

Colletotrichum species in Australia

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Abstract Forty-four species of *Colletotrichum* are confirmed as present in Australia based on DNA sequencing analyses. Many of these species were identified directly as a result of two workshops organised by the Subcommittee on Plant Health Diagnostics in Australia in 2015 that covered morphological and molecular approaches to identification of *Colletotrichum*. There are several other species of *Colletotrichum* reported from Australia that remain to be substantiated by DNA sequence-based methods. This body of work aims to provide a basis from which to critically examine a number of isolates of *Colletotrichum* deposited in Australian culture collections.

Keywords Biosecurity · Diagnostics · Molecular methods · Phylogenetic methods · Plant pathogens · Taxonomy

Introduction

Colletotrichum (Ascomycota, Sordariomycetes) is one of the most important genera of plant pathogenic fungi worldwide, having been voted as one of the top 10 fungal pathogens by 495 scientists associated with the journal *Molecular Plant Pathology* (Dean et al. 2012). Species of *Colletotrichum* affect a range of plants, often causing diseases known as anthracnose, on many field and horticultural crops (Hyde et al. 2009a). On some tropical fruits, anthracnose can cause post-harvest losses of up to 100 % in the absence of control measures. *Colletotrichum* species are also common endophytes, epiphytes and saprobes (Hyde et al. 2009b).

According to Taylor et al. (2000), the three most common ways to recognize species are the morphological, biological, and phylogenetic species concepts. The morphological species

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concept has been used for fungal species descriptions and diagnoses since 1753, when Linnaeus published *Species Plantarum* (McNeill et al. 2012, art. 13). However, between the 1880s and the 1950s, hundreds of new *Colletotrichum* species were described based on the premise that *Colletotrichum* species were host-specific. The first monograph of *Colletotrichum* was published by von Arx (1957), who accepted 11 species, based on morphological characteristics alone, disregarding the plant hosts. Later Sutton (1980) reviewed this genus and accepted 23 species, and subsequently, 39 species of *Colletotrichum* (Sutton 1992). The identification and classification of species of *Colletotrichum* has undergone a taxonomic revolution in the last decade through the application of molecular phylogenetic methods (Cai et al. 2009, Crouch et al. 2009b, Hyde et al. 2009a, b). This approach has resulted in the recognition that several single species were actually complexes or aggregates of closely related cryptic species that were morphologically indistinguishable (Cannon et al. 2012), including *C. acutatum* (Damm et al. 2012b), *C. boninense* (Damm et al. 2012a), *C. caudatum* (Crouch 2014), *C. destructivum* (Damm et al. 2014), *C. gloeosporioides* (Weir et al. 2012), *C. gigasporum* (Liu et al. 2014), *C. graminicola* (Crouch and Beim 2009, Du et al. 2005), *C. orbiculare* (Damm et al. 2013) and *C. truncatum* (Damm et al. 2009). Several major revisions of these species complexes have resulted in the formal description of many new species of *Colletotrichum* (Cai et al. 2009, Crouch et al. 2009a, b, Crouch 2014, Damm et al. 2009, 2012a, b, 2013, 2014, Hyde et al. 2009a, b, Shivas and Tan 2009, Weir et al. 2012). Cannon et al. (2012) summarised the history of the classification of *Colletotrichum*, which currently has over 100 accepted species. The importance of using a polyphasic approach to species delimitation in *Colletotrichum*, together with a large sample size, was emphasised in a recent study (Liu et al. 2016) that showed the apparent *C. siamense* species complex (Sharma et al. 2015) was a single species.

Very little is known about the biology, pathogenicity, host range and geographical distribution of many of the recently recognised species of *Colletotrichum*. This has created a dilemma for plant pathologists. Hyde et al. (2010) first noted that there was an urgent need to reassess inventories of many plant pathogenic genera, including *Colletotrichum*, in Australia where the effectiveness of biosecurity measures relies heavily on the accuracy of specimen-based databases of plant pathogens (Shivas et al. 2006). Hyde et al. (2010) recognised that the revision of checklists must be supported by examination of herbarium specimens, living cultures and DNA libraries.

Two complementary workshops were held in Australia in 2015 as part of annual training offered to the National Plant Biosecurity Diagnosticians Network in order to ensure Australian plant biosecurity diagnosticians were introduced to taxonomic changes and diagnostic challenges that surround recent changes in the taxonomy of *Colletotrichum* species. The workshops were funded by Plant Health Australia and arranged

by the Subcommittee on Plant Health Diagnostics. The first workshop introduced the species complexes with a focus on morphology and biology. The second workshop introduced molecular and phylogenetic methods as applied to DNA sequence data obtained from isolates of *Colletotrichum*. Both workshops emphasised practical methods with more than 80 isolates examined from several Australian culture collections. Information gathered from these two workshops formed the basis for an up-to-date inventory of *Colletotrichum* species in Australia based on molecular phylogenetic evidence.

Materials and methods

Specimens and species identification

Living cultures of 86 specimens were sourced from Australian plant pathogen culture collections, including BRIP (Queensland), DAR (New South Wales), VPRI (Victoria) and WAC (Western Australia). A literature and database search found a further 106 Australian specimens with publicly accessible evidence of DNA sequence data from previous studies. This DNA sequence data was sourced from GenBank (<http://www.ncbi.nlm.nih.gov>) (Benson et al. 2013), and the Q-bank Fungi database (a reference database for mycological phytopathology, <http://www.q-bank.eu/Fungi/>). Confirmation that species of *Colletotrichum* occurred in Australia required that the specimen had an unambiguous DNA sequence that matched data from the ex-type specimen.

DNA extraction, PCR amplification and DNA sequencing

Mycelia were collected from cultures grown on potato dextrose agar (Difco™, Becton, Dickinson and Company, New Jersey, USA) and macerated with 0.5 mm glass beads (Daintree Scientific) in a Tissue Lyser (QIAGEN). Genomic DNA was extracted with the Genra Puregene DNA Extraction kit (QIAGEN) or with ISOLATE II Plant DNA kit (Bioline) according to the manufacturers' instructions. Gene sequences were obtained from up to four nuclear gene regions for species identifications. These are *glyceraldehyde-3-phosphate dehydrogenase* (GAPDH), *glutamine synthetase* (GS), the *internal transcribed spacer* (ITS), and *β-tubulin 2* (TUB). Primers used in this study are shown in Table 1. Where the standard GS primers (Stephenson et al. 1997) sequenced poorly, the primers from Weir et al. (2012) were used instead. All gene regions were amplified with the Phusion High-Fidelity PCR Master Mix (New England Biolabs). The PCR products were purified with the QIAquick PCR Purification Kit (QIAGEN), and sequenced on the 3730xl DNA Analyzer (Applied Biosystems) by a commercial company (Macrogen Incorporated, Korea) using the amplifying primers. All sequences generated were assembled using Geneious v. 9.1

Table 1 Primers used in this study, with sequences and references

Gene	Product name	Primer	Direction	Sequence (5'-3')	Reference
GAPDH	<i>Glyceraldehyde-3-phosphate dehydrogenase</i>	GDF	Forward	GCC GTC AAC GAC CCC TTC ATT GA	Templeton et al. 1992
		GDR	Reverse	GGG TGG AGT CGT ACT TGA GCA TGT	Templeton et al. 1992
GS	<i>Glutamine synthetase</i>	GSF1	Forward	ATG GCC GAG TAC ATC TGG	Stephenson et al. 1997
		GSF3*	Forward	TCG CCC GCA CTG CTG CAG CCG G	Weir et al. 2012
		GSR1	Reverse	GAA CCG TCG AAG TTC CAG	Stephenson et al. 1997
		GSR2	Reverse	GAA CCG TCG AAG TTC CAC	Weir et al. 2012
ITS	<i>Internal transcribed spacer</i>	V9G	Forward	TTACGTCCCTGCCCTTTGTA	de Hoog and Gerrits van den Ende 1998
		ITS4	Reverse	TCC TCC GCT TAT TGA TAT GC	White et al. 1990
TUB	<i>β-tubulin 2</i>	T1	Forward	AAC ATG CGT GAG ATT GTA AGT	O'Donnell and Cigelnik 1997
		Bt2b	Reverse	ACC CTC AGT GTA GTG ACC CTT GGC	Glass and Donaldson 1995

* The sequence given by Weir et al. (2012) has been corrected here

(Biomatters Ltd), and deposited in GenBank (Table 2, in bold). These sequences were compared against those from type specimens using BLASTn (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>).

Results

The names of Australian *Colletotrichum* species that were verified by DNA sequence analysis are given in the following numbered list. This list contains only species that were verified by comparison of their DNA sequence data against type specimens. The GenBank accession numbers of sequences generated in this study are provided in Table 2 (in bold font) together with reference sequences generated in other studies (Roman font).

Australian species of *Colletotrichum* verified by DNA sequence data

1. *Colletotrichum acutatum* J.H. Simmonds, Queensland Journal of Agricultural and Animal Science 25: 178A (1968).

Colletotrichum acutatum was first described from Redlands Horticultural Research Station, Cleveland, Queensland on papaya (*Carica papaya*) by Simmonds (1965, 1968). This species has since been recognised as an important pathogen that causes anthracnose on a range of plants worldwide. However, *C. acutatum* has long been recognised as a complex of closely related species that have conidia with acute ends (Simmonds 1965). The chequered taxonomic history of *C. acutatum* was summarized by Damm et al. (2012b), who designated an epitype that has ultimately provided taxonomic stability for this species. In Australia, *C. acutatum* is widespread and known to cause diseases or be associated with disease symptoms on a range of plants, including papaya, strawberry, olives and pistachio.

The literature indicates that the *C. acutatum* species complex may have a much wider host range in Australia, where it has been reported to cause diseases on avocado, tomato (Simmonds 1965), grapes (Melksham et al. 2002, Whitelaw-Weckert et al. 2007), olive (Spooner-Hart et al. 2007) and almond (McKay et al. 2009). However, many of these records require verification. *Colletotrichum acutatum* can be distinguished from other species in the *C. acutatum* species complex by any of the six genes analysed in Damm et al. (2012b). The Australian specimens of *C. acutatum* were identified based on the 100 % identity of ITS, GS and/or TUB sequences to *C. acutatum* ex-type strain CBS 112996 (Table 2).

2. *Colletotrichum alcornii* J.A. Crouch, IMA Fungus 5: 27 (2014).

Colletotrichum alcornii belongs to the *Colletotrichum caudatum* species complex (Crouch 2014). *Colletotrichum alcornii* is only known from two native Australian grass species in south-east Queensland. Although morphologically similar to four other species in the *C. caudatum* species complex, *C. alcornii* is easily distinguished based on ITS sequences (Crouch 2014).

3. *Colletotrichum alienum* B.S. Weir & P.R. Johnst., Studies in Mycology 73: 139 (2012).

Colletotrichum alienum belongs to the *C. gloeosporioides* species complex (Weir et al. 2012). In Australia, *C. alienum* has been reported from avocado (Weir et al. 2012), *Grevillea* sp. (Liu et al. 2013b), *Protea* spp. (Crous et al. 2013) and *Nerium oleander* (oleander) (Schena et al. 2014). *Colletotrichum alienum* was identified as a serious anthracnose pathogen of Proteaceae in South Africa, Europe and Australia (Liu et al. 2013b, Crous et al. 2013). *Colletotrichum alienum* is best distinguished from other

Table 2 List of Australian species of *Colletotrichum* verified by DNA sequencing. Sequences derived from this study are in **bold**

Species	Culture ^{a b}	Host (symptoms)	Locality (State ^c , city/town)	GenBank Accessions				
				ITS	GAPDH	GS	TUB	TUB
<i>Colletotrichum acutatum</i>	VPRI 10179 = BRIP 52653	<i>Anemone</i> sp. (stem spot)	Vic, Geelong	GU183350				GU183309
	VPRI 10517	<i>Anemone</i> sp. (leaf spot)	Vic, Geelong	GU183353				GU183310
	= BRIP 52656							
	= IMI 223120							
	WAC 12421	<i>Boronia megastigma</i>	SA, Mt Barker					
	= BRIP 52695							
	IMI 117617 ^{HT}	<i>Carica papaya</i> (fruit rot)	Qld, Ormiston	AF411700				
	BRIP 4693 ^{IS}							
	CBS 112996							
	= ATCC 56816							
	= ICMP 1783 ^{3x-EP}							
	WAC 5416	<i>Fragaria x ananassa</i> (fruit rot)	WA, Wanneroo	GU183356				GU183312
	= BRIP 52691							
	= CBS 127602							
	BRIP 54038	<i>Malus domestica</i> (bitter rot)	Qld, Brisbane	KU498249				JQ950000
	CBS 111993	<i>Grevillea</i> sp.	NSW, Mt Annari	JQ948349				GU183307
	BRIP 27048	<i>Mangifera indica</i> (fruit lesion)	Qld, Ayr	GU183326				GU183313
WAC 5990 = BRIP 52692	<i>Olea europaea</i>	WA, Kalamunda	GU183357				KX069817	
WAC 12568			KX069826			KX069811		
= BRIP 63678								
VPRI 41821	<i>Pistacia vera</i> (leaf and fruit spots)	NSW, Kyalite	KU498251				KU221358	
= BRIP 62667								
VPRI 41430	<i>Pistacia vera</i>		KU498250				KU221356	
= BRIP 54786								
VPRI 41429	<i>Pistacia vera</i>	Vic, Robinvale					KU221357	
= BRIP 54787								
VPRI 41432							KU221355	
= BRIP 54785								
WAC 5506 = BRIP 52690								
IMI 336479	<i>Ranunculus</i> sp.							
VPRI 16304								
= BRIP 52652								
WAC 11064 = BRIP 62860	<i>Vicia faba</i> var. <i>minor</i>	WA, unknown	GU183355				GU183311	
IMI 176617 ^{PT}	<i>Bohrtochiolea bladnii</i>	WA, Perth	JQ948367				JQ950018	
= BRIP 8779 ^{PT}			GU183349				GU183308	
IMI 176619 ^{HT}	<i>Imperata cylindrica</i> var. <i>major</i>	Vic, Clayton South						
BRIP 8824 ^{IS}								
DAR 25578	<i>Fragaria x ananassa</i> (fruit rot)	WA, Dongara	KX069821		KX069805	KX069808	KX069812	
= BRIP 61805		Qld, Toogoolawah	JX076857					
DAR 32071	<i>Fragaria x ananassa</i>	Qld, Caboolture						
= BRIP 61799			JX076858					
CBS 111982	<i>Fragaria x ananassa</i>	NSW, Sydney	KU498254		KU221344	KU221335	KU221362	
CBS 132880	<i>Fragaria x ananassa</i>	NSW, Gosford	KU498253			KU221334	KU221361	
F263	<i>Grevillea</i> sp.	NSW, unknown						
DAR 37820	<i>Nerium oleander</i>	NSW, unknown	KC297069		KC296998	KC297021	KC297091	
= IMI 313842	<i>Persea americana</i>	Unknown	KC297075		KC297005	KC297022	KC297099	
		NSW, Murwillumbah	KC425654				KC435707	
			JX010217		JX010018	JX010074	JX010385	

Table 2 (continued)

Species	Culture ^{a,b}	Host (symptoms)	Locality (State ^c , city/town)	GenBank Accessions				
				ITS	GAPDH	GS	TUB	
<i>Colletotrichum aotearoa</i>	= ICMP 18691 CBS 132448 VPRI 40656 = BRIP 62670 IMI 313839 = ICMP 18696	<i>Banksia marginata</i> (leaf spot) <i>Hedyocarya angustifolia</i> (leaf spot)	Vic, Dunkeld Vic, Marysville	KC297064 KU498255	KC296997 KU221340	KC297020	KC297089 KU221363	
<i>Colletotrichum asianum</i>	BRIP 57972 BRIP 58110 BRIP 583266 BRIP 57882 BRIP 28734 BRIP 28451 BRIP 62535b CBS 131325 ^{PT} IMI 279189 ^{HT} BRIP 14026 ^{IS}	<i>Mangifera indica</i> <i>Mangifera indica</i> (stem dieback) <i>Mangifera indica</i> (stem dieback & leaf wilt) <i>Mangifera indica</i> (stem canker) <i>Mangifera indica</i> (anthracnose) <i>Mangifera indica</i> (fruit rot) <i>Mangifera indica</i> (fruit lesion) <i>Hakea</i> sp. <i>Axonopus fissifolius</i>	NSW, Sextonville NT, Bees Creek Qld, Brisbane Qld, Townsville Qld, Nangaba Qld, Gin Gin Qld, Mareeba Qld, Tolga WA, Alcoa Qld, Peregian Beach	JX010192 KF877314 KU498262 KU498261 JN191511 KU498256 KU498263 JQ948456 EU554086	JQ00973 JX010073	JX010384	JX010384	
<i>Colletotrichum australe</i>	CBS 131325 ^{PT}						JQ950107	
<i>Colletotrichum axonopodi</i>	IMI 279189 ^{HT} BRIP 14026 ^{IS}							
<i>Colletotrichum boninense</i>	CBS 112762	<i>Leucospermum</i> sp.	NSW, Nunkeri	AY376522			AY376570	
<i>Colletotrichum brevisporum</i>	BRIP 54898b BRIP 43898	<i>Carica papaya</i> (fruit rot) <i>Passiflora edulis</i> (vascular discoloration)	Qld, South Johnstone Qld, Gregory River	KU498265 KU498264				
<i>Colletotrichum brisbanense</i>	IMI 117622 ^{HT} BRIP 4684 ^{IS}	<i>Capsicum frutescens</i> (fruit rot)	Qld, Brisbane	JQ948291	JQ948621		JQ949942	
<i>Colletotrichum chlorophyti</i>	CBS 292.67 ^{ex-IS}	<i>Sylostanthes hamata</i>	Qld, Townsville	GU227895	GU228287		GU228189	
<i>Colletotrichum circinans</i>	VPRI 42617	<i>Alium porrum</i>	Vic, Devon Meadows	KU612898	KU612899			
<i>Colletotrichum coccodes</i>	DAR 32115 = BRIP 61800 WAC 8123 = BRIP 63677 VPRI 41585 = BRIP 54784	<i>Fragaria x ananassa</i> (fruit rot) <i>Iris</i> sp. <i>Solanum lycopersicum</i> (root rot & anthracnose)	NSW, Pimlico WA, Millendon Vic, Warragul	KU498266 KX069825		KX069810	KX069816 KU221372	
<i>Colletotrichum cymbidicola</i>	IMI 34792 ^{HT}	<i>Cymbidium</i> sp. (leaf lesion)	WA, Fremantle	JQ005166	JQ005253		JQ005600	
<i>Colletotrichum dematium</i>	IMI 350847	<i>Solanum tuberosum</i>	Tas, unknown	GU227825	GU228217		GU228119	
<i>Colletotrichum destructivum</i>	UQ349 UQ343	<i>Medicago scutellata</i> <i>Ornithopus compressus</i>	Qld, Kingsthorpe Qld, Leyburn	AF451909 AF451908				
<i>Colletotrichum dracaenophilum</i>	VPRI 41774 VPRI 41775	<i>Dracaena sanderiana</i> (stem lesion)	Vic, Narre Warren	KU612894 KU612896	KU612895 KU612897			
<i>Colletotrichum fiorintiae</i>	WAC 13265 = BRIP 52696 BRIP 28761 BRIP 20127 BRIP 29285 BRIP 29284 BRIP 52336 BRIP 52335 BRIP 28522 = ICMP 125688	<i>Acacia acuminata</i> <i>Mangifera indica</i> (endophyte) <i>Persea americana</i> (fruit rot)	Vic, Vermont WA, Manjimup Qld, Yarvun Qld, Brisbane Qld, Mt Tamborine WA, Pemberton Qld, Bli Bli	GU183320 GU183333 GU183320 GU183336 GU183335 GU183347 GU183346 JX01066			GU183268 GU183269 GU183268 GU183271 GU183270 GU183273 GU183272	
<i>Colletotrichum gloeosporioides</i>	DAR 76936 = ICMP 18736 F264 F265	<i>Persea americana</i> (fruit rot) <i>Carya illinoensis</i> <i>Citrus limon</i>	NSW, Tamworth unknown	JX010151 KC425621 KC425619	JX009946 JX009976		KC425696 KC425695	

Table 2 (continued)

Species	Culture ^{a,b}	Host (symptoms)	Locality (State ^c , city/town)	GenBank Accessions			
				ITS	GAPDH	GS	TUB
<i>Colletotrichum godetiae</i>	F266	<i>Citrus sinensis</i>	Vic, Lindenow	KC425620			KC425694
	VPRI 42335		WA, Gingin	KU612890	KU612891		KX069818
	WAC 12803 = BRIP 63680		Vic, Olinda	KX069827			KU612889
	VPRI 41712 = BRIP 54820		Vic, Mt Selby				
	VPRI 16247		NSW, Nemingham	KU612886	KU612888	JX010132	
	ICMP 18539 ^{ex-HT}		Qld, Palmwoods	JQ005204	JQ005291		JQ005638
	BRIP 29085 = CBS 127597		NSW, unknown	JQ005299			JQ005646
	CBS 111998		Qld, Brisbane	JQ005203	JQ005290		JQ005637
	BRIP 28443 = CBS 127596		unknown	JQ005178	JQ005265		JQ005612
	CBS 127595		unknown	JQ005186	JQ005273		JQ005620
<i>Colletotrichum kahawae</i> subsp. <i>ciggaro</i>	CBS 127591	<i>Sclerocroton integerrimus</i>	WA, Mingenew	KU974988	KU974972		KU975004
	WAC 10408	<i>Lupinus albus</i>					
	= BRIP 63847						
	WAC 9624 = IMI 375715		WA, Morawa	JQ948161	JQ948491		JQ949812
	WAC 12999			KU974991	KU974979		KU975012
	= BRIP 63855						
	WAC 10361			KU974985	KU974969		KU975001
	= BRIP 63844						
	WAC 10359		WA, Three Springs	KU974984	KU974968		KU975000
	= BRIP 63843						
<i>Colletotrichum lupini</i>	WAC 13001 =		WA, Yandanooka	KU974998	KU974981		KU975014
	BRIP 63857						
	WAC 12994	<i>Lupinus angustifolius</i>	WA, Dongara	KU974992	KU974975		KU975007
	= BRIP 63850						
	WAC 13000	<i>Lupinus cosentinii</i>	WA, Badgingarra	KU974997	KU974980		KU975013
	= BRIP 63856						
	WAC 13003		WA, Eneabba	KU974999	KU974983		KU975016
	= BRIP 63859						
	WAC 12997		WA, Geraldton	KU974995	KU974978		KU975010
	= BRIP 63853						
<i>Colletotrichum karstii</i>	WAC 10425		WA, Mingenew	KU974989	KU974973		KU975005
	= BRIP 63848						
	WAC 10427			KU974990	KU974974		KU975006
	= BRIP 63849						
	WAC 10404		WA, Morawa	KU974987	KU974971		KU975003
	= BRIP 63846						
	WAC 12998		WA, Mullewa	KU974996			KU975011
	= BRIP 63854						
	WAC 10398		WA, Three Springs	KU974986	KU974970		KU975002
	= BRIP 63845						
<i>Colletotrichum karstii</i>	WAC 12996		WA, Walkaway	KU974994	KU974977		KU975009
	= BRIP 63852						
	WAC 13002						
	= BRIP 63858						
	WAC 12995		WA, Mingenew	KU974993	KU974976		KU975008
	= BRIP 63851						

Table 2 (continued)

Species	Culture ^{a,b}	Host (symptoms)	Locality (State ^c , city/town)	GenBank Accessions			
				ITS	GAPDH	GS	TUB
<i>Colletotrichum musae</i>	BRIP 59413	<i>Musa acuminata</i> (crown rot)	Old, Tully WA, Camarvon	KU498267			
	WAC 6994 = BRIP 63674	<i>Musa acuminata</i> (leaf spot)		KX069823	KX069807		KX069814
	WAC 13307			KX069828			KX069819
	= BRIP 63681						
	BRIP 61880b	<i>Musa</i> sp. (endophyte)	Old, Atherton Old, Walkamin NSW, Macksville	KU498270			
	BRIP 61879b			KU498269			
	DAR 30640	<i>Musa</i> sp. (fruit rot)		KU498268			
	= BRIP 61798						
	VPRI 10347	<i>Citrus limon</i> (leaf spot)	Vic, unknown				KU221374
	= BRIP 54771						
VPRI 41914	<i>Fragaria x ananassa</i> (plant wilt)	Vic, Heatherton	KU498271	KU221346		KU221376	
= BRIP 62666							
VPRI 11206	<i>Protea compacta</i> (stem dieback)	Vic, Emerald				KU221375	
= BRIP 54791 = IMI 260254							
CBS 482.82	<i>Protea</i> sp. (leaf spot)	Vic, Burnley	JQ948213	JQ948543		JQ949864	
VPRI 17866		Vic, Doncaster				KU221373	
= BRIP 54770							
BRIP 49125	<i>Ocimum basilicum</i> (leaf spot)	Old, Brisbane NSW, Bringsely	KU498272			KF178590	
DAR 81253	<i>Benincasa hispida</i> (leaf lesion)		KF178469	KF178494		KF178614	
= CBS 129432							
VPRI 41776	<i>Dracaena sanderiana</i>	Vic, Blackburn	KU612982	KU612893			
WAC 15254 = BRIP 62862	<i>Phoromitum tenax</i> (anthracnose)	WA, Busselton	KU947422			KU947421	
VPRI 12013 = BRIP 62668	<i>Daphne odora</i> (leaf and tip dieback)	Vic, Mitcham	KU498273	KU221341		KU221364	
IMI 117612 ^{HT}	<i>Carica papaya</i> (fruit rot)	Old, Brisbane	JX010276	JZ009934	JX010104	JZ010414	
PDD 28797 ^{EP}							
ICMP 1778 ^{ex-EP}							
ICMP 1780			JX010186	JX010010			
BRIP 48744	<i>Litchi chinensis</i> (anthracnose)	Old, Mena Creek	JN191512			JN191526	
BRIP 57881	<i>Mangifera indica</i> (stem canker)	Old, Narangba	KU498275	KU221348		KU221378	
BRIP 57981	<i>Passiflora edulis</i> (anthracnose)	NT, Darwin				KF877320	
BRIP 55403	<i>Persea americana</i> (stem dieback)	NSW, Durambah	KU498274	KU221347		KU221377	
ICMP 12564		Old, Home Hill	JX010184	JX009919			
= BRIP 28418							
VPRI 32546	<i>Salix alba</i> subsp. <i>vitellina</i> (leaf lesion)	ACT, Brindabella	KU498279	KU221351		KU221381	
= BRIP 60908							
VPRI 32735	<i>Salix cinerea</i> (leaf lesion)	Vic, Cann River	KU498277	KU221349			
= BRIP 55537							
VPRI 32575	<i>Salix fragilis</i> (leaf lesion)	NSW, Tumut River	KU498276			KU221379	
= BRIP 55536							
VPRI 32736	<i>Salix purpurea</i> (leaf lesion)	Tas, Blackfish Creek	KU498278	KU221350		KU221380	
= BRIP 55538							
VPRI 41498	<i>Sansevieria</i> sp. (anthracnose)	Vic, Cranbourne					
BRIP 57967b	<i>Artocarpus heterophyllus</i> (endophyte)	NT, Middle Point				KF877321	
BRIP 57977	<i>Artocarpus serotocarpus</i> (endophyte)	NT, Bees Creek				KF877322	
BRIP 57970	<i>Coffea arabica</i> (leaf lesion)	NT, Bees Creek				KF877323	
BRIP 57963	<i>Coffea canephora</i> (endophyte)	NT, Middle Point				KF877324	

Table 2 (continued)

Species	Culture ^{a,b}	Host (symptoms)	Locality (State ^c , city/town)	GenBank Accessions			
				ITS	GAPDH	GS	TUB
	DAR 28343c = BRIP 61797	<i>Dionaea muscipula</i> (leaf blight)	NSW, Sydney	KU498283			KU221383
	BRIP 57980	<i>Eriobotrya japonica</i> (endophyte)	NT, Bees Creek				KF877327
	BRIP 57979	<i>Ficus carica</i> (endophyte)	NT, Bees Creek				KF877328
	F272	<i>Hibiscus</i> sp.	unknown	KC425649			KC425703
	BRIP 57975	<i>Mentha</i> sp. (leaf lesion)	NT, Bees Creek				KF877329
	F28	<i>Olea europaea</i>	unknown	KC425659			KC425702
	ICMP 12567 = BRIP 19772	<i>Persea americana</i> (fruit rot)	NSW, Murwillumbah	JX010250	JX00940	JX10076	JX010387
	ICMP 12565 = BRIP 28812	<i>Persea americana</i> (anthracnose)	Qld, Mt Tamborine	JX010249	JX009937		
	BRIP 57976	<i>Piper nigrum</i> (leaf lesion)	NT, Bees Creek				KF877330
	ICMP 18574 = DAR 76934	<i>Pistacia vera</i>	NSW, Muswellbrook	JX010270	JX010002		JX010391
	BRIP 57978	<i>Rosmarinus officinalis</i> (endophyte)	NT, Bees Creek				KF877331
	BRIP 57966	<i>Theobroma cacao</i> (endophyte)	NT, Middle Point				KF877318
	BRIP 24197	<i>Actinidia chinensis</i> (stem end rot)	Qld, Mt Tamborine	GU183323			GU183282
	BRIP 28487	<i>Averrhoa carambola</i> (fruit rot)	Qld, unknown	GU183328			GU183286
	VPRI 13891a = BRIP 54772	<i>Calothammus quadrifidus</i> (stem lesion)	Vic, Park Orchards				KU221386
	BRIP 11084 = CBS 294.67	<i>Carica papaya</i> (fruit rot)	Qld, Brisbane	JQ948277	JQ948607		JQ949928
	BRIP 28519 ^{HT} = CBS 122122	<i>Carica papaya</i> (anthracnose)	Qld, Yandina	GU183331	JQ948606		GU183289
	= ICMP 17298						
	BRIP 28420	<i>Cyphomandra betacea</i> (fruit rot)	Qld, Mt Tamborine	GU183327			GU183285
	BRIP 4704	<i>Fragaria x ananassa</i> (fruit rot)	Qld, Forest Glen	GU183317			GU183277
	BRIP 11086 = CBS 295.67		Qld, Nambour	GU183318	JQ948608		GU183278
	BRIP 4703		Qld, Townsville	GU183316			GU183276
	VPRI 10360 = BRIP 52654		Vic, Scoresby	GU183348			GU183301
	VPRI 10487 = BRIP 52655		Vic, Silvan	GU183351			GU183302
	WAC 2768		WA, Wanneroo	JQ948280	JQ948610		JQ949931
	= IMI 354381		unknown				KU221387
	VPRI 10449	<i>Fragaria x ananassa</i> (leaf spot)					
	= BRIP 54813						
	= CBS 132311						
	WAC 7989		WA, Baldiwin	GU183359			GU183306
	= BRIP 52694						
	BRIP 39473	<i>Litchi chinensis</i> (fruit pepper spot)	NSW, Byron Bay	GU183337			GU183292
	BRIP 24243	<i>Litchi chinensis</i> (anthracnose)	Qld, Atherton	GU183324			GU183283
	BRIP 48724		Qld, Mena Creek	GU183338			GU183263
	BRIP 60282	<i>Litchi chinensis</i> (endophyte)	Qld, Mareeba	KU498285			KU221388
	VPRI 15973 = BRIP 54768	<i>Mandevilla</i> sp. (leaf spot)	Vic, Park Orchards				KU221385
	BRIP 28832	<i>Mangifera indica</i> (fruit rot)	Qld, Ayr	GU183334			GU183291

Colletotrichum simmondsii

Table 2 (continued)

Species	Culture ^{a,b}	Host (symptoms)	Locality (State ^c , city/town)	GenBank Accessions				
				ITS	GAPDH	GS	TUB	
	BRIP 28533	<i>Mangifera indica</i> (anthracnose)	Unknown	GU183332				GU183290
	BRIP 24124	<i>Nephelium lappaceum</i> (fruit rot)	Qld, Kamerunga	GU183321				GU183280
	BRIP 54619	<i>Prunus domestica</i> (shoot dieback)	Qld, Lake Eacham	KU498284				KU221384
	DAR 62104	<i>Solanum lycopersicum</i> (fruit spot & rot)	NSW, Comboyne	KU498286				
	= BRIP 61803							
	VPRI 11088		NSW, Tweed Heads	GU183352				GU183303
	= IMI 260260							
	= BRIP 52657							
	BRIP 62284	<i>Vaccinium corymbosum</i> (fruit rot)	NSW, Brooklet	KU498287				
	BRIP 62285		NSW, Corindi	KU498288				
	VPRI 15627		Vic, Knoxfield	GU183300				
	= BRIP 52651							
<i>Colletotrichum sloanei</i>	BRIP 48742	<i>Litchi chinensis</i> (anthracnose)	Qld, Mena Creek	KU498289	KU221342			KU221365
<i>Colletotrichum spinaciae</i>	VPRI 31994	<i>Spinacia oleracea</i> (leaf spot)	Vic, Cora Lynn	KU498290	KU221353			
	= BRIP 52700							
	= CBS 125444							
	VPRI 32005			KU498291	KU221354			
	= BRIP 52701							
	= CBS 125446							
<i>Colletotrichum spinosum</i>	DAR 48942 ^{HT}	<i>Xanthium spinosum</i> (stem lesion)	NSW, Coolah	KF178474	KF178498	KF178620		KF178595
	= CBS 515.97 ^{ex-HT}							
<i>Colletotrichum tanacetii</i>	BRIP 57314 ^{HT}	<i>Tanacetum cinerariifolium</i> (leaf lesion)	Tas, Scottsdale	JX218228	JX218243			JX218233
	= CBS 132693 ^{ex-HT}							
<i>Colletotrichum theobromicola</i>	DAR 41929	<i>Aeschynomene falcata</i> (leaf spot & stem blight)	NSW, Grafton					KU221389
	= BRIP 13654							
	DAR 25000 = BRIP 61796	<i>Aeschynomene falcata</i> (stem lesion)	NSW, Shannon Brook	KU498293				KU221391
	BRIP 57969	<i>Coffea arabica</i> (flower lesion)	NT, Bees Creek	KF877316				
	BRIP 57984	<i>Coffea arabica</i> (leaf lesion)	NT, Melville Island	KF877317				
	BRIP 46055	<i>Olea europaea</i> (anthracnose)	Qld, Wondai	KU498292				KU221390
	BRIP 58448	<i>Punica granatum</i> (fruit rot)	Qld, Tolga	JX010291	JX009948	JX010067		JX010381
	ICMP 17958	<i>Syzygium guianensis</i>	Qld, Samford					
	= CBS 124250							
	BRIP 35000 ^{HT,d}	<i>Syzygium viscosa</i>	Qld, Townsville	JX010289	JX009962	JX010063		JX010380
	= ICMP 17957 ^{IS}							
	= CBS 124251 ^{IS}							
<i>Colletotrichum tofieldiae</i>	BRIP 63314c	<i>Grevillea crithmifolia</i>	WA, Bullsbrook	KX069822				KX069813
	WAC 7298 = BRIP 63676	<i>Iris germanica</i>	WA, Mundaring	KX069824				KX069815
<i>Colletotrichum truncatum</i>	BRIP 12563	<i>Arachis hypogaea</i> (leaf spot)	Qld, Gurgena	AF451899				
	DAR 33949b	<i>Glycine max</i> (pod blight)	NSW, Grafton	KU498297				
	= BRIP 62368							
	DAR 44534		NSW, Herons Creek	AF451906				
	BRIP 55638b		unknown	KU498294				
	BRIP 55662b		unknown	KU498295				
	BRIP 55697	<i>Helianthus annuus</i>	Qld, Ryeford	KU498296				
	CBS 141.79	<i>Syzygium hamata</i>	Qld, Townsville	GU227873	GU228265			GU228167
	DAR 67500	<i>Xanthium occidentale</i>	NT, Mainoru Station	AF451907				

Table 2 (continued)

Species	Culture ^{a,b}	Host (symptoms)	Locality (State ^c , city/town)	GenBank Accessions			
				ITS	GAPDH	GS	TUB
<i>Colletotrichum xanthorrhoeae</i>	WAC 8358 ^{HT} = BRIP 45094 ^{ex-HT} = CBS 127831 ^{IS} = ICMP 17903 ^{IS} IMI 350817 = ICMP 17820	<i>Xanthorrhoea preissii</i> (leaf spot)	WA, Perth	GU048667	JX009927	JX010138	JX010448
		<i>Xanthorrhoea</i> sp. (leaf spot)	Qld, Cunningham's Gap	JX010260	JX010008		

^a BRIP, Queensland Plant Pathology Herbarium, Brisbane, Queensland; CBS, Centraalbureau voor Schimmelcultures, Fungal Biodiversity Centre, Utrecht, the Netherlands; DAR, NSW Plant Pathology Herbarium, Orange, NSW; ICMP, International Collection of Microorganisms from Plants, Auckland, New Zealand; IMI, Culture collection of CAB International, Wallingford, UK; PDD, New Zealand Fungal and Plant Disease Collection, New Zealand; UQ, University of Queensland, St Lucia, Queensland; VPRI, Victorian Plant Pathology Herbarium, Bundoora, Victoria; WAC, Western Australian Culture Collection, South Perth, Western Australia

^b EP, epitype; HT, holotype; IS, isotype; PT, paratype

^c ACT, Australian Capital Territory, NSW, New South Wales; NT, Northern Territory; Qld, Queensland; SA, South Australia; Tas, Tasmania; Vic, Victoria; WA, Western Australia

^d holotype of *Colletotrichum gloeosporioides* f. *stylosanthidis*

species in the *C. gloeosporioides* species complex using GS sequences. The Australian specimens of *C. alienum* were identified based on the 99–100 % identity of GS sequences to *C. alienum* ex-type strain ICMP 12071 (Table 2).

4. *Colletotrichum aotearoa* B. Weir & P.R. Johnst. Studies in Mycology 73: 139 (2012).

Colletotrichum aotearoa belongs to the *C. gloeosporioides* species complex (Weir et al. 2012). *Colletotrichum aotearoa* was first reported from Australia on *Banksia marginata* (Liu et al. 2013b) in a study of *Colletotrichum* species on Proteaceae. *Colletotrichum aotearoa* can be distinguished from other species in the *C. gloeosporioides* species complex with GAPDH, GS and/or TUB sequences (Weir et al. 2012). The Australian specimens of *C. aotearoa* were identified based on the 99–100 % identity of GS sequences to the *C. aotearoa* ex-type strain ICMP 18537 (Table 2).

5. *Colletotrichum asianum* Prihastuti, L. Cai & K.D. Hyde, Fungal Diversity 39: 96 (2009).

Colletotrichum asianum belongs to the *C. gloeosporioides* species complex (Weir et al. 2012). In Australia, *C. asianum* has only been isolated from mango, on which it is associated with a range of disease symptoms (Weir et al. 2012; Anderson et al. 2013 as *C. gloeosporioides*, James et al. 2014). *Colletotrichum asianum* can be distinguished from other species in the *C. gloeosporioides* species complex by any of the eight genes analysed in Weir et al. (2012). The Australian specimens of *C. asianum* were identified based on the 99–100 % identity of ITS, and/or TUB sequences to *C. asianum* ex-type strain MFLU 090234 (Table 2).

6. *Colletotrichum australe* Damm, P.F. Cannon & Crous, Studies in Mycology 73: 57 (2012b).

Colletotrichum australe belongs to the *C. acutatum* species complex, and is distinguishable by either GAPDH, histone 3, ITS or TUB sequences (Damm et al. 2012b).

7. *Colletotrichum axonopodi* J.A. Crouch, B.B. Clarke, J.F. White & B.I. Hillman, Mycologia 101: 727 (2009).

Colletotrichum axonopodi belongs to the *C. graminicola* species complex, which comprises predominantly grass infecting species with falcate spores (Cannon et al. 2012). *Colletotrichum axonopodi* can be identified by its unique ITS sequence and association with leaf spots on some species of *Axonopus* (Crouch et al. 2009a).

8. *Colletotrichum boninense* Moriwaki, Toy. Sato & Tsukib., Mycoscience 44: 48 (2003).

Damm et al. (2012a) resolved the *C. boninense* species complex with multilocus phylogenetic analyses, describing several new species. *Colletotrichum boninense* is best identified by ITS and GAPDH sequences (Damm et al. 2012a). Damm et al. (2012a) reported *C. boninense* from *Leucospermum* sp. in Australia without specimen collection details (CBS 112115). *Colletotrichum boninense* has been reported as a pathogen and an endophyte on a range of plant species worldwide (Hyde et al. 2009a).

9. ***Colletotrichum brevisporum*** Noireung, Phouliv., L. Cai & K.D. Hyde, *Cryptogamie Mycologie* 33: 350 (2012).

This is the first record of *C. brevisporum* in Australia. *Colletotrichum brevisporum* has been recorded as an endophyte as well as a pathogen on a range of host plant species in several tropical countries. *Colletotrichum brevisporum* can be identified based on sequence identity of ITS, GAPDH and/or TUB (Noireung et al. 2012). The Australian specimens of *C. brevisporum* have been identified based on the 100 % identity of ITS sequences to *C. brevisporum* ex-type strain BCC 38876 (Table 2).

10. ***Colletotrichum brisbanense*** Damm, P.F. Cannon & Crous, *Studies in Mycology* 73: 59 (2012b).

Colletotrichum brisbanense belongs to the *C. acutatum* species complex (Damm et al. 2012b). This species is only known from the type specimen. *Colletotrichum brisbanense* was described from one of Simmonds' (1968) paratype specimens of *C. acutatum*, which Shivas & Tan (2009) had assigned to *C. simmondsii*. GAPDH and TUB sequences clearly separate *C. brisbanense* and *C. simmondsii* (Damm et al. 2012b).

11. ***Colletotrichum chlorophyti*** S. Chandra & Tandon [as *chlorophytum*], *Current Science* 34: 565 (1965).

In North America, *C. chlorophyti* has been reported as the cause of anthracnose in soybean (*Glycine max*) (Yang et al. 2012). The only Australian specimen was deposited in the CBS culture collection as *C. dematium* (Table 2). A multilocus phylogenetic analysis re-identified it as *C. chlorophyti* (Damm et al. 2009). The extent to which *C. chlorophyti* is responsible for anthracnose on *Stylosanthes hamata* in Australia is not known.

12. ***Colletotrichum circinans*** (Berk.) Voglino, *Annali della Reale Accademia d'Agricoltura di Torino* 49: 175 (1907).

Colletotrichum circinans belongs to the *C. dematium* clade (Cannon et al. 2012). There are records of *C. circinans* in Australia from onion, shallot and leeks (*Allium* spp.) with smudge and bulb rot (Hall et al. 2009, Persley et al. 2010). The only Australian specimen has been identified based on the

100 % identity of ITS and GAPDH sequences to *C. circinans* ex-type strain CBS 221.81 (Table 2).

13. ***Colletotrichum coccodes*** (Wallr.) S. Hughes, *Canadian Journal of Botany* 36: 754 (1958).

Colletotrichum coccodes was recently neotypified (Liu et al. 2011), which stabilized the taxonomic concept of this species. Prior to this, *C. coccodes* was reported as the causal agent of brown root rot of tomato (Golzar 2009b) and black dot of potato (Ben-Daniel et al. 2010). Most isolates identified as *C. coccodes* in Australian culture collections require validation by molecular phylogenetic analyses. The Australian specimens have been identified based on the 100 % identity of ITS, GAPDH and/or TUB sequences to *C. coccodes* ex-type strain CBS 164.49 (Table 2).

14. ***Colletotrichum cymbidiicola*** Damm, P.F. Cannon, Crous, P.R. Johnst. & B. Weir, *Studies in Mycology* 73: 19 (2012a).

Colletotrichum cymbidiicola belongs to the *C. boninense* species complex (Damm et al. 2012a). *Colletotrichum cymbidiicola* causes anthracnose on *Cymbidium* spp. in Australia, India, Japan, New Zealand and USA (Damm et al. 2012a, Chowdappa et al. 2014, Bethke 2014), and is likely present in all countries where *Cymbidium* orchids are grown. The only Australian specimen (Table 2) is the holotype of *C. cymbidiicola* (Damm et al. 2012a).

15. ***Colletotrichum dematium*** (Pers.) Grove, *Journal of Botany, British and Foreign Legion, London* 56: 341 (1918).

Damm et al. (2009) designated an epitype for *C. dematium* derived from a culture isolated from a dead leaf of *Eryngium campestre* collected in France. *Colletotrichum dematium* has a wide host range with pathogenic, saprobic and endophytic strains (Damm et al. 2009). Although *C. dematium* has been claimed to cause several economically important diseases, this has rarely been demonstrated. The only Australian specimen (Table 2) was included in the study by Damm et al. (2009). Most isolates identified as *C. dematium* in Australian culture collections require validation by molecular phylogenetic analyses.

16. ***Colletotrichum destructivum*** O'Gara, *Mycologia* 7: 38 (1915).

Colletotrichum destructivum is a species complex that was recently revised by Damm et al. (2014) who designated an epitype for *C. destructivum*. *Colletotrichum destructivum* can be identified by its ITS and TUB2 sequences. In Australia, two isolates from pasture legumes

(Ford et al. 2004) have ITS sequences identical to those of the epitype.

17. *Colletotrichum dracaenophilum* D.F. Farr & M.E. Palm, Mycological Research 110: 1401 (2006).

Colletotrichum dracaenophilum is a stem pathogen of *Dracaena* spp. (Farr et al. 2006). *Colletotrichum dracaenophilum* occupies a small clade that is basal to the entire genus, apart from the combined *C. orbiculare* and *C. cliviae* clade (Cannon et al. 2012). The Australian records have been identified based on the 100 % identity of ITS and GAPDH sequences to *C. dracaenophilum* ex-type strain CBS 118199 (Table 2).

18. *Colletotrichum fioriniae* (Marcelino & S. Gouli) R.G. Shivas & Y.P. Tan, Fungal Diversity 39: 117 (2009).

Colletotrichum fioriniae belongs to the *C. acutatum* species complex. *Colletotrichum fioriniae* was originally described as an entomopathogen from a scale insect in the USA, where it has also been reported as an endophyte in several plants (Marcelino et al. 2008). In Australia, *C. fioriniae* has been reported as the cause of leaf and stem blight on *Acacia acuminata* (Golzar 2009a as *C. acutatum*) and fruit rot of avocado (Shivas and Tan 2009). *Colletotrichum fioriniae* is readily identified by any of the six genes analysed by Damm et al. (2012b).

19. *Colletotrichum fructicola* Prihastuti, L.Cai & K.D. Hyde, Fungal Diversity 39: 158 (2009).

Colletotrichum fructicola belongs to the *C. gloeosporioides* species complex (Weir et al. 2012). Little is known about the host range and pathogenicity of this species in Australia. *Colletotrichum fructicola* can be distinguished from other species in the *C. gloeosporioides* species complex with GS sequences (Weir et al. 2012).

20. *Colletotrichum gloeosporioides* (Penz.) Penz. & Sacc., Atti del Reale Istituto Veneto di Scienze, Lettere ed Arti, Serie 6, 2: 670 (1884).

Colletotrichum gloeosporioides is a name well known to plant pathologists. A major revision of *Colletotrichum* by von Arx (1957) based on morphology rather than host association, reduced the number of accepted species from 750 to 11, with the vast majority reduced to synonymy with *C. gloeosporioides*. This left *C. gloeosporioides* as a biologically and genetically diverse species associated with at least 470 different host genera (Sutton 1980). Cannon et al. (2008) epitypified *C. gloeosporioides* with a specimen and living culture from *Citrus sinensis* collected from Italy. In a major revision, Weir et al. (2012) used multilocus sequence analyses

to resolve the *C. gloeosporioides* species complex into a number of segregate species. The species that gives its name to this complex, *C. gloeosporioides*, was found to be commonly associated with *Citrus*, although it also occurred on other host species (Weir et al. 2012).

Although *C. gloeosporioides* has been implicated as an important pathogen of a wide range of plants in Australia, there are relatively few published records that can be verified by DNA sequence data. The situation is similar for most applications of the name *C. gloeosporioides* in much of the plant pathology literature worldwide. The extent of the problem was highlighted by Cai et al. (2009), who estimated that more than 86 % of the records of *C. gloeosporioides* in GenBank had sequences that diverged considerably from the epitype and were likely to represent other species. Further, Phoulivong et al. (2010) considered that *C. gloeosporioides* was not a common pathogen on tropical fruits. In Australia, *C. gloeosporioides* has been recorded from cultivated *Citrus* spp. (Schena et al. 2014) and pecan (Weir et al. 2012). *Colletotrichum gloeosporioides* can be distinguished from other species in the *C. gloeosporioides* species complex by any of the eight genes analysed in Weir et al. (2012). The Australian records of *C. gloeosporioides* have been identified based on the 100 % identity of ITS, GAPDH and/or TUB sequences to the *C. gloeosporioides* ex-type strain IMI 356878 (Table 2).

21. *Colletotrichum godetiae* Neerg., Friesia 4: 72 (1950).

Colletotrichum godetiae belongs to the *C. acutatum* species complex (Damm et al. 2012b). *Colletotrichum godetiae* is recorded as a pathogen of fruit, leaves and stems of *Fragaria*, *Malus* and *Prunus*, mainly in Europe and Western Asia (Damm et al. 2012b). These are new records for Australia. *Colletotrichum godetiae* can be distinguished from other species in the *C. acutatum* species complex by any of the six genes analysed in Damm et al. (2012b). The Australian records of *C. godetiae* have been identified based on the 100 % identity of ITS and/or TUB sequences to *C. godetiae* ex-type strain CBS 133.44 (Table 2).

22. *Colletotrichum kahawae* subsp. *ciggaro* B. Weir & P.R. Johnst., Studies in Mycology 73: 158 (2012).

Colletotrichum kahawae subsp. *ciggaro* belongs to the *C. gloeosporioides* species complex (Weir et al. 2012). *Colletotrichum kahawae* subsp. *ciggaro* has a broad host range and worldwide distribution. It differs only in one gene, glutamine synthetase, from the African coffee berry disease pathogen, *C. kahawae* subsp. *kahawae*, which is not present in Australia (Weir et al. 2012). The only Australian record listed in Table 2 is the ex-type culture of *C. kahawae* subsp. *ciggaro* (Weir et al. 2012).

23. *Colletotrichum karsti* You L. Yang, Zuo Y. Liu, K.D. Hyde & L. Cai [as 'karstii'], Cryptogamie Mycologie 32: 241 (2011).

Colletotrichum karsti belongs to the *C. boninense* species complex (Damm et al. 2012a). Due to the high variation observed in morphology and sequences, a polyphasic approach is recommended to ensure accurate identification (Damm et al. 2012a). In Australia, *C. karsti* has a wide host range, although its role as a pathogen is not known.

24. *Colletotrichum lupini* (Bondar) Damm, P.F. Cannon & Crous, Studies in Mycology 73: 78 (2012).

Colletotrichum lupini belongs to the *C. acutatum* species complex (Damm et al. 2012b). *Colletotrichum lupini* is an economically important pathogen of lupin crops worldwide, including Australia, where it was first detected in Western Australia in 1994 (Sweetingham et al. 1995, Yang and Sweetingham 1998, as *C. gloeosporioides*). *Colletotrichum lupini* has established in Western Australia, where it has spread through wild populations of blue lupins (*Lupinus cosentinii*), as well as in parts of South Australia, where it is known to occur in *Lupinus albus*. *Colletotrichum lupini* is not known to occur in the lupin growing regions in New South Wales (Anon. 2015) or Victoria (Thomas 2010). *Colletotrichum lupini* can be distinguished from other species in the *C. acutatum* species complex by most of the six genes (excluding actin) analysed by Damm et al. (2012b), with TUB providing the best resolution. The Australian records of *C. lupini* have been identified based on the 100 % identity of ITS, GAPDH and/or TUB sequences to *C. lupini* ex-type strain CBS 109225 (Table 2).

25. *Colletotrichum musae* (Berk. & M.A. Curtis) Arx, Verhandelingen Koninklijke Nederlandse Akademie van Wetenschappen, Sect. 51: 107 (1957).

Colletotrichum musae belongs to the *C. gloeosporioides* species complex (Weir et al. 2012). Su et al. (2011) designated an epitype for *C. musae* from banana (*Musa* sp.) fruit in the USA, collected from the geographic locality of the type. In Australia, *C. musae* has been associated with several banana diseases, including anthracnose, fruit speckle, black end and crown rot (Cooke et al. 2009). However, many of these records require verification by molecular methods. *Colletotrichum musae* can be distinguished from other species in the *C. gloeosporioides* species complex by any of eight genes analysed by Weir et al. (2012). The Australian records of *C. musae* have been identified based on the 100 % identity of ITS, GAPDH and/or TUB sequences to *C. musae* ex-type strain CBS 116870 (Table 2).

26. *Colletotrichum nymphaeae* (Pass.) Aa, Netherlands Journal of Plant Pathology, Supplement 1 84: 110 (1978).

Colletotrichum nymphaeae belongs to the *C. acutatum* species complex (Damm et al. 2012b). *Colletotrichum nymphaeae* was determined as the causal agent of celery stunt disease in Japan (Yamagishi et al. 2015). This raises the possibility that celery leaf curl disease in Queensland, which was attributed to *C. acutatum sensu lato* (Heaton and Dullahide 1993), may be caused by *C. nymphaeae*. *Colletotrichum nymphaeae* is easily distinguished from other species in the *C. acutatum* species complex with TUB (Damm et al. 2012b). The Australian records of *C. nymphaeae* have been identified based on the 100 % identity of TUB sequences to *C. nymphaeae* ex-type strain CBS 515.78 (Table 2).

27. *Colletotrichum ocimi* Damm, Studies in Mycology 79: 70 (2014).

Colletotrichum ocimi belongs to the *C. destructivum* species complex (Damm et al. 2014). Persley et al. (2010) reported that black spot of basil, caused by *C. ocimi* (as *C. gloeosporioides*), is a minor disease in Australia. *Colletotrichum ocimi* is easily distinguished from other species in the *C. acutatum* species complex by its unique ITS and TUB sequences (Damm et al. 2014). The Australian record of *C. ocimi* was identified based on the 100 % identity of ITS sequence to *C. ocimi* ex-type strain CBS 298.94 (Table 2).

28. *Colletotrichum orbiculare* Damm, P.F. Cannon & Crous, Fungal Diversity 61: 39 (2013).

Colletotrichum orbiculare sensu Damm et al. (2013) was recently described and pragmatically given the same name as the earlier invalid name *C. orbiculare sensu* von Arx (1957), which is well known in the plant pathology literature worldwide as a pathogen of host plants in the Cucurbitaceae. In Australia, *C. orbiculare sensu lato* has been associated with diseases on several plant species in the Asteraceae, Cucurbitaceae and Fabaceae (Auld et al. 1994 as *C. lagenarium*, Simmonds 1965, Walker et al. 1991, Persley et al. 2010). However, most of these records require verification by molecular methods. The only verified Australian specimen (Table 2) was included in the study by Damm et al. (2013)

29. *Colletotrichum petchii* Damm, P.F. Cannon & Crous, Studies in Mycology 73: 29 (2012a).

Colletotrichum petchii belongs to the *C. boninense* species complex (Damm et al. 2012a). This fungus appears to be host

specific to *Dracaena* spp., which are native to Africa (Damm et al. 2012a). The Australian record of *C. petchii* was identified based on the 100 % identity of ITS and GAPDH sequences to *C. petchii* ex-type strain CBS 378.94 (Table 2).

30. *Colletotrichum phormii* (Henn.) D.F. Farr & Rossmann, Mycological Research 110: 1403 (2006).

Colletotrichum phormii belongs to the *C. acutatum* complex (Damm et al. 2012b). *Colletotrichum phormii* was first reported in Australia in 2010 from Perth, WA, based on the ITS sequence of isolate WAC 12416 (Golzar and Wang 2010). However, this record cannot be verified as there is no living culture available and the sequence data was not deposited in GenBank. *Colletotrichum phormii* is separated from other species in the *C. acutatum* species complex by sequences of secondary genes, namely TUB, GAPDH, histone and actin (Damm et al. 2012b). The Australian record of *C. phormii* was identified based on the 100 % identity of TUB sequences to *C. phormii* ex-type strain CBS 118194 (Table 2).

31. *Colletotrichum pyricola* Damm, P.F. Cannon & Crous, Studies in Mycology 73: 94 (2012b).

Colletotrichum pyricola belongs to the *C. acutatum* species complex (Damm et al. 2012b). This fungus was first described from fruit rot of pear (*Pyrus communis*) in New Zealand. *Colletotrichum pyricola* can be distinguished from other species in the *C. acutatum* species complex with GAPDH and TUB (Damm et al. 2012b). The Australian record of *C. pyricola* was identified based on the 100 % identity of GAPDH and TUB sequences to *C. pyricola* ex-type strain CBS 128531 (Table 2).

32. *Colletotrichum queenslandicum* B. Weir & P.R. Johnst., Studies in Mycology 73: 164 (2012).

Colletotrichum queenslandicum belongs to the *C. gloeosporioides* species complex (Weir et al. 2012). This fungus was first described from Queensland as *C. gloeosporioides* var. *minor* Simmonds (1968), who regarded this fungus as an important cause of fruit rot in avocado and papaya, as well as a wide range of other hosts (Simmonds 1965, 1966). A new name, *C. queenslandicum*, was given to this taxon as the orthographically correct species epithet *minus* was already occupied by *C. minus* Zimm. In Australia, *C. queenslandicum* has been reported from avocado (Weir et al. 2012), lychee (Anderson et al. 2013 as *C. gloeosporioides*), passionfruit (James et al. 2014) and pawpaw (Weir et al. 2012). *Colletotrichum queenslandicum* is best distinguished from other species in the *C. gloeosporioides* species complex by GAPDH, GS or TUB sequences (Weir et al. 2012). The Australian specimens of *C. queenslandicum*

were identified based on the 100 % identity of GAPDH and/or TUB sequences to *C. queenslandicum* ex-type strain ICMP 1778 (Table 2).

33. *Colletotrichum salicis* (Fuckel) Damm, P.F. Cannon & Crous, Studies in Mycology 73: 97 (2012b).

Colletotrichum salicis belongs to the *C. acutatum* species complex (Damm et al. 2012b). This fungus was first reported on *Salix* spp. from mainland Australia as *Glomerella miyabeana* (Cunnington et al. 2007), which is considered a synonym of *C. salicis* by Damm et al. (2012b). *Colletotrichum salicis* is best distinguished from other species in the *C. acutatum* species complex with GAPDH or TUB sequences (Damm et al. 2012b). The Australian specimens of *C. salicis* were identified based on the 99–100 % identity of GAPDH and/or TUB sequences to *C. salicis* ex-type strain CBS 607.94 (Table 2).

34. *Colletotrichum sansevieriae* Miho Nakam. & Ohzono, Journal of General Plant Pathology 72: 253 (2006).

Colletotrichum sansevieriae causes leaf anthracnose on *Sansevieria* spp., which are native to Africa and Asia. The only Australian specimen (Table 2) was collected in 2008 (Aldaoud et al. 2011).

35. *Colletotrichum siamense* Prihastuti, L. Cai & K.D. Hyde, Fungal Diversity 39: 98 (2009).

Colletotrichum siamense belongs to the *C. gloeosporioides* species complex (Weir et al. 2012). In Australia, *C. siamense* has a diverse host range (Weir et al. 2012, James et al. 2014, Schena et al. 2014), although its role as a pathogen is unclear. *Colletotrichum siamense* is best distinguished from other species in the *C. gloeosporioides* species complex with TUB (Weir et al. 2012). The Australian specimens of *C. siamense* were identified based on the 100 % identity of TUB sequences to *C. siamense* ex-type strain ICMP 18578 (Table 2).

36. *Colletotrichum simmondsii* R.G. Shivas & Y.P. Tan, Fungal Diversity 39: 119 (2009).

Colletotrichum simmondsii belongs to the *C. acutatum* species complex (Damm et al. 2014). In Australia, *C. simmondsii* has been associated with fruit rots of papaya, strawberry, tomato and blueberry (Shivas and Tan, 2009). *Colletotrichum simmondsii* is easily distinguished from other species in the *C. acutatum* species complex with ITS or TUB sequences (Damm et al. 2012b). The Australian specimens of *C. simmondsii* were identified based on the 100 % identity of ITS and/or TUB to *C. simmondsii* ex-type strain BRIP 28519 (Table 2).

37. *Colletotrichum sloanei* Damm, P.F. Cannon & Crous., *Studies in Mycology* 73: 103 (2012b).

Colletotrichum sloanei belongs to the *C. acutatum* species complex (Damm et al. 2012b). Little is known about the host range and pathogenicity of this species. *Colletotrichum sloanei* is best distinguished from other species in the *C. acutatum* species complex with GAPDH or TUB sequences (Damm et al. 2012b). The Australian specimens of *C. sloanei* were identified based on the 100 % identity of GAPDH and TUB sequences to *C. sloanei* ex-type strain IMI 364297 (Table 2).

38. *Colletotrichum spinaciae* Ellis & Halst., *Journal of Mycology* 6: 34 (1890).

Colletotrichum spinaciae belongs to the *C. dematium* clade (Cannon et al. 2012). In Victoria, *C. spinaciae* (as *C. dematium*) was shown to cause anthracnose on spinach when it was first detected in 2004 (Washington et al. 2006). This detection invoked a biosecurity response of interstate surveys, which found that anthracnose on spinach was widespread in eastern Australia, resulting in no further quarantine action (Washington et al. 2006). The Australian specimens of *C. spinaciae* were identified based on the 100 % identity of ITS and GAPDH sequences to *C. spinaciae* ex-epitype strain CBS 128.57 (Table 2).

39. *Colletotrichum spinosum* Damm & P.F. Cannon, *Fungal Diversity* 61:46 (2013).

Colletotrichum spinosum belongs to the *C. orbiculare* species complex (Damm et al. 2013). *Colletotrichum spinosum* is a common pathogen of *Xanthium spinosum* in eastern Australia, where it caused seedling blight and stem anthracnose (Veitch 1942, Butler 1951, Anderson and Walker 1962, Simmonds 1965, 1966, Walker et al. 1991). In pathogenicity tests, the ex-holotype isolate was found to be highly virulent on *Xanthium* spp. as well as some other Asteraceae, and further infected some species of Cucurbitaceae, Fabaceae and Myrtaceae (Walker et al. 1991). It has been evaluated as a mycoherbicide for the biological control of *Xanthium spinosum* in Australia (Auld et al. 1988, 1990, Auld & Say 1999). *Colletotrichum spinosum* can be easily identified based on GAPDH, GS and/or TUB2 sequences (Damm et al. 2013).

40. *Colletotrichum tanacetii* M. Barimani et al., *Plant Pathology* 62: 1252 (2013).

Colletotrichum tanacetii belongs to the *C. destructivum* species complex (Damm et al. 2014). *Colletotrichum tanacetii* is a serious pathogen of pyrethrum in northern Tasmania (Barimani et al. 2013). The only Australian specimen

(Table 2) is the holotype of *C. tanacetii*, which was distinguished from other *Colletotrichum* spp. by GAPDH, ITS and TUB sequences (Barimani et al. 2013)

41. *Colletotrichum theobromicola* Delacr., *Bull. Soc. Mycol. France* 31: 191 (1905).

Colletotrichum theobromicola belongs to the *C. gloeosporioides* species complex (Weir et al. 2012). *Colletotrichum theobromicola* has been reported on several diverse host species in Australia (James et al. 2014, Schena et al. 2014). The Type A and Type B isolates of *C. gloeosporioides* f. *stylosanthis* were shown to be pathogenic on *Stylosanthes* in northern Australia (Irwin & Cameron 1978). These isolates were subsequently named *C. gloeosporioides* f. *stylosanthis* “f. sp. *guianensis*” and *C. gloeosporioides* f. *stylosanthis* “f. sp. *stylosanthis*”, respectively, by Munaut et al. (2002). Weir et al. (2012) considered these names synonyms of *C. theobromicola*. *Colletotrichum theobromicola* can be distinguished from other species in the *C. gloeosporioides* species complex by any of the eight genes analysed in Weir et al. (2012). The Australian specimens of *C. theobromicola* have been identified based on the 100 % identity of ITS and/or TUB sequences to *C. theobromicola* ex-type strain CBS 124945 (Table 2).

42. *Colletotrichum tofieldiae* (Pat.) Damm, P.F. Cannon & Crous, *Fungal Diversity* 39: 77 (2009).

Colletotrichum tofieldiae was previously treated as a variant of *C. dematium* var. *minus*, but was confirmed as a distinct species after an epitype was designated (Damm et al. 2009). *Colletotrichum tofieldiae* has been reported as an endophyte (Tao et al. 2013, Hiruma et al. 2016). Its role as a pathogen is unknown. The Australian specimens of *C. tofieldiae* were identified based on the 100 % identity of ITS and TUB sequences to *C. tofieldiae* ex-type strain CBS 495.85 (Table 2).

43. *Colletotrichum truncatum* (Schwein.) Andrus & W.D. Moore, *Phytopathology* 25: 122 (1935).

In Australia, most records of this fungus (e.g. Ford et al. 2004) still require verification by molecular methods. Worldwide, *C. truncatum* causes anthracnose diseases of many hosts in the Solanaceae and Fabaceae (Damm et al. 2009). An epitype from *Phaseolus lunatus* in the USA was designated by Damm et al. (2009), who used multilocus phylogenetic analyses to show that *C. capsici* and *C. curvatum* were synonyms of *C. truncatum*. The Australian specimens of *C. truncatum* were identified based on the 100 % identity of ITS sequences to *C. truncatum* ex-type strain CBS 151.35 (Table 2).

44. *Colletotrichum xanthorrhoeae* R.G. Shivas, Bathgate & Podger, Mycological Research 102: 280 (1998).

Colletotrichum xanthorrhoeae belongs to the *C. gloeosporioides* species complex (Weir et al. 2012). *Colletotrichum xanthorrhoeae* was widespread in Western Australia, where it caused a leaf spot disease on *Xanthorrhoea* spp. (Shivas et al. 1998). *Colletotrichum xanthorrhoeae* can be easily identified by its distinctive morphology, very slow growth rate in culture, and its unique ITS sequence (Weir et al. 2012).

Discussion

The species listed above have been verified by molecular phylogenetic analyses. There are several other names of *Colletotrichum* species that have been either (i) reported in the scientific literature as occurring in Australia, or (ii) listed (often unpublished) in culture collections of Australian isolates, which have yet to be verified by DNA sequence analyses. Some omissions from the Australian list of verified names are worthy of comment, in particular, *C. lindemuthianum* (Sacc. & Magnnus) Briosi & Cavara, *C. graminicola* (Ces.) G.W. Wilson and *C. sublineola* Henn. ex Sacc. & Trotter. *Colletotrichum lindemuthianum*, belongs to the *C. orbiculare* species complex (Damm et al. 2013) and is the causal agent of anthracnose of common bean (*Phaseolus vulgaris*) (Cruickshank 1966), although it has been practically eradicated from commercial production in Australia (Persley et al. 2010). The name *C. lindemuthianum* was only recently stabilised with the designation of an epitype (Liu et al. 2013a). Liu et al. (2013a) also noted that very few studies of *C. lindemuthianum* incorporated DNA sequence analysis, which is the case in Australia and explains its absence from our list. *Colletotrichum graminicola* and *C. sublineola* belong to a monophyletic clade of species with falcate conidia that are mostly host specific on grasses (Crouch et al. 2009a, b, Cannon et al. 2012). Worldwide, *C. graminicola* and *C. sublineola* are important pathogens of maize (*Zea mays*) and corn (*Sorghum* spp.), respectively (Crouch and Beirn 2009), yet rarely important in Australia (Simmonds 1966, Plant Health Australia (2001) Australian Plant Pest Database, online database <http://appd.ala.org.au/> accessed 2 Sep. 2016). The graminicolous *Colletotrichum* isolates in Australian collections have yet, for the most part, to be identified and classified by molecular phylogenetic methods.

Other Australian species that remain to be verified by DNA sequence analyses include *C. acaciae* Gutner, *C. caudatum* (Peck ex Sacc.) Peck, *C. crassipes* (Speg.) Arx, *C. falcatum* Went, *C. fuscum* Laubert, *C. higginsianum* Sacc., *C. malvarum* (A. Braun & Casp.) Southw., *C. orchidearum* Allesch., *C. schizanthi* C.N. Jensen & V.B. Stewart,

C. trichellum (Fr.) Duke, *C. trifolii* Bain, and *C. xanthii* Halst. These species require verification by DNA sequencing to confirm their presence in Australia.

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