REVIEW ARTICLE

Options to reduce ranges in critical soil nutrient levels used in fertilizer recommendations by accounting for site conditions and methodology: A review

Siatwiinda M. Siatwiinda[1](http://orcid.org/0000-0003-0517-9604) · Gerard H. Ros1 [·](http://orcid.org/0000-0002-6062-9770) Olusegun A. Yerokun2 · Wim de Vries[1](http://orcid.org/0000-0001-9974-0612)

Accepted: 21 December 2023 / Published online: 30 January 2024 © The Author(s) 2024

Abstract

Fertilizer recommendations (FR) to improve yields and increase proftability are based on relationships between crop yields and soil nutrient levels measured via soil extraction methods. Within these FR, critical soil nutrient (CSN) levels are used to distinguish nutrient defcient from non-defcient soils. The variation in CSN levels is large, implying a risk of over- or under-fertilization. Here, we review and assess the factors infuencing the derivation of CSN levels in order to increase both their reliability and applicability within FR systems. The evaluated factors included site conditions, i.e., crop type and location as a surrogate for climate and soil properties, and methodological factors, i.e., the experimental approach (feld or pot experiments), and statistical methods and cut-of point. Results showed that the range of values used to defne the medium soil fertility classes coincided with the range of CSN levels derived from experimental data. We show that harmonizing methodological aspects can substantially reduce the uncertainty in the CSN levels $(> 50\%)$, implying a substantial enhancement of the reliability of FR systems. Inclusion of site conditions might further improve the reliability. To enable reduction in CSN levels requires well-documented feld experiments and standardization of data collection and analysis. We foresee the potential for generic FR systems that make use of reliable data, more process-based interpretation of nutrient pools and accounting for the interactions among nutrients.

Keywords Soil fertility classes · Critical nutrient levels · Extraction methods · Fertilizer recommendations

1 Introduction

Optimal fertilization in crop production has the potential to improve yields, nutrient use efficiencies, and consequently the proftability of agriculture (Schut and Giller [2020;](#page-20-0) Bado et al. [2018\)](#page-18-0). Additionally, optimal fertilization reduces the amount of non-point pollution from fertilizer and manure, attributed to injudicious nutrient management (Tandy et al. [2021\)](#page-21-0). In most countries, the optimum fertilizer dose and fertilization strategies are established by extension services, academic research, and fertilizer companies with the objective to maximize crop yield and profitability. These fertilizer recommendations (FR) are usually aided by

Lusaka, Zambia

plant (Izsaki [2009](#page-19-0); Weetman and Wells [1990;](#page-21-1) Bell [2023](#page-18-1)) or soil diagnostics (van Heerwaarden [2022\)](#page-21-2), using empirical algorithms that translate soil or plant nutrient concentrations into a desired nutrient dose in order to achieve the target crop yield. Both methods require insights into the relationship between concentrations in soils or plants and the corresponding growth and yield response curves, usually obtained in feld experiments using diferent rates of fertilizers (Bell [2023](#page-18-1)). How soil- and plant-based methods will complement each other is currently under debate; however, sensor-derived estimates (of both) will certainly contribute to more tailor-made fertilizer practices that can match crop demand with the actual soil nutrient supply or availability.

Plant diagnostics use whole shoot or plant part analysis to assess defciencies or toxicities in view of the nutrient requirements (Izsaki [2009;](#page-19-0) Weetman and Wells [1990\)](#page-21-1) and have been used in various in-season FR for both macro and micro nutrients (Olfs et al. [2005](#page-20-1)). The use of plant diagnostics is based on the idea that the plant itself is the best

 \boxtimes Siatwiinda M. Siatwiinda siatwiinda.siatwiinda@wur.nl

¹ Wageningen University and Research, PO Box 47, 6700 AA Wageningen, The Netherlands

indicator of defciencies in the nutrient supply from the soil (Lemaire et al. [2019](#page-19-1)). Although the result of a plant analysis can be used to decide about the necessity, the 'optimum' timing, and the 'optimum' fertilizer dose, its value for FR systems has been challenged because nutrient interactions and physiological growth stage infuence the critical level (Marchand et al. [2013;](#page-20-2) Martínez et al. [2002\)](#page-20-3). The actual nutrient level in the plant is the result of the interaction between nutrient supply, water availability, growth stage, and possible stressors afecting crop nutrient uptake (Cadot et al. [2018\)](#page-18-2). Hence, plant nutrient concentration alone is not an accurate proxy of nutrient requirement (Lemaire et al. [2019\)](#page-19-1).

Soil diagnostics is by far the most common method to optimize crop fertilization (van Heerwaarden [2022](#page-21-2)). This soil-based approach includes sampling and laboratory analysis of bioavailable nutrients, thereby relating the crop yield response to the nutrient dose applied while accounting for soil nutrient supply (Eckert [1987](#page-19-2); Jordan-Meille et al. [2012\)](#page-19-3). The relationships underling FR are by defnition empirical and only valid for the agroecosystem properties for which they have been derived, limiting their applicability for other regions and land uses (Lemaire et al. [2019\)](#page-19-1). Critical soil nutrient levels have been defned for each extraction method (soil test) to ensure that crop growth is not limited assuming a desired target yield level (Breitschuh et al. [2008](#page-18-3); Tunney et al. [1997\)](#page-21-3). These levels are often defned as the soil nutrient level that corresponds to a crop yield that is 90% or 95% of the maximum yield; it is used as the soil nutrient level that distinguishes defcient from non-defcient soils (Cox [1992](#page-19-4); Mortvedt [1977;](#page-20-4) Steinfurth et al. [2022](#page-21-4); Conyers et al. [2013\)](#page-18-4). Substantial variation in these critical levels has been found across regions, farming systems, and continents, being afected by soil sampling and laboratory procedures to measure the plant availability of nutrients (Mortvedt [1977;](#page-20-4) Ros et al. [2011;](#page-20-5) Jordan-Meille et al. [2012\)](#page-19-3), the methodology to derive critical nutrient levels from experimental data, crop and soil management practices (Lemaire et al. [2019](#page-19-1)), and the agroecosystem properties controlling crop growth (Bell et al. [2013a;](#page-18-5) Conyers et al. [2020](#page-19-5); Conyers et al. [2013](#page-18-4); Lemaire et al. [2019\)](#page-19-1). The latter properties include not only crop type and crop variety but also soil properties such as pH, texture, groundwater depth, and soil organic matter (Valkama et al. [2009;](#page-21-5) Schut and Giller [2020\)](#page-20-0). This has been confrmed by recent datadriven approaches showing that the crop yield response to nutrient inputs is afected by various agroecosystem properties (Ros et al. [2016](#page-20-6); Coulibali et al. [2020\)](#page-19-6).

Measured soil nutrient levels via soil testing classifes the soil fertility status as being low, medium, high, or very high, following an empirical relationship between crop yield and nutrient levels, defned here as soil test calibration (Jordan-Meille et al. [2012](#page-19-3); Mitchell and Huluka

 $2012a$, [b\)](#page-20-8). Both crop yield as well the agronomic efficiency of fertilizer nutrient inputs depend on this soil fertility status, with high crop yield responses to nutrient additions expected in soils with low nutrient availability (Voss [1998](#page-21-6)) and low responses in soils with a high fertility status (Babu et al. [2016\)](#page-18-6). As a result, soil tests enable farmers to enhance crop yield and improve the agronomic efficiency of fertilizers by optimizing nutrient additions. The determination of critical soil nutrient levels serves as a basis for the conventional agronomic build-up-and-maintenance approach (Locke and Hanson [1991;](#page-20-9) Zone et al. [2020](#page-21-7)). At low soil nutrient levels, nutrients are added in excess of crop nutrient removal at a given target yield, thus elevating the soil fertility level (build-up), while nutrient addition is equal to crop nutrient removal at an adequate soil nutrient level, thus sustaining this adequate soil fertility status over time (maintain). Apart from an appropriate assessment of the crop nutrient demand, based on a target crop yield and nutrient contents in harvested crop parts, the accuracy of FR systems thus depends on the correct derivation of these critical soil nutrient levels. However, substantial variation in those levels has been found across agroecosystems (Lemaire et al. [2019](#page-19-1)) and there is an urgent need for standardized approaches to derive critical soil nutrient levels. The current uncertainty on these critical nutrient levels is one of the reasons for inefficient use of fertilizers leading to environmental losses and associated costs (Abay et al. [2022;](#page-17-0) Conyers et al. [2013;](#page-18-4) van Heerwaarden [2022](#page-21-2); Fryer et al. [2019\)](#page-19-7). Given the fact that most fertilizer recommendations originate from feld experiments from the 60's and 70's, one might further question their robustness for current agricultural conditions (Zhang et al. [2021\)](#page-21-8). Taking into account the economic and environmental challenges for farmers to optimize crop yields, there is a clear need for improved and scientifcally sound fertilizer recommendations (Conyers et al. [2013](#page-18-4); Slaton et al. [2022](#page-20-10); Steinfurth et al. [2022](#page-21-4); Jordan-Meille et al. [2012\)](#page-19-3).

A critical review of the derivation of critical soil nutrient levels, their link to soil fertility classes and the underlying agronomic concepts has rarely been made. This review aims to unravel the impacts of site conditions and methodological factors (see Fig. [1\)](#page-2-0) on the accuracy of critical soil nutrients levels in order to evaluate the opportunities for improved soil based FR systems. We focus on phosphorus (P) , potassium (K) and zinc (Zn) and extending these insights to boron (B), iron (Fe), manganese (Mn) and copper (Cu) where possible. We excluded nitrogen (N) since the required external input of N is far more controlled by the crop N demand as compared to the aforementioned nutrients, since biological processes (mainly N mineralization) rather than chemical process determine the soil N supply, usually being a minor part of the total N taken up by crops in global agriculture.

Fig. 1 Two experimental approaches to determine the soil phosphorus critical levels. **A** A pot experiment and **B** a feld experiment with the similar experimental treatments.

The ranges derived in critical levels are compared with observed ranges in medium soil fertility status. The latter range is related to crop yields that equal approximately 80–95% of its maximum value, being linked to the cut-of point to derive critical soil nutrient levels (for details see Section [3.1](#page-6-0)).

In more detail, we compared the observed range in critical levels defning the median soil fertility class from FR systems with the observed range derived from peer reviewed pot and feld experiments. We hypothesized that the variation in critical soil nutrient values declines when one accounts for the methodological conditions under which these values have been derived. If this hypothesis is true, then there is a high potential for a build-up-and-maintenance approach using generally applicable soil tests being independent of region (climate), land use (crop type), and soil type. We also evaluated the impacts of site conditions in terms of region and crop type to see if they afect the critical levels. In this article, we frst review the methodological factors determining critical soil nutrient levels along with their impacts, advantages and disadvantages (Section [2](#page-2-1)). Next, we describe the critical soil nutrient levels for all common soil tests as being used in FR as well the experimental observations from literature and quantitatively assess the impact of site conditions and methodological aspects on these critical soil nutrient levels (Section [3\)](#page-6-1). Furthermore, we evaluate the value of soil tests to improve the efficiency of fertilization and describe the opportunities to do so as well as the main bottlenecks (Section [3\)](#page-6-1).

2 Methodological factors determining critical soil nutrient levels

Critical soil nutrient levels are by defnition associated with a soil nutrient test used (Jordan-Meille et al. [2012](#page-19-3)) and their relationship with crop yield responses to fertilization.

Important factors controlling this relationship include (i) soil sampling intensity, sampling depth, and sample preparation; (ii) soil extraction method; (iii) experimental type; and (iv) statistical approaches applied to link crop response to soil nutrient levels (Jordan-Meille et al. [2012](#page-19-3); Colomb et al. [2007](#page-18-7); Heckman et al. [2006\)](#page-19-8).

2.1 Soil sampling and sample preparation

Although soil nutrient levels usually decline with soil depth, fertilizer trials linking crop nutrient responses to soil nutrient concentrations are usually limited to the top soil. The sampling depth varies from $<$ 5 cm up to 60 cm or more, depending on nutrient, cropping system, and climate (Bell et al. [2013b](#page-18-8); Speirs et al. [2013](#page-21-9); Brennan and Bell [2013](#page-18-9); Agegnehu et al. [2015](#page-17-1); Dodd and Mallarino [2005;](#page-19-9) Holford and Doyle [1992](#page-19-10); Cox and Barnes [2002\)](#page-19-11). A soil depth of 0–30 cm is most often used, assuming that the majority of the nutrients are taken up from the top layer and ignoring the contribution of nutrient uptake from the subsoil (Conyers et al. [2020](#page-19-5)). The soil depth afects the relationship between soil nutrient levels and the crop nutrient response because soil management practices and fertilizer application strategies afect the distribution of nutrients over depth. For example, minimum tillage coupled with broadcast application might reduce the amount of available nutrients in the subsoil (Bell et al. [2013b\)](#page-18-8). Others showed that broadcasting P and K fertilizers increased soil nutrient levels over the ploughing layer whereas deep banding application of fertilizers increased P and K in the deeper soil layer only (Yuan et al. [2020](#page-21-10)). Consequently, the derived critical soil nutrient levels are valid for the analyzed soil depth only, limiting the applicability of FR to situations that have comparable nutrient distributions over depth (Cox [1992\)](#page-19-4).

Soil sample preparation and handling, including storage conditions, homogenization, drying, grinding, sieving, and moisture content (Savoy [2013](#page-20-11); Slaton et al. [2022](#page-20-10); Jordan-Meille et al. [2012\)](#page-19-3), also infuence the nutrient concentrations

thereby afecting the relationships between (critical) soil nutrient levels and crop yield (Slaton et al. [2022](#page-20-10); Barbagelata and Mallarino [2013](#page-18-10)). Consequently, aspects of soil preparation and processing, historical management with regards to tillage, and fertilizer application need to be considered when studies show variation in critical soil nutrient levels (Slaton et al. [2022](#page-20-10)).

2.2 Extraction methods

There is a large a number of soil tests available to extract plant available nutrients (Jordan-Meille et al. [2012;](#page-19-3) Mortvedt [1977;](#page-20-4) Srivastava et al. [2008;](#page-21-11) Csathó et al. [2002\)](#page-19-12). Most of the soil tests use chemical extractants to determine the amount and nutrient species that are readily available or could become available for crop uptake throughout the growing season, also referred to as bioavailable nutrients (Nafu et al. [2012\)](#page-20-12) since only a small fraction of the total nutrients in soil is being available for crop uptake (Alva [1993\)](#page-18-11). Conceptually, three interconnected soil nutrient pools can be distinguished including (i) the actual available nutrient pool in soil solution; (ii) a potentially available pool that can become available due to chemical and biological processes, often called the labile or reactive nutrient pool; and (iii) a non-available or fxed nutrient pool (Harmsen et al. [2005](#page-19-13)). The nutrients in solution are readily available for plant uptake whereas the concentration is controlled by mineralization and immobilization for nutrients that are mediated by soil biology and sorption, desorption and precipitation, and dissolution for nutrients whose concentration is dominated by soil chemical processes (Wang et al. [2004](#page-21-12); Islam et al. [2017;](#page-19-14) Bilias and Barbayiannis [2019\)](#page-18-12). Soil tests strongly vary in methodology, including factors such as ionic strength, molarity, pH, soil solution ratio, shaking time, and subsequent fltration or centrifugation. As a consequence, the observed nutrient concentration might refect the actual concentration in soil solution (as being the case for $0.01M$ CaCl₂ method (Sánchez-Alcalá et al. [2014;](#page-20-13) Houba et al. [2000\)](#page-19-15)) or exceeds the concentration in soil solution with several orders of magnitude (Neyroud and Lischer [2003](#page-20-14)). Extraction methods can be classifed as single nutrient extractions like Olsen P (Olsen [1954\)](#page-20-15) or as multi-nutrient extractions like Mehlich 3 (Mehlich [1984\)](#page-20-16) and $CaCl₂$ (Houba et al. [2000](#page-19-15)). Multi-nutrient soil extractants have proven advantageous due to practical, budgetary, and environmental reasons (Ussiri et al. [1998](#page-21-13); Bortolon and Gianello [2012,](#page-18-13) [2010](#page-18-14)). More recently alternatives such as soil sensors using spectroscopy have emerged to potentially replace these soil extractant (Mohamed et al. [2018\)](#page-20-17).

Regarding the major plant nutrients, most soil tests developed were focused on P and to a lesser extent for the cations such as K, Ca, and Mg and micronutrients such as Zn, Cu, Mn, Mo, and Fe (Table [1\)](#page-4-0). The soil tests for P include extractions with $0.01M$ CaCl₂ (Houba et al. [2000\)](#page-19-15), ammonium lactate (Egnér et al. [1960\)](#page-19-16), sodium bicarbonate (Olsen [1954](#page-20-15)), calcium acetate lactate (Schüller [1969\)](#page-20-18), Mehlich 1, 2, 3 (Mehlich [1953;](#page-20-19) Mehlich [1978](#page-20-20); Mehlich [1984](#page-20-16)), and Bray 1 and 2 (Bray and Kurtz [1945](#page-18-15)), with the lowest P concentrations found in the $0.01M$ CaCl₂ method and the highest P concentrations found in the Bray 2 method (Wuenscher et al. [2015\)](#page-21-14). For the cations K, Ca, and Mg, common extractants include ammonium acetate (Doll and Lucas [1973;](#page-19-17) Simard [1993\)](#page-20-21) and multi-extractants such as Colwell (Colwell [1963;](#page-18-16) Colwell and Esdaile [1968](#page-18-17)) and Mehlich 1, 2, and 3 (Mehlich [1978](#page-20-20); Mehlich [1984\)](#page-20-16). Micronutrient concentrations have been estimated using the diethylenetriaminepentaacetic acid (DTPA) (Lindsay and Norvell [1978\)](#page-19-18), ethylenediamine tetra-acetic acid (EDTA) (Trierweiler and Lindsay [1969\)](#page-21-15), and also Mehlich 1, 2, 3 method. Methods for Boron include hot water (Berger and Truog [1939\)](#page-18-18), CaCl2 (Houba et al. [2000](#page-19-15)), DTPA (Lindsay and Norvell [1978\)](#page-19-18), and Mehlich extractions (Mehlich [1978;](#page-20-20) Mehlich [1984\)](#page-20-16). The soil test selected has strong impacts on nutrient levels as illustrated by Steinfurth et al. ([2021\)](#page-21-16).

There are no general guidelines to select the most appropriate soil test given agroecosystem properties such as land use, climate, soil texture, and soil acidity (Mashayekhi et al. [2014\)](#page-20-22). In addition, practical/logistical aspects such as ease of method and available equipment also afect the choice of the soil tests to be applied. Logistically, DTPA is preferred over Mehlich 3, HCl, and EDTA simply due to the time needed to perform the analysis (Mortvedt [1977](#page-20-4)). In the case of P extractants (Wuenscher et al. [2015](#page-21-14)), water extraction and CaCl₂ have been proposed for high-intensity farming systems given the fact that those soils are characterized by high available P levels in soil solution, whereas the P_{CACT2} concentration is often below detection limit in extensive agricultural and natural systems. In addition, the P_{OLSEN} test is due to its properties better applicable for calcareous soils whereas P_{BRAY} is more suitable for acidic soils and Mehlich 3 works across a broad range of soils (Locke and Hanson [1991](#page-20-9); Jordan-Meille et al. [2012;](#page-19-3) Neyroud and Lischer [2003](#page-20-14); Fixen et al. [1990\)](#page-19-19). More importantly, none of them has a mechanistic underpinned concept in relation to the processes controlling the actual nutrient supply or bufering, making their relationship with crop nutrient uptake and agronomic efficiency (the change in yield per kg of nutrient added) largely empirical.

To reduce the uncertainty in observed critical soil nutrient levels, it is necessary to standardize soil testing procedures across laboratories. This is partly tackled by various initiatives such as the Global Soil Partnership by the Food Agriculture Organization and Wageningen Evaluating Programmes for Analytical Laboratories (WEPAL) though the agronomic derivation of critical soil nutrient levels

² Springer INRAC

is lacking a standardized protocol. Note that the analysis protocol in the laboratory might also afect the soil nutrient level determined, as shown for the difference in P by methods using colorimetry or inductively coupled plasma analytical procedures (Heckman et al. [2006](#page-19-8)).

2.3 Experimental type

Critical soil nutrients levels have been derived from shortterm manipulation experiments (pot or feld or combined), long-term field experiments, and extensive monitoring datasets from agricultural felds (Mortvedt [1977;](#page-20-4) Ayodele and Agboola [1985](#page-18-20)). Soil amendment experiments include experiments where the soil is amended by fertilizer to attain a certain soil fertility status after which the crop response to nutrient inputs is determined. To adapt the soil fertility status, nutrients are either added (Cox [1996;](#page-19-21) Srivastava et al. [2006;](#page-21-21) Cox and Barnes [2002](#page-19-11)) or the soil nutrient levels are diluted by mixing the soil with quartz (Corrales et al. [2007](#page-19-22); Brtnicky et al. [2021](#page-18-21)). A diference in crop yield response between the manipulated and unmanipulated soil can be used to quantify the soil nutrient supply. The advantage of this controlled experimental approach is that all factors controlling crop yield remain equal except for the soil nutrient level, such as soil type, microbial community structure, temperature, and water availability. Since these experiments are usually carried out as pot experiments, their direct applicability for feld applicable fertilizer recommendations remains limited (Mortvedt [1977](#page-20-4)).

Field experiments can both be short-term (one growing season) or long-term (several years) and have the highest potential to refect the actual conditions controlling the crop response to nutrient availability. Nevertheless, results from feld experiments are still highly afected by local soil and weather conditions, complicating the upscaling to regional or county level. Similar aspects limit the applicability from long-term feld experiments, though the impact of weather might be included in the time series of observed crop yield responses to nutrient availability. In the last case, averaged and more generally applicable relationships between (criti cal) soil nutrient concentrations and crop yield responses can be established, independent of the actual weather con - ditions of the growing season (Zhang et al. [2021](#page-21-8)), in particular when experiments are done for more than 10 years (Mortvedt [1977\)](#page-20-4).

Lastly, critical soil nutrient levels can be derived from large monitoring datasets where crop yields and extractable soil nutrient contents are determined (Speirs et al. [2013](#page-21-9); Anderson et al. [2013;](#page-18-22) Bell et al. [2013b\)](#page-18-8). Their advantage is the use of real feld data from many plots and the possibility to include (or assess) the impact of site-specifc conditions such as water availability, soil management, fertilizer strategy, and crop rotation. This approach has recently gained attention by new data-driven approaches tested for precision farming purposes, but it is rarely implemented in any fertilizer recommendation system yet. As with all empirical methods, data pre-processing and analysis should follow transparent and scientifcally sound procedures to enable robust application in real-world situation thereby avoiding issues like overftting, heteroscedasticity, and multicollinearity (Conyers et al. [2020](#page-19-5)).

2.4 Statistical models

A critical soil nutrient level can be derived from any experiment where both crop yields and soil nutrient levels have been determined by ftting a mathematical model on those observations (Ayodele and Agboola [1985](#page-18-20); Hauser [1973\)](#page-19-23). The critical level is classically defned as the cutoff value above which the crop yield does not respond to added nutrient inputs, mostly set at 90–95% of the maximum yield. The crop response is commonly given as relative yield (RY) in view of the maximum yield observed (Bai et al. [2013](#page-18-23)). Liu et al. [\(2017](#page-20-24)) has defned RY as "the proportion between nutrient-limited yield and attainable yield with optimal fertilization." The common RY cut-off values range between 80 and 99% (Fageria et al. [1997](#page-19-24); Colomb et al. [2007](#page-18-7); Agboola and Ayodele [1987](#page-17-2)) and only in exceptional cases the maximum yield has been used. Lowering the cutoff value also lowers the critical soil nutrient level. The wide range in cut-off values is partly related to the crop type with 95% cut-off values mostly being used for high value crops such as vegetables and a value of 80% for low value crops, especially in places where fertilizer use is limited, but the selection remains rather subjective and sometimes even arbitrary. The fact that the majority of studies fail to report the cut-off values as well the crop yield data supports the idea that none of the soil tests has been developed for generic purposes but rather for specifc and regional applications.

Various mathematical models have been applied to relate the crop yield response to nutrient availability (Cox [1992;](#page-19-4) Hochmuth et al. [2011\)](#page-19-25) and subsequently the critical soil nutrient level needed to achieve the desired yield. These include a (i) a linear model where the yield increases continually and linearly with a change in nutrient availability, often being applied when the range in soil nutrient levels is insufficient to achieve maximum yield; (ii) quadratic model where the yield increases in response to increased soil nutrient availability until a threshold is reached, beyond which the crop response begins to decline; (iii) linear-plateau model (Waugh et al. [1973](#page-21-23)) in which the response is approximated by a linear response upon an increase in soil nutrient levels until a point (shoulder point) where the yield stabilizes (plateau); and (iv) exponential Mitscherlich model where soil nutrient level is exponentially related to yield (Melsted and Peck [1977](#page-20-25); Mitscherlich et al. [1913](#page-20-26)). In addition to these most common fve models, some are also using clustering algorithms such as the Cate-Nelson method (Mallarino and Blackmer [1992;](#page-20-27) Cate and Nelson [1971](#page-18-24)), alternative exponential models (e.g., used in the Better Fertilizer Decisions for Cropping Systems in Australia projects) or arcsine–log models (Dyson and Conyers [2013\)](#page-19-26). The calibration procedure to defne the critical soil nutrient level varies from least squared diferences to graphical methods (Nelsen and Anderson, 1977). Variation in the mathematical models certainly contribute to the observed range in critical soil nutrient levels (Perrin [1976](#page-20-28)) where the Mitscherlich model often results in higher critical values than the Cate-Nelson model (Colomb et al. [2007\)](#page-18-7) and where the linear-plateau also gives higher values than curvilinear models (Perrin [1976\)](#page-20-28). In addition, the Cate-Nelson often performs better than the Mitscherlich and quadratic models (Agboola and Ayodele [1987\)](#page-17-2). There is currently no broad consensus on which model should be preferred as well as the conditions determining the selection of the most appropriate one. One advantage of the Cate-Nelson model is that the critical level is not determined by relative yield level as opposed to the Mitscherlich model (Colomb et al. [2007\)](#page-18-7). Melsted and Peck [\(1977\)](#page-20-25) highlight that the crop response to variation in immobile nutrients such as P, K, Ca, and Mg can best be explained by exponential or quadratic models whereas linear models are usually better for mobile nutrients such as N and B. Despite the advantages of the Cate-Nelson methods, it has failed in some cases to identify critical soil nutrient levels (Heckman et al. [2006\)](#page-19-8). A broad statistical analysis of crop yields and soil nutrient levels available from various feld experiments across multiple sites would be needed to assess which model is best (in terms of explained variation) to derive a critical level.

3 Comparison of ranges in critical soil nutrient levels derived from experiments and those in medium soil fertility classes

3.1 Overall approach

The critical soil nutrient levels that are used to distinguish soil fertility classes are based on cut-off values for the relative yield. An illustration of the relationship between relative yield and soil nutrient levels, distinguishing low, medium and high soil fertility classes is given in Fig. [2](#page-7-0). In this example the critical level is set near a relative yield of ca 95% at the border of the medium and high soil fertility class, while the relative yield at the border between the low and medium class is set at 70%. In agronomy there are no fxed guidelines for the delineation of these classes, making the derivation prone to subjective and arbitrary choices of the researchers involved (Cate and Nelson [1971](#page-18-24)). For instance

Fig. 2 An illustration of the relationship between relative yield and soil nutrient levels, distinguishing low, medium, and high soil fertility classes. In this example the critical level is set near a relative yield of 100% but this may vary in literature from 80–95%. This level is generally near the upper range of the medium fertility class.

(Fageria et al. [1997](#page-19-24)) classifed soil fertility classes as follows: very low (< 70% RY); low (70–95% RY); medium (95–100% RY); and high (> 100%) whereas (Fageria and Santos [2008\)](#page-19-27) defned the medium class as 90–100% of RY. Ayodele and Agboola [\(1985\)](#page-18-20) classifed fertility classes as low for yield levels at 74%, medium at 94% and high at yield levels exceeding 97% of the maximum yield. On the other hand, Vieira et al. ([2016\)](#page-21-24) classifed medium class at those critical levels where the yield varies between ca 70 and 90% of the maximum yield. Overall, the medium soil fertility class is mostly related to crop yields that fall within 90 to 95% of the maximum yield, sometimes being as low as 70%. This is well linked to the cut-off values used to derive critical levels.

We compared the range in published critical soil nutrient values for the "medium" soil fertility status with the specifc critical values reported in peer reviewed feld trials. We hypothesized that the ranges should be comparable, meaning that all observed critical soil nutrient values from feld experiments should fall within the upper and lower critical value defning the "medium" soil fertility class. The critical soil nutrient level is thus assumed to coincide with the medium soil fertility class. We then assessed whether the range in soil nutrient levels in the "medium" soil fertility status can be reduced by accounting for diferences in site conditions and methodology.

3.2 Data collection

We frst searched the reported ranges in nutrient levels for diferent soil fertility classes, as being used in fertilizer recommendation systems, usually published in grey literature. Most of the studies distinguish three diferent soil nutrient classes being defned as "low," "medium," and "high" fertility. For some cases two addition classes

were defned being "very low" and "very high. We focus our discussion on the critical soil nutrient level represented by the "medium" soil fertility status class. This is achieved by establishing the lower and upper values associated with a specifc element and extraction method. The selected studies originated mainly from extension service providers, and university and agricultural departments. The studies were in some cases complemented with peer reviewed journal articles based on a google search using the keywords "fertilizer recommendations." All selected studies should at least contain the critical nutrient levels defning the soil fertility status (being classifed as very low, low, medium, high, and very high) and a description of the soil test done. Relevant studies and experiments were selected if the data consisted of a range that represented a soil fertility class for a specifc element and specifed extraction method. Where needed, units were converted to mg kg^{-1} . A total number of 36 publications were selected covering 12 nutrients, 17 soil tests, and 1 to 5 soil fertility status classes. Table [2](#page-8-0) provides a summarized overview of the range in critical soil nutrient values used to defne the medium soil fertility class. A detailed overview of the critical thresholds defning all soil fertility classes per nutrient and extraction method has been included in the supplementary material, Table S1.

A systematic search for critical soil nutrient levels in peer reviewed literature was additionally conducted by via SCOPUS on 7th September, 2020 using the following search parameters: "(TITLE-ABS-KEY (crop OR plant OR maize OR zea AND mays OR corn OR rice OR oryza AND sativa OR wheat OR triticum AND aestivum)) AND (((response OR growth OR yield OR " Nutrient Use efficiency" OR efect OR uptake OR "soil*test*correlat*" OR "soil* test* calibrati*" OR "soil*test*interpr*" OR "cate-nelson*" OR " mistcherlich*")) AND (phosphorus OR p OR potassium OR k OR zinc OR zn OR boron OR b OR sulfur OR sulphur

Table 2 Summary of the range in medium soil fertility classes and in critical level (the 90% interval) for each element for the given extraction methods.

	Element Extraction method	Range in medium soil fertility class, ppm (mg/kg)	Range in critical levels (mg/kg)
P	Bray 1	$12 - 41$	$9.3 - 40$
	Bray 2	$10 - 25$	$12 - 56$
	Olsen	$7 - 25$	$4.9 - 21$
	Colwell	$20 - 100$	$15 - 54$
	Mehlich 1	$8.8 - 13$	$13 - 58$
	Mehlich 3	$15 - 83$	$16 - 55$
K	Ammonium acetate	$75 - 280$	48-284
	Bray	91-110	$175 - 175$
	Colwell	80-120	$28 - 61$
	Mehlich 1	$47 - 90$	$48 - 129$
	Mehlich ₃	$35 - 201$	$32 - 72$
Zn	Mehlich 3	$0.6 - 5$	$1.8 - 13$
	DTPA	$0.25 - 3$	$0.3 - 13$
B	Hot water	$0.26 - 2$	$0.2 - 0.5$
Cu	DTPA	$0.1 - 2.59$	$1.3 - 1.7$
Fe	DTPA	$2.5 - 11.1$	$4.7 - 8.5$
Mn	Mehlich 3	$30 - 200$	$11 - 11$
	DTPA	$0.5 - 10$	$8.0 - 11$

OR s OR iron OR fe OR manganese OR mn OR copper OR cu)) AND (LIMIT-TO (EXACTKEYWORD , "crop yield")) AND (LIMIT-TO (LANGUAGE , "English")) AND (LIMIT-TO (SRCTYPE , "j"))."

A total number of 123 articles were selected. After excluding publications with incomplete data on soil nutrient levels and related crop response, only 61 were left for further analysis. For each of them, experiment details, site properties, crop type, soil nutrient level and response model used were retrieved. Each critical level was considered a data point, resulting in 448 data points (for details, see Table S2 in SI). A summary of the selected studies is given in Table [3.](#page-9-0) Again, all units were converted to mg kg^{-1} where needed.

Phosphorus was the most studied nutrient followed by K and Zn (Fig. 3), probably because P is a major nutrient whose availability is highly controlled by soil properties (in contrast to nitrogen which is driven by crop demand) while also being a nutrient often limiting crop production (Tandy et al. [2021\)](#page-21-0). Micronutrients such as B, Mn, Mo, and Cu are less frequently studied despite their relevance in the magnitude and nutritional quality of yield, probably due to methodological issues (requiring high laboratory skills and accurate equipment) or simply due to the absence of appropriate micronutrient fertilizers. Nevertheless, micronutrient defciencies are widespread limiting crop yield and nutritional quality for human intake (Mortvedt [1977](#page-20-4); Berkhout et al. [2017;](#page-18-25) Kihara et al. [2020;](#page-19-28) Kihara et al. [2017](#page-19-29); Graham and Welch [2000;](#page-19-30) Wakeel et al. [2018;](#page-21-25) Breure et al. [2023\)](#page-18-26). Generally, cereals are the most studied crop with wheat being the most assessed for multiple elements (Fig. [3](#page-10-0)) followed by maize. This dominance for P and the limited number of crops being analyzed shows that the scientific basis for most of other nutrients in the fertilizer recommendation systems is rather limited. The limited experimental data on nutrients other than P limits the quantitative analysis of the impact of methodology and site conditions (see Section 3.2) to phosphorus only. Given the similarities in soil and crop uptake processes, we might assume that the conclusions for P are also valid

3.3 Statistics used to evaluate factors controlling variation in critical levels

for the other nutrients.

We evaluated the impact of both methodology and agroecology on the variation in critical soil nutrient levels including site conditions. These include crop type and location (with location being a surrogate for various climate and soil properties) and methodological factors such as soil extraction method (see Section [2.2\)](#page-3-0), experimental type (see Section [2.3](#page-5-0)), and statistical model applied and cut-off value used (see Section [2.4\)](#page-6-2). Data were checked for normality and log-transformed if needed. Critical soil nutrient levels were standardized to unit variance for each extraction method to allow a proper intercomparison. The standardized data was subsequently used for the statistical analysis. For visualization purposes, data is shown in original scale in the supplementary material. To reduce the number of options for each of the categorial variables, we combined options that had low number of observations $(n < 5)$ and those where the group means were not statistically diferent (tested via a simple *t*-test).

Impact of location, crop type, extractant, experimental approach, statistical model, and cut-off value on critical values were analyzed for individual and combined efect on critical levels using multiple linear regression and ANOVA. We frst performed a main factor analysis to evaluate the impact of each factor on the variation in critical soil nutrient levels using the explained variance and the RMSE. We additionally evaluated their combined impact via twoway and three-way interactions assuming that the observed variation in critical soil nutrient levels across studies can be explained by the aforementioned factors.

3.4 Variation in critical levels as compared to ranges in medium soil fertility class

Table [2](#page-8-0) provides an overview of the range in critical soil nutrient values defning the medium soil fertility classes as compared to the range in critical soil nutrient values derived

Table 3 Summary of the analyzed results on critical levels from the systematic literature review.

Element	Number of Crop data points		Classification of experiment	Extraction method	Statistical method	Cut-off point
Phosphorus (P) 341		Alfafa (1),	Large dataset field experiments (202)	$0.1N$ AHDF (12)	Arcsine-log calibration 80% (25) curve (ALCC) (101)	
		Barley (16)	Long-term field experiments (42)	Modified Olsen (4)	Cate-Nelson (31)	85% (6)
		Canola (2)	Short-term field experiments (46)	AB-DTPA (1)	Exponential function (30)	87% (1)
		Clover (6)	Long-term amended field experiments (9)	Alkaline fluoride (2)	Fisher-Woodruff (3)	90\% (104)
		Cotton (1)	Short-term amended field experiments (4)	$AL-P(5)$	Linear model (12)	94% (3)
		Field Peas (1)	Short-term combined field and pot experiments (24)	Bray 1 (48)	Combined methods- linear; logarithmic; quadratic; Cate & Nelson etc. (26)	95% (83)
		Lupin (4)	Short-term pot experiment (14)	Bray 2 (9)	Linear plateau (40)	99% (7)
		Maize (120)		BSES (3)	Mistcherlich (73)	$94-96\%$ (8)
		Multiple crops (3)		CaCl2(3)	Quadratic plateau (15)	Economic (15)
		Peanuts (1)		Colwell (111)	Unspecified (10)	Max profit (3)
		Rice(5)		DGT(5)		Max yield (3)
		Sorghum (4)		Mehlich $1(7)$		Unspecified (83)
		Soybean (29)		Mehlich $2(2)$		
		Sunflower (3)		Mehlich 3 (77)		
		Wheat (144)		New Mehlich (4)		
				Olsen (41)		
				Resin (3)		
				Soltanpour (1)		
				Truog (2)		
				saturation extract (1)		
Potassium (K)	47	Barley (2)	Large dataset field experiments (5)	Ammonium acetate (20)	Cate-Nelson (11)	90% (19)
		Canola (1)	Short-term field experiments (34)	Blanchet and Perigaud (1)	Linear-plateau (9)	$93\% (7)$
		Cotton (1)	Long-term amended field experiments (3)	Bray (1)	Mitscherlich (18)	94% (1)
		Lupin (1)	Short-term pot experiment (5)	Colwell (5)	Quadratic plateau (6)	96% (4)
		Maize (15)		Hanway and Heidal (1) Unspecified (3)		Unspecified (16)
		Multiple crops (2)		Hunter and Pratt (1)		
		Peanuts (1)		Mehlich 1 (14)		
		Plantain (1)		Mehlich $3(4)$		
		Rice(7)				
		Soybean (8)				
		Wheat (6)				
		White Oats (2)				
$\text{Zinc}(\text{Zn})$	37	Canola (1)	Short-term field experiments (9)	Magnesium chloride (MgCl2)	Arcsine-log calibration 80% (12) curve (ALCC)ALCC method (1)	
		Maize (6)	Short-term pot experiment (28)	DTPA $(18) + 5$ (DTPA Cate-Nelson (14)) modified)		90% (6)

Element	Number of Crop data points		Classification of experiment	Extraction method	Statistical method	Cut-off point
		Rice(21)		EDTA $(5) + 5$ EDTA with ammonium carbonate	Column order procedure (1)	Unspecified (19)
		Wheat (9)		Mehlich $3(3)$	Interaction Chi-square (1)	
					Mistcherlich (4)	
					Unspecified (16)	
Iron (Fe) 6		Wheat (6)	Short-term field experiment (6)	DTPA(6)	Cate-Nelson (2)	80% (3)
					Interaction Chi-square (1)	Unspecified (3)
					Column order procedure (1)	
					Mitscherlich (2)	
Copper (Cu)	6	Wheat (6)	Short-term field experiment (6)	DTPA(6)	Cate-Nelson (2)	80% (3)
					Interaction Chi-square (1)	Unspecified (3)
					Column order procedure (1)	
					Mitscherlich (2)	
Boron (B)	4	Wheat (4)	Short-term field experiment (4)	Hot water	Interaction Chi-square (1)	80% (2)
					Column order procedure (1)	Unspecified (2)
					Mitscherlich (2)	

Table 3 (continued)

from feld experimental data. A detailed overview of the critical soil nutrient values for all soil fertility classes is given in Table S1 while additional details on the variation in critical levels are available in the shared dataset in excel.

Table [2](#page-8-0) shows that the critical soil nutrient values defning the medium soil fertility status varied from 7 to

Fig. 3 Overview of the proportion and total number of crops studied per nutrient.

100 mg P kg⁻¹, from 35 to 280 mg K kg⁻¹, from 0.25 to 5 mg Zn kg^{-1} , from 0.26 to 2 mg B kg^{-1} , from 0.1 to 2.59 mg Cu kg⁻¹, from 2.5 to 11 mg Fe kg⁻¹, and from 0.5 to 200 mg Mn $kg⁻¹$ for the nutrients included in fertilizer recommendation systems. Critical soil nutrient levels from experimental data varied from 0.33 to 96 mg kg⁻¹ for P, from 25 to 301 mg kg⁻¹ for K, and from 0.17 to 14.1 mg kg^{-1} for Zn. We conclude that for most of the nutrients, the critical soil nutrient levels are within the range of those used to defne the medium soil fertility status as being used in fertilizer recommendation systems (Table [2\)](#page-8-0).

For phosphorus (Fig. [4](#page-11-0)), the observed range in critical soil nutrient values for the Bray 1 soil test (varying from 3.5 to 45 mg P kg⁻¹) fits within the median soil fertility class ranging from 12 to 41 mg P kg^{-1}). Similarly, for P_{OLSEN} the critical values ranged from 3.9 to 24 mg P kg⁻¹ being similar to the critical values defning the fertility class (ranging between 7 and 25 mg P kg^{-1}). Same results are found for $P_{MEHLICH3}$ and $P_{COLWELL}$ with only a few outliers in both cases. For potassium determined via extraction with ammonium acetate, the critical soil nutrient values ranged between 30 and 301 mg K kg^{-1} , being mostly within the boundaries of the medium soil K fertility status (75 and 280 mg K kg⁻¹). Similar findings were found for $K_{\text{MEHI,ICHI}}$

Fig. 4 Comparison of critical levels for P across all crops for various extraction methods derived from experiments and those in medium soil fertility classes. Each point represents a critical level derived from an experiment. The extraction method is **a** Bray 1, **b** Bray 2, **c** Olsen, **d** Mehlich 1, **e** Mehlich 3, and **f** Colwell. The green dotted line indicates the lower threshold and the red line indicates the upper threshold of the medium soil fertility class.

with critical soil nutrient values varied from 30 to 129 mg K kg⁻¹ and the median K fertility class being defined as soils with $K_{MEHLICH1}$ varying between 47 and 90 mg K $kg⁻¹$. While there is quite some agreement between critical soil nutrient levels derived from feld experiments and the levels defning the soil fertility class, there is also a wide range in critical soil nutrient levels across the analyzed studies that warrants further exploration. This variation is at least dependent on the soil test analyzed, as shown by the small ranges found for P_{OLSEN} and $P_{COLWELL}$ (from experimental data) and P_{OLSEN} , $P_{MEHLICH1}$, P_{BRAY1} , and P_{BRAY2} (from fertilizer recommendation systems). For potassium (Fig. [5](#page-12-0)) as well as the micronutrients (Fig. [6\)](#page-13-0) was the observed range in experimental derived critical soil nutrient levels consistently smaller than those derived from fertilizer recommendation systems.

Our analysis confrms that the critical value underpinning FR systems matches those derived from experimental data, in particular for P, K, and Zn. However, we also note that this is partly due to the high variation observed in the critical soil nutrient levels defning the lower and upper boundary of the medium soil fertility status class (Figs. [4,](#page-11-0) [5](#page-12-0), and [6\)](#page-13-0). This agrees with earlier studies done for single nutrients like phosphorus (Jordan-Meille et al. [2012\)](#page-19-3) and has been explained by variation in crop type, climate, laboratory procedures, and soil properties (Jordan-Meille et al. [2012;](#page-19-3) Colomb et al. [2007\)](#page-18-7).

3.5 Site conditions and methodological aspects afecting critical levels

The variation in critical soil nutrient values was huge due to variation in site conditions and methodology. The following factors were analyzed: crop type, location, experimental approach, and statistical models.

3.5.1 Crop type

The impact of crop type on the variation in critical soil P levels are given in Fig. [7.](#page-13-1) Using critical levels

Fig. 5 Comparison of critical levels for K across all crops for various extraction methods derived from experiments and those in medium soil fertility classes. The extraction method is **a** ammonium acetate, **b** Mehlich 1, and **c** Mehlich 3. The green dotted line indicates the lower threshold and the red line indicates the upper threshold of the medium soil fertility class.

standardized by extraction method, wheat has generally higher critical levels as compared to maize and soybean while maize has lower critical levels. The absolute critical levels for crops are reported shown in supplementary material Figure S1. The critical levels for barley, soybean, clover, and wheat were not significantly different while these levels significantly varied between canola, maize, cotton, lupin, peanuts, rice, sorghum, and sunflower. Overall, the crop factor was significant in determining the critical level. Differentiating the P recommendation per crop type is therefore logic in order to sustain crop development for crops varying in rooting density and their ability to take up P from soil. Compared to other site conditions, crop type could explain 5% of the variation in critical limits. Highest critical soil P levels when standardized by extraction methods are required for barley and wheat and lowest for maize and rice. The observed differences in critical soil P levels among crops is not only associated with differences in rooting systems but also in the ability of crops to deal with water and nutrient stresses during the growing season (Brouder and Volenec [2008](#page-18-27)). Burak et al. ([2021\)](#page-18-28) found that barley had longer roots than maize; however, the maize had wider roots. Crops with more intensive rooting systems allow therefore lower critical soil nutrient levels than crops where the nutrient uptake is limited by soil diffusion. However, the critical level is not only dependent on the root volume but other factors including crop demand.

3.5.2 Location

About 29% of the variation in the critical soil P levels can be explained by the location ($p < 0.001$). The experiments done were located in various countries including Australia, USA, Brazil, Canada, China, Ethiopia, France, Hungarian, Kenya, Morocco, Nigeria, Tanzania, Madagascar, and Vietnam, confirming the variation in agroecological conditions related to these countries, such as rainfall, temperature, and soil properties. For instance, Feiziasl et al. ([2009\)](#page-19-31) highlight that a reduction in precipitation and temperature increases the critical nutrient level where others Conyers et al. ([2020\)](#page-19-5) showed that soil type and planting date also affects the critical level. If correct, this would imply that the derived critical soil nutrient levels are very specific for the regions where they have been determined. Since detailed soil and climatic variables for the experiments are unknown, we could not confirm or deny this conclusion. Other data-driven machine learning models have shown their ability to relate the variation in agronomic efficiency and nutrient use efficiency to site properties and management (Coulibali et al. [2020;](#page-19-6) Kirchmann et al. [2020](#page-19-32); Qin et al. [2018](#page-20-29)), suggesting that a more generic critical

Fig. 6 Comparison of critical levels for **a** Zn, **b** Cu, **c** Fe, and **d** Mn across all crops based on DTPA extraction method derived from experiments and those in medium soil fertility classes. The green dotted line indicates the lower threshold and the red line indicates the upper threshold of the medium soil fertility class.

Fig. 7 Impact of crop type on critical soil P levels, standardized for the various extraction methods. Critical levels are standardized to unit variance per extraction method.

soil nutrient level can be determined. All data-driven statistical approaches are by definition limited to the range in site conditions for which the models have been calibrated. Nevertheless, recent innovations in precision farming technologies suggest that by smart combination of sensor-derived estimates of soil properties and crop yield measurements on field and farm level can lead to tailor-made and efficient fertilizer recommendations (Guerrero et al. [2021;](#page-19-33) Maleki et al. [2007;](#page-20-30) Zhang et al. [2018](#page-21-26); Ros et al. [2021\)](#page-20-31).

3.5.3 Experimental approach

Seven experimental approaches have been used to related crop yield responses to soil nutrient availability, including large dataset based on feld experiments (LDF, *n* = 207); long-term amended feld experiments (LTAF, *n* = 12); longterm feld experiments (LTF, *n* = 42); short-term amended field experiments (STAF, $n = 4$); short-term field experiments (STF, $n = 112$); short-term pot experiments (STP, $n = 47$); and short-term pot and field experiments (STPF, $n = 24$). The experimental approach explains 24% of the variation in critical soil nutrient levels for P ($p < 0.05$). Combined with the crop type, however, it did not explain much additional variation in soil critical nutrient levels, suggesting that these factors were partly correlated. On average, the STF approach gave higher critical soil nutrient levels than the other approaches (Fig. [8\)](#page-14-0) and the absolute critical levels are shown in supplementary material Figure S2. Overall, feld experiments had relatively higher critical values (Fig. [8](#page-14-0)) compared to the pot experiments contrary to the findings of Ayodele and Agboola ([1985](#page-18-20)). However, Mortvedt ([1977](#page-20-4)) argues that achieving 90% relative yield in a pot versus in the field may lead to different nutrient requirements, with less nutrients needed to reach 90% in the pot than in the field. The field conditions imply that the greater a plant's potential for growth, the higher the minimum soil test level required to support its growth. Furthermore,

Fig. 8 Impact of experimental approach on the critical level for **a** P, **b** K, and **c** Zn across all crops, standardized for the various extraction methods. LDF: large dataset based on feld experiments; LTAF: longterm amended feld experiments; LTF: long-term feld experiments; STAF: short-term amended feld experiments; STF: short-term feld experiments; STP: short-term pot experiments; STPF: short-term pot and feld experiments.

interactive factors that might have affected the trial period such as climatic factors, management thereby altering the comparison of critical levels between the pot and field experiments. At the same time, it is also possible that there are usually more loses in the field than in a controlled environment thus leading to a higher critical level. Based on our results, amended field experiments (soil fertility classes) are a more suitable approach to determine the critical level than short-term experiments.

3.5.4 Statistical models

As expected, the statistical model used has substantial impact on the critical soil nutrient level derived from feld experimental data. About 20% of the variation in the critical soil nutrient levels could be explained by the model applied. Figure [9](#page-14-1) highlights the diferences observed among critical soil nutrient levels for P, K, and Zn being derived by various statistical models used and the absolute critical levels are shown in supplementary material Figure S3. For instance, for P, the mean (and standardized to unit variance) critical soil nutrient level declined in the order Mitscherlich, exponential, quadratic, linear, alcc and Cate-Nelson (Fig. [9](#page-14-1)a). A similar trend was observed for potassium (Fig. [9b](#page-14-1)). Diferences in critical soil nutrient values due to Cate-Nelson, exponential, and

Fig. 9 Impact of the model used on the standardized critical levels of **a** P, **b** K, and **c** Zn across all crops, standardized for the various extraction methods.

Mitscherlich models were therefore significant ($p < 0.05$). These fndings are similar to earlier conclusions derived from Colomb et al. [\(2007\)](#page-18-7) and Perrin [\(1976\)](#page-20-28). For example, Colomb et al. ([2007\)](#page-18-7) found that Mitscherlich (exponential) models resulted in critical soil nutrient levels being 1.3 to 1.8 times higher than levels being derived from Cate-Nelson models. In addition, linear plateau models have been suggested to result in lower fertilizer recommendations (and hence higher critical soil nutrient levels) than the curvilinear models when applied to the same dataset (Perrin [1976\)](#page-20-28). In three locations for soybean and maize, the exponential model led to higher critical levels followed by the quadratic plateau and linear plateau respectively (Dodd and Mallarino [2005\)](#page-19-9). In addition, nonlinear models are often preferred due to the underlying processes controlling crop development and studies show that their explained variance outcompetes the linear models (Alivelu et al. [2003](#page-18-29); Cox [1992](#page-19-4)). Recent studies are promoting the use of the Cate-Nelson model it looks at higher yields in positive quadrants thus aligns with the law of optimum better refects the actual situation in the feld than the historical law of the minimum (Lemaire et al. [2019](#page-19-1)).

Except for the Cate-Nelson model, all models use a specified cut-off values for the desired yield response. The cut-off point has substantial influence on the critical level across all elements, extraction method, and crop and explained about 14% of the variation in the critical soil nutrient levels. In any case, the relative yield depends on the agronomic intensity of the production system and the choice of the RY also depends on the acceptable economic risk level (Bell et al. [2013c\)](#page-18-30). For instance, 90% of 10 tons/ ha yield target and 90% of 5 ton/ha target are diferent but cannot be diferentiated by looking at the relative yield.

3.6 Disentangling the impacts of site conditions and methodological aspects on critical levels

The ranges of threshold soil nutrient levels defning the boundaries of the medium soil fertility class as well as the range in critical soil nutrient levels observed in feld

Fig. 10 The **a**–**c** explained variance and **d**–**f** standard error of P in the models. $V0 =$ accounts for no factors; $V1 =$ location; $V2 = location + soil extractant;$ $V3 =$ location + soil extractant + experimental approach; $V4 = location + soil extractant$ + experimental approach + statistical model; V5 = location + soil extractant + experimental approach + statistical $model + crop$; $V6 = location$ + soil extractant + experimental approach + statistical $model + crop + cut-off point.$ $MO =$ accounts for no factors; $M1 =$ soil extractant ; $M2 =$ soil extractant + experimental approach ; M3 = soil extractant + experimental approach + statistical model ; M4= soil extractant + experimental approach + statistical model + $cut-off$ point. $SO =$ accounts for no factors; S1= crop; S2= loca $tion + crop.$

experimental data was huge. This limits their applicability in FR systems outside the situation for which the critical levels have been derived. We analyzed the contribution of site conditions approximated by the location of the experiment and crop type and methodological factors being the soil test, the experimental approach, the statistical model used, and the cut-off value used. Combined site conditions and methodological aspects explained 51% of the variation in critical soil phosphorus levels observed in a wide range of experiments (Fig. [10](#page-15-0)a). However, the methodological aspects alone also explained 51% of the variation (Fig. [10b](#page-15-0)) while site conditions alone explained 30% of the variation (Fig. [10](#page-15-0)c). The contribution of individual factors explaining the variation in critical P limits was 29% for location, 5% for crop type, 24% for experimental approach, 20% for statistical model, and 14% for cut-off point and methodological factors emerged as the main driver of variations in critical levels, surpassing the infuence of site conditions. This is highlighted by the fact that incorporating site conditions did not notably enhance explained variation once methodological aspects were considered (see Fig. [10](#page-15-0)a, b). Even though location exerted the single most signifcant impact on the observed variations in critical soil P levels, it did not substantially contribute when difference in methodological aspect were considered.

Considering both the methodological aspects and site conditions, the mean standard error on prediction reduced by 53%, from ca 0.74 to ca 0.35 (Fig. [10](#page-15-0)d), but a similar reduction was found when considering the methodological aspects alone (Fig. [10e](#page-15-0)), whereas the site conditions alone reduced the mean standard error by 18% only, from ca 0.74 to 0.61 (Figure [10f](#page-15-0)). This results implies that correcting for methodological aspects can cause a potential reduction of ca 50% in the range of critical values. Imputation of this reduction on the range in critical P levels, as given in Table [2,](#page-8-0) implies that it would quite strongly reduce the range $($ = uncertainty $)$ for the critical soil nutrient levels defning the median soil fertility class as being used in fertilizer recommendations. Reducing the range around the mean by 50% implies that the range of P_{BRAY1} is reduced from 9.3–40 to 17–32 mg P kg⁻¹, while P_{BRAY2} is reduced from 12–56 to 23–45 mg P kg⁻¹, P_{OLSEN} from 4.9–21 to 8–17 mg P kg⁻¹, P_{COLWELL} from 15–54 to 25–44 mg P kg^{-1} , $P_{MEHLICH1}$ from 13–58 to 24–47 mg P kg⁻¹, and P_{MEHLICH3} from 16–55 to 26–45 mg P kg-1. A more accurate measure of the critical soil P level will evidently improve the nutrient use efficiency and avoid unnecessary build-up of P stocks in the soil and associated P losses via leaching, runoff, and erosion.

Although other attributes such as soil properties including the soil organic matter, clay content could have further reduced the critical level ranges (based on expectations from (Wuenscher et al. [2015\)](#page-21-14)), limited data availability did not allow a more detailed assessment. We hypothesized that location could be used as a surrogate for site conditions, including soil properties and climate, but our analysis shows that it is highly correlated with methodological aspects. Without original feld data of agro-ecological site conditions for the diferent experiments, it is impossible to disentangle their impact from methodological aspects affecting the derived crop nutrient response to nutrient availability. It is likely that even a stronger decline in the uncertainty of the estimated critical P levels is possible when the exact site properties controlling the crop response are quantifed. Supporting evidence can be deduced from recent initiatives where machine learning algorithms are trained to explain the crop response to variation in soil nutrient levels while accounting for agroecological site conditions such as weather, soil quality, and crop management measures (Timsina et al. [2021;](#page-21-27) Jayashree et al. [2022\)](#page-19-34). Therefore, establishing a minimum dataset requirement for soil test correlation and calibration studies as proposed by Slaton et al. ([2022\)](#page-20-10) and Conyers et al. [\(2013](#page-18-4)) would be a step towards improving fertilizer decision-making using evidence based information.

4 Conclusions and outlook

This study confrmed that the range in critical soil nutrient levels is comparable to the range in medium soil fertility classes used in fertilizer recommendation systems. This range is large. Our study thus aimed to unravel factors infuencing the derivation of critical levels in order to reduce this high uncertainty. Strong variation in observed critical values originate from methodological aspects and site conditions, both explaining 51% of the variation in the critical levels found for phosphorus. The geographical location explained most of the variation in critical P levels followed by experimental approach, extraction method, the statistical model used, its cut-off value to assess yield level, and crop type. The uncertainty in the critical soil P level declined with a similar percentage when accounting for methodological aspects and site conditions, showing that there is potential to develop fertilizer recommendation systems with more robust estimates (more limited ranges) for the critical nutrient level above which further fertilizer increase hardly increase the crop yield. A similar reduction in the range in critical levels might be expected for the other nutrients (K, Mg, B, Mn, Mo, Cu, and Zn) but the low availability of experimental data limited the approach to derive more robust critical soil nutrient values.

This review highlights that there is a clear potential to reduce the uncertainty (in particular the observed range) in critical soil nutrient levels by correcting diferences in methodological aspects. Their impact on the variation in critical nutrient levels was much bigger than the impact of location. This confrms our hypothesis that the variation in critical soil nutrient values declines when one accounts for

the methodological conditions under which these values have been derived. Our results for phosphorus indicate that a reduction of 50% in the uncertainty of critical levels is possible by harmonizing methodological aspects, implying that more generic and broadly applicable soil (P) tests are possible when such harmonization is practiced. We assumed that location proxied site conditions, such as climate and soil properties, but unfortunately location appears to be entangled with methodology in our study. Consequently, the impact of variation in site conditions on critical soil nutrient levels could not well be derived.

As long as a correction for methodology is not implemented, the current FR systems remains limited to the conditions for which the critical soil nutrient levels have been determined. For that reason, Lemaire et al. ([2019\)](#page-19-1) proposed an alternative FR system using plant diagnostics to optimize fertilization practices in view of crop demand. In addition, when site conditions afect the actual plant availability of nutrients or the risk for nutrient deficiency, a sustainable fertilizer recommendation system might differentiate per nutrient for those factors. In that way one accounts for the site conditions controlling the variation in critical soil nutrient levels required for optimum crop yield. For example, the soil P status in the German FR system is diferently evaluated for six soil texture classes and two land use categories. On the long term, we see potential for generic soil tests with narrow ranges defining the medium soil fertility class. There are currently two potential approaches to improve the reliability of soil-based FR systems: (i) statistical data driven approaches where the crop response is predicted in view of all site conditions (Chlingaryan et al. [2018](#page-18-31); Radočaj et al. [2022](#page-20-32); Barbedo [2019](#page-18-32)) and (ii) replacement of the empirical selected soil tests by soil test methods that reflect the mechanistic processes in soil controlling the plant availability (van Doorn et al. [2023\)](#page-21-28).

The use of standard data formats for documenting experiments and modelling crop yield responses to nutrient inputs will certainly facilitate the exchange of information and the correct derivation of critical soil nutrient levels (Slaton et al. [2022](#page-20-10)). Furthermore, more attention for the interaction between nutrients, including interactions between macro and micronutrients is needed, considering that micronutrients are often limiting yields in great areas of Africa (Rietra et al. [2017;](#page-20-33) Kihara et al. [2017](#page-19-29); Berkhout et al. [2017](#page-18-25); Kihara et al. [2020](#page-19-28)). The focus of one element when deriving relationships between crop yields and soil nutrient levels ignores those nutrient interactions, which are relevant in influencing crop response. Though establishing a reliable database will take time and require multi-stakeholder collaboration (Lyons et al. [2021](#page-20-34)), we foresee high potential for more generic fertilizer recommendation systems that make use of reliable data, more process-based interpretation of

nutrient pools and accounting for the interactions among nutrients as well as site conditions controlling the actual plant availability of nutrients (Lemaire et al. [2019](#page-19-1)).

Supplementary Information The online version contains supplementary material available at<https://doi.org/10.1007/s13593-023-00943-3>.

Authors' contributions SS: conceptualization, data collection and analysis, and writing—original draft; Gerard H. Ros: conceptualization, data analysis, and writing (reviewing and editing); Olusegun Yerokun: conceptualization and writing (reviewing and editing—original draft); and Wim de Vries: conceptualization and writing (reviewing and editing).

Funding This work is part of a PhD fellowship funded by Wageningen University and Research.

Data availability The data are readily available on request and all data analyzed during this study are included as part of the supplementary material.

Code availability The scripts used in data analysis are readily available on request.

Declarations

Conflict of interest The authors declare that there are no conficts of interest.

Ethical approval Not applicable.

Consent for publication Not applicable.

Consent to participate Not applicable.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit<http://creativecommons.org/licenses/by/4.0/>.

References

- Abay KA, Abay MH, Amare M, Berhane G, Aynekulu E (2022) Mismatch between soil nutrient defciencies and fertilizer applications: implications for yield responses in Ethiopia. Agr Econ 53(2):215–230.<https://doi.org/10.1111/agec.12689>
- Agboola A, Ayodele O (1987) Soil test calibration for upland rice in south western Nigeria. Fert Res 14(3):227-234. [https://doi.org/](https://doi.org/10.1007/BF01050667) [10.1007/BF01050667](https://doi.org/10.1007/BF01050667)
- Agegnehu G, Nelson PN, Bird MI, Van Beek C (2015) Phosphorus response and fertilizer recommendations for wheat grown on Nitisols in the central Ethiopian highlands. Commun Soil Sci Plant Anal 46(19):2411–2424. [https://doi.org/10.1080/00103624.](https://doi.org/10.1080/00103624.2015.10819222411) [2015.10819222411](https://doi.org/10.1080/00103624.2015.10819222411)
- Alivelu K, Srivastava S, Subba Rao A, Singh K, Selvakumari G, Raju N (2003) Comparison of modifed Mitscherlich and response plateau models for calibrating soil test based nitrogen recommendations for rice on Typic Ustropept. Commun Soil Sci Plant Anal 34(17–18):2633–2643.<https://doi.org/10.1081/CSS-120024790>
- Alva A (1993) Comparison of mehlich 3, mehlich 1, ammonium bicarbonate-DTPA, 1.0 M ammonium acetate, and 0.2 M ammonium chloride for extraction of calcium, magnesium, phosphorus, and potassium for a wide range of soils. Commun Soil Sci Plant Anal 24(7–8):603–612.<https://doi.org/10.1080/00103629309368826>
- Anderson GC, Peverill KI, Brennan RF (2013) Soil sulfur—crop response calibration relationships and criteria for feld crops grown in Australia. Crop Pasture Sci 64(5):523–530. [https://doi.](https://doi.org/10.1071/CP13244) [org/10.1071/CP13244](https://doi.org/10.1071/CP13244)
- Ayodele O, Agboola A (1985) Calibration of available P in soils from derived savannah zone of western Nigeria. Fert Res 6(2):121– 129. <https://doi.org/10.1007/BF01051006>
- Babu T, Tubana B, Paye W, Kanke Y, Datnoff L (2016) Establishing soil silicon test procedure and critical silicon level for rice in Louisiana soils. Commun Soil Sci Plant Anal 47(12):1578–1597
- Bado VB, Djaman K, Mel VC (2018) Developing fertilizer recommendations for rice in Sub-Saharan Africa, achievements and opportunities. Paddy Water Environ 16(3):571–586. [https://doi.](https://doi.org/10.1080/00103624.2016.1194996) [org/10.1080/00103624.2016.1194996](https://doi.org/10.1080/00103624.2016.1194996)
- Bai Z, Li H, Yang X, Zhou B, Shi X, Wang B, Li D, Shen J, Chen Q, Qin W (2013) The critical soil P levels for crop yield, soil fertility and environmental safety in diferent soil types. Plant Soil 372(1–2):27–37.<https://doi.org/10.1007/s11104-013-1696-y>
- Bansal K, Bhadoria U, Dube J (1985) Effect of applied potassium on nutrient contents of rice grown in three soils. Plant Soil 84(2):275–278. <https://doi.org/10.1007/BF02143190>
- Barbagelata PA, Mallarino AP (2013) Field correlation of potassium soil test methods based on dried and feld-moist soil samples for corn and soybean. Soil Sci Soc Am J 77(1):318–327. <https://doi.org/10.2136/sssaj2012.0253>
- Barbedo JGA (2019) Detection of nutrition defciencies in plants using proximal images and machine learning: a review. Comput Electron Agric 162:482–492. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.compag.2019.04.035) [compag.2019.04.035](https://doi.org/10.1016/j.compag.2019.04.035)
- Bell MJ, Moody PW, Anderson GC, Strong W (2013) Soil phosphorus—crop response calibration relationships and criteria for oilseeds, grain legumes and summer cereal crops grown in Australia. Crop Pasture Sci 64(5):499–513. [https://doi.org/](https://doi.org/10.1071/CP13016) [10.1071/CP13016](https://doi.org/10.1071/CP13016)
- Bell MJ, Strong W, Elliott D, Walker C (2013) Soil nitrogen—crop response calibration relationships and criteria for winter cereal crops grown in Australia. Crop Pasture Sci 64(5):442–460. <https://doi.org/10.1071/CP12431>
- Bell R, Reuter D, Scott B, Sparrow L, Strong W (2013) Soil phosphorus–crop response calibration relationships and criteria for winter cereal crops grown in Australia. Crop Pasture Sci 64(5):480–498. <https://doi.org/10.1071/CP12428>
- Bell R (2023) Diagnosis and prediction of deficiency and toxicity of nutrients. In: Marschner's mineral nutrition of plants. Elsevier, pp 477-495. [https://doi.org/10.1016/B978-0-12-819773-8.](https://doi.org/10.1016/B978-0-12-819773-8.00008-3) [00008-3](https://doi.org/10.1016/B978-0-12-819773-8.00008-3)
- Berger K, Truog E (1939) Boron determination in soils and plants. Ind Eng Chem 11(10):540–545. [https://doi.org/10.1021/ac501](https://doi.org/10.1021/ac50138a007) [38a007](https://doi.org/10.1021/ac50138a007)
- Berkhout E, Malan M, Kram T (2017) Micronutrients for agricultural intensifcation. Is Sub-Saharan Africa at risk?: policy study. PBL Netherlands Environmental Assessment Agency, The Hague, Netherlands
- Bilias F, Barbayiannis N (2019) Potassium availability: an approach using thermodynamic parameters derived from quantity-intensity

relationships. Geoderma 338:355–364. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.geoderma.2018.12.026) [geoderma.2018.12.026](https://doi.org/10.1016/j.geoderma.2018.12.026)

- Bortolon L, Gianello C (2010) Copper and zinc extracted by multielement solution for Southern Brazilian soils/Extracao de cobre e de zinco por solucoes multielementares em solos do sul do Brasil. Cienc Rural 40(3):670–674
- Bortolon L, Gianello C (2012) Multielement extraction from Southern Brazilian soils. Commun Soil Sci Plant Anal 43(12):1615–1624. <https://doi.org/10.1080/00103624.2012.681733>
- Bray RH, Kurtz L (1945) Determination of total, organic, and available forms of phosphorus in soils. Soil Sci 59(1):39–46
- Breitschuh G, Loide V, Timbare R, Staugaitis G, Spiegel H, Pikula D, Kotvas F, Ceh B, Cermak P, Loch J (2008) Soil testing methods and fertilizer recommendations in Central-Eastern European countries. Nawozy I Nawożenie (30)
- Brennan RF, Bell MJ (2013) Soil potassium—crop response calibration relationships and criteria for feld crops grown in Australia. Crop Pasture Sci 64(5):514–522.<https://doi.org/10.1071/CP13006>
- Breure MS, Njoroge S, Pasley HR, Hoffland E (2023) Exploring options for increasing maize yields and grain Zn concentrations in sub-Saharan Africa. Plant Soil:1-12. [https://doi.org/10.1007/](https://doi.org/10.1007/s11104-023-05998-5) [s11104-023-05998-5](https://doi.org/10.1007/s11104-023-05998-5)
- Brouder SM, Volenec JJ (2008) Impact of climate change on crop nutrient and water use efficiencies. Physiologia Plantarum 133(4):705–724. [https://doi.org/10.1111/j.1399-3054.2008.](https://doi.org/10.1111/j.1399-3054.2008.01136.x) [01136.x](https://doi.org/10.1111/j.1399-3054.2008.01136.x)
- Brtnicky M, Hammerschmiedt T, Elbl J, Kintl A, Skulcova L, Radziemska M, Latal O, Baltazar T, Kobzova E, Holatko J (2021) The potential of biochar made from agricultural residues to increase soil fertility and microbial activity: impacts on soils with varying sand content. Agronomy 11(6):1174. [https://doi.org/10.3390/](https://doi.org/10.3390/agronomy11061174) [agronomy11061174](https://doi.org/10.3390/agronomy11061174)
- Burak E, Dodd IC, Quinton JN (2021) Do root hairs of barley and maize roots reinforce soil under shear stress? Geoderma 383:114740.<https://doi.org/10.1016/j.geoderma.2020.114740>
- Cadot S, Bélanger G, Ziadi N, Morel C, Sinaj S (2018) Critical plant and soil phosphorus for wheat, maize, and rapeseed after 44 years of P fertilization. Nutr Cycl Agroecosys 112(3):417–433. <https://doi.org/10.1007/s10705-018-9956-0>
- Cate RB, Nelson LA (1971) A simple statistical procedure for partitioning soil test correlation data into two classes 1. Soil Sci Soc Am J 35(4):658–660. [https://doi.org/10.2136/sssaj1971.](https://doi.org/10.2136/sssaj1971.03615995003500040048x) [03615995003500040048x](https://doi.org/10.2136/sssaj1971.03615995003500040048x)
- Chlingaryan A, Sukkarieh S, Whelan B (2018) Machine learning approaches for crop yield prediction and nitrogen status estimation in precision agriculture: a review. Comput Electron Agric 151:61–69. [https://doi.org/10.1016/j.compag.2018.05.](https://doi.org/10.1016/j.compag.2018.05.012) [012](https://doi.org/10.1016/j.compag.2018.05.012)
- Colomb B, Debaeke P, Jouany C, Nolot J (2007) Phosphorus management in low input stockless cropping systems: crop and soil responses to contrasting P regimes in a 36-year experiment in southern France. Eur J Agron 26(2):154–165. [https://doi.org/](https://doi.org/10.1016/j.eja.2006.09.004) [10.1016/j.eja.2006.09.004](https://doi.org/10.1016/j.eja.2006.09.004)
- Colwell J (1963) The estimation of the phosphorus fertilizer requirements of wheat in southern New South Wales by soil analysis. Aust J Exp Agr 3(10):190–197. [https://doi.org/10.1071/EA963](https://doi.org/10.1071/EA9630190) [0190](https://doi.org/10.1071/EA9630190)
- Colwell J, Esdaile R (1968) The calibration, interpretation, and evaluation of tests for the phosphorus fertilizer requirements of wheat in northern New South Wales. Soil Res 6(1):105–120. <https://doi.org/10.1071/SR9680105>
- Conyers M, Bell M, Wilhelm N, Bell R, Norton R, Walker C (2013) Making better fertiliser decisions for cropping systems in Australia (BFDC): knowledge gaps and lessons learnt. Crop Pasture Sci 64(5):539–547.<https://doi.org/10.1071/CP13068>

- Conyers M, Bell R, Bell M (2020) Factors infuencing the soil-test calibration for Colwell P and wheat under winter-dominant rainfall. Crop Pasture Sci 71(2):113–118. [https://doi.org/10.](https://doi.org/10.1071/CP19375) [1071/CP19375](https://doi.org/10.1071/CP19375)
- Corrales I, Amenós M, Poschenrieder C, Barceló J (2007) Phosphorus efficiency and root exudates in two contrasting tropical maize varieties. J Plant Nutr 30(6):887–900. [https://doi.org/](https://doi.org/10.1080/15226510701375085) [10.1080/15226510701375085](https://doi.org/10.1080/15226510701375085)
- Coulibali Z, Cambouris AN, Parent S-É (2020) Site-specifc machine learning predictive fertilization models for potato crops in Eastern Canada. PLoS One 15(8):e0230888. [https://doi.org/](https://doi.org/10.1371/journal.pone.0230888) [10.1371/journal.pone.0230888](https://doi.org/10.1371/journal.pone.0230888)
- Cox F (1992) Range in soil phosphorus critical levels with time. Soil Sci Soc Am J 56(5):1504–1509. [https://doi.org/10.2136/sssaj](https://doi.org/10.2136/sssaj1992.03615995005600050028x) [1992.03615995005600050028x](https://doi.org/10.2136/sssaj1992.03615995005600050028x)
- Cox F (1996) Economic phosphorus fertilization using a linear response and plateau function. Commun Soil Sci Plant Anal 27(3–4):531–543.<https://doi.org/10.1080/00103629609369575>
- Cox F, Barnes J (2002) Peanut, corn, and cotton critical levels for phosphorus and potassium on Goldsboro soil. Commun Soil Sci Plant Anal 33(7–8):1173–1186.<https://doi.org/10.1081/CSS-120003880>
- Csathó P, Magyar M, Debreczeni K, Sárdi K (2002) Correlation between soil P and corn leaf P contents in a network of Hungarian long-term feld trials. Commun Soil Sci Plant Anal 33(15– 18):3085–3103.<https://doi.org/10.1081/CSS-120014508>
- Dodd JR, Mallarino AP (2005) Soil-test phosphorus and crop grain yield responses to long-term phosphorus fertilization for cornsoybean rotations. Soil Sci Soc Am J 69(4):1118–1128. [https://](https://doi.org/10.2136/sssaj2004.0279) doi.org/10.2136/sssaj2004.0279
- Doll E, Lucas R (1973) Testing soils for potassium, calcium and magnesium. Soil testing and plant analysis 3
- Dyson C, Conyers M (2013) Methodology for online biometric analysis of soil test–crop response datasets. Crop Past Sci 64(5):435–441. <https://doi.org/10.1071/CP13009>
- Eckert DJ (1987) Soil test interpretations: basic cation saturation ratios and sufficiency levels. Soil Test Sampl Correlat Calibr Interpret 21:53–64.<https://doi.org/10.2136/sssaspecpub21.c6>
- Egnér H, Riehm H, Domingo W (1960) Untersuchungen über die chemische Bodenanalyse als Grundlage für die Beurteilung des Nährstofzustandes der Böden. II. Chemische Extraktionsmethoden zur Phosphor-und Kaliumbestimmung Kungliga Lantbrukshögskolans Annaler 26:199–215
- Fageria N, Santos A (2008) Lowland rice response to thermophosphate fertilization. Commun Soil Sci Plant Anal 39(5–6):873–889. <https://doi.org/10.1080/00103620701881071>
- Fageria NK, Santos AB, Baligar VC (1997) Phosphorus soil test calibration for lowland rice on an inceptisol. Agron J 89(5):737–742. <https://doi.org/10.2134/agronj1997.00021962008900050005x>
- Feiziasl V, Jafarzadeh J, Pala M, Mosavi S (2009) Determination of critical levels of micronutrients by plant response column order procedure for dryland wheat (T. aestivum L.) in Northwest of Iran. Int J Soil Sci 4(1):14–26. <https://doi.org/10.3923/ijss.2009.14.26>
- Fixen P, Grove J, Westerman R (1990) Soil testing and plant analysis. Test Soils Phos Soil Sci Soc Am Book 3:141–180. [https://doi.](https://doi.org/10.2136/sssabookser3.3ed.c7) [org/10.2136/sssabookser3.3ed.c7](https://doi.org/10.2136/sssabookser3.3ed.c7)
- Fryer MS, Slaton NA, Roberts TL, Ross WJ (2019) Validation of soiltest-based phosphorus and potassium fertilizer recommendations for irrigated soybean. Soil Sci Soc Am J 83(3):825–837. [https://](https://doi.org/10.2136/sssaj2019.02.0032) doi.org/10.2136/sssaj2019.02.0032
- Graham RD, Welch RM (2000) Plant food micronutrient composition and human nutrition. Commun Soil Sci Plant Anal 31(11– 14):1627–1640.<https://doi.org/10.1080/00103620009370529>
- Guerrero A, De Neve S, Mouazen AM (2021) Chapter one - current sensor technologies for in situ and on-line measurement of soil nitrogen for variable rate fertilization: a review. In: Sparks DL

(ed) Advances in Agronomy, vol 168. Academic Press, pp 1-38. <https://doi.org/10.1016/bs.agron.2021.02.001>

- Harmsen J, Rulkens W, Eijsackers H (2005) Bioavailability: concept for understanding or tool for predicting? Land Contamin Reclam 13(2):161–172
- Hauser G (1973) Guide to the calibration of soil tests for fertilizer recommendations. FAO Soils Bulletin. Food and Agriculture Organization of the United Nations, Rome
- Heckman J, Jokela W, Morris T, Beegle D, Sims J, Coale F, Herbert S, Griffin T, Hoskins B, Jemison J (2006) Soil test calibration for predicting corn response to phosphorus in the northeast USA. Agron J 98(2):280–288.<https://doi.org/10.2134/agronj2005-0122>
- Hochmuth G, Hanlon E, Overman A (2011) Fertilizer experimentation, data analyses, and interpretation for developing fertilization recommendations—examples with vegetable crop research. vol SL345. Florida Cooperative Extension Service, Institute of Food and Agricultural Sciences, Florida
- Holford I, Doyle A (1992) Infuence of intensity/quantity characteristics of soil phosphorus tests on their relationships to phosphorus responsiveness of wheat under feld conditions. Soil Res 30(3):343–356.<https://doi.org/10.1071/SR9920343>
- Holford I, Morgan J, Bradley J, Cullis BR (1985) Yield responsiveness and response curvature as essential criteria for the evaluation and calibration of soil phosphate tests for wheat. Soil Res 23(2):167–180.<https://doi.org/10.1071/SR9850167>
- Houba V, Temminghoff E, Gaikhorst G, Van Vark W (2000) Soil analysis procedures using 0.01 M calcium chloride as extraction reagent. Commun Soil Sci Plant Anal 31(9–10):1299–1396. [https://](https://doi.org/10.1080/00103620009370514) doi.org/10.1080/00103620009370514
- Islam A, Karim AJMS, Solaiman ARM, Islam MS, Saleque MA (2017) Eight-year long potassium fertilization efects on quantity/intensity relationship of soil potassium under double rice cropping. Soil Till Res 169:99–117.<https://doi.org/10.1016/j.still.2017.02.002>
- Izsaki Z (2009) Efect of nitrogen supply on nutritional status of maize. Commun Soil Sci Plant Anal 40(1–6):960–973. [https://doi.org/](https://doi.org/10.1080/00103620802693482) [10.1080/00103620802693482](https://doi.org/10.1080/00103620802693482)
- Jayashree D, Pandithurai O, Paul Jasmin Rani L, Menon PK, Beria MV, Nithyalakshmi S (2022) Fertilizer recommendation system using machine learning. In: Disruptive technologies for big data and cloud applications: Proceedings of ICBDCC 2021. Springer, pp 709-716. https://doi.org/10.1007/978-981-19-2177-3_66
- Jordan-Meille L, Rubæk GH, Ehlert PAI, Genot V, Hofman G, Goulding K, Recknagel J, Provolo G, Barraclough P (2012) An overview of fertilizer-P recommendations in Europe: soil testing, calibration and fertilizer recommendations. Soil Use Manage 28(4):419–435. <https://doi.org/10.1111/j.1475-2743.2012.00453.x>
- Kihara J, Sileshi GW, Nziguheba G, Kinyua M, Zingore S, Sommer R (2017) Application of secondary nutrients and micronutrients increases crop yields in sub-Saharan Africa. Agron Sustain Dev 37(4):25.<https://doi.org/10.1007/s13593-017-0431-0>
- Kihara J, Bolo P, Kinyua M, Rurinda J, Piikki K (2020) Micronutrient defciencies in African soils and the human nutritional nexus: opportunities with staple crops. Environ Geochem Health 42(9):3015–3033.<https://doi.org/10.1007/s10653-019-00499-w>
- Kirchmann H, Börjesson G, Bolinder MA, Kätterer T, Djodjic F (2020) Soil properties currently limiting crop yields in Swedish agriculture–an analysis of 90 yield survey districts and 10 long-term feld experiments. Eur J Agron 120:126132. [https://doi.org/10.](https://doi.org/10.1016/j.eja.2020.126132) [1016/j.eja.2020.126132](https://doi.org/10.1016/j.eja.2020.126132)
- Lemaire G, Sinclair T, Sadras V, Bélanger G (2019) Allometric approach to crop nutrition and implications for crop diagnosis and phenotyping. A review. Agron Sustain Dev 39(2):1–17. <https://doi.org/10.1007/s13593-019-0570-6>
- Lindsay WL, Norvell WA (1978) Development of a DTPA soil test for zinc, iron, manganese, and copper1. Soil Sci Soc Am J

42(3):421–428. [https://doi.org/10.2136/sssaj1978.0361599500](https://doi.org/10.2136/sssaj1978.03615995004200030009x) [4200030009x](https://doi.org/10.2136/sssaj1978.03615995004200030009x)

- Liu D-Y, Zhang W, Yan P, Chen X-P, Zhang F-S, Zou C-Q (2017) Soil application of zinc fertilizer could achieve high yield and high grain zinc concentration in maize. Plant Soil 411(1–2):47–55. <https://doi.org/10.1007/s11104-016-3105-9>
- Locke MA, Hanson RG (1991) Calibration of corn response to Bray I, Bray II, and Mehlich II extractable soil phosphorus. Commun Soil Sci Plant Anal 22(11–12):1101–1121. [https://doi.org/10.](https://doi.org/10.1080/00103629109368478) [1080/00103629109368478](https://doi.org/10.1080/00103629109368478)
- Lyons SE, Arthur DK, Slaton NA, Pearce AW, Spargo JT, Osmond DL, Kleinman PJ (2021) Development of a soil test correlation and calibration database for the USA. Agr Environ 6(4):e20058. <https://doi.org/10.1002/ael2.20058>
- Maleki MR, Mouazen AM, Ramon H, De Baerdemaeker J (2007) Optimisation of soil VIS–NIR sensor-based variable rate application system of soil phosphorus. Soil Till Res 94(1):239–250. [https://](https://doi.org/10.1016/j.still.2006.07.016) doi.org/10.1016/j.still.2006.07.016
- Mallarino A, Blackmer A (1992) Comparison of methods for determining critical concentrations of soil test phosphorus for corn. Agron J 84(5):850–856. [https://doi.org/10.2134/agronj1992.](https://doi.org/10.2134/agronj1992.00021962008400050017x) [00021962008400050017x](https://doi.org/10.2134/agronj1992.00021962008400050017x)
- Marchand S, Parent S-É, Deland J-P, Parent L-É (2013) Nutrient signature of Quebec (Canada) cranberry (Vaccinium macrocarpon Ait.). Revista Brasileira de Fruticultura 35:292–304
- Martínez G, Snyder V, Vázquez M, González-Vélez A, Guzmán JL (2002) Factors afecting magnesium availability to plantains in highly weathered soils. J Agric Uni Pue. [https://doi.org/10.](https://doi.org/10.46429/jaupr.v86i1-2.3179) [46429/jaupr.v86i1-2.3179](https://doi.org/10.46429/jaupr.v86i1-2.3179)
- Mashayekhi P, Abbasi Z, Tatari M, Mahmoodi-Eshkaftaki M (2014) Determination of soil feature effects on plant-available phosphorus extraction using response surface and Cate-Nelson methodology. Commun Soil Sci Plant Anal 45(15):2046–2057. [https://doi.](https://doi.org/10.1080/00103624.2014.923437) [org/10.1080/00103624.2014.923437](https://doi.org/10.1080/00103624.2014.923437)
- Mehlich A (1978) New extractant for soil test evaluation of phosphorus, potassium, magnesium, calcium, sodium, manganese and zinc. Commun Soil Sci Plant Anal 9(6):477–492. [https://doi.org/](https://doi.org/10.1080/00103627809366824) [10.1080/00103627809366824](https://doi.org/10.1080/00103627809366824)
- Mehlich A (1984) Mehlich 3 soil test extractant: a modification of Mehlich 2 extractant. Commun Soil Sci Plant Anal 15(12):1409–1416
- Mehlich A (1953) Determination of P, Ca, mg, K, Na, and NH4. North Carolina Soil Test Division (Mimeo 1953):23-89
- Melsted S, Peck T (1977) The Mitscherlich-Bray growth function. Soil Test Correlat Interpreting Anal Results 29:1–18. [https://doi.org/](https://doi.org/10.2134/asaspecpub29) [10.2134/asaspecpub29](https://doi.org/10.2134/asaspecpub29)
- Mitchell CC, Huluka G (2012) Basis of soil testing in Alabama (Agronomy and Soils Departmental Series), vol 324A. Auburn University, Alabama
- Mitchell CC, Huluka G (2012) Nutrient recommendation tables for alabama crops (Agronomy and Soils Departmental Series), vol 324B. Auburn University, Alabama
- Mitscherlich E, Fischer H, Bowser L, Strigel A, Dodt J, Schmitz B (1913) Zur quantitativen Bestimmung des Kaliums. Z Anal Chem 52(9):587–590.<https://doi.org/10.1007/BF01347169>
- Mohamed ES, Saleh AM, Belal AB, Gad AA (2018) Application of near-infrared refectance for quantitative assessment of soil properties. Egy J Rem Sens Spa Sc 21(1):1–14. [https://doi.org/10.](https://doi.org/10.1016/j.ejrs.2017.02.001) [1016/j.ejrs.2017.02.001](https://doi.org/10.1016/j.ejrs.2017.02.001)
- Mortvedt J (1977) Micronutrient soil test correlations and interpretations. Soil Test Corre Interpret Anal Results 29:99–117. [https://](https://doi.org/10.2134/asaspecpub29.c7) doi.org/10.2134/asaspecpub29.c7
- Nafu A, Abiodun M, Okpara I, Chude V (2012) Soil fertility evaluation: a potential tool for predicting fertilizer requirement for crops in Nigeria. Afr J Agr Res 7(47):6204–6214. [https://doi.](https://doi.org/10.5897/AJAR12.210) [org/10.5897/AJAR12.210](https://doi.org/10.5897/AJAR12.210)
- Neyroud J-A, Lischer P (2003) Do diferent methods used to estimate soil phosphorus availability across Europe give comparable results? J Plant Nutr Soil Sci 166(4):422–431. [https://doi.org/](https://doi.org/10.1002/jpln.200321152) [10.1002/jpln.200321152](https://doi.org/10.1002/jpln.200321152)
- Olfs HW, Blankenau K, Brentrup F, Jasper J, Link A, Lammel J (2005) Soil-and plant-based nitrogen-fertilizer recommendations in arable farming. J Plant Nutr Soil Sci 168(4):414–431. [https://doi.](https://doi.org/10.1002/jpln.200520526) [org/10.1002/jpln.200520526](https://doi.org/10.1002/jpln.200520526)
- Olsen SR (1954) Estimation of available phosphorus in soils by extraction with sodium bicarbonate. United States Department Of Agriculture, Washington
- Ostatek-Boczynski ZA, Lee-Steere P (2012) Evaluation of Mehlich 3 as a universal nutrient extractant for Australian sugarcane soils. Commun Soil Sci Plant Anal 43(4):623–630. [https://doi.org/10.](https://doi.org/10.1080/00103624.2012.643845) [1080/00103624.2012.643845](https://doi.org/10.1080/00103624.2012.643845)
- Perrin RK (1976) The value of information and the value of theoretical models in crop response research. Am J Agric Econ 58(1):54–61. <https://doi.org/10.2307/1238577>
- Qin Z, Myers DB, Ransom CJ, Kitchen NR, Liang SZ, Camberato JJ, Carter PR, Ferguson RB, Fernandez FG, Franzen DW (2018) Application of machine learning methodologies for predicting corn economic optimal nitrogen rate. Agron J 110(6):2596–2607. <https://doi.org/10.2134/agronj2018.03.0222>
- Radočaj D, Jurišić M, Gašparović M (2022) The role of remote sensing data and methods in a modern approach to fertilization in precision agriculture. Remote Sens-Basel 14(3):778. [https://doi.org/](https://doi.org/10.3390/rs14030778) [10.3390/rs14030778](https://doi.org/10.3390/rs14030778)
- Rietra RP, Heinen M, Dimkpa CO, Bindraban PS (2017) Efects of nutrient antagonism and synergism on yield and fertilizer use efficiency. Commun Soil Sci Plant Anal 48(16):1895–1920. <https://doi.org/10.1080/00103624.2017.1407429>
- Ros G, Temminghoff E, Hoffland E (2011) Nitrogen mineralization: a review and meta-analysis of the predictive value of soil tests. Eur J Soil Sci 62(1):162–173. [https://doi.org/10.1111/j.1365-](https://doi.org/10.1111/j.1365-2389.2010.01318.x) [2389.2010.01318.x](https://doi.org/10.1111/j.1365-2389.2010.01318.x)
- Ros G, Van Rotterdam A, Bussink D, Bindraban P (2016) Selenium fertilization strategies for bio-fortifcation of food: an agroecosystem approach. Plant Soil 404(1):99–112. [https://doi.org/](https://doi.org/10.1007/s11104-016-2830-4) [10.1007/s11104-016-2830-4](https://doi.org/10.1007/s11104-016-2830-4)
- Ros GH, Luleva M, de Vries W (2021) Soil analysis is pivotal for fertilizer recommendations: Comment on "Soil based, feld specifc fertilizer recommendations are a pipe-dream" by A.G.T. Schut and K.E. Giller. Geoderma [\(https://doi.org/10.1016/j.](https://doi.org/10.1016/j.geoderma.2020.114680) [geoderma.2020.114680\)](https://doi.org/10.1016/j.geoderma.2020.114680). Geoderma 387:114861. [https://doi.](https://doi.org/10.1016/j.geoderma.2020.114861) [org/10.1016/j.geoderma.2020.114861](https://doi.org/10.1016/j.geoderma.2020.114861)
- Sánchez-Alcalá I, Del Campillo M, Torrent J (2014) Extraction with 0.01 M CaCl2 underestimates the concentration of phosphorus in the soil solution. Soil Use Manage 30(2):297–302. [https://](https://doi.org/10.1111/sum.12116) doi.org/10.1111/sum.12116
- Savoy H (2013) Procedures used by state soil testing laboratories in the southern region of the United States. Southern Cooperative Series Bulletin 409. Clemson Experiment Station
- Schüller H (1969) Die CAL-Methode, eine neue Methode zur Bestimmung des pfanzenverfügbaren Phosphates in Böden. Zeitschrift für Pfanzenernährung und Bodenkunde 123(1):48– 63. <https://doi.org/10.1002/jpln.19691230106>
- Schut AG, Giller KE (2020) Soil-based, feld-specifc fertilizer recommendations are a pipe-dream. Geoderma 380:114680. <https://doi.org/10.1016/j.geoderma.2020.114680>
- Simard R (1993) Ammonium acetate-extractable elements. In: M.R. C, E.G. G (eds) Soil sampling and methods of analysis, vol 1. Lewis Publishers, Florida, USA, pp 39-42
- Slaton NA, Lyons SE, Osmond DL, Brouder SM, Culman SW, Drescher G, Gatiboni LC, Hoben J, Kleinman PJ, McGrath JM (2022) Minimum dataset and metadata guidelines for

soil-test correlation and calibration research. Soil Sci Soc Am J 86(1):19–33.<https://doi.org/10.1002/saj2.20338>

- Soltanpour P (1985) Use of ammonium bicarbonate DTPA soil test to evaluate elemental availability and toxicity. Commun Soil Sci Plant Anal 16(3):323–338. [https://doi.org/10.1080/00103](https://doi.org/10.1080/00103628509367607) [628509367607](https://doi.org/10.1080/00103628509367607)
- Soltanpour P, Workman S (1979) Modifcation of the NH4 HCO3- DTPA soil test to omit carbon black. Commun Soil Sci Plant Anal 10(11):1411–1420.<https://doi.org/10.1080/00103627909366996>
- Speirs SD, Scott BJ, Moody PW, Mason SD (2013) Soil phosphorus tests II: a comparison of soil test-crop response relationships for diferent soil tests and wheat. Crop Pasture Sci 64(5):469– 479.<https://doi.org/10.1071/CP13111>
- Srivastava S, Subba Rao A, Alivelu K, Singh K, Raju N, Rathore A (2006) Evaluation of crop responses to applied fertilizer phosphorus and derivation of optimum recommendations using the Mitscherlich-Bray equation. Commun Soil Sci Plant Anal 37(05– 06):847–858. <https://doi.org/10.1080/00103620600564182>
- Srivastava PC, Naresh M, Srivastava P (2008) Appraisal of some soil tests for zinc availability to late-sown wheat grown in Mollisols. Commun Soil Sci Plant Anal 39(3–4):440–449. [https://doi.org/](https://doi.org/10.1080/00103620701826563) [10.1080/00103620701826563](https://doi.org/10.1080/00103620701826563)
- Steinfurth K, Hirte J, Morel C, Buczko U (2021) Conversion equations between Olsen-P and other methods used to assess plant available soil phosphorus in Europe – a review. Geoderma 401:115339. <https://doi.org/10.1016/j.geoderma.2021.115339>
- Steinfurth K, Börjesson G, Denoroy P, Eichler-Löbermann B, Gans W, Heyn J, Hirte J, Huyghebaert B, Jouany C, Koch D (2022) Thresholds of target phosphorus fertility classes in European fertilizer recommendations in relation to critical soil test phosphorus values derived from the analysis of 55 European long-term feld experiments. Agric Ecosyst Environ 332:107926. [https://](https://doi.org/10.1016/j.agee.2022.107926) doi.org/10.1016/j.agee.2022.107926
- Tandy S, Hawkins JM, Dunham SJ, Hernandez-Allica J, Granger SJ, Yuan H, McGrath SP, Blackwell MS (2021) Investigation of the soil properties that afect Olsen P critical values in diferent soil types and impact on P fertiliser recommendations. Eur J Soil Sci 72(4):1802–1816.<https://doi.org/10.1111/ejss.13082>
- Tiller K, Honeysett J, De Vries M (1972) Soil zinc and its uptake by plants. II Soil chemistry in relation to prediction of availability. Soil Res 10(2):165–182. <https://doi.org/10.1071/SR9720165>
- Timsina J, Dutta S, Devkota KP, Chakraborty S, Neupane RK, Bishta S, Amgain LP, Singh VK, Islam S, Majumdar K (2021) Improved nutrient management in cereals using Nutrient Expert and machine learning tools: productivity, proftability and nutrient use efficiency. Agr Syst 192:103181. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.agsy.2021.103181) [agsy.2021.103181](https://doi.org/10.1016/j.agsy.2021.103181)
- Trierweiler J, Lindsay W (1969) EDTA-ammonium carbonate soil test for zinc. Soil Sci Soc Am J 33(1):49–54. [https://doi.org/10.2136/](https://doi.org/10.2136/sssaj1969.03615995003300010017x) [sssaj1969.03615995003300010017x](https://doi.org/10.2136/sssaj1969.03615995003300010017x)
- Truog E (1930) Determination of the readily available phosphorus of soils. J Am Soc Agr. [https://doi.org/10.2134/agronj1930.00021](https://doi.org/10.2134/agronj1930.00021962002200100008x) [962002200100008x](https://doi.org/10.2134/agronj1930.00021962002200100008x)
- Tunney H, Breeuwsma A, Withers P, Ehlert P (1997) Phosphorus fertilizer strategies: present and future. In: H. Tunney OTC, P. C. Brookes, & A. E. Johnston (ed) Phosphorus loss from soil to water. Phosphorus loss from soil to water. CAB International, Wallingford, pp 177-203
- Ussiri DA, Mnkeni PNS, MacKenzie AF, Semoka JMR (1998) Soil test calibration studies for formulation of phosphorus fertilizer recommendations for maize in Morogoro district, Tanzania. I Evaluation of soil test methods. Commun Soil Sci Plant Anal 29(17–18):2801–2813
- Valkama E, Uusitalo R, Ylivainio K, Virkajärvi P, Turtola E (2009) Phosphorus fertilization: a meta-analysis of 80 years of research in Finland. Agric Ecosyst Environ 130(3–4):75–85. [https://doi.](https://doi.org/10.1016/j.agee.2008.12.004) [org/10.1016/j.agee.2008.12.004](https://doi.org/10.1016/j.agee.2008.12.004)
- van Heerwaarden J (2022) The theoretical potential for tailored fertilizer application The case of maize in Sub-Saharan Africa. Field Crops Res 288:108677.<https://doi.org/10.1016/j.fcr.2022.108677>
- van Doorn M, van Rotterdam D, Ros GH, Koopmans G, Smolders E, de Vries W (2023) The phosphorus saturation degree as a universal agronomic and environmental soil P test. Crit Rev Env Sci Tec. <https://doi.org/10.1080/10643389.2023.2240211>
- Vieira RCB, Fontoura SMV, Bayer C, Moraes RPd, Carniel E (2016) Potassium fertilization for long term no-till crop rotation in the Central-Southern region of Paraná, Brazil. Revista Brasileira de Ciência do Solo 40. [https://doi.org/10.1590/18069657rbcs201](https://doi.org/10.1590/18069657rbcs20150193) [50193](https://doi.org/10.1590/18069657rbcs20150193)
- Voss R (1998) Fertility recommendations: past and present. Commun Soil Sci Plant Anal 29(11–14):1429–1440. [https://doi.org/10.](https://doi.org/10.1080/00103629809370040) [1080/00103629809370040](https://doi.org/10.1080/00103629809370040)
- Wakeel A, Farooq M, Bashir K, Ozturk L (2018) Micronutrient malnutrition and biofortifcation: recent advances and future perspectives. Plant Micronutr Effic. 225-243. [https://doi.org/10.1016/](https://doi.org/10.1016/B978-0-12-812104-7.00017-4) [B978-0-12-812104-7.00017-4](https://doi.org/10.1016/B978-0-12-812104-7.00017-4)
- Wang JJ, Harrell DL, Bell PF (2004) Potassium bufering characteristics of three soils low in exchangeable potassium. Soil Sci Soc Am J 68(2):654–661.<https://doi.org/10.2136/sssaj2004.6540>
- Waugh D, Cate Jr R, Nelson L (1973) Discontinuous models for rapid correlation, interpretation, and utilization of soil analysis and fertilizer response data. International Soil Fertility Evaluation and Improvement Program, Raleigh
- Weetman G, Wells C (1990) Plant analyses as an aid in fertilizing forests. Soil Test Plant Anal 3:659–690. [https://doi.org/10.2136/](https://doi.org/10.2136/sssabookser3.3ed.c25) [sssabookser3.3ed.c25](https://doi.org/10.2136/sssabookser3.3ed.c25)
- Wuenscher R, Unterfrauner H, Peticzka R, Zehetner F (2015) A comparison of 14 soil phosphorus extraction methods applied to 50 agricultural soils from Central Europe. Plant Soil Environ 61(2):86–96.<https://doi.org/10.17221/932/2014-PSE>
- Yuan M, Fernández FG, Pittelkow CM, Greer KD, Schaefer D (2020) Soil and crop response to phosphorus and potassium management under conservation tillage. Agron J 112(3):2302–2316. <https://doi.org/10.1002/agj2.20114>
- Zhang H, Davison W, Gadi R, Kobayashi T (1998) In situ measurement of dissolved phosphorus in natural waters using DGT. Anal Chim Acta 370(1):29–38. [https://doi.org/10.1016/S0003-2670\(98\)00250-5](https://doi.org/10.1016/S0003-2670(98)00250-5)
- Zhang Y, Chen D, Wang S, Tian L (2018) A promising trend for feld information collection: an air-ground multi-sensor monitoring system. Inform Process Agric 5(2):224–233. [https://doi.org/10.](https://doi.org/10.1016/j.inpa.2018.02.002) [1016/j.inpa.2018.02.002](https://doi.org/10.1016/j.inpa.2018.02.002)
- Zhang H, Antonangelo J, Grove J, Osmond D, Slaton NA, Alford S, Florence R, Huluka G, Hardy DH, Lessl J (2021) Variation in soil-test-based phosphorus and potassium rate recommendations across the southern USA. Soil Sci Soc Am J 85(4):975–988. <https://doi.org/10.1002/saj2.20280>
- Zone PP, Culman SW, Haden VR, Lindsey LE, Fulford AM, Zhao K (2020) Do soil test levels and fertilization with phosphorus and potassium impact feld crop tissue concentrations? Agron J 112(4):3024–3036. <https://doi.org/10.1002/agj2.20243>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.