BIOLOGICAL CONTROL

Parasitism Performance of Tetrastichus brontispae Ferriere over the Coconut Hispine Beetle, Brontispa longissima (Gestro)

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Keywords **Abstract**

Biological control, choice test, host, parasitoid

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In this study, the effect of host density, host, and parasitoid ages in choice and no-choice tests on the parasitism performance of Tetrastichus brontispae Ferriere, one of the major parasitoid of Brontispa longissima (Gestro), was investigated in the laboratory. The results revealed that an increased host density resulted in no increased parasitism of B. longissima by T. brontispae; the optimal host density was three host pupae per parasitoid when considering the costs for mass rearing. Moreover, parasitoid age was quite crucial for effective parasitism and affected the emergence rate. Although 2-h to 4-day-old parasitoids successfully parasitized the host pupae, younger parasitoids (within 2-day-old) presented higher parasitism capacity than older parasitoids. More importantly, both choice and no-choice tests confirmed that all host stages tested from 2-h to 4-day-old were suitable for T. brontispae parasitization, although 2-h to 2-day-old hosts were preferred. We also demonstrated that sex ratio, emergence rate, and egg to adult developmental time were not influenced by host density, parasitoid, and host age in both choice and no-choice tests. Our data will allow for more accurate prediction and interpretation on the parasitization by T. brontispae, supporting mass-production initiatives and mass release in programs of B. longissima.

Introduction

The coconut hispine beetle, Brontispa longissima (Gestro) (Coleoptera: Chrysomelidae), is native to Indonesia and Papua New Guinea and is one of the most invasive and economically threatening insects in the region, where it is a major pest on Palmae plants (Chiu & Chien [1985,](#page-6-0) Waterhouse [1987\)](#page-6-0). The species was accidentally introduced into Southeast Asia and has since dispersed throughout the area, including the countries of Thailand, Vietnam, and the Philippines, and the Hainan Province of China (Chiu & Chien [1985](#page-6-0), Waterhouse [1987,](#page-6-0) Nakamura et al [2006](#page-6-0), [2008,](#page-6-0) Lu et al [2008,](#page-6-0) Chen et al [2010](#page-6-0), Liu et al [2014\)](#page-6-0). This particular beetle damages plants mainly by feeding on their leaves, resulting in losses to the coconut and tropical tourism industry (Nakamura et al [2006](#page-6-0), [2008](#page-6-0), Lu et al [2008](#page-6-0), Chen et al

[2010](#page-6-0), Liu et al [2014\)](#page-6-0). Therefore, the beetle's invasion and infestations have raised great concern internationally, and the species was listed in the Global Invasive Species Database in 2010 (Nguyen et al [2012,](#page-6-0) Liu et al [2014](#page-6-0)). In China, the pest was first discovered in Haikou (Hainan Province) in 2002, after which it quickly spread throughout the island to become the most serious threat to the tourism and coconut industry of Hainan.

In order to control B. longissima, chemical pesticides have been frequently applied in most of the countries where the pest is located. However, insecticides are always deemed undesirable and impracticable, primarily because the individuals who apply it face great risk when scaling tall coconut trees and because its application is highly likely to lead to insect resistance (Nakamura et al [2008](#page-6-0)). Previously, an entomopathogenic fungus was used for controlling B. longissima on young coconut seedlings (Nakamura et al [2006\)](#page-6-0), but its application was difficult and impracticable in areas with tall coconut plants.

Considering the cost-benefit and sustainability of biological control, two promising biological control agents—Asecodes hispinarum Boucek (Hymenoptera: Eulophidae) and Tetrastichus brontispae Ferrière (Hymenoptera: Eulophidae) have been introduced to control B. longissima (Waterhouse [1987](#page-6-0), Voegele [1989](#page-6-0), Lu et al [2008](#page-6-0)). Previous studies have demonstrated that these two parasitoids were capable of controlling B. longissima because of their high host specificity (Lu et al [2008](#page-6-0), Chen et al [2010](#page-6-0), Ichiki et al [2011,](#page-6-0) Nguyen et al [2012](#page-6-0), Liu et al [2014](#page-6-0)). Successful biological control of B. longissima was achieved in the countries of Samoa, Sulawesi, Tahiti, the Solomon Islands, and Taiwan (Chiu & Chien [1985](#page-6-0), Voegele [1989,](#page-6-0) Nguyen et al [2012](#page-6-0)). Hence, environmentally friendly biological control techniques seem to be feasible and reasonable.

Tetrastichus brontispae is one of the most promising endoparasitoids of B. longissima and has been successfully introduced into several countries, primarily because of its ability to locate hosts (i.e., parasitization specificity) (Chen et al [2010,](#page-6-0) Ichiki et al [2011,](#page-6-0) Nguyen et al [2012\)](#page-6-0). Existing details on the biological traits of T. brontispae associated with different factors have been well-documented in recent years (Ma et al [2006,](#page-6-0) Zhou et al [2006](#page-6-0), Tang et al [2009](#page-6-0), Chen et al [2010,](#page-6-0) Ichiki et al [2011](#page-6-0), Nguyen et al [2012](#page-6-0), Liu et al [2014](#page-6-0)), providing useful information for its mass rearing in the laboratory before mass release.

Determining the characteristics of the parasitoid-host relationship is also critical and essential for biological control, and such works have been widely conducted on other parasitoid wasps (Duan & Messing [1999,](#page-6-0) Hill & Hoy [2003,](#page-6-0) Haye et al [2005,](#page-6-0) Sheppard et al [2005](#page-6-0), Ayvaz et al [2008\)](#page-6-0). In this regard, Chen et al [\(2010](#page-6-0)) reported how temperature, relative humidity, photoperiod, and food sources affect the parasitism rate of T. brontispae; Nguyen et al [\(2012\)](#page-6-0) collected limited data to describe the effect of host age and parasitoid age on parasitization by T. brontispae. In addition, Tang et al [\(2014a](#page-6-0), [b](#page-6-0)) studied the parasitism of T. brontispae on another host, Octodonta nipae. Our recently published works mainly investigated the biological characteristics and parasitism of T. brontispae on B. longissima in relation to different temperatures and cold storage (Liu et al [2014\)](#page-6-0). However, little information exists on the particular characteristics of parasitism of T. brontispae on B. longissima, and no study has examined how host density and choice and no-choice tests influence the parasitism performance of T. brontispae.

Parasitism specificity testing within the confines of the controlled conditions of a laboratory is the most common way to obtain data on the parasitoid-host relationship; results may then be extrapolated to the natural environment. Hence, the main objective of this study was to investigate the effect of host density and host and parasitoid age on the

characteristics of the parasitoid-host relationship in relation to T. brontispae parasitism on B. longissima. Furthermore, in an effort to improve the basic knowledge and accurate assessment of parasitism specificity, choice and non-choice tests were conducted to determine the parasitism performance of T. brontispae on B. longissima.

Material and Methods

Host insect cultures

Adult B. longissima were originally collected in 2010 from Haikou (Hainan Province, China; 20°1′N, 110°19′E) and later reared in a laboratory to establish colonies. Rearing conditions consisted of a temperature of $26 \pm 1^{\circ}$ C, $75 \pm 5\%$ RH, and a 12-h light and 12 h of scotophase. Approximately 300 B. longissima adults were used for establishing the initial cultures. Males and females were kept together, to allow for mating, in plastic containers (17 length \times 11 width \times 5 height, in cm) with a screened window in the lid. Feral individuals were added to the colonies twice a year to maintain genetic diversity. B. longissima were continuously reared on fresh coconut leaves, and a piece of tissue paper soaked in a 10% honey solution (weight/volume) was fixed on the wall of the boxes. When offspring larvae in the same generation emerged, they were transferred into other boxes. Afterwards, newly emerged pupae were collected daily for maintaining the parasitoids and for use in the experiment.

Parasitoid insect cultures

The colony of T. brontispae was originally obtained from Taiwan in 2012 and was maintained using 48-h-old host pupae. Both males and females were kept together in Petri dishes (6.0 mm in diameter, 2.5 mm in height), fed with 10% honey solution soaked tissue paper, and maintained at the laboratory conditions as described above. For mass rearing, approximately 200 one-day-old pupae were placed in each box together with 400 adult wasps. The boxes were removed after the adult wasps died. The newly emerged adults were collected daily and used in the experiments.

Effects of host density on parasitism

One-day-old female and male parasitoid wasps were kept together for 24 h to ensure mating. Next, mated female parasitoid were randomly selected and one was introduced per tube (2 cm in diameter), with a 10% honey solution soaked piece of tissue paper. Two-hour-old B. longissima host pupae were then selected and divided into six groups (treatments) based on tested host densities of 2, 3, 4, 5, 6, and 7 individuals. For each treatment, host pupae were

placed into the tubes together with the female parasitoid and maintained in the cabinets at conditions similar to those described above. After 24 h of parasitism activity, each female wasp was removed from the tubes. After the fifth day, the pupae were dissected to confirm parasitism status, and number of parasitized pupae counted and recorded. Fifty replicates were tested for each host density treatment. Based on observations from previous experiments and the present experiment, we could distinguish the parasitized pupae from the non-parasitized by color changes. Once parasitized, the color of the cuticle changes to a deep blackbrown (5-day-old pupae), which gets darker with time. In contrast, the non-parasitized pupae remain pale yellow until emergence.

Effects of parasitoid age on parasitism

Female and male parasitoids emerged the same day and were collected and kept together for 24 h to permit mating. Afterwards, 1-, 2-, 3-, and 4-day-old female parasitoids were randomly selected and introduced into tubes (2 cm in diameter, 0.5 cm in length) with a 10% honey solution provided. Fifty female parasitoid wasps replicates were tested for each parasitoid age. Two-hour-old B. longissima pupae in groups of five were placed in each tube with one female parasitoid. After 24 h of parasitism activity, new groups of five host pupae were again transferred into the tubes with the original female parasitoid for continued parasitism. These procedures were repeated until the parasitoid wasps died. After the fifth day, the progeny emerged, the pupae were dissected to confirm parasitism status, and the number of parasitized pupae was counted. To determine the effect of parasitoid age on the performance of T. brontispae, 30 parasitized pupae (30 replicates) for each parasitoid age were transferred into new tubes and maintained under the same laboratory conditions. After emergence, the number of adult parasitoids was counted, and the sex ratio (male/female), the emergence rate, and the egg to adult development time were determined.

Choice and no-choice tests

Choice and no-choice tests were conducted to determine T. brontispae host preference and performance. Before the tests, B. longissima pupae were randomly selected from five host age classes: 2-h, and 1-, 2-, 3-, and 4-day-old. For the choice test, we used one round container (50.5 mm in diameter, 5.5 mm in height) with five small enclosed areas (45 mm in diameter) each with an open door. Figure 1 is a diagram of the choice test equipment used. A group of ten pupae from each of the five host age groups was transferred into the small rooms with a 10% honey solution soaked piece of tissue paper. After that, one mated female parasitoid was placed in

Fig 1 The diagram of the choice test equipment used for determining host preference of Tetrastichus brontispae over Brontispa longissima.

the central position of the box, and the box was maintained in a climate cabinet under the conditions described above. For the no-choice tests, five tubes (2 cm in diameter, 0.5 cm in length) were used in which ten B. longissima pupae for of each of the five host ages were selected and paired with one mated female parasitoid and later transferred into each tube with the already-established ten host pupae.

For both the choice and no-choice tests, 50 female parasitoid wasps (replicates) were tested for each host age. All female parasitoids were removed after 24 h of parasitism activity. After the fifth day, the progeny emerged and the parasitized pupae were dissected to confirm parasitism status, the parasitized pupae were counted, and the parasitism rate was calculated. Furthermore, 30 parasitized pupae for each host age for both tests were removed and placed in another new tube. Once adults emerged, we recorded the developmental time from egg to adult, the number of emerged females and males, the sex ratio, and the emergence rate.

Data analysis

For the choice tests, a choice index (SI) was used to assess parasitism preferences. SI was calculated using the following equation: SI = $R_i / \sum_{i=1}^{m} R_i$, where R_i is the percentage of para $i=1$ sitized pupae in all tested pupae for each treatment, and m is the number of different host categories used in the test.

The experimental design was completely randomized and balanced. The data presented as percentages were normalized using a logarithmic transformation. All statistical tests

were performed using the GLIMMIX procedure in SAS version 9.13 [\(www.sas.com](http://www.sas.com/)). Data were subjected to an analysis of variance for determination of differences between means by using one-way ANOVA and the Tukey's HSD test. Differences in treatments at a probability level of $p < 0.05$ were considered to be significant.

Results

Effects of host density

The effect of host density on parasitism of T. brontispae is shown in Fig 2. A similar number of parasitized pupae were observed when the host pupae density was 3, 4, 5, and 7, whereas the number significantly declined when the host density was 2 and 6 ($p <$ 0.05, Tukey's test).

Effects of parasitoid age

Generally, the parasitism performance of T. brontispae was highly influenced by wasp age, and the parasitism capacity decreased with an increase in parasitoid age. Younger parasitoids (1- to 2-day-old) parasitized a similar number of host pupae and had a higher parasitism than older females (p < 0.05, Tukey's test, Table [1](#page-4-0)). Parasitoid age also influenced the adult emergence rate and offspring production, which were higher in younger wasps. However, the sex ratio and developmental time from egg to adult were not affected by parasitoid age, ranging from 84.7 to 87.3% and 18.0 to 18.6 days, respectively (Table [1](#page-4-0)).

Choice tests and the effects of host age

There was female T. brontispae parasitization of pupae in each host age group (Table [2\)](#page-4-0). The parasitoids selected relatively more 2-h-, 1-day-, and 2-day-old host pupae to parasitize. However, for 3- and 4-day-old host pupae, parasitism

declined to an average of 0.23 and 0.10 pupae, respectively, when they were exposed to the parasitoids together with the other treatment in the choice test (Table [2\)](#page-4-0). The above results were also observed with respect to the parasitism rate of T. brontispae in consideration of the tested host stages of B. longissima pupae (Table [2](#page-4-0)). Taken together, all tests indicated that T. brontispae had a greater preference for parasitization of younger B. longissima pupae; thus, the 2-h to 2 day-old pupae had higher parasitism than the 3- and 4-dayold pupae. However, the sex ratio, the emergence rate, and the developmental time were not influenced by the host stages ($p > 0.05$, Tukey's test).

No-choice tests and the effects of host age

In the no-choice tests, a similar number of 2-h and 3-day-old parasitized pupae were found, ranging from an average of 2.0 and 2.4 pupae, respectively, whereas the number significantly declined to only 0.6 pupae for the 4-day-old pupae treatment (Table [3](#page-5-0)). These same results were also observed for the rate of T. brontispae parasitism for the tested host stages of B. longissima pupae (Table [3\)](#page-5-0). However, the host stages did not affect the parasitoid sex ratio, the emergence rate, or the developmental time of offspring, as there were no differences among the treatments ($p > 0.05$, Tukey's test) (Table [3](#page-5-0)).

Discussion

In biological control programs, the most significant challenges are using an effective parasitoid rearing technique and the parasitism specificity tests in the laboratory. The present study of the characteristics of the parasitoid-host relationship of T. brontispae parasitism on B. longissima is essential for promoting an understanding of the successful mass rearing of T. brontispae. This research may also be used to accurately interpret and provide predictions regarding these characteristics in relation to natural settings, before the actual release.

Fig 2 Number of parasitized pupae under different host densities (2 to 7 in of Brontispa longissima pupae) at $26 \pm 1^{\circ}$ C and 75 ± 5% RH under 12:12-L/D photoperiod in the laboratory conditions. Data are presented as means ± SE. For each set, bars having the same letter are not significantly different at $p = 0.05$ by Tukey's HSD test.

Parasitoid age	No. of parasitized pupae	No. of emerged female	Sex ratio (% female)	% Adult emergence rate	Development time (days)
1-day-old	2.9 ± 0.33 a	28.7 ± 3.48 a	84.8 ± 4.22 a	91.2 ± 6.08 a	18.6 ± 0.11 a
2-day-old	2.0 ± 0.18 a	24.9 ± 1.28 a	86.1 ± 4.16 a	90.9 ± 6.15 a	$18.3 \pm 0.11 a$
3-day-old	0.8 ± 0.16 b	8.7 ± 0.95 b	84.8 ± 4.55 a	84.6 ± 5.87 b	18.5 ± 0.12 a
4-day-old	0.2 ± 0.17 b	7.2 ± 1.62 b	87.3 ± 5.02 a	81.2 ± 5.24 bc	18.1 ± 0.11 a

Table 1 Parasitism performance and reproduction of 1-, 2-, 3-, and 4-day-old Tetrastichus brontispae on Brontispa longissima under controlled laboratory conditions ($26 \pm 1^{\circ}$ C; $75 \pm 5\%$ RH; 12:12-L/D photoperiod).

Data are presented as means ± SE. Means in the same column followed by different small letters are significantly different at $p < 0.05$ level by Tukey's HSD test.

Successful biological control of B. longissima by mass release of T. brontispae has been reported in several countries (Voegele [1989](#page-6-0), Chiu & Chien [1985,](#page-6-0) Nakamura et al [2006\)](#page-6-0); however, knowledge on host specificity of the parasitoid needs to be identified and improved. Our previous study demonstrated that T. brontispae parasitism is largely influenced by temperature and the cold storage period (Liu et al [2014\)](#page-6-0); this study increases the amount of information on the characteristics of T. brontispae parasitism on B. longissima.

In the current study, more parasitized pupae were observed when there were five host pupae exposed to one parasitoid (Fig [2\)](#page-3-0). Moreover, no statistically significant differences were observed with densities of 3, 4, 5, or 7 host pupae per parasitoid (Fig [2\)](#page-3-0). Therefore, considering the costs for mass rearing in biological control programs, we recommend three host pupae per parasitoid. In contrast, Chen et al [\(2010\)](#page-6-0) demonstrated that T. brontispae oviposition frequency became higher as host density increased because the parasitoid would have had more opportunities to encounter a host. Tang et al ([2014a](#page-6-0), [b\)](#page-6-0) also demonstrated that the fecundity of T. brontispae increased as the host density of *O. nipae* increased. Furthermore, the current study revealed that the number of parasitized pupae declined with low host density, especially when the host density was only two pupae. These results are consistent with the results of Godfray [\(1994\)](#page-6-0) and Quicke [\(1997\)](#page-6-0), who reported that other parasitoid species avoided previously parasitized hosts. It is possible that superparasitism occurs with lower host density (Ichiki et al [2011,](#page-6-0) Nguyen et al [2012\)](#page-6-0), but this behavior requires further study. Another likely reason is that the capacity for parasitism might be restricted because of few opportunities to encounter a host in a lower host density environment.

The female parasitoid age is rather important for successful parasitism (Aung et al [2010\)](#page-5-0), and Amalin et al [\(2005\)](#page-5-0) showed the importance of parasitoid age to parasitism using a field release experiment. Research of parasitism in relation to parasitoid age has been widely documented for other species (Hentz [1998,](#page-6-0) Honda et al [1998,](#page-6-0) Amalin et al [2005,](#page-5-0) Ayvaz et al [2008](#page-6-0)). In this study, younger parasitoids (1- to 2-day-old) had a higher parasitism performance than older parasitoids (3- to 4day-old). Our results are consistent with those of Nguyen et al [\(2012\)](#page-6-0), who also detected a decrease in T. brontispae parasitism on B. longissima in older parasitoids. Similarly, younger T. brontispae were also more effective at attacking the host O. nipae (Tang et al [2014a,](#page-6-0) [b](#page-6-0)). However, Pitcher et al [\(2002](#page-6-0)) demonstrated that parasitism efficiency of another parasitoid species was higher for older parasitoids than for younger ones, primarily owing to the higher searching speed, activity rate, and oviposition frequency of the former. Meanwhile, Ayvaz et al [\(2008](#page-6-0)) reported that middled-aged Trichogramma evanescens had a higher parasitism rate than younger and older individuals.

The current study reveals that host age also played an important role in the parasitism of T. brontispae. Both choice and no-choice tests confirmed that T. brontispae has a strong preference for parasitization of younger B. longissima pupae (from 2-h to 2-day-old) than older pupae (Tables 2 and [3\)](#page-5-0). This result is consistent with Chen et al ([2010\)](#page-6-0) and Nguyen

Table 2 Parasitism and reproduction of Tetrastichus brontispae on different stages (2-h to 4-day-old) of Brontispa longissima pupae in choice experiments under controlled laboratory conditions (26 ± 1°C; 75 ± 5% RH; 12:12-L/D photoperiod).

	Host age No. of parasitized pupae Parasitism rate (%) Choice indexes Sex ratio (% female) % Adult emergence rate Development time (days)					
2-h-old	0.8 ± 0.14 a	15.3 ± 2.83 a	0.28 ± 0.06 a	85.3 ± 4.25 a	$89.9 \pm 6.02 a$	18.7 ± 0.12 a
	1-day-old 0.9 ± 0.14 a	18.0 ± 2.77 a	0.29 ± 0.04 a	84.5 ± 4.09 a	88.2 ± 5.94 a	$18.6 \pm 0.11 a$
	2 -dav-old 0.8 ± 0.18 a	17.3 ± 3.68 a	0.29 ± 0.06 a	$86.7 \pm 5.21 a$	88.7 ± 5.48 a	18.3 ± 0.13 a
	3 -day-old 0.2 ± 0.10 b	4.8 ± 2.08 b	0.07 ± 0.03 b	87.7 ± 5.06 a	$84.4 \pm 5.31 a$	$18.9 \pm 0.11 a$
4-day-old	0.1 ± 0.06 C	2.0 ± 1.11 C	0.03 ± 0.01 C	84.9 ± 4.54 a	81.3 ± 5.23 ab	18.9 ± 0.13 a

Data are presented as means \pm SE. Means in the same column followed by different small letters are significantly different at $p < 0.05$ level by Tukey's HSD test.

Host ages	No. of parasitized pupae	Parasitism rate (%)	Sex ratio (% female)	% Adult emergence rate	Development time (days)
2-h-old	2.2 ± 0.17 a	44.1 ± 3.49 a	86.7 ± 4.02 a	90.2 ± 6.03 a	18.5 ± 0.11 a
1-day-old	2.0 ± 0.21 a	40.7 ± 4.23 a	84.9 ± 4.05 a	90.1 ± 6.14 a	18.5 ± 0.12 a
2-day-old	2.3 ± 0.17 a	46.0 ± 3.48 a	85.3 ± 4.26 a	90.0 ± 6.25 a	18.2 ± 0.12 a
3-day-old	2.4 ± 0.18 a	48.0 ± 3.66 a	$87.0 \pm 4.41 a$	89.3 ± 5.74 a	18.8 ± 0.10 a
4-day-old	0.6 ± 0.13 b	12.0 ± 2.64 b	86.3 ± 5.27 a	88.5 ± 5.62 a	18.7 ± 0.12 a

Table 3 Parasitism and reproduction of Tetrastichus brontispae on different stages (2-h to 4-day-old) of Brontispa longissima pupae in non-choice experiments under controlled laboratory conditions (26 ± 1°C; 75 ± 5% RH; 12:12-L/D photoperiod).

Data are presented as means ± SE. Means in the same column followed by different small letters are significantly different at $p < 0.05$ level by Tukey's HSD test.

et al ([2012](#page-6-0)), who demonstrated that the optimum host age in B. longissima for parasitism was 1-day-old pupae. Recently, Tang et al ([2014a](#page-6-0), [b](#page-6-0)) reported that T. brontispae also preferred 1-day-old O. nipae pupae, rather than older pupae. The primary reason why T. brontispae prefer younger pupae is that older pupae are less suitable for oviposition, primarily because it is more difficult for the parasitoid to penetrate the hard cuticle of older pupae. In addition, it is possible that older pupae are more capable of generating a physical or physiological defense response to parasitism, or perhaps they only provide adequate nourishment for parasitoid development for a limited time. Such immune responses should be explored following the example of Tang et al ([2014a](#page-6-0), [b\)](#page-6-0), who investigated the effects of T. brontispae parasitization on the immune system of the host, O. nipae. Nevertheless, some studies obtained opposite results on other parasitoids, such as one on the parasitism of Lariophagus distinguendus (Foerster) which was shown to positively correlated with the host age of Sitophilus oryzae (Line) (Van den Assem [1971](#page-6-0)). Kitt & Keller [\(1998\)](#page-6-0) also indicated a decrease in the suitability of younger hosts, which they suggested was most likely due to their smaller size.

Conducting both choice and no-choice tests resulted in a more accurate assessment of host range, host preference, and parasitism performance of parasitoids (Ayvaz et al [2008,](#page-6-0) Mansfield & Mills [2004,](#page-6-0) Murray et al [2010](#page-6-0)). The results of the current study show that both choice and no-choice tests perform the same in terms of determining the preferences of T. brontispae in relation to the range of host ages. The role and the value of using choice versus no-choice host specificity tests for other parasitoids have been debated previously (Van Driesche & Murray [2004,](#page-6-0) Withers & Mansfield [2005,](#page-6-0) Murray et al [2010\)](#page-6-0). In nature, parasitoids that cannot locate their preferred hosts may disperse, rather than accept, lower quality hosts, yet hosts across the widest possible range of quality may be accepted in no-choice tests. However, this implies the risk of false positives with the tests because of severely overestimating the potential range in host quality, whereas in nature parasitoids may reject lower quality hosts (Goldson & Phillips [1990](#page-6-0), Withers & Browne [2004](#page-6-0)); this may explain a

higher parasitism rate in no-choice tests compared to choice tests (Tables [2](#page-4-0) and 3). If so, choice tests arguably are useful for modeling the host selection processes including the location of host habitat, host attack, and host suitability (Kitt & Keller [1998,](#page-6-0) Withers & Mansfield [2005\)](#page-6-0) because they provide a reasonable opportunity to display host preferences. However, choice tests must also be viewed with caution, specifically in regards to whether parasitoids do in fact "choose" among the different host ages under natural conditions. Most parasitoids are unlikely to encounter more than one host age at a time in nature. Otherwise, Edwards ([1999\)](#page-6-0) and Barratt ([2004](#page-6-0)) reported that a parasitoid accepts or rejects encountered hosts based on stimuli present at the time, and the previous experience and physiological state of the parasitoid.

In summary, results of the present study lead to the conclusion that a host density of three host pupae per parasitoid is ideal when the costs for mass rearing are also considered. Parasitoid age was critical for successful parasitism, with younger T. brontispae performing better than older T. brontispae. Both choice and no-choice tests confirmed that the parasitoid had a stronger preference to parasitize the 2-h to 2-day-old host pupae. Taken together, the results of the present study have increased knowledge of the parasitoid-host relationship of T. brontispae on B. longissima.

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