

CrossMark

INTERSTITIAL LUNG DISEASE (E ALHAMAD, SECTION EDITOR)

The role of serological testing in idiopathic interstitial pneumonia: a rheumatologist perspective

Mohammed A. Omair¹

Published online: 27 June 2015 © Springer Science+Business Media New York 2015

Abstract The diagnosis of idiopathic interstitial pneumonias (IIPs) is complex and requires multidisciplinary approach. Identifying an underlying connective tissue disease (CTD) is associated with a better prognosis and should be routinely evaluated. Serological testing is an important tool in detecting patients with CTDs and differentiating them from patients with IIP. This article will highlight the importance of serological testing in patients with IIPs and the role of the rheumatologist in evaluating these patients.

Keywords Idiopathic interstitial pneumonias · Connective tissue diseases · Autoantibodies · Lung dominant connective tissue disease

Introduction

Idiopathic interstitial pneumonias (IIPs) are a group of interstitial lung diseases with no known etiology. There are several entities with varying patterns of inflammation and fibrosis. In 2002, the American Thoracic Society (ATS) and the European Respiratory Society (ERS) have developed an international multidisciplinary classification of the IIPs [1]. This statement was recently updated in 2013 [2]. One important addition in the classification of IIPs is the acceptance of nonspecific interstitial pneumonia (NSIP) as a clinic-pathological entity [3],

This article is part of the Topical Collection on Interstitial Lung Disease

which is more commonly associated with underlying connective tissue disease (CTD) [4]. It has been observed that CTDrelated interstitial lung disease (CTD-ILD) carries a better prognosis than the IIPs [5, 6]. Autoantibody testing is recommended to evaluate patients with symptoms suggestive of CTD such as joint pain, oral ulcers, skin rash, and many others [7]. Pulmonary involvement is variable in CTDs [8..]. Currently; there are no strong data to confidently recommend serological testing in patients with IIPs. Despite the lack of evidence, the ATS, ERS, the Japanese Respiratory Society (JRS), and the Latin American Thoracic Association (ALAT) have conjointly recommended that serologic testing for CTDs should be performed in the evaluation of the majority of patients with idiopathic pulmonary fibrosis [9]. The antinuclear antibody (ANA) and rheumatoid factor (RF) are the most commonly used autoantibodies for screening of patients with suspected CTDs. They are found in the normal population [10, 11] as well as in a variety of non-rheumatic causes [12] including idiopathic pulmonary fibrosis [13•].

Routine evaluation might sometime not be adequate to identify the underlying CTD which would wrongly lead to labeling a patient as having IIP. Solutions to this problem would include the use of nailfold capillaroscopy [14•], extensive serological testing [15], and minor salivary gland biopsy [16]. In this report, we will discuss why would an extensive serological testing might lead to identify more patients with underlying CTDs and the role of the rheumatologist in identifying it.

The advances in autoantibody detection in patients with CTDs

Antibodies to specific antigens have been discovered in the last three decades that proved to be important in the diagnosis and management of CTDs. They are useful in screening and

Mohammed A. Omair momair@ksu.edu.sa

¹ Division of Rheumatology, Department of Medicine (38), King Khalid University Hospital, College of Medicine, King Saud University, PO Box 2925, Riyadh 11461, Saudi Arabia

predicting disease subset, complications, and even survival. Many of these autoantibodies were detected in the sera of patients before the onset of symptoms [17–20]. The idiopathic inflammatory myopathies (IIMs) include dermatomyositis (DM) and polymyositis (PM). Autoantibodies against the cytoplasmic aminoacyl-tRNA synthetase (ARS) enzymes are the most frequently detected in patients with IIMs [21]. The classical and most widely tested is the Jo-1 (histidyl tRNAsynthetase) [22] which is only found in 20 % of patients [23, 24]. To date, seven other anti-ARS molecules have been identified and are collectively found in 20 % of IIM patients which includes PL-7 (threonyl) [25], PL-12 (alanyl) [26], EJ (glycyl) [27], OJ (isoleucyl) [28], KS (asparaginyl) [29], Ha (tyrosyl) [30], and most recently Zo (phenylalanyl) [31]. Most of these new antibodies were detected in the anti-synthetase syndrome which commonly includes interstitial pneumonia and might be the predominant feature [32-34]. In a series of 34 patients with clinically proven anti-synthetase syndrome with lung involvement, 9 (24 %) of them were ANA and Jo-1 negative but positive for PL-7 and PL-12 [35]. The anti-CADM-140 is a non-ARS antibody which is usually associated with cutaneous manifestations and minimal muscle involvement that can be complicated by a rapidly progressive interstitial pneumonia especially in the Asian ethnicity [36, 37]. Ye et al. reported the 6 months survival of patients with predominant ILD with a positive anti-CADM-140 antibody to be 40.8 % [38]. Other rare autoantibodies include the anti-SRP, anti-Mi-2, and antip155/140, all have been also demonstrated to be present in IIMs but do not have a clear association with lung disease [39].

Similarly, in the field of systemic sclerosis (SSc), the two classical autoantibodies frequently used are topoisomerase I antibody which has been linked to the diffuse subset and lung involvement [40-42] and anti-centromere [43] which is usually found in the limited subset and predicts pulmonary hypertension. The estimated frequencies at its best of topoisomerase I and anticentromere in the SSc population are 15-20 % and 20-30 %, respectively [44]. While other autoantibodies such as RNA polymerase III [45, 46], Th/To [47], and PM-Scl [48] which can be detected in patients tested negative for the former two antibodies can be of significant importance and predict progression and lung involvement [49, 50] are not commonly used in daily clinical practice. In one study of patients with a clinical diagnosis of IPF and surgical lung biopsy-proven usual interstitial pneumonia (UIP), the presence of a nucleolar ANA pattern and anti-Th/To was associated with the presence of many symptoms suggestive of SSc sine scleroderma (ssSSc) and limited SSc that most did not meet the classification criteria [51]. Interestingly, the prognosis in Th/To positive and Th/To negative patients was similar. This finding was similarly demonstrated from our center where the presence of autoantibodies in patients with IPF was not associated with an improved survival [hazard ratio (HR) 0.68, 95 % CI 0.202–2.347; P=0.550], rather the presence of autoimmune symptoms conveyed a statistically significant improved survival (HR 0.27; 95 % CI 0.09–0.82; P=0.020) [13•]. This finding could make us believe that the presence of autoantibodies by themselves does not improve survival, but their role is by improving the detection of underlying CTD.

Limitations of current classification criteria

Initial classification criteria published for CTDs have been designed from the 1980s forward and primarily used to include patients into clinical trials, cohorts, and registries. Many of these criteria have undergone extensive revisions because of the following limitations. First, they were developed from patients with established diseases leading to inclusion of late specific components with a lower sensitivity. This issue has been solved by extraction of new criteria from a mixed population of early and late. Additionally, different methodologies have been innovated to develop more robust and clinically meaningful criteria. As an example, the 2010 American College of Rheumatology (ACR)-European League Against Rheumatism (EULAR) for RA [52] were developed by different methodologies compared to the 1987 ACR criteria [53]. Second, new investigational tools have been recognized as useful and feasible to be used for the diagnosis of CTDs. A clear example is the addition of nailfold capillaroscopy to the 2013 ACR-EULAR classification criteria for systemic sclerosis [54] compared to the preliminary 1988 criteria [55]. Third (as discussed above), the discovery and addition of new autoantibodies have helped to stratify more patients. Two important examples are RNA polymerase III to the 2013 ACR-EULAR classification criteria for SSc, and the addition of cyclic citrullinated peptide (CCP) antibodies to the 2010 ACR-EULAR classification criteria for RA. Finally, weight has been given to the autoantibody titer such as the 2010 ACR-EULAR classification criteria for RA (2 points for low positive RF or anti-CCP and 3 points for high positive RF or anti-CCP). As a result, the new criteria classified more patients to have the disease and deemed more sensitive [56, 57]. Despite of all of these measures, classification criteria are still not perfect and their application on patients presenting with predominant respiratory symptoms or findings suggestive of IIPs is still unknown.

Rare syndromes in CTDs with lack of predominant features

Some patients with CTDs lack the predominant feature of their primary disease which deviates clinicians from suspecting these conditions which might lead to a significant delay in diagnosis and subsequent organ failure [58, 59]. An important aspect in diagnosing these patients is the identification of the subtle clinical findings and the detection of specific autoantibodies. Two important syndromes are ssSSc and amyopathic DM (ADM). In 1986, Giordano et al. proposed the term ssSSc as a distinct subtype of SSc with visceral involvement and no skin manifestations. It has been shown that these patients have similar visceral involvement to the limited subtype but has more respiratory symptoms [60]. ADM was associated with a poorer ILD survival in comparison with classic DM and PM [36] with a mortality reaching 67 % when complicated by acute/subacute ILD [37].

Lung dominant CTDs

The lung is a major target for CTDs. In the presence of an established CTD and lung manifestations, the causal relation is clear assuming that infectious causes and drug adverse events have been properly excluded. In some conditions, lung involvement precedes other symptoms. Homma et al. prospectively followed 68 patients with a diagnosis of IIPs at inclusion for a duration of 1-11 years. Thirteen (19 %) of them developed systemic manifestations of CTD adequate to label them as having CTD-related IIP [61]. In 2007, Kinder et al. have discussed the matter that idiopathic NSIP might be the lung manifestation of undifferentiated CTD (UCTD) and suggested that the occurrence of one prespecified symptom or signs of CTDs plus the presence of systemic inflammation (either autoantibody or acute phase reactant) would be highly suggestive of UCTD [62]. Later on, Fischer et al. have discussed the challenges and limitations of evaluating patients with IPs (not confined to one histological subtype) and findings suggestive of an underlying CTD that do not meet classification criteria of CTD. They have proposed the concept of lung-dominant CTDs (LD-CTD) and suggested provisional criteria [15]. The serological testing suggested in these criteria is much more extensive than that proposed by Kinder et al. [62].

Pereira et al. reported the prevalence and characteristics of patients fulfilling the LD-CTD criteria (proposed by Fischer et al.) in 1998 ILD patients. Fifty-two (2.6 %) fulfilled the criteria. After a median follow-up of 61 months, 8 (15.3 %) fulfilled the classification criteria of a specific CTD [63••]. Omote et al. reported the most common histological pattern of LD-CTD is the usual interstitial pneumonia (UIP) followed by NSIP. Patients with NSIP had a significant improvement in the annual change in percent predicted forced vital capacity and better survival compared to patients with UIP [64••].

The rheumatologist as part of the IIP multidisciplinary team

The multidisciplinary team evaluating patients with suspected IIPs consists of pulmonologists, radiologists, and pathologists [2]. The role of rheumatologists is still not fully explored. Many rheumatologists are not familiar with the concept of LD-CTD and might hesitate to accept it as a form of UCTD or as a specific disease subtype because of the exclusive presentation of lung pathology with the complete absence of extrathoracic CTD manifestations. As an example, if a 40-year-old lady is evaluated for respiratory symptoms, CT scan findings of NSIP and histological lung features include lymphoid aggregates with germinal centers, extensive pleuritis, prominent plasmacytic infiltration and dense perivascular collagen with the presence of a positive ANA and topoisomerase I antibody without any other clinical symptoms. The pulmonologist would be reluctant to label her as suffering from idiopathic NSIP; on the other hand, the rheumatologist has no adequate evidence to diagnose this patient as having SSc. This lack of consensus would lead to confusion and undesirable consequences affecting patient management and prognosis. Solutions to this problem are increasing the awareness of LD-CTD concept in the rheumatology society and incorporating the rheumatologist into the multidisciplinary team evaluating patients with suspected LD-CTD.

Conclusion

Routine evaluation of patients with IPs is not adequate and can lead to misclassification and mismanagement. The presence of autoantibodies in patients with IIPs would not improve prognosis in all subtypes, but a more extensive serological testing can lead to a better detection of underlying CTDs which might eventually offer better therapeutic option and have a more favorable prognosis. Increasing awareness of rheumatologists about the concept of LD-CTD and involving them in evaluation of these patients might lead to a better informed decision and patient care.

Compliance with Ethics Guidelines

Conflict of Interest The authors declare that they have no competing interests.

Human and Animal Rights and Informed Consent This article contains no studies with human or animal subjects performed by the author.

References

Papers of particular interest, published recently, have been highlighted as:

- Of importance
- •• Of major importance
- American Thoracic Society/European Respiratory Society International Multidisciplinary Consensus Classification of the Idiopathic Interstitial Pneumonias. This joint statement of the American Thoracic Society (ATS), and the European Respiratory Society (ERS) was adopted by the ATS board of directors, June 2001 and by the ERS executive committee, June 2001. Am J Respir Crit Care Med. 2002;165(2):277–304.

- Travis WD, Costabel U, Hansell DM, King Jr TE, Lynch DA, Nicholson AG, et al. An official American Thoracic Society/ European Respiratory Society statement: update of the international multidisciplinary classification of the idiopathic interstitial pneumonias. Am J Respir Crit Care Med. 2013;188(6):733–48.
- Travis WD, Hunninghake G, King Jr TE, Lynch DA, Colby TV, Galvin JR, et al. Idiopathic nonspecific interstitial pneumonia: report of an American Thoracic Society project. Am J Respir Crit Care Med. 2008;177(12):1338–47.
- Romagnoli M, Nannini C, Piciucchi S, Girelli F, Gurioli C, Casoni G, et al. Idiopathic nonspecific interstitial pneumonia: an interstitial lung disease associated with autoimmune disorders? Eur Respir J. 2011;38(2):384–91.
- Flaherty KR, Colby TV, Travis WD, Toews GB, Mumford J, Murray S, et al. Fibroblastic foci in usual interstitial pneumonia: idiopathic versus collagen vascular disease. Am J Respir Crit Care Med. 2003;167(10):1410–5.
- Park JH, Kim DS, Park IN, Jang SJ, Kitaichi M, Nicholson AG, et al. Prognosis of fibrotic interstitial pneumonia: idiopathic versus collagen vascular disease-related subtypes. Am J Respir Crit Care Med. 2007;175(7):705–11.
- Habash-Bseiso DE, Yale SH, Glurich I, Goldberg JW. Serologic testing in connective tissue diseases. Clin Med Res. 2005;3(3): 190–3.
- Vij R, Strek ME. Diagnosis and treatment of connective tissue disease-associated interstitial lung disease. Chest. 2013;143(3): 814–24. A review article covering aspects of connective tissue disease-associated interstitial lung disease.
- Raghu G, Collard HR, Egan JJ, Martinez FJ, Behr J, Brown KK, et al. An official ATS/ERS/JRS/ALAT statement: idiopathic pulmonary fibrosis: evidence-based guidelines for diagnosis and management. Am J Respir Crit Care Med. 2011;183(6):788–824.
- Dequeker J, Van Noyen R, Vandepitte J. Age-related rheumatoid factors. Incidence and characteristics. Ann Rheum Dis. 1969;28(4): 431–6.
- Guo YP, Wang CG, Liu X, Huang YQ, Guo DL, Jing XZ, et al. The prevalence of antinuclear antibodies in the general population of China: a cross-sectional study. Curr Ther Res Clin Exp. 2014;76: 116–9.
- Lane SK, Gravel Jr JW. Clinical utility of common serum rheumatologic tests. Am Fam Physician. 2002;65(6):1073–80.
- 13.• Alhamad EH, Cal JG, AlBoukai AA, Shaik SA, Omair MA. Autoimmune symptoms in idiopathic pulmonary fibrosis: clinical significance. Clin Respir J. 2014;13. A study demonstrating an improved survival in IPF patients with the presence of symptoms of connective tissue diseases rather than autoantibodies.
- 14.• Cakmakci Karadogan D, Balkarli A, Onal O, Altinisik G, Cobankara V. The role of nailfold capillaroscopy in interstitial lung diseases—can it differentiate idiopathic cases from collagen tissue disease associated interstitial lung diseases? Tuberk Toraks. 2015;63(1):22–30. A study demonstrating the role of nailfold capillaroscopy in detecting occult connective tissue diseases.
- Fischer A, West SG, Swigris JJ, Brown KK, du Bois RM. Connective tissue disease-associated interstitial lung disease: a call for clarification. Chest. 2010;138(2):251–6.
- Fischer A, Swigris JJ, du Bois RM, Groshong SD, Cool CD, Sahin H, et al. Minor salivary gland biopsy to detect primary Sjogren syndrome in patients with interstitial lung disease. Chest. 2009;136(4):1072–8.
- Aho K, Heliovaara M, Maatela J, Tuomi T, Palosuo T. Rheumatoid factors antedating clinical rheumatoid arthritis. J Rheumatol. 1991;18(9):1282–4.
- Aho K, von Essen R, Kurki P, Palosuo T, Heliovaara M. Antikeratin antibody and antiperinuclear factor as markers for subclinical rheumatoid disease process. J Rheumatol. 1993;20(8):1278–81.

- Nielen MM, van Schaardenburg D, Reesink HW, van de Stadt RJ, van der Horst-Bruinsma IE, de Koning MH, et al. Specific autoantibodies precede the symptoms of rheumatoid arthritis: a study of serial measurements in blood donors. Arthritis Rheum. 2004;50(2): 380–6.
- Rantapaa-Dahlqvist S, de Jong BA, Berglin E, Hallmans G, Wadell G, Stenlund H, et al. Antibodies against cyclic citrullinated peptide and IgA rheumatoid factor predict the development of rheumatoid arthritis. Arthritis Rheum. 2003;48(10):2741–9.
- Gunawardena H, Betteridge ZE, McHugh NJ. Myositis-specific autoantibodies: their clinical and pathogenic significance in disease expression. Rheumatology (Oxford). 2009;48(6):607–12.
- Nishikai M, Reichlin M. Heterogeneity of precipitating antibodies in polymyositis and dermatomyositis. Characterization of the Jo-1 antibody system. Arthritis Rheum. 1980;23(8):881–8.
- Mimori T, Imura Y, Nakashima R, Yoshifuji H. Autoantibodies in idiopathic inflammatory myopathy: an update on clinical and pathophysiological significance. Curr Opin Rheumatol. 2007;19(6): 523–9.
- 24. Brouwer R, Hengstman GJ, Vree Egberts W, Ehrfeld H, Bozic B, Ghirardello A, et al. Autoantibody profiles in the sera of European patients with myositis. Ann Rheum Dis. 2001;60(2):116–23.
- Mathews MB, Reichlin M, Hughes GR, Bernstein RM. Antithreonyl-tRNA synthetase, a second myositis-related autoantibody. J Exp Med. 1984;160(2):420–34.
- Bunn CC, Bernstein RM, Mathews MB. Autoantibodies against alanyl-tRNA synthetase and tRNAAla coexist and are associated with myositis. J Exp Med. 1986;163(5):1281–91.
- Targoff IN, Trieu EP, Plotz PH, Miller FW. Antibodies to glycyltransfer RNA synthetase in patients with myositis and interstitial lung disease. Arthritis Rheum. 1992;35(7):821–30.
- Targoff IN, Trieu EP, Miller FW. Reaction of anti-OJ autoantibodies with components of the multi-enzyme complex of aminoacyl-tRNA synthetases in addition to isoleucyl-tRNA synthetase. J Clin Invest. 1993;91(6):2556–64.
- Hirakata M, Suwa A, Nagai S, Kron MA, Trieu EP, Mimori T, et al. Anti-KS: identification of autoantibodies to asparaginyl-transfer RNA synthetase associated with interstitial lung disease. J Immunol. 1999;162(4):2315–20.
- Hashish L TE, Sadanandan P, Targoff, IN. Identification of autoantibodies to tyrosyl-tRNA synthetase in dermatomyositis with features consistent with antisynthetase syndrome. Arthritis Rheum. [Abstract]. 2005;52(S312).
- Betteridge Z, Gunawardena H, North J, Slinn J, McHugh N. Antisynthetase syndrome: a new autoantibody to phenylalanyl transfer RNA synthetase (anti-Zo) associated with polymyositis and interstitial pneumonia. Rheumatology (Oxford). 2007;46(6):1005–8.
- Targoff IN, Arnett FC. Clinical manifestations in patients with antibody to PL-12 antigen (alanyl-tRNA synthetase). Am J Med. 1990;88(3):241–51.
- Hirakata M, Suwa A, Takada T, Sato S, Nagai S, Genth E, et al. Clinical and immunogenetic features of patients with autoantibodies to asparaginyl-transfer RNA synthetase. Arthritis Rheum. 2007;56(4):1295–303.
- Sato S, Kuwana M, Hirakata M. Clinical characteristics of Japanese patients with anti-OJ (anti-isoleucyl-tRNA synthetase) autoantibodies. Rheumatology (Oxford). 2007;46(5):842–5.
- 35. Fischer A, Swigris JJ, du Bois RM, Lynch DA, Downey GP, Cosgrove GP, et al. Anti-synthetase syndrome in ANA and anti-Jo-1 negative patients presenting with idiopathic interstitial pneumonia. Respir Med. 2009;103(11):1719–24.
- Kang EH, Lee EB, Shin KC, Im CH, Chung DH, Han SK, et al. Interstitial lung disease in patients with polymyositis, dermatomyositis and amyopathic dermatomyositis. Rheumatology (Oxford). 2005;44(10):1282–6.

- Suda T, Fujisawa T, Enomoto N, Nakamura Y, Inui N, Naito T, et al. Interstitial lung diseases associated with amyopathic dermatomyositis. Eur Respir J. 2006;28(5):1005–12.
- Ye S, Chen XX, Lu XY, Wu MF, Deng Y, Huang WQ, et al. Adult clinically amyopathic dermatomyositis with rapid progressive interstitial lung disease: a retrospective cohort study. Clin Rheumatol. 2007;26(10):1647–54.
- Casciola-Rosen L, Mammen AL. Myositis autoantibodies. Curr Opin Rheumatol. 2012;24(6):602–8.
- Catoggio LJ, Bernstein RM, Black CM, Hughes GR, Maddison PJ. Serological markers in progressive systemic sclerosis: clinical correlations. Ann Rheum Dis. 1983;42(1):23–7.
- Weiner ES, Earnshaw WC, Senecal JL, Bordwell B, Johnson P, Rothfield NF. Clinical associations of anticentromere antibodies and antibodies to topoisomerase I. A study of 355 patients. Arthritis Rheum. 1988;31(3):378–85.
- Douvas AS, Achten M, Tan EM. Identification of a nuclear protein (Scl-70) as a unique target of human antinuclear antibodies in scleroderma. J Biol Chem. 1979;254(20):10514–22.
- 43. Tan EM, Rodnan GP, Garcia I, Moroi Y, Fritzler MJ, Peebles C. Diversity of antinuclear antibodies in progressive systemic sclerosis anti-centromere antibody and its relationship to CREST syndrome. Arthritis Rheum. 1980;23(6):617–25.
- 44. Ho KT, Reveille JD. The clinical relevance of autoantibodies in scleroderma. Arthritis Res Ther. 2003;5(2):80–93.
- Kuwana M, Kaburaki J, Mimori T, Tojo T, Homma M. Autoantibody reactive with three classes of RNA polymerases in sera from patients with systemic sclerosis. J Clin Invest. 1993;91(4):1399–404.
- 46. Hirakata M, Okano Y, Pati U, Suwa A, Medsger Jr TA, Hardin JA, et al. Identification of autoantibodies to RNA polymerase II occurrence in systemic sclerosis and association with autoantibodies to RNA polymerases I and III. J Clin Invest. 1993;91(6):2665–72.
- Hardin JA, Rahn DR, Shen C, Lerner MR, Wolin SL, Rosa MD, et al. Antibodies from patients with connective tissue diseases bind specific subsets of cellular RNA-protein particles. J Clin Invest. 1982;70(1):141–7.
- Reichlin M, Maddison PJ, Targoff I, Bunch T, Arnett F, Sharp G, et al. Antibodies to a nuclear/nucleolar antigen in patients with polymyositis overlap syndromes. J Clin Immunol. 1984;4(1):40–4.
- Hanke K, Bruckner CS, Dahnrich C, Huscher D, Komorowski L, Meyer W, et al. Antibodies against PM/Scl-75 and PM/Scl-100 are independent markers for different subsets of systemic sclerosis patients. Arthritis Res Ther. 2009;11(1):R22.
- Mitri GM, Lucas M, Fertig N, Steen VD, Medsger Jr TA. A comparison between anti-Th/To- and anticentromere antibody-positive systemic sclerosis patients with limited cutaneous involvement. Arthritis Rheum. 2003;48(1):203–9.
- Fischer A, Meehan RT, Feghali-Bostwick CA, West SG, Brown KK. Unique characteristics of systemic sclerosis sine scleroderma-associated interstitial lung disease. Chest. 2006;130(4):976-81.
- 52. Aletaha D, Neogi T, Silman AJ, Funovits J, Felson DT, Bingham 3rd CO, et al. 2010 rheumatoid arthritis classification criteria: an American College of Rheumatology/European League Against

Rheumatism collaborative initiative. Arthritis Rheum. 2010;62(9): 2569–81.

- 53. Arnett FC, Edworthy SM, Bloch DA, McShane DJ, Fries JF, Cooper NS, et al. The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. Arthritis Rheum. 1988;31(3):315–24.
- 54. van den Hoogen F, Khanna D, Fransen J, Johnson SR, Baron M, Tyndall A, et al. 2013 classification criteria for systemic sclerosis: an American College of Rheumatology/European League against rheumatism collaborative initiative. Arthritis Rheum. 2013;65(11): 2737–47.
- 55. Subcommittee for scleroderma criteria of the American Rheumatism Association Diagnostic and Therapeutic Criteria Committee. Preliminary criteria for the classification of systemic sclerosis (scleroderma). Arthritis Rheum. 1980;23(5):581–90.
- Hoffmann-Vold AM, Gunnarsson R, Garen T, Midtvedt O, Molberg O. Performance of the 2013 American College of Rheumatology/European League Against Rheumatism classification Criteria for Systemic Sclerosis (SSc) in large, well-defined cohorts of SSc and mixed connective tissue disease. J Rheumatol. 2015;42(1):60–3.
- Fautrel B, Combe B, Rincheval N, Dougados M. Level of agreement of the 1987 ACR and 2010 ACR/EULAR rheumatoid arthritis classification criteria: an analysis based on ESPOIR cohort data. Ann Rheum Dis. 2012;71(3):386–9.
- Rodnan GP, Fennell Jr RH. Progressive systemic sclerosis sine scleroderma. JAMA. 1962;180:665–70.
- Horswell RR, Hargrove Jr MD, Peete WP, Ruffin JM. Scleroderma presenting as the malabsorption syndrome. A case report. Gastroenterology. 1961;40:580–2.
- Poormoghim H, Lucas M, Fertig N, Medsger Jr TA. Systemic sclerosis sine scleroderma: demographic, clinical, and serologic features and survival in forty-eight patients. Arthritis Rheum. 2000;43(2):444–51.
- Homma Y, Ohtsuka Y, Tanimura K, Kusaka H, Munakata M, Kawakami Y, et al. Can interstitial pneumonia as the sole presentation of collagen vascular diseases be differentiated from idiopathic interstitial pneumonia? Respiration. 1995;62(5):248–51.
- 62. Kinder BW, Collard HR, Koth L, Daikh DI, Wolters PJ, Elicker B, et al. Idiopathic nonspecific interstitial pneumonia: lung manifestation of undifferentiated connective tissue disease? Am J Respir Crit Care Med. 2007;176(7):691–7.
- 63.•• Pereira DA, Dias OM, de Almeida GE, Araujo MS, Kawano-Dourado LB, Baldi BG, et al. Lung-dominant connective tissue disease among patients with interstitial lung disease: prevalence, functional stability, and common extrathoracic features. J Bras Pneumol. 2015;41(2):151–60. A study describing a cohort of patients with lung dominant connective tissue disease and their outcome.
- 64.•• Omote N, Taniguchi H, Kondoh Y, Watanabe N, Sakamoto K, Kimura T, et al. Lung-dominant connective tissue disease: clinical, radiologic and histologic features. Chest. 2015;7. A study describing a cohort of patients with lung dominant connective tissue disease.