RESEARCH ARTICLE-CHEMICAL ENGINEERING



# Optimization Study of *Leucaena leucocephala* Seed Extract as Natural Coagulant on Decolorization of Aqueous Congo Red Solutions

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### Abstract

Coagulation is a water and wastewater treatment technology that has been widely used due to its high efficiency and effectiveness. Various inorganic salts are commonly used as coagulant; however, there are some negative impacts of utilization of these chemical coagulants, such as high costs, large sludge volume, and also potential negative impact to human health. To overcome these problems, natural-based coagulants have been explored to reduce and substitute chemical coagulants. In this study we investigated utilization of *Leucaena leucocephala* seed protein, extracted using 0.5 M MgCl<sub>2</sub> solution as natural coagulant to treat synthetic Congo red wastewater. From the protein solubility profile at various pH, it could be observed that leucaena protein's isoelectric point was around pH 4. The optimization of coagulation condition was done by using Response Surface Method (RSM)—Central Composite Design (CCD) with variations of pH (2.93–4.12), coagulant dose (183.52–496.58 mg eq BSA/L), and dye concentration (44.1–56 mg/L). This experimental design was used to obtain the optimum condition that gave maximum dye removal percentage and minimum sludge volume. It was found that pH, coagulant dose, and dye concentration were significant for dye removal and sludge volume. Furthermore, it became evident that charge neutralization was the coagulation mechanism in this study. Based on the obtained model, 88.61% dye removal with 16.93 mL/L sludge volume was obtained at the optimum condition. The predicted optimum condition was in a good agreement with experimental data with relatively low error value. The obtained result implies good potency of leucaena as natural coagulant.

Keywords Congo red · Crude extract · Leucaena leucocephala · Natural coagulant · Response surface method

# **1** Introduction

Textile industry is classified as one of the largest and rapidly growing industries in many countries. It is known that the textile industry consumed a large number of water for its production thus generating a large volume of wastewater. Textile wastewater is high in biological oxygen demand (BOD), chemical oxygen demand (COD), total suspended solid (TSS), and other hazardous contents, namely heavy metal substances, halogenated hydrocarbons, pigments, dyes, and organic solvents [1]. According to Verma et al. (2012), typical textile wastewater had a wide range of BOD and COD value, namely 110–5500 mg/L and 50–18,000 mg/L, respectively,

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depending on the materials used during textile processing. It is also known that textile dyes have high-molecular weight and complex structure that making it non-biodegradable [2]. According to the regulations in Indonesia, the maximum BOD and COD in the discharge water for textile industry are 35–60 mg/L and 115–150 mg/L, depending on the wastewater debit, while the maximum organic content in drinking water is 10 mg/L. Direct discharging of textile wastewater into the river body should be avoided, as it could damage the aquatic ecosystem, as well as cause various diseases in human due to its mutagenic and carcinogenic nature.

Several methods to treat textile wastewater have been extensively studied, such as adsorption [3, 4], membrane filtration [5], electrocoagulation-flocculation [6] and advanced oxidation processes (AOPs) such as Fenton oxidation [7] and photo catalytic oxidation [8]. These previously mentioned technologies have their own advantages and drawbacks in wastewater treatment. However the coagulation—flocculation methods for treating wastewater are still widely used as



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primary chemical treatment due to its high efficiency and low cost [9]. Chemical coagulants such as alum (AlCl<sub>3</sub>), ferric chloride (FeCl<sub>3</sub>), and polyaluminum chloride (PAC) are commonly used [1]. While the effectiveness of these chemical coagulants is well-recognized, there are, nonetheless, disadvantages associated with usage of these coagulants such as: ineffectiveness in low-temperature water, relatively high-procurement costs, detrimental effects on human health (Alzheimer's, dementia), production of large less-degradable sludge and alteration of the water's pH [10]. It is therefore desirable to reduce and replace the chemical coagulants with natural-based coagulants to counteract the aforementioned drawbacks.

Natural coagulants have been used for more than 2000 years for traditional water treatment in India, Africa, and China [11]. Natural coagulants can be produced from plants (seeds, leaves, and roots), animals, and microorganisms. There are some advantages of natural coagulant such as non-toxic and non-corrosive nature, and also less volume sludge generation [10]. Based on these facts, utilization of natural coagulant may become an alternative for sustainable water and wastewater treatment, especially in the aspect of waste minimization and prevention, less hazardous chemicals, and its renewability [12]. Some plant-based natural coagulants such as *Moringa oleifeira* [13], *Vicia faba* [14], *Carica papaya* [15], and *Plantago ovata* [16, 17] have been investigated to treat various wastewaters.

In this study, Leucaena leucocephala, also known as white popinac or petai cina in Indonesian, was investigated its performance as natural coagulant. Leucaena is a native tree that can be easily found in tropical countries [18]. Leucaena is a species of legume that has high content of protein, namely 57-64% w of dry basis [19], making it a potential candidate as natural coagulant [20]. Furthermore, according to Sethi and Kulkarni (1993) 43.5% of total leucaena protein is globulin [21], a protein fraction that is soluble in salt solutions. Direct utilization of leucaena seed in powder form as natural coagulant is possible [22]. However, the active coagulating agent is commonly used in extract form to minimize the existence of organic substances mixed with active coagulant substances that could cause increase COD (Chemical Oxygen Demand) and DOC (Dissolved Organic Carbon) in the treated water [23, 24]. Solution of MgCl<sub>2</sub> was used to prepare the crude extract, to investigate utilization of divalent salt that has not been widely used, in comparison with previous studies where NaCl solution was used as solvent [25, 26]. Although the coagulation using leucaena has been previously investigated, it is crucial to investigate the effect of various variables, namely coagulant dose, pH, and dye concentration to the coagulation performance. Furthermore, maximum color removal and minimum sludge volume produced should be optimized for an efficient coagulation process using response surface methodology (RSM). RSM



with central composite design (CCD) are commonly used to investigate statistically relation between input variables and response with minimum number of experimental points [27, 28].

## 2 Materials and Methods

#### 2.1 Extraction of Protein from Leucaena Seed

Leucaena seed was procured from local market in Probolinggo, Indonesia. The seed was de-husked and the kernel was then oven dried (Memmert Un110) at 65 °C for 4 h to obtained kernel with water content 7-10%w. The dried kernel was then pulverized to a fine powder using a commercial food processor (Panasonic MX GX1462). The fine powder was then sieved through a standard 80 mesh and stored in an airtight container for further use. The extraction was carried out at various pH to determine the effect of pH extraction to the protein extractability, as it is known that protein extracted is usually following a "U" shaped curve with minimum value at its isoelectric point, and higher solubility at both acid and basic condition [29]. Five grams of dried leucaena powder was dispersed in 100 mL of 0.5 M MgCl<sub>2</sub> (Merck p.a.) with pH adjustments over the range of 2-9 by adding 0.1 M NaOH or HCl. The suspension was rigorously mixed for 60 min and separated by mean of centrifugation at 5000 rpm for 5 min. The protein concentration in the extract was determined using Bradford method [30], in which Bovine serum albumin (BSA, Merck p.a) was used as standard in this analysis method. The protein concentration was then stated as mg equivalent BSA per mL (mg eq BSA/mL).

#### 2.2 Coagulation Study

Congo red (Sigma-Aldrich) was used as a model substance of textile wastewater. A stock solution of 1 g/L was prepared by dissolving Congo red powder in distilled water. This stock solution was diluted as needed to obtain desired concentration. The coagulation study was done by using jar test apparatus. Congo red solution with concentration of 44.1–56 mg/L was adjusted to the desired pH value (2.93–4.12) using either 0.1 M HCl or NaOH. The leucaena crude extract was then added varying from 183.52 to 496.58 mg eq BSA/L according to the experimental design. The volume of extract that was added into the coagulation experiment was calculated using Eq. 1, where  $M_1$  is the initial protein concentration,  $V_1$  is the volume of extract added,  $M_2$  final concentration in the coagulation system, and  $V_2$  is the volume of wastewater treated.

 $M_1 \times V_1 = M_2 \times V_2 \tag{1}$ 

**Table 1** Range and level ofexperiment variables

Table 2 Design of experiments

Variable		Unit	Symbol	Range a	Range and level				
				$-\alpha$	- 1	0	+ 1	+α	
pН		_	А	2.52	2.93	3.53	4.12	4.53	
Coagulant dose		mg eq BSA/mL	, B	76.8	183.52	340.05	496.58	603.3	
Dye con	centration	mg/mL	С	40.04	44.10	50.00	56.00	60.00	
Run	А	В	С	Y1 Dye remov	al (%)	Y2 Slud	ge volume (	mL/L)	
1	0	0	0	-	ui (,c)		Ge volume (		
1	0	0	0	84.9		31			
2 3	+ 1	+ 1 - 1	+ 1 - 1	87.86 48.72		40			
3 4	+ 1 - 1		-	48.72 95.55		15 54			
4 5	$-1 \\ 0$	+ 1 0	+1 -α	93.33 77.68		21			
6	0	0 + α	$-\alpha$ 0	86.12		35			
7	0	$+ \alpha$	0	85.06		33 27			
8	+ 1	+ 1	- 1	85.8		35			
9	0	0	0	84.65		30			
10	- 1	+ 1	- 1	92.84		30			
11	- 1	- 1	- 1	90.7		19			
12	0	0	+α	86.53		32			
13	0	$-\alpha$	0	83.38		13			
14	+α	0	0	58.42		30			
15	- 1	- 1	+ 1	94.5		31			
16	0	0	0	85.22		30			
17	0	0	0	84.56		28			
18	$-\alpha$	0	0	90.88		50			
19	+ 1	- 1	+ 1	56.36		19			
20	0	0	0	84.94		31			

The mixture of Congo red solution and leucaena crude extract was rapidly mixed at 200 rpm for 2 min followed by slow mixing at 30 rpm for 30 min. The mixture was then allowed to settle for 1 h. The sludge volume was measured by using 1 L Imhoff cone after 1 h settling, calculated using Eq. 2.

sludge volume 
$$\left(\frac{mL}{L}\right) = \frac{V \text{sludge}(mL)}{V \text{solution}(L)}$$
 (2)

The initial (Co) and final concentration (Ci) of Congo red was measured using a spectrophotometer at its maximum wavelength, and the dye removal was calculated according to Eq. 3.

Dye Removal(%) = 
$$\frac{(Co - Ci)}{Co} \times 100$$
 (3)

## 2.3 Design of Experiments

A response surface methodology (RSM) based on central composite design (CCD) was employed in order to study the optimum condition of coagulation using Leucaena crude extract. RSM is a significant, fast and economical statistical technique for the determination of the interactive effects of parameters on experimental data [31]. Meanwhile, CCD is used for modeling the optimization process so that to obtain maximum dye removal with minimum sludge volume. The range and level of each variable is presented in Table 1. The experimental design in coded value with the responses is shown in Table 2.

The regression and graphical analysis with statistical significance were done using Design-Expert software (ver 7.0.0, Stat-Ease, Inc). In order to visualize the relationship between the experimental variables and responses, the response surface and contour plots were generated from the models. The validity of predicted optimum condition was confirmed by conducting triplicate experiments at the optimum val-



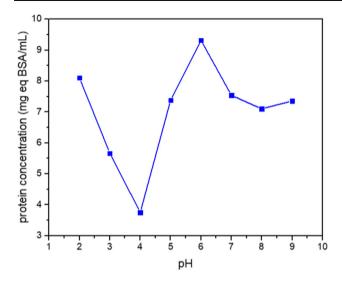


Fig. 1 Extracted protein concentration as the function of pH

ues of pH, dose, and Congo red concentration. Furthermore, the morphology of sludge flocs at the optimum condition was observed using scanning electron microscope (SEM; HITACHI SU3500).

## **3 Results and Discussions**

## 3.1 Effect of pH on Leucaena Seed Extraction

This research was conducted to determine the best pH to extract protein from Leucaena seed. Based on Fig. 1, it could be observed that at acidic pH (pH<4), the extracted protein concentration increases along with decreasing pH. It was possible due to the presence more positively charged protein molecules at low pH, making electrostatic repulsion between protein molecules were dominant. With the presence of dissociated salts in water, namely Mg<sup>2+</sup> and Cl<sup>-</sup>, positively charged protein molecules become more charged, resulting in more protein molecules repelling each other and subsequently bound to the salt ions, thus increasing the extracted protein [32, 33].

Proteins reached its minimum solubility at pH 4. At this condition, the charges of protein molecules were neutral, which is known as isoelectric point [34]. In this condition, the intermolecular forces between protein molecules would be dominant and the proteins would form peptide bonds, subsequently precipitate due to their increased molecular weight [35]. Above the protein isoelectric point, protein concentration in the solution tends to increase. This phenomenon could happen as the surface of the protein molecules would be negatively charged, making electrostatic repulsion between the protein molecules was dominant and with the presence of salt



ions, the proteins' charge would increase, thus increasing the solubility of the protein in the salt solution [32, 33].

The obtained isolectric point for leucaena protein was important information to determine the possible interaction between colloids and protein, which act as natural coagulant. The obtained isoelectric point was similar with other legumes, such as common beans (*Phaseolus vulgaris*) and fava beans (*Vicia faba*) with isoelectric point around 4–5 [36, 37], and soybeans (*Glycine max*) and winged beans (*Psophocarpus tetragonolobus*) with isoelectric point around 3.5–5 [38].

## 3.2 Congo Red Coagulation

#### 3.2.1 Analysis of Variance (ANOVA) and Model Fitting

The results of the ANOVA in Tables 3 and 4 show that the quadratic model is significant with 95% confidence level ( $\alpha < 0.05$ ). Moreover, the pH (*A*), coagulant dose (*B*), and dye concentration (*C*) were all significant. The effect of those variables and their interactions will create quadratic model equations, respectively, presented in Eqs. (4) and (5).

$$\operatorname{Ln}(Y1) = -2.7 + 4.7A + 1.0 \times 10^{-2}B + 4.5 \times 10^{-3}C - 1.0 \times 10^{-2}AB + 1.0 \times 10^{-2}AC + 6.9 \times 10^{-5}BC - 0.9A^{2} + 7.5 \times 10^{-6}B^{2} - 3.5 \times 10^{-4}C^{2} - 2.5 \times 10^{-5}ABC + 2.2 \times 10^{-3}A^{2}B - 2.1 \times 10^{-6}AB^{2}$$
(4)

$$\frac{1}{\text{Sqrt}(Y2)} = 0.6 + 0.2A - 5.9 \times 10^{-4}B$$
  
- 2.0 × 10<sup>-2</sup>C - 6.9 × 10<sup>-5</sup>AB  
+ 1.4 × 10<sup>-3</sup>AC + 5.3 × 10<sup>-6</sup>BC - 2.0  
× 10<sup>-2</sup>A<sup>2</sup> + 5.24 × 10<sup>-7</sup>B<sup>2</sup>  
+ 1.1 × 10<sup>-4</sup>C<sup>2</sup> (5)

Lack of fit and  $R^2$  describe the suitability of the model with the response data [39]. High *p* value (>0.05) of the lack of fit was obtained for both of responses, indicating insignificant lack of fit and significant model correlation between variables (pH, coagulant dose, and dye concentration) and responses (dye removal and sludge volume). While for  $R^2$ , the values were close to 1, indicating the model and data were well fitted. Furthermore, the difference between adjusted and predicted  $R^2$  was less than 0.2, indicating both models were valid and could be used for the optimization process [39]. Table 3 ANOVA for dye

Table 4 ANOVA for sludge

volume

removal

Source	Sum of squares	df	Mean square	F Value	<i>p</i> value Prob>F
Model	$6.3 \times 10^{-1}$	12	$5.2  imes 10^{-2}$	6,219.89	< 0.0001
A-pH	$9.8\times10^{-2}$	1	$9.8  imes 10^{-2}$	11,635.78	< 0.0001
B-coagulant dose	$1.0 \times 10^{-3}$	1	$1.0 \times 10^{-3}$	62.3	< 0.0001
C-Dye concentration	$1.3\times 10^{-2}$	1	$1.3\times 10^{-2}$	1,544.31	< 0.0001
AB	$1.2  imes 10^{-1}$	1	$1.2\times 10^{-1}$	14,178.09	< 0.0001
AC	$1.0 \times 10^{-3}$	1	$1.0  imes 10^{-3}$	147.73	< 0.0001
BC	$2.0 \times 10^{-3}$	1	$2.0 \times 10^{-3}$	268.36	< 0.0001
$A^2$	$4.2\times 10^{-2}$	1	$4.2\times 10^{-2}$	5,023.08	< 0.0001
$B^2$	$7.08\times10^{-6}$	1	$7.08\times10^{-6}$	0.84	0.39
$C^2$	$2.0 \times 10^{-3}$	1	$2.0 \times 10^{-3}$	263.19	< 0.0001
ABC	$2.0 \times 10^{-3}$	1	$2.0 \times 10^{-3}$	179.2	< 0.0001
$A^2B$	$4.8\times 10^{-2}$	1	$4.8\times10^{-2}$	5,774.9	< 0.0001
$AB^2$	$3.0 \times 10^{-3}$	1	$3.0 \times 10^{-3}$	385.97	< 0.0001
Residual	$5.87 \times 10^{-5}$	7	$8.39\times10^{-6}$		
Lack of Fit	$1.61\times 10^{-5}$	2	$8.07\times10^{-6}$	0.947	0.4480
Pure Error	$4.26\times 10^{-5}$	5	$8.52\times10^{-6}$		
Cor Total	$6.3  imes 10^{-1}$	19			

 $R^2 = 0.9999$ ; Adjusted  $R^2 = 0.9997$ ; Predicted  $R^2 = 0.9987$ , Adequate Precision = 289.192

Source	Sum of squares	df	Mean square	F Value	<i>p</i> value Prob> <i>F</i>
Model	$2.4 \times 10^{-2}$	9	$3.0 \times 10^{-3}$	119.25	< 0.0001
A-pH	$2.0  imes 10^{-3}$	1	$2.0  imes 10^{-3}$	97.86	< 0.0001
B-Coagulant Dose	$1.5\times 10^{-2}$	1	$1.5\times 10^{-2}$	654.42	< 0.0001
C-Dye Concentration	$3.0 \times 10^{-3}$	1	$3.0 \times 10^{-3}$	113.04	< 0.0001
AB	$3.0 \times 10^{-4}$	1	$3.0 \times 10^{-4}$	14.8	0.0032
AC	$2.0\times 10^{-4}$	1	$2.0  imes 10^{-4}$	8.1	0.0173
BC	$2.0 \times 10^{-4}$	1	$2.0 \times 10^{-4}$	8.5	0.0154
$A^2$	$1.0  imes 10^{-3}$	1	$1.0  imes 10^{-3}$	48.9	< 0.0001
$B^2$	$2.0 \times 10^{-3}$	1	$2.0 \times 10^{-3}$	104.37	< 0.0001
$C^2$	$2.0  imes 10^{-4}$	1	$2.0  imes 10^{-4}$	8.94	0.0136
Residual	$2.0  imes 10^{-4}$	10	$2.28\times 10^{-5}$		
Lack of Fit	$8.94 \times 10^{-5}$	5	$1.79\times 10^{-5}$	0.65	0.6784
Pure Error	$1.0 \times 10^{-4}$	5	$2.77 \times 10^{-5}$		
Cor Total	$2.5  imes 10^{-2}$	19			

 $R^2 = 0.9908$ ; Adjusted  $R^2 = 0.9825$ ; Predicted  $R^2 = 0.9644$ , Adequate Precision = 40.964

The suitability between data and model could also be seen in the predicted versus actual graphs, presented in Fig. 2.

#### 3.2.2 Effect of Coagulation Operating Parameters

The profile of the interaction between pH and coagulant dose to dye removal and sludge volume is presented in Fig. 3. Based on Fig. 3, it could be observed that with the increase of the coagulant dose accompanied by decreasing pH value could increase the dye removal and sludge volume. Based on Fig. 1, it could be seen that the extracted leucaena protein had isoelectric point around pH 4, where it exhibited the lowest solubility. On the other hand, Congo red's isoelectric point was known to be 3 [40], therefore the molecules would be negatively charged at pH>3, and vice versa. Thus, at pH below 4 the protein became positively charged and was readily available to neutralize anionic Congo red colloids. It is commonly observed that more destabilized colloids would



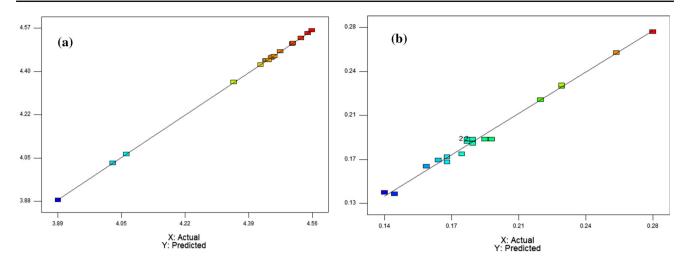


Fig. 2 Predicted versus actual plot for dye removal (a) and sludge volume (b)

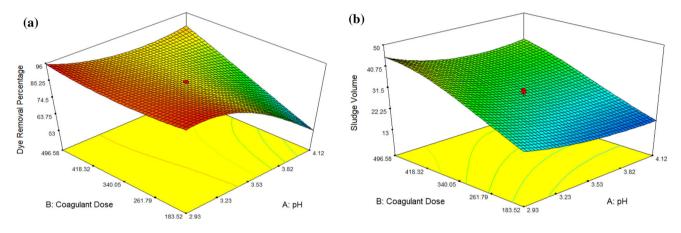


Fig. 3 Response surface plots of the interaction between pH and coagulant dose in dye removal (a); sludge volume (b)

significantly increase dye removal and sludge volume [25]. On the other hand, with increase of pH to 4.12, the dye removal and sludge volume were also decreased. It was possible as at the pH above protein isoelectric point, the protein molecules would be negatively charged, thus hindering the interaction between protein and Congo red molecules [41].

It could also be observed at pH 2.93; coagulant dose of 183.52 mg/L gave 23.25 mL/L sludge volume, while increasing the dose to 496.58 mg/L gave sludge volume increase to 43.75 mL/L with similar removal around 90%. There are generally three possible conditions of dosing in coagulation, namely low, optimum, and overdosing [42]. At low coagulant dose, there was not enough coagulant to neutralize the Congo red molecules, thus resulting in low removal and low sludge formation. Optimum condition could happen when the right amount of coagulant was added, and over addition of coagulant could give similar, or slightly lower removal but with higher sludge volume. This phenomenon could happen due to the presence of excessive coagulant; the colloid molecules became re-charged and re-stabilized. At this con-

re-charged and re-stabl

dition, the growth of flocs became hindered thus making the sludge became porous and with higher volume [25].

The profile of the interaction between pH and dye concentration to dye removal and sludge volume is presented in Fig. 4. It could be observed that with increase of dye concentration from 44.1 to 56 mg/L, the dye removal and sludge formation were also increased. This trend was possible as with the increase of dye concentration, more colloidal particles was contained in the solution, causing more colloids adsorbed to the protein [43]. On the other hand, the lower coagulation pH, the higher dye removal and sludge formation was observed. The observed trend confirmed the effect of pH on coagulation process in this study as previously explained.

The profile of the interaction between coagulant dose and dye concentration to dye removal and sludge volume is presented in Fig. 5. It could be observed that with the increase of coagulant dose and dye concentration, the dye removal (Fig. 5a–c) and sludge volume (Fig. 5d–f) were also increased. With the higher dye concentration; the more colloidal particles could be adsorbed to the protein. When the



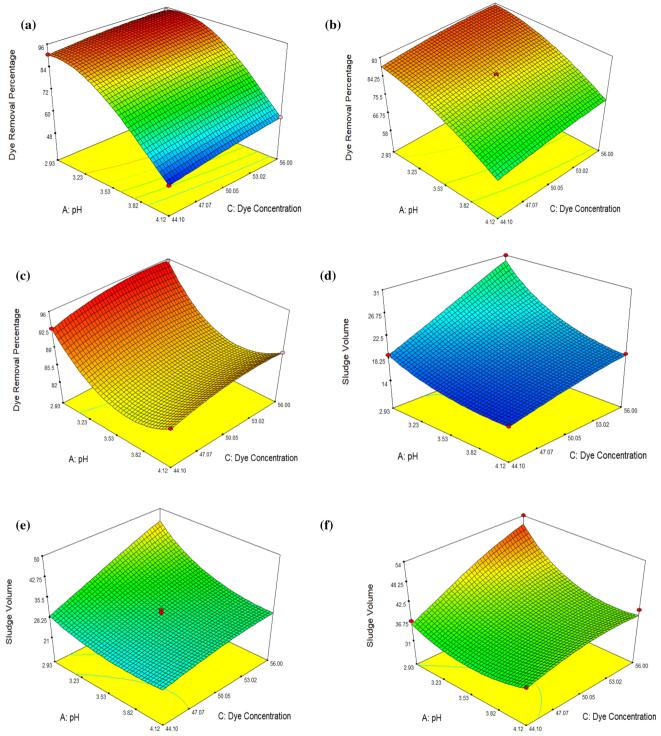


Fig. 4 Response surface plots of the interaction between pH and dye concentration to dye removal at low (a), center (b), and high (c) level coagulant dose; and to sludge volume at low (d), center (e), and high (f) level coagulant dose

dye concentration was too low, less colloidal particles could be destabilized by cationic proteins, making the excessive presence of protein could cause charge re-stabilization. As the result, colloidal particles became repulsive and inhibited coagulation [44]. Similar result was also reported by previous researchers when using protein extracted from *Moringa oleifera* [45], *Cocos nucifera* [46], and pine cone [47] as natural coagulant. Furthermore, it could be observed in Fig. 5 that at every level of pH, the addition of coagulant dose and dye concentration had not reached optimum conditions, thus



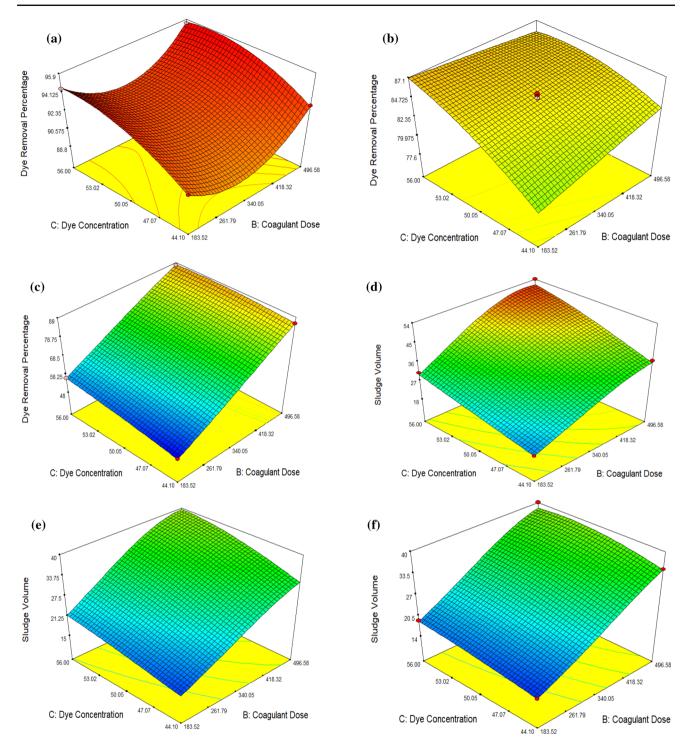


Fig. 5 Response surface plots of the interaction between coagulant dose and dye concentration to dye removal at low (a), center (b), and high (c) level pH; and to sludge volume at low (d), center (e), and high (f) level pH

further study, i.e., expanding the range of coagulant dose as it showed a tendency toward the optimum point, is needed.

 $B^2$  for sludge volume formation had high significance, and B, AC, BC, ABC,  $AB^2$  for the dye removal; A, AB, AC, BC,  $A^2$ ,  $C^2$  for sludge volume had moderate significance.

Based on these results, it could be concluded that the coagulation is influenced by pH (A), coagulant dose (B), and dye concentration (C). Furthermore, based on the ANOVA, parameters A, C, AB,  $A^2$  and  $A^2B$  for the dye removal; B, C,



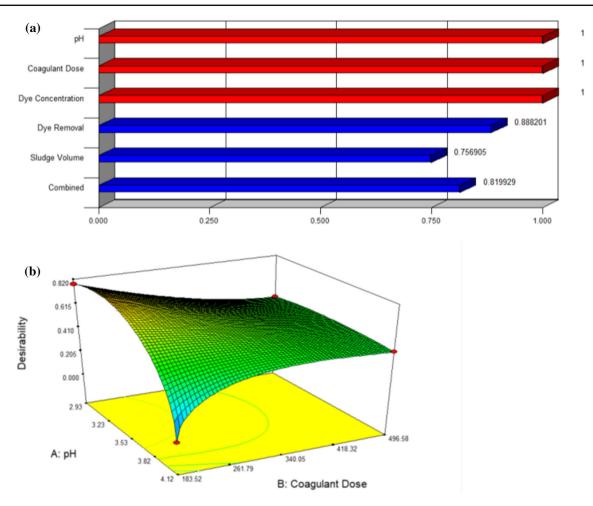


Fig. 6 Desirability value for each individual parameter and response (a) and desirability curve for the optimization study at low level of dye concentration

 Table 5 Results comparison between model and experiment of optimum condition

Parameter	Leucaena crude extract
рН	3.22
Coagulant dose (mg eq BSA/L)	183.52
Dye concentration (mg/L)	44.1
Dye removal (model) (%)	88.6
Dye removal (experiment) (%)	$88.5 \pm 1.08$
Error (%)	0.11
Sludge volume (model) (mL/L)	16.93
Sludge volume (experiment) (mL/L)	$17.33 \pm 0.5$
Error (%)	2.33

#### 3.2.3 Optimum Condition of Coagulation

Optimization was done in objective to obtain operating conditions with maximum dye removal and minimum sludge volume, while other variables (pH, coagulant dose, dye concentration) was set in range and importance for all variables and responses was set equal. According to the desirability value for each parameter, presented in Fig. 6a, it could be seen that all parameters, namely pH, coagulant dose, and dye concentration had desirability value 1.0, while dye removal was 0.89 and sludge volume was 0.76, resulted on the overall desirability of 0.82 (Fig. 6b). The desirability value should lie between 0 and 1, where higher desirability value; closer to 1, corresponds to more desirable response [48]. Based on the models, the optimum condition was found at pH 3.22, coagulant dose 183.53 mg eq BSA/L, and dye concentration 44.1 mg/L. Confirmation of the optimum condition result was done in triplicate, presented in Table 5. Based on this result, it could be observed that the model was valid and could predict optimum condition with low error, calculated using Eq. 6.

$$\operatorname{error}(\%) = \frac{|\operatorname{response}(\operatorname{experiment}) - \operatorname{response}(\operatorname{model})|}{\operatorname{rensponse}(\operatorname{experiment})} \times 100\%$$
(6)



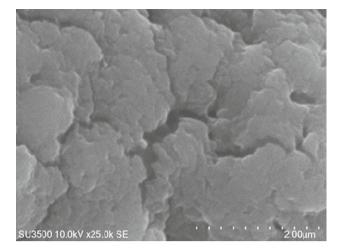


Fig. 7 Morphology of flocs from Leucaena seeds extract at its optimum condition (magnification of  $25,000 \times$ )

The sludge volume produced from leucaena extract coagulant was much lower than alum (40 mL/L) with similar coagulation condition [25]. This was possible due to bigger sludge size compared to alum making the sludge more compact thus occupying smaller volume [13]. The sludge observation is presented in Fig. 7. It could be observed that the flocs structure was dispersed with crack openings. Similar structure was also observed by previous study using protein as active coagulating agent [49]. According to [49], a compact-porous structure as observed in Fig. 6 was desirable due to the presence of large surface area of coagulant could facilitate coagulation of particles.

Based on the findings obtained in this study, it could be seen that further studies of utilization of leucaena seed's protein as natural coagulant is needed. According to Yin (2010), purification of crude extract using various technologies could improve the coagulant activity and efficiency. Various technologies that could be used such as ion exchange [50], dialysis [51], ultrafiltration [52], spray drying [53], and lyophilization [54]. Furthermore, a combination between natural coagulants with other water-wastewater treatment technologies, such as electrochemical techniques [55], membrane filtration [56], and adsorption [57], could also be explored.

## **4** Conclusion

In this study, we have investigated the utilization of MgCl<sub>2</sub> extracted leucaena protein as natural coagulant to treat synthetic Congo red wastewater. Furthermore, the effect various parameters, namely pH, coagulant dose, and dye concentration, to the dye removal and sludge volume formation was studied using response surface method (RSM)—central composite design (CCD). Based on the experiments, it was found



that all parameters are significant to the dye removal and sludge formation. Moreover, pH was found to be highly significant in dye removal as both of the protein and Congo red molecules' charges were pH sensitive. This phenomenon was in relation with the charge neutralization mechanism that is commonly found in protein-based natural coagulant. It also became evident that coagulant dose and dye concentration was correlated to each other. With the increase of coagulant dose at the same dye concentration, both dye removal and sludge volume also increased until optimal dye removal. Over addition of coagulant resulted on the increase of sludge volume, possibly due to colloid re-stabilization, with similar or slightly lower removal. The model obtained in this study was well fitted, with the optimum condition was achieved at pH 3.22, coagulant dose 183.52 mg eq BSA/L, and dye concentration 44.1 mg/L resulting in dye removal of 88.61% and sludge volume of 16.93 mL/L. In the light of the results obtained in this study, leucaena seed protein showed a great potential to be used as natural coagulant in water-wastewater treatment. Further study such as active coagulating agent purification and its combination with other technologies are needed for effective and efficient implementation as commercial natural coagulant.

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## **Compliance with Ethical Standards**

**Conflict of interest** The authors declare that they have no conflict of interest.

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