### **REVIEW**



# Krüppel-like factor 17, a novel tumor suppressor: its low expression is involved in cancer metastasis

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**Abstract** Krüppel-like factor (KLF) family is highly conserved zinc finger transcription factors that regulate cell proliferation, differentiation, apoptosis, and migration. KLF17 is a member of the KLF family. Recent studies have demonstrated that KLF17 low expression and inactivation are caused by microRNA, gene mutation, and loss of heterozygosity in human tumors, which participates in tumor progression. KLF17 low expression increases cancer metastatic viability; its mechanism is that low KLF17 mediates epithelial-mesenchymal transition (EMT) through regulating EMT-related genes expression; the reduced-KLF17 also increases cancer metastasis though upregulating inhibitor of DNA binding 1 (ID1). Additionally, mutant p53 proteins are capable of developing a complex with KLF17, which mediate the depletion of KLF17 inhibiting EMT gene transcription and increases cancer metastasis. KLF17 downregulation also mediates the activation of TGF-\beta pathway.

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**Keywords** Krüppel-like factor  $17 \cdot \text{Cancer metastasis} \cdot \text{Epithelial-mesenchymal transition} \cdot \text{TGF-}\beta \text{ pathway} \cdot \text{p53}$  pathway

### Introduction

Krüppel-like factor (KLF) family is highly conserved zinc finger transcription factors, which are critical regulator of essential biological cellular processes, including proliferation, differentiation, apoptosis, and migration [1–3]. Structurally, the C-terminal region of the KLF family members is highly conserved, which is composed of triple tandem zinc fingers evenly spaced by conserved linker regions, while the Nterminal regions are highly divergent [4]. Accordingly, KLF members can recognize similar target sequences, while their N-terminus can bind to different factors leading to diverse functions [2, 4]. To date, 17 members of the KLF protein family, KLF1-17, have been described in mammals, and an increasing number of studies have demonstrated that KLF 1-17 are involved in the pathobiology of tumor progression [1, 5, 6]. KLF17, also known as zinc finger protein 393 (Zfp393), was first identified as a germ cell-specific gene in mouse [7]. van Vliet and colleagues [8] identified and renamed the hypothetical protein FLJ40160 as KLF17; KLF17 is the human homologue of murine Zfp393, and KLF17 gene is mapped to chromosome 1p34.1. They reported that KLF17 was a novel member of the Sp/KLF family of transcription factors and was more closely related to the KLF subfamily. Sharing similarity with *Drosophila* Krüppel gene, human Sp/KLF family is characterized by a triple-C2H2 DNA-binding domain [9, 10]. Recently, many reports have focused on KLF17 functions in tumorigenesis and found that KLF17 plays an important role in cancer development. In the present study, we summarized KLF17's function in cancer process and its mechanism.

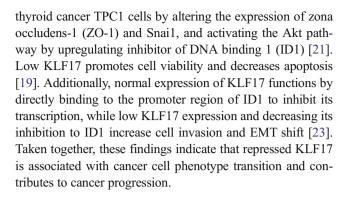


KLF17 is downregulated and correlated with tumor progression in various human cancers. Recent studies have demonstrated that low KLF17 is involved in transforming growth factor  $\beta$  (TGF- $\beta$ ) pathway and p53 pathway in human cancer and regulates epithelial-mesenchymal transition (EMT) and participates in metastases.

# The low expression of KLF17 is involved in tumor process

KLF17 lowly expresses in human tumors KLFs are a family containing highly conserved zinc finger transcription factors, which contains 17 members KLF1-17 in. KLF family genes are mapped to chromosome 1p34.1. The short arm of human chromosome 1 is one of the most studied genomic intervals in human cancer; allelic deletions in the 1p36 and 1p32 regions correlate with poor survival [11]. KLF6 gene is mutated in a subset of human prostate cancer and involved in human prostate cancer [12]; it is also inactivated by loss of heterozygosity (LOH) [13]. Additionally, Evi-1 oncoprotein binds to the zinc finger gene and regulates KLFs' gene expression [14]. HBx also binds to the zinc finger transcription factor and inactivates KLF gene expression in Escherichia coli [15]. KLF17 deficiency in tumors may also be from gene mutation and oncoprotein or virus protein inactivating. KLF17 has a transactivation activity both in embryonic chickens and humans [8, 16]. It is demonstrated that KLF17 is frequently downexpressed in the majority of human cancers, including breast cancer, lung adenocarcinoma, hepatocellular carcinoma (HCC), gastric cancer, papillary thyroid carcinoma (PTC), and non-small cell lung cancer (NSCLC) [17-22]. The expression level of KLF17 in lung adenocarcinoma cells and primary tumor tissues was lower than in immortal human bronchial epithelial cells and tumor-adjacent lung tissues, respectively [17]. The survival rate is higher in the high KLF17 expression group than in the low KLF17 expression group of patients with HCC, and the downregulated KLF17 expression is associated with the poor prognosis of HCC [18]. Peng and colleagues [20] reported that the expression level of KLF17 was significantly decreased in 98 of 158 gastric adenocarcinoma cases. Expression of KLF17 is also decreased in PTC tissues compared with the adjacent normal tissues [21].

Low expression of KLF17 contributes to cancer cell phenotype The forced expression of KLF17 leads to the inhibition of cell growth [23]. Silencing of KLF17 increases the transcription of CD44, plasminogen activator inhibitor 1 (PAI-1), and Cyclin-D1, while overexpression of KLF17 decreases the transcription of these genes [19]. Moreover, overexpression of KLF17 leads to cellular morphological changes and inhibits cell invasion significantly [24]. The repressed KLF17 promotes the motility and proliferation of human



KLF17 expression predicts survival and is associated with tumor progression The reduced expression of KLF17 is an independent prognostic indicator of the majority of human tumors, and it is significantly associated with tumor progression. Low expression of KLF17 is also an independent predictor of lymph node metastasis in breast cancer [23]. The clinical studies showed that low KLF17 is associated with a reduced survival time in lung adenocarcinoma patients, and the distant tumor metastasis is significantly increased [17]. KLF17 expression level is an independent prognostic indicator, and it is correlated with the tumor stage and size in lung adenocarcinoma and HCC [17, 18]. It has been identified that reduced expression of KLF17 is strongly associated with tumor size, pN stage, and lymphovascular invasion in gastric adenocarcinoma [20]. Moreover, KLF17 expression is an independent prognostic factor for both overall survival and disease-free survival in gastric adenocarcinoma [20]. KLF17 expression is correlated with clinical-pathological parameters and affects the prognosis of PTC patients [21].

### KLF17 is involved in cancer metastasis

KLF17 low expression in the metastatic cancer The first research on KLF17 in breast cancer conducted by Gumireddy et al. [23] in 2009 reported that lower expression of KLF17 was involved in breast cancer metastasis; lower expression of KLF17 was found in breast cancer cell lines with an invasive phenotype, and lower expression of KLF17 was found in the patients with lymph node metastases compared in the patients without metastases. Repressed KLF17 is also found in metastatic HCC [18]; KLF17 is posttranscriptionally inhibited by microRNA-9 (miR-9) in HCC and implicated in miR-9-mediated HCC metastasis [25]. Low KLF17 expression is significantly associated with metastasis in lung adenocarcinoma, gastric cancer, PTC, and NSCLC [17, 20–22]. Gastric cancer studies showed that the reduced expression of KLF17 protein is correlated with its lymphovascular invasion [20]. In lung adenocarcinoma, low expression of KLF17 is also related to tumor growth and poor prognosis [17]. The downregulation of KLF17 may play a role



in initiation and/or progression as well as the metastasis of esophageal squamous cancer [26]. These data indicate that KLF17 lowly expresses in metastatic tumor.

KLF17 low expression increases cancer metastatic viabil-

ity Cancer metastasis is a complex and multistep process, which consists of a series of discrete biological processes including malignant cell spread from the primary tumor to distant foci and subsequently adaptation to distant tissue environments [27-29]. Gumireddy et al. reported that the silence of KLF17 increased cell viability in metastatic breast cancer [23]. Further studies elucidated that the low expression of KLF17 was associated with carcinoma progression, and suppression of KLF17 expression promotes tumor cell migration, invasion, and EMT shift [19, 23, 30]. KLF17 exerts its tumor suppressor function by interacting with the promoters of EMT-related genes; it was at first identified as a novel tumor suppressor from a forward genetic screen in a mouse model [23]. In recent years, a growing number of studies have demonstrated that the repressed expression of KLF17 contributes to metastasis. However, KLF17 has not always been shown to suppress metastasis, which implies that it may exist as a context dependence of suppressive pathways [31].

## KLF17 is a negative regulator of EMT

Uncontrolled cell survival, growth, angiogenesis, invasion, and metastasis are essential hallmarks of cancer [32]. Metastasis is the primary cause of cancer deaths, including a complex multistep process. In the past decades, a growing number of studies have demonstrated that EMT plays a critical role in promoting metastasis [33, 34]. KLF17 is one of the negative regulators of EMT and metastasis via regulating EMT-related genes such as E-cadherin, ID1, ZO-1, β-catenin, Snai1, vimentin, and fibronectin [5, 35]. KLF17 is the human orthologue of murine Zfp393 and can activate transcription from CACCC-box by binding to a typical G/C-rich site via its zinc fingers [8].

Epithelial cells are connected laterally via several types of cellular junctions, including adherens junctions, desmosomes, and tight junctions [36]. In addition, the basal epithelial cells are firmly anchored to the underlying basement membrane via hemidesmosomes to maintain their apical-basal polarity. Altered local microenvironment and gene promote the malignant conversion of epithelial cells to activate the EMT process. Biomarkers for EMT include the increased expression of transcription factors and proteolysis [36, 37]. When EMT shift, epithelial cells lose epithelial characteristics and acquire mesenchymal characteristics, and transdifferentiate into motile mesenchymal cells, which is essential to allow carcinoma cells to lose cell-cell junctions and depart from each other for single-cell migration and invasion [36, 38]. This switch of this

program is mediated by key transcription factors, including Snail, zinc finger E-box-binding (ZEB), and basic helix-loop-helix (bHLH) transcription factors, the functions of which are finely regulated at the transcriptional, translational, and post-translational levels [33, 39–41]. The decreased level of KLF17 was correlated with reduced survival span, and the expressions of EMT-related genes were altered in HCC patient. In addition, KLF17 inhibits HCC cell invasion and migration possibly via counteracting EMT [18, 25].

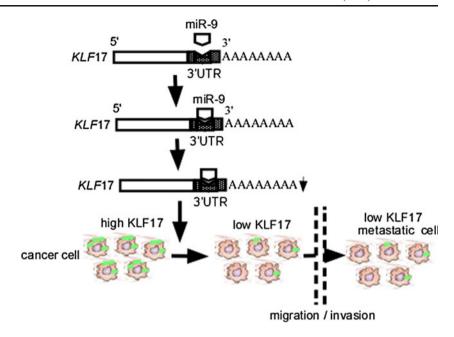
Gumireddy et al. [23] reported that knockdown KLF17 led to apparent migratory phenotype, including spindle-like and fibroblastic morphology, the major characteristics of EMT in both mouse breast and human breast cancer cells. After the knockdown of the expression of KLF17, breast cancer cells displayed the major characteristics of EMT. In addition, significantly reduced expression of epithelial markers was found in KLF17 knockdown cells, while the dramatic increase of mesenchymal markers was found in KLF17 knockdown cells [23]. KLF17 can inhibit the transcription of ID1, which is the gene of encoding a key metastasis regulator in breast cancer, via directly binding to its promoter region [23]. When transfected by siKLF17, the expression of EMT-related genes, E-cadherin, ZO-1, and vimentin, changed dramatically in HepG2 cells [18]. Sun et al. [25] found that KLF17 can bind directly to the promoter regions of ZO-1, vimentin, and fibronectin and regulate ZO-1, vimentin, and fibronectin expression, so thought that ZO-1, vimentin, and fibronectin are downstream gene targets of KLF17.

# Dyregulated expression KLF17 activates signaling transduction

miR-9 mediates KLF17 low expression As an upstream regulator of KLF17, miR-9 inhibits KLF17 expression via directly targeting its 3' untranslated region (3' UTR), resulting in migration and invasion in HCC (Fig. 1). miR-9 upregulation facilitates tumor progression in diverse human cancer, including HCC [42], Hodgkin lymphoma (HL) [43], breast cancer [44], cervical cancer [45], colon cancer [46], acute myeloid leukemia (AML) [47], and gastric cancer (GC) [48]. In contrast, miR-9 undergone hypermethylation-associated silencing is correlated with metastasis in various cancers, including colorectal cancer (CRC) [49], clear cell renal cell carcinoma (ccRCC) [50], lung cancer [51], neuroblastoma [52], and nasopharyngeal carcinoma (NPC) [53]. miR-9 is regulated by prospero homeobox 1 (PROX1), a tumor suppressor. Lu and colleagues [46] further confirmed that PROX1 promotes EMT by inhibiting E-cadherin via binding to miR-9 promoter in colon cancer cells. Taken together, these studies show that miR-9 could act as an oncogene and promote the progression of HCC via KLF17.



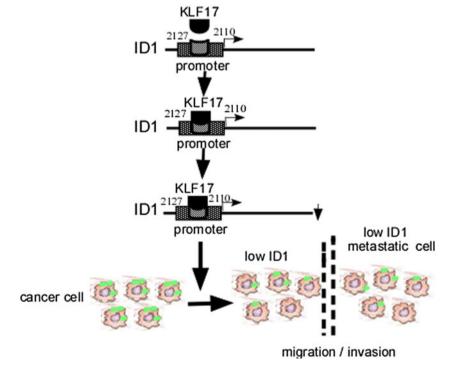
Fig. 1 Schematic illustration of miR-9-mediated KLF17 low expression. miR-9 downregulates KLF17 expression through binding to 3' UTR of *KLF*17 gene, increases cell migration and invasion. KLF17, Krüppel-like factor 17; 3'UTR, 3' untranslated region; miR-9, microRNA-9



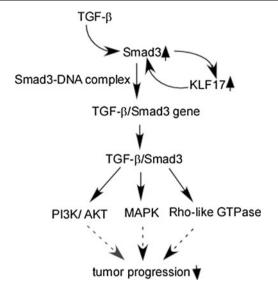
Low KLF17 mediates ID1 increase ID1 is one member of the vertebrate inhibitors of differentiation family and a negative regulator of bHLH transcription factors. ID1 expression exhibits a unique spatio-temporal pattern during development and malignancy [54, 55]. ID1 has been shown to play a critical role in the diverse biological process, including cell cycle, proliferation, apoptosis, senescence, and metastasis [56–58]. Gumireddy reported that KLF17 could directly bind to the mouse ID1 promoter region at -2127 to -2110 from the

transcriptional initiation site and suppress the expression of ID1 [23] (Fig. 2). Elevated levels of ID1 protein have been reported in a variety of human cancers and are capable of promoting invasion and metastasis [58–60]. On the other hand, the expression of ID1 is significantly correlated with high grade and poor prognosis of human cancer [61]. ID1 plays a critical role in tumor maintenance of high-grade glioma (HGG), and deletion of ID1 and ID3 in vitro reduces invasiveness of HGG [62]. ID proteins are able to interact with

Fig. 2 Schematic illustration of KLF17-mediated ID1 low expression. KLF17 downregulates ID1 expression through binding to *ID*1 gene promoter and increases cell migration and invasion. *ID1*, inhibitor of DNA binding 1

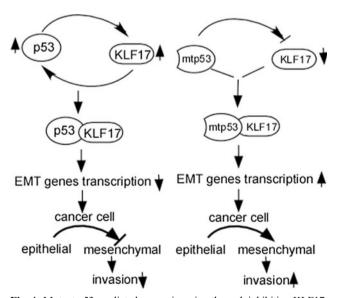




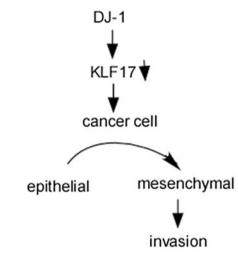


**Fig. 3** TGF-β signaling pathway mediates KLF17 expression. TGF-β enhances KLF17 expression through Smad3 as well KLF17 induces Smad3 to generate a positive feedback loop, regulates a panel of TGF-β/Smad3-dependent target genes by modulating Smad3-DNA complex formation, and finally decreases tumor progression though PI3K/AKT, MAPK, and Rho-like GTPase. TGF-β transforming growth factor β, PI3K phosphatidylinositol-3-kinase, MAPK mitogen-activated protein kinase

bHLH proteins as homodimer or heterodimer and negatively regulate bHLH proteins [63]. Overexpression of ID1 enhances metastatic potential human thyroid tumors, which lets the thyroid tumor cells acquire the mesenchymal features [64]. Moreover, Gumireddy and colleagues found that ID1



**Fig. 4** Mutant p53-mediated cancer invasion through inhibiting KLF17. p53 enhances KLF17 transcription and KLF17 and enhances p53 transcription to generate a positive feedback loop. Mutant p53 potentiates cancer progression through inhibition of KLF17 expression. Mutant p53 protein develops a complex with KLF17, leading to inhibit KLF17-mediated EMT gene transcription. *mtp53* mutant p53, *EMT* epithelial-mesenchymal transition



**Fig. 5** Schematic illustration of DJ-1-mediated low KLF17 expression. DJ-1 decreases KLF17 expression, promotes cell transition from epithelial to mesenchymal, and increases cell invasion. *DJ-1* parkinson protein 7

interacted with transcription factor AP-2α (TFAP2A) to suppress S100A9 expression, leading to migratory and invasive phenotypes of cancer cells [65]. ID1 is also able to inhibit mp53-mediated endothelial cell migration and tube formation [66]. ID1 upregulates mouse double minute 2 (MDM2) expression, a key negative regulator of p53, in esophageal cancer cells [67]. Cooperating with oncogenic Ras, ID1 triggers metastatic transformation of mammary carcinoma [68]. By inducing the expression of matrix metalloproteinase (MMP) 9, ID1 promotes the invasiveness of breakpoint cluster region/ Abelson (BCR/ABL) leukemia cells [69]. The overexpression of ID1 induces a significantly increased secretion of the active form of MMP2 [70] and upregulates vascular endothelial growth factor (VEGF) to promote angiogenesis in prostate cancer [71]. Additionally, ID1 can downregulate zinc finger binding protein 89 (ZBP-89), leading to mesenchymal markers' expression and finally promoting NSCLC metastasis [72]. Taken together, these studies indicate that suppressed KLF17 in human cancers promotes metastasis through inducing ID1 (Fig. 2).

**KLF17** is a downstream mediator of the TGF- $\beta$  signaling pathway TGF- $\beta$  pathway is one of the most deregulated pathways, which is intimately associated with the induction of EMT during heart development, renal fibrosis, and cancer [73–75]. TGF- $\beta$  signaling can switch breast cancer cells from cohesive to single-cell motility and ultimately contribute to metastasis [76]. A recent study shows that KLF17 is a key regulator of TGF- $\beta$  signaling pathway, and it has the capacity to suppress tumor progression through potentiating TGF- $\beta$ /Smad3-dependent signaling pathway [30]. TGF- $\beta$  enhances KLF17 expression in multiple cancer cells via Smad3; KLF17 induces Smad3 to generate a positive feedback loop, which



regulates a panel of TGF-β/Smad3-dependent target genes by modulating Smad3-DNA complex formation [30] (Fig. 3). So far, three types of Smads have been identified, such as receptor-regulated Smads (R-Smads), common-partner Smads (Co-Smads), and inhibitory Smads (I-Smads) [77]. In the canonical pathway, R-Smads are directly phosphorylated by the activated type I TGF-β receptors (TβRI). Receptormediated phosphorylation of R-Smads together with Smad4 induces their accumulation in the nucleus, where they interact with other transcription factors to regulate transcriptional responses [73, 78]. As a tumor suppressor pathway, TGF-β signaling is well illustrated by modulation of receptors and Smads in cancers, which is further supported by studies of cancer development in mouse models [79]. In the noncanonical pathway, TGF-β signal is transducted via three pathways including mitogen-activated protein kinase (MAPK) pathways, Rho-like GTPase signaling pathways, and phosphatidylinositol-3-kinase (PI3K)/Akt pathways [80] (Fig. 3).

### KLF17 functions via p53-dependent pathway

Wild-type p53, a major tumor suppressor, participates in diverse cellular stress stimuli, including ribosomal stress, nutrient depletion, viral infection, oncogenes activation and hypoxia, and heat shock [81, 82]. As a critical regulator of metastasis, p53 directly regulates the transcription of metastatic genes including EMT and stemness genes, interacts with ECM and anoikis, and inhibits cancer metastasis [83] (Fig. 4). However, mutant p53 loses tumor suppressor activity and gains functions, and contributes to malignant progression [84–86]. Recent studies have shown that KLF17 is involved in p53 pathway. Ali and colleagues [22] showed that KLF17 exerted an anti-EMT effect via the p53-dependent pathway in NSCLC. KLF17 mRNA levels were induced in a dosedependent manner in siRNA targeting p53 A549 cells with Nutlin-3 treatment, but there was not KLF17 transcription in p53 depleted cells [22]. p53 enhances KLF17 transcription, and KLF17 enhances p53 transcription to generate a positive feedback loop (Fig. 4). Furthermore, p53 interacts with KLF17 promoter via p53 consensus responsive element (p53RE) and recruits p300 in response to chemotherapy [22]. In contrast, mutant p53 potentiates cancer progression through KLF17 inhibition via recruiting to the upstream of KLF17 promoter in metastatic breast cancer. In addition, endogenous mutant p53 proteins are capable of developing a complex with KLF17, which inhibits KLF17-mediating EMT gene transcription (Fig. 4). The metastasis suppressor ability of KLF17 to EMT target genes is enhanced when mutant p53 is depleted [19]. Taken together, these studies suggest that suppressed KLF17 leads to weaken the tumor suppressor strength of p53.



DJ-1 [anti-oxidant protein encoded by *PARK7* gene (Parkinson protein 7 gene)] is a conserved protein ubiquitously expressed in human tissues, which is a human oncogene identified in 1997 [87]. DJ-1 is overexpressed in a variety of human cancers and correlated with poor prognosis [88]. The increased levels of DJ-1 are detected in the nipple fluid of breast cancer patients [89]. It has been shown that DJ-1 is also implicated in multiple cellular processes, including cell proliferation, invasion, and metastasis [90, 91]. Recently, it is reported that DJ-1 is highly expressed in invasive breast cancer cell and is able to re-

press the expression of KLF17 to promote breast cancer cell

invasion by downregulating E-cadherin and increasing Snail

expression (Fig. 5); moreover, DJ-1 could directly regulate

KLF17 by binding to the ID1 promoter [24]. Taken together,

DJ-1 could promote the invasion of breast cancer cells via

KLF17 may be a downstream signaling molecule of DJ-1

### **Conclusions**

regulating the KLF17/ID1 pathway.

Since KLF17 was at first identified as a tumor suppressor, an increasing number of studies have reported that KLF17 is frequently downregulated, which is correlated with tumor progression in various human cancers. KLF17 low expression promotes metastasis; its mechanism is to directly increase cell invasion and initiate EMT shift though regulating EMT gene expression. Additionally, the reduced-KLF17 in human cancer is involved in TGF- $\beta$  pathway and p53 pathway. However, tumor metastasis is a result of many genes' concerted action, the precise mechanism of KLF17-involved metastasis is still incomplete, and much more challenges for researchers need to be overcome.

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**Author contributions** Shanzhou and Xiaowei Tang contributed to the analysis and interpretation of data and drafting of the manuscript. Faqing Tang contributed to the study concept and design, analysis, and drafting of the manuscript.

#### Compliance with ethical standards

Conflicts of interest None

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#### References

- Limame R, de Op Beeck K, Lardon F, De Wever O, Pauwels P. Kruppel-like factors in cancer progression: three fingers on the steering wheel. Oncotarget. 2014;5:29–48.
- Tetreault MP, Yang Y, Katz JP. Kruppel-like factors in cancer. Nat Rev Cancer. 2013;13:701–13.
- Carlson CM, Endrizzi BT, Wu J, Ding X, Weinreich MA, Walsh ER, et al. Kruppel-like factor 2 regulates thymocyte and T-cell migration. Nature. 2006;442:299–302.
- Kaczynski J, Cook T, Urrutia R. Sp1- and Kruppel-like transcription factors. Genome Biol. 2003;4:206.
- Lu XJ, Shi Y, Chen JL, Ma S. Kruppel-like factors in hepatocellular carcinoma. Tumour Biol. 2015;36:533

  –41.
- Ghaleb AM, Nandan MO, Chanchevalap S, Dalton WB, Hisamuddin IM, Yang VW. Kruppel-like factors 4 and 5: the yin and yang regulators of cellular proliferation. Cell Res. 2005;15:92–
- Yan W, Burns KH, Ma L, Matzuk MM. Identification of zfp393, a germ cell-specific gene encoding a novel zinc finger protein. Mech Dev. 2002;118:233–9.
- van Vliet J, Crofts LA, Quinlan KG, Czolij R, Perkins AC, Crossley M. Human KLF17 is a new member of the Sp/KLF family of transcription factors. Genomics. 2006;87:474–82.
- Iuchi S. Three classes of c2h2 zinc finger proteins. Cell Mol Life Sci. 2001;58:625–35.
- Schuh R, Aicher W, Gaul U, Cote S, Preiss A, Maier D, et al. A conserved family of nuclear proteins containing structural elements of the finger protein encoded by Kruppel, a Drosophila segmentation gene. Cell. 1986;47:1025–32.
- Tsukamoto K, Ito N, Yoshimoto M, Kasumi F, Akiyama F, Sakamoto G, et al. Allelic loss on chromosome 1p is associated with progression and lymph node metastasis of primary breast carcinoma. Cancer. 1998;82:317–22.
- Narla G, Heath KE, Reeves HL, Li D, Giono LE, Kimmelman AC, et al. KLF6, a candidate tumor suppressor gene mutated in prostate cancer. Science. 2001;294:2563–6.
- Camacho-Vanegas O, Narla G, Teixeira MS, DiFeo A, Misra A, Singh G, et al. Functional inactivation of the KLF6 tumor suppressor gene by loss of heterozygosity and increased alternative splicing in glioblastoma. Int J Cancer. 2007;121:1390–5.
- Takahashi S, Licht JD. The human promyelocytic leukemia zinc finger gene is regulated by the Evi-1 oncoprotein and a novel guanine-rich site binding protein. Leukemia. 2002;16:1755–62.
- Jo BB, Jeong MS, Park SY, Cheong J, Jang SB. The binding of hepatitis B virus x protein to glioma-associated oncogene homologue 1 and its biological characterization in vitro. Appl Biochem Biotechnol. 2011;165:109–22.
- Antin PB, Pier M, Sesepasara T, Yatskievych TA, Darnell DK. Embryonic expression of the chicken Kruppel-like (KLF) transcription factor gene family. Dev Dyn. 2010;239:1879–87.
- Cai XD, Zhou YB, Huang LX, Zeng QL, Zhang LJ, Wang QQ, et al. Reduced expression of Kruppel-like factor 17 is related to tumor growth and poor prognosis in lung adenocarcinoma. Biochem Biophys Res Commun. 2012;418:67–73.
- Liu FY, Deng YL, Li Y, Zeng D, Zhou ZZ, Tian DA, et al. Downregulated KLF17 expression is associated with tumor invasion and poor prognosis in hepatocellular carcinoma. Med Oncol. 2013;30: 425.
- Ali A, Shah AS, Ahmad A. Gain-of-function of mutant p53: mutant p53 enhances cancer progression by inhibiting KLF17 expression in invasive breast carcinoma cells. Cancer Lett. 2014;354:87–96.
- Peng JJ, Wu B, Xiao XB, Shao YS, Feng Y, Yin MX. Reduced Kruppel-like factor 17 (KLF17) expression correlates with poor

- survival in patients with gastric cancer. Arch Med Res. 2014;45: 394-9
- Ye WC, Gao L, Huang J, Fang XM, Xie G. Suppressed Kruppel like factor 17 expression induces tumor proliferation, metastasis and a poor prognosis in papillary thyroid carcinoma. Mol Med Rep. 2014;10:2087–92.
- Ali A, Bhatti MZ, Shah AS, Duong HQ, Alkreathy HM, Mohammad SF, et al. Tumor-suppressive p53 signaling empowers metastatic inhibitor KLF17-dependent transcription to overcome tumorigenesis in non-small cell lung cancer. J Biol Chem. 2015;290:21336–51.
- Gumireddy K, Li A, Gimotty PA, Klein-Szanto AJ, Showe LC, Katsaros D, et al. KLF17 is a negative regulator of epithelialmesenchymal transition and metastasis in breast cancer. Nat Cell Biol. 2009;11:1297–304.
- Ismail IA, Kang HS, Lee HJ, Kim JK, Hong SH. Dj-1 upregulates breast cancer cell invasion by repressing KLF17 expression. Br J Cancer. 2014;110:1298–306.
- Sun Z, Han Q, Zhou N, Wang S, Lu S, Bai C, et al. MicroRNA-9 enhances migration and invasion through KLF17 in hepatocellular carcinoma. Mol Oncol. 2013;7:884–94.
- Wang N, Liu ZH, Ding F, Wang XQ, Zhou CN, Wu M. Downregulation of gut-enriched Kruppel-like factor expression in esophageal cancer. World J Gastroenterol. 2002;8:966–70.
- Nguyen DX, Bos PD, Massague J. Metastasis: from dissemination to organ-specific colonization. Nat Rev Cancer. 2009;9:274

  –84.
- Steeg PS. Tumor metastasis: mechanistic insights and clinical challenges. Nat Med. 2006;12:895–904.
- Weinberg RA. Mechanisms of malignant progression. Carcinogenesis. 2008;29:1092–5.
- Ali A, Zhang P, Liangfang Y, Wenshe S, Wang H, Lin X, et al. KLF17 empowers TGF-beta/Smad signaling by targeting Smad3dependent pathway to suppress tumor growth and metastasis during cancer progression. Cell Death Dis. 2015;6, e1681.
- Dong P, Kaneuchi M, Xiong Y, Cao L, Cai M, Liu X, et al. Identification of KLF17 as a novel epithelial to mesenchymal transition inducer via direct activation of TWIST1 in endometrioid endometrial cancer. Carcinogenesis. 2014;35:760–8.
- Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. 2011;144:646–74.
- Tsai JH, Yang J. Epithelial-mesenchymal plasticity in carcinoma metastasis. Genes Dev. 2013;27:2192–206.
- Thiery JP, Acloque H, Huang RY, Nieto MA. Epithelialmesenchymal transitions in development and disease. Cell. 2009;139:871–90.
- 35. Iwanicki MP, Brugge JS. Transcriptional regulation of metastatic [id]entity by KLF17. Genome Biol. 2009;10:244.
- Nelson WJ. Remodeling epithelial cell organization: transitions between front-rear and apical-basal polarity. Cold Spring Harb Perspect Biol. 2009;1, a000513.
- Zeisberg M, Neilson EG. Biomarkers for epithelial-mesenchymal transitions. J Clin Invest. 2009;119:1429–37.
- Huang RY, Guilford P, Thiery JP. Early events in cell adhesion and polarity during epithelial-mesenchymal transition. J Cell Sci. 2012;125:4417–22.
- Puisieux A, Brabletz T, Caramel J. Oncogenic roles of EMTinducing transcription factors. Nat Cell Biol. 2014;16:488–94.
- Lamouille S, Xu J, Derynck R. Molecular mechanisms of epithelial-mesenchymal transition. Nat Rev Mol Cell Biol. 2014;15:178–96.
- De Craene B, Berx G. Regulatory networks defining EMT during cancer initiation and progression. Nat Rev Cancer. 2013;13:97– 110.
- Tan HX, Wang Q, Chen LZ, Huang XH, Chen JS, Fu XH, et al. MicroRNA-9 reduces cell invasion and e-cadherin secretion in SK-Hep-1 cell. Med Oncol. 2010;27:654–60.



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- Leucci E, Zriwil A, Gregersen LH, Jensen KT, Obad S, Bellan C, et al. Inhibition of miR-9 de-represses HuR and DICER1 and impairs Hodgkin lymphoma tumour outgrowth in vivo. Oncogene. 2012;31:5081–9.
- Ma L, Young J, Prabhala H, Pan E, Mestdagh P, Muth D, et al. Mir-9, a myc/mycn-activated microRNA, regulates e-cadherin and cancer metastasis. Nat Cell Biol. 2010;12:247–56.
- Wilting SM, Snijders PJ, Verlaat W, Jaspers A, van de Wiel MA, van Wieringen WN, et al. Altered microRNA expression associated with chromosomal changes contributes to cervical carcinogenesis. Oncogene. 2013;32:106–16.
- Lu MH, Huang CC, Pan MR, Chen HH, Hung WC. Prospero homeobox 1 promotes epithelial-mesenchymal transition in colon cancer cells by inhibiting e-cadherin via miR-9. Clin Cancer Res. 2012;18:6416–25.
- Chen P, Price C, Li Z, Li Y, Cao D, Wiley A, et al. Mir-9 is an essential oncogenic microRNA specifically overexpressed in mixed lineage leukemia-rearranged leukemia. Proc Natl Acad Sci U S A. 2013;110:11511–6.
- Rotkrua P, Akiyama Y, Hashimoto Y, Otsubo T, Yuasa Y. Mir-9 downregulates CDX2 expression in gastric cancer cells. Int J Cancer. 2011;129:2611–20.
- Bandres E, Agirre X, Bitarte N, Ramirez N, Zarate R, Roman-Gomez J, et al. Epigenetic regulation of microRNA expression in colorectal cancer. Int J Cancer. 2009;125:2737–43.
- Hildebrandt MA, Gu J, Lin J, Ye Y, Tan W, Tamboli P, et al. HSA-miR-9 methylation status is associated with cancer development and metastatic recurrence in patients with clear cell renal cell carcinoma. Oncogene. 2010;29:5724

  –8.
- Heller G, Weinzierl M, Noll C, Babinsky V, Ziegler B, Altenberger C, et al. Genome-wide miRNA expression profiling identifies miR-9-3 and miR-193a as targets for DNA methylation in non-small cell lung cancers. Clin Cancer Res. 2012;18:1619–29.
- Zhang H, Qi M, Li S, Qi T, Mei H, Huang K, et al. MicroRNA-9 targets matrix metalloproteinase 14 to inhibit invasion, metastasis, and angiogenesis of neuroblastoma cells. Mol Cancer Ther. 2012;11:1454–66.
- Lu J, Luo H, Liu X, Peng Y, Zhang B, Wang L, et al. Mir-9 targets CXCR4 and functions as a potential tumor suppressor in nasopharyngeal carcinoma. Carcinogenesis. 2014;35:554–63.
- Jen Y, Manova K, Benezra R. Each member of the ID gene family exhibits a unique expression pattern in mouse gastrulation and neurogenesis. Dev Dyn. 1997;208:92–106.
- Perk J, Gil-Bazo I, Chin Y, de Candia P, Chen JJ, Zhao Y, et al. Reassessment of ID1 protein expression in human mammary, prostate, and bladder cancers using a monospecific rabbit monoclonal anti-ID1 antibody. Cancer Res. 2006;66:10870–7.
- Ling F, Kang B, Sun XH. Id proteins: small molecules, mighty regulators. Curr Top Dev Biol. 2014;110:189–216.
- Mern DS, Hoppe-Seyler K, Hoppe-Seyler F, Hasskarl J, Burwinkel B. Targeting ID1 and ID3 by a specific peptide aptamer induces e-box promoter activity, cell cycle arrest, and apoptosis in breast cancer cells. Breast Cancer Res Treat. 2010;124:623–33.
- Perk J, Iavarone A, Benezra R. ID family of helix-loop-helix proteins in cancer. Nat Rev Cancer. 2005;5:603–14.
- Nair R, Teo WS, Mittal V, Swarbrick A. ID proteins regulate diverse aspects of cancer progression and provide novel therapeutic opportunities. Mol Ther. 2014;22:1407–15.
- Lasorella A, Benezra R, Iavarone A. The ID proteins: Master regulators of cancer stem cells and tumour aggressiveness. Nat Rev Cancer. 2014;14:77–91.
- Sharma P, Patel D, Chaudhary J. ID1 and ID3 expression is associated with increasing grade of prostate cancer: ID3 preferentially regulates CDKN1b. Cancer Med. 2012;1:187–97.

- Niola F, Zhao X, Singh D, Sullivan R, Castano A, Verrico A, et al. Mesenchymal high-grade glioma is maintained by the ID-RAP1 axis. J Clin Invest. 2013;123:405–17.
- Benezra R, Davis RL, Lockshon D, Turner DL, Weintraub H. The protein ID: a negative regulator of helix-loop-helix DNA binding proteins. Cell. 1990;61:49–59.
- Ciarrocchi A, Piana S, Valcavi R, Gardini G, Casali B. Inhibitor of DNA binding-1 induces mesenchymal features and promotes invasiveness in thyroid tumour cells. Eur J Cancer. 2011;47:934

  45.
- Gumireddy K, Li A, Kossenkov AV, Cai KQ, Liu Q, Yan J, et al. ID1 promotes breast cancer metastasis by s100a9 regulation. Mol Cancer Res. 2014;12:1334–43.
- Qiu J, Wang G, Hu J, Peng Q, Zheng Y. ID1-induced inhibition of p53 facilitates endothelial cell migration and tube formation by regulating the expression of beta1-integrin. Mol Cell Biochem. 2011;357:125–33.
- Hui CM, Cheung PY, Ling MT, Tsao SW, Wang X, Wong YC, et al. ID-1 promotes proliferation of p53-deficient esophageal cancer cells. Int J Cancer. 2006;119:508–14.
- Swarbrick A, Roy E, Allen T, Bishop JM. ID1 cooperates with oncogenic Ras to induce metastatic mammary carcinoma by subversion of the cellular senescence response. Proc Natl Acad Sci U S A. 2008;105:5402–7.
- Nieborowska-Skorska M, Hoser G, Rink L, Malecki M, Kossev P, Wasik MA, et al. ID1 transcription inhibitor-matrix metalloproteinase 9 axis enhances invasiveness of the breakpoint cluster region/ Abelson tyrosine kinase-transformed leukemia cells. Cancer Res. 2006;66:4108–16.
- Coppe JP, Itahana Y, Moore DH, Bennington JL, Desprez PY. ID-1 and ID-2 proteins as molecular markers for human prostate cancer progression. Clin Cancer Res. 2004;10:2044–51.
- Ling MT, Lau TC, Zhou C, Chua CW, Kwok WK, Wang Q, et al. Overexpression of ID-1 in prostate cancer cells promotes angiogenesis through the activation of vascular endothelial growth factor (VEGF). Carcinogenesis. 2005;26:1668–76.
- Pillai S, Rizwani W, Li X, Rawal B, Nair S, Schell MJ, et al. ID1 facilitates the growth and metastasis of non-small cell lung cancer in response to nicotinic acetylcholine receptor and epidermal growth factor receptor signaling. Mol Cell Biol. 2011;31:3052–67.
- Derynck R, Muthusamy BP, Saeteurn KY. Signaling pathway cooperation in TGF-beta-induced epithelial-mesenchymal transition. Curr Opin Cell Biol. 2014;31:56–66.
- 74. Massague J. TGF beta in cancer. Cell. 2008;134:215–30.
- Derynck R, Akhurst RJ. Differentiation plasticity regulated by TGF-beta family proteins in development and disease. Nat Cell Biol. 2007;9:1000–4.
- Giampieri S, Manning C, Hooper S, Jones L, Hill CS, Sahai E. Localized and reversible TGFbeta signalling switches breast cancer cells from cohesive to single cell motility. Nat Cell Biol. 2009;11: 1287–96.
- 77. Samanta D, Datta PK. Alterations in the Smad pathway in human cancers. Front Biosci (Landmark Ed). 2012;17:1281–93.
- Shi Y, Massague J. Mechanisms of TGF-beta signaling from cell membrane to the nucleus. Cell. 2003;113:685–700.
- Bae E, Sato M, Kim RJ, Kwak MK, Naka K, Gim J, et al. Definition of Smad3 phosphorylation events that affect malignant and metastatic behaviors in breast cancer cells. Cancer Res. 2014;74:6139–49.
- van Leeuwen I, Lain S. Sirtuins and p53. Adv Cancer Res. 2009;102:171–95.
- Meek DW. Regulation of the p53 response and its relationship to cancer. Biochem J. 2015;469:325–46.
- Zilfou JT, Lowe SW. Tumor suppressive functions of p53. Cold Spring Harb Perspect Biol. 2009;1, a001883.
- Powell E, Piwnica-Worms D, Piwnica-Worms H. Contribution of p53 to metastasis. Cancer Discovery. 2014;4:405–14.



- Muller PA, Vousden KH. Mutant p53 in cancer: new functions and therapeutic opportunities. Cancer Cell. 2014;25:304–17.
- 85. Freed-Pastor WA, Prives C. Mutant p53: one name, many proteins. Genes Dev. 2012;26:1268–86.
- Oren M, Rotter V. Mutant p53 gain-of-function in cancer. Cold Spring Harb Perspect Biol. 2010;2:a001107.
- 87. Nagakubo D, Taira T, Kitaura H, Ikeda M, Tamai K, Iguchi-Ariga SM, et al. Dj-1, a novel oncogene which transforms mouse NIH3t3 cells in cooperation with Ras. Biochem Biophys Res Commun. 1997;231:509–13.
- Cao J, Lou S, Ying M, Yang B. DJ-1 as a human oncogene and potential therapeutic target. Biochem Pharmacol. 2015;93:241–50.
- 89. Oda M, Makita M, Iwaya K, Akiyama F, Kohno N, Tsuchiya B, et al. High levels of DJ-1 protein in nipple fluid of patients with breast cancer. Cancer Sci. 2012;103:1172–6.
- He X, Zheng Z, Li J, Ben Q, Liu J, Zhang J, et al. DJ-1 promotes invasion and metastasis of pancreatic cancer cells by activating SRC/ERK/uPA. Carcinogenesis. 2012;33:555–62.
- Chang YH, Lee SH, Chang HC, Tseng YL, Lai WW, Liao CC, et al. Comparative secretome analyses using a hollow fiber culture system with label-free quantitative proteomics indicates the influence of PARK7 on cell proliferation and migration/invasion in lung adenocarcinoma. J Proteome Res. 2012;11:5167–85.

