

Concentrations, Health Risks and Sources of Polycyclic Aromatic Hydrocarbons in Nigerian Honey

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Abstract

The concentrations of the US EPA 16 priority polycyclic aromatic hydrocarbons (PAHs) were measured in honey samples collected from different regions of Nigeria with a view to providing information on the extent of contamination, regional profiles, sources and risks of PAHs in this food type. The concentrations of the PAHs were determined by using gas chromatography-mass spectrometry after extraction by ultra-sonication with *n*-hexane and dichloromethane. The concentrations of $\Sigma 16$ PAHs in the honey samples ranged from 169–522 $\mu\text{g kg}^{-1}$, 97.2–1980 $\mu\text{g kg}^{-1}$, 180–641 $\mu\text{g kg}^{-1}$ and 122–357 $\mu\text{g kg}^{-1}$ for South-East, South-West, Niger Delta, and North Central regions respectively. The compositional patterns of PAHs in the analyzed honey samples followed the order: 5-rings > 4-rings > 3-rings > 6-rings > 2-rings. The estimated daily intake values from consumption of these honey samples ranged from not detected (nd) to 1.9 $\text{ng kg}^{-1} \text{bw day}^{-1}$, nd to 5.9 $\text{ng kg}^{-1} \text{bw day}^{-1}$, nd to 18 $\text{ng kg}^{-1} \text{bw day}^{-1}$ and 0.6 to 33 $\text{ng kg}^{-1} \text{bw day}^{-1}$ for BaP, PAH2, PAH4 and PAH8 respectively. The incremental life cancer risk and margin of exposure values for the majority of the samples indicate

that there is no risk associated with the consumption of these honey samples. The PAH isomeric ratios and principal component analysis indicated that combustion of fossil fuels, natural gas and biomass, and automobile emissions were the main sources of PAHs in these samples from the different regions of Nigeria.

Keywords: Honey, Polycyclic aromatic hydrocarbons, Risk assessment, Nigeria

Introduction

Honey is a natural product produced by *Apis mellifera* bees from the nectar or secretions of plants, and does not usually contain any additives or preservatives¹. It contains a number of nutritionally valuable compounds and has healing, prophylactic, anti-oxidative, anti-bacterial and immune-enhancing properties^{2,3}. Honey is made up of a mixture of carbohydrates, such as fructose (25–45% m/m), glucose (25–37% m/m), maltose (2–12% m/m) and sucrose (0.5–3% m/m) with traces of other sugars, and water (14–18% m/m)⁴ as well as small amounts of a wide array of vitamins, mineral substances (0.1 to 1.0% m/m)⁵, amino acids and antioxidants¹. The colour, flavour, carbohydrate composition and mineral, amino acid and antioxidant content of honey often varies with floral types¹. The concentrations of contaminants in honey reflect the conditions of the environment and bee-keeping practices. Since the forage area of the bee hive is very large (more than 7 km²) and the bees come in contact with the basic components of the environment (air, soil and water), the concentrations of contaminants in honey reflect their amounts in the whole region⁴. Thus, honey can serve as a useful environmental marker and bioindicator for monitoring environmental contaminants^{6–9}. Despite the known nutritional and therapeutic properties of honey, the occurrence of xenobiotics, such as metals, pesticides and polycyclic aromatic hydrocarbons, in honey may constitute a serious threat to consumers.

Polycyclic aromatic hydrocarbons (PAHs) are a diverse group of over one hundred (100) organic compounds consisting of two or more fused aromatic rings¹⁰ and/or pentacyclic rings in linear, angular or cluster formations^{11–13}. PAHs are primarily produced by the

incomplete combustion or heat-induced decomposition of matter¹⁴⁻¹⁶ and natural combustion processes, such as forest fires, and volcanic eruptions. They have been classified as hazardous compounds of environmental and health concern because of the fact that a number of them have been found to exhibit carcinogenic, genotoxic and mutagenic properties¹⁷, and long-range transportation and deposition capabilities. Consequently, the United States Environmental Protection Agency (USEPA) has listed 16 PAHs as priority environmental pollutants in order to facilitate environmental monitoring. The International Agency for Research on Cancer (IARC) has classified the 16 US EPA PAHs as follows. Benzo[*a*]pyrene (BaP) is carcinogenic (group 1), dibenzo[*a,h*]anthracene (DahA) is probably carcinogenic (group 2A), whereas naphthalene (Nap), benzo[*a*]anthracene (BaA), chrysene (Chry), benzo[*b*]fluoranthene (BbF), benzo[*k*]fluoranthene (BkF) and indeno[1,2,3-*c,d*]pyrene (IndP) are classified as possible human carcinogens (group 2B) while others are not classified as carcinogenic to humans¹⁸.

Human exposure to PAHs occurs mainly through contact with air, water and soil, and the consumption of contaminated products¹⁹ in which PAHs may accumulate in the lipid components²⁰. Dietary sources are by far the most important route of human exposure to PAHs, apart from smoking and occupational exposures. There are a number of published studies on the concentrations of PAHs in honey but most of these are restricted to European countries^{9,21-25}. To the best of our knowledge, there is no published data on the PAH concentrations in honey from the different regions of Nigeria. The objective of this study was to determine the concentrations of PAHs in honey samples from different regions of Nigeria with a view to providing information on the compositional patterns, regional profiles, sources and risks associated with the consumption of these honey samples. Such information is useful for environmental and food quality management.

Results and Discussion

The concentrations of the $\Sigma 16$ PAHs from different regions of Nigeria are presented in Tables 1 and 2. The concentrations of $\Sigma 16$ PAHs in the analyzed honey samples ranged from 169-522 $\mu\text{g kg}^{-1}$, 97-1980 $\mu\text{g kg}^{-1}$, 180-641 $\mu\text{g kg}^{-1}$ and 122-357 $\mu\text{g kg}^{-1}$ for South-East, South-West, Niger Delta, and North Central regions respectively. The differences observed in the $\Sigma 16$ PAH concentrations in these samples were significant ($p < 0.05$). There were also significant differences ($p < 0.05$) in the regional mean concentrations of $\Sigma 16$ PAHs. However, no significant difference was observed in

the mean concentrations of $\Sigma 16$ PAHs observed in the South-East and Niger Delta regions. The concentration pattern of the $\Sigma 16$ PAHs in the honey samples followed the order: South-West > South-East > Niger Delta > North Central. The regional concentration patterns reflect the distribution pattern of industries in Nigeria. For instance, the honey samples from the south-western part of Nigeria, that is more industrialised, showed higher concentrations of the $\Sigma 16$ PAHs.

A wide concentration range for PAHs has been reported in honey and honey products in the literature. For example, Dobrinas *et al.*⁹ reported PAH concentrations in the range of 56 to 2410 $\mu\text{g kg}^{-1}$ in honey from Romania. In Italy, Moret *et al.*²³ reported total PAH concentrations in the range of 38 to 41300 $\mu\text{g kg}^{-1}$ in raw propolis and 0.9 to 1790 $\mu\text{g kg}^{-1}$ in propolis extracts. Ciemark *et al.*²⁶ reported $\Sigma 23$ PAH concentrations up to 305 $\mu\text{g kg}^{-1}$ in blossom honey. In Spain, the concentrations of 15 PAHs in 30 samples of honey collected from different regions were found to be below the limit of detection²⁵. The PAH content in the honey samples in our study corresponds with that of Dobrinas *et al.*⁹ from Romania and Moret *et al.*²³ from Italy but was lower than values reported for honey in Italy²¹, France²², Czech Republic²⁴ and Spain²⁵. The contamination of these honey samples by PAHs could be due to bush burning and industrial and automobile emissions. In Nigeria, a bush fallowing farming method is commonly practiced, which involves clearing and burning of the cleared biomass at each farming season. In Nigeria, honeys are harvested mainly by the use of smoke which can also be a source of PAHs in the honey samples. Post-harvest contamination from improper packaging and processing also cannot be ruled out. Most frequently honeys are displayed for sale in open plastic containers along the major highways.

In this study, the naphthalene concentrations in the samples from the different regions were below the limit of quantification (LOQ) in all samples except for SW19 (313 $\mu\text{g kg}^{-1}$). In SW19 naphthalene constituted 28.4% of the $\Sigma 16$ PAHs. The non-detectability of naphthalene in these samples may be due to its high volatility.

The 3-ringed PAH concentrations ranged from less than the LOQ to 390 $\mu\text{g kg}^{-1}$, which constituted up to 65% of the $\Sigma 16$ PAHs in some of these samples. Acenaphthene (Ace) and fluorene (Flu) were the dominant 3-ringed PAHs in these samples in terms of frequency of occurrence. Ace and Flu were detected in 35% of the honey samples at concentrations in the range of 27 to 146 $\mu\text{g kg}^{-1}$ and 26 to 264 $\mu\text{g kg}^{-1}$ respectively. The concentrations of Flu were higher than the other 3-ringed PAHs in these honey samples. Phenanthrene (Phen) and anthracene (Ant) were detected in 28% and 22% of these honey samples at concentrations of 12 to 136

Table 1. PAH concentrations ($\mu\text{g kg}^{-1}$) from different regions in Nigeria.

	Nap	Acy	Ace	Flu	Phen	Ant	Flt	Pyr	BaA	Chry	BbF	BkF	BaP	IndP	DahA	BghiP	Total	2R	3R	4R	5R	6R	PAH2	PAH4	PAH8
SE1	ND	ND	33.2	33.6	41.6	ND	ND	ND	24.7	ND	29.6	191	ND	ND	27.3	ND	381	ND	108	24.7	248	ND	ND	54.3	272
SE2	ND	ND	45.8	28.6	ND	ND	ND	ND	ND	95.7	141	113	27.3	ND	ND	ND	451	ND	74.4	95.7	281	ND	123	264	377
SE3	ND	ND	ND	ND	ND	ND	ND	153.5	ND	35.4	27.4	ND	ND	ND	ND	27.7	244	ND	ND	189	27.4	27.7	35.4	62.8	90.5
SE4	ND	ND	145.6	25.7	ND	ND	ND	ND	27.5	44.7	60.6	110	26.2	ND	ND	ND	440	ND	171	72.2	197	ND	70.9	159	269
SE5	ND	ND	26.7	30.4	ND	ND	ND	ND	37.3	ND	ND	74.3	ND	ND	ND	ND	169	ND	57.1	37.3	74.3	ND	ND	37.3	112
SE6	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	86.6	147	ND	ND	59.3	ND	293	ND	ND	ND	293	ND	ND	86.6	293
SE7	ND	105	54.8	28.4	ND	ND	ND	ND	177	4.5	93.1	34.8	25.3	ND	ND	ND	522	ND	188	181	153	ND	29.8	299	334
SE8	ND	124	ND	ND	ND	ND	ND	ND	40.8	91.6	80.8	114	ND	ND	ND	ND	451	ND	124	132	195	ND	91.6	213	327
SW9	31.3	35.9	32.2	ND	67.2	ND	ND	61.7	77.5	ND	117	131	85.1	ND	87.1	92.9	1100	313	135	139	420	92.9	85.1	279	591
SW10	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	52	ND	ND	45.1	ND	97.2	ND	ND	ND	97.1	ND	ND	ND	97.1
SW11	ND	ND	ND	43	ND	ND	ND	ND	25.9	25.7	ND	46.2	ND	ND	39.3	ND	180	ND	43	51.6	85.5	ND	25.7	51.6	137
SW12	ND	60.5	32.1	ND	ND	ND	ND	ND	98.4	ND	ND	30.5	ND	ND	25.5	ND	247	ND	92.6	98.4	56	ND	ND	98.4	154
SW13	ND	ND	ND	263.6	62.9	63.9	44.4	97.1	371	198	141	63.3	59.8	67	430	122.3	1980	ND	390	710	694	189	257	769	1450
SW14	ND	ND	ND	ND	ND	59.7	ND	210	ND	104	ND	41.9	ND	ND	ND	ND	415	ND	59.7	314	41.9	ND	104	104	149
SW15	ND	ND	ND	38.3	ND	60.8	ND	57.8	ND	31.3	ND	70.9	31.8	ND	28.6	27.4	347	ND	99.1	89.1	131	27.4	63.1	63.1	190
SW16	ND	ND	ND	81.5	136	ND	ND	416	44.3	28.5	57.2	42.4	ND	ND	ND	ND	868	ND	218	551	99.6	ND	28.5	130	172
SW17	ND	40	26.1	65.3	ND	ND	ND	ND	24.4	ND	75.2	197	ND	ND	63.5	ND	360	ND	131	ND	81.4	ND	ND	ND	81.4
SW18	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	80.5	240	154	29	26	ND	530	ND	ND	80.5	424	26	110	350	530
SS20	ND	ND	ND	ND	ND	ND	ND	142	ND	95.8	74	80.3	ND	28	29.2	449	ND	ND	238	182	29.2	176	176	307	307
SS21	ND	62.4	48.1	ND	ND	ND	28.1	ND	111	73.1	149	92.7	21.3	ND	ND	56	641	ND	111	212	263	56	94.4	354	502
SS22	ND	ND	45.8	28.6	ND	ND	ND	ND	95.7	34.9	60.5	27.3	ND	ND	ND	ND	293	ND	74.4	95.7	123	ND	123	158	218
SS23	ND	ND	ND	ND	55.8	53.8	82.5	29.6	26.7	17.9	25.1	ND	ND	ND	ND	ND	291	ND	110	157	25.1	ND	17.9	69.7	69.7
SS24	ND	ND	ND	ND	ND	59.7	ND	104	ND	157	ND	52.4	ND	ND	ND	ND	378	ND	59.7	261	52.4	ND	157	157	209
SS25	ND	ND	ND	31.5	ND	67.1	ND	47.2	ND	31.3	ND	7.1	31.8	ND	2.9	0.3	230	ND	98.6	47.2	157	36.4	6.0	61.7	193
SS26	ND	ND	ND	38.3	ND	60.8	ND	57.7	ND	31.3	ND	198	46	ND	ND	ND	245	ND	99.1	89	41.8	0.3	63.1	63.1	73.4
SS27	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	198	46	ND	ND	ND	ND	245	ND	ND	ND	244	ND	ND	198	244
SS28	ND	ND	ND	ND	ND	ND	ND	ND	28.1	ND	7.5	90	ND	ND	54.6	ND	180	ND	28.1	152	152	ND	ND	35.6	180
SS29	ND	ND	37.8	30.4	ND	ND	ND	ND	37.3	ND	127	ND	26.8	ND	ND	ND	233	ND	68.2	37.3	127	ND	ND	37.3	165
SS30	ND	ND	ND	ND	53.8	ND	106	ND	62.7	38.6	23.9	59.4	26.8	ND	ND	46.1	417	ND	53.8	207	110	46.1	65.4	152	258
NC31	ND	ND	62	ND	115.8	ND	42.1	28.4	ND	ND	ND	24.3	ND	ND	ND	ND	273	ND	178	70.5	24.3	ND	ND	ND	24.3
NC32	ND	ND	ND	ND	24.2	24.1	35	81.1	ND	ND	ND	ND	28.5	ND	ND	37.1	230	ND	48.3	116	28.5	37.1	28.5	28.5	65.6
NC33	ND	ND	ND	ND	ND	ND	ND	141.9	ND	42.9	ND	74	27.4	ND	28	29.2	343	ND	ND	185	129	29.2	70.3	70.3	202
NC34	ND	ND	ND	ND	ND	38.7	64.7	30.2	ND	ND	9.2	90.6	0.5	24.5	39.8	58.9	357	ND	38.7	94.9	140	83.4	0.5	9.7	224
NC35	ND	ND	ND	ND	62.8	ND	33.8	ND	ND	ND	ND	25.3	ND	ND	ND	ND	122	ND	62.8	33.8	25.3	ND	ND	ND	25.3
NC36	ND	ND	62	ND	11.6	ND	42.1	28.4	ND	ND	ND	24.3	ND	ND	ND	ND	168	ND	73.6	70.5	24.3	ND	ND	ND	24.3
NC37	ND	63.5	ND	ND	ND	ND	32.6	ND	ND	ND	26.4	30.3	ND	ND	ND	35.3	188	ND	63.5	32.6	56.7	35.3	ND	26.4	92
NC38	ND	ND	ND	ND	ND	ND	ND	ND	28.2	29.1	36.2	10.9	ND	50.2	ND	155	ND	ND	28.2	126	126	39.1	68.2	155	155
NC39	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	25.5	38.1	ND	ND	38.3	33.6	136	ND	ND	ND	102	33.6	ND	25.5	134
NC40	ND	ND	26.8	ND	49.2	ND	ND	27.8	ND	27.8	ND	41.3	25.5	ND	ND	193	ND	76	55.6	66.8	ND	53.3	53.3	94.6	

Naphthalene (Nap), Acenaphthylene (Acy), Acenaphthene (Ace), Fluorene (Flu), Phenanthrene (Phen), Anthracene (Ant), Fluoranthene (Flt), Pyrene (Pyr), Benzo[a]anthracene (BaA), Chrysene (Chry), Benzo[b]fluoranthene (BbF), Benzo[k]fluoranthene (BkF), Benzo[e]pyrene (BeP), Indeno[1,2,3-c,d]pyrene (IndP), Dibenzol[a,h]anthracene (DahA) and Benzo[ghi]perylene (BghiP), Rings (R), South-East (SE), South-West (SW), Niger Delta (SD), North Central (NC), Below limit of quantification (LOQ) (ND).

Table 2. Mean \pm SD (Median) concentrations ($\mu\text{g kg}^{-1}$) of PAHs in honey samples.

PAH Compound	South-East	South-West	Niger Delta	North Central
Naphthalene	0.0 \pm 0.0 (0.0) ND	31.3 \pm 98.9 (0.0) (ND-312.7)	0.0 \pm 0.0 (0.0) ND	0.0 \pm 0.0 (0.0) ND
Acenaphthylene	28.5 \pm 53.1 (0.0) (ND-124)	13.6 \pm 22.8 (0.0) (ND-60.5)	5.2 \pm 18.0 (0.0) (ND-62.4)	6.4 \pm 20.1 (0.0) (ND-63.5)
Acenaphthene	38.3 \pm 48.4 (30) (ND-145.6)	9.0 \pm 14.6 (0.0) (ND-32.2)	11.0 \pm 20.0 (0.0) (ND-48.1)	15.2 \pm 6.1 (0.0) ND-62.0
Fluorene	18.3 \pm 15.3 (27.1) (ND-33.6)	49.2 \pm 47.0 (0.0) (ND-264)	10.7 \pm 16.0 (0.0) (ND-38.3)	0.0 \pm 0.0 (0.0) ND
Phenanthrene	5.2 \pm 14.7 (0.0) (ND-41.6)	26.6 \pm 47.0 (0.0) (ND-136.0)	9.1 \pm 21.3 (0.0) (ND-55.8)	26.4 \pm 38.8 (5.8) (ND-62.8)
Anthracene	0.0 \pm 0.0 (0.0) ND	18.4 \pm 29.7 (0.0) (ND-63.9)	20.1 \pm 29.9 (0.0) (ND-67.1)	6.3 \pm 13.7 (0.0) (ND-38.7)
Fluoranthene	0.0 \pm 0.0 (0.0) ND	46.2 \pm 131 (0.0) (ND-417.5)	18.1 \pm 36.9 (0.0) (ND-106)	25.0 \pm 23.3 (33.2) (ND-64.7)
Pyrene	19.2 \pm 54.3 (0.0) (ND-153.5)	48.7 \pm 67.1 (28.9) (ND-210)	31.7 \pm 48.0 (0.0) (ND-141.9)	33.8 \pm 45.5 (28.1) (ND-142)
Benzo[<i>a</i>]anthracene	38.4 \pm 58.3 (26.1) (ND-40.8)	64.1 \pm 113 (25.2) (ND-371)	22.1 \pm 34.7 (0.0) (ND-111)	0.0 \pm 0.0 (0.0) ND
Chrysene	34.0 \pm 40.7 (20.0) (ND-95.7)	38.7 \pm 64.4 (12.9) (ND-198)	49.2 \pm 51.0 (35.0) (ND-157)	9.9 \pm 16.4 (0.0) (ND-42.9)
Benzo[<i>b</i>]fluoranthene	64.9 \pm 45.0 (70.7) (ND-140.8)	39.0 \pm 54.9 (0.0) (ND-141)	61.1 \pm 85.1 (24.5) (ND-240.2)	9.0 \pm 12.8 (0.0) (ND-29.1)
Benzo[<i>k</i>]fluoranthene	98.0 \pm 60.7 (112) (ND-191)	75.7 \pm 51.4 (57.7) (30.5-197)	71.6 \pm 44.6 (67.3) (ND-154)	38.4 \pm 26.1 (33.3) (ND-90.6)
Benzo[<i>a</i>]pyrene	9.9 \pm 13.6 (0.0) (ND-27.3)	17.7 \pm 31.1 (0.0) (ND-85.1)	18.5 \pm 23.6 (13.7) (ND-80.3)	9.3 \pm 12.8 (0.3) (ND-28.5)
Indeno[1,2,3- <i>c,d</i>]pyrene	0.0 \pm 0.0 (0.0) ND	6.7 \pm 21 (0.0) (ND-67.0)	2.2 \pm 7.5 (0.0) (ND-26.0)	2.5 \pm 7.7 (0.0) (ND-24.5)
Dibenz[<i>a,h</i>]anthracene	10.8 \pm 21.8 (0.0) (ND-59.3)	72.0 \pm 129 (34.0) (ND-430)	7.1 \pm 17.0 (0.0) (ND-54.6)	15.6 \pm 20.8 (0.0) (ND-50.2)
Benzo[<i>g,h,i</i>]perylene	3.5 \pm 9.8 (0.0) (ND-27.7)	24.3 \pm 45.3 (0.0) (ND-122.3)	14.0 \pm 21.5 (0.0) ND-56.0	19.4 \pm 21.9 (14.6) (ND-58.9)
Σ 16 PAHs	368.9 \pm 121.6 (411) (169-522)	581.1 \pm 586.2 (353.5) (180-1980)	352 \pm 137 (316) (180-641)	216.4 \pm 83.1 (191) (122-357)
2 Rings	0.0 \pm 0.0 (0.0) ND	31.3 \pm 98.9 (0.0) (ND-313)	\pm 0.0 (0.0) ND	0.0 \pm 0.0 (0.0) ND
3 Rings	90.3 \pm 70.9 (91.4) (ND-171.3)	116.9 \pm 117 (95.9) (ND-390)	56.2 \pm 45.3 (64.0) (ND-111)	54.1 \pm 53.1 (55.6) (ND-178)
4 Rings	91.5 \pm 71.0 (84.0) (ND-188.9)	197.7 \pm 248 (93.8) (ND-710)	121 \pm 90.0 (92.4) (ND-261)	68.7 \pm 53.2 (63.1) (ND-185)
5 Rings	184 \pm 95.0 (196) (27.4-293.2)	204 \pm 213 (98.4) (41.9-694.2)	158 \pm 111 (140) (25.1-424)	72.4 \pm 47.9 (61.8) (24.3-140)
6 Rings	3.5 \pm 9.8 (0.0) (ND-27.7)	31.0 \pm 62.9 (0.0) (ND-189.3)	16.2 \pm 21.3 (0.2) (ND-56.0)	21.9 \pm 27.4 (14.6) (ND-83.4)
PAH2	43.8 \pm 46.8 (32.6) (ND-123)	56.4 \pm 80.2 (27.1) (ND-257.3)	67.7 \pm 64.2 (64.3) (ND-176)	19.2 \pm 26.8 (0.3) (ND-70.3)
PAH4	147.1 \pm 102.0 (123) (37.3-299)	160 \pm 228 (99.0) (ND-769)	151 \pm 110 (155) (35.6-354)	28.2 \pm 27.4 (26.0) (ND-70.3)
PAH8	259 \pm 104 (283) (90.5-377)	338 \pm 420 (163) (81.4-1450)	246 \pm 144 (214) (69.0-530)	104 \pm 73.0 (93.3) (24.3-224)

$\mu\text{g kg}^{-1}$ and 24 to 67 $\mu\text{g kg}^{-1}$ respectively. Ant was below the LOQ in the honey samples from the South-East region of Nigeria. Acenaphthylene (Acy) was detected in 15% of the samples at concentrations in the range of 36 to 124 $\mu\text{g kg}^{-1}$.

The 4-ringed PAHs were found in 88% of the sam-

ples at concentrations varying from 24 to 710 $\mu\text{g kg}^{-1}$. Chry was the dominant 4-ringed PAH in the honey samples in terms of concentration and frequency of occurrence. In these samples, Chry was detected at concentrations in the range of 4.5 to 198 $\mu\text{g kg}^{-1}$ which constituted 0.9 to 42.1% of the Σ 16 PAHs. Pyrene (Pyr)

and BaA were detected in 43% and 35% of the samples at concentrations in the range of 30 to 210 $\mu\text{g kg}^{-1}$ and 25 to 371 $\mu\text{g kg}^{-1}$ respectively. BaA and fluoranthene (Flt) were not detected in the honey samples from the North Central and South-East parts of Nigeria respectively. However, Flt was detected in 28% of the total samples examined at concentrations between 28 and 418 $\mu\text{g kg}^{-1}$ which constituted 2.2 to 48% of the $\Sigma 16$ PAHs in these samples.

The 5-ringed PAH concentrations ranged from 25.1 to 694 $\mu\text{g kg}^{-1}$ which constituted 8.9 to 100% of the $\Sigma 16$ PAHs in these samples. The 5-ring PAHs were the dominant PAHs in terms of concentrations and frequency of occurrence compared with 2-, 3- and 6-ringed PAHs. The dominant 5-ringed PAH was BkF which had 95% occurrence in the honey samples. BkF was detected at concentrations that varied from 7.1 to 197 $\mu\text{g kg}^{-1}$. BbF was detected in 58% of the honey samples at concentrations of 7.5 to 240 $\mu\text{g kg}^{-1}$ which constituted up to 45.3% of the $\Sigma 16$ PAHs in some of these samples. BaP was detected in 45% of the honey samples at concentrations ranging from 0.5 to 85 $\mu\text{g kg}^{-1}$. There is no legislation regulating the concentration of BaP in honey and honey products. Consequently, these results were compared with the limit for BaP specified for infant food. The concentrations of BaP in 45% of the samples examined were above the 1 $\mu\text{g kg}^{-1}$ limit specified for BaP in infant food²⁷. The concentrations of BaP in this study are comparable to those found in honey from Romania (0.5 to 141 $\mu\text{g kg}^{-1}$)²³ and in propolis and propolis-based dietary supplements in Italy (0.8 to 42 $\mu\text{g kg}^{-1}$)¹⁹ but were higher than BaP concentrations reported in honey from France²², Czech Republic²⁴, Spain²⁵ and Poland²⁶.

The 6-ringed PAHs were detected in 38% of the honey samples in the range of 0.3 to 189 $\mu\text{g kg}^{-1}$, which constituted up to 25% of the $\Sigma 16$ PAHs in some of the samples. Benzo[ghi]perylene (BghiP) was the most abundant 6-ringed PAH compound in the honey samples. Indeno[1,2,3-c,d]pyrene (IndP) was detected in three samples at concentrations of 25 to 67 $\mu\text{g kg}^{-1}$. IndP was not detected in any of the samples from South-East Nigeria.

Although, BaP is considered as a suitable indicator for occurrence and effect of PAHs in food, this relation is not fully convincing. Therefore, the European Food Safety Authority (EFSA) has suggested the use of PAH2 (Chry + BaP), PAH4 (PAH2 + BaA + BbF) and PAH8 (PAH4 + BkF + DahA + IndP + BghiP) as indicators for occurrence and effects of PAHs in food²⁸. The indicators: PAH2, PAH4 and PAH8 were detected in 60, 88 and 100% of the samples at concentrations in the range of 0.5 to 257 $\mu\text{g kg}^{-1}$, 9.7 to 769 $\mu\text{g kg}^{-1}$, and 24 to 1450 $\mu\text{g kg}^{-1}$ respectively. The average concen-

trations of PAH8 in the four regions were 259 $\mu\text{g kg}^{-1}$ (South-East), 338 $\mu\text{g kg}^{-1}$ (South-West), 246 $\mu\text{g kg}^{-1}$ (Niger Delta) and 104.1 $\mu\text{g kg}^{-1}$ (North Central). The results indicated that honey samples from south-western Nigeria contained higher concentrations of PAH8 than the other regions.

Dietary Intake and Risk Assessment

The estimated dietary intake of PAHs based on BaP and the EFSA suggested indicators for occurrence and effect of PAHs in food are displayed in Table 3. The maximum estimated daily intake values for BaP, PAH2, PAH4 and PAH8 were 1.9 $\text{ng kg}^{-1} \text{bw day}^{-1}$, 5.9 $\text{ng kg}^{-1} \text{bw day}^{-1}$, 18 $\text{ng kg}^{-1} \text{bw day}^{-1}$ and 33 $\text{ng kg}^{-1} \text{bw day}^{-1}$ respectively. The results of the present study revealed that the intake values of these indicators of PAHs were higher in the South-West region than the Niger Delta, South-East and North Central regions of Nigeria. The dietary intakes of PAHs in this study are lower than intake values reported from consumed canned fish¹³, smoked/grilled fish²⁹, fresh and smoked fish^{30,31}, milk³², biscuits³³, chocolates and candies³⁴ and ready to eat foods in Nigeria³⁵ but were comparable to intake values obtained from consumption of tea in Nigeria³⁶. Moret *et al.*²³ reported intake values for BaP and PAH8 of 0.0 to 1.3 $\text{ng kg}^{-1} \text{bw day}^{-1}$ and 0.01 to 47 $\text{ng kg}^{-1} \text{bw day}^{-1}$ respectively through consumption of raw propolis and propolis extracts used as dietary supplements. The estimated margin of exposure (MOE) based on BaP and the EFSA suggested indicators for effects and occurrence of PAHs in foods are displayed in Table 3. The BaP-MOE, PAH2-MOE, PAH4-MOE and PAH8-MOE values were greater than 10,000 which indicate no health concern associated with the consumption of these honeys at the current ingestion rate.

The computed BaP_{TEQ} and BaP_{MEQ} concentrations from ingestion of these honey samples are displayed in Table 4. BaP_{TEQ} is directly associated with carcinogenicity, whereas BaP_{MEQ} (mutagenic activity) may not be directly associated with cancer^{37,38} and may have to do with other non-cancerous adverse health effects such as pulmonary disease, birth defects, impotency, low IQ, etc.³⁹⁻⁴¹. The BaP_{TEQ} for the honey samples ranged from 0.2 to 549 $\mu\text{g kg}^{-1}$. The main contributors to $\Sigma \text{BaP}_{\text{TEQ}}$ values of these honey samples were BaA, BaP, BbF, and DahA, while the contribution of IndP, Chry and BkF to the $\Sigma \text{BaP}_{\text{TEQ}}$ was minimal. BaP_{MEQ} values obtained in this study ranged from 2.67 to 281 $\mu\text{g kg}^{-1}$ with significant contributions from BbF, BkF and BaP. The honey samples from the South-West region had higher mean BaP_{TEQ} and BaP_{MEQ} values than the other regions.

The estimated incremental life cancer risk associated with the consumption of these honey samples ranged

Table 3. Estimated daily intakes ($\text{ng kg}^{-1} \text{bw day}^{-1}$) and margin of exposure for honey samples.

	Daily intake				Margin of exposure			
	BaP	PAH2	PAH4	PAH8	BaP	PAH2	PAH4	PAH8
SE1	0.0	0.0	1.2	6.2	–	–	274000	78800
SE2	0.6	2.8	6.0	8.6	112000	60500	56400	56900
SE3	0.0	0.8	1.4	2.1	0	210000	237000	237000
SE4	0.6	1.6	3.6	6.1	117000	105000	93700	79800
SE5	0.0	0.0	0.9	2.5	–	–	399000	192000
SE6	0.0	0.0	2.0	6.7	–	–	172000	73200
SE7	0.6	0.7	6.8	7.6	121000	245000	49700	64200
SE8	0.0	2.1	4.9	7.5	–	81300	69800	65600
SW9	1.9	1.9	6.4	13.5	36000	87500	53300	36300
SW10	0.0	0.0	0.0	2.2	–	–	–	221000
SW11	0.0	0.6	1.2	3.1	–	290000	289000	157000
SW12	0.0	0.0	2.2	3.5	–	–	151000	139000
SW13	1.4	5.9	18	33	51300	28900	19400	14800
SW14	0.0	2.4	2.4	3.3	–	71700	143000	147000
SW15	0.7	1.4	1.4	4.3	96400	119000	236000	113000
SW16	0.0	0.7	3.0	3.9	–	261000	115000	125000
SW17	0.0	0.0	0.0	1.9	–	–	–	264000
SW18	0.0	0.0	2.3	8.2	–	–	150000	59600
SS19	0.7	2.5	8.0	12	106000	68000	42600	40500
SS20	1.8	4.0	4.0	7.0	38200	42300	84600	69800
SS21	0.5	2.2	8.1	12	144000	78900	42100	42700
SS22	0.6	2.8	3.6	5.0	112000	60500	94300	98300
SS23	0.0	0.4	1.6	1.6	–	416000	214000	308000
SS24	0.0	3.6	3.6	4.8	–	47500	94900	103000
SS25	0.1	0.1	1.4	4.4	51100	1240000	241000	111000
SS26	0.7	1.4	1.4	1.7	96400	118000	236000	292000
SS27	0.0	0.0	4.5	5.6	–	–	75300	88000
SS28	0.0	0.0	0.8	4.1	–	–	418000	119000
SS29	0.0	0.0	0.9	3.8	–	–	399000	130000
SS30	0.6	1.5	3.5	5.9	114000	114000	98000	83300
NC31	0.0	0.0	0.0	0.6	–	–	–	883000
NC32	0.7	0.7	0.7	1.5	108000	261000	523000	327000
NC33	0.6	1.6	1.6	4.6	112000	106000	212000	107000
NC34	0.0	0.0	0.2	5.1	6130000	14900000	1540000	96000
NC35	0.0	0.0	0.0	0.6	–	–	–	848000
NC36	0.0	0.0	0.0	0.6	–	–	–	883000
NC37	0.0	0.0	0.6	2.1	–	–	564000	233000
NC38	0.2	0.9	1.6	3.5	281000	19000	218000	139000
NC39	0.0	0.0	0.6	3.1	–	–	584000	158000
NC40	0.6	1.2	1.2	2.2	120000	140000	279000	227000

between 8.0×10^{-10} to 1.8×10^{-6} (Table 5). The results indicated that these samples gave incremental life cancer risk (ILCR) values lower than the acceptable risk level of one in a million chance of additional cancer over a 52 year lifetime ($1/10^6$). This indicates that there is no additional risk arising from the consumption of these honey samples. The ILCR values are in strong agreement with the MOE values.

Source Analysis from Isomeric Ratios

Different isomeric ratios of PAHs have been used to identify the proportion of pyrogenic and petrogenic PAHs in environmental matrices⁴². These diagnostic

ratios include $\text{Ant}/(\text{Ant} + \text{Phen})$, $\text{Flt}/(\text{Flt} + \text{Pyr})$, $\text{BaA}/(\text{BaA} + \text{Chry})$, $\text{IndP}/(\text{IndP} + \text{BghiP})$, Phen/Ant , Flt/Pyr and LMW/HMW . The values obtained for the various diagnostic ratios in this work are shown in Table 6. In this study, the ratio of low molecular weight (LMW) to high molecular weight (HMW) PAHs ranged from 0.00 to 1.88. $\text{LMW}/\text{HMW} > 1$ indicates petrogenic while $\text{LMW}/\text{HMW} < 1$ indicates pyrogenic sources. Three samples showed LMW/HMW ratios greater than 1 which indicates that the sources of PAHs in these samples were pyrogenic in nature. A ratio of $\text{Ant}/(\text{Ant} + \text{Phen}) < 0.1$ and $\text{Flt}/(\text{Flt} + \text{Pyr}) < 0.4$ is indicative of petroleum sources, while $\text{Ant}/(\text{Flt} + \text{Pyr}) > 0.1$ implies

Table 4. BaP_{TEQ} and BaP_{MEQ} ($\mu\text{g kg}^{-1}$) in honey samples.

	BaA	Chry	BbF	BkF	BaP	IndP	DahA	BaP _{TEQ}	BaA	Chry	BbF	BkF	BaP	IndP	DahA	BaP _{MEQ}
SE1	2.5	ND	3.0	1.9	ND	ND	27	34.6	2.0	ND	7.4	21	ND	ND	7.92	38.3
SE2	ND	0.1	14.1	1.1	27.3	ND	ND	42.6	ND	1.6	35.2	12.5	27.3	ND	ND	76.6
SE3	ND	0.04	2.7	ND	ND	ND	ND	2.78	ND	0.6	6.9	ND	ND	ND	ND	7.5
SE4	2.8	0.04	6.1	1.1	26.2	ND	ND	36.2	2.3	0.8	15.2	12.1	26.2	ND	ND	56.4
SE5	3.7	ND	ND	0.7	ND	ND	ND	4.47	3.1	ND	ND	8.2	ND	ND	ND	11.2
SE6	ND	ND	8.7	1.5	ND	ND	59	69.4	ND	ND	21.7	16.2	ND	ND	17.2	55.1
SE7	17.7	0.00	9.3	0.6	25.3	ND	ND	52.6	14.5	0.08	23.3	3.8	25.3	ND	ND	67
SE8	4.1	0.1	8.1	1.1	ND	ND	ND	13.4	3.4	1.6	20.2	12.5	ND	ND	ND	37.6
SW9	7.8	ND	11.7	1.3	85.1	ND	87	193	6.4	ND	29.2	14.5	85.1	ND	25.3	160
SW10	ND	ND	ND	0.5	ND	ND	45	45.6	ND	ND	ND	5.7	ND	ND	13.1	18.8
SW11	2.6	0.03	ND	0.5	ND	ND	39	42.4	2.1	0.4	ND	5.1	ND	ND	11.4	19.0
SW12	9.8	ND	ND	0.3	ND	ND	25	35.7	8.1	ND	ND	3.4	ND	ND	7.4	18.8
SW13	37.1	0.2	14.1	0.6	59.8	6.7	430	549	30.4	3.4	35.2	7.0	59.8	20.8	125	281
SW14	ND	0.1	ND	0.4	ND	ND	ND	0.52	ND	1.8	ND	4.6	ND	ND	ND	6.4
SW15	ND	0.03	ND	0.7	31.8	ND	29	61.1	ND	0.5	ND	7.8	31.8	ND	8.3	48.4
SW16	4.4	0.03	5.7	0.4	ND	ND	ND	10.6	3.6	0.5	14.3	4.7	ND	ND	ND	23.1
SW17	ND	ND	ND	0.8	ND	ND	ND	0.81	ND	ND	ND	9.0	ND	ND	ND	9.0
SW18	2.4	ND	7.5	2.0	ND	ND	64	75.4	2.0	ND	18.8	21.7	ND	ND	18.4	60.9
SS19	ND	0.1	24.0	1.5	29	2.6	ND	57.2	ND	1.4	60.1	17	29	8.1	ND	116
SS20	ND	0.1	ND	0.7	80.3	ND	28	109	ND	1.6	ND	8.1	80.3	ND	8.1	98.2
SS21	11.1	0.1	14.9	0.9	21.3	ND	ND	48.2	9.1	1.2	37.1	10.2	21.3	ND	ND	79
SS22	ND	0.1	3.5	0.6	27.3	ND	ND	31.5	ND	1.6	8.7	6.7	27.3	ND	ND	44.3
SS23	2.7	0.02	2.5	ND	ND	ND	ND	5.20	2.2	0.3	6.3	ND	ND	ND	ND	8.8
SS24	ND	0.2	ND	0.5	ND	ND	ND	0.68	ND	2.7	ND	5.8	ND	ND	ND	8.4
SS25	ND	ND	5.6	1.0	6	ND	ND	12.5	ND	ND	13.9	10.5	6	ND	ND	30.4
SS26	ND	0.03	ND	0.1	31.8	ND	2.9	34.8	ND	0.5	ND	0.8	31.8	ND	0.8	34
SS27	ND	ND	19.9	0.5	ND	ND	ND	20.2	ND	ND	49.5	5.1	ND	ND	ND	54.5
SS28	2.8	ND	0.8	0.9	ND	ND	55	59.1	2.3	ND	1.9	9.9	ND	ND	15.8	29.9
SS29	3.7	ND	ND	1.3	ND	ND	ND	5.0	3.1	ND	ND	14	ND	ND	ND	17.1
SS30	6.3	0.04	2.4	0.6	26.8	ND	ND	36.1	5.1	0.7	6.0	6.5	26.8	ND	ND	45.1
NC31	ND	ND	ND	0.2	ND	ND	ND	0.24	ND	ND	ND	2.7	ND	ND	ND	2.7
NC32	ND	ND	ND	ND	28.5	ND	ND	28.5	ND	ND	ND	ND	28.5	ND	ND	28.5
NC33	ND	0.04	ND	0.7	27.4	ND	28	56.2	ND	0.7	ND	8.1	27.4	ND	8.1	44.4
NC34	ND	ND	0.9	0.9	0.5	2.45	40	44.6	ND	ND	2.3	10	0.5	7.6	11.6	31.9
NC35	ND	ND	ND	0.3	ND	ND	ND	0.3	ND	ND	ND	2.8	ND	ND	ND	2.8
NC36	ND	ND	ND	0.2	ND	ND	ND	0.2	ND	ND	ND	2.7	ND	ND	ND	2.7
NC37	ND	ND	2.6	0.3	ND	ND	ND	2.9	ND	ND	6.6	3.3	ND	ND	ND	9.9
NC38	ND	0.03	2.9	0.4	10.9	ND	50	64.4	ND	0.5	7.3	4.0	10.9	ND	14.6	37.2
NC39	ND	ND	2.6	0.3	ND	ND	38.3	41.2	ND	ND	6.4	4.2	ND	ND	11.1	21.7
NC40	ND	0.03	ND	0.4	25.5	ND	ND	25.9	ND	0.5	ND	4.5	25.5	ND	ND	30.5

Benzo[a]anthracene (BaA), Chrysene (Chry), Benzo[b]fluoranthene (BbF), Benzo[k]fluoranthene (BkF), Benzo[a]pyrene (BaP), Indeno[1,2,3-cd]pyrene (IndP), Dibenz[a,h]anthracene (DahA) and Benzo[ghi]perylene (BghiP), South-East (SE), South-West (SW), Niger Delta (SS), North Central (NC), Below limit of quantification (ND).

biomass and coal combustion. A Flt/(Flt + Pyr) ratio between 0.4 and 0.5 implies liquid fossil fuel combustion and Flt/(Flt + Pyr) > 0.5 suggests biomass and coal combustion. Ratios of BaA/(BaA + Chry) < 0.2 and IndP/(IndP + BghiP) > 0.5 and BaA/(BaA + Chry) > 0.35 indicate that the source of PAHs is biomass and coal combustion⁴². In this study, the ratio of Flt/(Flt + Pyr) ranged from 0.30 to 1.00, Ant/(Ant + Phen) ranged from 0.37 to 1.00; BaA/(BaA + Chry) ranged from 0.31 to 1.00 and IndP/(IndP + BghiP) > 0.5 indicates that sources of PAHs in these honey samples were mainly due to biomass and fossil fuel combustion.

The ratio of $\Sigma \text{COMB}/\Sigma \text{PAHs}$ provides useful information on the degree to which the origins of PAHs are related to combustion of typical organics⁴³. ΣCOMB is the sum of Flt, Pyr, Chry, BkF, BaP, BghiP and IndP, while ΣPAH is the sum of the concentrations of the 16 PAHs measured in this study. The ratio of $\Sigma \text{COMB}/\Sigma \text{PAH}$ values in this study ranged from 0.19 to 0.94 indicating high fractions of combustion origin in these sites. In addition, the total index¹⁵ was also estimated as the sum of single indices (discussed earlier) normalized for the limit value (low temperature sources-high temperature sources) reported in the literature⁴².

Table 5. Excess cancer risk of PAHs in honey samples.

	BaA	Chry	BbF	BkF	BaP	IndP	DahA	Excess cancer risk
SE1	8.1×10^{-9}	0.0	9.7×10^{-9}	6.2×10^{-9}	0.0	0.0	8.9×10^{-8}	1.1×10^{-7}
SE2	0.0	3.1×10^{-10}	4.6×10^{-8}	3.7×10^{-9}	8.9×10^{-8}	0.0	0.0	1.4×10^{-7}
SE3	0.0	1.2×10^{-10}	2.0×10^{-9}	0.0	0.0	0.0	0.0	9.1×10^{-9}
SE4	9.0×10^{-9}	1.5×10^{-10}	2.0×10^{-8}	3.6×10^{-9}	8.6×10^{-8}	0.0	0.0	1.2×10^{-7}
SE5	1.2×10^{-8}	0.0	0.0	2.4×10^{-9}	0.0	0.0	0.0	1.5×10^{-8}
SE6	0.0	0.0	2.8×10^{-8}	4.8×10^{-9}	0.0	0.0	1.9×10^{-7}	2.3×10^{-7}
SE7	5.8×10^{-8}	1.5×10^{-11}	3.1×10^{-8}	1.1×10^{-9}	8.3×10^{-8}	0.0	0.0	1.7×10^{-7}
SE8	1.3×10^{-8}	3.0×10^{-10}	2.7×10^{-8}	3.7×10^{-9}	0.0	0.0	0.0	4.4×10^{-8}
SW9	2.5×10^{-8}	0.0	3.8×10^{-8}	4.3×10^{-9}	2.8×10^{-7}	0.0	2.9×10^{-7}	6.3×10^{-7}
SW10	0.0	0.0	0.0	1.7×10^{-9}	0.0	0.0	1.5×10^{-7}	1.5×10^{-7}
SW11	8.5×10^{-9}	8.4×10^{-11}	0.0	1.5×10^{-9}	0.0	0.0	1.3×10^{-7}	1.4×10^{-7}
SW12	3.2×10^{-8}	0.0	0.0	1.0×10^{-9}	0.0	0.0	8.4×10^{-8}	1.2×10^{-7}
SW13	1.2×10^{-7}	6.5×10^{-10}	4.6×10^{-8}	2.1×10^{-9}	2.0×10^{-7}	2.2×10^{-8}	1.4×10^{-6}	1.8×10^{-6}
SW14	0.0	3.4×10^{-10}	0.0	1.4×10^{-9}	0.0	0.0	0.0	1.7×10^{-9}
SW15	0.0	1.0×10^{-10}	0.0	2.3×10^{-9}	1.0×10^{-7}	0.0	9.4×10^{-8}	2.0×10^{-7}
SW16	1.5×10^{-8}	9.3×10^{-11}	1.9×10^{-8}	1.4×10^{-9}	0.0	0.0	0.0	3.5×10^{-8}
SW17	0.0	0.0	0.0	2.7×10^{-9}	0.0	0.0	0.0	2.7×10^{-9}
SW18	8.0×10^{-9}	0.0	2.5×10^{-8}	6.5×10^{-9}	0.0	0.0	2.1×10^{-7}	2.5×10^{-7}
SS19	0.0	2.6×10^{-10}	7.9×10^{-8}	5.1×10^{-9}	9.5×10^{-8}	8.5×10^{-9}	0.0	1.9×10^{-7}
SS20	0.0	3.1×10^{-10}	0.0	2.4×10^{-9}	2.6×10^{-7}	0.0	9.2×10^{-8}	3.6×10^{-7}
SS21	3.6×10^{-8}	2.4×10^{-10}	4.9×10^{-8}	3.0×10^{-9}	7.0×10^{-8}	0.0	0.0	1.6×10^{-7}
SS22	0.0	3.1×10^{-10}	1.1×10^{-8}	2.0×10^{-9}	8.9×10^{-8}	0.0	0.0	1.0×10^{-7}
SS23	8.8×10^{-9}	5.9×10^{-11}	8.2×10^{-9}	0.0	0.0	0.0	0.0	1.7×10^{-8}
SS24	0.0	5.14×10^{-10}	0.0	1.7×10^{-9}	0.0	0.0	0.0	2.2×10^{-9}
SS25	0.0	0.0	1.8×10^{-8}	3.1×10^{-9}	2.0×10^{-8}	0.0	0.0	4.1×10^{-8}
SS26	0.0	1.0×10^{-10}	0.0	2.3×10^{-10}	1.0×10^{-7}	0.0	9.5×10^{-9}	1.1×10^{-7}
SS27	0.0	0.0	6.5×10^{-8}	1.5×10^{-9}	0.0	0.0	0.0	6.6×10^{-8}
SS28	9.2×10^{-9}	0.0	2.5×10^{-9}	3.0×10^{-9}	0.0	0.0	1.8×10^{-7}	1.9×10^{-7}
SS29	1.2×10^{-8}	0.0	0.0	4.2×10^{-9}	0.0	0.0	0.0	1.6×10^{-8}
SS30	2.1×10^{-8}	1.3×10^{-10}	7.8×10^{-9}	2.0×10^{-9}	8.8×10^{-8}	0.0	0.0	1.2×10^{-7}
NC31	0.0	0.0	0.0	8.0×10^{-10}	0.0	0.0	0.0	8.0×10^{-10}
NC32	0.0	0.0	0.0	0.0	9.3×10^{-8}	0.0	0.0	9.3×10^{-8}
NC33	0.0	1.4×10^{-10}	0.0	2.4×10^{-9}	9.0×10^{-8}	0.0	9.2×10^{-8}	1.8×10^{-7}
NC34	0.0	0.0	3.1×10^{-9}	3.0×10^{-9}	1.6×10^{-9}	8.0×10^{-9}	1.3×10^{-7}	1.5×10^{-7}
NC35	0.0	0.0	0.0	8.3×10^{-10}	0.0	0.0	0.0	8.3×10^{-10}
NC36	0.0	0.0	0.0	8.0×10^{-10}	0.0	0.0	0.0	8.0×10^{-10}
NC37	0.0	0.0	8.7×10^{-9}	9.9×10^{-10}	0.0	0.0	0.0	9.6×10^{-9}
NC38	0.0	9.2×10^{-11}	9.5×10^{-9}	1.2×10^{-9}	3.6×10^{-8}	0.0	1.6×10^{-7}	2.1×10^{-7}
NC39	0.0	0.0	8.4×10^{-9}	1.3×10^{-9}	0.0	0.0	1.3×10^{-7}	1.4×10^{-7}
NC40	0.0	9.1×10^{-11}	0.0	1.4×10^{-9}	8.4×10^{-8}	0.0	0.0	8.5×10^{-8}

Benzo[a]anthracene (BaA), Chrysene (Chry), Benzo[b]fluoranthene (BbF), Benzo[k]fluoranthene (BkF), Benzo[a]pyrene (BaP), Indeno[1,2,3-cd]pyrene (IndP), Dibenz[a,h]anthracene (DahA); South-East (SE), South-West (SW), Niger Delta (SS), North Central (NC).

$$\text{Total index} = \frac{\text{Ant}/(\text{Ant} + \text{Phen})}{0.1} + \frac{\text{Flt}/(\text{Flt} + \text{Pyr})}{0.4} + \frac{\text{BaA}/(\text{BaA} + \text{Chry})}{0.2} + \frac{\text{IndP}/(\text{IndP} + \text{BghiP})}{0.5}$$

It should be noted that PAHs associated with high temperature processes (combustion) have a total index that is greater than 4, while PAHs originating from low temperature processes (petroleum products) have a total index that is less than 4. The total index values in our samples ranged from 1.49 to 10.7. Eight samples had total index values less than 4 (low temperature process-

es) while 20 samples had total index values greater than 4 which confirms that most of the PAHs in the honey samples originated predominantly from combustion processes.

Multivariate Analysis

The results of the principal component analysis of PAHs in honey from the four geographical zones of Nigeria are displayed in Table 7. In the Niger Delta (South-South), three components were extracted. Factor 1 accounts for 23.38% of the variance and is dominated by high loadings in Acy, BaA and BghiP, and weak loadings in Flt and Ace. Acy and BaA are mark-

Table 6. Diagnostic ratios of PAHs in honey samples.

	LMW/ HMW	Ant/ (Ant + Phe)	BaA/ (BaA + Chry)	Flt/ (Flt + Pyr)	IndP/ (IndP + BghiP)	BaP/ BghiP	CombPAH/ Sum PAH	Total index
SE1	0.40	0.00	1.00	0.00	0.00	0.00	0.57	5.00
SE2	0.20	0.00	0.00	0.00	0.00	0.00	0.52	0.00
SE3	0.00	0.00	0.00	0.00	0.00	0.00	0.89	0.00
SE4	0.64	0.00	0.38	0.00	0.00	0.00	0.47	1.90
SE5	0.51	0.00	1.00	0.00	0.00	0.00	0.66	5.00
SE6	0.00	0.00	0.00	0.00	0.00	0.00	0.50	0.00
SE7	0.56	0.00	0.98	0.00	0.00	0.00	0.46	4.88
SE8	0.38	0.00	0.31	0.00	0.00	0.00	0.55	1.54
SW9	0.69	0.00	1.00	0.00	0.00	0.92	0.41	5.00
SW10	0.00	0.00	0.00	0.00	0.00	0.00	0.53	0.00
SW11	0.31	0.00	0.50	0.00	0.00	0.00	0.54	2.51
SW12	0.60	0.00	1.00	0.00	0.00	0.00	0.52	5.00
SW13	0.25	0.59	0.65	0.31	0.35	0.49	0.52	10.7
SW14	0.17	1.00	0.00	0.00	0.00	0.00	0.86	10.0
SW15	0.40	1.00	0.00	0.00	0.00	1.16	0.63	10.0
SW16	0.33	0.00	0.61	0.87	0.00	0.00	0.68	5.23
SW17	1.61	0.00	0.00	0.00	0.00	0.00	0.38	0.00
SW18	0.00	0.00	1.00	0.00	0.00	0.00	0.61	5.00
SS19	0.00	0.00	0.00	0.00	1.00	0.00	0.55	2.00
SS20	0.00	0.00	0.00	0.00	0.00	2.75	0.94	0.00
SS21	0.21	0.00	0.60	1.00	0.00	0.38	0.60	5.51
SS22	0.34	0.00	0.00	0.00	0.00	0.00	0.63	0.00
SS23	0.60	0.39	0.60	0.74	0.00	0.00	0.54	8.78
SS24	0.19	1.00	0.00	0.00	0.00	0.00	0.84	10.0
SS25	0.41	1.00	0.00	0.00	0.00	0.16	0.55	10.0
SS26	0.76	1.00	0.00	0.00	0.00	106	0.56	10.0
SS27	0.00	0.00	0.00	0.00	0.00	0.00	0.19	0.00
SS28	0.00	0.00	1.00	0.00	0.00	0.00	0.66	5.00
SS29	0.41	0.00	1.00	0.00	0.00	0.00	0.71	5.00
SS30	0.15	0.00	0.62	1.00	0.00	0.58	0.81	5.59
NC31	1.88	0.00	0.00	0.60	0.00	0.00	0.35	1.49
NC32	0.27	0.41	0.00	0.30	0.00	0.77	0.79	4.83
NC33	0.00	0.00	0.00	0.00	0.00	0.94	0.92	0.00
NC34	0.12	0.37	0.00	0.68	0.29	0.01	0.75	6.03
NC35	1.06	0.00	0.00	1.00	0.00	0.00	0.48	2.50
NC36	0.78	0.00	0.00	0.60	0.00	0.00	0.56	1.49
NC37	0.51	0.00	0.00	1.00	0.00	0.00	0.52	2.50
NC38	0.00	0.00	0.00	0.00	0.00	0.00	0.49	0.00
NC39	0.00	0.00	0.00	0.00	0.00	0.00	0.53	0.00
NC40	0.62	0.00	0.00	0.00	0.00	0.00	0.63	0.00

Phenanthrene (Phen), Anthracene (Ant), Fluoranthene (Flt), Pyrene (Pyr), Benzo[a]anthracene (BaA), Chrysene (Chry), Benzo[a]pyrene (BaP), Indeno[1,2,3-cd]pyrene (IndP), Benzo[ghi]perylene (BghiP), South-East (SE), South-West (SW), Niger Delta (SS), North Central (NC).

ers for gasoline vehicle related sources^{44,46}. Factor 2, accounting for 19.89% of the total variance, was dominated by high positive loadings in BbF, BkF and IndP, with high negative bipolar loadings in Phen and Flt. Because BbF and IndP are known marker compounds of oil combustion^{43,47}, this factor may represent oil combustion related sources emanating from the activities of the oil and gas industries in the Niger Delta. Factor 3, accounting for 14.48% of the variance, is dominated by Pyr, Chry and BaP. Because Pyr, Flt and BaP are typical markers of coal combustion^{44,48} they could represent coal combustion related sources.

In the South-East region, three components were extracted accounting for 70.9% of the total variance. Factor 1 accounted for 29.59% and was dominated by BkF and DahA with negative loadings in Pyr and BghiP. BkF and DahA are representative compounds for diesel emissions^{43,49,50}. Factor 2 accounted for 25.25% of the total variance with high positive loadings for Ace, Flu, BaA, and BaP. Acy and Flu are tracers for wood combustion sources⁴⁴.

For the South-West region, three components were extracted accounting for 76.27% of the total variance. Factor 1 constituted 42.80% of the total variance, and

Table 7. PCA analysis of PAHs in honey samples.

	South East			South West			Niger Delta			North Central			
	Factor 1	Factor 2	Factor 3	Factor 1	Factor 2	Factor 3	Factor 1	Factor 2	Factor 3	Factor 1	Factor 2	Factor 3	Factor 4
Nap					.836								
Acy			.605		.695		.816				.585		-.561
Ace		.775			.828		.476	.369			-.800		
Flu	.332	.780	-.383	.824			-.422				-.799		
Phen	.420		-.646			.932		-.692	-.442				
Ant				.637	-.486	-.307	-.454	-.536		.902			
Flt						.935	.490	-.628	-.424	.843		-.300	
Pyr	-.876	-.337		.420	-.355			-.382	.845			.914	
BaA		.639		.910			.900						
Chry			.769	.864	-.445				.674	-.518			.468
BbF	.355		.795	.745	.382	.426		.671			.740	.618	
BkF	.914				.479			.804		.334		-.546	.754
BaP		-.784	.410	.707	.569				.743			.926	
IndP				.940				.637		.829			.490
DahA	.533	-.596		.952					.310		.446		.844
BghiP	-.876	-.337		.904	.373		.799			.701	.626		
Variance %	29.59	25.25	16.04	42.80	21.44	12.03	23.28	19.89	14.48	30.45	24.51	19.66	10.80

Napthalene (NaP), Acenaphthylene (Acy), Acenaphthene (Ace), Fluorene (Flu), Phenanthrene (Phen), Anthracene (Ant), Fluoranthene (Flt), Pyrene (Pyr), Benzo[*a*]anthracene (BaA), Chrysene (Chry), Benzo[*b*]fluoranthene (BbF), Benzo[*k*]fluoranthene (BkF), Benzo[*a*]pyrene (BaP), Indeno[1,2,3-*cd*]pyrene (IndP), Dibenzo[*a,h*]anthracene (DahA) and Benzo[*ghi*]perylene (BghiP).

was dominated by Flu, Ant, BaA, Chry, BbF, BaP, IndP, DahA and BghiP. Diesel emissions are characterized by BaA, Chry, BbF, IndP, and DahA^{43,44,49-51}. BghiP and BaP have been identified as tracers for automobile emissions because these compounds were found to be enriched in traffic tunnels^{47,52-54}. Factor 2 accounts for 21.44% of the total variance, with high loadings in Nap, Acy, Ace and BaP. Acy, Ace and Phen are characteristic of fossil fuel/biomass combustion^{45,49,55,56}. Factor 3 has high loadings in Phen and Flt.

The PCA factor loadings for the North Central region indicate that four components were extracted accounting for 85.42% of the total variance. Factor 1 has high loadings in Ant, Flt, IndP and BghiP. Flt, IndP, and BghiP are typical markers for coke production^{52,53}, while Ant is a product of wood combustion. This factor probably represents sources from coal, straw and wood combustion. Factor 2 has high positive loadings in Acy, BbF, and BghiP with high negative bipolar loadings in Ace and Flu. BbF and BghiP are markers for gasoline combustion, while Acy is a product of wood combustion. This factor represents a mixture derived from gasoline and wood combustion. Factor 3 accounts for 19.66% of the total variance with high loadings in Pyr, BbF and BaP. Factor 3 is related to exhaust emissions from stationary sources^{44,57,58}. Factor 4 has high positive loadings in BkF and DahA, and weak positive loadings in Chry and IndP. BkF and DahA are tracers of diesel emissions^{43,49,50}.

Conclusions

The regional distribution patterns of $\Sigma 16$ PAHs in these honeys followed the order: South-West > South-East > Niger Delta > North Central. The 5-ringed PAHs were the dominant PAH compounds in these honey samples. The incremental life cancer risk values for the majority of the samples were above the acceptable risk level of one in a million chance of additional cancer over a 52 year lifetime ($1/10^6$), while the margin of exposure values, based on the EFSA suggested indicators for occurrence and effects of PAHs in food, were greater than 10000. This indicated there is no additional risk associated with the consumption of these honeys based on the current ingestion rate, however, a considerable risk may arise with excessive consumption of these products. The PAH isomeric ratios and principal component analysis indicated that combustion of fossil fuels, natural gas and biomass, and automobile emissions were the main sources of PAHs in these honey samples from the different regions of Nigeria. This study provided useful information for environmental and food quality management in Nigeria.

Materials and Methods

Samples and Sample Collection

The sampling procedures have been previously de-

scribed by Iwegbue *et al.*⁵⁹ Briefly, a total of 40 samples of honey, consisting of raw samples from beehives and bottled samples from vendors, were collected from different locations across Nigeria. The honey samples were categorized into four geographical regions: (i) south-west region (latitude 6° N to 9° N; longitude 2° E to 5° E), (ii) Niger Delta (latitude 2° N to 8° N; longitude 5° E to 9° E), (iii) south-east region (latitude 5° N to 7° N; longitude 6° E to 8° E), and (iv) north central region (9° N to 14° N; longitude 2° E to 12° E). The major activities in these regions are agriculture, urbanization and industrial development. The south-western and south-eastern regions have higher concentrations of manufacturing industries than the northern region where there is predominantly mining and large-scale farming. The Niger Delta region, and some parts of the eastern region, house the crude oil production facilities with their associated multiple gas flaring units. Within a given location in a region at least 2 to 4 samples were collected and mixed together. Most vendors claimed that the honey samples collected were sourced from the wild except for 5 samples collected from bee farms in different locations. The bottled samples were labeled “undiluted pure natural honey” although the practice of enhancing honey with sugar syrup cannot be completely ruled out⁵⁹.

Reagents

All chemicals and reagents used were of analytical grade. Acetone and *n*-hexane were purchased from Riedel-de Haën (Seelze, Germany, with a purity of 99.8%) while dichloromethane (LC grade), anhydrous sodium sulfate (purity 99%), alumina and silica gel were purchased from BDH Chemicals (Poole, UK). A PAH standard mixture containing the US EPA 16 priority PAHs was purchased from Supelco (Bellefonte, PA, USA). Working mixed standard solutions containing all the PAHs were prepared by dilution of the stock solution with acetone and stored at -20°C in the dark to avoid volatilization and photodegradation.

Sample Preparation, Extraction and Clean-up

A mass of 5.0 g of the honey sample was mixed with the same amount of anhydrous Na₂SO₄, until the mixture became free-flowing. A 30 mL aliquot of hexane/dichloromethane (1 : 1 v/v) was added to the resulting material and placed in an ultra-sonic bath, and sonicated at 30°C for 30 minutes. The organic extract was filtered and the process was repeated three times by sonication of the residue with a fresh mixture of hexane/dichloromethane each time as described above. The extracts for each sample were combined and reduced to 1 mL by using a rotary evaporator, and subsequently cleaned-up by solid phase extraction with 2 g of alumi-

num oxide. The PAHs were eluted with 15 mL of hexane, 15 mL hexane and dichloromethane (9 : 1 v/v) and 20 mL of hexane and dichloromethane (4 : 1 v/v). The eluted fractions were combined and evaporated to approximately 0.5 mL with a gentle stream of nitrogen.

Chemical Analysis

The PAHs in the eluted fractions were measured with a gas chromatograph (HP 6890 Palo Alto, CA) equipped with a J&W Durabond 5 (cross-linked phenyl methyl siloxane) column (0.25 µm film thickness, 0.25 µm × 30 m) and a HP 5973 series mass-selective detector. The mass spectrometer was operated in the electron impact ionization mode (ionizing energy of 70 eV) scanning from *m/z* 50 to 450 at 3.6 scans/s. The ion source and quadrupole temperatures were 230°C and 150°C respectively. The operating conditions were as follows: the injection port and the GC/MS interface temperatures were 290°C and 250°C respectively. The column temperature was initially held at 80°C for 0.5 min and then increased to 230°C at 80°C/min and from 230°C to 280°C at 5°C/min, and held at 280°C for 18 min; the solvent delay was 6 min. The injection volume was 2 µL in pulsed splitless mode and the carrier gas was helium with a linear velocity of 1 mL/min.

Quality Control/Quality Assurance and Statistical Analysis

The quantification was carried out by the use of external calibrations which were obtained with PAH solutions at five concentration levels. To evaluate the extraction efficiency for the target compounds, known concentrations of standard PAH mixtures were added to fresh portions of already analyzed samples at three concentration levels and all analysis steps from extraction to chromatographic analysis were repeated. The recoveries for the PAH compounds were in the range of 66 to 103%. The relative standard deviations for replicate analyses (*n* = 3) were less than 6%. The *r*² values for the calibration lines for the PAH compounds ranged from 0.9994 to 0.9999 while the limits of detection and quantification for the PAH compounds ranged from 0.03 to 0.2 µg kg⁻¹ and from 0.1 to 0.7 µg kg⁻¹ respectively. The average inter-day and intra-day precision ranged between 1.8 and 6.9%. The performance characteristics of the present method meet the criteria specified in European Commission Regulation 836/2011 (recovery between 50 and 120%)⁶⁰. Analysis of variance (ANOVA) and Tukey multiple-comparison tests were used to determine whether the concentrations of the PAHs varied significantly within and between the regions respectively. Differences with *p* values less than 0.05 (*p* < 0.05) were considered to be statistically significant. The statistical calculations were performed

with SPSS version 20.5.

Estimation of Dietary Intakes and Risk Assessment

The estimated daily intake (EDI) of PAHs from the consumption of these honey samples was evaluated by using the formula:

$$EDI (\mu\text{g kg}^{-1} \text{ bw day}^{-1}) = \frac{MI \times C_{\text{PAH}}}{BW} \quad (1)$$

where MI is the mass of product ingested per day. In this study, an ingestion rate of 1.4 g/day was used based on the per capita consumption of 0.5 kg per annum per person. C_{PAH} is the concentration of the various indicators for the occurrence and effects of PAHs in foods such as BaP, PAH2, PAH4 and PAH8²⁸.

Margin of Exposure

The margin of exposure (MOE) approach was adopted to assess the risk of PAHs in these honey samples by using the various indicators of occurrence and effects of PAHs in foods (BaP, PAH2, PAH4 and PAH8)²⁸. The MOE is the ratio of a defined point on the dose response curve (reference point) for the adverse effect of the compound in the animal carcinogenicity study to the estimated average daily intake of the compound by humans⁶¹.

$$MOE = \frac{BMDL_{10} \times 10^6}{EDI} \quad (2)$$

The Benchmark Dose Lower Limit (BMDL₁₀) is the reference point that was derived from mathematical modelling of experimental tumour data within the observed range. The BMDL₁₀ values for BaP, PAH2, PAH4 and PAH8 are 0.07, 0.17, 0.34 and 0.49 mg kg⁻¹ bw day⁻¹, respectively²⁸.

Toxic Equivalency Factor

PAHs occur as mixtures and the risk to human health from various PAH exposures can be established by the toxicity or carcinogenic potency of the individual PAH compound relative to BaP. The risks of PAHs in soil, dust and foods have been assessed using the BaP toxic equivalent factor (BaP_{TEQ}) and the BaP mutagenic equivalent factor (BaP_{MEQ})⁶²⁻⁶⁷.

The BaP carcinogenic equivalent (BaP_{TEQ}) for the individual PAHs is given by the formula:

$$BaP_{\text{TEQ}} = \sum C_i \times BaP_{\text{TEF}} \quad (3)$$

where BaP_{TEF} is the cancer potency relative to BaP and C_i is the individual PAH concentration.

The BaP mutagenic equivalent (BaP_{MEQ}) for the individual PAHs is given by the equation:

$$BaP_{\text{MEQ}} = \sum C_i \times BaP_{\text{MEF}} \quad (4)$$

where BaP_{MEF} is the mutagenic potency relative to BaP and C_i is the individual PAH concentration. The BaP carcinogenic equivalency factors (BaP_{TEFs}) of the seven carcinogenic PAHs used were: BaP (1), BaA (0.1), BbF (0.1), BkF (0.01), Chry (0.001), DahA (1) and IndP (0.1)⁶⁴. The BaP mutagenic potency factors (BaP_{MEFs}) were BaP (1), BaA (0.082), BbF (0.25), BkF (0.11), Chry (0.017), DahA (0.29) and IndP (0.31)⁶⁶.

Estimation of Excess Cancer Risk

The excess cancer risk was estimated by using the general equation:

$$\text{Excess cancer risk} = \frac{EI \times ED \times CSF}{BW \times AT} \times 10^{-6} \quad (5)$$

where EI is the estimated intake, ED is the exposure duration in years (adults = 30 years), CSF is the oral cancer slope factor (mg kg⁻¹ d⁻¹), BW is the human body weight (assuming 60 kg weight), AT is the averaging time for carcinogens in years (assuming 52 years for the average Nigerian) and 10⁻⁶ is the conversion factor. The CSF data for individual PAHs are BaA = 0.73, Chry = 0.0073, BbF = 0.73, BkF = 0.073, BaP = 7.3, IndP = 0.73 and DahA = 7.3⁶⁴.

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References

1. dos Santos, J. S. *et al.* Honey classification from semi-arid, Atlantic and transitional forest zones in Bahia, Brazil. *J. Brazilian Chem. Soc.* **19**, 502-508 (2008).
2. Bilandžić, N. *et al.* Determination of trace elements in Croatian floral honey originating from different regions. *Food Chem.* **128**, 1160-1164 (2011).
3. Ru, Q. M., Feng, Q. & He, J. Z. Risk assessment of heavy metals in honey consumed in Zhejiang province, southeastern, China. *Food Chem. Toxicol.* **53**, 256-262 (2013).
4. Ioannidou, M. D., Zachariadis, G. A., Anthemidis, A. N. & Stratis, J. A. Direct determination of toxic trace metals in honey and sugars using inductively coupled plasma atomic absorption spectrometry. *Talanta* **65**, 92-97 (2005).
5. Lachman, J. *et al.* Analysis of minority honey components: Possible use for the evaluation of honey quality. *Food Chem.* **101**, 973-979 (2007).
6. Conti, M. E. & Botre, F. Honeybees and their products

- as potential bio-indicators of heavy metals contamination. *Environ. Monit. Assess.* **69**, 267-282 (2001).
7. Blasco, C. *et al.* Assessment of pesticide residues in honey samples from Portugal and Spain. *J. Agric. Food Chem.* **51**, 8132-8138 (2003).
 8. Hammel, Y. A. *et al.* Multi-screening approach to monitor and quantify 42 antibiotic residues in honey by liquid chromatography-tandem mass spectrometry. *J. Chromat. A* **1177**, 58-76 (2008).
 9. Dobrinas, S., Birghila, S. & Coatu, V. Assessment of polycyclic aromatic hydrocarbons in honey and propolis produced from various flowering trees and plants in Romania. *J. Food Comp. Anal.* **21**, 71-77 (2008).
 10. Yu, A.-R. & Lee, M.-Y. Preventive effects of Korean medicinal herbs on the phenanthrene-induced oxidative DNA damages. *Toxicol. Environ. Health. Sci.* **2**, 99-103 (2010).
 11. Martorell, I. *et al.* Polycyclic aromatic hydrocarbons (PAHs) in foods and estimated PAH intake by the population of Catalonia, Spain: Temporal trend. *Environ. Inter.* **36**, 424-432 (2010).
 12. Tuteja, G., Rout, C. & Bishnoi, N. R. Quantification of polycyclic aromatic hydrocarbons in leafy and underground vegetables. A case study around Panipat City, Haryana, India. *J. Environ. Sci. Technol.* **4**, 611-620 (2011).
 13. Iwegbue, C. M. A. *et al.* Polycyclic aromatic hydrocarbon profiles of some brands of canned fish in the Nigerian market. *Human Ecol. Risk Assess.* **21**, 157-168 (2015).
 14. Singh, S. & Vashishth, A. V. PAHs in some brands of tea. *Environ. Monit. Assess.* **177**, 35-38 (2011).
 15. Barreca, S. *et al.* Determination of selected polycyclic aromatic hydrocarbons by gas chromatography-mass spectrometry for the analysis of wood to establish the cause of sinking of an old vessel (Scauri wreck) by fire. *Microchemical. J.* **117**, 116-121 (2014).
 16. Olawoyin, R., Grayson, R. L. & Okareh, O. T. Eco-toxicological and epidemiological assessment of human exposure to polycyclic aromatic hydrocarbons in the Niger Delta, Nigeria. *Toxicol. Environ. Health. Sci.* **4**, 173-185 (2012).
 17. Orecchio, S. Polycyclic aromatic hydrocarbons (PAHs) in indoor emission from decorative candles. *Atmos. Environ.* **45**, 1888-1895 (2011).
 18. International Agency for Research on Cancer (IARC). Some non-heterocyclic polycyclic aromatic hydrocarbons and some related exposures. IARC Monograph on the Evaluation of Carcinogenic Risks to Humans. Monograph 2012, 92, World Health Organization International Agency for Research on Cancer, Lyon. 868 p. available from monographs.iarc.fr/ENG/monographs/vol92/index.php.
 19. Orecchio, S. & Papuzza, V. Levels, fingerprint and daily intake of polycyclic aromatic hydrocarbons (PAHs) in bread baked using wood as fuel. *J. Hazard Mat.* **164**, 876-883 (2009).
 20. Aguinaga, N., Campillo, N., Vinas, P. & Hernandez-Cordoba, M. Determination of 16 polycyclic aromatic hydrocarbons in milk and related products using solid phase microextraction coupled to gas chromatography-mass spectrometry. *Anal. Chimica Acta.* **596**, 285-290 (2007).
 21. Perugini, M. *et al.* Monitoring of polycyclic aromatic hydrocarbons in bees (*Apis mellifera*) and honey in urban areas and wildlife reserves. *J. Agric. Food Chem.* **57**, 7440-7444 (2009).
 22. Lambert, O. *et al.* Polycyclic aromatic hydrocarbons: Bees, honey and pollen as sentinels for environmental chemicals. *Chemosphere* **86**, 98-104 (2010).
 23. Moret, S., Purcaro, G. & Conte, L. S. Polycyclic aromatic hydrocarbons (PAHs) levels in propolis and propolis-based dietary supplement from the Italian market. *Food Chemistry* **122**, 333-338 (2010).
 24. Batelková, P. *et al.* Polycyclic aromatic hydrocarbons and risk elements from the South Moravian region (Czech Republic). *Acta Vet. Brno* **81**, 169-174 (2012).
 25. Corredera, L. *et al.* Evaluation of heavy metals and polycyclic aromatic hydrocarbons in honeys from different regions. *J. Food Prot.* **77**, 504-509 (2014).
 26. Ciemiak, A., Witzak, A. & Mocek, K. Assessment of honey contamination with polycyclic aromatic hydrocarbons. *J. Environ. Sci. Health* **48**, 993-998 (2013).
 27. Commission of the European Communities. Commission Regulation (EC) No.1881/2006 of 19 December 2006 setting maximum levels for certain contaminants in food stuffs, official Journal of the European Union L365/5 (2006).
 28. European Food Safety Authority (EFSA), Scientific Opinion of the Panel on Contaminants in the Food Chain on a request from the European Commission on Polycyclic Aromatic Hydrocarbons in Food. *The EFSA Journal* **724**, 1-114 (2008).
 29. Akpambang, V. O. E. *et al.* Determination of polycyclic aromatic hydrocarbons (PAHs) in commonly consumed Nigerian smoked/grilled fish and meat. *Food Addit. Contam.* **26**, 1096-103 (2009).
 30. Iwegbue, C. M. A. *et al.* Concentrations and profiles of polycyclic aromatic hydrocarbons in some popular fish species. *J. Food Prot.* **78**, 554-560 (2015).
 31. Iwegbue, C. M. A., Basse, F. I., Iwekumo, A. & Obi, G. Concentrations and health risks of polycyclic aromatic hydrocarbons in smoke-cured fish products in Nigeria. *Int. J. Environ. Stud.* <http://dx.doi.org/10.1080/00207233.2016.1155400> (2016).
 32. Iwegbue, C. M. A. & Basse, F. I. Concentration and health hazards of polycyclic aromatic hydrocarbons in some brands of milk in Nigerian Market. *Food Measure. Charact.* **763**, 177-184 (2013).
 33. Iwegbue, C. M. A. *et al.* Concentrations and health risk of polycyclic aromatic hydrocarbons in some brands of biscuits in the Nigerian Market. *Human. Ecol. Risk Assess.* **21**, 338-357 (2015).
 34. Iwegbue, C. M. A. *et al.* Concentrations of polycyclic aromatic hydrocarbons in some commercial brands of candies and chocolates in Nigeria. *Quality Assur. Crop.*

- Food Saf.* **7**, 661-667 (2015).
35. Iwegbue, C. M. A. Concentrations of Polycyclic aromatic hydrocarbons in hawked baked ready-to-eat foods consumed in southern Nigeria. *Acta Alimen.* DOI:10.1556/AAlim.2015.0004 (2015).
 36. Iwegbue, C. M. A. *et al.* Determination of polycyclic aromatic hydrocarbons in water-based and gin-based tea infusions of selected tea brands in Nigeria. *Poly. Arom. Compds* DOI:10.1080/10406638.2015.1033433 (2015).
 37. Zeiger, E. Identification of rodent carcinogens and non-carcinogens using genetic toxicity tests: premises, promises, and performance. *Regul. Toxicol. Pharm.* **28**, 85-95 (1998).
 38. Zeiger, E. Mutagens that are not carcinogens: faulty theory or faulty tests? *Mutat. Res.-Gen. Tox. En.* **492**, 29-33 (2001).
 39. DeMarini, D. M. *et al.* Bioassay-directed fractionation and salmonella mutagenicity of automobile and forklift diesel exhaust particles. *Environ. Health Persp.* **112**, 814-819 (2004).
 40. Seagrave, J. *et al.* Mutagenicity and in vivo toxicity of combined particulate and semi volatile organic fractions of gasoline and diesel engine emissions. *Toxicol. Sci.* **70**, 212-216 (2002).
 41. Essumang, D. K., Adokoh, C. K., Afriyie, J. & Mensah, E. Source assessment and analysis of polycyclic aromatic hydrocarbon (PAH's) in the Oblogo waste disposal sites and some water bodies in and around the Accra metropolis of Ghana. *J. Water Resource and Protection* **1**, 456-468 (2009).
 42. Yunker, M. B. *et al.* PAHs in the Franser River basin: a critical appraisal of PAH ratios as indicators of PAH source and composition. *Organic Geochemistry* **22**, 489-515 (2002).
 43. Lee, B. K. & Dong, T. T. Toxicity and source assignment of polycyclic aromatic hydrocarbons in road dust from urban residential and industrial areas in a typical industrial city in Korea. *J. Mater. Cycles Waste* **13**, 34-42 (2011).
 44. Fang, G. C. *et al.* Characterization, identification of ambient air and road dust polycyclic aromatic hydrocarbons in central Taiwan, Taichung. *Sci. Total Environ.* **327**, 135-146 (2004).
 45. Khalili, N. R., Scheff, P. A. & Holsen, T. M. PAH source fingerprints for coke ovens, diesel and gasoline engines, highway tunnels, and wood combustion emissions. *Atmosphere Environment* **29**, 533-542 (1995).
 46. Guo, H. *et al.* Particle-associated polycyclic aromatic hydrocarbons in urban air of Hong Kong. *Atmospheric Environment* **37**, 5307-5317 (2003).
 47. Harrison, R. M., Smith, D. J. T. & Luhana, L. Source apportionment of atmospheric polycyclic aromatic hydrocarbons collected from an urban location in Birmingham, U.K. *Environ. Sci. Tech.* **30**, 825-832 (1996).
 48. Dong, T. T. T. & Lee, B. K. Characteristics, toxicity, and source apportionment of polycyclic aromatic hydrocarbons (PAHs) in road dust of Ulsan, Korea. *Chemosphere* **74**, 1245-1253 (2009).
 49. Wang, X.-T. *et al.* Polycyclic aromatic hydrocarbons (PAHs) in urban soils of the megacity Shanghai: occurrence, source apportionment and potential human health risk. *Sci. Total Environ.* **447**, 80-89 (2013).
 50. Yang, B. *et al.* Source apportionment of polycyclic aromatic hydrocarbons in soils of Huanghuai Plain, China: comparison of three receptor models. *Sci. Total Environ.* **443**, 31-39 (2013).
 51. Kwon, H. O. & Choi, S. D. Polycyclic aromatic hydrocarbons (PAHs) in soils from a multi-industrial city, South Korea. *Sci. Total Environ.* **470-471**, 1494-1501 (2014).
 52. Larsen, R. K. & Baker, J. E. Source apportionment of polycyclic aromatic hydrocarbon in urban atmosphere: a comparison of three methods. *Environ Sci. Tech.* **37**, 1873-1881 (2003).
 53. Boonyatumanond, R. *et al.* Source of polycyclic aromatic hydrocarbons (PAHs) in street dust in a tropical Asian mega-city, Bangkok, Thailand. *Sci. Total Environ.* **384**, 420-432 (2007).
 54. Liu, Y. *et al.* Source apportionment of polycyclic aromatic hydrocarbons (PAHs) in surface sediments of the Huangpu River, Shanghai, China. *Sci. Total Environ.* **407**, 2931-2938 (2009).
 55. Simcik, M. F., Eisenreich, S. J. & Liroy, P. J. Source apportionment and source/sink relationships of PAHs in the coastal atmosphere of Chicago and Lake Michigan. *Atmos. Environ.* **33**, 5071-5079 (1999).
 56. Ravindra, K., Sokhi, R. & Van Grieken, R. Atmospheric polycyclic aromatic hydrocarbons; source attribution, emission factors and regulation. *Atmos. Environ.* **42**, 2895-2921 (2008).
 57. Yang, H. H., Lee, W. J., Chen, S. J. & Lai, S. O. PAH emission from various industrial stacks. *J. Hazard Mater.* **60**, 159-174 (1998).
 58. Kulkarni, P. & Venkataraman, C. Atmospheric polycyclic aromatic hydrocarbons in Mumbai, India. *Atmos. Environ.* **34**, 2785-2790 (2000).
 59. Iwegbue, C. M. A., Obi-Iyeke, G. E., Tesi, G. O. & Bassey F. I. Concentrations of selected metals in honey consumed in Nigeria. *Inter. J. Environ. Stud.* **72**, 713-722 (2015).
 60. European Commission (EC). Commission Regulation (EU) No 836/2011 of 19 August 2011 amending Regulation (EC) No 333/2007 laying down the methods of sampling and analysis for the official control of the levels of lead, cadmium, mercury, inorganic tin, 3-MCPD and benzo(a)pyrene in foodstuffs, http://www.iss.it/binary/lcdr2/cont/CR_836_2011.pdf (2011).
 61. Larsen, J. C. Risk assessment of chemicals in European traditional foods. *Trends Food Sci. Technol.* **17**, 471-481 (2006).
 62. Thompson, T. S., Clement, R. E., Thornton, N. & Luyt, J. Foundation and emission of PCDDs/PCDFs in the petroleum refining industry. *Chemosphere* **20**, 1525-1532 (1992).
 63. Nisbet, I. C. T. & LaGoy, P. K. Toxic equivalency fac-

- tors (TEFs) for polycyclic aromatic hydrocarbons (PAHs). *Regul. Toxicol. Pharmacol.* **16**, 290-300 (1992).
64. US EPA. Risk-based concentration Table. Environmental Protection Agency, Region 111 (Third Quarter) (1993).
65. Larsen, J. C. & Larsen, P. B. in *Air Pollution and Health* (eds Hester, E. E. & Harrison, R.R.) 33-36 (The Royal Society of Chemistry. Cambridge, UK, 1991).
66. Durant, J. *et al.* Human cell mutagenicity of oxygenated, nitrated and unsubstituted polycyclic aromatic hydrocarbons associated with urban aerosols. *Mut. Res.-Gen. Toxicol.* **371**, 123-157 (1996).
67. Durant, J. *et al.* Mutagenicity of C₂₄H₁₄ PAH in human cells expressing CYP1A1. *Mut. Res.-Gen. Toxicol.* **446**, 1-14 (1999).