laticlavia* (Fig. 11, Supplementary information). We use a matrix of 39 characters (23 from the wings, 1 from the head, and 15 from male genitalia; Table 1 in the Supplementary information). This character matrix resulted in a single, more parsimonious tree of 55 steps with a consistency index (CI) equal to 0.8182 and a retention index (RI) equal to 0.6154. The informative characters were band shape, spot color in cell $Sc + R1$, spines on the distal part of the hypandrium, proximal portion of the uncus in lateral view, distal portion of the uncus bifurcated in dorsal view, proximal portion of the uncus in dorsal view, tegumen in dorsal view, saccus in lateral view (size and shape), and valvae in ventral view (size) (Table 1 and Figs. 4–9, Supplementary Information). Of the characters with the best CI ft and apomorphies, ECO 03 presents a postdiscal red dot in cell CuA1 of the anterior wing in dorsal view. The point size in cell $Sc + R1$ is not vestigial, and the color of this point is not totally red (it contains white scales). The basal points do not present white scales. The proximal portion of the hypandrium is slightly narrow in the center. The tegumen in the lateral view is slightly curved, and the proximal portion of the valvae in the lateral view is fattened. ECO 02, the thin red band pattern, is present in ventral view of the posterior wing (as in the type specimen of *B. aganisa*), and the spot size in the cell $Sc + R1$ can be medium and white. The saccus in lateral view can be straight or curved. ECO 01 does not present a clear broadband pattern, the cilia on the labial palps in the middle segment in the inner part only with few white cilia. The distal portion of the hypandrium is semi-rectangular, the valvae in ventral view are slender, and the distal portion of the valvae in lateral view is wider than the width of the valvae. A comparison of these diagnostic characters of the groups can be seen in more detail in Table 3 (Supplementary Information).

Geographical Distribution

The map shows the records of the three groups, i.e., ECO 01, ECO 02, and ECO 03, in México (Fig. [4](#page-8-0)) highlighting a broad distribution for *B. aganisa* ECO 02 (green circles), predominantly in southern and southeastern Mexico with records in the Yucatán Peninsula (Quintana Roo, Yucatán and Campeche states), Chiapas, and Oaxaca and only two record from Queretaro and Zacatecas. Meanwhile, the records for the *B. aganisa* ECO 01 (blue circles) have a restricted distribution in the southern region of the Yucatan Peninsula (Quintana Roo and Campeche states), near the border with Belize and Guatemala. Finally, *B. aganisa* ECO 03 (pink circles) presents a restricted distribution in the north of Oaxaca and Sinaloa in northern Mexico.

The map indicates several sympatric areas between groups. The ECO 01 and ECO 02 groups present a sympatric distribution in the southern part of the Yucatan Peninsula, more precisely in the Calakmul region of Campeche. Furthermore, ECO 02 and ECO 03 are in sympatry in the north of Oaxaca (Fig. [4](#page-8-0)).

Discussion

Previous studies with molecular traits using COI in *B. hyperia* have documented the separation of *B. hyperia*, establishing the possibility that it is a complex of cryptic species. Prado et al. (2011) suggested the division into two species when analyzing the specimens in the Yucatan Peninsula. Later, Zhang et al. (2021) (2021) recognized that the typical subspecies is diferent at a specifc level from the taxon of north and central America, renaming it *B. aganisa*. Our study confrms the result of Zhang et al. [\(2021](#page-12-0)) when comparing all our samples with the *B. aganisa* and *B. hyperia* sequences, which are grouped into diferent clades. Moreover, all the samples of the ECO 02 group are the same as *B. aganisa*. Our research confrms previous fndings by Prado et al. [\(2011](#page-11-8)) by studying a more signifcant number of samples; in addition, a third lineage (ECO 03) is recognized (Fig. [1\)](#page-4-0). Furthermore, in GenBank/BoldSystem, Janzen and Hajibabaei [\(2009](#page-10-15)) recognize three lineages from Costa Rica (DHJ01, DHJ02, and DHJ03), two of which are genetically close to the lineages of Mexico (ECO 01 with DHJ02 and ECO 02 with DHJ01) (Fig. [1\)](#page-4-0). The three lineages discovered

Fig. 4 Location of *Biblis* groups: blue circles for ECO 01, green circles for ECO 02, and pink circles for ECO 03. The areas where the groups overlap are zones of sympatry

with specimens from Mexico are strongly supported by the Bayesian analysis using COI. The genetic distances according to the Kimura model (Kimura [1980](#page-10-24)) between groups or species, found both by Prado et al. [\(2011\)](#page-11-8) and by Zhang et al. ([2021](#page-12-0)), established that there is a valid diference at a specifc level, having a threshold above 4% with COI for *Biblis* and above 3% for Lepidoptera as suggested by Hebert et al. [\(2003\)](#page-10-4). In our study, genetic distances between ECO 01 and ECO 02 were similar to Prado et al. [\(2011](#page-11-8)). In addition, the new group (ECO 03) presents a distance of 5.87% from ECO 01 and 4.12% from ECO 02; this is not surprising since their biogeographic areas (Fig. [4](#page-8-0)) are diferent from their evolutionary history (Morrone [2019\)](#page-11-23). Additionally, the results of the haplotype network, ASAP, and ABGD analysis corroborated the groups formed based on the genetic distances and tree topology.

The use of discriminant analysis in other groups of Lepidoptera using measurements of parts of the wings and structures of the male genitalia has been previously used to delimit species (Kolev [2005;](#page-11-24) Hernández-Roldán and Munguira [2008;](#page-10-28) Prieto et al. [2009;](#page-11-25) Núñez et al. [2021](#page-11-26)) and even to confrm cryptic species (Dincâ et al. [2011\)](#page-10-29). According to the discriminant analyses in which linear measurements were used in wing characters (Fig. [3A](#page-6-1), [B](#page-6-1)), the results were consistent with the Bayesian tree. This method was efective for species discrimination, as there was no overlap between the three groups. The male genitalia of the three groups are similar, but when applying this analysis, the diference is clear, as there is no overlap between the groups (Fig. [3C,](#page-6-1) [D](#page-6-1)). Hypandrium examination has been helpful for reliable diagnosis in the delimitation of species within Biblidinae, despite only shape and proportions being used (Jenkins [1990](#page-10-30); Leite et al. [2017](#page-11-27); Zubek et al. [2015](#page-12-4)). In our study, the length and angle of the hypandrium were two fundamental measures to support the separation. The impossibility of differentiation by a simple observation means that these three species must be considerate cryptic species.

Phylogenetic studies where molecular and morphological characters are included are crucial to resolve relationships between species, particularly in unresolved nodes that become more robust when more characters are added to the analysis (Wahlberg and Nylin [2003\)](#page-11-28). Moreover, if there is a good selection of characters, the results may be consistent with genetic analyzes (Shi et al. [2015](#page-11-29)). The cladistics analysis where we included 39 morphological characters (qualitative characteristics) of wings and genitalia of a single more parsimonious tree produced results that were consistent with the molecular phylogeny, since the same relationships were recovered between groups (ECO 01, ECO 02+ECO 03).

Some of the characters that contribute to the formation of these groups, for example in the ECO 02 group, which is the most variable in its coloring pattern, present an exclusive characteristic that coincides with that presented in the type specimen of *B. aganisa* described by Boisduval of "Java"

in 1836, specifcally that in the ventral view, the submarginal band is thin and red scales predominate. However, this unique feature is rare since it was present in only 8% of all individuals. In addition, this group presents another variant of the submarginal band, which is wide clear, with a few red scales, which resembles the type specimen of *Didonis pasira* Doubleday, which is currently considered synonymous with *B*. *aganisa*. Therefore, we suggest that ECO 02 is the group that represents *B*. *aganisa*. However, this form of the submarginal band is not exclusive to the group since it is also observed in individuals of ECO 03.

The ECO 01 group shares the pattern of the intermediate submarginal band with both groups (Fig. 3, Supplementary Information); however, the pinkish scales are absent in the submarginal area of the anterior wing in ventral view, a characteristic that is also shared with the other two groups. We also observed a red spot in cell CuA1 of the forewings in dorsal view within the ECO 03 group; however, this is not present in all individuals. When examining the qualitative characteristics of the three groups individually, we consider that it is difficult to identify any group since they share several characteristics with the other groups; therefore, it is not very reliable to use only qualitative characters to discriminate species.

The data on the distributions of the sequenced specimens was necessary to detect sympatry areas and delimitation of the groups. One specimen from Campinas, Sao Paulo, Brazil (NW106-3), is related to ECO 01 from Mexico. Nevertheless, the shape, size, and position of the red band on the hindwing is diferent in Mexico, Costa Rica, and Brazil. So, we suggest that there are three subspecies with the following distribution range: one subspecies is widely distributed from Mexico to Nicaragua (ECO 01), the other represented by DHJ02 from Nicaragua to Brazil, and the last one represented by NW106-3 in Brazil. Our results on the actual distributions supported ECO 01 as a diferent group and more distant from ECO 03, particularly as it did not present areas of sympatry and was in a diferent historic biogeographical area (Fig. [4\)](#page-8-0). Similarly, there was no overlap of the groups using quantitative characteristics in both wings and genitalia (Fig. [3\)](#page-6-1). There are other groups of Lepidoptera that do not extend their distribution beyond the Yucatan Peninsula (Llorente-Bousquets et al. [2006](#page-11-30)), such as *Heraclides rogeri rogeri* (Boisduval) and *Hamadryas julitta* Fruhstorfer, while ECO 02 and ECO 03 have wider distributions in Mexico. The distribution of ECO 02 is probably towards the Gulf slope. ECO 03 exists towards the Mexican Pacifc slope, with the zone of sympatry for both groups in northern Oaxaca, as demonstrated in this study and for other groups of Lepidoptera (e.g., Lasaia, Arellano-Covarrubias et al. [2019](#page-9-7)).

This paper provides evidence that that the genus *Biblis* is more diverse in Mexico than was supposed in recent times.

We also confirm the proposal of Zhang et al. (2021) to reinstall the Boisduval nomination of *Biblis aganisa* and report the existence of two new species in Mexico in sympatry with *B. aganisa*.

To conclude, we consider these species as cryptic species.

Supplementary Information The online version contains supplementary material available at<https://doi.org/10.1007/s13744-022-00969-4>.

Acknowledgements We thank Alma Estrella García Morales and Manuel Elías Gutiérrez from Node of MEXBOL from ECOSUR for their help in processing and sending the samples for sequencing; to Humberto Bahena Basave for his help in taking photographs to make part of the fgures; to Holger Weissenberger from ECOSUR Department of Observation and Study of the Earth, the Atmosphere and the Ocean for his help in making the map; and to the Consejo Nacional de Ciencia y Tecnología (CONACYT) for the scholarship awarded to the frst author.

Author Contribution All authors contributed to the study conception and design. Material preparation, data collection, and analysis were performed by Hugo Álvarez García and Carmen Pozo. The frst draft of the manuscript was written by Hugo Álvarez García, and all authors commented on previous versions of the manuscript. All authors read and approved the fnal manuscript.

Declarations

Conflict of Interest The authors declare no competing interests.

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