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Synthesis, molecular docking, and in silico ADME prediction of some fused pyrazolo[1,5‑*a***]pyrimidine and pyrazole derivatives as potential antimicrobial agents**

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Abstract

Twenty compounds of pyrazolo[1,5-*a*]pyrimidines **14a–j** and pyrazole derivatives as Schif bases **16a–j** have been synthesized by the reaction of 5-amino-pyrazole derivatives **9a**, **b** with 2-(arylidene)malononitriles **10a–e** or various aldehydes **15a–e**, respectively. The biological activity of the pyrazolo[1,5-*a*]pyrimidines **14a–j** and pyrazole derivatives **16a–j** was evaluated and showed a variation in antimicrobial inhibitory activity. In particular, the three pyrazolo[1,5-*a*]pyrimidines (**14b**, **14e,** and **14j**) and four pyrazole Schif bases (**16c**, **16d**, **16 h,** and **16i**) have possessed a broad spectrum of activity with the inhibition zone range from 15 to 20 mm. Also, the structure–activity relationship study was done. Furthermore, physicochemical properties, drug-likeness model score, bioactivity scores, ADME, and toxicity properties were predicted in silico and some products possessed acceptable and a good results. Finally, the molecular docking studies performed inside the active site of DNA gyrase (PDB: 2XCT), the secreted aspartic protease from *C. albicans* (PDB: 1ZAP), and exhibited lower binding energy with diferent types of binding mode. Besides, the electrostatic potential maps were generated to determine the charge distribution around a molecule and therefore determine the regions that could form hydrogen bonding donors and acceptors that confrmed the binding mode in the docking study. The results suggested that these derivatives could act as antimicrobial agents.

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Graphic abstract

Introduction

The important tasks of the researchers in the medical, pharmaceutical, and chemical felds are the design and preparation of bioactive compounds to overcome some health problems such as cancer and bacterial infectious diseases. Therefore, there is a growing interest in the preparation of pyrazolo[1,5-*a*]pyrimidines derivatives due to their biological applications as promising antimicrobial, anticancer, antitubercular, and enzyme inhibitors [[1](#page-22-0)–[6](#page-22-1)]. For example, pyrazolo[1,5-*a*]pyrimidine-3-carbonitrile derivative **1** was synthesized by Attia et al., as a promising anti-proliferative agent against the MCF-7 cell line [\[7](#page-22-2)]. The two water-soluble pyrazolo[1,5-*a*]pyrimidine derivatives (**2** and **3**) were synthesized and characterized with antibacterial activity against *Escherichia coli* [[8](#page-22-3)]. Also, from our previous studies, the two pyrazolo[1,5-*a*]pyrimidine derivatives {5,7-dimethylpyrazolo[1,5-*a*]pyrimidine derivative **4** and 7-phenylpyrazolo[1,5-*a*]pyrimidine derivative **5**} were prepared and displayed a promising antimicrobial agent [[9\]](#page-23-0) and antitumor agent against HepG-2 cells [[10\]](#page-23-1), respectively (Fig. [1\)](#page-2-0). Moreover, the pyrazolopyrimidine structure is present in drugs such as Dinaciclib, Anagliptin, and Ocinaplon [[11](#page-23-2)].

ties such as antimicrobial, antitumor, immunomodulatory, analgesic, DNA binding, and DHFR/DNA gyrase inhibitors [\[12–](#page-23-3)[18](#page-23-4)]. In 2013, Malladi and co-workers have described a synthesis of compound triazole-pyrazole Schiff base **6** which exhibited promising antibacterial activity against *Staphylococcus aureus* [[19\]](#page-23-5). Recently, in 2020, Hassan et al*.,* prepared two furan-pyrazole Schiff bases **7** and **8** as *Nitrofurantoin*® analogues which exhibited antibacterial activities against *Escherichia coli* and *Salmonella typhimurium* bacteria [[20](#page-23-6)] (Fig. [1\)](#page-2-0). Because of the biological importance of pyrazolo[1,5-

Besides, pyrazole Schiff bases (pyrazole-azomethine) are known to have a vast spectrum of biological activi-

a]pyrimidines, pyrazole Schif bases, and in continuation of our program for synthesis bioactive compounds [[21–](#page-23-7)[40\]](#page-23-8). In this work, we have synthesized 5-amino-*N*-aryl-1*H*-pyrazole-4-carboxamides **9a**, **b** [[41](#page-23-9)] as starting materials for the preparation of new 5-aryl-7-aminopyrazolo[1,5-*a*]pyrimidines **14a–j** and pyrazole Schif bases **16a–j** for evaluation of their antimicrobial properties as well as the structure–activity relationship (SAR), the molecular docking, the electrostatic potential maps (MEP), and in silico properties prediction were studied (Fig. [2\)](#page-3-0).

Fig. 1 Biological applications of pyrazolopyrimidines **1–5** and pyrazole Schif bases **6–8**

Result and discussion

Chemistry

The target compounds, pyrazolo[1,5-*a*]pyrimidines **14a–j** and pyrazole Schif bases **16a–j**, are shown in Schemes [1](#page-4-0) and [2.](#page-5-0) The starting materials, 5-amino-pyrazole derivatives **9a**, **b** and 2-(arylidene)malononitriles **10a–e,** were prepared according to previously reported methods [[42,](#page-23-10) [43](#page-23-11)] and are described in Scheme [3](#page-5-1).

A series of 7-amino-pyrazolo[1,5-*a*]pyrimidines **14a–j** were prepared via the reaction of 5-amino-*N*-aryl-1*H*-pyrazole-4-carboxamides **9a**, **b** with 2-(arylidene)malononitriles **10a–e** in refuxing ethanol (Scheme [1](#page-4-0)). This reaction involves nucleophilic addition of amino group of **9a, b** to double bond of 2-(arylidene)malononitriles **10a–e** to produce the intermediate 5-((2,2-dicyano-1-phenylethyl) amino)-1*H*-pyrazole derivative **11a–j**. Furthermore, the cyclic imino group in intermediate 1*H*-pyrazole derivative **11a–j** acts as nucleophile to cyano group to obtain 7-imino-pyrazolo[1,5-*a*]pyrimidine-3-carboxamide

Fig. 2 The target compounds **(14a–j** and **16a–j)** and their studies

derivatives **12a–j** that underwent proton shift and oxidation to afford the corresponding 7-amino-pyrazolo^[1,5-*a*] pyrimidine derivatives **14a–j**.

The structure of 7-amino-pyrazolo[1,5-*a*]pyrimidines structures **14a–j** was confrmed by spectral data and elemental analysis. The IR spectrum of 7-aminopyrazolo[1,5-*a*]pyrimidine derivative **14a** was characterized by appearing new band for C≡N group at ν 2213 cm⁻¹ and two bands at ν 3428 and 3304 cm⁻¹ for NH₂ and 2NH groups. Also, the C=O amide group appeared at *ν* 1641 cm−1. The mass spectrum of pyrazolo[1,5-*a*]pyrimidine derivative **14a** was exhibited a molecular ion pack at *m/z* 489 with intensity 17.65% corresponding to the formula $C_{28}H_{23}N_7O_2$ (489.53) and the base peak at m/z 396 (100%). The ¹H NMR (δ /ppm) spectrum of **14a** exhibited two signals at *δ* 2.43 and 3.74 ppm assigned to the protons of CH_3 and OCH₃ groups, respectively. Besides, the NH₂ group appeared as a broad signal at *δ* 9.00 ppm, and two

exchangeable singlet signals at *δ* 9.23 and 10.14 ppm corresponding to two NH groups.

The three aromatic rings (Figure S1, Supplementary Material) were interpreted as the following:

- (i) Phenyl ring (fve protons) appeared as two triplets at *δ* 7.09 ppm (1H, *J*=7.5 Hz) and *δ* 7.37 ppm (2H, $J = 7.8$ Hz) and one doublet at δ 7.60 ppm (2H, *J*=7.7 Hz).
- (ii) 4-Methoxyphenyl ring (4 protons, AB-system) appeared as two doublets at δ 6.90 (2H) and 7.84 (2H) ppm (each with *J* (coupling constant) = 9.0 Hz).
- (iii) 4-Methylphenyl ring (4 protons, *AB-system*) appeared as two doublets at *δ* 7.44 (2H) and 7.90 (2H) ppm (each with J (coupling constant) = 8.2 Hz).

Additionally, the 13C NMR spectrum of **14a** exhibited characterized signals of CH_3 and $-OCH_3$ carbon at δ 21.19

Scheme 1 Synthesis of pyrazolo[1,5-*a*]pyrimidines **14a–j**

and 55.26 ppm, respectively. The rest of carbon atoms (26 carbon) of 7-amino-pyrazolo[1,5-*a*]pyrimidine derivative **14a** have appeared ranged between *δ* 74.88–145.70 ppm as well as signals at *δ* 149.49, 153.98, 156.32, 161.47, and 162.29 ppm corresponding to two C=N, carbonyl group, carbon attached to methoxy (C–OMe), and carbon attached to an amino group $(C-NH₂)$, respectively.

Moreover, the 5-(2,5-dimethoxyphenyl)-pyrazolo[1,5-*a*] pyrimidine-3-carboxamide derivative **14j** was another example of the ¹H NMR spectra, exhibited three singlet signals at *δ* 3.70, 3.75, and 3.78 ppm for the three methoxy groups. The aromatics protons appeared at δ 6.86 (d, 2H, $J=9.5$ Hz, *p*-methoxyphenyl ring), 7.10–7.17 (m, 3H, 2, 5-dimethoxyphenyl ring), 7.34 (d, $2H$, $J=9.2$ Hz, *p*-chlorophenyl ring), 7.48 (d, 2H, *J* = 9.2 Hz, *p*-chlorophenyl ring), and 7.78 (d, 2H, *J*=9.3 Hz, *p*-methoxyphenyl ring). Further, three exchangeable singlet signals appeared as the broad peak

at *δ* 8.97 ppm, and two signals at *δ* 9.11 and 10.04 ppm refer to the protons of the amino group $(NH₂)$ as well as two NH groups, respectively. The 13C NMR spectrum of **14j** exhibited characterized signals of three methoxy carbons at *δ* 55.12, 55.68, and 55.93 ppm. The rest of carbon atoms (26 carbon) of **14j** have appeared at *δ* 78.02, 88.84, 104.95, 113.21, 114.13, 115.17, 117.23, 118.96, 120.38, 126.91, 128.90, 133.25, 137.20, 145.72, 148.51, 150.65, 153.07, 153.91, 156.07, 159.97, 159.99, and 162.09 ppm. Also, the structure of 7-amino-pyrazolo[1,5-*a*]pyrimidine derivatives **14a–j** and reaction mechanism pathway was proved from our previous study through X-ray single-crystal analysis of analogues [\[44](#page-23-12)].

Schif bases **16a–j** were prepared via the reaction of 5-amino-pyrazoles **9a**, **b** with various aldehydes **15a–e** in refuxing absolute ethanol (Scheme [2](#page-5-0)). Schif bases structure **16a–j** was confrmed by spectral analysis. The IR spectrum

Scheme 2 Synthesis of Schif bases **16a–j**

of azomethine derivative **16d** displayed bands at *ν* 3431, 3235, 1635, and 1615 cm−1 characteristic for 3NH, carbonyl $(C=O)$, and imino $(C=N)$ groups, respectively. The ${}^{1}H$ NMR (*δ*/ppm) spectrum of 5-(4-chloro-3-nitrobenzylideneamino)- 1*H*-pyrazole-4-carboxamide derivative **16d** exhibited one signal at *δ* 3.70 ppm for the methoxy group. The aromatic

protons have appeared at δ 6.89 (d, 2H, $J = 8.3$ Hz, *p*-methoxyphenyl ring), 7.06 (t, 1H, $J = 7.3$ Hz, phenyl ring), 7.31–7.34 (m, 4H, phenyl ring), 7.67 (d, 2H, *J*=7.9 Hz, *p*-methoxyphenyl ring), 8.00 (d, 1H, *J*=8.3, 4-chloro-3-nitrophenyl ring), 8.30 (d, 1H, *J*=8.3, 4-chloro-3-nitrophenyl ring), and 8.67 (s, 1H, 4-chloro-3-nitrophenyl ring). The methine proton (–CH=N–) has appeared at δ 8.76 ppm. Besides, the three exchangeable signals appeared at *δ* 9.13, 9.79, and 12.80 ppm related to three NH protons.

Additionally, the IR spectrum of pyrazole Schif base derivative **16f** was characterized by bands for 3NH, C=O, C=N, and C=C groups at *ν* 3434, 3229, 1656, 1612, and 1587 cm−1, respectively. The 1 H NMR (*δ*/ppm) spectrum of pyrazole derivative **16f** exhibited two signals at *δ* 3.73 and 3.87 ppm for the protons of two methoxy groups. Besides, the proton of $-CH=N$ – function appeared at δ 8.66 ppm and the protons of the three NH appeared as single signals at *δ* 9.03, 10.02, and 12.78 ppm. The spectrum has exhibited the protons of the three aromatic rings (Figure S2, Supplementary Material) as.

- (i) Two doublets appeared at δ 6.90 (2H, J = 8.9 Hz) and 7.69 (2H, *J*=8.9 Hz) for the *p*-methoxyphenyl ring.
- (ii) One doublet at *δ* 7.40 (4H) for the *p*-chlorophenyl ring with $J=8.9$ Hz.
- (iii) *m*-Methoxyphenyl ring; one proton as a doublet–doublet (dd) at δ 7.23 ($J=8.2 \& 1.8$ Hz), one proton as a triplet at *δ* 7.53 (*J*=7.9 Hz), single signal at *δ* 7.59 for one proton, and one doublet for one proton at *δ* 7.62 $(J=2.4$ Hz).

The 13C NMR spectrum of **16f** exhibited characterized signals of two methoxy carbons at *δ* 55.19 and 55.23 ppm. The rest of 23 carbon atoms appeared at *δ* 92.61, 112.24, 114.39, 119.22, 119.53, 120.41, 122.48, 126.69, 128.34, 128.73, 130.28, 133.79, 136.17, 137.46, 151.21, 154.04, 159.74, 162.03, and 162.65 ppm.

Antimicrobial evaluation

Pyrazolo[1,5-*a*]pyrimidines **14a–j** and pyrazole Schif bases **16a–j** were evaluated in vitro for their antimicrobial activity against four microbes using the agar difusion method [[45,](#page-23-13) [46](#page-23-14)], and the results are listed in Table [1](#page-7-0) and represented in Fig. [3](#page-8-0)**.**

From the results obtained in Table [1](#page-7-0) and Fig. [3](#page-8-0), it was found that most of the pyrazolo[1,5-*a*]pyrimidines **14a–j** and pyrazole Schif bases **16a–j** have a reasonable antimicrobial undertaking impact against most of the tested pathogens in this study. Besides, most of the pyrazolopyrimidines **14a–j** and pyrazole Schif bases **16a–j** confrmed a diverse inhibitory efect against all pathogenic microorganisms used.

In the case of Bacillus subtilis (G+) bacteria: the three pyrazolopyrimidines [**14b**, **14e** (inhibition zone $(IZ) = 20$ mm), and **14j** $(IZ = 21$ mm)] and the pyrazole Schiff base $16i$ ($IZ = 19$ mm) confirmed a strong inhibition efect more than the antibacterial standard used (Tetracycline, IZ=18 mm). Furthermore, pyrazolopyrimidine **14d** and the pyrazole Schiff bases $16a$ (IZ = 18 mm) showed comparable antibacterial activity of the Tetracycline drug $(IZ = 18$ mm). Both the pyrazolopyrimidine derivatives (**14c** and **14g–i**) and pyrazole Schif bases (**16c–h** and **16j**) showed a good inhibition with range $=15-17$ mm.

In the case of Escherichia coli (G−) bacteria: all the pyrazolo[1,5-*a*] pyrimidines **14a–j** and pyrazole Schif bases **16a–j** ($IZ = 13$ to 16 mm) presented from good to an excellent inhibition compared to the Novobiocin ($IZ = 16$ mm).

Aspergillus niger (Filamentous fungi): Pyrazolopyrimidine **14f** and four pyrazole Schif bases **(16c, 16d, 16h,** and **16i**) revealed IZ from (18 mm) to (20 mm) and confrmed a robust inhibition impact*,* while, four pyrazolopyrimidines (**14d, 14e, 14g,** and **14i**) and three Schif bases **(16a, 16e**, and $16j$) with $IZ = 15$ mm showed a reasonable inhibitory activity compared to Clotrimazole ($IZ=23$ mm) as a positive control. It is worth mentioning that *Aspergillus niger* is one of the fungal pathogens that can afect the respiratory tract. *A. niger* is a causative agent of pulmonary diseases inclusive of aspergillosis, bronchial asthma, and acute allergic alveolitis. The fungus colonizes ancient tuberculosis or bronchi static cavities, in which its varieties a large colony (aspergilloma); or it can also without a doubt invade the lung tissue to produce hemorrhagic and necrotizing pneumonia [[47\]](#page-23-15).

In the case of Candida albicans (Yeast): the pyrazolopyrimidines (**14b**, **14d,** and **14f**) and Schif base **16i** (IZ $rang = 17–20$ mm) showed a good inhibition effect. Moreover, the pyrazolopyrimidines (**14c, 14e, 14g,** and **14i**) and Schif bases (**16b, 16c, 16d, 16e, 16g,** and **16h**) showed a moderate inhibition with inhibition zone $(IZ) = 15$ mm. Further, the pyrazolopyrimidine $14a$ ($IZ = 13$ mm) and two Schiff bases $16a$ (IZ=13 mm) and $16j$ (IZ=12 mm) showed a weak efect in contrast to the antibacterial standard antibiotic Cyclohexamide (IZ=32 mm).

From the above, the three pyrazolo[1,5-*a*]pyrimidines (**14b**, **14e**, and **14j**) and four pyrazole Schif bases (**16c**, **16d**, **16h,** and **16i**) proved to be potent against the four microbes tested, possessed a broad spectrum of activity, and in several cases, their activities exceeded the reference drugs used.

Structure activity relationship study

Depending on the structure represented on Schemes [1,](#page-4-0) [3,](#page-5-1) and Fig. [4,](#page-9-0) our synthesis strategy depends on the preparation of two 2-cyano-*N*-(aryl)acetamide, and the aryl represented as phenyl or 4-chlorophenyl. The cyanoacetanilide derivatives reacted with 4-methoxyphenyl isothiocyante and fnally refuxed with hydrazine hydrate to produce 5-aminopyrazole derivatives **9a, b** with two variable aryl groups. The 5-amino-pyrazole derivatives **9a, b** obtained used as key starting materials to form pyrazolo[1,5-*a*]pyrimidine derivatives **14a–j** and azomethine pyrazole derivatives **16a–j**. Generally, most of the target compounds **14a–j** and

Compounds	Ar	Ar ₁	Ar ₂	Bacteria		Fungi	
				(G^+) B. subtilis ATCC-6633	(G^-) E. coli ATCC-25922	Filamentous A. niger NRRL-3	Unicellular C. albicans ATCC-10231
14a	Ph	4 -CH ₃ OC ₆ H ₄	4 -CH ₃ C ₆ H ₄	12	14	$\overline{0}$	13
14 _b	Ph	4 -CH ₃ OC ₆ H ₄	3 -CH ₃ C ₆ H ₄	20	16	$\overline{0}$	17
14c	Ph	4 -CH ₃ OC ₆ H ₄	3 -CH ₃ OC ₆ H ₄	15	14	Ω	15
14d	Ph	4 -CH ₃ OC ₆ H ₄	$3-CIC6H4$	18	15	15	17
14e	Ph	4 -CH ₃ OC ₆ H ₄	$2,5$ -di-CH ₃ OC ₆ H ₃	20	16	15	15
14f	$4-CIC6H4$	4 -CH ₃ OC ₆ H ₄	4 -CH ₃ C ₆ H ₄	13	14	18	18
14 _g	$4-CIC6H4$	4 -CH ₃ OC ₆ H ₄	3 -CH ₃ C ₆ H ₄	16	15	15	15
14 _h	$4-CIC6H4$	4 -CH ₃ OC ₆ H ₄	3 -CH ₃ OC ₆ H ₄	15	15	$\mathbf{0}$	$\overline{0}$
14i	$4-CIC6H4$	4 -CH ₃ OC ₆ H ₄	$3-CIC6H4$	16	15	15	15
14j	$4-CIC6H4$	4 -CH ₃ OC ₆ H ₄	$2,5$ -di-CH ₃ OC ₆ H ₃	21	16	$\overline{0}$	$\overline{0}$
16a	Ph	4 -CH ₃ OC ₆ H ₄	3 -CH ₃ OC ₆ H ₄	18	15	15	13
16b	Ph	$4\text{-}\mathrm{CH}_3\mathrm{OC}_6\mathrm{H}_4$	$4-(CH_3)_2 N C_6H_4$	12	13	$\overline{0}$	15
16c	Ph	4 -CH ₃ OC ₆ H ₄	$2,5$ -di-CH ₃ OC ₆ H ₃	15	14	20	15
16d	Ph	4 -CH ₃ OC ₆ H ₄	4-Cl-3-NO ₂ -C ₆ H ₃	17	16	20	15
16e	Ph	4 -CH ₃ OC ₆ H ₄	Thiophen-2-yl	15	15	15	15
16f	$4-CIC6H4$	4 -CH ₃ OC ₆ H ₄	3 -CH ₃ OC ₆ H ₄	15	15	$\mathbf{0}$	$\overline{0}$
16g	$4-CIC6H4$	4 -CH ₃ OC ₆ H ₄	4- $\left({\rm CH}_3\right)_2$ N ${\rm C}_6{\rm H}_4$	15	14	$\overline{0}$	15
16h	$4-CIC6H4$	4 -CH ₃ OC ₆ H ₄	$2,5$ -di-CH ₃ OC ₆ H ₃	16	15	20	15
16i	$4-CIC6H4$	4 -CH ₃ OC ₆ H ₄	4-Cl-3-NO ₂ -C ₆ H ₃	19	15	20	20
16j	$4-CIC6H4$	4 -CH ₃ OC ₆ H ₄	Thiophen-2-yl	16	14	15	12
A				18	$\overline{}$	$-$	
B					16		
C					$\overline{}$	23	
D						$\overline{}$	32

Table 1 Antimicrobial activities represented by the mean of Inhibition Zone (IZ) in (mm) of pyrazolo[1,5-*a*]pyrimidines **14a–j,** pyrazole Schif bases **16a–j,** and standard antibiotics using the agar difusion method

Antibacterial Standard; A=Tetracycline (30 µg); B=Novobiocine (30 µg), Antifungal Standard; C=Clotrimazole (50 µg); D=Cyclohexamide $(50 \,\mu g)$

16a–j showed moderate to favorable activity against the tested pathogens.

Firstly, the 7-amino-6-cyano-2-((4-methoxyphenyl) amino)-*N*-phenyl-5-(aryl)pyrazolo[1,5-*a*]pyrimidine-3-carboxamide **14a–e** displayed good antibacterial results, moderate antifungal activity, and the activity depends on the aryl group at position fve and aromatic amide at position three of pyrazolo[1,5-*a*]pyrimidine derivatives **14**. The presence of a dimethoxy group at positions two and fve in the aryl group and *meta* methyl group in pyrazolo[1,5-*a*]pyrimidine **14e** & **14b** exhibited antibacterial activity with IZ ranged between 16–20 mm compared with Tetracycline 18 mm and Novobiocin 16 mm. Additionally, these two derivatives displayed moderate antifungal activity with IZs ranged between 15–17 mm in comparison with Clotrimazole $(IZ = 23$ mm) and Cycloheximide ($IZ = 32$ mm). Further, replacement *N*-phenyl by *N*-(4-chlorophenyl) exhibited equipotent in case of $5-(2,5-di-CH_3O-C₆H_3)$ pyrazolo^{[1},5-*a*] pyrimidine-3-carboxamide derivative **14j** with IZs ranged between

16–21 mm, while $5-(3-CH_3-C_6H_4)$ pyrazolo $[1,5-a]$ pyrimidine-3-carboxamide derivative **14g** revealed slight decrease with IZs 16 mm, and 15 mm against *B. subtilis* and *E. Coli*, respectively. Additionally, it was found that the phenyl ring in pyrazolo[1,5-*a*]pyrimidine-3-carboxamide derivative **14d** had higher activity than the corresponding member incorporating chloro group as *N*-(4-chlorophenyl)-pyrazolo[1,5-*a*] pyrimidine-3-carboxamide derivative **14j** against *B. subtilis* strain. Moreover, replacement *N*-phenyl by *N*-(4-chlorophenyl) of 3-carboxamide group in pyrazolo[1,5-*a*]pyrimidine derivatives **14h** and **14j** demonstrated no activity against fungal strains. Besides, addition of the electron-withdrawn group as chlorine atom in pyrazolo[1,5-*a*]pyrimidine-3-carboxamide derivatives **14f** and **14g** displayed increased the activity with IZ values from zero to 18 and 15 mm, respectively.

Similarly, the azomethine of 5-amino-pyrazole derivatives **16a–j** showed variation in activity related to the different substituents in both two-aryl groups at positions

Fig. 3 Showed the antimicrobial activities (Inhibition Zone (IZ) in mm) of pyrazolo[1,5-*a*]pyrimidines **14a–j,** pyrazole Schif bases **16a–j,** and standard antibiotics **a** bacteria, **b** fungi

three and fve in pyrazole derivatives **16a–j**. Regarding the series of 5-((substituted benzylidene)amino)-1*H*-pyrazole-4-carboxamide derivatives **16a–j**, changing *N*-(phenyl) or *N*-(4-chlorophenyl) has little efect on antimicrobial activity. Furthermore, introducing more electron-withdrawing group as *N*-(4-chlorophenyl) to pyrazole derivatives **16a–j** increase activity against *B. subtilis* from 12–18 mm for pyrazoles **16a–e** to 15–19 mm for pyrazoles **16f–j**. Additionally, changing of 5-((substituted benzylidene)amino) by heterocyclic aldehyde as 5-((thiophen-2-ylmethylene)amino) in 1*H*-pyrazole-4-carboxamide derivatives **16a–d** and **16f–i** not cause signifcant enhancement in antimicrobial activity. The most active azomethine pyrazole derivatives **16a–j** among the synthesized derivatives that having 5-((4-chloro-3-nitrobenzylidene)amino) and *N*-(4-chlorophenyl)-4-carboxamide derivative is pyrazole derivative **16i** demonstrated antimicrobial activity IZs ranging between 15–19 mm and antifungal activity IZs 20 mm. Unfortunately, the pyrazole derivatives **16b**, **16f**, and **16g** are inactive against *A. Niger* except for moderate activity for other pyrazole derivatives. While the pyrazole derivative **16f** showed no activity against *C. albicans*.

Finally, in the general presence of the chloro atom in amide as *N*-(4-chlorophenyl) in position three in **Fig. 4** Illustrate the structure of diferent substituent in both pyrazolo[1,5-*a*]pyrimidine derivatives **14a–j** and pyrazoleazomethine derivatives **16a–j**

pyrazolo[1,5-*a*]pyrimidine derivatives **14a–j** and position four in pyrazole derivatives **16a–j** causes increases the activity with signifcant results against *B. subtilis*, while in the case of *E. Coli* nearly results were obtained. Besides, *N*-(phenyl)-1*H*-pyrazolo[1,5-*a*]pyrimidin-3-carboxamide derivatives **14a–j** and *N*-(phenyl)-1*H*-pyrazole-4-carboxamide derivatives **16a–j** revealed abroad antifungal activity. Moreover, it can be concluded that the pyrazolo[1,5-*a*] pyrimidine derivatives **14a–j** more sensitive to antibacterial activity while 5-((substituted benzylidene)amino)-1*H*-pyrazole-4-carboxamide derivatives **16a–j** displayed better antifungal activity.

In silico studies

Physicochemical properties and drug‑likeness model score

The physicochemical properties and drug-likeness (Lipinski and Veber rules) of the pyrazolo[1,5-*a*]pyrimidines (**14b**, **14e,** and **14j**) and Schif bases (**16c**, **16d**, **16h,** and **16i**) were predicted by using the SwissADME web [\(http://swissadme.](http://swissadme.ch/index.php#undefined) [ch/index.php#undefined](http://swissadme.ch/index.php#undefined)) [[48](#page-23-16), [49\]](#page-23-17). Also, the drug-likeness model score was predicted by using the Molsoft web ([https://](https://www.molsoft.com/mprop/) [www.molsoft.com/mprop/\)](https://www.molsoft.com/mprop/) [\[50\]](#page-23-18). The predicted results are listed in Table [2.](#page-10-0)

From Table [2](#page-10-0), the pyrazolo[1,5-*a*]pyrimidine **14b** and Schiff bases (**16c, 16d, 16h**, and **16i**) agreement with Lipinski's rule of fve and with Veber's rule. While the two pyrazolo[1,5-*a*]pyrimidines (**14e** and **14j**) do not match with each of Lipinski's rule of fve and Veber's rule by the violation in molecular weight and TPSA.

The rule for the drug-likeness model score estimation is based on positive and negative values, when the druglikeness score was positive value; the compound was like a drug. But, if the drug-likeness score was negative value; the compound was non-drug [[51](#page-23-19)].

Accordingly, all the tested compounds except **16c** showed positive values in the range from 0.05 to 0.44; therefore, these compounds may be considered to be druglike. Figure [5](#page-10-1) represents the drug-likeness model score of the two pyrazolo[1,5-*a*]pyrimidines **14b** and **14j**.

The bioactivity scores prediction

The bioactivity scores of the pyrazolo[1,5-*a*]pyrimidines (**14b**, **14e,** and **14j**) and Schif bases (**16c**, **16d**, **16h,** and **16i**) toward G protein-coupled receptor (GPCR) ligand, ion channel modulator, a kinase inhibitor, nuclear receptor ligand, protease inhibitors, and enzyme inhibitor were predicted by using the Molinspiration bioactivity score v2018.03 web ([https://www.molinspiration.com/cgi-bin/](https://www.molinspiration.com/cgi-bin/properties) [properties\)](https://www.molinspiration.com/cgi-bin/properties) [[52](#page-23-20)]. The predicted results are written in Table [3](#page-10-2).

The rule for the bioactivity scores estimation is the following: when the bioactivity score was more than 0.00; the compound was considered active. While if the bioactivity score in the range between -0.50 and 0.00; the compound was moderately active. But if the bioactivity score was less than -0.50 , the compound was inactive.

The following observations are concluded from Table [3](#page-10-2):

Physicochemical proper-	Pyrazolo[1,5-a]pyrimidines				Pyrazole Schiff bases			
ties and drug-likeness score	14 _b	14e	14j	16c	16d	16h	16i	
Molecular weight ≤ 500	489.53	535.55	570.00	471.51	490.90	505.95	525.34	
#H-bond acceptors ≤ 10	5			6	6	6	6	
#H-bond donors ≤ 5	3	3	3	3	3	3	3	
Lipophilicity $MLOGP \leq 4.15$	3.26	2.46	2.92	3.03	3.28	3.50	3.75	
Violations Lipinski's rule (Yes; 0 or 1)	Yes: 0 No: 2	$MW > 500$, N or $O > 10$	No: 2 $MW > 500$, N or $O > 10$	Yes: 0	Yes: 0		Yes; 1: MW > 500 Yes; 1: MW > 500	
#Rotatable bonds ≤ 10	7	9	9	10	9	10	9	
$TPSA^a \leq 140$		130.36 148.82	148.82	109.86	137.22	109.86	137.22	
Violations Veber's rule (Yes; 0)		Yes: 0 No: 1: TPSA > 140	No: 1: $TPSA > 140$	Yes: 0	Yes: 0	Yes: 0	Yes: 0	
Drug-likeness model score	0.38	0.06	0.44	-0.23	0.05	0.22	0.15	

Table 2 Physicochemical properties and drug-likeness model score of the pyrazolo[1,5-*a*]pyrimidines (**14b**, **14e,** and **14j**) and pyrazole Schif bases (**16c**, **16d**, **16h,** and **16i**)

a *TPSA* Topological polar surface area

(MolSoft)

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Fig. 5 Displayed the drug-likeness model score plotting of two pyrazolo[1,5-*a*]pyrimidines **14b** and **14j**. The green curve represents non-drug, and the blue curve represents drug

Table 3 The bioactivity scores prediction of the pyrazolo[1,5-*a*]pyrimidines (**14b**, **14e**, and **14j**) and pyrazole Schif bases (**16c**, **16d**, **16h,** and **16i**)

^aG protein-coupled receptor (GPCR) ligand

- G protein-coupled receptor (GPCR) ligand: all the pyrazolo[1,5-*a*]pyrimidines (**14b**, **14e,** and **14j**) and Schif bases (**16c**, **16d**, **16h,** and **16i**) were found to be moderately active with the bioactivity scores in the range from -0.10 to -0.44 .
- Ion channel modulator: the two pyrazolo[1,5-*a*]pyrimidines **14b** and **14e** were found to be moderately active with the bioactivity scores -0.32 and -0.44 , respectively, while the other compounds were found to be inactive.
- Kinase inhibitor: the three pyrazolo[1,5-*a*]pyrimidines (**14b**, **14e,** and **14j**) and the two Schif bases (**16c** and **16h**) were found to be active with bioactivity scores in the range from 0.06 to 0.045. While the two Schiff bases (**16d** and **16i**) were found to be moderately active with the bioactivity scores -0.02 and -0.03 , respectively.
- Nuclear receptor inhibitor: all the pyrazolo[1,5-*a*]pyrimidines (**14b**, **14e,** and **14j**) and Schif bases (**16c**, **16d**, **16h,** and **16i**) were found to be inactive with the bioactivity scores less than -0.50 (the range from -0.59 to -0.77).
- Protease inhibitor: the three pyrazolo^{[1}], 5-*a*] pyrimidines (**14b**, **14e,** and **14j**) and the two Schif bases (**16c** and **16h**) were found to be moderately active (range from −0.37 to −0.42). But, the two Schif bases (**16d** and

16i) were found to be inactive with the bioactivity scores -0.56 and -0.55, respectively.

• Enzyme inhibitor: all the pyrazolo[1,5-*a*]pyrimidines (**14b**, **14e,** and **14j**) and Schif bases (**16c**, **16d**, **16h,** and **16i**) were found to be moderately active (range from -0.23 to -0.38).

From the above, the pyrazolo[1,5-*a*]pyrimidine **14b** almost shows the highest bioactivity score compared to the other compounds.

ADME and toxicity properties prediction

ADME properties and toxicity properties prediction of the pyrazolo[1,5-*a*]pyrimidines (**14b**, **14e**, and **14j**) and Schif bases (**16c**, **16d**, **16h,** and **16i**) were predicted by using the Pre-ADMET web (<https://preadmet.bmdrc.kr/>) [[53\]](#page-23-21). The predicted results are written in Table [4.](#page-11-0)

The following observations are concluded from Table [4](#page-11-0):

• Cytochrome P450 isoforms determine the pharmacokinetic properties, bioactivation, and safety persistence of drugs. In the case of all the pyrazolo[1,5-*a*]pyrimidines (**14b**, **14e,** and **14j**) and Schif bases (**16c**, **16d**, **16h,** and **16i**) are an inhibitor of the two enzymes (CYP2C9 and

Table 4 ADME and toxicity properties prediction of the pyrazolo[1,5-*a*]pyrimidines (**14b**, **14e,** and **14j**) and pyrazole Schif bases (**16c**, **16d**, **16h,** and **16i**)

= Inhibitor, Non-mutagen, Negative carcinogenicity, and low risk

= Non-inhibitor, Mutagen, Positive carcinogenicity, and high risk

 $=$ Ambiguous, $\qquad \qquad$ = Medium risk

CYP3A4). But all the compounds are non-inhibitor of the two enzymes (CYP2C19 and CYP2D6).

- P-glycoprotein (P-gp) has two functions the frst is a barrier for delivering drugs properly and it is responsible for extruding toxins. All the compounds (**14b**, **14e**, **14j**, **16c**, **16d**, **16h,** and **16i**) were found to be efective.
- The rule for the plasma protein binding (PPB) estimation is the following: when the value was less than 90%; the compound was low activity. While if the value was more than or equal to 90%; the compound was high activity. The two pyrazolo[1,5-*a*]pyrimidines (**14b** and **14e**) and two Schif bases (**16d** and **16i**) had plasma protein binding (PPB) more than 90% (range from 91.17 to 98.23%, highly active), which affects the excretion of the drugs and its efficiency.
- Ames test (mutagenicity), the three pyrazolo[1,5-*a*]pyrimidines (**14b**, **14e**, and **14j**) and three Schif bases (**16c**, **16d**, and **16h**) were predicted to be mutagens.
- Carcinogenicity (mouse) test and the two Schiff bases (**16d** and **16i**) were predicted as positive, while the three pyrazolo[1,5-*a*]pyrimidines (**14b**, **14e**, and **14j**) and the two Schif bases (**16c** and **16h**) presented negatively.
- Carcinogenicity (rat) test, the pyrazolo[1,5-*a*]pyrimidine **14j,** and the three Schif bases (**16d**, **16h,** and **16i**) were predicted as negative.
- In the case of the hERG inhibition, the two compounds **14e** and **16c** showed high risk and the three compounds **14j**, **16h**, and **16i** presented a medium risk. While compounds **14b** and **16d** presented an ambiguous risk.

From the above, all compounds are an inhibitor of the two cytochrome P450 isoforms (CYP2C9 and CYP3A4 enzymes) and P-glycoprotein (P-gp). Besides, some compounds showed highly active toward plasma protein binding (PPB), were predicted as negative against Carcinogenicity (mouse and rat) test, and presented medium risk in the case of the hERG inhibition.

Computational study of the binding mode

One of the most important tools in structure-based drug discovery is molecular docking simulation. It can describe newly designed derivatives' behavior in the binding site of target proteins [[54](#page-23-22)]. Docking study plays an important role and considers one of the most applied simulated screening methods due to its ability to predict and get insights into a protein–ligand complex structure, which is useful information for the successful development of drug targeting and lead optimization [[55](#page-23-23)]. The preferring structure selected from one to another in the docking study related to the highest binding afnity between the ligand and the protein [[56](#page-23-24)]. Our study involved simulation of the most

active derivatives inside both DNA gyrase for bacterial and secreted aspartic protease for fungal promising derivatives using Molecular Operating Environmental (MOE) software version 2008.10. The docking result is represented in Table [5](#page-13-0) and Fig. [6a](#page-14-0)–d.

Study the most active derivatives against bacterial strains inside DNA gyrase

The most active derivatives **14b**, **14e**, **14j**, and **16i** against *Bacillus subtilis* and *Escherichia coli* were docked inside the active site of *S. auras* DNA gyrase (PDB: 2XCT) [[57,](#page-24-0) [58](#page-24-1)]. The co-crystallized ligand (Ciprofloxacin) redocked inside *S. aureus* Gyrase's active site revealed binding energy $S = -9.87$ kcal mol⁻¹. The ligand formed one hydrogen bond donor between Tyr580 and nitrogen of piperazine ring as well as hydrogen bond acceptor between His1081 and oxygen of carboxylate group with bond length 2.35, 2.30 Å, respectively. The most promising pyrazolo[1,5-*a*]pyrimidine-3-carboxamide derivative **14j** with inhibition zone (IZ) ranged between 16-21 mm showed binding energy $S = -25.35$ kcal mol⁻¹, through two hydrogen bond and one arene–cation interaction between His1061 and pyrazole ring. The residue Asp512 formed a side-chain hydrogen bond donor with the amino group of pyrimidine as well as a side-chain hydrogen bond acceptor between Ser1028 and oxygen of methoxy group with bond length and strength 2.83 \AA (11%), 2.99 \AA (38%), respectively Fig. [6](#page-14-0)a.

Furthermore, *N*-phenyl-5-(3-methylphenyl)pyrazolo[1,5 *a*]pyrimidine-3-carboxamide derivative **14b** exhibited binding affinity $S = -23.05$ kcal mol⁻¹, through two hydrogen bonds from a side-chain. These two bonds created between the Ser1028 with the cyano in pyrimidine ring (H-acceptor), Ser1028 with the amino of pyrimidine (H-donor) with bond length 2.89 Å, and 3.05 Å. Besides, arene–cation interaction between His1061 and pyrazole ring Fig. [6b](#page-14-0). Similarly, *N*-phenyl-5-(2,5-dimethoxyphenyl)-pyrazolo[1,5-*a*] pyrimidine-3-carboxamide derivative **14e** displayed binding energy $S = -22.76$ kcal mol⁻¹, by one hydrogen bond side-chain acceptor between Ser1028 and cyano group attached to pyrimidine ring as well as one arene–cation interaction between His1061 and pyrazole ring. Moreover, *N*-(4-chlorophenyl)-1*H*-pyrazole-4-carboxamide **16i** displayed one hydrogen bond donor between Asp510 and NH of pyrazole ring with bond length 2.76 Å and strength 45%.

Finally, the active derivatives **14b**, **14e**, **14j**, and **16i** showed a good binding affinity inside the pocket sites of DNA gyrase ranged between -25.35 to -20.18 kcal mol⁻¹ with diferent binding modes and therefore suggested that these derivatives act as gyrase inhibitors.

			Cpd. No S (kcal mol ⁻¹) Residue Interacting groups			Distance (\AA) Strength (%) Hydrophobic interaction		
			Docking inside the <i>S. aureus</i> DNA gyrase (PDB: 2XCT)					
14 _b	-23.05		Ser1028 CN of pyrimidine	2.89	57	Arg1033, Val511, Ala1032, Asp510, Asp512, Glu435,		
			Ser1028 NH2 of pyrimidine	3.05	15	Tyr1150, Pro1080, Tyr580, Lys581		
			His1061 Pyrazole ring		-			
14e	-22.76	Ser1028	CN OF Pyrimidine	2.88	69	Arg1033, Val511, Asp512, Asp510, Pro1080, Lys581,		
			His1061 Pyrazole ring		$\overline{}$	Tyr1150, Tyr580, Gly582, Lys581		
14j	-25.35	Asp512	NH ₂ of pyrimidine	2.59	50	Asp508, Asp510, Lys581,		
		Ser1028	Oxygen of methoxy			Pro1080, Gly582, Glu585		
			His1061 Pyrazole ring					
16i	-20.18	Asp510	NH of pyrazole	2.76	45	Ser438, Asp437, Gly436, Glu435, Gly582, Lys581, Gly545, Tyr580, Tyr1150		
Cip	-9.87 His1081	Oxygen of carboxylate 2.30		37	Ser1028, Arg1033, Val511, Asp510, Pro1080,			
		Tyr580	Nitrogen of piprazine	2.35	48	Tyr1150, Lys581, Gly545		
			Docking inside the secreted aspartic protease from c. albicans (PDB: IZAP)					
16c	-27.52	Asp218	NH of anisidine der	2.41	37	Tyr225, Gly85, Thr221, Iie305, Gly220, Tyr84,		
		Asp32	Nitrogen of pyrazole	2.72	28	Asn301, Leu216, Glu193, Gly34, Thr13, Ile30,		
		Ser88	Oxygen of methoxy gr	3.20	16	Ile119		
16d	-24.91	Gly34	NH of anisidine der	2.97	22	Gly83, Thr221, Glu193,		
	Asp218		NH of pyrazole	2.56	27	Ala303, Gly85, Asp86, Ile305, Asn301		
		Arg 195	Phenyl of benzamide	$\overline{}$	-			
		Tyr225	Aryl of aldehyde der	$ -$	--			
16h	-27.66	Asp 32	Nitrogen of pyrazole	2.84	24	Gly34, Leu216, Asn301, Thr221, Ile305, Tyr225,		
		Asp32	Nitrogen of pyrazole	2.80	12	Ile119, Ile30, Thrr84, Asp86, Val12, Ser88, Asp32,		
		Asp218	NH of anisidine der	2.41	39	Gly85		
16i	-28.03	Asn301	Oxygen of methoxy	3.18	13	Tyr225, Ile305, Gly85, Asp86, Asp218, Ser35, Gly34,		
		Tyr ₈₄	Aryl of benzamide der –		$\overline{}$	Asn131, Asp32, Thr221, Gly220, Gly83, Glu132, $Ile82$, $Ile133$		
L.Nor	-30.98	Thr221	NH of amide der	3.05	22	Ile223, Tyr84, Leu216, Asp86, Gly220, Glu193,		
		Asp218	OH group	2.31	23	Asp32, Ile123, Ala303, Gly85, Ile305, Asn301, Tyr225, Thr222		

Table 5 Molecular modeling study of the most promising derivatives inside the active sites of *S. aureus* DNA gyrase (PDB: 2XCT) and secreted aspartic protease from *c. albicans* (PDB: IZAP)

Cip.=Ciprofoxacin; L.Nor.=Norleucinamide; (–) arene–cation interaction, (– –) arene–arene interaction

Study the most active derivatives against fungal strains inside secreted aspartic protease

As new possible antifungal drug goals, secreted aspartic protease (Saps) obviously hold promise. Saps are important virulence factors because it includes a wide variety of physiological processes of fungal as well as the various aspects of the interactions between host and fungal [\[59](#page-24-2)]. Similarly, the most active four pyrazole derivatives **16c**, **16d**, **16h**, and **16i** against *A. niger* and *C. albicans* were docked inside the active site of secreted aspartic protease (Saps) (PDB: 1ZAP) [\[58](#page-24-1)]. The co-crystallized ligand (L-Norleucinamide derivative) showed binding energy *S*=−30.98 kcal mol−1 through two hydrogen bond side-chain donor between the residue Thr221 and NH of amide derivative as well as Asp218 with hydroxyl group with bond length 3.05 Å (22%) and 2.31 Å (23%), respectively. The most promising derivative against fungal strains 1*H*-pyrazole-4-carboxamide derivative **16i** exhibited binding energy *S*=−28.03 kcal mol−1 through one side-chain hydrogen bond acceptor. The pyrazole derivative **16i** indicated that a hydrogen bond was formed between the oxygen of the methoxy group and the residue Asn301 (distance 3.18 Å and strength 13%). Meanwhile, an arene–arene interaction was observed between the 4-chlorobenzamide derivative and Tyr84 (Fig. [6c](#page-14-0)). (All docking study figures in supplementary material, Figures S3-S22).

Additionally, the NH of pyrazole and NH of anisidine derivatives in pyrazole **16d** could form two hydrogen bonds donor between Asp218 and Gly34, respectively. Besides, the aryl group of aldehyde derivative created an arene–cation interaction with Tyr225. Further, the docking pose of 5-((2,5-dimethoxybenzylidene)amino)-3-((4 methoxyphenyl)amino)-*N*-(4-chlorophenyl)-1*H*-pyrazole-4-carboxamide (**16h)** demonstrated three favorable

Fig. 6 a 2D and 3D binding mode of pyrazolo[1,5-*a*]pyrimidine-3-carboxamide derivative **14j** inside the active site of *S. auras* DNA gyrase (PDB: 2XCT). **b** 2D and 3D binding mode of pyrazolo[1,5 *a*]pyrimidine-3-carboxamide derivative **14b** inside the active site of *S. auras* DNA gyrase (PDB: 2XCT). **c** 2D and 3D binding mode of

1*H*-pyrazole-4-carboxamide derivative **16i** inside the active site of secreted aspartic protease (Saps) (PDB: 1ZAP). **d** 2D and 3D binding mode of 1*H*-pyrazole-4-carboxamide derivative **16c** inside the active site of secreted aspartic protease (Saps) (PDB: 1ZAP)

hydrogen bonds with binding energy *S*=−27.66 kcal mol−1. Two of them between the nitrogen of pyrazole derivative with the oxygen of carboxylate and oxygen of carbonyl of Asp32 (distance 2.81, 2.84 Å). Besides, one hydrogen bond side-chain donor between the NH of anisidine moiety and Asp218 with bond length 2.41 Å. Furthermore,

Fig. 6 (continued)

N-phenyl-1*H*-pyrazole-4-carboxamide **16c** displayed binding energy *S*=−27.52 kcal mol−1 with three hydrogen bonds between Asp218, Asp32, and Ser88 with NH of anisidine, the nitrogen of pyrazole, and oxygen of methoxy group, respectively (Fig. [6](#page-14-0)d**)**. (All docking study fgures in supplementary material, Figures S3-S22).

Finally, it can be concluded that the docking results revealed that the 1*H*-pyrazole-4-carboxamide derivative **16c**, **16d**, **16h**, and **16i** binding to the active site of secreted aspartic protease (Saps) (PDB: 1ZAP) and may act as inhibitors. Besides, the other derivatives **14b**, **14e**, **14j**, and **16i** were suggested to be DNA gyrase inhibitors with good bind energy scores and diferent binding modes. Overall, these derivatives are suggested to be inhibitors for bacterial and fungal by a diferent mode of actions and thus may participate in antimicrobial activity.

Molecular electrostatic potential (MEP) maps

Our work was extended to investigate the reasons behind a good binding of the most promising derivatives inside the two active pockets. Molecular electrostatic potential (MEP) maps were generated to determine the charge distribution around a molecule. MEP maps provide information about the regions with electron-rich regions (red) and electron deficiency regions, i.e., hydrophobic regions that responsible for forming interactions (hydrogen bonds, arene–arene, and arene–cation interactions) with the residue inside the active site. The MEP was performed using MOE 2008.10, where the active derivatives' geometry optimized using the MMFF94x force feld according to the previously reported method [[60](#page-24-3)]. As represented in Fig. [7](#page-16-0), the positive charge (blue) was located on the nitrogen of pyrimidine and pyrazoles as well as amino, cyano, and NH of both benzamide and anisidine derivatives. Simultaneously, the negative charge (red) was displayed on the oxygen of carbonyl and methoxy groups. Finally, the presence of nucleophiles (red) and electrophile (blue) regions showed that these regions could form hydrogen bonding donors and acceptors and illustrate the binding mode in the docking study (All Molecular Electrostatic Potential (MEP) maps fgures in supplementary material, Figures S23-S29).

Fig. 7 Showed molecular electrostatic potential maps of most promising derivatives to determine the charge distribution around molecules

Conclusion

In summary, new bioactive pyrazolo[1,5-*a*]pyrimidines **14a–j** and pyrazole Schif bases **16a–j** have been designed and synthesized by the reaction of 5-amino-pyrazoles **9a**, **b** with 2-(arylidene)malononitriles **10a–e** or various aldehydes **15a–e**, respectively. The structures of new bioactive pyrazolo[1,5-*a*]pyrimidines **14a–j** and pyrazole Schif bases **16a–j** were confrmed using diferent spectroscopic techniques. The pyrazolo[1,5-*a*]pyrimidines **14a–j** and pyrazole Schif bases **16a–j** were screened for their in vitro antimicrobial activities against four microbes. The three pyrazolo[1,5-*a*]pyrimidines (**14b**, **14e,** and **14j**) and four pyrazole Schif bases (**16c**, **16d**, **16h,** and **16i**) proved to be potent against the four microbes tested, have a wide spectrum efect, and in several cases, their activities exceeded that of the reference drugs used. The structure–activity relationship (SAR) was performed. As well as, physicochemical properties, drug-likeness model score, bioactivity scores, ADME, and toxicity properties were predicted in silico. The prediction result showed the fve compounds (**14b, 16c, 16d, 16h**, and **16i**) agreement with Lipinski's rule of fve and with Veber's rule. Also, in the case of a drug-likeness model score, all the tested compounds (**14b**, **14e, 14j**, **16d**, **16h,** and **16i**) except **16c** showed positive values (range 0.05 to 0.44). Therefore, these compounds may be considered to be drug-like. The pyrazolo[1,5-*a*] pyrimidine **14b** almost shows the highest bioactivity score compared to the other compounds. All compounds are an inhibitor of CYP2C9, CYP3A4 enzymes, and P-glycoprotein (P-gp). Also, some compounds showed highly active $(\geq 90\%)$ toward plasma protein binding (PPB), were predicted as negative against carcinogenicity (mouse and rat) test, and presented medium risk in the hERG inhibition test. Furthermore, molecular docking studies were performed for the most active derivatives against *S. auras* DNA gyrase (PDB: 2XCT) and secreted aspartic protease (Saps) (PDB: 1ZAP) as antibacterial and antifungal targets, respectively, to determine the binding patterns. The most active antibacterial derivatives **14b**, **14e**, **14j**, and **16i** against DNA Gyrase (PDB:2XCT) showed binding energy $S = -25.35$ to -20.18 kcal mol⁻¹. Besides, for promising fungal derivatives **16c**, **16d**, **16h**, and **16i** displayed binding energy $S = -28.03$ to -24.91 kcal mol⁻¹, and both of them showed diferent types of binding mode. The results suggested that these derivatives could act as good antimicrobial agents. MEP maps were performed to provide information about the regions with electron-rich regions (red) and electron defciency regions that responsible for forming interactions with the residue inside the active site and therefore confrmed the binding mode in the docking study. In the future, these compounds are possibly suitable

temples for the development of a novel antibacterial and antifungal therapeutic drug.

Materials and methods

Chemistry

All melting points were measured on a Gallenkamp melting point apparatus and are uncorrected. The IR spectra were recorded (KBr disk) on a PerkinElmer 1650 FTIR instrument. ¹H NMR (500 or 400 MHz) and ¹³C NMR (125 or 100 MHz) spectra were recorded on a JEOL spectrometer using DMSO d_6 as solvent and tetramethylsilane (TMS) as an internal standard. Chemical shifts (*δ*-values) are reported in ppm. Mass spectra were recorded on a Varian MAT 112 spectrometer at 70 eV. Elemental analyses were performed at the Micro Analytical Center, Cairo University, Egypt.

The progress of the reactions was monitored by thin-layer chromatography (TLC) using aluminum sheets coated with silica gel F_{254} (Merck), viewing under a short-wavelength UV lamp effected detection. All evaporations were performed under reduced pressure at 40 °C.

3-Methoxybenzaldehyde (**15a**), 4-(dimethylamino)benzaldehyde (**15b**), 2,5-dimethoxybenzaldehyde (**15c**), 4-chloro-3-nitrobenzaldehyde (**15d**), and thiophene-2-carbaldehyde (**15e**) were of Merck AR grade, Germany.

Synthesis of 5-amino-N-aryl-3-(4-methoxyphenylamino)- 1H-pyrazole-4-carboxamide (**9a, b**) [\[41](#page-23-9)] and *2-(arylidene) malononitriles* (**10a–e**) [\[42,](#page-23-10) [43](#page-23-11)] were prepared according to the literature procedure.

Synthesis of 7‑amino‑6‑cyano‑5‑aryl‑2‑(4‑methoxyphenyl amino)‑*N***‑aryl‑pyrazolo[1,5‑***a***]pyrimidine‑3‑carboxamides (14a–j)**

A mixture of 5-amino-*N*-aryl-1*H*-pyrazole-4-carboxamide derivatives **9a**, **b** (0.01 mol) with 2-(arylidene)malononitriles **10a–e** (0.01 mol) as [2-(4-methylbenzylidene)malononitrile (**10a**), 2-(3-methylbenzylidene)malononitrile (**10b**), 2-(3-methyoxybenzylidene)malononitrile (**10c**), 2-(3-chlorobenzylidene)malononitrile (**10d**), and 2-(2,5-dimethoxybenzylidene)malononitrile (**10e**)] and a catalytic amount of triethylamine (four drops) in absolute ethanol (30 mL) was refuxed for 6 h. After cooling, the solvent was concentrated under reduced pressure and the solid obtained was collected and recrystallized from ethanol to give the corresponding pyrazolo[1,5-*a*]pyrimidines **14a–j**.

7‑Amino‑6‑cyano‑2‑(4‑methoxyphenylamino)‑*N***‑phe‑ nyl‑5‑(4‑methylphenyl)pyrazolo[1,5‑***a***]pyrimidine‑3‑car‑ boxamide (14a)**

Yellow crystals; M.p.: > 300 °C; Yield (80%); IR (KBr) $v_{\text{max}}/$ cm⁻¹ 3428, 3304 (NH₂, 2NH), 2213 (C≡N), 1641 (C=O), 1597 (C=N), 1551 (C=C); ¹H NMR (500 MHz, DMSO d_6 , *δ*/ppm): 2.43 (s, 3H, CH₃), 3.74 (s, 3H, OCH₃), 6.90 (d, 2H, *J*= 9.0 Hz, *p*-methoxyphenyl ring), 7.09 (t, 1H, *J*=7.5 Hz, phenyl ring), 7.37 (t, 2H, *J*=7.8 Hz, phenyl ring), 7.44 (d, 2H, *J*=8.2 Hz, *p*-methylphenyl ring), 7.60 (d, 2H, *J*=7.7 Hz, phenyl ring), 7.84 (d, 2H, *J*=9.0 Hz, *p*-methoxyphenyl ring), 7.90 (d, 2H, *J*=8.2 Hz, *p*-methylphenyl ring), 9.00 (s, br, 2H, NH₂ exchangeable by D₂O), 9.23, 10.14 (2s, 2H, 2NH exchangeable by D_2O); ¹³C NMR (125 MHz, DMSO-*d*₆, δ ppm): 21.15 (C, CH₃), 55.26 (C, OCH₃), 74.88, 89.09, 114.26, 118.87, 119.06, 123.54, 123.78, 128.69, 129.24, 129.34, 133.40, 133.98, 138.46, 141.08, 145.70, 149.49, 153.98, 156.32, 161.47, 162.29 (26C); MS (*m/z*, %): 489 (M+, 17.65%), 396 (base peak, 100%); Anal. Calcd. (%) for $C_{28}H_{23}N_7O_2$ (489.53): C, 68.70; H, 4.74; N, 20.03. Found: C, 68.98; H, 4.57; N, 20.29%.

7‑Amino‑6‑cyano‑2‑(4‑methoxyphenylamino)‑*N***‑phe‑ nyl‑5‑(3‑methylphenyl)pyrazolo[1,5‑***a***]pyrimidine‑3‑car‑ boxamide (14b)**

Yellow crystals; M.p.: > 300 °C; Yield (77%); IR (KBr) $v_{\text{max}}/$ cm⁻¹ 3428, 3319 (NH₂, 2NH), 2216 (C≡N), 1656 (C=O), 1599 (C=N), 1554 (C=C); ¹H NMR (500 MHz, DMSO d_6 , *δ*/ppm): 2.48 (s, 3H, CH₃), 3.79 (s, 3H, OCH₃), 6.94 (d, 2H, *J*=9.5 Hz, *p*-methoxyphenyl ring), 7.13 (d, 1H, *J*=7.7 Hz, *m*-methylphenyl ring), 7.39 (t, 2H, *J*=8.2 Hz, phenyl ring), 7.50 (d, 1H, *J*=7.9 Hz, *m*-methylphenyl ring), 7.56 (t, 1H, *J*=7.9 Hz, phenyl ring), 7.62 (d, 4H, *J*=8.6 Hz, *m*-methylphenyl ring + phenyl ring), 7.86 (d, 2H, $J = 9.2$ Hz, *p*-methoxyphenyl ring), 9.12 (s, br, 2H, NH₂ exchangeable by D₂O), 9.23, 10.21 (2s, 2H, 2NH exchangeable by D₂O); Anal. Calcd. (%) for $C_{28}H_{23}N_7O_2$ (489.53): C, 68.70; H, 4.74; N, 20.03. Found: C, 68.53; H, 4.48; N, 20.27%.

7‑Amino‑6‑cyano‑5‑(3‑methoxyphenyl)‑2‑(4‑methoxyphe nylamino)‑*N***‑phenylpyrazolo[1,5‑***a***]pyrimidine‑3‑carboxa‑ mide (14c)**

Yellow crystals; M.p.: 258–260 °C; Yield (72%); IR (KBr) $v_{\text{max}}/\text{cm}^{-1}$ 3433, 3332 (NH₂, 2NH), 2212 (C≡N), 1652 $(C=0)$, 1595 $(C=N)$, 1547 $(C=C)$; ¹H NMR (500 MHz, DMSO-*d*₆, *δ*/ppm): 3.74, 3.86 (2s, 6H, 2OCH₃), 6.90 (d, 2H, *J*=9.0 Hz, *p*-methoxyphenyl ring), 7.09 (t, 1H, *J*=7.4 Hz, phenyl ring), 7.18–7.21 (m, 1H, *m*-methoxyphenyl ring), 7.36 (t, 2H, *J*=7.9 Hz, phenyl ring), 7.54–7.56 (m, 3H, *m*-methoxyphenyl ring), 7.59 (d, 2H, *J* =7.7 Hz, phenyl

ring), 7.85 (d, 2H, *J*=9.0 Hz, *p*-methoxyphenyl ring), 9.09 (s, br, 2H, NH₂ exchangeable by D_2O), 9.23, 10.14 (2s, 2H, 2NH exchangeable by D_2O ; ¹³C NMR (125 MHz, DMSO*d*6, *δ*/ppm): 55.43, 56.11 (2C, 2OCH3), 75.09, 89.19, 113.86, 114.24, 115.98, 116.90, 118.89, 119.06, 120.98, 123.51, 129.14, 129.87, 133.30, 137.95, 138.38, 145.57, 149.40, 154.00, 156.36, 159.26, 160.78, 162.17 (26C); MS (*m/z*, %): 505 (M^+ , 41.43%), 93 (base peak, 100%); Anal. Calcd. (%) for $C_{28}H_{23}N_7O_3$ (505.53): C, 66.52; H, 4.59; N, 19.39. Found: C, 66.40; H, 4.31; N, 19.65%.

7‑Amino‑5‑(3‑chlorophenyl)‑6‑cyano‑2‑(4‑methoxyphenyl amino)‑*N***‑phenylpyrazolo[1,5‑***a***]pyrimidine‑3‑carboxamide (14d) [[61\]](#page-24-4)**

Orange crystals; M.p. > 300 °C; Yield (74%); IR (KBr) $v_{\text{max}}/$ cm⁻¹ 3442, 3299 (NH₂, 2NH), 2212 (C≡N), 1665 (C=O), 1632 (C=N), 1598 (C=C); ¹H NMR (400 MHz, DMSO- d_6 , *δ*/ppm) 3.78 (s, 3H, OCH3), 6.89 (d, 2H, *J*=9.0 Hz, *p*-methoxyphenyl ring), 7.07 (t, 1H, *J*=7.4 Hz, phenyl ring), 7.33 (t, 2H, *J*=8.3 & 7.5 Hz, phenyl ring), 7.60 (d, 2H, *J*=8.0 Hz, phenyl ring), 7.62 (d, 1H, *J*=2.9 Hz, *m*-chlorophenyl ring), 7.76 (d, 2H, *J*=9.0 Hz, *p*-methoxyphenyl ring), 7.93–7.96 (m, 1H, *m*-chlorophenyl ring), 8.02 (s, 1H, *m*-chlorophenyl ring), 8.06 (d, 1H, *J*=2.8 Hz, *m*-chlorophenyl ring), 8.93 (s, br, 2H, NH₂ exchangeable by D₂O), 9.28, 10.06 (2s, 2H, 2NH exchangeable by D_2O ; ¹³C NMR (100 MHz, DMSO d_6 CDCl₃, *δ*/ppm): 55.45 (1C, OCH₃), 75.14, 89.98, 114.40, 115.97, 119.20, 119.29, 123.66, 127.23, 128.77, 129.21, 130.48, 130.86, 133.59, 134.16, 138.73, 138.78, 145.75, 149.60, 154.38, 157.00, 159.45, 162.53 (26C); MS (*m/z*, %): 511 (M^{+2} , 2.69), 509 (M^{+} , 8.45), 368 (base peak, 100%); Anal. Calcd. (%) for $C_{27}H_{20}CIN_7O_2$ (509.95): C, 63.59; H, 3.95; N, 19.23. Found: C, 63.50; H, 4.00; N, 19.27%.

7‑Amino‑6‑cyano‑5‑(2,5‑dimethoxyphenyl)‑2‑(4‑methoxy‑ phenylamino)‑*N***‑phenylpyrazolo[1,5‑***a***]pyrimidine‑3‑car‑ boxamide (14e)**

Orange crystals; M.p. 210–212 °C; Yield (75%); IR (KBr) $v_{\text{max}}/\text{cm}^{-1}$ 3429, 3321 (NH₂, 2NH), 2217 (C≡N), 1654 $(C=0)$, 1595 $(C=N)$, 1552 $(C=C)$; ¹H NMR (500 MHz, DMSO- d_6 , $δ$ /ppm): 3.72, 3.82, 3.88 (3s, 9H, 3OCH₃), 6.90 (d, 2H, *J*=7.5 Hz, *p*-methoxyphenyl ring), 7.07–7.22 (m, 4H, 2,5-dimethoxyphenyl ring+phenyl ring), 7.34 (t, 2H, *J*=7.3 Hz, phenyl ring), 7.51 (d, 1H, *J*=7.8 Hz, 2,5-dimethoxyphenyl ring), 7.65 (d, 2H, *J*=7.8 Hz, *p*-methoxyphenyl ring), 7.85 (d, 1H, *J*=8.3 Hz, 2,5-dimethoxyphenyl ring), 8.68 (s, br, 2H, NH₂ exchangeable by D₂O), 9.34, 10.06 (2s, 2H, 2NH exchangeable by D_2O); Anal. Calcd. (%) for $C_{29}H_{25}N_7O_4$ (535.55): C, 65.04; H, 4.71; N, 18.31. Found: C, 65.24; H, 4.45; N, 18.65%.

7‑Amino‑*N***‑(4‑chlorophenyl)‑6‑cy‑ ano‑2‑(4‑methoxyphenylamino)‑5‑(4‑methylphenyl) pyrazolo[1,5‑***a***]pyrimidine‑3‑carboxamide (14f)**

Yellow crystals; $M.p.: > 300 °C$; Yield (76%); IR (KBr) $v_{\text{max}}/\text{cm}^{-1}$ 3439, 3296, 3234 (NH₂, 2NH), 2214 (C≡N), 1639 (C=O), 1595 (C=N), 1564 (C=C); ¹H NMR (500 MHz, DMSO- d_6 , δ /ppm): 2.38 (s, 3H, CH₃), 3.70 (s, 3H, OCH3), 6.85 (d, 2H, *J* = 7.3 Hz, *p*-methoxyphenyl ring), 7.36 (d, 4H, *J* = 7.5 Hz, ArH), 7.55 (d, 2H, *J*=6.2 Hz, ArH), 7.74 (d, 2H, *J*=7.1 Hz, ArH), 7.83 (d, 2H, *J* = 7.7 Hz, *p*-methoxyphenyl ring), 8.76 (s*, br*, 2H, NH2), 9.06, 10.15 (2s, 2H, NH); MS (*m/z*, %): 525 (M+2, 25.41), 523 (M+, 76.85), 93 (base peak, 100%); Anal. Calcd. (%) for $C_{28}H_{22}C1N_7O_2$ (523.97): C, 64.18; H, 4.23; N, 18.71. Found: C, 64.35; H, 4.05; N, 18.48.

7‑Amino‑*N***‑(4‑chlorophenyl)‑6‑cy‑ ano‑2‑(4‑methoxyphenylamino)‑5‑(3‑methylphenyl) pyrazolo[1,5‑***a***]pyrimidine‑3‑carboxamide (14g)**

Yellow crystals; M.p.: > 300 °C; Yield (71%); IR (KBr) $v_{\text{max}}/$ cm⁻¹ 3419, 3335 (NH₂, 2NH), 2216 (C≡N), 1651 (C=O), 1592 (C=N), 1554 (C=C); ¹H NMR (500 MHz, DMSO- d_6 , *δ*/ppm): 2.44 (s, 3H, CH₃), 3.74 (s, 3H, OCH₃), 6.89 (d, 2H, *J*=8.9 Hz, *p*-methoxyphenyl ring), 7.40 (d, 2H, *J*=8.9 Hz, *p*-chlorophenyl ring), 7.44 (d, 1H, *J*=7.7 Hz, *m*-methylphenyl ring), 7.49 (t, 1H, *J*=7.6 Hz, *m*-methylphenyl ring), 7.58 (d, 2H, *J*=8.8 Hz, *p*-chlorophenyl ring), 7.77 (d, 1H, *J*=7.7 Hz, *m*-methylphenyl ring), 7.80 (s, 1H, *m*-methylphenyl ring), 7.82 (d, 2H, *J*=8.9 Hz, *p*-methoxyphenyl ring), 9.00 (s, br, 2H, NH₂ exchangeable by D₂O), 9.12, 10.21 (2s, 2H, 2NH exchangeable by D_2O); Anal. Calcd. (%) for $C_{28}H_{22}CIN_{7}O_{2}$ (523.97): C, 64.18; H, 4.23; N, 18.71. Found: C, 64.02; H, 4.54; N, 18.52%.

7‑Amino‑*N***‑(4‑chlorophenyl)‑6‑cy‑ ano‑5‑(3‑methoxyphenyl)‑2‑(4‑methoxyphenylamino) pyrazolo[1,5‑***a***]pyrimidine‑3‑carboxamide (14h)**

Yellow crystals; M.p.: > 300 °C; Yield (73%); IR (KBr) $v_{\text{max}}/$ cm⁻¹ 3436, 3307 (NH₂, 2NH), 2213 (C≡N), 1643 (C=O), 1603 (C=N), 1544 (C=C); ¹H NMR (500 MHz, DMSO- d_6 , *δ*/ppm): 3.74, 3.86 (2s, 6H, 2OCH3), 6.90 (d, 2H, *J*=9.1 Hz, *p*-methoxyphenyl ring), 7.18–7.20 (m, 1H, *m*-methoxyphenyl ring), 7.42 (d, 2H, *J*=8.8 Hz, *p*-chlorophenyl ring), 7.51–7.55 (m, 3H, *J* = 7.7 Hz, *m*-methoxyphenyl ring), 7.62 (d, 2H, *J*=8.9 Hz, *p*-chlorophenyl ring), 7.84 (d, 2H, $J=9.0$ Hz, *p*-methoxyphenyl ring), 8.99 (s, br, 2H, NH₂) exchangeable by D_2O , 9.17, 10.18 (2s, 2H, 2NH exchangeable by D₂O); Anal. Calcd. (%) for $C_{28}H_{22}CIN_{7}O_{3}$ (539.97):

C, 62.28; H, 4.11; N, 18.16. Found: C, 62.35; H, 4.05; N, 18.34%.

7‑Amino‑5‑(3‑chlorophenyl)‑*N***‑(4‑chlorophenyl)‑6‑cy‑ ano‑2‑(4‑methoxyphenylamino)pyrazolo[1,5‑***a***]pyrimi‑ dine‑3‑carboxamide (14i) [[61](#page-24-4)]**

Yellow crystals; M.p. 252–254 ºC; Yield (71%); IR (KBr) $v_{\text{max}}/\text{cm}^{-1}$ 3397, 3308 (NH₂, 2NH), 2211 (C≡N), 1652 $(C=0)$, 1613 $(C=N)$, 1547 $(C=C)$; ¹H NMR (400 MHz, DMSO-*d*₆, CDCl₃, *δ*/ppm): 3.73 (s, 3H, OCH₃), 6.82 (d, 2H, *J*=8.8 Hz, ArH), 7.16–7.22 (m, 2H, ArH), 7.52 (d, 2H, *J*=7.8 Hz, ArH), 7.60 (d, 2H, *J*=8.8 Hz, ArH), 7.66 (d, 1H, *J*=8.8 Hz, ArH), 7.81–7.91 (m, 3H, ArH), 8.63 (s*, br*, 2H, NH₂ exchangeable by D₂O), 9.14, 10.02 (2s, 2H, 2NH exchangeable by D₂O); ¹³C NMR (100 MHz, DMSO- d_6) CDCl₃, $δ$ /ppm): 55.52 (1C, OCH₃), 75.48, 89.92, 114.36, 115.87, 119.28, 120.51, 127.13, 127.89, 128.34, 129.01, 130.38, 131.00, 133.49, 134.24, 134.93, 137.49, 145.89, 149.54, 154.42, 156.98, 161.21, 163.20 (26C); Anal. Calcd. (%) for $C_{27}H_{19}Cl_2N_7O_2$ (544.39): C, 59.57; H, 3.52; N, 18.01. Found: C, 59.49; H, 3.60; N, 17.95%.

7‑Amino‑*N***‑(4‑chlorophenyl)‑6‑cy‑**

ano‑5‑(2,5‑dimethoxyphenyl)‑2‑(4‑methoxyphenylamino) pyrazolo[1,5‑*a***]pyrimidine‑3‑carboxamide (14j)**

Yellow crystals; M.p.: > 300 °C; Yield (74%); IR (KBr) $v_{\text{max}}/$ cm⁻¹ 3446, 3310 (NH₂, 2NH), 2215 (C≡N), 1637 (C=O), 1596 (C=N), 1568 (C=C); ¹H NMR (500 MHz, DMSO*d*6, *δ*/ppm): 3.70, 3.75, 3.78 (3s, 9H, 3OCH3), 6.86 (d, 2H, *J* = 9.5 Hz, *p*-methoxyphenyl ring), 7.10–7.17 (m, 3H, 2,5-dimethoxyphenyl ring), 7.34 (d, 2H, *J*=9.2 Hz, *p*-chlorophenyl ring), 7.48 (d, 2H, *J*=9.2 Hz, *p*-chlorophenyl ring), 7.78 (d, 2H, *J*=9.3 Hz, *p*-methoxyphenyl ring), 8.97 (s*, br*, 2H, NH₂ exchangeable by D₂O), 9.11, 10.04 (2s, 2H, 2NH exchangeable by D_2O); ¹³C NMR (125 MHz, DMSO- d_6) *δ*/ppm): 55.12, 55.68, 55.93 (3C, 3OCH3), 78.02, 88.84, 104.95, 113.21, 114.13, 115.17, 117.23, 118.96, 120.38, 126.91, 128.90, 133.25, 137.20, 145.72, 148.51, 150.65, 153.07, 153.91, 156.07, 159.97, 159.99, 162.09 (26C); Anal. Calcd. (%) for $C_{29}H_{24}CN_7O_4$ (570.00): C, 61.11; H, 4.24; N, 17.20. Found: C, 60.90; H, 4.39; N, 17.48%.

Synthesis of Schif bases 16a‑j

A mixture of 5-amino-*N*-aryl-1*H*-pyrazole-4-carboxamide derivatives **9a**, **b** (0.01 mol) and various aldehydes **15a–e** (0.01 mol) {namely: 3-methoxybenzaldehyde (**15a**), 4-(dimethylamino)benzaldehyde (**15b**), 2,5-dimethoxybenzaldehyde (**15c**), 4-chloro-3-nitrobenzaldehyde (**15d**), or thiophene-2-carbaldehyde **(15e)**} with a catalytic amount of glacial acetic acid (1 mL) in absolute ethanol (25 mL) was refuxed for 2 h and then left to cool. The solid product was filtered off, dried and finally recrystallized from ethanol to aford the corresponding Schif bases **16a–j**.

5‑(3‑Methoxybenzylideneamino)‑3‑(4‑methoxyphenylamin o)‑*N***‑phenyl‑1***H***‑pyrazole‑4‑carboxamide (16a)**

Yellow crystals; M.p.: 201–203 °C; Yield (75%); IR (KBr) $v_{\text{max}}/\text{cm}^{-1}$ 3434, 3229 (3NH), 1656 (C=O), 1612 (C=N), 1587 (C=C); ¹H NMR (500 MHz, DMSO- d_6 , δ /ppm): 3.73, 3.88 (2s, 6H, 2OCH3), 6.91 (d, 2H, *J*=8.3 Hz, *p*-methoxyphenyl ring), 7.08 (t, 1H, *J* =7.4 Hz, phenyl ring), 7.23 (d, 1H, *J*=7.5 Hz, *m*-methoxyphenyl ring), 7.36 (t, 2H, *J*=7.9 Hz, phenyl ring), 7.55 (t, 1H, *J*=7.8 Hz, *m*-methoxyphenyl ring), 7.62–7.64 (m, 4H, phenyl ring+*m*-methoxyphenyl ring), 7.68 (d, 2H, *J*=8.0 Hz, *p*-methoxyphenyl ring), 8.71 (s*,* 1H, –CH=N–), 9.05, 9.99, 12.80 (3s, 3H, 3NH exchangeable by D₂O); ¹³C NMR (125 MHz, DMSO- d_6 , δ / ppm): 55.27, 55.34 (2C, 2OCH3), 83.17, 112.33, 114.51, 118.98, 119.82, 123.30, 128.54, 129.08, 129.37, 130.53, 135.02, 138.54, 146.21, 152.25, 155.38, 156.89, 159.83, 161.70, 162.69 (23C); MS (*m/z*, %): 441 (M+, 25.67%), 83 (base peak, 100%); Anal. Calcd. (%) for $C_{25}H_{23}N_5O_3$ (441.48): C, 68.01; H, 5.25; N, 15.86. Found: C, 68.17; H, 5.06; N, 15.69%.

5‑(4‑(Dimethylamino)benzylideneamino)‑3‑(4‑methoxyphe nylamino)‑*N***‑phenyl‑1***H***‑pyrazole‑4‑carboxamide (16b) [\[62](#page-24-5)]**

Yellow crystals; M.p.: 216–218 °C; Yield (87%); IR (KBr) $v_{\text{max}}/\text{cm}^{-1}$ 3445, 3292 (3NH), 1652 (C=O), 1614 (C=N), 1592 (C=C); ¹H NMR (400 MHz, DMSO- d_6 , δ/ppm): 3.01 (s, 6H, N(CH₃)₂), 3.66 (s, 3H, OCH₃), 6.83 (d, 4H, *J*=8.9 Hz, ArH), 7.02 (t, 1H, ArH), 7.33 (t, 2H, ArH), 7.48 (2H, ArH), 7.60 (d, 2H, *J*=8.4 Hz, ArH), 7.80 (d, 2H, *J*=8.7 Hz, ArH), 8.67 (s, 1H, –N=CH–), 9.03, 10.07, 12.59 (3s, 3H, 3NH exchangeable by D_2O); ¹³C NMR (100 MHz, DMSO- d_6 , δ /ppm) δ 40.10 (2C, N (CH₃)₂ under solvent DMSO), 55.48 (OCH₃), 92.74, 112.34, 115.12, 120.08, 121.82, 126.58, 128.07, 128.43, 129.19, 132.02, 132.84, 135.21, 152.78, 153.40, 153.84, 154.45, 160.57, 162.06 (23C); Anal. Calcd. (%) for $C_{26}H_{26}N_6O_2$ (454.52): C, 68.70; H, 5.77; N, 18.49. Found: C, 68.75; H, 5.70; N, 18.55%.

5‑(2,5‑Dimethoxybenzylideneamino)‑3‑(4‑methoxyphenyla mino)‑*N***‑phenyl‑1***H***‑pyrazole‑4‑carboxamide (16c)**

Orange crystals; M.p.: 234–236 °C; Yield (69%); IR (KBr) $v_{\text{max}}/\text{cm}^{-1}$ 3389, 3320 (3NH), 1654 (C=O), 1594 (C=N), 1552 (C=C); ¹H NMR (500 MHz, DMSO- d_6 , δ/ppm): 3.72, 3.83, 3.89 (3s, 9H, 3OCH3), 6.91 (d, 2H, *J*=8.0 Hz, *p*-methoxyphenyl ring), 7.08–7.35 (m, 7H, Ar–H), 7.50 (s, 1H, 2,5-dimethoxyphenyl ring), 7.65 (d, 2H, *J*=8.3 Hz,

p-methoxyphenyl ring), 8.69 (s*,* 1H, –CH=N–), 9.47, 10.18, 13.01 (3s, 3H, 3NH exchangeable by D_2O); Anal. Calcd. (%) for $C_{26}H_{25}N_5O_4$ (471.51): C, 66.23; H, 5.34; N, 14.85. Found: C, 66.62; H, 5.17; N, 14.68%.

5‑(4‑Chloro‑3‑nitrobenzylideneamino)‑3‑(4‑methoxypheny lamino)‑*N***‑phenyl‑1***H***‑pyrazole‑4‑carboxamide (16d)**

Buf crystals; M.p.: 252–254 °C; Yield (72%); IR (KBr) $v_{\text{max}}/\text{cm}^{-1}$ 3431, 3235 (3NH), 1653 (C=O), 1615 (C=N), 1513 (C=C); ¹H NMR (500 MHz, DMSO- d_6 , *δ*/ppm): 3.70 (s, 3H, OCH3), 6.89 (d, 2H, *J*=8.3 Hz, *p*-methoxyphenyl ring), 7.06 (t, 1H, *J*=7.3 Hz, phenyl ring), 7.31–7.34 (m, 4H, phenyl ring), 7.67 (d, 2H, *J*=7.9 Hz, *p*-methoxyphenyl ring), 8.00 (d, 1H, *J*=8.3, 4-chloro-3-nitrophenyl ring), 8.30 (d, 1H, *J*=8.3, 4-chloro-3-nitrophenyl ring), 8.67 (s, 1H, 4-chloro-3-nitrophenyl ring), 8.76 (s*,* 1H, –CH=N–), 9.13, 9.79, 12.80 (3s, 3H, 3NH exchangeable by D₂O); ¹³C NMR (125 MHz, DMSO- d_6 , $δ$ /ppm): 55.23 (1C, OCH₃), 96.78, 114.61, 118.92, 119.73, 121.51, 123.06, 125.26, 126.27, 128.87, 132.72, 133.81, 134.09, 138.39, 147.84, 152.02, 153.37, 157.62, 160.86, 162.42 (23C); Anal. Calcd. (%) for $C_{24}H_{19}CIN_6O_4$ (490.90): C, 58.72; H, 3.90; N, 17.12; Found: C, 58.99; H, 3.79; N, 17.28%.

3‑(4‑Methoxyphenylamino)‑*N***‑phenyl‑5‑(thiophen‑2‑yl‑ methyleneamino)‑1***H***‑pyrazole‑4‑carboxamide (16e)**

Yellow crystals; M.p.: 212–214 °C; Yield (67%); IR (KBr) $v_{\text{max}}/\text{cm}^{-1}$ 3433, 3272 (3NH), 1652 (C=O), 1594 (C=N), 1547 (C=C); ¹H NMR (500 MHz, DMSO- d_6 , *δ*/ppm): 3.73 (s, 3H, OCH3), 6.90 (d, 2H, *J*=7.1 Hz, *p*-methoxyphenyl ring), 7.09 (d, 1H, *J*=5.3 Hz, thiophene ring), 7.33–7.40 (m, 5H, phenyl ring), 7.72 (d, 2H, *J*=7.0 Hz, *p*-methoxyphenyl ring), 7.91 (d, 1H, *J*=3.6 Hz, thiophene ring), 8.05 (s, 1H, thiophene ring), 8.68 (s*,* 1H, –CH=N–), 9.19, 9.98, 12.50 (3s, 3H, 3NH exchangeable by D_2O); ¹³C NMR (125 MHz, DMSO-*d*₆, $δ$ /ppm): 55.79 (1C, OCH₃), 89.70, 115.01, 119.23, 123.68, 129.62, 129.74, 133.89, 134.14, 134.29, 136.99, 139.23, 141.83, 155.79, 156.45, 157.46, 159.54, 163.28 (21C); MS (*m/z*, %): 417 (M+, 18.85%), 186 (base peak, 100%); Anal. Calcd. (%) for $C_{22}H_{19}N_5O_2S$ (417.48): C, 63.29; H, 4.59; N, 16.78; Found: C, 63.45; H, 4.39; N, 16.85%.

*N***‑(4‑Chlorophenyl)‑5‑(3‑methoxybenzylideneamino)‑3‑(4‑ methoxyphenylamino)‑1***H***‑pyrazole‑4‑carboxamide (16f)**

Yellow crystals; M.p.: 205–207 $°C$; Yield (73%); IR (KBr) $v_{\text{max}}/\text{cm}^{-1}$ 3434, 3229 (3NH), 1656 (C=O), 1612 $(C=N)$, 1587 $(C=C)$; ¹H NMR (500 MHz, DMSO- d_6 , δ / ppm): 3.73, 3.87 (2s, 6H, 2OCH3), 6.90 (d, 2H, *J*=8.9 Hz, *p*-methoxyphenyl ring), 7.21, 7.23 (dd, 1H, *J*=8.2, 1.8 Hz, *m*-methoxyphenyl ring), 7.40 (d, 4H, *J*=8.9 Hz, *p*-chlorophenyl ring), 7.53 (t, 1H, *J*=7.9 Hz, *m*-methoxyphenyl ring), 7.59 (s, 1H, *m*-methoxyphenyl ring), 7.62 (d, 1H, *J*=2.4 Hz, *m*-methoxyphenyl ring), 7.69 (d, 2H, *J*=8.9 Hz, *p*-methoxyphenyl ring), 8.66 (s*,* 1H, –CH=N–), 9.03, 10.02, 12.78 (3s, 3H, 3NH exchangeable by D₂O); ¹³C NMR (125 MHz, DMSO-*d*₆, *δ*/ppm): 55.19, 55.23 (2C, 2OCH₃), 92.61, 112.24, 114.39, 119.22, 119.53, 120.41, 122.48, 126.69, 128.34, 128.73, 130.28, 133.79, 136.17, 137.46, 151.21, 154.04, 159.74, 162.03, 162.65 (23C); Anal. Calcd. (%) for $C_{25}H_{22}CIN_{5}O_{3}$ (475.93): C, 63.09; H, 4.66; N, 14.72; Found: C, 63.34; H, 4.53; N, 14.43%.

*N***‑(4‑Chlorophenyl)‑5‑(4‑(dimethylamino)benzylideneamin o)‑3‑(4‑methoxyphenylamino)‑1***H***‑pyrazole‑4‑carboxamide (16g) [[62\]](#page-24-5)**

Yellow crystals; M.p.: 209–211 °C, Yield (86%); IR (KBr) $v_{\text{max}}/\text{cm}^{-1}$ 3431, 3275 (3NH), 1662 (C=O), 1609 (C=N), 1581 (C=C); ¹H NMR (400 MHz, DMSO- d_6 , δ ppm): 3.38 $(s, 6H, N(CH_3), 3.71 (s, 3H, OCH_3), 6.85 (d, 2H, J=9 Hz,$ *p*-dimethylaminophenyl ring), 6.88 (d, 2H, $J = 8.9$ Hz, *p*-methoxyphenyl ring), 7.40 (d, 2H, *J*=8.6 Hz, *p*-chlorophenyl ring), 7.45 (d, 2H, *J*=8.5 Hz, *p*-chlorophenyl ring), 7.68 (d, 2H, *J*=8.6 Hz, *p*-methoxyphenyl ring), 7.82 (d, 2H, *J*=8.5 Hz, *p*-dimethylaminophenyl ring), 8.66 (s*,* 1H, –CH=N–), 8.74, 10.24, 12.24 (3s, 3H, 3NH exchangeable by D₂O); ¹³C NMR (100 MHz, DMSO- d_6 , δ ppm): 40.06 (2C, N (CH₃)₂ under solvent DMSO), 55.67 (1C, OCH₃), 92.48, 112.27, 114.78, 120.87, 121.08, 122.12, 127.29, 129.43, 132.02, 133.35, 135.00, 152.88, 153.70, 153.89, 154.25, 161.19, 163.66 (23C); Anal. Calcd. (%) for $C_{26}H_{25}CIN_6O_2$ (488.97): C, 63.86; H, 5.15; N, 17.19; Found: C, 63.95; H, 5.10; N, 17.25%.

*N***‑(4‑Chlorophenyl)‑5‑(2,5‑dimethoxybenzylideneamino) ‑3‑(4‑methoxyphenylamino)‑1***H***‑pyrazole‑4‑carboxamide (16h)**

Orange crystals; M.p.: 250–252 °C; Yield (74%); IR (KBr) $v_{\text{max}}/\text{cm}^{-1}$ 3432, 3251 (3NH), 1653 (C=O), 1597 (C=N), 1544 (C=C); ¹H NMR (500 MHz, DMSO- d_6 , δ /ppm): 3.73, 3.82, 3.88 (3s, 9H, 3OCH3), 6.89, 6.91 (dd, 2H, *J*=2.8 & 9.5 Hz, *p*-methoxyphenyl ring), 7.18 (d, 1H, *J*=2.8 Hz, 2,5-dimethoxyphenyl ring), 7.21 (d, 1H, *J* = 2.7 Hz, 2,5-dimethoxyphenyl ring), 7.38 (d, 2H, *J*=9.2 Hz, *p*-chlorophenyl ring), 7.42 (d, 2H, *J*= 9.0 Hz, *p*-chlorophenyl ring), 7.61 (s, 1H, 2,5-dimethoxyphenyl ring), 7.66, 7.68 (dd, 2H, *J*=9.2, 2.7 Hz, *p*-methoxyphenyl ring), 8.64 (s*,* 1H, –CH=N–), 9.34, 10.13, 11.74 (3s, 3H, 3NH exchangeable by D₂O); ¹³C NMR (125 MHz, DMSO- d_6 , δ /ppm): 55.26, 55.59, 56.47 (3C, 3OCH3), 92.55, 109.44, 114.21, 114.42, 119.00, 120.58, 121.94, 123.30, 126.66, 128.83,

134.18, 137.62, 153.41, 153.81, 154.87, 156.70, 156.90, 160.72, 162.88 (23C); Anal. Calcd. (%) for $C_{26}H_{24}CIN_5O_4$ (505.95): C, 61.72; H, 4.78; N, 13.84; Found: C, 61.90; H, 4.51; N, 13.98%.

5‑(4‑Chloro‑3‑nitrobenzylideneamino)‑*N***‑(4‑chlorophenyl)‑3‑(4‑methoxyphenylamino)‑1***H***‑pyrazole‑4‑carboxamide (16i)**

Yellow crystals; M.p.: 280–282 °C; yield (71%); IR (KBr) $v_{\text{max}}/\text{cm}^{-1}$ 3440, 3280, 3230 (3NH), 1653 (C=O), 1615 $(C=N)$, 1561 $(C=C)$; ¹H NMR (500 MHz, DMSO- d_6 , δ / ppm): 3.72 (s, 3H, OCH3), 6.89 (d, 2H, *J*=8.7 Hz, *p*-methoxyphenyl ring), 7.35 (d, 4H, *J*=8.8 Hz, *p*-chlorophenyl ring), 7.70 (d, 2H, *J*=8.8 Hz, *p*-methoxyphenyl ring), 7.96 (d, 1H, *J*=8.4 Hz, 4-chloro-3-nitrophenyl ring), 8.27 (d, 1H, *J*=8.3 Hz, 4-chloro-3-nitrophenyl ring), 8.26 (s, 1H, 4-chloro-3-nitrophenyl ring), 8.72 (s*,* 1H, –CH=N–), 9.14, 9.87, 12.94 (3s, 3H, 3NH exchangeable by D_2O); Anal. Calcd. (%) for $C_{24}H_{18}Cl_2N_6O_4$ (525.34): C, 54.87; H, 3.45; N, 16.00; Found: C, 55.05; H, 3.28; N, 16.19%.

*N***‑(4‑Chlorophenyl)‑3‑(4‑methoxyphenylamino)‑5‑(thiophe n‑2‑ylmethyleneamino)‑1***H***‑pyrazole‑4‑carboxamide (16j)**

Yellow crystals; M.p.: 221–223 °C; yield (68%); IR (KBr) $v_{\text{max}}/\text{cm}^{-1}$ 3432, 3278 (3NH), 1653 (C=O), 1598 (C=N), 1562 (C=C); ¹H NMR (500 MHz, DMSO- d_6 , *δ*/ppm): 3.73 (s, 3H, OCH3), 6.91 (d, 2H, *J*=8.1 Hz, *p*-methoxyphenyl ring), 7.39–7.45 (m, 5H, *p*-chlorophenyl ring+thiophene ring), 7.74 (d, 2H, *J*=8.0 Hz, *p*-methoxyphenyl ring), 7.91 (d, 1H, *J*=2.3 Hz, thiophene ring), 8.05 (d, 1H, *J*=4.8 Hz, thiophene ring), 8.64 (s*,* 1H, –CH=N–), 9.19, 10.04, 12.97 (3s, 3H, 3NH exchangeable by D₂O); Anal. Calcd. (%) for $C_{22}H_{18}CIN_5O_2S$ (451.93): C, 58.47; H, 4.01; N, 15.50; Found: C, 58.58; H, 4.19; N, 15.21%.

In vitro antimicrobial evaluation

Microorganisms

The microbes used in this study were Gram-positive bacterial strain (*Bacillus subtilis* ATCC-6633), Gram-negative bacterial strain (*Escherichia coli* ATCC-25922), yeast (*Candida albicans* ATCC-10231), and *Aspergillus niger* NRRL-3 as fungi. These microorganisms were obtained from the American Type Culture Collection (ATCC, Rockville, MD, USA) and Northern Utilization Research and Development Division, United State Department of Agriculture, Peoria, Illinois, USA (NRRL). The bacterial strains were revived for bonsai by sub-culturing fresh nutrient agar (NA) medium (Merck, Darmstadt, Germany) for 24 h before the test, while, fungi were cultured on potato dextrose agar (PDA) (Lab M., Bury, Lancashire, UK) for 7 days at 28 °C before the experiment was carried out.

Inoculum preparation

Stock cultures were maintained at 4 °C on slopes of nutrient agar and potato dextrose agar. Active cultures for experiments were prepared by transferring a lapful of cells from the stock cultures to test tubes of Mueller–Hinton broth (MHB) (Lab M Limited, Bury, Lancashire, UK) for bacteria and Sabouraud dextrose broth (SDB) (Lab M., Bury, Lancashire, UK) for fungi that were incubated without agitation for 24 h at 37 °C and 25 °C, respectively. To 5 mL of MHB and SDB, 0.2 mL of culture was inoculated and incubated (or diluted) till it reached the turbidity equal to that of the standard 0.5 McFarland solution at 625 nm $(A = 0.08 \text{ to } 0.1)$ which is equivalent to 1.5×10^8 cfu.mL⁻¹.

Bioassay method

Agar difusion technique as described by Perez et al. [[45\]](#page-23-13) was used to determine the in vitro antibacterial activity of the powder samples. A 0.1 mL aliquot of 18 h broth culture of the above-mentioned bacteria that had been adjusted to the turbidity equivalent of 0.5 McFarland standards [[63](#page-24-6)] was dispensed into sterile Petri dishes previously labeled with the test bacteria. Molten sterile Muller-Hinton was aseptically poured into the plates and gently rotated for the bacteria to be homogeneously distributed in the medium. The agar plates were allowed to solidify. The antibacterial screening bioassay was made by the agar well difusion method described by Jorgensen and Turnidge [[64\]](#page-24-7) using Mueller–Hinton agar (Lab M Limited, Bury, Lancashire, UK), then the plates allowed to difuse for two hours at 4 °C. The experiment was conducted in triplicates. All plates were incubated at 37 °C for 24 h for bacterial strains and 28–30 °C for 48 h for fungal strain. Clearance zones around the wells were noted and measured in millimeters [[65\]](#page-24-8). Standard bacterial antibiotics as Tetracycline (30 µg) and Novobiocine (30 μ g) and fungal antibiotic as Clotrimazole (50 μ g), and Cyclohexamide (50 µg) were used as a positive control for bacteria and fungi, respectively.

Molecular docking

All the molecular docking studies were performed using Molecular Operating Environmental (MOE) software 2008.10. The most active derivatives against the tested bacterial strains (**14b**, **14e**, **14j**, and **16i**) and fungal pathogens (**16c**, **16d**, **16h**, and **16i**) were builders using Chem-Draw 2014. The structure of the most active derivatives was protonated and minimized energy using MMFF94X force feld with a root mean standard deviation (RMSD) gradient of 0.05 kcal mol⁻¹ Å as well as the partial charged calculated. For promising antibacterial derivatives *S. aureus* Gyrase complex with Ciprofoxacin (PDB:2XCT), and for promising fungal derivatives secreted aspartic protease from *C. albicans* (PDB: 1ZAP) were selected for this study [[57,](#page-24-0) [58](#page-24-1)]. The enzyme with its co-crystallized ligand was retrieved from the protein data bank. For DNA gyrase (PDB:2XCT), the docking process performed using only one chain G chain only according to the reported method. For fungal, the protein structure (PDB: 1ZAP) contains only one chain (A). The enzyme was prepared according to default protocol by removing the water molecules, hydrogen atom added. The alpha site fnder was used to generate the active site. For docking studies, the trigonal matcher placement and the London dG scoring function were selected after redocking the co-crystalized ligand in both enzymes.

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Declarations

Conflict of interest The authors declare that they have no confict of interest.

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